Rapid species recognition favors greater avian-perceived plumage dichromatism in true thrushes (genus: *Turdus*)

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Abstract

Keywords

8 dichromatism, plumage, species recognition

Background

10 Methods

n Plumage sexual dichromatism

A total of N=77 *Turdus* thrush species (approximately ~89% of all known true thrush species) were sampled for plumage spectral reflectance using prepared bird skin specimens at the American Museum of Natural History in New York City and the Field Museum in Chicago. Reflectance measurements spanning 300-700nm were taken in triplicate from the belly, breast, throat, crown and mantle plumage patches [1] of each individual. N=3 male and N=3 female individuals were measured for most species (exceptions: *T. lawrencii*, N=2 males and N=2 females; *T. swalesi*, N=1 male and N=1 female). Reflectance spectra were measured using a 400 µm fiber optic reflection probe fitted with a rubber stopper to maintain a consistent measuring distance of 3 mm and area of 2 mm2 at a 90° angle to the surface of the feather patch. Measurements were taken using a JAZ spectrometer with a pulsed-xenon light source (Ocean Optics, Dunedin, USA) and we used a diffuse 99% reflectance white standard (Spectralon WS-1-SL, Labsphere, North Sutton NH, USA).

We applied a receptor-noise limited visual model [2] of the European Blackbird (*T. merula*) visual sys-

We applied a receptor-noise limited visual model [2] of the European Blackbird (*T. merula*) visual system [3] in the *pavo* [4] package in R v4.0.0 [5] to calculate avian-perceived chromatic and achromatic visual contrast (in units of "Just-Noticeable Differences",or JNDs) of male vs. female plumage patches for all sampled *Turdus* species. Chromatic and achromatic JNDs were calculated for male-female pairs within each species (i.e., N=9 JND values calculated per patch for each species where N=3 males and N=3 females sampled), and then JND values were averaged for each species' respective plumage patches. Under

ideal laboratory conditions, 1 JND is generally considered to be the discriminable threshold past which
an observer is predicted to be able to perceive the two colors as different. However, natural light environments vary both spatially and temporally [6], bringing into question the accuracy of a 1 JND threshold for generalizing visual contrast under natural conditions. Therefore, we calculated the total number
of sexually-dichromatic plumage patches per species (out of N=5 measured patches) as the number of
plumage patches with average JND values > 1, 2, or 3 to account for uncertainty in visual discrimination
thresholds due to variation in psychophysical and ambient lighting conditions affecting the strength of
between-sex plumage visual contrast [7].

37 Life History Data

38 Breeding Timing Model

We collected data on migration behavior and breeding season length from *Thrushes* [8] and the *Handbook* of the Birds of the World [9]. We assigned three different kinds of migratory behavior: 1) full migration when a species description clearly stated that a species "migrates", 2) partial migration when a species was described to have "altitudinal migration", "latitudinal migration" or "movement during non-breeding season", or 3) sedentary when when a species was described as "resident" or "sedentary". Breeding season length was defined as the number of months the species breeds each year.

45 Breeding Sympatry Model

Species' breeding ranges were acquired from *BirdLife International* [10]. We calculated congener breeding range overlaps (as percentages) using the *letsR* package in R [11]. We then calculated the number of sympatric species as the number of congeners with breeding ranges that overlap >30% with the focal species' breeding range [12].

50 Breeding Spacing Model

- 51 Species' breeding range sizes (in km2) were acquired using the BirdLife International breeding range maps.
- Species' island vs. mainland residence was also determined using breeding ranges from BirdLife Interna-
- tional. Mainland residence was assigned if the species had a breeding range on any continent and Japan.
- Island residence was assigned to species having a breeding range limited to a non-continental landmass
- entirely surrounded by an oceanic body of water.

56 Statistical Modeling

We used phylogenetically-corrected Bayesian multilevel logistic regression models using the *brms* v2.13.0 package [13] in R v4.0.0 [5] where responses, the number of sexually-dichromatic patches >1, 2, and 3 chromatic and achromatic JNDs, were modeled as binomial trials (N=5 plumage patch "trials") to test for associations with breeding timing, breeding sympatry and breeding spacing. For all phylogenetically-corrected models, we used the *Turdus* phylogeny from Nylander et al. (2008) [14]to create a covariance

matrix of species' phylogenetic relationships. All models used a dataset of N=67 out of the *Turdus* species for which all the types of data (see above) were available.

Our breeding timing models included the following predictors: z-scores of breeding season length 64 (mean centered and divided by one standard deviation), migratory behavior (full migration as the reference 65 category versus partial migration or sedentary), and their interaction. Breeding sympatry models included the number of sympatric species with greater than 30% breeding range overlap as the only predictor of 67 the probability of having a sexually-dichromatic plumage patch. Breeding spacing models included log_e transformed breeding range size (km2) and breeding landmass (mainland as the reference category versus island). We also ran null models (intercept only) for all responses. All models' intercepts and response 70 standard deviations were assigned a weak prior (Student T: df = 3, location = 0, scale = 10), and predictor coefficients were assigned flat priors. We ran each model for 6,000 iterations across 6 chains and assessed 72 Markov Chain Monte Carlo (MCMC) convergence using the Gelman-Rubin diagnostic (Rhat) [15]. We then performed k-fold cross-validation [16] to refit each model K=16 times. For each k-fold, the training dataset included a randomly selected set of $N-Nrac{1}{K}$ or N≈63 species, and the testing dataset included $N\frac{1}{K}$ or Npprox4 species not included in the training dataset. Finally, we compared differences between the models' expected log pointwise predictive densities (ELPD) to assess which model(s) best predicted the probability of having a sexually-dichromatic plumage patch. [16]. 78

Models' predictor effects were assessed using 90% highest-density intervals of the posterior distributions and probability of direction, the proportion of the posterior distribution that shares the same sign (positive or negative) as the posterior median [17], to provide estimates of the probability of that a predictor has an entirely positive or negative effect on the presence of sexually-dimorphic plumage patches.

Results

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We obtained N ≥ 4000 effective samples for each model parameter and all models' Markov Chains (MCMC) successfully converged (Rhat = 1 for all models' parameters) (Supplementary Figure). All *breeding* sympatry, *breeding timing*, and *breeding spacing* models performed similarly well and substantially better than *intercept only* models in predicting the probability of having a sexually dimorphic plumage patch with achromatic JND values > 1, 2, or 3 (Table 1; all models predicting achromatic plumage patches had ELPD values within 4, following the convention of Burnham and Anderson (2002)[18]). Among models predicting the probability of having a sexually-dichromatic plumage patch with chromatic JND values > 1, 2, or 3, all models performed much better than *intercept only* models, and *breeding sympatry* models had the best predictive performance (Table 1; *breeding sympatry* models all have ELPD = 0, only the *breeding spacing* models predicting dichromatic plumage patches with had similar predictive performance).

Among predictors of achromatically sexually-dimorphic plumage patches

Table 1: Expected log pointwise predictive densities (ELPD) differences and kfold information criterion values of models (ELPD Difference \pm standard error (kfold IC \pm standard error)). Lower values indicate greater model prediction performance.

		Model			
Plumage Metric	JND Threshold	Breeding Sympatry	Breeding Timing	Breeding Spacing	Intercept Only
Achromatic					
	1 JND	0 ± 0 (-122.17 ± 0.67)	-2.51 ± 2.49 (-124.68 ± 2.38)	-2.59 ± 1.01 (-124.76 ± 1.04)	-21.69 ± 7.36 (-143.87 ± 7.51)
	2 JND	0 ± 0 (-98.94 ± 7.56)	-1.19 ± 3.95 (-100.13 ± 9.22)	-0.7 ± 1.34 (-99.64 ± 7.92)	-52.42 ± 12.67 (-151.36 ± 13.4)
	3 JND	-0.04 ± 1.4 (-85.4 ± 8.91)	-1.7 ± 4.41 (-87.07 ± 10.71)	0 ± 0 (-85.37 ± 8.76)	-28.54 ± 10.02 (-113.91 ± 13.65
Chromatic					
	1 JND	0 ± 0 (-115.75 ± 2.95)	-5.67 ± 3.55 (-121.42 ± 2.28)	-2.73 ± 3.4 (-118.49 ± 2.67)	-14.8 ± 7.22 (-130.55 ± 7.05)
	2 JND	0 ± 0 (-88.47 ± 8.77)	-3.8 ± 4.46 (-92.27 ± 10.01)	-3.32 ± 5.29 (-91.79 ± 10.91)	-50.53 ± 14.49 (-139 ± 16.77)
	3 JND	0 ± 0 (-62.77 ± 10.41)	-8 ± 4.32 (-70.77 ± 12.29)	-4.43 ± 3.9 (-67.2 ± 11.72)	-47.63 ± 15.34 (-110.4 ± 20.01)

Table 2: Model predictor effect estimates (posterior median log-odds and 90% highest-density interval) on the presence of a plumage patch with achromatic or chromatic visual contrast values > 1, 2, and 3 JND. Model effects with a probability of direction (pd) value \geq 0.90 are bolded in red for a negative effect and blue for a positive effect on plumage dichromatism.

Model	Parameter	Achromatic, 1 JND	Achromatic, 2 JND	Achromatic, 3 JND	Chromatic, 1 JND	Chromatic, 2 JND	Chromatic, 3 JND
Breeding Timing							
	Intercept	-1.16 (-3.87, 1.67), pd = 0.76	-1.16 (-3.87, 1.67), pd = 0.76 -8.36 (-16.28, -0.62), pd = 0.98 -7.81 (-14.83, -1.66), pd = 0.99 -0.88 (-2.98, 1.03), pd = 0.78 -7.21 (-15.29, 0.55), pd = 0.95 -1.271 (-28.03, 0.31), pd = 0.96	-7.81 (-14.83, -1.66), pd = 0.99	-0.88 (-2.98, 1.03), pd = 0.78	-7.21 (-15.29, 0.55), pd = 0.95	-12.71 (-28.03, 0.31), pd = 0.96
	Breeding Season Length	-0.06 (-0.62, 0.56), pd = 0.57 -2.26 (-4.72, 0.05), pd = 0.97	-2.26 (-4.72, 0.05), pd = 0.97	-1.39 (-3.56, 0.4), pd = 0.91	-0.12 (-0.59, 0.34), pd = 0.66	-0.12 (-0.59, 0.34), pd = 0.66 -1.99 (-4.64, 0.35), pd = 0.94	-2.5 (-8.12, 2.21), pd = 0.83
	Partial Migration vs. No Migration	-0.04 (-1.16, 1.01), pd = 0.53	1.41 (-1.2, 4.12), pd = 0.83	1.29 (-0.82, 3.57), pd = 0.85	0.79 (-0.06, 1.59), pd = 0.94	1.9 (-0.87, 4.9), pd = 0.88	4.26 (-1.13, 10.99), pd = 0.92
	Full Migration vs. No Migration	1.6 (-0.05, 3.19), pd = 0.96	4.2 (1.16, 7.5), pd = 0.99	3.11 (0.46, 5.73), pd = 0.98	0.83 (-0.37, 1.99), pd = 0.88	4.39 (1.03, 8.14), pd = 0.99	5.46 (-0.68, 12.61), pd = 0.95
	Breeding Season Length x Partial Migration	0.29 (-0.73, 1.37), pd = 0.68	3.03 (-0.14, 6.38), pd = 0.96	2.11 (-0.27, 4.69), pd = 0.94	0.33 (-0.43, 1.14), pd = 0.76	2.2 (-0.82, 5.53), pd = 0.9	3.54 (-2.58, 11.13), pd = 0.85
	Breeding Season Length \times Full Migration	1.58 (-0.4, 3.68), pd = 0.9	4.19 (-0.53, 9.34), pd = 0.93	2.8 (-1.3, 6.72), pd = 0.89	0.52 (-1.16, 2.12), pd = 0.7	5.08 (-0.18, 11.12), pd = 0.95	6.07 (-4.27, 17.43), pd = 0.85
	Phylogenetic Signal λ, Median (90% Credible Interval)	0.29 (0.16, 0.43)	0.72 (0.56, 0.86)	0.61 (0.42, 0.8)	0.17 (0.08, 0.28)	0.74 (0.57, 0.88)	0.89 (0.77, 0.97)
Breeding Spacing							
	Intercept	-1.94 (-6.01, 2.01), pd = 0.8	-9.77 (-20.11, 0.89), pd = 0.95	-10.31 (-19.2, -1.98), pd = 0.98 -0.67 (-3.63, 2.27), pd = 0.65 -8.32 (-18.86, 2.03), pd = 0.92 -12.87 (-30.57, 4.41), pd = 0.91	-0.67 (-3.63, 2.27), pd = 0.65	-8.32 (-18.86, 2.03), pd = 0.92	-12.87 (-30.57, 4.41), pd = 0.91
	Island vs. Mainland	0.08 (-1.38, 1.57), pd = 0.54	-0.64 (-4.43, 2.88), pd = 0.61	-0.09 (-3.02, 2.96), pd = 0.52	-1.3 (-2.45, -0.12), pd = 0.97	-3.39 (-8.67, 1.38), pd = 0.89	-3.26 (-12.57, 4.21), pd = 0.77
	Breeding Range Size	0.08 (-0.13, 0.28), pd = 0.75	0.21 (-0.27, 0.7), pd = 0.77	0.26 (-0.14, 0.66), pd = 0.87	0.02 (-0.14, 0.18), pd = 0.58	0.21 (-0.29, 0.72), pd = 0.77	0.23 (-0.62, 1.1), pd = 0.69
	Phylogenetic Signal λ, Median (90% Credible Interval)	0.27 (0.15, 0.41)	0.71 (0.56, 0.85)	0.6 (0.42, 0.77)	0.15 (0.07, 0.25)	0.72 (0.55, 0.86)	0.85 (0.71, 0.95)
Breeding Sympatry							
	Intercept	-0.9 (-3.45, 1.76), pd = 0.72	-6.89 (-14.7, -0.02), pd = 0.95	-6.74 (-13.39, -1.09), pd = 0.98 -1.38 (-3.25, 0.3), pd = 0.91	-1.38 (-3.25, 0.3), pd = 0.91	-6.34 (-13.61, 0.11), pd = 0.95	-6.34 (-13.61, 0.11), pd = 0.95 -11.29 (-22.79, -1.24), pd = 0.98
	Number of Sympatric Species (≥ 30% Breeding Range Overlap) 0.03 (-0.18, 0.24), pd = 0.61	0.03 (-0.18, 0.24), pd = 0.61	0.14 (-0.31, 0.56), pd = 0.71	0.12 (-0.27, 0.49), pd = 0.71	0.34 (0.17, 0.51), pd = 0.99	0.46 (0.01, 0.92), pd = 0.96	0.75 (0.03, 1.5), pd = 0.97
	Phylogenetic Signal λ, Median (90% Credible Interval)	0.26 (0.14, 0.39)	0.7 (0.54, 0.83)	0.59 (0.41, 0.77)	0.13 (0.06, 0.23)	0.69 (0.52, 0.83)	0.82 (0.67, 0.94)

Discussion

97 Conclusions

Acknowledgements

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