## Introduction to metabarcoding

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## Which is our question?

- ▶ Who is in there?
- ► What can they do?
- ► What are they doing?



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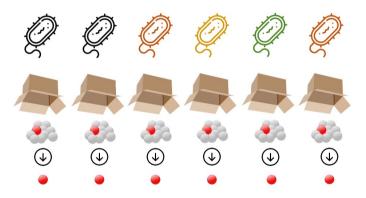




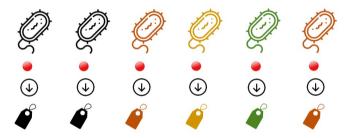




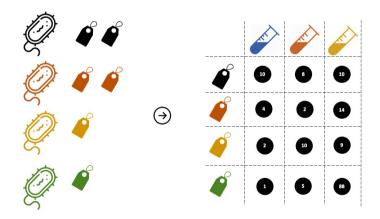
# Metabarcoding



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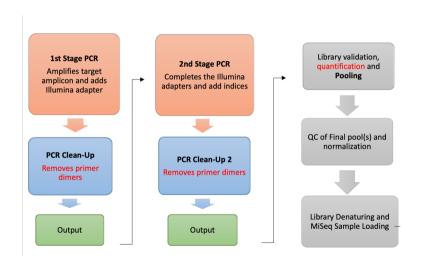
# Metabarcoding



#### General workflow

- 1. Sample collection
- 2. DNA extraction
- 3. PCR amplification of our target
- 4. Adaptor ligation
- 5. Sequencing
- 6. Data analysis

### Library preparation workflow

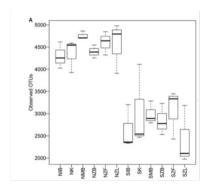


## Technical considerations - experimental design

- ► Target gene (specificity, resolution)
- ► Replicates (technical/biological)
- ► Sequencing depth
- Databases for taxonomy assignment
- Bias

#### Technical considerations - DNA extraction

- ► DNA extraction introduces a bias in the final dataset
- A recent investigation on 322 studies shows they used 72 different methods. 14 did not report such info!

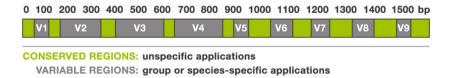


#### Technical considerations - DNA extraction

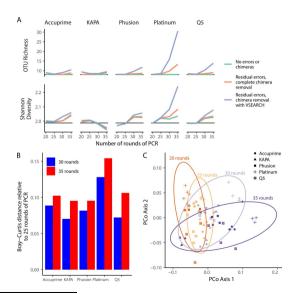
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#### Technical considerations - PCR

- Target gene
  - ▶ Who is our target? Bacteria, fungi, insects, fish, specific genus...
  - Is the resolution optimal for our question?
  - ▶ Are PCR primers available or we have to design them?
  - Is the taxonomy database available or we have to build a custom one?
- PCR bias
  - Use a Hi-Fi polymerase
  - Use optimal annealing temperature
  - ▶ Do not exaggerate with PCR cycles
  - Run multiple PCRs on the same sample



#### Technical considerations - PCR



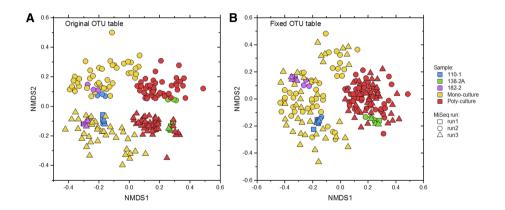
<sup>&</sup>lt;sup>1</sup>Sze and Schloss 2019 mSphere



### Technical considerations - Library prep

- Which sequencing methodology will be a good fit for your research question? (read length/depth of sampling)
- ► How diverse if your system?

## Technical considerations - Sequencing run





<sup>&</sup>lt;sup>1</sup>Song et al. 2018 Phytobiomes

### Control samples

- ▶ **Negative control 1.** Run molecular biology grade water throughout the pipeline. Pool it with the other samples even if you do not see amplification.
- ▶ **Negative control 2.** This is the negative control from your first PCR. If you see a band, discard the entire batch of samples and start again. If no band is observed, sequence anyway.
- ► **Mock community.** Pre-built or custom.

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