

Towards Understanding Plant Reactions to Human Movement

- Machine Learning Analysis of the Electrical Signal of
 - Plant Species Exposed to Eurythmy Letters



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Introduction

Background needed to understand the project.

02

Methodology

Detailed explanation of the process.

03

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Evaluation of features and models.

04

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Interpretation of the results, limitations, and future work.

01

Introduction (Related Work)

What does a plant do?

Photosynthesis



Intelligence



Respiration



Nutrition



Tropisms



Transpiration



Dormancy



Electrophysiology

Electrical Responses

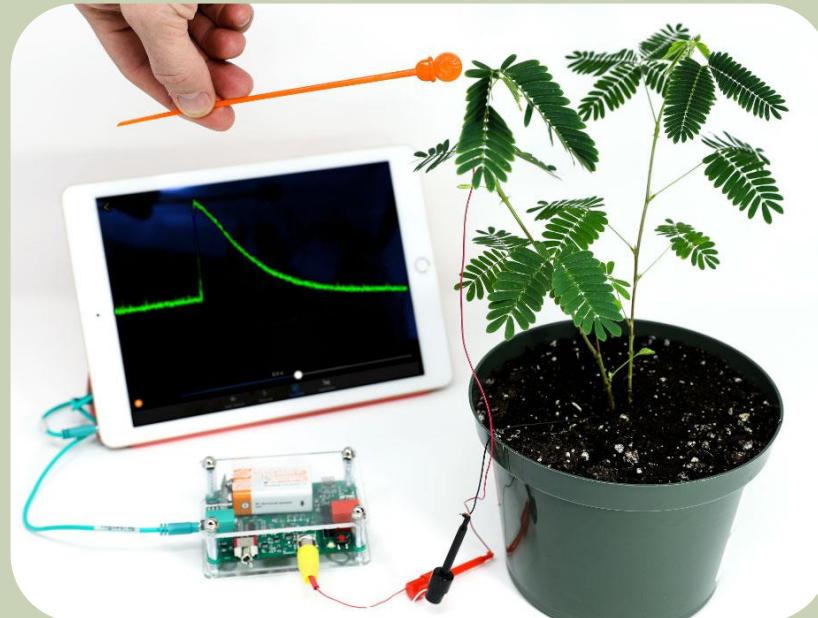
Sanderson, 1873

Pickard, 1973

Marzullo, 2012



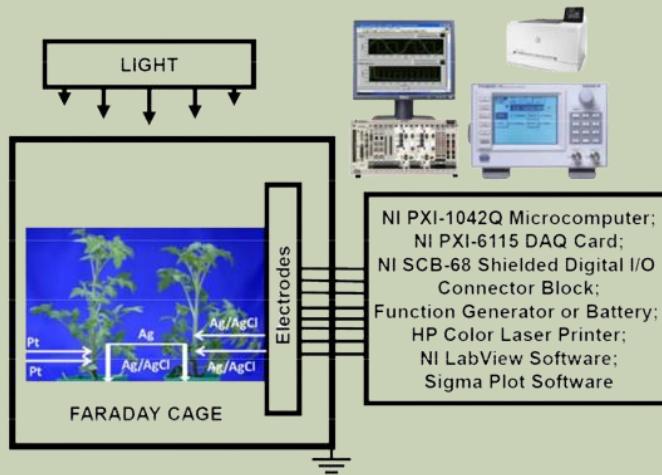
Plant Spikerbox



Communication

Volkov et al, 2019

1. Volatile Organic Compounds (chemicals)
2. Mycorrhizal Networks (fungi)
3. Rhizosphere (chemical)
4. Natural Grafting (merge roots)
5. Electrostatic or Electromagnetic
6. Acoustic
7. **Electrical Signal through the Soil**



Light Stimulus

Prediction from Electrophysiological Response



Chatterjee et al, 2014



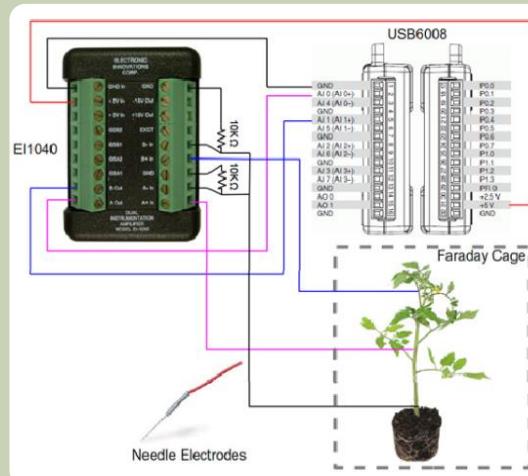
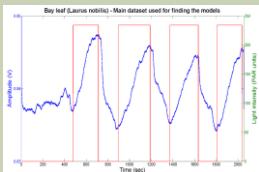
bay leaf



cucumber



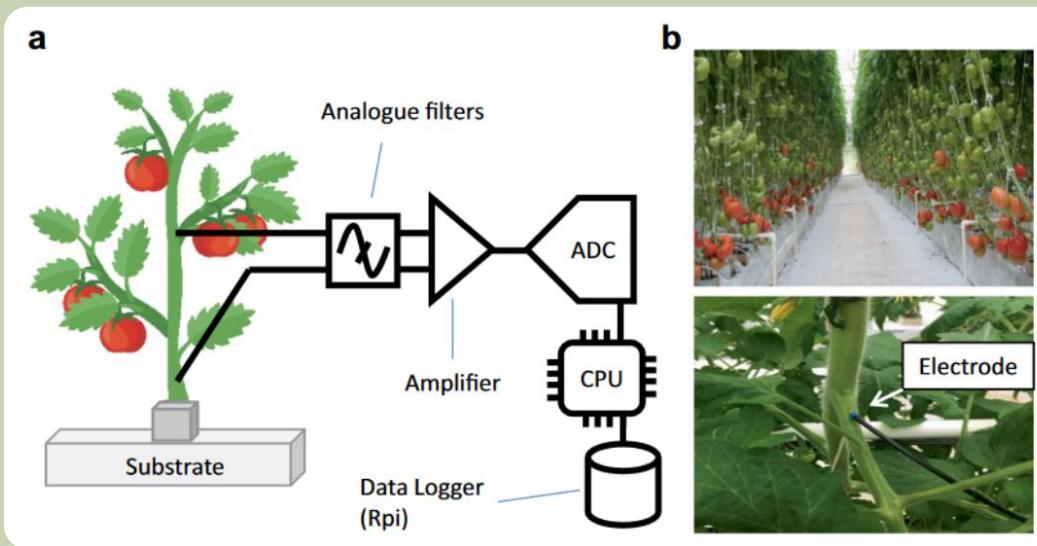
Photon
Flux
Density



Water status

By electrophysiological assessment

Tran et al, Nature 2019



Monitoring

2 weeks

Output
Day/Night
Water Deficit

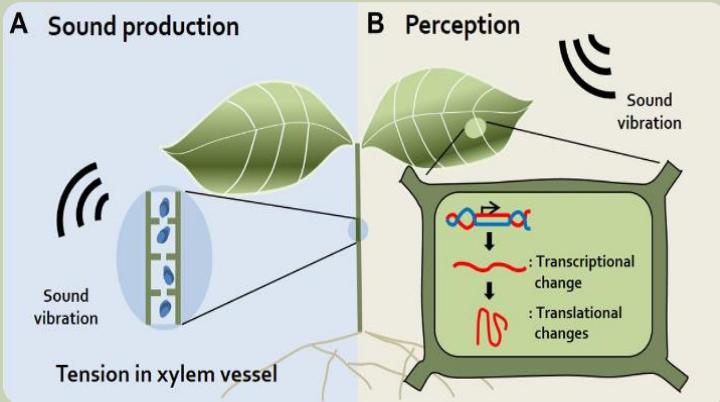
Accuracy

0.94
0.98

Sound Production

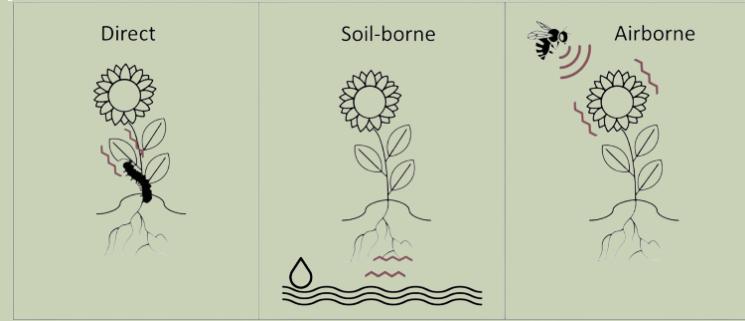
Jung et al, 2018

- Spontaneous sounds
- Gas bubbles in xylem vessels
- Audible and ultrasonic (20-105kHz)

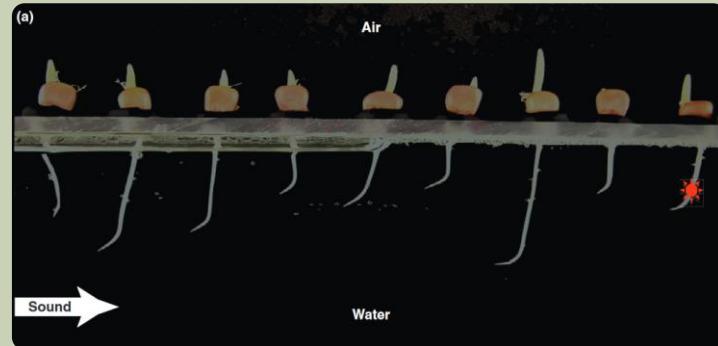


Perception

Khait et al, 2019



Gagliano et al, 2012



Music

Peter & Gloor, 2021



Location
Indoor greenhouse

Species
Dancing plant

Output
Control | 200 Hz | 600 Hz

Accuracy
0.72

Mood Person Detector

Oezkaya & Gloor, 2020

Species
Mimosa Pudica

Output
6 people
+/- mood



Accuracy
0.66
0.86





Eurythmy

- Expressive movement art
- Originated by Rudolf Steiner
- Used in education, anthroposophic medicine, and in Biodynamic agriculture
- The melody is conveyed through expressing the **arm gestures** of the actual letters

Research Questions

01

Do plants react to eurythmy?

02

Do plants react differently between
different eurythmy gestures?

03

Do the electrical signals of different plant
species contain unique characteristics?

02

Methodology

Methodology

01

Experiments

Explanation of the data collection process

02

Preparation

Labeling, trimming and normalization.

03

Featurization

Extraction of wave characteristics

04

Modeling

Selection, modeling and fine-tuning

Methodology

01

Experiments

How to Measure

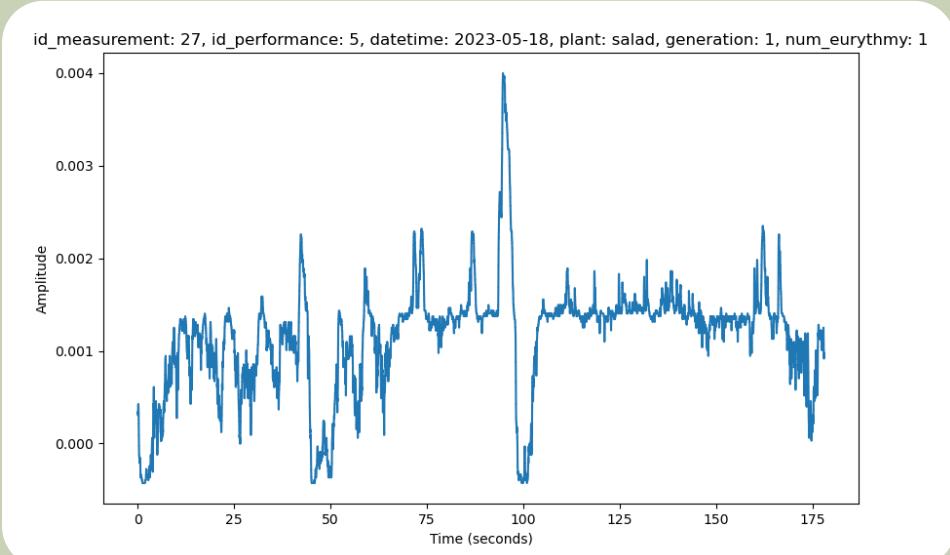
Extracellular Recording

- Surface measurements (EEG)
- Total sum of bioelectrical activity in large groups of cells
- Placement of electrodes on the plant surface and soil

<https://backyardbrains.com/products/plantspikerbox>



Electrical Plant Recording



Measurements

Salad



A-G-D

Tomato



A-G-D-O

Basil



A-G-D-L

Eurythmy Gestures
(4x times each)

Procedure



Control

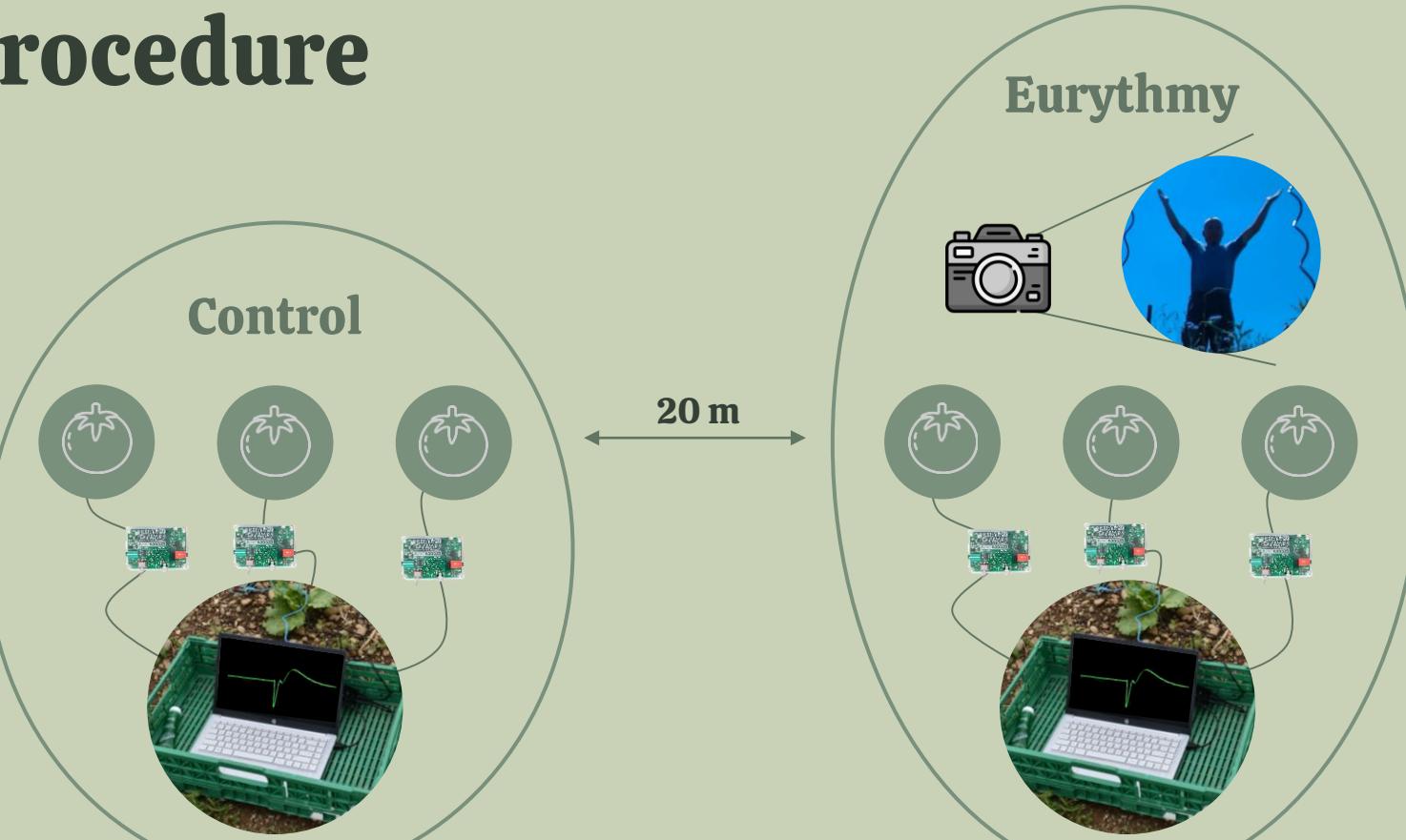
The tension of 3 plants far away from the expert but in the same orchard is measured



Eurythmy

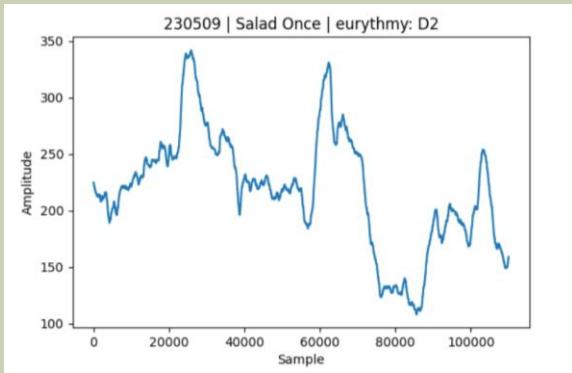
The voltage of 3 plants is measured while an expert performs the eurythmy gestures on them

Procedure



Files

Wav



The Wav files are datapoints and represent the plant voltage over time

625 files: 42 hours

Mp4



The Mp4 files help us to detect the gesture and timing of eurythmy

107 files: 7 hours

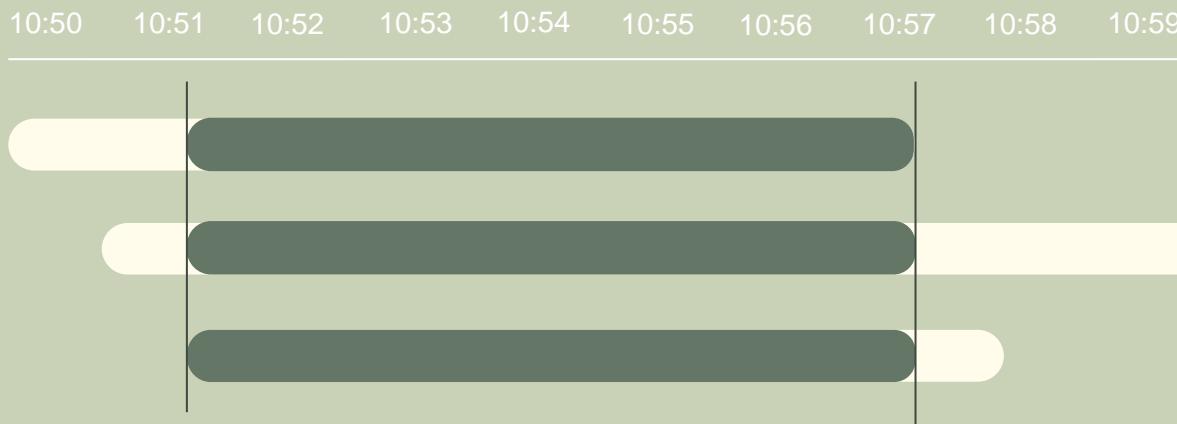
Methodology

02

Preparation

Synchronization

Intersection

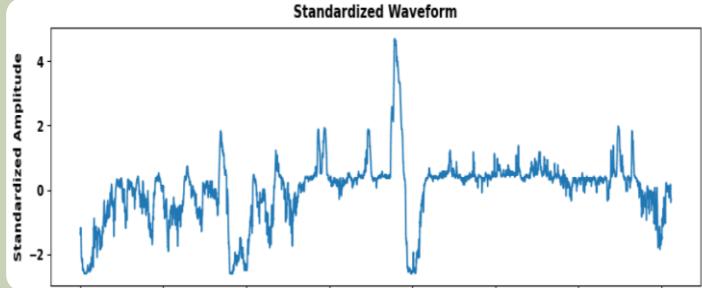
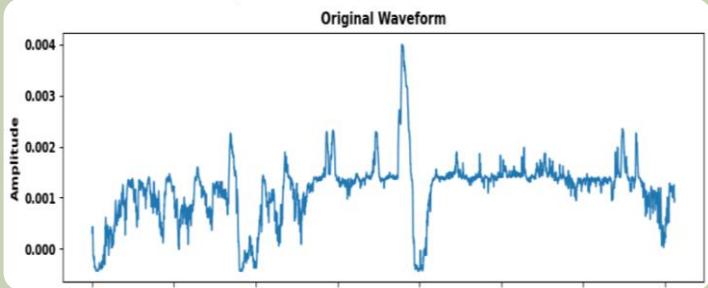


...

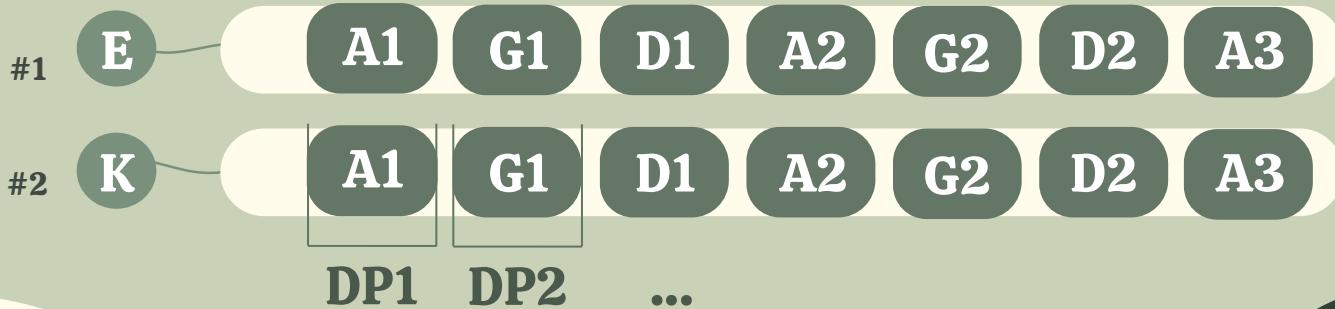
Standardization

Z-Score Normalization

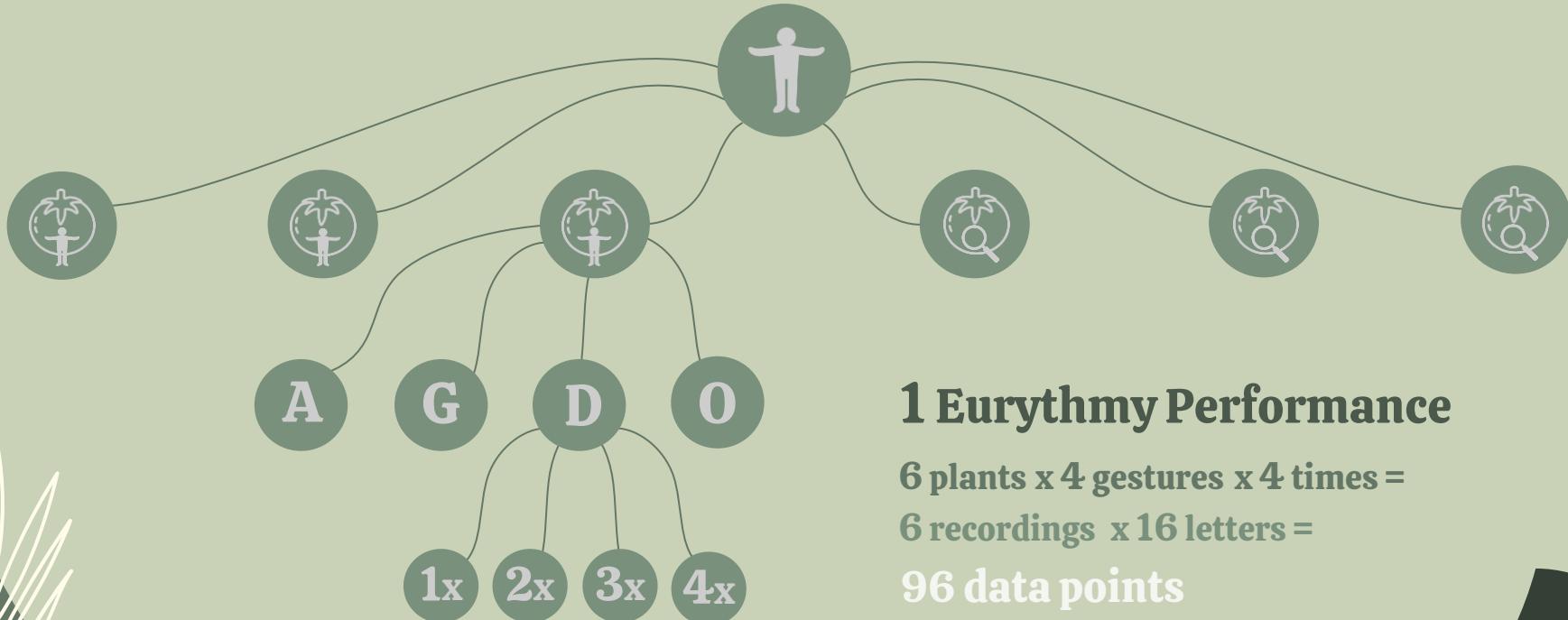
```
def standardize_wave_zscore(waveform):
    mean = np.mean(waveform)
    std_dev = np.std(waveform)
    return (waveform - mean) / std_dev
```



Different Gestures

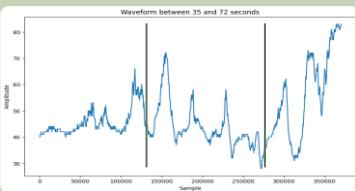


Data Points



Trim

Labelling the video



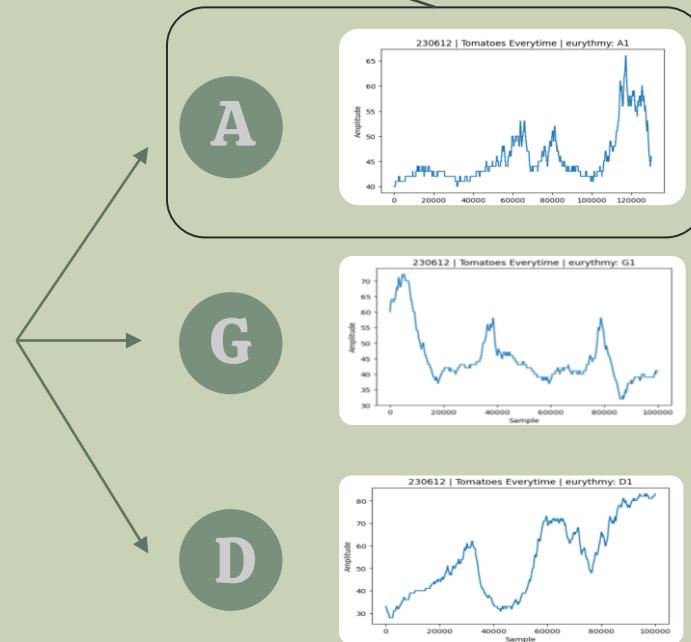
+

230612 TJ

File Edit View

| id, start, end |
|-----------------|
| A1, 00:35,00:48 |
| G1, 00:50,01:00 |
| D1, 01:02,01:12 |
| A2, 01:16,01:27 |
| G2, 01:28,01:37 |
| D2, 01:39,01:49 |
| A3, 01:53,02:03 |
| G3, 02:04,02:15 |
| D3, 02:17,02:27 |
| A4, 02:30,02:39 |
| G4, 02:40,02:49 |
| D4, 02:51,03:01 |

Data Point



Data Count

Letter data points:

8878

Control

4395

Eurythmy

4483

A

1261

G

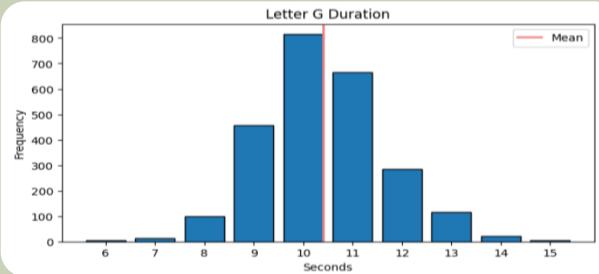
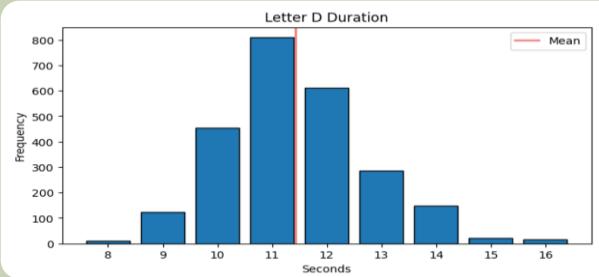
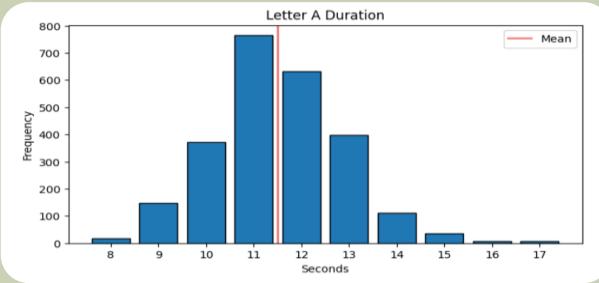
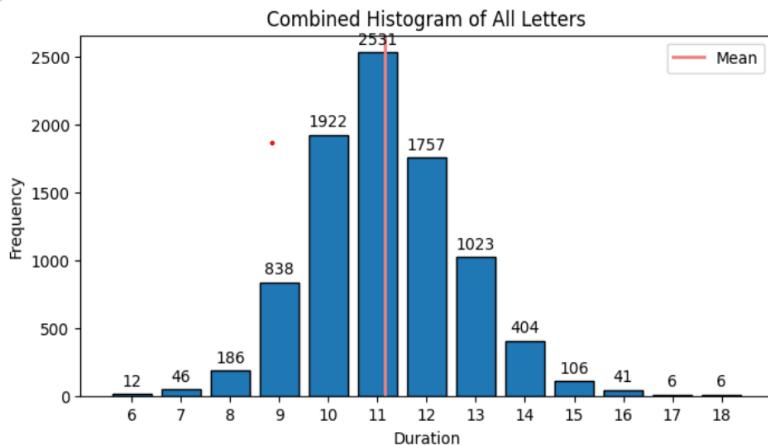
1255

D

1255

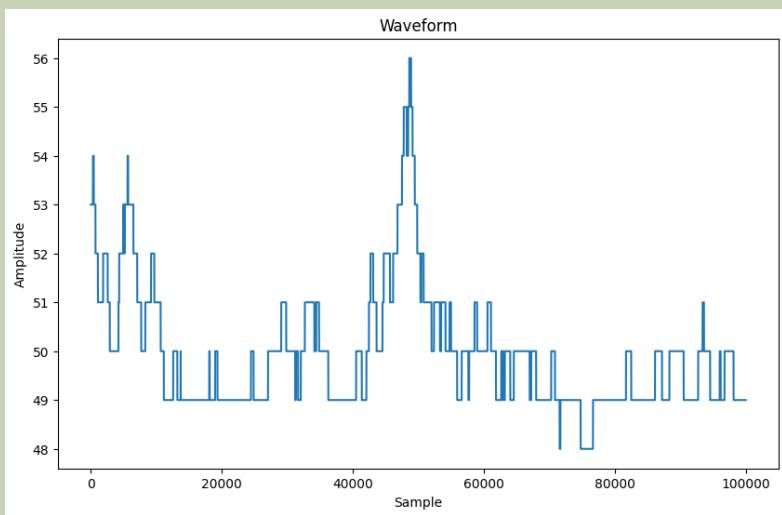
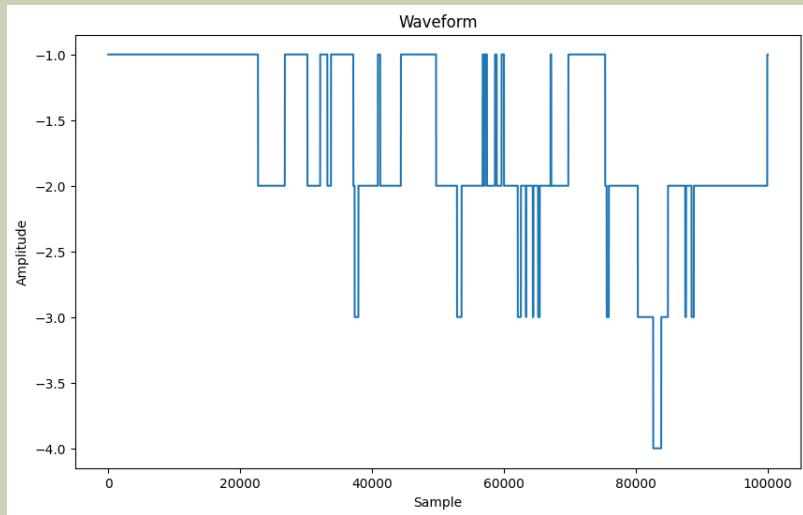
EDA

Letter Duration



Outliers

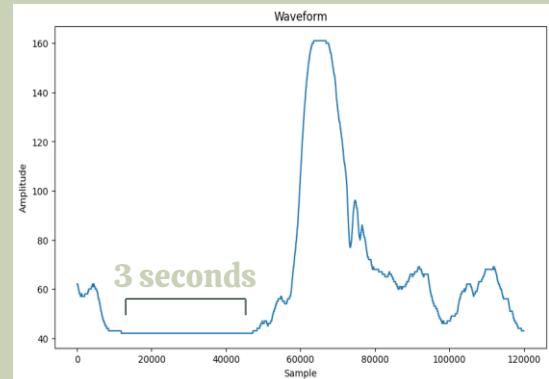
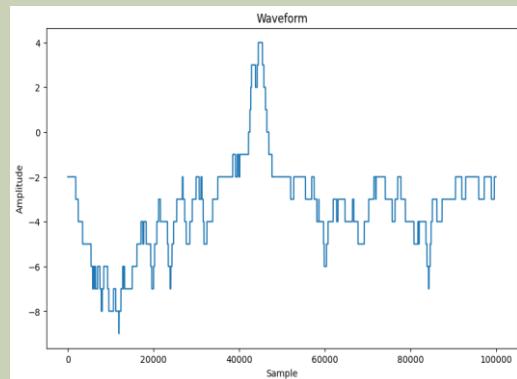
How to detect them systematically?



Outliers

Constant Value for X seconds

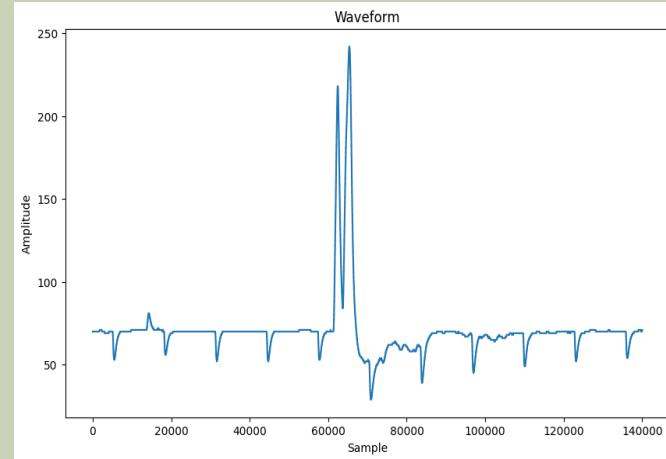
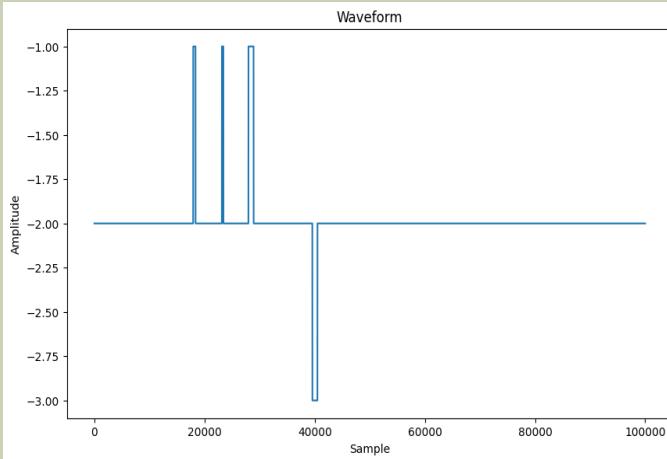
1st Approach



Outliers

Kurtosis threshold

2nd Approach

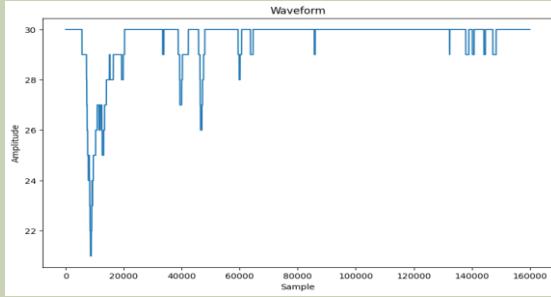


Outliers

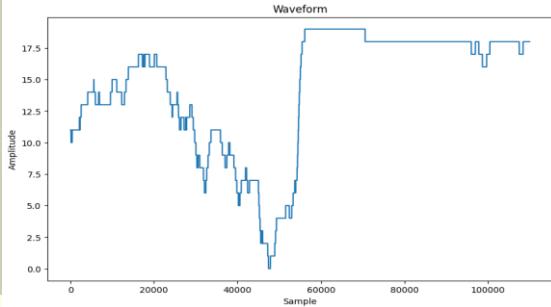
Flatness Ratio

3rd Approach

90%

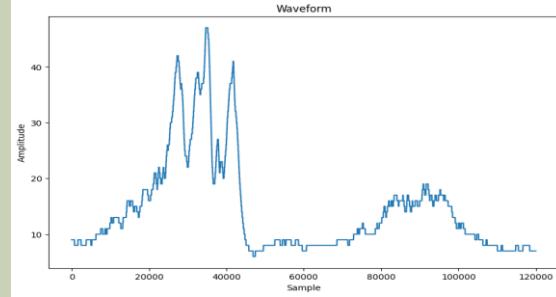


70%

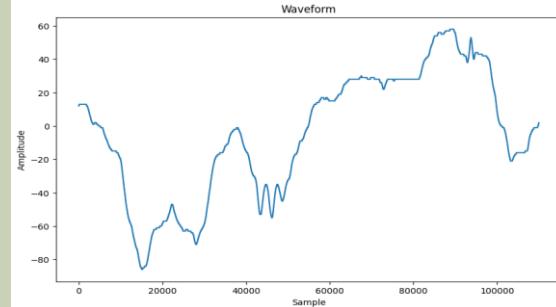


20% dataset is > 50% Flatness Ratio

40%



20%



Flatness Ratio

Definition

Formula

$$R = \frac{1}{n} \sum_{i=1}^m l_i \cdot 1(l_i > T)$$

Given an array A , the flatness ratio R is defined as the ratio between the count of consecutive elements exceeding a certain threshold T and the total number of elements:

Where

- $A = [a_1, a_2, \dots, a_n]$: The array of n elements.
- $S = [s_1, s_2, \dots, s_m]$: A sequence where each s_i represents a segment of consecutive occurrences of the same value in A .
- l_i : The length of the i^{th} segment s_i in S .
- T : The threshold for considering a segment's contribution to the flatness ratio.
- $1(\cdot)$: The indicator function, which is 1 if the condition inside the parentheses is true, and 0 otherwise.
- n : The total number of elements in A .
- m : The total number of segments in S .

Explanation

- $\sum_{i=1}^m l_i \cdot 1(l_i > T)$: This summation iterates through each segment s_i in S , multiplies its length l_i by 1 if $l_i > T$, and sums up these products.
- $\frac{1}{n}$: This term normalizes the sum by dividing it by the total number of elements n in A , yielding the flatness ratio R .

Flatness Ratio

Using Programming

Method

1. It iterates through the array, tracking sequences of identical values.
2. If a sequence length exceeds the threshold, it is added to the total 'flat' length.
3. The ratio of the total 'flat' length to the array's length is returned

```
def extract_flatness_ratio(array, threshold):
    """
    Calculates the flatness ratio of an array.

    The flatness ratio is defined as the proportion of the array where consecutive values remain the same and exceed a specified threshold length.

    :param array: The input array to analyze.
    :param threshold: The minimum length of consecutive values to be considered 'flat'.
    :return: The flatness ratio of the array.

    The function works as follows:
    - It iterates through the array, tracking sequences of identical values.
    - If a sequence length exceeds the threshold, it is added to the total 'flat' length.
    - The ratio of the total 'flat' length to the array's length is returned.
    """

    current_value = None # Tracks the current value being compared
    current_length = 0 # Tracks the length of the current sequence of identical values
    total_length = 0 # Accumulates the total length of all 'flat' sequences

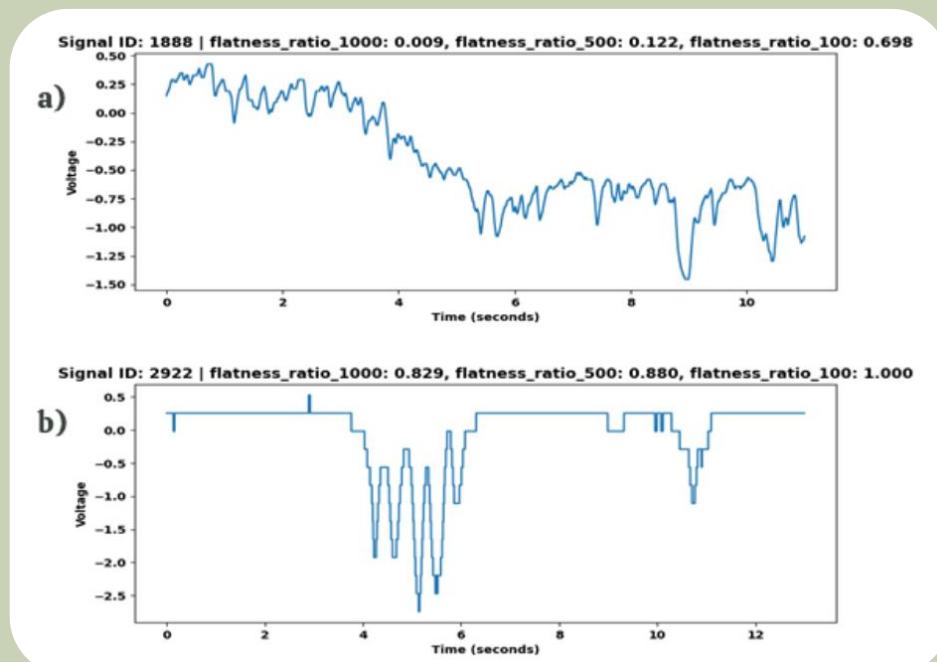
    for value in array:
        if value == current_value:
            # If the current value matches the previous one, increment the sequence length
            current_length += 1
        else:
            # If the current value is different, reset the sequence
            current_value = value
            # If the previous sequence was long enough, add its length to the total
            if current_length > threshold:
                total_length += current_length
            current_length = 1 # Start a new sequence

    # Check the last sequence in the array
    if current_length > threshold:
        total_length += current_length

    # Calculate and return the flatness ratio
    return total_length / len(array)
```

Outlier Identification

Flatness Ratios



Flatness Ratio

In Datasets

| FR | 1000 | 500 | 100 | |
|--------|------|-----|-----|---|
| Alvaro | 27% | 44% | 83% | + |
| Luis | 24% | 40% | 81% | |
| Jakob | 7% | 10% | 23% | |
| Saad | 3% | 7% | 22% | - |

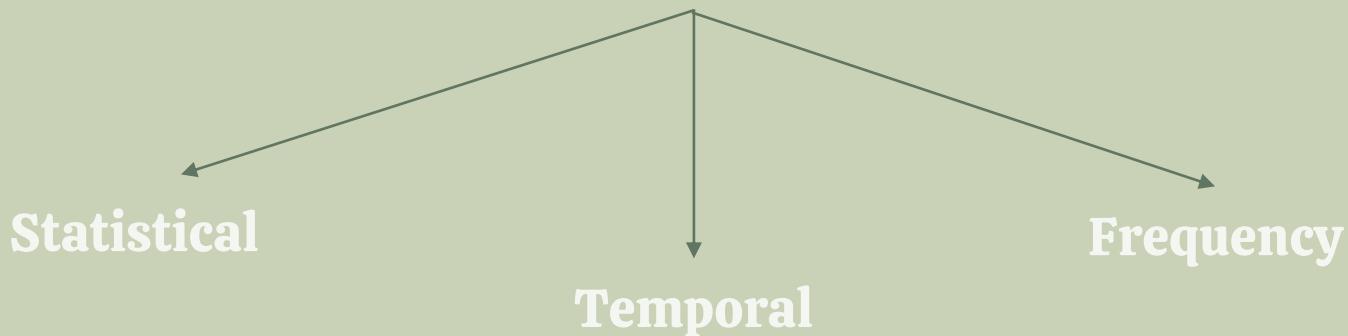
Methodology

03

Featurization

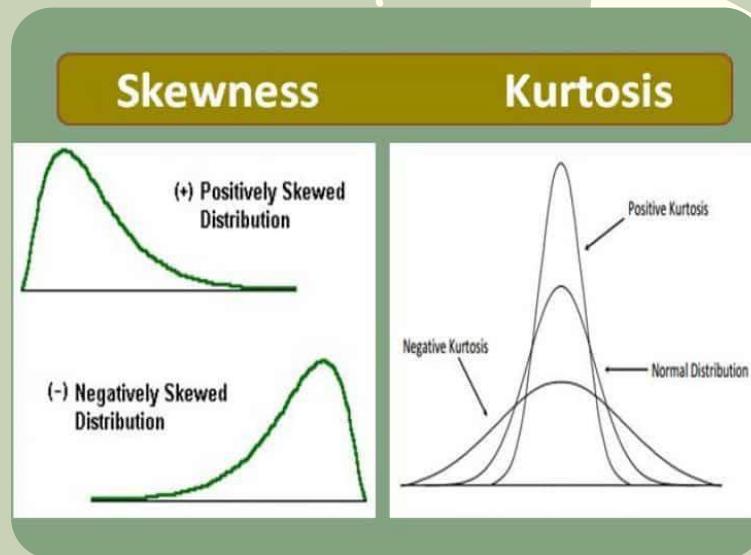
Featurization

Type of Features



Statistical Features

- Mean
- Variance
- Standard Deviation
- Interquartile Range
- Skewness
- Kurtosis
- Hjorth mobility
- Hjorth Complexity
- DFA



Temporal Features

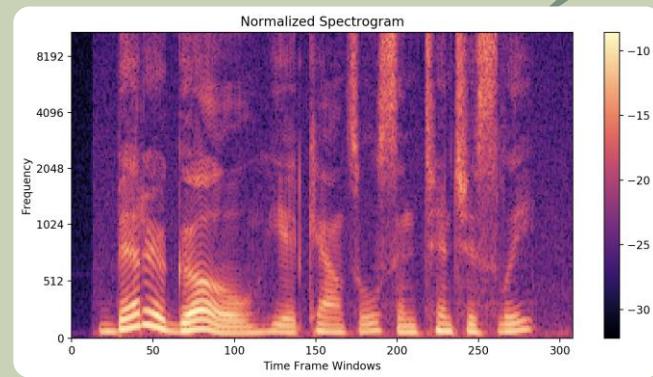


- Zero-Crossing Rate
- Root Mean Square Energy
- Slope Sign Changes Ratio

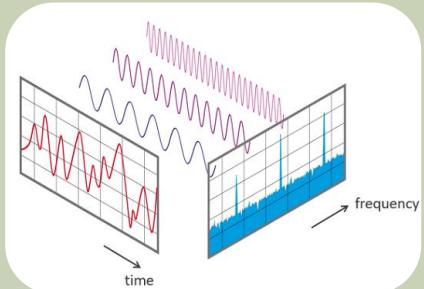
Frequency Processing

Spectrogram

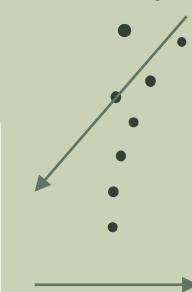
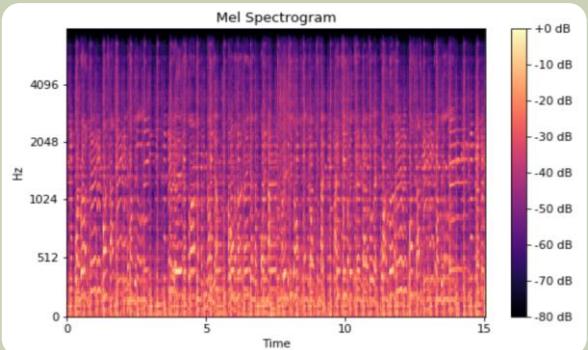
Normalized Spectrogram



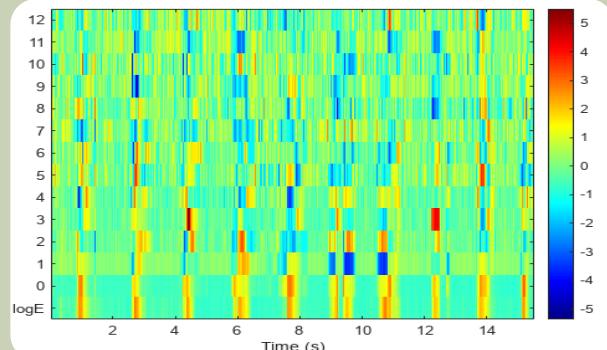
FFT



Mel Spectrogram



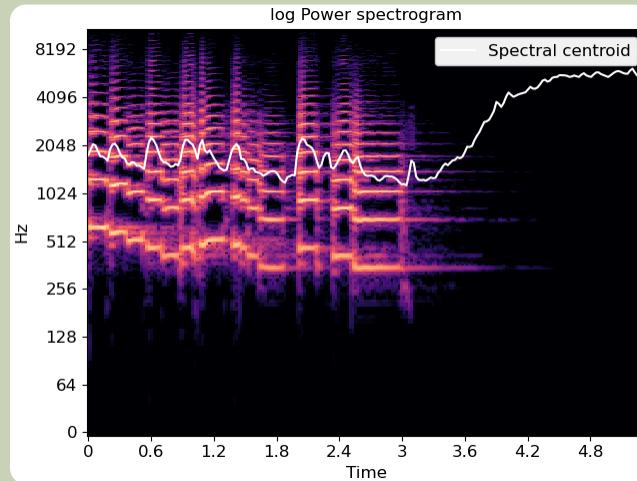
MFCC



Frequency Features

Mid-Term Features

- MFCCs
- Delta MFCCs
- Chromas
- Spectral Centroid
- Bandwith
- Energy Entropy



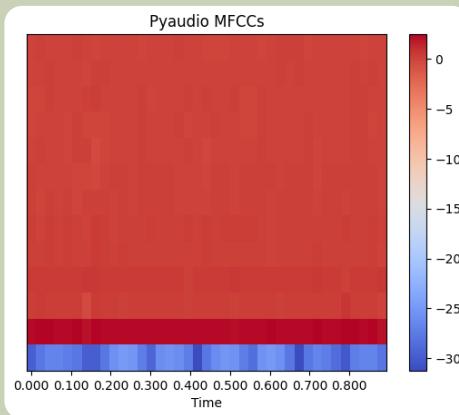
Comparison of libraries

PyAudioAnalysis

+ deltas

+ chromas

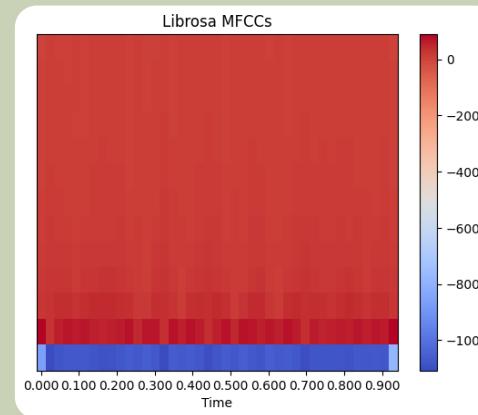
documentation



Librosa

5x faster
+ param MFCCS

documentation



```
librosa.feature.mfcc(*, y=None, sr=22050, S=None, n_mfcc=20, dct_type=2, norm='ortho', lifter=0, **kwargs)
```

PyAudio: `def feature_extraction(signal, sampling_rate, window, step, deltas=True):`

Methodology

04

Modeling

Models used

Table 8. Synthesis of the different models and hyperparameters used for the classification task.

| Model name | Parameter | Values |
|-------------------|-------------------|---------------------|
| AdaBoost | n_estimators | 50, 100 |
| | Learning Rate | 0.1, 1 |
| Extra Trees | n_estimators | 100, 200 |
| | Max Depth | None, 20 |
| | Min Samples Split | 2, 5 |
| Gaussian NB | Var Smoothing | 1e-09, 1e-08, 1e-10 |
| Gradient Boosting | n_estimators | 100, 200 |
| | Learning Rate | 0.1, 0.5 |
| | Max Depth | 3, 5 |
| K-Neighbors | n_neighbors | 5, 10, 15 |
| | Weights | uniform, distance |
| LGBM | n_estimators | 100, 200 |
| | Learning Rate | 0.1, 0.05 |
| | Num Leaves | 31, 64 |
| Random Forest | n_estimators | 100, 200, 300 |
| | Max Depth | None, 10, 20 |
| | Min Samples Split | 2, 5 |
| XGB | n_estimators | 100, 200 |
| | Learning Rate | 0.1, 0.05 |
| | Max Depth | 3, 6 |

03

Results

Results

RQ1

Eurythmy/ Control

Feature Analysis

Kruskal-Wallis

Features

Slope Sign Changes Ratio

Mean

Variance

PyaudioAnalysis

...

Correlation Treshold: 0.8

Table 9. Kruskal-Wallis test of the feature differences between Control and Eurythmy Groups.

| Feature | Control Avg | Eurythmy Avg | p-Value |
|-----------------------------|-------------|--------------|-----------|
| slope_sign_changes_ratio | 0.0297 | 0.0087 | 0.00e+00 |
| mfcc_9_mean | 0.0534 | 0.0431 | 1.81e-239 |
| mfcc_12_mean | 0.0426 | 0.0344 | 5.81e-225 |
| mfcc_13_mean | 0.0303 | 0.0241 | 2.29e-207 |
| mfcc_11_mean | 0.0400 | 0.0322 | 6.01e-206 |
| delta_spectral_spread_std | 0.0632 | 0.0766 | 8.10e-135 |
| dfa | 1.6961 | 1.6382 | 1.89e-123 |
| delta_energy_std | 0.0509 | 0.0433 | 1.66e-35 |
| variance | 0.6927 | 0.4800 | 8.63e-33 |
| interquartile_range | 0.8215 | 0.6964 | 6.49e-27 |
| energy_mean | 0.1320 | 0.1526 | 2.93e-25 |
| cepstra_8_std | 0.7639 | 0.7646 | 2.38e-23 |
| delta_chroma_8_std | 4.51e-05 | 4.13e-05 | 6.85e-17 |
| delta_mfcc_12_std | 0.1189 | 0.1205 | 5.45e-09 |
| mean | -0.0001 | -0.0366 | 1.95e-08 |
| delta_zcr_mean | -3.15e-07 | 7.33e-08 | 2.22e-06 |
| delta_chroma_4_mean | 2.49e-08 | 2.47e-08 | 3.38e-06 |
| chroma_std_mean | 0.0551 | 0.0550 | 8.27e-06 |
| delta_energy_entropy_mean | 2.60e-05 | -3.41e-05 | 1.01e-05 |
| delta_chroma_3_mean | 1.04e-06 | 1.19e-06 | 3.20e-05 |
| delta_spectral_rolloff_mean | -2.56e-08 | 1.32e-07 | 0.0003 |
| cepstra_1_std | 1.4264 | 1.4421 | 0.0017 |
| delta_spectral_centroid_std | 0.0379 | -0.0376 | 0.0023 |
| delta_chroma_std_mean | 1.01e-06 | -6.95e-07 | 0.0033 |
| energy_std | 0.0833 | 0.0819 | 0.0083 |

Canonical Comparison

Average

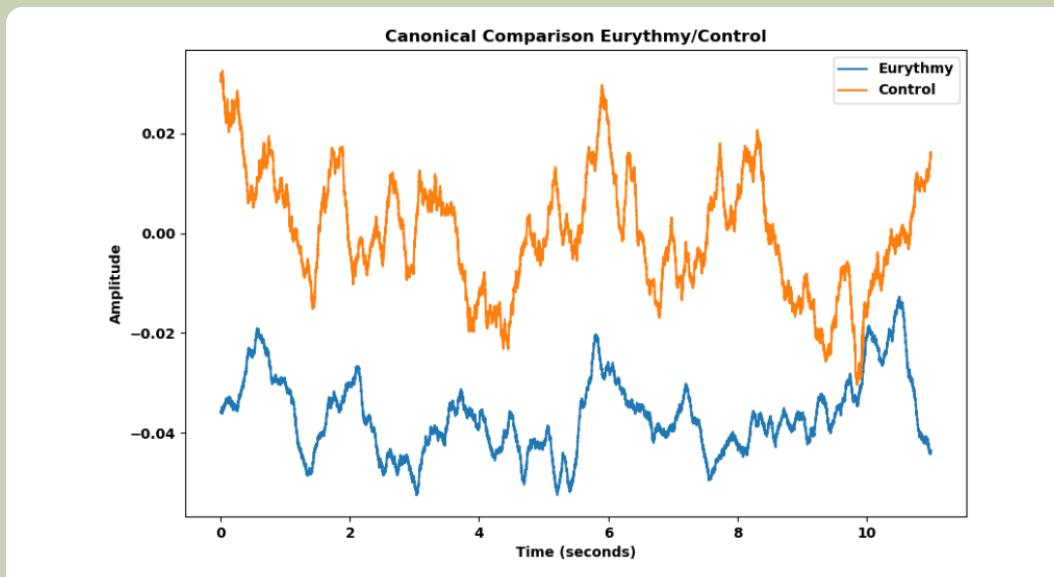


Figure 7. Eurythmy and Control average electrical signals when eurythmy gestures were performed.

Model Results

Table 10. Performance metrics for models in classifying eurythmy and control samples.

| Model | F1 | Accuracy | Precision | Recall |
|------------------|---------------|---------------|---------------|---------------|
| baseline | - | 0.5038 | - | - |
| adaboost | 0.7134 | 0.7153 | 0.7211 | 0.7153 |
| extratrees | 0.7251 | 0.7257 | 0.7277 | 0.7257 |
| gaussiannb | 0.4531 | 0.5260 | 0.5558 | 0.5260 |
| gradientboosting | 0.7389 | 0.7396 | 0.7422 | 0.7396 |
| kneighbors | 0.5179 | 0.5179 | 0.5179 | 0.5179 |
| lgbm | 0.7488 | 0.7494 | 0.7521 | 0.7494 |
| randomforest | 0.7337 | 0.7344 | 0.7367 | 0.7344 |
| xgb | 0.7381 | 0.7390 | 0.7422 | 0.7390 |

Best Model

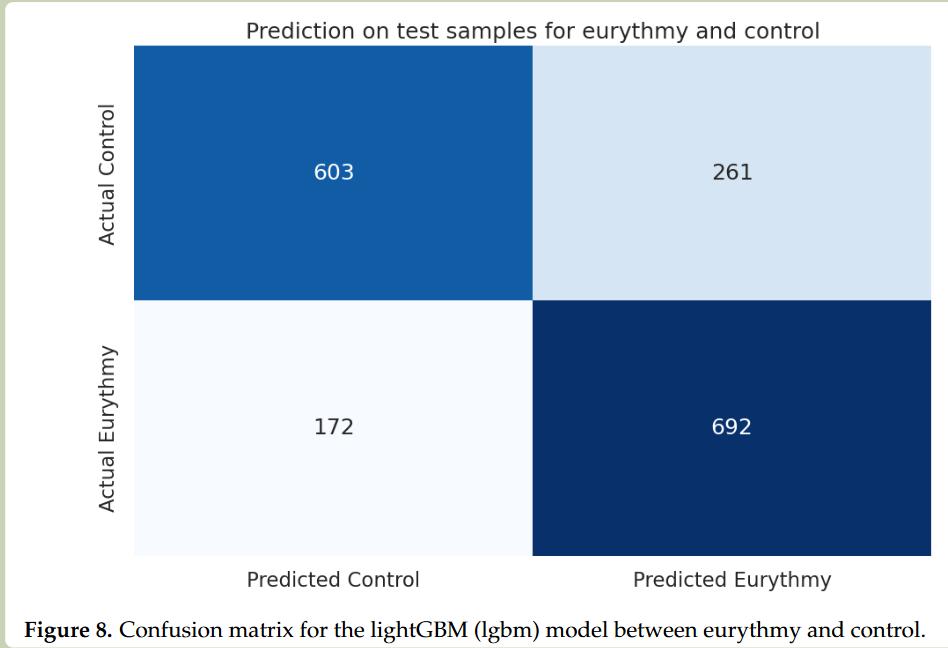


Figure 8. Confusion matrix for the lightGBM (lgbm) model between eurythmy and control.

lightGBM
Test Accuracy: 0.75
+49% baseline
F1-Score: 0.75

Results

RQ2

A/G/D

Letters

A



G



D



Feature Analysis

Kruskal-Wallis

Features

Cepstrals

Mean

Energy

Hjorth

...

Correlation Treshold: 0.9

Table 11. Statistical significances of Kruskal-Wallis between letter groups.

| Feature | A Avg | G Avg | D Avg | p-Value |
|-----------------------------|----------|----------|----------|----------|
| mean | 0.0757 | -0.0056 | -0.0448 | 2.59e-30 |
| root_mean_square_energy | 0.7505 | 0.7186 | 0.6903 | 1.40e-09 |
| cepstra_4_avg | 0.2641 | 0.2623 | 0.2582 | 2.23e-09 |
| cepstra_1_std | 1.3481 | 1.3422 | 1.3313 | 9.00e-09 |
| zero_crossing_rate | 4.70e-05 | 4.71e-05 | 4.89e-05 | 0.0011 |
| dfa | 1.5778 | 1.5745 | 1.5777 | 0.0040 |
| delta_chroma_3_std | 0.0140 | 0.0142 | 0.0137 | 0.0069 |
| spectral_spread_std | 0.0356 | 0.0362 | 0.0357 | 0.0073 |
| slope_sign_changes_ratio | 0.0096 | 0.0102 | 0.0096 | 0.0082 |
| spectral_flux_std | 0.0719 | 0.0727 | 0.0710 | 0.0113 |
| spectral_centroid_std | 0.0235 | 0.0238 | 0.0235 | 0.0116 |
| mfcc_3_mean | 0.2130 | 0.2112 | 0.2132 | 0.0118 |
| delta_mfcc_5_std | 0.0944 | 0.0953 | 0.0952 | 0.0123 |
| delta_spectral_spread_std | 0.0621 | 0.0632 | 0.0624 | 0.0129 |
| cepstra_2_std | 1.2621 | 1.2697 | 1.2412 | 0.0198 |
| mfcc_5_mean | 0.1373 | 0.1362 | 0.1374 | 0.0231 |
| mfcc_8_mean | 0.0784 | 0.0782 | 0.0791 | 0.0317 |
| mfcc_7_mean | 0.0769 | 0.0765 | 0.0776 | 0.0340 |
| delta_spectral_centroid_std | 0.0389 | 0.0394 | 0.0390 | 0.0387 |
| mfcc_6_mean | 0.1447 | 0.1438 | 0.1450 | 0.0500 |

Canonical Comparison

Interpolation Resampling and Average

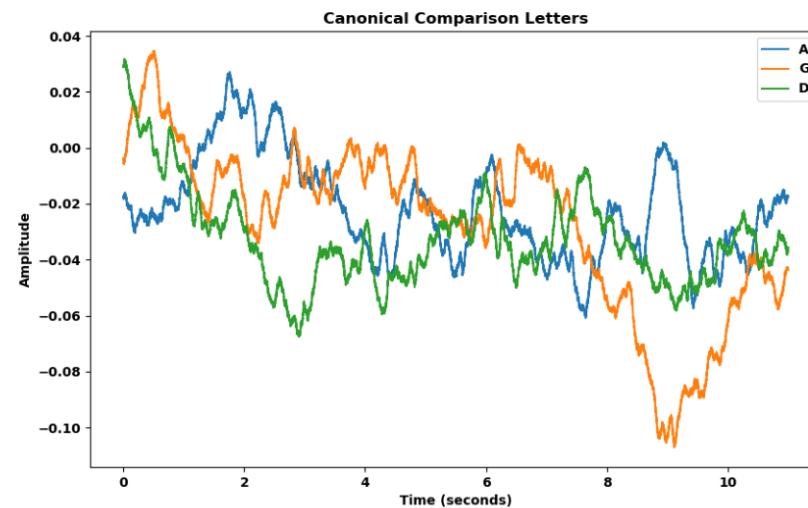


Figure 9. Canonical representation of the average potential curve of each group (A,G,D) after resampling and averaging.

Model Results

Table 12. Performance metrics for models in classifying eurythmy gestures.

| Model | F1 | Accuracy | Precision | Recall |
|-------------------------|---------------|---------------|---------------|---------------|
| baseline | - | 0.3467 | - | - |
| adaboost | 0.4388 | 0.4392 | 0.4415 | 0.4392 |
| extratrees | 0.4349 | 0.4392 | 0.4363 | 0.4392 |
| gaussiannb | 0.4186 | 0.4185 | 0.4192 | 0.4185 |
| gradientboosting | 0.4536 | 0.4586 | 0.4539 | 0.4586 |
| kneighbors | 0.3738 | 0.3757 | 0.3750 | 0.3757 |
| lgbm | 0.4320 | 0.4350 | 0.4307 | 0.4350 |
| randomforest | 0.4444 | 0.4475 | 0.4435 | 0.4475 |
| xgb | 0.4534 | 0.4558 | 0.4530 | 0.4558 |

Best Model

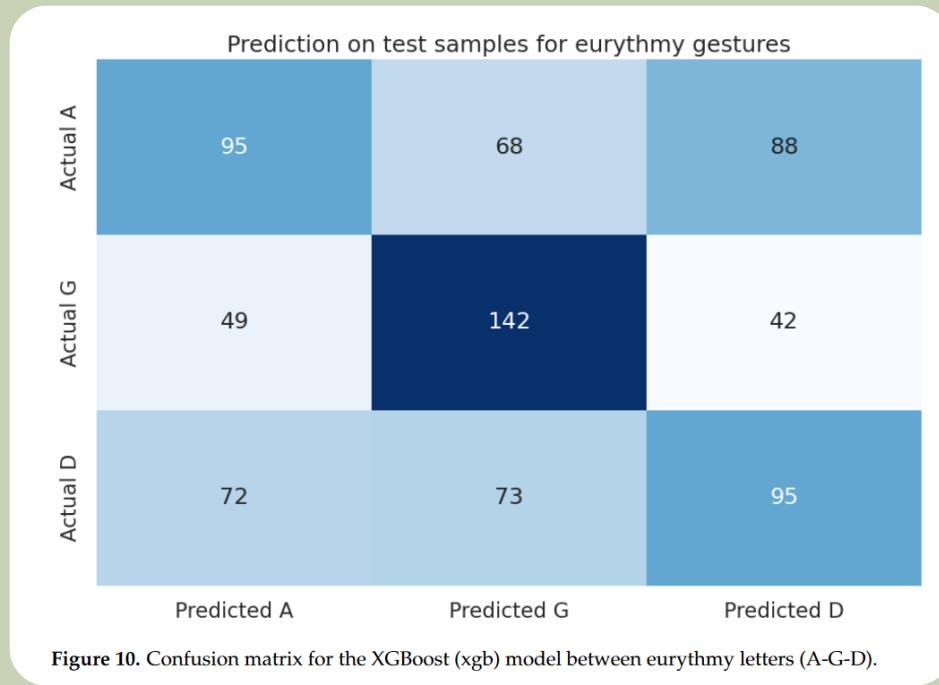


Figure 10. Confusion matrix for the XGBoost (xgb) model between eurythmy letters (A-G-D).

Gradient Boosting

Test Accuracy: 0.46

+32% baseline

F1-Score: 0.45

Results

RQ3

salad/tomato/basil

Feature Analysis

Kruskal-Wallis

Features

Dfa

Slope Sign Changes Ratio

MFCC

Variance

...

Correlation Treshold: 0.8

Table 13. Statistical Analysis of Feature Differences across Plant Species.

| Feature | Lettuce Avg | Tomato Avg | Basil Avg | p-Value |
|----------------------------|-------------|------------|------------|-----------|
| dfa | 1.6519 | 1.6271 | 1.6783 | 0.00e+00 |
| slope_sign_changes_ratio | 0.0220 | 0.0152 | 0.0260 | 3.54e-198 |
| delta_mfcc_3_std | 0.1085 | 0.1136 | 0.1062 | 1.15e-118 |
| mfcc_6_std | 0.0531 | 0.0554 | 0.0526 | 1.11e-105 |
| mfcc_5_std | 0.0510 | 0.0531 | 0.0505 | 1.92e-95 |
| zcr_mean | 0.0004 | 0.0004 | 0.0007 | 4.92e-91 |
| mfcc_10_std | 0.0472 | 0.0494 | 0.0470 | 3.37e-83 |
| energy_entropy_std | 0.1260 | 0.1342 | 0.1303 | 9.85e-83 |
| delta_mfcc_8_std | 0.0837 | 0.0870 | 0.0827 | 7.63e-80 |
| delta_mfcc_7_std | 0.0795 | 0.0828 | 0.0790 | 5.93e-75 |
| variance | 0.1940 | 0.1602 | 0.1780 | 1.48e-74 |
| delta_mfcc_12_std | 0.0729 | 0.0762 | 0.0726 | 2.25e-74 |
| mfcc_9_std | 0.0463 | 0.0484 | 0.0462 | 2.65e-74 |
| mfcc_13_std | 0.0385 | 0.0404 | 0.0389 | 4.10e-72 |
| delta_mfcc_11_std | 0.0770 | 0.0804 | 0.0764 | 9.50e-72 |
| delta_energy_std | 0.0788 | 0.0833 | 0.0820 | 2.63e-65 |
| delta_chroma_11_std | 0.0025 | 0.0027 | 0.0025 | 1.76e-61 |
| delta_spectral_rolloff_std | 0.0040 | 0.0043 | 0.0042 | 1.23e-57 |
| delta_chroma_2_std | 7.66e-05 | 8.14e-05 | 7.80e-05 | 7.79e-56 |
| delta_chroma_3_std | 0.0131 | 0.0139 | 0.0137 | 5.93e-55 |
| delta_spectral_flux_std | 0.1126 | 0.1178 | 0.1108 | 3.45e-51 |
| cepstra_1_std | 1.3283 | 1.3552 | 1.3269 | 2.29e-50 |
| interquartile_range | 0.3874 | 0.3811 | 0.4151 | 1.00e-49 |
| energy_mean | 0.2299 | 0.2210 | 0.2145 | 1.10e-43 |
| cepstra_4_avg | 0.2565 | 0.2648 | 0.2555 | 7.12e-38 |
| spectral_spread_std | 0.3321 | 0.3032 | 0.3156 | 1.14e-37 |
| root_mean_square_energy | 0.7415 | 0.7562 | 0.7344 | 4.99e-37 |
| energy_std | 0.0612 | 0.0632 | 0.0623 | 2.83e-32 |
| hjorth_mobility | 0.0001 | 0.0001 | 0.0001 | 4.94e-32 |
| delta_spectral_flux_mean | 0.0048 | 0.0051 | 0.0050 | 3.33e-26 |
| hjorth_complexity | 11097.4615 | 11059.9986 | 11611.5156 | 5.48e-17 |
| chroma_std_mean | 0.0550 | 0.0550 | 0.0552 | 2.80e-14 |
| mean | 0.0076 | 0.0029 | 0.0126 | 7.16e-06 |
| mfcc_10_mean | 0.0568 | 0.0574 | 0.0565 | 6.16e-05 |
| mfcc_11_mean | 0.0382 | 0.0388 | 0.0385 | 0.0022 |
| mfcc_3_mean | 0.2311 | 0.2326 | 0.2317 | 0.0169 |
| mfcc_9_mean | 0.0514 | 0.0519 | 0.0514 | 0.0204 |
| delta_mfcc_5_mean | 3.17e-05 | -7.96e-05 | 1.78e-05 | 0.0241 |
| delta_mfcc_8_mean | 4.80e-05 | -3.96e-05 | -1.37e-05 | 0.0394 |

Canonical Comparison

Average

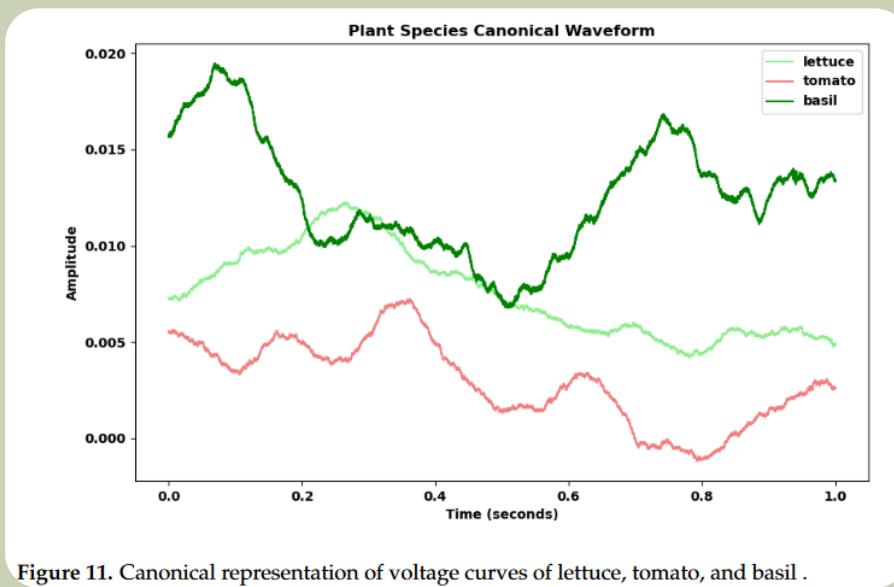


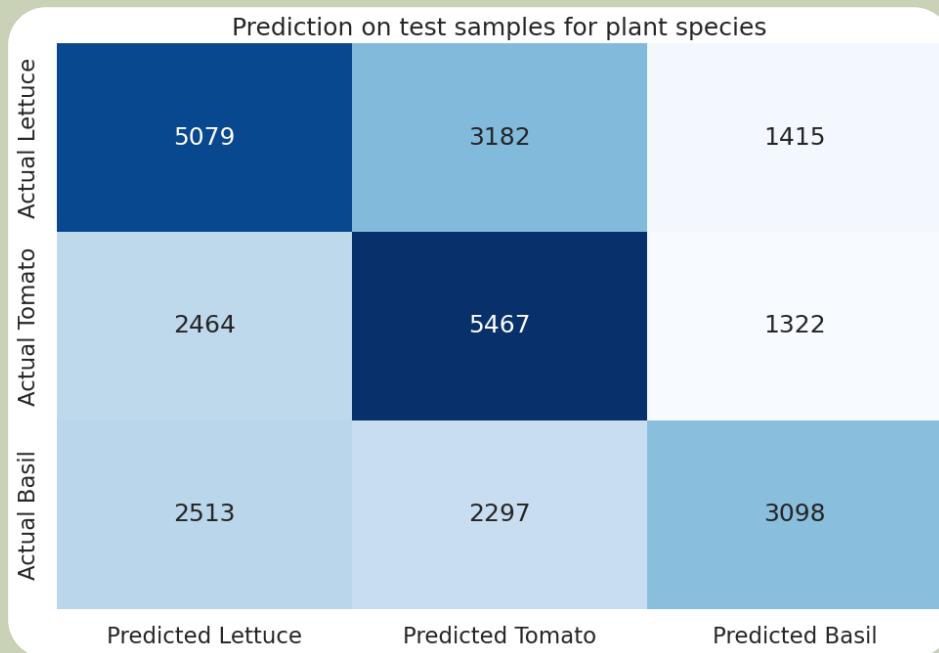
Figure 11. Canonical representation of voltage curves of lettuce, tomato, and basil .

Model Results

Table 14. Model Evaluation Metrics.

| Model | F1 | Accuracy | Precision | Recall |
|------------------|--------|----------|-----------|--------|
| baseline | - | 0.3605 | - | - |
| adaboost | 0.4136 | 0.4180 | 0.4186 | 0.4180 |
| extratrees | 0.4225 | 0.4299 | 0.4355 | 0.4299 |
| gaussiannb | 0.2759 | 0.3670 | 0.3870 | 0.3670 |
| gradientboosting | 0.4949 | 0.4981 | 0.5005 | 0.4981 |
| kneighbors | 0.3382 | 0.3515 | 0.3443 | 0.3515 |
| lgbm | 0.4849 | 0.4868 | 0.4881 | 0.4868 |
| randomforest | 0.4474 | 0.4534 | 0.4601 | 0.4534 |
| xgb | 0.5051 | 0.5084 | 0.5108 | 0.5084 |

Best Model



XGBoost

Test Accuracy: 0.5

+41% baseline

F1-Score: 0.5

04

Discussion

Discussion

01

Featurization

Processing Possibilities

Downsampling



What rate should we choose?

Filter



Should we remove high frequencies?

Offset



When should we center the wave?

Processing



What should we input in the model?

Iterations: Sample Rate | Filter Threshold | Offset Moment | Input

MFCC parameters

01

Framing

Window Size: 20ms - 25 ms (audio)

02

Windowing

Hop Length: 0.50ws – 0.75ws

03

FFT

Window Function: Hamming|Hann

04

Mel Filterbank

Nº of Points FFT: 512|1024|2048

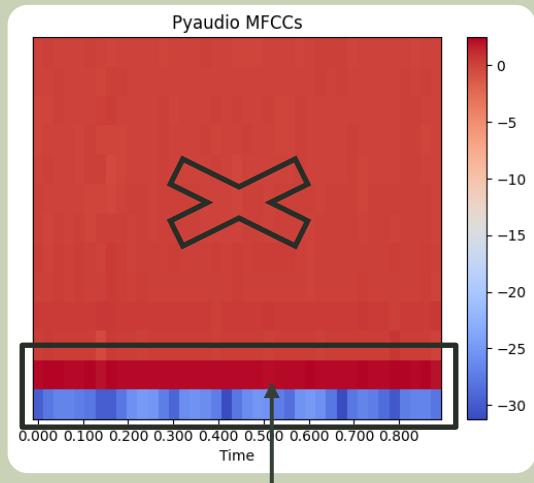
05

MFCCs

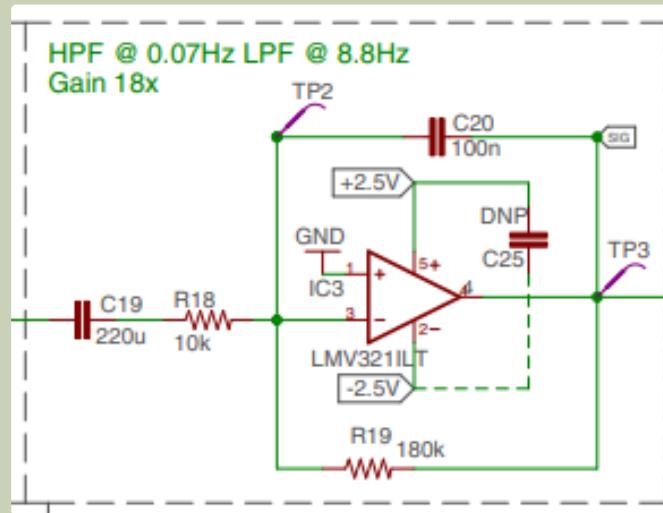
Nº of Filterbanks: 20 - 40

Nº of MFCCs: Type-II|None

MFCC idea



Look closer here



Discussion 02 Modeling

Input Approaches

01

MFCCs

- Machine Learning (+ features)
- MFCC- CNN (+ResNet)
- FCNN

02

Spectrogram

- MFCC- CNN (+ResNet)

03

Signal

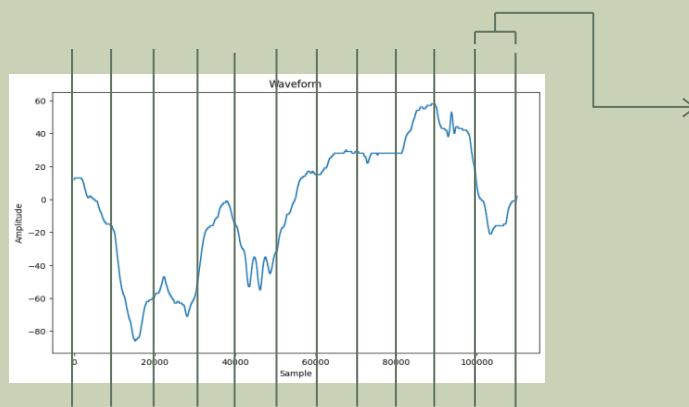
- LSTM Classifier

x2 time series

This was Tried

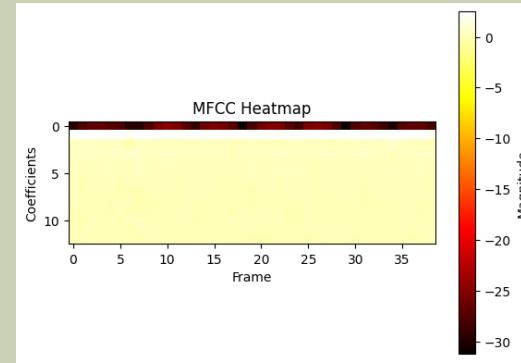
Neural Networks (Didn't Work)

Similar Data Input



Trim wave in 1s segments

Output Matrix



Size: 13x39 (507 elem)

Data points

6.7k
↓
75k

FCNN & MFCC-CNN

Discussion

03

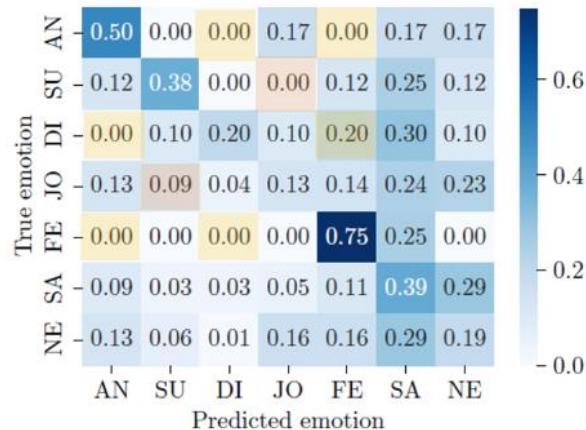
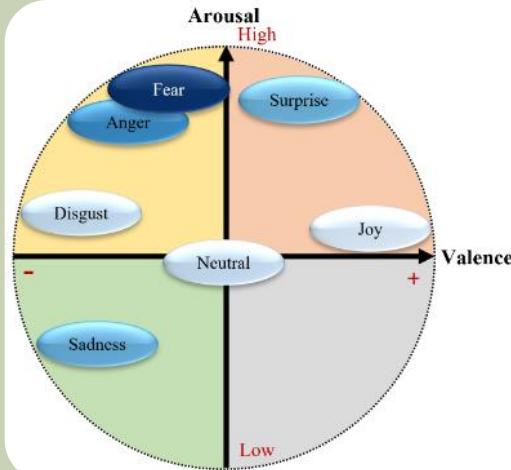
Future of Work

Human emotion

Kruse et al, 2023

Species
Basil Output
7 emotions Accuracy
0.32

| Model | Test set Accuracy | Test set Recall |
|--------------|-------------------|-----------------|
| MLP | 0.399 | 0.220 |
| biLSTM | 0.260 | 0.351 |
| MFCC-CNN | 0.377 | 0.275 |
| MFCC-RestNet | 0.318 | 0.324 |



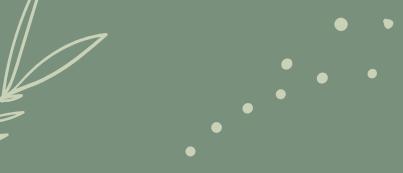
Vision Bio-Lingo



Elings, 2016 Effects of plants on People

- physical
 - 1. Stress and mental fatigue relief
 - 2. Treatment of mental health issues
 - 3. Lower blood pressure and heart rate
 - 4. Faster recovery from stress
 - 5. Reduction in risk factors for diseases
 - 6. Improved self-esteem and responsibility
 - 7. Enhanced tranquility and enjoyment
 - 8. Promotion of relaxation and reflection
 - 9. Positive correlation with well-being
 - 10. Stimulation of social cohesion
 - 11. Enhanced social interaction
 - 12. Reduction in loneliness
- mental
 - 1. Stress and mental fatigue relief
 - 2. Treatment of mental health issues
 - 3. Lower blood pressure and heart rate
 - 4. Faster recovery from stress
 - 5. Reduction in risk factors for diseases
 - 6. Improved self-esteem and responsibility
 - 7. Enhanced tranquility and enjoyment
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 - 11. Enhanced social interaction
 - 12. Reduction in loneliness
- social
 - 1. Stress and mental fatigue relief
 - 2. Treatment of mental health issues
 - 3. Lower blood pressure and heart rate
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 - 5. Reduction in risk factors for diseases
 - 6. Improved self-esteem and responsibility
 - 7. Enhanced tranquility and enjoyment
 - 8. Promotion of relaxation and reflection
 - 9. Positive correlation with well-being
 - 10. Stimulation of social cohesion
 - 11. Enhanced social interaction
 - 12. Reduction in loneliness

Dissemination



Deciphering Human Motion

Presentation



Date
Dec 12, 6pm-7pm

Location
26 Trowbridge St

<https://rcc.harvard.edu/event/made-upm>



GitHub Project

<https://github.com/alvaro-francisco-gil/Plant-Reactivity-Analysis>



Reproducible
Explainable
Scalable

Can plants percieve human gestures?

Article



Article

Can plants perceive human gestures? Using AI to track eurythmic human-plant interaction

Alvaro Francisco Gil ^{1,2}, Moritz Weinbeer ³ and Peter A. Gloor ^{1,*}

¹ MIT Center for Collective Intelligence, Cambridge MA, USA

² UPM Technical University of Madrid, Spain

³ Foundation Fintan, Rheinau, Switzerland

* Correspondence: pgloor@mit.edu

Abstract: This paper explores if plants are capable of responding to human movement by changes in their electrical signals. Towards that goal we conducted a series of experiments, where humans over a period of 6 months were performing different types of Eurythmic gestures in proximity of garden plants, namely salad, basil, and tomatoes. To measure plant perception, we used the plant spikerbox, a device that measures changes in voltage differentials of plants between roots and leaves. Using machine learning, we find that the voltage differentials over time of the plant predict if (a) Eurythmy has been performed, and (b) which kind of Eurythmy gestures has been performed. We also find that the signals are different based on the species of the plant. In other words, the perception of a salad, tomato, or basil might differ just as perception of different species of animals differ. This opens new ways of studying plant ecosystems, while also paving the way to use plants as biosensors for analyzing human movement.

Keywords: plant-human interaction; signal processing; plant action potentials; machine learning; eurythmy; plant bio-sensors

Thanks!



Do you have any questions?

alvaro.francisco.gil@alumnos.upm.es

alvaro_f@mit.edu



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