

Ecological conditions favoring species recognition and rapid mate pairing are associated with greater plumage sexual dichromatism in true thrushes (genus: *Turdus*)

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Abstract

Keywords

Background

Methods

Plumage sexual dichromatism

A total of N=77 *Turdus* thrush species were sampled for plumage spectral reflectance using from bird skins at the American Museum of Natural History in New York City and the Field Museum in Chicago. Reflectance measurements from 300-700nm were taken in triplicate for the belly, breast, throat, crown and mantle plumage patches [1] of each individual bird skin. N=3 male and N=3 female individuals were measured for most species (exceptions: *Turdus lawrencii*, N=2 males and N=2 females; *Turdus swalesi*, N=1 male and N=1 female). Reflectance spectra were measured using a 400 µm fiber optic reflection probe fitted with a rubber stopper to maintain a consistent measuring distance of 3 mm and area of 2 mm² at a 90° angle to the surface of the feather patch. Measurements were taken using a JAZ spectrometer with a pulsed-xenon light source (Ocean Optics, Dunedin, USA) and all measurements were made relative to a diffuse reflectance white standard (Spectralon WS-1-SL, Labsphere, North Sutton NH, USA).

We used a receptor-noise limited visual model [2] of the European Blackbird (*Turdus merula*) visual system [3] in the *pavo* [4] package in R v4.0.0 [5] to calculate avian-perceived chromatic and achromatic visual contrast (in units of “Just-Noticeable Differences”, or JNDs) of male vs. female plumage patches for all sampled *Turdus* species. Chromatic and achromatic JNDs were calculated for male-female pairs within each species (i.e., N=9 JND values calculated per patch for each species where N=3 males and N=3 females sampled), and then JND values were averaged for each species’ respective plumage patches. Under ideal laboratory conditions, a JND value of 1 is generally considered to be the discriminable threshold past which an observer is predicted to be able to perceive the two colors as different. However, nat-

ural light environments vary both spatially and temporally [6], bringing into question the accuracy of a JND=1 threshold for generalizing visual contrast under natural conditions. Therefore, we calculated the total number of sexually-dichromatic plumage patches per species (out of N=5 measured patches) as the number of plumage patches with average JND values > 1, 2, or 3 to account for uncertainty in visual discrimination thresholds due to variation in psychophysical and ambient lighting conditions affecting the strength of between-sex plumage visual contrast [7].

Life History Data

Breeding Timing Model

We collected data on migration behavior and breeding season length from *Thrushes* [8] and the *Handbook of the Birds of the World* [9]. We assigned three different kinds of migratory behavior: 1) *full migration* when a species description clearly stated that a species “migrates”, 2) *partial migration* when a species was described to have “altitudinal migration”, “latitudinal migration” or “movement during non-breeding season”, or 3) *sedentary* when a species was described as “resident” or “sedentary”. Breeding season length was defined as the number of months the species breeds.

Breeding Sympatry Model

Species’ breeding ranges were acquired from *BirdLife International* [10]. We calculated congener breeding range overlaps (as percentages) using the *letsR* package in R [11]. We then calculated the number of sympatric species as the number of congeners with breeding ranges that overlap >30% with the focal species’ breeding range [12].

Breeding Spacing Model

Species’ breeding range sizes (in km²) were acquired using the *BirdLife International* breeding range maps. Species’ island vs. mainland residence was also determined using breeding ranges from *BirdLife International*. Mainland residence was assigned if the species had a breeding range on any continent and Japan. Island residence was assigned to species having a breeding range limited to a non-continental landmass entirely surrounded by an oceanic body of water.

Statistical Modeling

We used phylogenetically-corrected Bayesian multilevel logistic regression models using the *brms* v2.13.0 package [13] in R v4.0.0 [5] where responses, the number of sexually-dichromatic patches >1, 2, and 3 chromatic and achromatic JNDs, were modeled as binomial trials (N=5 plumage patch “trials”) to test for associations with breeding timing, breeding sympatry and breeding spacing. For all phylogenetically-corrected models, we used the *Turdus* phylogeny from Nylander et al. (2008) [14] to create a covariance matrix of species’ phylogenetic relationships. All models used a dataset of N=67 *Turdus* species for which all data were available.

Our *breeding timing* models included the following predictors: z-scores of breeding season length (mean centered and divided by one standard deviation), migratory behavior (full migration as the reference category versus partial migration or sedentary), and their interaction. *Breeding sympatry* models included the number of sympatric species with greater than 30% breeding range overlap as the only predictor of the number of sexually-dichromatic plumage patches. *Breeding spacing* models included \log_e transformed breeding range size (km²) and breeding landmass (mainland as the reference category versus island). We also ran null models (intercept only) for all responses. All models' intercepts and response standard deviations were assigned a weak prior (Student T: df = 3, location = 0, scale = 10), and predictor coefficients were assigned flat priors. We ran each model for 6,000 iterations across 6 chains and assessed Markov Chain Monte Carlo (MCMC) convergence using the Gelman-Rubin diagnostic (Rhat) [15]. We then performed k-fold cross-validation [16] to refit each model K=16 times. For each k-fold, the training dataset included a randomly selected set of $N - N \frac{1}{K}$ or N≈63 species, and the testing dataset included $N \frac{1}{K}$ or N≈4 species not included in the training dataset. Finally, we compared differences between the models' expected log pointwise predictive densities (ELPD) to assess which model(s) best predicted the number of sexually-dichromatic plumage patches [16].

Results

We obtained $N \geq 4000$ effective samples for each model parameter and all models' Markov Chains (MCMC) successfully converged (Rhat = 1 for all models' parameters).

Table 1: Expected log pointwise predictive densities (ELPD) differences and kfold information criterion values of models.

		Model			
Plumage Metric	JND Threshold	Breeding Sympatry	Breeding Timing	Breeding Spacing	Intercept Only
Achromatic					
	1 JND	0 ± 0 (-122.17 ± 0.67)	-2.51 ± 2.49 (-124.68 ± 2.38)	-2.59 ± 1.01 (-124.76 ± 1.04)	-21.69 ± 7.36 (-143.87 ± 7.51)
	2 JND	0 ± 0 (-98.94 ± 7.56)	-1.19 ± 3.95 (-100.13 ± 9.22)	-0.7 ± 1.34 (-99.64 ± 7.92)	-52.42 ± 12.67 (-151.36 ± 13.4)
	3 JND	-0.04 ± 1.4 (-85.4 ± 8.91)	-1.7 ± 4.41 (-87.07 ± 10.71)	0 ± 0 (-85.37 ± 8.76)	-28.54 ± 10.02 (-113.91 ± 13.65)
Chromatic					
	1 JND	0 ± 0 (-115.75 ± 2.95)	-5.67 ± 3.55 (-121.42 ± 2.28)	-2.73 ± 3.4 (-118.49 ± 2.67)	-14.8 ± 7.22 (-130.55 ± 7.05)
	2 JND	0 ± 0 (-88.47 ± 8.77)	-3.8 ± 4.46 (-92.27 ± 10.01)	-3.32 ± 5.29 (-91.79 ± 10.91)	-50.53 ± 14.49 (-139 ± 16.77)
	3 JND	0 ± 0 (-62.77 ± 10.41)	-8 ± 4.32 (-70.77 ± 12.29)	-4.43 ± 3.9 (-67.2 ± 11.72)	-47.63 ± 15.34 (-110.4 ± 20.01)

Table 2: Model predictor effect estimates (posterior median log-odds) on the number of achromatic and chromatic plumage patches with visual contrast values > 1, 2, and 3 JND. Model effects with a probability of direction (pd) value ≥

0.90 are bolded in red for a negative effect and blue for a positive effect on plumage dichromatism.

Discussion

Conclusions

Acknowledgements

References

1. Andersson S, Prager M. 2006 Quantifying Colors. In *Bird coloration, Volume 1: Mechanisms and Measurements* (eds GE Hill, KJ McGraw), pp. 76–77. Cambridge, MA: Harvard University Press.
2. Vorobyev M, Osorio D. 1998 Receptor noise as a determinant of colour thresholds. *Proceedings. Biological sciences / The Royal Society* **265**, 351–8. (doi:[10.1098/rspb.1998.0302](https://doi.org/10.1098/rspb.1998.0302))
3. Hart NS, Partridge JC, Cuthill IC, Bennett AT. 2000 Visual pigments, oil droplets, ocular media and cone photoreceptor distribution in two species of passerine bird: The blue tit (*Parus caeruleus* L.) And the blackbird (*Turdus merula* L.). *Journal of comparative physiology. A, Sensory, neural, and behavioral physiology* **186**, 375–387. (doi:[10.1007/s003590050437](https://doi.org/10.1007/s003590050437))
4. Maia R, Gruson H, Endler JA, White TE. 2019 Pavo 2: New tools for the spectral and spatial analysis of colour in r. *Methods in Ecology and Evolution* **10**, 1097–1107. (doi:[10.1111/2041-210X.13174](https://doi.org/10.1111/2041-210X.13174))
5. R Core Team. 2020 *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing.
6. Endler JA, Monographs E, Feb N. 1993 The Color of Light in Forests and Its Implications. *Ecological Monographs* **63**, 1–27. (doi:[10.2307/2937121](https://doi.org/10.2307/2937121))
7. Kemp DJ, Herberstein ME, Fleishman LJ, Endler JA, Bennett ATD, Dyer AG, Hart NS, Marshall J, Whiting MJ. 2015 An Integrative Framework for the Appraisal of Coloration in Nature. *The American Naturalist* **185**, 705–724. (doi:[10.1086/681021](https://doi.org/10.1086/681021))
8. Clement P, Hathway R. 2000 *Thrushes*. London: A&C Black Publishers Ltd.
9. del Hoyo J, Elliott A, Sargatal J, Christie DA, de Juana E. 2017 *Handbook of the birds of the world alive*.
10. BirdLife International and Handbook of the Birds of the World. 2018 *Bird species distribution maps of the world. Version 2018.1*.
11. Vilela B, Villalobos F. 2015 letsR: A new R package for data handling and analysis in macroecology. *Methods in Ecology and Evolution* **6**, 1229–1234. (doi:[10.1111/2041-210X.12401](https://doi.org/10.1111/2041-210X.12401))
12. Cooney CR, Tobias JA, Weir JT, Botero CA, Seddon N. 2017 Sexual selection, speciation and constraints on geographical range overlap in birds. *Ecology Letters* **20**, 863–871. (doi:[10.1111/ele.12780](https://doi.org/10.1111/ele.12780))

- 113 13. Bürkner PC. 2017 Brms: An R package for Bayesian multilevel models using Stan. *Journal of Statis-*
114 *tistical Software* **80**, 1–28. (doi:[10.18637/jss.v080.i01](https://doi.org/10.18637/jss.v080.i01))
- 115 14. Nylander JAA, Olsson U, Alström P, Sanmartín I. 2008 Accounting for phylogenetic uncertainty in
biogeography: A bayesian approach to dispersal-vicariance analysis of the thrushes (Aves: Turdus).
116 *Systematic Biology* **57**, 257–268. (doi:[10.1080/10635150802044003](https://doi.org/10.1080/10635150802044003))
- 117 15. Gelman A, Carlin JB, Stern HS, Dunson DB, Vehtari A, Rubin DB. 2013 *Bayesian data analysis, third*
118 *edition*. Third. Boca Raton, FL: CRC Press. (doi:[10.1201/b16018](https://doi.org/10.1201/b16018))
- 119 16. Vehtari A, Gelman A, Gabry J. 2017 Practical Bayesian model evaluation using leave-one-out cross-
120 validation and WAIC. *Statistics and Computing* **27**, 1413–1432. (doi:[10.1007/s11222-016-9696-4](https://doi.org/10.1007/s11222-016-9696-4))