## CondAlt 2.1 Manual

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#### Introduction

The *CondAlt 2.1* Software Package predicts alternative structures of an RNA sequence based on conditional base-pair probabilities, as follows. First, the computed MFE structure  $S_1^*$  predicts the dominant structure. Then, the *UNAFold* Software [2, 4-7] computes McCaskill base-pair probabilities conditioned on excluding base pairs close to (i.e., with endpoints near) base pairs in  $S_1^*$ . A calculation of McCaskill conditional base-pair probabilities yields a longest bulge-containing stem seed  $L^*$ , in which the conditional probability of every base-pair is higher than 0.5. Finally, we predict an alternative structure  $S_2^*$  as the lowest-energy structure containing  $L^*$ . the Materials and Methods Section of "Structural Prediction of RNA Switches using Conditional Base-Pair Probabilities" article gives algorithmic details.

# Folding-energy Parameters

Calculation of base-pair probabilities as well as conditional base-pair probabilities are computed via version 3.0 energy parameters of *UNAfold* (version 4.0.0) [8].

#### Installation

UNAfold [2] has to be already installed and available in the main path. The downloadable zip file contains the stand-alone main program in Perl 'altmfe.pl' and accompanying java programs required for the main program. All input and output files are to be in the same folder as 'altmfe.pl'.

#### **Input Parameters**

- 1. -i <fasta file>: Input RNA sequence. The name of file MUST have a ".seq" extension. The input must be in fasta format and start with '>' in its first line. There must be only one RNA sequence in the following line and it must contain unambiguous nucleotides; A, C, G, and U. The end of file must contain at most one newline. The length of the sequence must be in the range of [1,500].
- 2. -d <distance threshold>: Dissimilarity Threshold  $\tau$ . This parameter is used to define the size of the meta-stable structure to be eliminated in conditional probabilities calculations; i.e.,  $\mathcal{E}$ . Higher  $\tau$  leads to higher number of eliminated base pairs. Default is set to 5.
- 3. -t <temperature>: Folding temperature T.  $S_2^*$  can be predicted at different temperatures. For a given T,  $S_1^*$  is still predicted at the default temperature 37°C, but quantities  $\mathcal P$ ,  $L^*$ , and  $S_2^*$  are all calculated at T. See Materials and Methods.

## **Output Files**

Both the alternative structures and the seed locations are available to user. After running the Perl file with the above input parameters, two output files: [input.seq].out and [input.seq]MFE1.aux.

- 1. [input.seq].out contains for the dominant structure  $S_1^*$  and alternative structure  $S_2^*$  (in that order): (1) the free energy; (2) the sequence; and (3) the resulting structures in Stockholm format.
- 2. [input.seq]MFE1.aux contains the base-pairs in the stem seed  $L^*$ .

## A Simple Example:

Consider the following input RNA sequence and parameters:

```
<fasta file>=input.seq
>seq
CGUACGUAGCUAGUCGUACGUACGUACUGA
<distance threshold> = 5
<temperature> = 37
```

Because of the defaults, the following commands are equivalent:

```
perl -i altmfe.pl input.seq -d 5 -t 37
perl -i altmfe.pl input.seq -d 5
perl -i altmfe.pl input.seq -t 37
perl -i altmfe.pl input.seq
```

## They yield:

#### input.seq.out:

```
>-17.5
CGUACGUAGCUAGUCGUACGUACGUACUGA
.((((((((((((.....))))))))))...
>-6.4
CGUACGUAGCUAGUCGUACGUACGUACGUACUGA
.....(((((((....)))))))...
```

#### And input.fastaMFE1.aux:

```
F 16 31 1
F 17 30 1
F 18 29 1
F 19 28 1
F 20 27 1
F 21 26 1
```

#### References

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- 4. Jaeger JA, Turner DH, Zuker M. Predicting optimal and suboptimal secondary structure for RNA. Methods in enzymology. 1990;183:281-306.
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