

Addressing The Scientific Reproducibility Crisis Through Educational Software
Integration

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Table of Contents

Introduction	1
Chapter 1: An Introduction to Reproducibility	3
1.1 What Is Reproducibility?	3
1.2 The Reproducibility Crisis	4
1.3 The Components of Reproducible Research	5
1.4 Current Attempts to Address Reproducibility in Scientific Publishing	7
1.4.1 Case Studies Across The Sciences	7
1.4.2 Case Studies In The Statistical And Data Sciences	9
1.4.3 The Bigger Picture	10
1.4.4 Assessing the Success of Academic Reproducibility Policies	14
1.5 Limitations on Achieving Reproducibility in Scientific Publishing	15
1.5.1 Challenges for Authors	15
1.5.2 Challenges for Journals	16
1.6 Attempts to Address These Limitations	17
1.6.1 Through Education	17
1.6.2 Through Software	19
1.7 Understanding The Gaps In Existing Reproducibility Solutions	21
1.7.1 In Education	21
1.7.2 In Software	22
1.7.3 What We Need Moving Forward	22
Chapter 2: <code>fertile</code>: My Contribution To Addressing Reproducibility	25
2.1 <code>fertile</code> , An R Package Creating Optimal Conditions For Reproducibility	25
2.1.1 Component 1: Accessible Project Files	27
2.1.2 Component 2: Organized Project Structure	28
2.1.3 Component 3: Documentation	31
2.1.4 Component 4: File Paths	34
2.1.5 Component 5: Randomness	37
2.1.6 Component 6: Readability and Style	38
2.1.7 Summary of Reproducibility Components	40
2.1.8 User Customizability	40
2.1.9 Educational Features	50
2.2 How <code>fertile</code> Works	52
2.2.1 Shims	52

2.2.2	Hidden Files	54
2.2.3	Environment Variables	56
2.2.4	The Dots (...)	57
2.2.5	Parameterized Reports	59
2.3	Summary	60
Chapter 3: Incorporating <code>fertile</code> Into The Greater Data Science Com-		
	munity	63
3.1	Potential Applications of <code>fertile</code>	64
3.1.1	In Journal Review	64
3.1.2	For Teaching Reproducibility	67
3.1.3	In Other Areas	70
3.2	Testing <code>fertile</code> in the Real World: Experimental Design and Analysis .	71
3.2.1	Background	71
3.2.2	Experimental Design	71
3.2.3	Results	74
3.2.4	Limitations	86
3.2.5	Implications	89
3.2.6	Future Work	89
Conclusion		91
Appendix A: The First Appendix		95
Appendix B: The Second Appendix, for Fun		97
References		99

List of Figures

1.1	Reproducibility vs. Replicability	4
1.2	Reproducibility Policies of Top Statistical and Data Sciences Journals .	10
1.3	The TOP Factor Rubric	13
1.4	Popular Reproducibility Packages in R	20
1.5	Popular Continuous Integration Tools	21
2.1	fertile's Package Logo	25
2.2	A Sample R Project	26
2.3	Summary of Reproducibility Components and the Related Functionalities in 'fertile'	40
2.4	The 6 Reproducibility Badges Available in 'fertile'	44
2.5	Output: Badges and Reasons for Failure	45
2.6	Output: System/User Information and File List	46
2.7	List of Functions Shimmed by 'fertile'	52
3.1	Potential 'fertile' Journal Review Process	65
3.2	Another Potential 'fertile' Journal Review Process	66
3.3	The Replication Assignment From Harvard Professor Gary King's Gov 2001 Graduate Course	68
3.4	Sample Question From The Section On RStudio Projects	75
3.5	Another Sample Question From The Section On RStudio Projects . .	75
3.6	Sample Question From The Section On File Paths	76

Abstract

The preface pretty much says it all.

Second paragraph of abstract starts here.

Introduction

If it weren't for Professor Ben Baumer, I would not be here writing this thesis today. The Introduction to Data Science course I took with him in my first semester at Smith College spurred on my love for the field, prompting me to declare a major—with Ben as my advisor—soon after.

That course—SDS 192—was my first introduction to the topic of scientific reproducibility. In Ben's class, one of the first undergraduate courses in the country to implement reproducibility as part of the curriculum, I was introduced to a variety of tools that could help make my work more reproducible. I learned how to write effective narrative reports with RMarkdown, how to implement version control on GitHub, how to write well-styled code, and how to use R projects to structure my files in a way that was easy to manage.

Through this work, I began to see the benefits of emphasizing a reproducible workflow. When I worked on projects with other students, the tools I learned in class enabled us to collaborate with one another easily and keep track of the progress we had made. And when I tried to look back at my own work from the past, my attention to reproducibility meant that it was much easier for me to re-compute and understand what I had done previously.

Additionally, my work in Introduction to Data Science helped build a strong interest in R. I developed a love for programming in the language and was curious to learn more about how it worked—both behind the scenes and practice. This interest prompted me to pursue research in the field. I wanted to expand my knowledge and challenge myself by learning about and solving cutting-edge issues in the data science domain.

As soon as I felt that I had built up enough coding experience to be able to tackle challenging issues, I reached out to the data science faculty to see if there were any opportunities. Ben enthusiastically welcomed me in, telling me that he had recently started developing an R package focused on reproducibility and offering me a position assisting him with writing it. That was when I first learned about **fertile**.

As part of his interest in reproducibility, Ben had been working—with help from Hadley Wickham and Jenny Bryan of RStudio—to develop a package designed to help users create optimal conditions for reproducible work in R (titled “fertile”).

When it was presented to me, the package was incomplete. Several of the major functions that still exist now were there at the time, but there were not many features beyond the basics. Additionally, several functions were not fully written or throwing errors, the package had little to no documentation, and the majority of tests that had

been written to test the functions were failing.

I was given the opportunity to join in on the project and help move **fertile** to a functional state: to fix the errors that were present, document the package, and write new functions to expand the operation beyond the barebones structure that was present.

The **fertile** project was an excellent fit for me. The package was not yet in a functional state, but had an excellent base of code to work off of. Given that it already had some basic functions and a strong structure, **fertile** provided a perfect opportunity for me to learn about the process of writing R packages.

The more I worked on **fertile**, the more interested in the project I became. The package was unlike anything else I had been exposed to in data science. Most of my course work had been in the form of front-end coding: conducting data analyses and visualizing information. **fertile** is nothing like that. It operates completely behind the scenes, conducting a kind of meta-analysis to determine the reproducibility of a data science project. Rather than writing code to analyze data, I had to learn how to write code to analyze code that *other people* wrote to analyze data.

This was an incredibly complex problem. It involved capturing information about the user's file system and their RStudio environment, both of which I knew nothing about. It also involved tracking and recording user behavior—something definitely not taught in data analysis courses. Even experienced R users deemed some of the goals associated with the package to be incredibly lofty. On the question of whether it was possible to test whether R code contained uncontrolled randomness, one coder said it was “probably impossible”, while another said there was “fundamentally no way” to achieve it.

The challenge of the problems at hand, however, was what made them interesting. Most of the time, there was no way to use my past coding knowledge to solve them—rather, I had to research and test out new solutions. While the process was at times incredibly frustrating, particularly given that some of the errors I came across had likely never been seen by any other R user, it was an incredible learning opportunity. I was exposed to a wide variety of R features I would likely have never seen otherwise and greatly broadened my problem-solving and troubleshooting skills.

Every problem I solved made me more confident in my abilities and more excited to work on the **fertile**. I delved more into the topic of reproducibility and looked at some of the work by other researchers and code developers. This research introduced me to new aspects of reproducibility I wanted to include in the package and inspired me to expand its scope.

This paper is the culmination of the work I have been doing with **fertile** for the past two years.

Chapter 1

An Introduction to Reproducibility

1.1 What Is Reproducibility?

In the field of data science, research is considered fully *reproducible* when the requisite code and data files produce identical results when run by another analyst, or more generally, when a researcher can “duplicate the results of a prior study using the same materials as were used by the original investigator” (Bollen et al. (2015)).

This term was first coined in 1992 by computer scientist Jon Claerbout, who associated it with a “software platform and set of procedures that permit the reader of a paper to see the entire processing trail from the raw data and code to figures and tables” (Claerbout & Karrenbach (1992)).

Since its inception, the concept of reproducibility has been applied across many different data-intensive fields, including epidemiology, computational biology, economics, clinical trials, and, now, the more general domain of statistical and data sciences (Goodman, Fanelli, & Ioannidis (2016)).

Reproducible research has a wide variety of benefits in the scientific community. When researchers provide the code and data used for their work in a well-organized and reproducible format, readers are more easily able to determine the veracity of any findings by following the steps from raw data to conclusions. The creators of reproducible research can also more easily receive more specific feedback (including bug fixes) on their work. Moreover, others interested in the research topic can use the code to apply the methods and ideas used in one project to their own work with minimal effort.

Although often confused, the concept of *reproducibility* is distinct from the similar idea of *replicability*: the ability of a researcher to duplicate the results of a study when following the original procedure but collecting new data. Replicability has larger-scale implications than reproducibility; the findings of research studies can not be accepted unless a variety of other researchers come to the same conclusions through independent work.

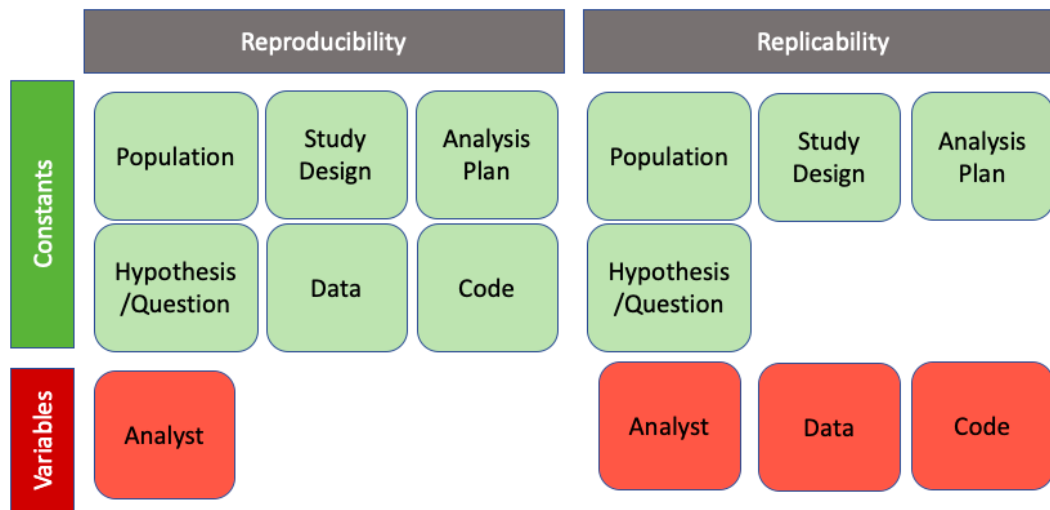


Figure 1.1: Reproducibility vs. Replicability

Reproducibility and replicability are both necessary to the advancement of scientific research, but they vary significantly in terms of their difficulty to achieve. Reproducibility, in theory, is somewhat simple to attain in data analyses—because code is inherently non-random (excepting applications involving random number generation) and data remain consistent, variability is highly restricted. The achievement of replicability, on the other hand, is a much more complex challenge, involving significantly more variability and requiring high quality data, effective study design, and incredibly robust hypotheses.

1.2 The Reproducibility Crisis

Despite the relative simplicity of achieving reproducibility, a significant proportion of the work produced in the scientific community fails to meet reproducibility standards. 52% of respondents in a 2016 Nature survey believed that science was going through a “crisis” of reproducibility. Additionally, the vast majority of researchers across all fields studied reported having been unable to reproduce another researcher’s results, while approximately half reported having been unable to reproduce their own (Baker (2016)). Other studies paint an even bleaker picture: a 2015 study found that over 50% of studies psychology failed reproducibility tests and research from 2012 found that figure closer to 90% in the field of cancer biology (Baker (2015), Begley & Ellis (2012)).

In the past several years, this “crisis” of reproducibility has risen toward the forefront of scientific discussion. Without reproducibility, the scientific community cannot properly verify study results. This makes it difficult to identify which information should be believed and which should not and increases the likelihood that studies sharing misleading information will be dispersed. The rise of data-driven technologies, alongside our newly founded ability to instantly share knowledge worldwide, has made

reproducibility increasingly critical to the advancement of scientific understanding, necessitating the development of solutions for addressing the issue.

Academics have recognized this, and publications on the topic appear to have increased significantly in the last several years (Eisner (2018); Fidler & Wilcox (2018); Gosselin (2020); McArthur (2019); Wallach, Boyack, & Ioannidis (2018)).

1.3 The Components of Reproducible Research

In order to see why there is an issue with reproducibility and gain a sense of how to solve it, it is important to first understand the components of reproducibility. Essentially, to answer, “What parts does researcher need to include, or what steps do they need to take, to be able to declare their work reproducible?”

Publications attempting to answer this can be found in all areas of scientific research. However, as Goodman et al. (2016) argue, the language and conceptual framework used to describe research reproducibility varies significantly across the sciences, and there are no clear standards on reproducibility agreed upon by the scientific community as a whole.

At a minimum, according to Goodman et al. (2016), achieving reproducibility requires the sharing of data (raw or processed), relevant metadata, code, and related software. However, according to other authors, the full achievement of reproducibility may require additional components.

Kitzes, Turek, & Deniz (2017) present a collection of case studies on reproducibility practices from across the data-intensive sciences, illustrating a variety of recommendations and techniques for achieving reproducibility. Although their work does not come to a consensus on the exact standards of reproducibility that should be followed, several common trends and principles emerge from their case studies that extend beyond the minimum recommendations of Goodman et al. (2016):

- 1) use clear separation, labeling, and documentation in provided code,
- 2) automate processes when possible, and
- 3) design the data analysis workflow as a sequence of small steps glued together, with outputs from one step serving as inputs into the next. This is a common suggestion within the computing community, originating as part of the Unix philosophy (Gancarz (2003)).

Cooper et al. (2017) focus on data analysis completed in R and identify a similar list of important reproducibility components, reinforcing the need for clearly labeled, well-documented, and well-separated files. In addition, they recommend publishing a list of software dependencies and using version control to track project changes over time.

Broman (2019) reiterates the need for clear naming and file separation while sharing several additional suggestions: keep the project contained in one directory, use relative paths when accessing the file system, and include a `README` file describing the project.

Wilson et al. (2017) argue that to follow good practice, an analysis must be located in a well-organized directory, be supplemented by a file containing information about the project (i.e., a README), and provide an explicit list of dependencies.

Wilson et al. (2014) emphasize communication, recommending that code be human-readable and consistently styled for ease of understanding once shared.

The reproducibility recommendations from R OpenSci, a non-profit initiative founded in 2011 to make scientific data retrieval reproducible, share similar principles to those discussed previously. They focus on a need for a well-developed file system, with no extraneous files and clear labeling. They also reiterate the need to note dependencies and use automation when possible, while making clear a suggestion not present in the previously-discussed literature: the need to use seeds, which allow for the saving and restoring of the random number generator state, when running code involving randomness (Martinez et al. (2018)).

Although these recommendations differ from one another, when considered in combination they provide a well-rounded picture of the components important to research reproducibility across the scientific community:

1. The basic project components are made accessible to the public:
 - Data (raw and/or processed)
 - Metadata
 - Code
 - Related Software
2. The file structure of project is well-organized:
 - Separate folders for different file types.
 - No extraneous files.
 - Minimal clutter.
3. The project is documented well:
 - Files are clearly named, preferably in a way where the order in which they should be run is clear.
 - A README is present.
 - Code contains comments.
 - Software dependencies are noted.
4. File paths used in code are not system- or user-dependent:
 - No absolute paths.
 - No paths leading to locations outside of a project's directory.
 - Only relative paths, pointing to locations within a project's directory, are permitted.
5. Randomness is accounted for:

- If randomness is used in code, a seed must also be set.
6. Code is readable and consistently styled:
- Though not mentioned in the sources described previously, it is also important that code be written in a coherent style. This is because code that conforms to a style guide or is written in a consistent dialect is easier to read, simplifying the process of following a researcher's work from beginning to end (Hermans & Aldewereld (2017)).

1.4 Current Attempts to Address Reproducibility in Scientific Publishing

In an attempt to increase reproducibility, leaders from academic journals around the world have taken steps to create new standards and requirements for submitted articles. These standards attempt to address the components of reproducibility listed previously, requesting that authors provide certain materials necessary for reproducing their work when they submit an article. However, these standards are highly inconsistent, varying significantly both across and within disciplines, and many only cover one or two of the six primary components, if any at all.

To illustrate this point, we will consider several case studies from journals publishing research on a variety of scientific fields.

1.4.1 Case Studies Across The Sciences

The journal whose requirements appear to align most closely with those components defined previously in Section 3 is the *American Journal of Political Science* (AJPS). In 2012, the AJPS became the first political science journal to require authors to make their data openly accessible online, and the publication has instituted stricter requirements since. AJPS now requires that authors submit the following alongside their papers (American Journal of Political Science (2016)).

- The dataset analyzed in the paper and information about its source. If the dataset has been processed, instructions for manipulating the raw data to achieve the final data must also be shared.
- Detailed, clear code necessary for reproducing all of the tables and figures in the paper.
- Documentation, including a README and codebook.
- Information about the software used to conduct the analysis, including the specific versions and packages used.

These standards are quite thorough and contain mandates for the inclusion of the vast majority of components necessary for complete reproducibility. Most journals, however, do not come close to meeting such high standards in their reproducibility statements.

For example, in the biomedical sciences, a group of editors representing over 30 major journals met in 2014 to address reproducibility in their field, coming to a consensus on a set of principles they wanted to uphold (National Institutes of Health (2014)). Listed below are those relating specifically to the use of data and statistical methods:

- 1) Journals in the biomedical sciences should have a mechanism to check the statistical accuracy of submissions.
- 2) Journals should have no (or generous) limit on methods section length.
- 3) Journals should use a checklist to ensure the reporting of key information, including:
 - The article meets nomenclature/reporting standards of the biomedical field.
 - Investigators report how often each experiment was performed and whether results were substantiated by repetition under a range of conditions.
 - Statistics must be fully reported in the paper (including test used, value of N, definition of center, dispersion and precision measures).
 - Authors must state whether samples were randomized and how.
 - Authors must state whether the experiment was blinded.
 - Authors must clearly state the criteria used for exclusion of any data or subjects and must include all results, even those that do not support the main findings.
- 4) All datasets used in analysis must be made available on request and should be hosted on public repositories when possible. If not possible, data values should be presented in the paper or supplementary information.
- 5) Software sharing should be encouraged. At the minimum, authors should provide a statement describing if software is available and how to obtain it.

Even though these principles seem well-developed on the surface, they fail to meet even the basic requirements defined by Goodman et al. (2016) previously. Several of the principles are purely recommendations; there is no requirement that code be shared, nor metadata. Additionally, software requirements are quite loose, requiring no information about dependencies or software version.

We see a similar issue even in journals designed specifically for the purpose of improving scientific reproducibility. *Experimental Results*, a publication created by Cambridge University Press to address some of the reproducibility and open access issues in academia, also falls short of meeting high standards. The journal, which showcases articles from a variety of scientific disciplines, states in their transparency and openness policy:

Whenever possible authors should make evidence and resources that underpin published findings, such as data, code, and other materials, available to readers without undue barriers to access.

The inclusion of code and data are only recommended and no definition of what “other materials” may mean is provided. No components of reproducibility extending beyond those required at a minimum are even considered (Cambridge University Press (2020)).

The *American Economic Review*, the first of the top economics journals to require the inclusion of data alongside publications, has stronger guidelines than several of those mentioned previously, though not as strong as the *American Journal of Political Science*. Their Data and Code Availability Policy states the following (American Economic Association (2020)):

It is the policy of the American Economic Association to publish papers only if the data and code used in the analysis are clearly and precisely documented, and access to the data and code is clearly and precisely documented and is non-exclusive to the authors.

These requirements are quite strict, prohibiting exceptions for papers using data or code not available to the public in the way that many other journals claiming to promote reproducibility do.

1.4.2 Case Studies In The Statistical And Data Sciences

When considering reproducibility policy, the field of Statistical and Data Sciences performs relatively well. The majority of highly ranked journals in the field contain statements on reproducibility. Some of these are quite robust, surpassing the requirements of many of the other journals discussed previously, while others are lacking.



















The *Journal of the American Statistical Association* stands out as having relatively robust requirements. The publication’s guidelines require that data be made publicly available at the time of publication except for reasons of security or confidentiality. It is strongly recommended that code be deposited in open repositories. If data is used in a process form, the provided code should include the necessary cleaning/preparation steps. Data must be in an easily understood form and a data dictionary should be included. Code should also be in a form that can be used and understood by others, including consistent and readable syntax and comments. Workflows involving more than one script should also contain a master script, Makefile, or other mechanism that makes it clear what each component does, in what order to run them, and what the inputs and outputs to each area (American Statistical Association (2020)).

The *Journal of Statistical Software* also has strong guidelines, though less thorough. Authors must provide *commented* source code for their software; all figures, tables, and output must be exactly reproducible on at least one platform, and random number generation must be controlled; and replication materials (typically in the form of a script) must be provided (Journal of Statistical Software (2020)).

The expectations of the *Journal of Computational and Graphical Statistics* are notably weaker, requiring only that authors “submit code and datasets as online supplements to the manuscript,” with exceptions for security or confidentiality, but providing no further detail (Journal of Computational and Graphical Statistics (2020)).

The *R Journal* has the same requirements, but with no exceptions on the data provision policy, stating that authors should “not use such datasets as examples” (R Journal Editors (2020)).

Perhaps the weakest reproducibility policies come from *The American Statistician* and the *Annals of Statistics*. The former appears to have no requirements, stating only that it “strongly encourages authors to submit datasets, code, other programs, and/or appendices that are directly relevant to their submitted articles,” while the latter appears to have no statement on reproducibility at all (The American Statistician (2020)).

Journal Name	Code Sharing Required?	Data Sharing Required?	Other Components Required?
Journal of the American Statistical Association			
Journal of Statistical Software			
Journal of Computational and Graphical Statistics			
The R Journal			
The American Statistician			
The Annals of Statistics			

★ Component recommended, but not required.

◆ Component required, but exceptions granted.

Figure 1.2: Reproducibility Policies of Top Statistical and Data Sciences Journals

1.4.3 The Bigger Picture

The journals mentioned here are just some of the many academic publishers with reproducibility policies. While they provide a sense of the specific wording and requirements of some policies, they do not necessarily serve as a representative sample of all academic publishing. It is important to also consider the bigger picture, exploring the state of reproducibility policy in academic publishing as a whole.

Given the scale of the academic publishing network and the sheer number of journals around the world, this is not necessarily an easy task.

In order to simplify this process, academics at the Center for Open Science (COS) attempted to create a metric, called the TOP Factor. The TOP Factor reports the steps that a journal is taking to implement open science practices. It has been calculated for a wide variety of journals, though the COS is still far from scoring all of the publications that are currently available.

The TOP Factor is calculated as follows. Publications are scored on a variety of categories associated with open science and reproducibility. For each category, they receive a score between 0 (poor) and 3 (excellent) based on the degree to which they emphasize each category in their submission/publication policies. A journal's final score, which can range from 0 to 30, is the sum of the individual scores in each of the categories.

	Not Implemented	Level I	Level II	Level III
Citation standards	Journal encourages citation of data, code and materials or says nothing.	Journal describes citation of data in guidelines to authors with clear rules and examples	Article provides appropriate citation for data and materials used consistent with journal's author guidelines	Article is not published until providing appropriate citation for data and materials following journal's author guidelines
Data transparency	Journal encourages data sharing or says nothing	Article states whether data are available and, if so, where to access them	Data must be posted to a trusted repository. Exceptions must be identified at article submission	Data must be posted to a trusted repository, and reported analyses will be reproduced independently prior to publication
Code transparency	Journal encourages code sharing or says nothing	Article states whether code is available and, if so, where to access it	Code must be posted to a trusted repository. Exceptions must be identified at article submission	Code must be posted to a trusted repository, and reported analyses will be reproduced independently prior to publication
Research materials transparency	Journal encourages materials sharing or says nothing	Article states whether materials are available and, if so, where to access them	Materials must be posted to a trusted repository. Exceptions must be identified at article submission	Materials must be posted to a trusted repository, and reported analyses will be reproduced independently prior to publication

Design and analysis transparency	Journal encourages design and analysis transparency or says nothing	Journal articulates design transparency standards	Journal requires adherence to design transparency standards for review and publication	Journal requires and enforces adherence to design transparency standards for review and publication
Study preregistration	Journal says nothing	Article states whether preregistration of study exists and, if so, where to access it	Article states whether preregistration of study exists and, if so, allows journal access during peer review for verification	Journal requires preregistration of studies and provides link and badge in article to meeting requirements
Analysis plan preregistration	Journal says nothing	Article states whether preregistration of study exists and, if so, where to access it	Article states whether preregistration with analysis plan exists and, if so, allows journal access during peer review for verification	Journal requires preregistration of studies with analysis plans and provides link and badge in article to meeting requirements
Replication	Journal discourages submission of replication studies or says nothing	Journal encourages submission of replication studies	Journal encourages submission of replication studies and conducts results blind review	Journal uses registered reports as a submission option for replication studies with peer review prior to observing the study outcomes

Figure 1.3: The TOP Factor Rubric

When looking at the overall distribution of TOP Factor scores, we see a relatively grim picture: Around 50% of journals score as low as 0-5 overall, while only just over 5% score more than 15, just half of the maximum possible score. Over 40 journals failed to score a single point (Woolston (2020)).

Although it is clear that some journals have relatively strong reproducibility and openness policies, that is clearly not the norm. And many that do appear to have policies lacking in robustness, including exceptions for data privacy and security concerns or phrasing guidelines as recommendations rather than requirements. The field of data science stands out among the rest, with the majority of top journals

having relatively robust policies.

1.4.4 Assessing the Success of Academic Reproducibility Policies

We have seen that, although not necessarily the standard, some journals across the sciences have enacted reproducibility policies. The simple implementation of a policy, however, does not ensure that its goals will be achieved. Reproducibility can only be addressed when both authors *and* journal reviewers actively implement publishing standards in practice. Without participation and dedication from all involved, reproducibility guidelines serve more as a theoretical goal than a practical achievement.

It is important to ask, then, whether academic reproducibility standards *actually* result in a greater number of reproducible publications.

Let us consider the case of the journal *Science*. *Science* instituted a reproducibility policy in 2011 and has maintained it ever since. In its original form, their policy stated the following:

All data necessary to understand, assess, and extend the conclusions of the manuscript must be available to any reader of *Science*. All computer codes involved in the creation or analysis of data must also be available to any reader of *Science*. After publication, all reasonable requests for data and materials must be fulfilled. Any restrictions on the availability of data, codes, or materials... must be disclosed to the editors upon submission...

This policy is similar to many of the others considered previously, requiring the publishing of code and data with exceptions permitted when necessary.

Stodden et al. (2018a) tested the efficacy of this policy in practice, emailing corresponding authors of 204 articles published in the year after *Science* first implemented its policy to request the data and code associated with their articles. The researchers only received (at least some of) the requested material from 36% of authors. This low rates were due to several factors:

- 26% of authors did not respond to email contact.
- 11% of authors were unwilling to provide the data or code without further information regarding the researchers' intentions.
- 11% asked the researchers to contact someone else and that person did not respond.
- 7% refused to share data and/or code.
- 3% directed the researchers back to their paper's supplemental information section.
- 3% of authors made a promise to follow up and then did not follow through.
- 3% of emails bounced.
- 2% gave reasons why they could not share for ethical reasons, size limitations, or some other reason.

Of the 56 papers they deemed likely reproducible, the authors randomly selected 22 and were able to replicate the results for all but 1, which failed due to its reliance on software that was no longer available.

Hardwicke et al. (2018) conducted a study on the journal *Cognition*, where researchers compared the reproducibility of published work both before and after the journal instituted an open data policy, which required that authors make relevant research data publicly available prior to publication of an article.

The researchers found a considerable increase in the proportion of data available statements (in contrast to ‘data not available’ statements, which could be present due to privacy or security concerns) since the implementation of the policy. Pre-open data policy, only 25% of articles had data available, while that number was a much higher 78% after the policy was put in place.

While the institution of an open data policy appears to have been associated with a significant increase in the percentage of studies with data available, further research indicates that the policy was perhaps not as effective as intended. Many of the datasets were usable in theory, but not in practice. Only 62% of the articles with data available statements had truly reusable datasets—in this case, meaning that the data were accessible, complete, and understandable. Though this is an increase from the pre-policy period, which saw 49% of articles with data availability statements as reusable in practice, it is still far from ideal.

Combining these two data points indicates that *less than half* of articles published after the open data policy was instituted actually contained truly usable data.

In this small sample of cases, we see that purely having a reproducibility statement does not necessarily mean that all, or even a majority, of published work will truly be reproducible.

1.5 Limitations on Achieving Reproducibility in Scientific Publishing

There are several reasons for this apparent divide between journal reproducibility standards and the true proportion of submitted articles that are truly reproducible. Some of these are challenges faced by the article authors, while others are faced by the journal editors.

1.5.1 Challenges for Authors

Stodden et al. (2018b) conducted a survey asking over 7,700 researchers about one of the key characteristics of reproducibility – open data – and gathered information about the reasons why authors found difficulties in making their data available to the public

The main challenges listed by respondents were as follows:

- 46% identified “Organizing data in a presentable and useful way” to be difficult.
- 37% had been “Unsure about copyright and licensing.”

- 33% had problems with “Not knowing which repository to use.”
- 26% cited a “Lack of time to deposit data.”
- 19% found the “Costs of sharing data” to be high.

The relative frequency of these issues varied across several characteristics, including author seniority, subject area, and geographical location, though authors in all categories faced some issues.

Beyond technical challenges, other reasons may lead authors to not place their focus on reproducibility. For example, some researchers might fear damage to their reputation if a reproduction attempt fails after they have provided the necessary materials (Lupia & Elman (2014)).

Given the relatively high frequency of concern over achieving reproducibility, it follows that researchers will not make the necessary effort to do so if journal guidelines provide a way out. Policies that *recommend* the inclusion of data or that allow exceptions to open data for certain reasons are likely to be associated with a lower proportion of reproducible articles than those that make open data mandatory.

1.5.2 Challenges for Journals

In addition to the challenges faced on the part of the authors, journal reviewers face their own difficulties in ensuring reproducibility.

In order to make sure that all submitted articles comply with reproducibility guidelines, reviewers must go through them one by one and reproduce all of the results by hand using the provided materials.

This is an incredibly intensive process, as we will see in the example of the *American Journal for Political Science* (AJPS), whose reproducibility policy was discussed previously in Chapter 1.4.1.

Jacoby, Lafferty-Hess, & Christian (2017) describe the AJPS process in detail:

Acceptance of an article for publication in the AJPS is contingent on successful reproducibility of any empirical analyses reported in the article.

After an article is submitted, staff from a third party vendor hired by AJPS go through the provided materials to ensure that they can be preserved, understood, and used by others. They then run all of the analyses in the article using the code, instructions, and data provided by the authors and compare their results to the submitted articles. Authors are then given an opportunity to resolve any issues that come up. This process is repeated until reproducibility is ensured.

Although providing a significant benefit to the scientific community, this thorough process is associated with high costs.

The verification process slows down the journal review process significantly, adding a median 53 days to the publication workflow, as many submitted articles require one or more rounds of resubmission (the average number of resubmissions is 1.7). It is also quite labor intensive, taking an average of 8 person-hours per manuscript to reproduce the analyses and prepare the materials for public release and adding significant monetary cost to AJPS.

Journals are often reluctant to take on such an intensive task due to the drastically

increased burden it places on reviewers and on the publication's financial resources. This is particularly true given that the number of submitted articles per year has been increasing over time (Leopold (2015)). Every additional submission increases the burden of achieving reproducibility, and with a large enough volume, the challenge can quickly become seemingly impossible to manage reasonably.

As a result, journals often encourage reviewers to consider authors' compliance with data sharing policies, but do not formally require that they ensure it as a criterion for acceptance (Hrynaszkiewicz (2020)).

1.6 Attempts to Address These Limitations

The previous discussion makes clear that, although reproducibility is critically important to scientific progress and academic journals are taking steps to encourage it, the scientific community is far from achieving the desired level of widespread reproducibility. In large part, this appears due to the challenge and complexity of actually achieving reproducibility. Those attempting to improve the reproducibility of work can face issues with concerns over legality of sharing data, large commitments of time or money, challenges finding a good repository, and organizing all of the many components of their work in an understandable way, among other things.

Additionally, science faces the additional challenge that many publishers do not emphasize reproducibility at all, providing many opportunities for all authors except those personally dedicated to producing reproducible work to leave reproducibility by the wayside. Many journals have no reproducibility requirements, and those that do often do not take the necessary steps to ensure that they are actually met.

These issues, however, are not impossible to overcome. Proponents of reproducibility have taken action to help address them, both through education on reproducibility and through software that helps simplify the process of achieving it.

1.6.1 Through Education

One way to address the reproducibility crisis is to educate data analysts on the topic so that they are aware of both the concept of reproducibility and how to achieve it in their own work. A natural place to focus this education is early on in the data science training pipeline as part of introductory or early-intermediate courses in undergraduate and graduate data science programs (Horton, Baumer, & Wickham (2014)). This sort of educational integration has a variety of benefits:

- Bringing reproducibility into the discussion early on gives students the tools to add knowledge to their field in the best way possible before they actually conduct any substantive analysis on their own (Janz (2016)). This produces many long run benefits, helping lessen the burden on promoting reproducibility placed on journals and increasing the number (and percentage) of researchers doing and promoting reproducible work.
- If covered in detail as part of the data science curriculum, reproducibility will

eventually come easily to students. If learned independently, without effective tools, reproducibility can be challenging and even disheartening to try to understand and succeed at. Practicing in the classroom gives students the ability to fail without damaging their reputation, giving a great opportunity to truly learn and understand the concepts so that they feel capable of handling them when they begin their own research.

- The application of grading to the topic provides an incentive for students to pay attention, learn, and absorb the information. This same incentive does not exist when researchers attempt to learn about reproducibility independently. In that situation, internal motivation, which may be weak in some individuals, is the only factor present to help promote success.

Several educators, primarily at the graduate level, have realized the opportunity and taken steps to introduce reproducibility into their courses. According to Janz (2016), the primary way of achieving this integration is through the assignment of “replication studies” in standard methods choices. In these assignments, students are given a published study and its supporting materials and asked to reproduce the results. The most famous course of this kind is Government 2001, taught by Gary King at Harvard University. In King’s course, students team up in small groups to reproduce a previous study. To help ensure that their workflow is reproducible, students are required to hand over their data and code to another student team who then tries to reproduce their work once again.

In Thomas M. Carsey’s intermediate statistics course at the University of North Carolina at Chapel Hill, students must reproduce the findings of a study by re-collecting the data from the original sources, then must extend the study by building on the analysis.

Christopher Fariss of Penn State University asks his students to replicate a research paper published in the last five years, noting that students must describe the article and the ease in which the results replicate.

The University of California at Berkeley has a similar course to Gary King’s Harvard course, where students each take a different piece of an existing study to work on reproducing and have to ensure that their piece fits with the piece of the next student (Hillenbrand (2014)).

At the undergraduate level, rather than assign replication studies the way many graduate schools tend to do, Smith College and Duke University have both integrated reproducibility into their introductory courses through the requirement that assignments be completed in the `RMarkdown` code + narration format (Baumer, Cetinkaya-Rundel, Bray, Loi, & Horton (2014)).

Another way to provide education on reproducibility is through the creation of workshops that focus solely on the topic, rather than through integration as just one part of a class (@Janz (2016)).

For example, the University of Cambridge conducts a Replication Workshop, where graduate students are asked to replicate a paper in their field over eight weekly sessions. When students encounter challenges, such as authors not responding to queries for

data or steps of the analysis being poorly defined and explained, they gain a first hand understanding of the consequences of poor transparency.

Workshops such as these are typically optional and not included as part of the primary curriculum, however, so while they may cover the topic of reproducibility in more detail than traditional courses, they often reach fewer students.

In spite of all of the advantages that these educational tools provide, “reproducibility training and assessment in data science education is largely neglected, especially among undergraduates and Master’s students in professional schools. . . , probably because the students are usually considered to be non-research oriented” (Yu & Hu (2019)). While some examples of reproducibility education exist, they are certainly not commonplace. However, given the increased discussion and emphasis on reproducibility in academia over the past several years, it is likely that this will change, particularly if methods are provided to educators to make the integration of reproducibility into their courses simple and relatively unburdensome.

1.6.2 Through Software

Several researchers and members of the Statistical and Data Sciences community have taken action to develop software focused on reproducibility which removes some of the load on data analysts by automating reproducibility processes and checking whether certain components are achieved.

Much of this software has been written for users of the coding and data analysis language R. R is very popular in the data science community due to its open-source nature, accessibility, extensive developer and user base, and statistical analysis-specific features.

Some of the existing software solutions are listed below:

rrtools (Marwick (2019)) addresses many of the issues discussed in Marwick, Boettiger, & Mullen (2018) by creating a basic reproducible structure based on the R package format for a data analysis project. In addition, it allows for isolation of the computer environment using **Docker**, provides a method to capture information about the versions of packages used in a project, contains tools for generating a README file, and provides an option for users to write tests to check that their functions operate as intended.

The **orderly** (FitzJohn et al. (2020)) package also focuses on file structure, requiring the user to declare a desired project structure (typically a step-by-step structure, where outputs from one step are inputs into the next) at the beginning and then creating the files necessary to achieve that structure. Its principal aim is to automate many of the basic steps involved in writing analyses, making it simple to:

- 1) Track all inputs into an analysis.
- 2) Store multiple versions of an analysis where it is repeated.
- 3) Track outputs of an analysis.
- 4) Create analyses that depend on the outputs of previous analyses.

When projects have a variety of components, **orderly** makes it easy to see inputs and outputs change with each re-run.

workflowr's (Blischak, Carbonetto, & Stephens (2019)) functionality is based around version control and making code easily available online. It works to generate a website containing time-stamped, versioned, and documented results. In addition, it manages the session and package information of each analysis and controls random number generation.

checkers (Ross, DeCicco, & Randhawa (2018)) allows you to create custom checks that examine different aspects of reproducibility. It also contains some pre-built checks, such as seeing if users reference packages that are less preferred to other similar ones and ensuring that the project is under version control. **renv** (Ushey & RStudio (2020)) (formerly **packrat**) helps to make projects more isolated, portable, and reproducible. It gives every project its own private package library, makes it easy to install the packages the project depends on if it is moved to another computer.

drake (OpenSci (2020)) analyzes workflows, skips steps where results are up to date, utilizes optimized computing to complete the rest of the steps, and provides evidence that results match the underlying code and data.

Lastly, the **reproducible** (McIntire & Chubaty (2020)) package focuses on the concept of caching: saving information so that projects can be run faster each time they are re-completed from the start.



Figure 1.4: Popular Reproducibility Packages in R

There have also been several **Continuous integration** tools developed outside of R which can be used by those coding in almost any language. These provide more general approaches to automated checking, which can enhance reproducibility with minimal code.

For example, **wercker**—a command line tool that leverages Docker—enables users to test whether their projects will successfully compile when run on a variety of operating systems without access to the user's local hard drive (Oracle Corporation (2019)).

GitHub Actions is integrated into GitHub and can be configured to do similar checks on projects hosted in repositories.

Travis CI and **Circle CI** are popular continuous integration tools that can also be used to check R code.

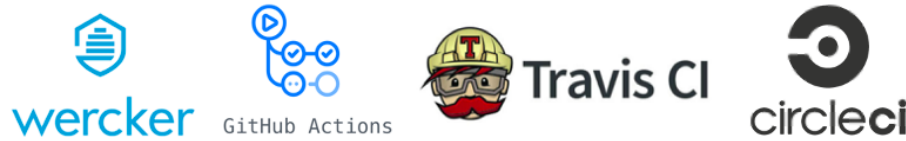


Figure 1.5: Popular Continuous Integration Tools

1.7 Understanding The Gaps In Existing Reproducibility Solutions

Although the current state of reproducibility in academia is quite poor, it is not an impossible challenge to overcome. The relative simplicity of addressing reproducibility, particularly when compared with replicability, makes it an ideal candidate for solution-building. Although significant progress on addressing reproducibility on a widespread scale is a long-term challenge, impactful forward progress—if on a smaller scale—can be achieved in the short-term.

As we have seen, software developers, data scientists, and educators around the world have realized this potential, taking steps to help address the current crisis of reproducibility. Journals have put in place guidelines for authors, statisticians have developed R packages that help structure projects in a reproducible format, and educators have begun integrate reproducibility exercises into their courses.

We have already explored the issues with journal policies, both for authors and reviewers, in-depth. Educational and software-based strategies attempt to address these policy issues by spreading ideas of reproducibility to more people and simplifying the process of achieving it, therefore resulting (in theory) in an improvement in the reproducibility of published scientific articles.

However, the reality is not quite so rosy. Many current educational and software-based solutions face their own challenges that limit them from achieving the desired outcome. In this section, we will consider these issues.

1.7.1 In Education

The two primary concerns about the integration of reproducibility in data science curricula revolve around time and difficulty.

As noted previously, the primary mode of teaching reproducibility is through the assignment of replication studies where students must take an existing study and go through the process of reproducing it themselves, including contacting the author for all necessary materials, rerunning code and analysis, and problem-solving when issues almost certainly come up.

In addition to the time required for the professor to collect all of the studies that students will be working on, the inclusion of such an assignment places a significant burden on educators by taking up time where they could be teaching other important material. Replication studies, if done correctly, can take weeks for students to

successfully complete. The choice to give such assignments is therefore associated with a significant opportunity cost which many professors are unwilling to take.

Additionally, both replication studies assigned in class and replication workshops outside of normal coursework require a working knowledge of how to successfully complete and understand research. This makes them inaccessible to individuals who are still in their undergraduate career and may not yet have had an opportunity to conduct research or those who are studying in non-research-focused technical programs.

In order to reach the widest variety of students possible, it is necessary to develop a new method of teaching reproducibility that is neither time consuming nor dependent on a prior understanding of the research process.

1.7.2 In Software

Previously, we considered several different types of software solutions: packages designed for users of R and continuous integration programs that can be used alongside a variety of coding languages. Although these solutions have their advantages, they also have significant drawbacks in terms of their ability to address reproducibility on a widespread scale.

Many of the packages designed for R are incredibly narrow in scope, with each effectively addressing a small component of reproducibility: file structure, modularization of code, version control, etc. They often succeed in their area of focus, but at the cost of accessibility to a wider audience. Their functions are often quite complex to use, and many steps must be completed to achieve the required reproducibility goal. This cumbersome nature means that most reproducibility packages currently available are not easily accessible to users with minimal R experience, nor particularly useful to those looking for quick and easy reproducibility checks. The significant learning curve associated with them can also detract potential users who may be interested in reproducibility but not willing to dedicate an extensive amount of time to understanding the intricacies of software operation.

Due to their generalized design, Continuous Integration tools do not face the same issues with narrowness or complexity that R packages struggle with. However, this generalizability provides its own additional challenge. Since Continuous Integration tools are designed to be accessible to a wide variety of users with different coding preferences, they are not particularly customizable and lack the ability to address features specific to certain programming languages.

Neither of these different software solutions appear to adequately address the challenge of reproducibility.

1.7.3 What We Need Moving Forward

While a variety of attempts to address reproducibility have been made, they all face their own set of challenges. Most focus on only one area of reproducibility, are too time consuming and burdensome to attempt, or require an extensive amount of background knowledge.

In order to truly improve scientific reproducibility, a better solution is needed.

The optimal solution should combine the positive aspects of the previously discussed educational and software methods, while remaining simple and easy to use. It should also have a variety of potential applications—journal reviewers should benefit from it, as should authors, as should educators and students and even those outside of academia.

The full list of necessary features for an effective reproducibility tool are provided below:

- 1) Be simple, with a small library of functions/tools that are straightforward to use.
- 2) Be accessible to a variety of users, with a relatively small learning curve.
- 3) Be able to address a wide variety of aspects of reproducibility, rather than just one or two key issues.
- 4) Have features specific to a particular coding language that can address that language's unique challenges.
- 5) Be customizable, allowing users to choose for themselves which aspects of reproducibility they want to focus on.
- 6) Be educational, teaching those that use it about why their projects are not reproducible and how to correct that in the future.
- 7) Be applicable to a wide variety of domains.

While this seems like a lot to ask, such a task is not impossible. In the next chapter, we will consider one potential solution: **fertile**, an R package focused on reproducibility that I have worked to develop.

Chapter 2

fertile: My Contribution To Addressing Reproducibility

2.1 `fertile`, An R Package Creating Optimal Conditions For Reproducibility

What if it were possible to address the existing issues with both educational and software reproducibility solutions simultaneously?

That is where my work comes in. In an attempt to produce meaningful change in the field of reproducibility, I have been developing `fertile`, a software package designed for R which helps users create optimal conditions for achieving reproducibility in their projects.



Figure 2.1: `fertile`'s Package Logo

`fertile` attempts to address the gaps in existing reproducibility solutions by combining software and education in one product. The package provides a set of simple, easy-to-learn tools that, rather than focus intensely on a specific area like other software programs, provide some information about all six major components of reproducibility. It is also designed to be incredibly flexible, offering benefits to users at any stage in the data analysis workflow and providing users with the option to

select which aspects of reproducibility they want to focus on.

fertile also contains several R-specific features, which address certain aspects of reproducibility that can be missed by external project development tools. It is designed primarily to be used on data analyses organized as R Projects (i.e. directories containing an `.Rproj` file) and contains several associated features to ensure that the project structure meets the standards discussed in the R community.

In addition, **fertile** is designed to be educational, teaching its users about the components of reproducibility and how to achieve them in their work. The package provides users with detailed reports on the aspects of reproducibility where their projects fell short, identifying the root causes and, in many cases, providing a recommended solution.

fertile is structured in such a way as to be understandable and operable to individuals of any skill level, from students in their first undergraduate data science course to experienced PhD statisticians. The majority of its tools can be accessed in only a handful of functions with minimal required arguments. This simplicity makes the process of achieving and learning about reproducibility accessible to a wide audience in a way that complex software programs or graduate courses requiring an advanced knowledge of research methods do not.

Reproducibility is significantly easier to achieve when all of the tools necessary to do so are located in one place. **fertile** provides this optimal all-inclusive structure, addressing all six of the primary components of reproducibility discussed in Chapter 1. We will consider **fertile**'s treatment of each of these components in turn, exploring its analysis of the sample R project shown below, titled `project_miceps`:

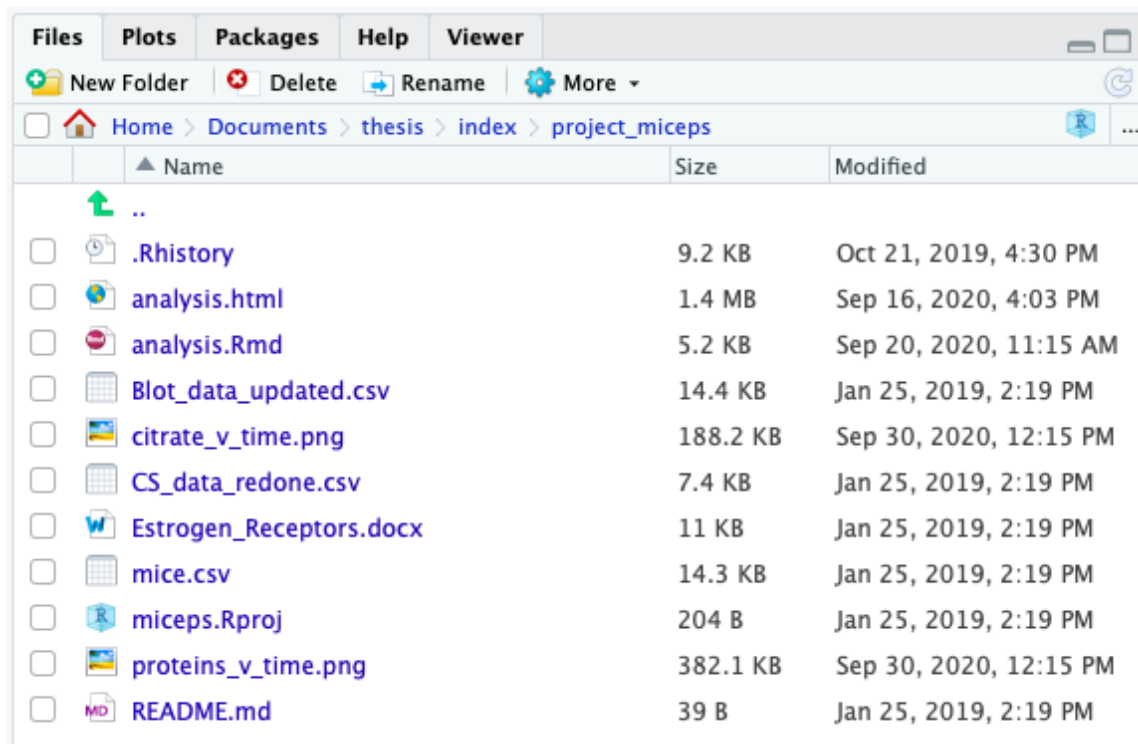


Figure 2.2: A Sample R Project

2.1.1 Component 1: Accessible Project Files

```
library(fertile)
```

`fertile` takes several steps to help users ensure that all of files necessary to run an analysis are provided in the project folder.

File Overview

One of the fastest ways to gain an overview of the existing files (including data, code, and metadata) is with the `proj_analyze_files()` function. It lists all of the files in the project, along with their size, type, and relative path within the directory. This can help users quickly produce an overview of how many code, data, and auxilliary (image or text) files they have.

```
proj_analyze_files("project_miceps")
```

```
# A tibble: 11 x 5
```

	file	size	ext	mime	path_rel
	<fs::path>	<fs::by>	<chr>	<chr>	<fs::path>
1	project_miceps/Blot~	14.43K	csv	text/csv	Blot_data_upd~
2	project_miceps/CS_d~	7.39K	csv	text/csv	CS_data_redon~
3	project_miceps/Estr~	10.97K	docx	application/vnd.openxmlfo~	Estrogen_Rece~
4	project_miceps/READ~	39	md	text/markdown	README.md
5	project_miceps/anal~	5.21K	Rmd	text/x-markdown	analysis.Rmd
6	project_miceps/anal~	1.41M	html	text/html	analysis.html
7	project_miceps/citr~	187.1K	png	image/png	citrate_v_tim~
8	project_miceps/mice~	14.33K	csv	text/csv	mice.csv
9	project_miceps/mice~	204	Rproj	text/rstudio	miceps.Rproj
10	project_miceps/prot~	378.74K	png	image/png	proteins_v_ti~
11	project_miceps/soft~	5.34K	txt	text/plain	software-vers~

Users can also check for the existence of a README description file with `has_readme()`:

```
has_readme("project_miceps")
```

```
v Checking for README file(s) at root level
```

Testing For Self-Containment

In order to truly check that a project is self contained, however, is is necessary to remove it from the environment it is located, ensuring that it can still run when cut off from the main file system.

The `sandbox()` function is designed to help facilitate this. `sandbox()` allows the user to make a copy of their project in a temporary directory that is isolated from the file system.

```
fs::dir_ls('project_miceps') %>% head(3)
```

```
project_miceps/Blot_data_updated.csv    project_miceps/CS_data_redone.csv
project_miceps/Estrogen_Receptors.docx
```

```
temp_dir <- sandbox('project_miceps')
temp_dir
```

```
/var/folders/v6/f62qz88s0sd5n3yqw9d8sb300000gn/T/RtmpRlc45x/project_miceps
```

```
fs::dir_ls(temp_dir) %>% head(3)
```

```
/var/folders/v6/f62qz88s0sd5n3yqw9d8sb300000gn/T/RtmpRlc45x/project_miceps/Blot_data_updated.csv
/var/folders/v6/f62qz88s0sd5n3yqw9d8sb300000gn/T/RtmpRlc45x/project_miceps/CS_data_redone.csv
/var/folders/v6/f62qz88s0sd5n3yqw9d8sb300000gn/T/RtmpRlc45x/project_miceps/Estrogen_Receptors.docx
```

Once a directory is sandboxed, users can run the `proj_render()` function, which checks all `.R` and `.Rmd` code files in the directory to ensure that they can compile properly, to ensure that their project is self-contained.

```
proj_render(temp_dir)
```

```
-- Rendering R scripts... ----- fertile 1.1.9003 --
```

If `proj_render()` executes without error, this indicates that all the necessary files are present. Users can also confirm this by checking the time stamp of the last successful render, captured in the `render_log_report()` function. If the time stamp matches the current time, the project successfully compiled.

```
slice_tail(render_log_report(temp_dir))
```

```
Reading from /var/folders/v6/f62qz88s0sd5n3yqw9d8sb300000gn/T/RtmpRlc45x/project_miceps/.f
```

```
# A tibble: 1 x 4
```

	path	path_abs	func	timestamp
	<chr>	<chr>	<chr>	<dtm>
1	LAST RENDERED	<NA>	proj_render	2021-02-19 21:57:20

2.1.2 Component 2: Organized Project Structure

fertile provides a wide variety of features for managing the file system of a project. Nine of the package's fifteen primary reproducibility checks relate to file structure.

Clear Project Root

Two of these are focused on the R `project` aspect of the file system. `has_proj_root()` ensures that there is a single `.Rproj` file indicating a clear root directory for the project, while `has_no_nested_proj_root()` ensures that there are no sub-projects within. The recognition of a clear root directory is necessary to allow for file structure analysis and project restructuring as it provides a baseline directory to define relative file paths from.

No File Clutter

Six of the major checks, whose names begin with `has_tidy_` focus on file clutter, addressing one of the components of a clean file structure. They check to make sure that no audio/video, image, source, raw data, `.rda`, or `.R` files are found in the root directory of the project.

The last check that focuses on the file system is `has_only_used_files()`. One of the more complicated checks in *fertile*, this function checks to make sure that there are no extraneous files serving no purpose (that are not “used”) included alongside the analysis. For this function to work properly, a clear definition of which files are considered as “used” is needed. In *fertile*, that definition includes the following:

- `.R`, `.Rmd`, and `.Rproj` files
- README files
- Data/text/image files whose file path is referenced, either as an input or output, in any of the `.R` or `.Rmd` files
- Outputs created by knitting any `.Rmd` files (`.html`, `.pdf`, or `.docx` files with the same name as the `.Rmd`)
- Files created by *fertile*. In an effort to capture information about dependencies, *fertile* creates a text file capturing the session information when a project is run and provides an option to generate an `.R` script that can install all of the packages referenced in a project’s code. These files are considered “used.”

```
# List the files in the directory
fs::dir_ls("project_miceps")
```

```
project_miceps/Blot_data_updated.csv    project_miceps/CS_data_redone.csv
project_miceps/Estrogen_Receptors.docx  project_miceps/README.md
project_miceps/analysis.Rmd             project_miceps/analysis.html
project_miceps/citrate_v_time.png       project_miceps/mice.csv
project_miceps/miceps.Rproj             project_miceps/proteins_v_time.png
project_miceps/software-versions.txt
```

```
# Check to see which are "used"
has_only_used_files("project_miceps")
```

```
Joining, by = "path_abs"
```

* Checking to see if all files in directory are used in code

Problem: You have files in your project directory which are not being used.

Solution: Use or delete files.

See for help: `?fs::file_delete`

A tibble: 2 x 1

path_abs

<chr>

1 /Users/audreybertin/Documents/thesis/index/project_miceps/Estrogen_Receptors.~

2 /Users/audreybertin/Documents/thesis/index/project_miceps/mice.csv

Reorganizing File Structure

There are also several functions help with reshaping the entire project structure to a more reproducible format. For example, `proj_suggest_moves()` provides suggestions for how to reorganize the files into folders.

```
files <- proj_analyze_files("project_miceps")
```

```
proj_suggest_moves(files)
```

A tibble: 9 x 8

	file	size	ext	mime	path_rel	dir_rel	path_new	cmd
	<fs::path>	<fs::by>	<chr>	<chr>	<fs::path>	<fs::path>	<fs::path>	<chr>
1	project_mic~	14.43K	csv	text/csv	Blot_data~	data-raw	project_mi~	file_m~
2	project_mic~	7.39K	csv	text/csv	CS_data_r~	data-raw	project_mi~	file_m~
3	project_mic~	10.97K	docx	applica~	Estrogen_~	inst/other	project_mi~	file_m~
4	project_mic~	5.21K	Rmd	text/x~	analysis.~	vignettes	project_mi~	file_m~
5	project_mic~	1.41M	html	text/ht~	analysis.~	inst/text	project_mi~	file_m~
6	project_mic~	187.1K	png	image/p~	citrate_v~	inst/image	project_mi~	file_m~
7	project_mic~	14.33K	csv	text/csv	mice.csv	data-raw	project_mi~	file_m~
8	project_mic~	378.74K	png	image/p~	proteins_~	inst/image	project_mi~	file_m~
9	project_mic~	5.34K	txt	text/pl~	software~	inst/text	project_mi~	file_m~

These suggestions are based on achieving the optimal file structure design for reproducibility, argued by Marwick et al. (2018) to be that of an R package (R-Core-Team (2020), Wickham (2015)), with an R folder, as well as `data`, `data-raw`, `inst`, and `vignettes`. Such a structure keeps all of the files separated and in clearly labeled directories where they are easy to find. Additionally, the extension of the R package structure to data analysis projects promotes standardization of file structure within the R community.

Users can execute these suggestions individually, using the code snippets provided next to each file name when running `proj_suggest_moves()`, but they can also

run them all together. `proj_move_files()` executes all of the suggestions from `proj_suggest_moves()` at once, building an R package style structure and sorting files into the correct folders by type.

Combined together, all of these functions make it simple to ensure that projects fall under a standard, simple, and reproducible structure with minimal clutter.

2.1.3 Component 3: Documentation

High quality documentation, including the presence of a README file, comments explaining the code, a list of software packages an analysis is dependent on, and a method in which to understand which order to run code files in, is incredibly important to reproducibility. Without written guidance, individuals looking to reproduce results may not understand how to take all of the project components and combine them in the right way to produce the desired outcome. Additionally, code used without the proper dependencies and software versions, even if it is perfectly functional and correctly written, will often result in errors when compiled.

`fertile` contains a variety of functions to ensure that projects are well documented.

README

One important component of documentation is the inclusion of a README file. A README is a text file that introduces and explains a project. Commonly, a project README contains background information necessary to understand a project. Some of the components that might be contained within are:

1. Background on the purpose of the project and the questions of interest.
2. Background on where the data used in the project came from and what they contain.
3. Information about installing and setting up the software necessary to run it.
4. A list of included files.
5. A description of how to run the project—for example, a summary of the order in which the included files should be run.
6. Contact information for the author of the project.

`fertile` ensures that a README is included alongside the user's project with the `has_readme()` function:

```
has_readme("project_miceps")
```

v Checking for README file(s) at root level

Clear File Order

Another important component of documentation is that it must be clear in which order provided files should be run. While this information can be included as part of the README, it is sometimes provided via other methods—for example, through

a `makefile/drakefile` (make-style file generated by the `Drake` package in R) or through a standardized naming or numbering convention of files.

`has_clear_build_chain()` checks for these non-README methods of ensuring that file ordering is clear. `project_miceps`, since it only contains one code file, passes this check easily:

```
has_clear_build_chain("project_miceps")
```

v Checking for clear build chain

Software Dependencies

There are also several features focused on software dependencies. Every time the code contained within a project is rendered by `fertile`, the package generates a file capturing the `sessionInfo()` just after the code was run. This file contains information about the R version in which the code was run, the list of packages that were loaded, and their specific versions.

The dependency information from `project_miceps` looks as follows:

```
-- Rendering R scripts... ----- fertile 1.1.9003 --
```

```
session_info <- "project_miceps/.software-versions.txt"
cat(readChar(session_info, 1e5))
```

The R project located at `'/Users/audreybertin/Documents/thesis/index/project_miceps'` was 1

R version 4.0.2 (2020-06-22)

Platform: x86_64-apple-darwin17.0 (64-bit)

Running under: macOS 10.16

Matrix products: default

BLAS: /Library/Frameworks/R.framework/Versions/4.0/Resources/lib/libRblas.dylib

LAPACK: /Library/Frameworks/R.framework/Versions/4.0/Resources/lib/libRlapack.dylib

locale:

en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8

attached base packages:

stats graphics grDevices utils datasets methods base

other attached packages:

stargazer_5.2.2 skimr_2.1.2 purrr_0.3.4

ggplot2_3.3.3 tidyrr_1.1.2 readr_1.4.0 dplyr_1.0.3

Interactively, users can access this dependency summary using the function `proj_dependency_report()`.

The `proj_pkg_script()` function builds off of this behavior, providing a method to simplify the process of installing the packages seen in the dependency report.

When run on an R project directory, the function creates a `.R` script file that contains the code needed to install all of the packages referenced in the project, differentiating between packages located on CRAN and those located on GitHub.

```
install_script <- proj_pkg_script("project_miceps")
cat(readChar(install_script, 1e5))

# Run this script to install the required packages for this
# R project.
# Packages hosted on CRAN...
install.packages(c( 'broom', 'dplyr', 'fs', 'ggplot2',
'purrr', 'readr', 'rmarkdown', 'skimr', 'stargazer',
'tidyr' ))
# Packages hosted on GitHub...
```

Users interested in ensuring that their project is reproducible can provide alongside it both:

1. The text-based summary of dependencies produced by `proj_dependency_report()`.
2. The `.R` dependency installation script.

Then, anyone wanting to re-create the results would have the documentation necessary to do so alongside a quick and easy method to set up the required software.

Code Commenting

In addition to README and dependency documentation, *fertile* also analyzes the quality of documentation provided directly alongside code.

Code commenting—the process of placing human-readable descriptions next to code to explain what the code is doing—is incredibly important for ensuring that a project can be understood by someone other than its author. To ensure that their code is understandable, data analysts must regularly include comments throughout their project code files.

`has_well_commented_code()` is designed to check for this. The function reads through all `.R` and `.Rmd` files in a project directory and, for each, calculates the fraction of lines in the file that are comments. Files for which comments make up less than 10% of the written lines are flagged as poorly commented, warning users to make corrections and increase the number of comments in their code.

The primary code file in `project_miceps`, “analysis.Rmd”, is a great example of a poorly commented file. It only contains 6 lines of comments, while including over 100 lines of code. As such, it is flagged by `has_well_commented_code()`:

```
has_well_commented_code("project_miceps")
```

* Checking that code is adequately commented

Problem: Suboptimally commented .R or .Rmd files found

Solution: Add more comments to the files below. At least 10% of the lines should be com

See for help: <https://intelligea.wordpress.com/2013/06/30/inline-and-block-comments-in->

A tibble: 1 x 2

file_name	fraction_lines_commented
<chr>	<dbl>
1 project_miceps/analysis.Rmd	0.04

2.1.4 Component 4: File Paths

fertile contains a variety of features designed to address issues with file paths. Several of these features happen proactively, warning users that they are entering a non-reproducible file path as it happens, while others can be accessed after a project has already been written.

Proactive Warnings When Coding

Proactively, *fertile* catches when an absolute path, or one leading outside of the project directory, is referenced in an input or output function and throws an error.

This interactive behavior is one of the educational features of the package, as it immediately provides an informative message explaining to the user that such a path is not reproducible, giving the programmer an opportunity to learn and course-correct from there.

One of the important components of R package development, however, is that a package should not interfere too much with user behavior. Users who are trying to code in their console, for example, might still want to open a file if they know the path is not reproducible if they are just trying to take a quick look at some data.

fertile accounts for this. Even though the package throws an error when non-reproducible paths are used, it also provides a solution for users to override this behavior and execute the command anyway. This is done through the error messaging system. The message provided when an error is thrown is custom created, responding to the command and path which triggered the error in the first place. For example, in the code below, `read.csv("~/Desktop/my_data.csv")`, gives a different message than `read_csv("../.../Desktop/my_data.csv")`.

```
library(fertile)
```

Reading from /Users/audreybertin/Documents/thesis/index/.fertile_log.csv

```
file.exists("~/Desktop/my_data.csv")
```

```
[1] TRUE
```

```
read.csv("~/Desktop/my_data.csv")
```

```
Reading from /Users/audreybertin/Documents/thesis/index/.fertile_log.csv
```

```
Checking for absolute paths...
```

```
Error: Detected absolute paths. Absolute paths are not reproducible and will likely
```

```
read_csv("../.../Desktop/my_data.csv")
```

```
Reading from /Users/audreybertin/Documents/thesis/index/.fertile_log.csv
```

```
Checking for absolute paths...
```

```
Checking for paths outside project directory...
```

```
Error: Detected paths that lead outside the project directory. Such paths are not re
```

In addition to catching non-reproducible file paths, *fertile* also stops the user when they try to change the working directory. Unlike with input and output functions like `read_csv()`, which can take a variety of different file paths—some which may be reproducible and some not, `setwd()` is essentially guaranteed to break reproducibility, no matter what directory it is set to. As a result, *fertile* is particularly strict with it. When a call to `setwd()` is caught, *fertile* throws an error and does not provide an option to override. Rather, the user is given an alternative function (`here::here()`) to use that achieves the same behavior in a much more reproducible way.

```
getwd()
```

```
[1] "/Users/audreybertin/Documents/thesis/index"
```

```
setwd("~/Desktop")
```

```
Error: setwd() is likely to break reproducibility. Use here::here() instead.
```

```
getwd()
```

```
[1] "/Users/audreybertin/Documents/thesis/index"
```

Project Path Summaries

fertile also contains functionality that analyzes code for path information after it is written. For example, `proj_analyze_paths` goes through all of the paths used in a project's `.R` and `.Rmd` files and produces a report detailing which ones failed to meet reproducibility standards, explaining the problem, and providing a solution.

```
proj_analyze_paths('project_miceps')
```

```
-- Generating reproducibility report... ----- fertile 1.1.9003 --
```

```
Checking for absolute paths...
```

```
Checking for paths outside project directory...
```

```
# A tibble: 2 x 5
```

	path	path_abs	func	problem	solution
	<chr>	<chr>	<chr>	<chr>	<chr>
1	/Users/audreyber~	/Users/audreybertin~	readr:~	Absolute path~	Use a relative ~
2	/Users/audreyber~	/Users/audreybertin~	readr:~	Path is not c~	Move the file a~

Paths can also be checked individually, or in groups, using `check_path()`.

```
# Path inside the directory
check_path("project_miceps")
```

```
# A tibble: 0 x 3
```

```
# ... with 3 variables: path <chr>, problem <chr>, solution <chr>
```

```
# Absolute path (current working directory)
check_path(getwd())
```

```
Error: Detected absolute paths. Absolute paths are not reproducible and will likely only w
```

```
# Path outside the directory
check_path("../README.md")
```

```
Error: Detected paths that lead outside the project directory. Such paths are not reproduc
```

These functions, together, cover all of the reproducibility subcomponents involving file paths. Users are warned not to use absolute paths or paths pointing outside of the directory, both while they are coding and after the fact, and provided with a recommended solution to the problem if they do so. Additionally, users are prevented from using functions that will certainly break reproducibility and provided with safer alternatives.

2.1.5 Component 5: Randomness

The component of randomness is addressed by `fertile` in a reproducibility check function titled `has_no_randomness()`, which does the following: First, it checks code files to determine whether they have employed random number generation. This is confirmed by recording the random number generation seed prior to running the files, rendering the files, and then checking to see whether the seed has changed. If the seed has changed, that indicates that random number generation has occurred. Then, it determines whether any randomness present was semi-random (i.e. controlled and repeatable) or truly random, by checking whether the `set.seed()` function was called. If the randomness was semi-random, it is considered permissible and reproducible. Truly random code is flagged as dangerous.

Let us consider an example from `project_miceps`. The primary code file, `analysis.Rmd` contains the following code. In this code, we see that a data set is read in via `read_csv` and reformatted. Then, the `sample_frac()` function takes a random sample of 50% of the rows of the data.

```
mice <- read_csv("Blot_data_updated.csv") %>%
  transmute(time = `Time point`,
            sex = Sex,
            id = `Mouse #`,
            erb = `ERB Normalized Final`,
            era = `ERA Normalized Final`,
            gper = `GPER Normalized Final`) %>%
  mutate(day = case_when(
    time == 0 ~ -1,
    time == 1 ~ 0,
    time == 2 ~ 1,
    time == 3 ~ 3,
    time == 4 ~ 5,
    time == 5 ~ 7
  ),
  type = ifelse(time == 0, "control", "treatment")
)
mice_tidy <- mice %>%
  select(-time) %>%
  gather(key = "protein", value = "amount", -day, -sex, -id, -type) %>%
  mutate(protein_label = factor(protein,
    labels = c("paste(ER, alpha)", "paste(ER, beta)", "GPER")))

sample_frac(mice, 0.5)
```

However, there is no seed set in the file. As a result, `has_no_randomness()` returns as a failure for the project. Had a seed been set, however, the project would have passed.

```
has_no_randomness('project_miceps')
```

* Checking for no randomness

Problem: Your code uses randomness

Solution: Set a seed using `set.seed()` to ensure reproducibility.

See for help: `?set.seed`

```
# A tibble: 1 x 2
  culprit expr
  <chr>    <glue>
1 ?      Example: set.seed(1)
```

fertile's randomness-centered feature helps analysts know that their use of randomness is controlled, ensuring that functions involving random number generation will always produce the same output each time they are run.

2.1.6 Component 6: Readability and Style

Though not an absolutely necessary component of reproducibility, code style can have a significant impact on how easy it is to follow the steps being taken in an analysis. The use of consistent and highly-readable style in code greatly simplifies and speeds up the process of understanding a data analysis project and repeating the steps included within.

fertile addresses code style via the function `has_no_lint()`. This function checks code for compatibility with RStudio Chief Scientist Hadley Wickham's "tidy" code format, which promotes the following best practices:

- Line length should not exceed 80 characters
- There should not be trailing whitespace
- All infix operators (`=`, `+`, `-`, `<-`, etc) should have spaces on either side
- All commas should be followed by spaces
- There is no code left in the file that is commented out
- `<-` should be used instead of `=` to assign variables
- Opening curly braces should never go on their own line and should always be followed by a new line
- There should always be a space between right parenthesis and an open curly brace
- Closing curly braces should always be on their own line, unless followed by an `else` statement

Any issues with incompatibility that are found by `has_no_lint()` appear in a window by the RStudio console. This window lists the style inconsistencies found in each code document, showing both the description of the error and the line number where it

occurred for each issue. This window is interactive; double-clicking on an error brings the user to exact location in the code where it occurred.

Let us consider an example. `project_miceps` contains an RMarkdown file titled `analysis.Rmd`, which contains some code involved with a data analysis. Some of the code is in tidy style but not all of it is.

For example, line 71, part of a `ggplot` call, contains the following code:

```
# Line 71 of `project_miceps/analysis.Rmd`,
# Test has been wrapped so it can all be seen at once

geom_hline(aes(yintercept = estimate + 1.96*std.error, color = sex),
linetype = 3) +
```

For this line, `has_no_lint()` finds the following inconsistencies with tidy style:

- Line 71: Lines should not be more than 80 characters.
- Line 71: Put spaces around all infix operators.
- Line 71: Trailing whitespace is superfluous.

The first comes up because the true line length, including spaces, is 84 characters. The second comes up because the `*` between `1.96` and `std.error` is not surrounded by spaces. The third is flagged because there is an empty space after the `+` at the end of the line.

An even less tidy piece of code is that found on line 189, written as follows:

```
# Line 189 of `project_miceps/analysis.Rmd`

if (length(mice) > 1){ holder_var = 1 }
```

For this line, `has_no_lint()` finds the following errors:

- Line 189: There should be a space between right parenthesis and an opening curly brace.
- Line 189: Opening curly braces should never go on their own line and should always be followed by a new line.
- Line 189: Use `<-`, not `=`, for assignment.
- Line 189: Closing curly braces should always be on their own line, unless it's followed by an else.

The first issue flagged is the fact that `)``{`, in the middle of the line, has no space in the middle. The second issue is flagged because the code to execute when the `if` statement is true (`holder_var = 1`) is on the same line as the `if` statement, rather than on its own line. The third error occurs because `holder_var` is defined using an

=. And the fourth occurs because the closing curly brace is on the same line as other code rather than by itself.

These informative error messages provided by *fertile* help teach users to use consistent, tidy style in their code. They do so while making the learning process incredibly simple, allowing users to click and see exactly which symbol or character caused the error. In addition to providing a detailed explanation of the was in which the provided code is not tidy, *fertile* also suggests a simple and fast solution for resolving the inconsistencies all at once, recommending that users apply the `usethis::use_tidy_style()` function to format their code automatically.

2.1.7 Summary of Reproducibility Components

As described in the previous sections, *fertile* addresses all six major components of reproducibility via a variety of functions. Some complex components have many associated functions attached to them, while others only have one.

A summary of the six components of reproducibility and the functionalities in *fertile* associated with them is provided below.

Component	Associated Tools in <i>fertile</i>
Accessible project files	<i>proj_analyze_files()</i> : returns a report of project files (including size, type, path) <i>sandbox()</i> + <i>proj_render()</i> : test that files are self-contained
Organized project structure	<i>has_proj_root()</i> + <i>has_no_nested_proj_root()</i> : check for clear project structure <i>has_tidy_xxx()</i> : series of functions checking for file clutter of different types <i>has_only_used_files()</i> : checks that there are no excess/unused files <i>proj_suggest_moves()</i> + <i>proj_move_files()</i> : restructure the file system
Documentation	<i>has_readme()</i> : checks for the existence of a README file <i>has_clear_build_chain()</i> : checks to make sure code files are clearly ordered <i>proj_dependency_report()</i> : returns info on project software dependencies <i>proj_pkg_script()</i> : generates an install script for dependencies <i>has_well_commented_code()</i> : checks to see that code is adequately commented
File paths	<i>shimmed functions</i> (read_csv/setwd/etc) throw errors with bad paths <i>proj_analyze_paths()</i> : checks that all paths in a project are reproducible <i>check_path()</i> : checks a single path for reproducibility
Randomness	<i>has_no_randomness()</i> : checks to make sure any randomness is controlled
Readability and style	<i>has_no_lint()</i> : ensures code follows tidy style guidelines

Figure 2.3: Summary of Reproducibility Components and the Related Functionalities in 'fertile'

2.1.8 User Customizability

fertile does not force its users into a box, instead allowing for a great deal of user choice in terms of which aspects of reproducibility to focus on. Users can run reproducibility tests at three different scales:

- 1) Comprehensively, where all checks are run within a single function or two.
- 2) In groups, where functions focused on similar aspects of reproducibility are run together.
- 3) Individually, where only one reproducibility check is run at a time.

Comprehensive Features

Users who want comprehensive behavior can access it through three primary functions: `proj_check()`, `proj_analyze()`, and `proj_badges()`.

The `proj_check()` function runs all sixteen reproducibility tests in the package, noting which ones passed, which ones failed, the reason for failure, a recommended solution, and a guide to where to look for help. These tests, many of which were described in detail previously, include: looking for a clear build chain, checking to make sure the root level of the project is clear of clutter, confirming that there are no files present that are not being directly used by or created by the code, and looking for uses of randomness that do not have a call to `set.seed()` present. A full list is provided below:

```
list_checks()
```

```
-- The available checks in `fertile` are as follows: ----- fertile 1.1.9003 --

[1] "has_tidy_media"           "has_tidy_images"
[3] "has_tidy_code"           "has_tidy_raw_data"
[5] "has_tidy_data"           "has_tidy_scripts"
[7] "has_readme"              "has_no_lint"
[9] "has_proj_root"           "has_no_nested_proj_root"
[11] "has_only_used_files"     "has_clear_build_chain"
[13] "has_no_absolute_paths"   "has_only_portable_paths"
[15] "has_no_randomness"       "has_well_commented_code"
```

The `proj_analyze()` function creates a report documenting the structure of a data analysis project, combining four key functions from `fertile` into one complete unit:

- `proj_analyze_packages()`, which captures a list of all packages referenced in the source code and which files they were referenced in
- `proj_analyze_files()`, which captures a list of all of the files located in the directory along with their type and size
- `proj_suggest_moves()`, which provides suggestions for how to reorganize files to create a more reproducible file structure
- `proj_analyze_paths()`, which captures a list of the non-reproducible file paths referenced in source code

```
proj_analyze('project_miceps')
```

```
-- Analysis of reproducibility for project_miceps ----- fertile 1.1.9003 --

-- Packages referenced in source code ----- fertile 1.1.9003 --

# A tibble: 10 x 3
  package      N used_in
  <chr>      <int> <chr>
1 broom          1 project_miceps/analysis.Rmd
2 dplyr          1 project_miceps/analysis.Rmd
3 fs             1 project_miceps/analysis.Rmd
4 ggplot2        1 project_miceps/analysis.Rmd
5 purrr          1 project_miceps/analysis.Rmd
6 readr          1 project_miceps/analysis.Rmd
7 rmarkdown      1 project_miceps/analysis.Rmd
8 skimr          1 project_miceps/analysis.Rmd
9 stargazer       1 project_miceps/analysis.Rmd
10 tidyr          1 project_miceps/analysis.Rmd

-- Files present in directory ----- fertile 1.1.9003 --

# A tibble: 11 x 4
  file                ext      size mime
  <fs::path>         <chr> <fs::byt> <chr>
1 Estrogen_Receptor~ docx    10.97K application/vnd.openxmlformats-officedocu~
2 citrate_v_time.png png     188.04K image/png
3 proteins_v_time.p~ png     378.25K image/png
4 Blot_data_updated~ csv      14.43K text/csv
5 CS_data_redone.csv csv       7.39K text/csv
6 mice.csv          csv     14.33K text/csv
7 analysis.html     html     1.41M text/html
8 README.md         md        39 text/markdown
9 software-versions~ txt      5.34K text/plain
10 miceps.Rproj      Rproj     204 text/rstudio
11 analysis.Rmd      Rmd      5.21K text/x-markdown

-- Suggestions for moving files ----- fertile 1.1.9003 --

# A tibble: 9 x 3
  path_rel      dir_rel  cmd
  <fs::path>    <fs::path> <chr>
1 Blot_data_updated~ data-raw  file_move('project_miceps/Blot_data_updated.csv~
2 CS_data_redone.csv data-raw  file_move('project_miceps/CS_data_redone.csv', ~
3 Estrogen_Receptor~ inst/other file_move('project_miceps/Estrogen_Receptors.do~
```

```

4 analysis.Rmd      vignettes  file_move('project_miceps/analysis.Rmd', fs::di~
5 analysis.html     inst/text   file_move('project_miceps/analysis.html', fs::d~
6 citrate_v_time.png inst/image  file_move('project_miceps/citrate_v_time.png', ~
7 mice.csv          data-raw    file_move('project_miceps/mice.csv', fs::dir_cr~
8 proteins_v_time.p~ inst/image  file_move('project_miceps/proteins_v_time.png',~
9 software-versions~ inst/text   file_move('project_miceps/software-versions.txt~

-- Problematic paths logged ----- fertile 1.1.9003 --

NULL

```

Similar to `proj_check()`, `proj_badges()` runs all sixteen reproducibility checks—`has_tidy_media`, `has_readme`, `has_no_absolute_paths`, etc. However, the way the function displays the resulting information is different. Rather than report the results of each check individually alongside detailed information about why each failed, `proj_badges()` provides a higher level summary, reporting on the *groups* of checks that either passed or failed.

In `proj_badges()`, the reproducibility checks are grouped into six categories corresponding to the six major components of reproducibility. These categories, alongside the checks which belong to them, are listed below:

- Project Structure
 - `has_proj_root()`
 - `has_no_nested_proj_root()`
- Tidy Files
 - `has_tidy_media()`
 - `has_tidy_images()`
 - `has_tidy_code()`
 - `has_tidy_raw_data()`
 - `has_tidy_data()`
 - `has_tidy_scripts()`
 - `has_only_used_files()`
- Documentation
 - `has_readme()`
 - `has_clear_build_chain()`
 - `has_well_commented_code()`
- File Paths
 - `has_no_absolute_paths()`
 - `has_only_portable_paths()`
- Randomness
 - `has_no_randomness()`
- Code Style

```
– has_no_lint()
```

When users pass all of the checks associated with one of the categories, they receive a badge certifying their success in that area. If any fewer than *all* available checks in a category are passed, then the user fails that category.

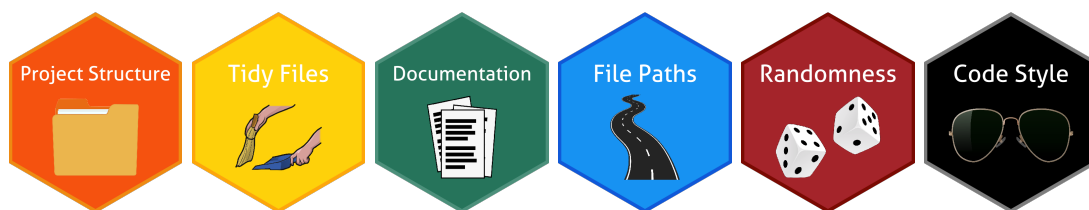


Figure 2.4: The 6 Reproducibility Badges Available in ‘fertile’

`proj_badges()` identifies the badges that a data analysis project should receive and then outputs a summary of this information—in the form of an `html` document—to the RStudio **Viewer** pane. This document—designed to produce a summary of a project that can be easily shared in places outside of RStudio (unlike the summaries produced by most other functions, which just produce output in the R console)—contains the following primary pieces of information:

1. The name of the folder where the project is stored.
2. The reproducibility badges awarded to the project.
3. The reproducibility badges failed by the project.
4. If failures occurred, the reasons why. These are broken down by badge and list the checks associated with that badge that did not pass.
5. A date/time/timezone stamp of when the `html` summary was created.
6. Information about the user who generated the `html` summary (their full name, computer username, email, and GitHub username, if available).
7. Information about the R version, platform, and operating system of the computer used to generate the `html` at the time that it was created.
8. The names of files in the project that the `html` found the badges for as well as their date of last modification.

The `html` output for `project_miceps` can be seen below:



Figure 2.5: Output: Badges and Reasons for Failure

Code Style

```
## # A tibble: 1 x 1
##   check_name
##   <chr>
## 1 has_no_lint
```

Output Generation Details:

This project summary was generated on 2020-10-29 at 14:00:51 (America/New_York) by a user with the following information:

- Full name: Audrey Bertin
- Username: audreybertin
- Email: N/A
- GitHub Username: N/A

The computer used to generate this file was running R version 4.0.2 (2020-06-22) on the x86_64-apple-darwin17.0 (64-bit) platform and the macOS Catalina 10.15.5 operating system.

The files analyzed in the creation of this summary, as well as their last-modified timestamp, are provided below:

```
## # A tibble: 9 x 2
##   file_name      last_modified
##   <chr>         <dtm>
## 1 Blot_data_updated.csv 2020-10-12 14:25:17
## 2 CS_data_redone.csv    2020-10-12 14:25:22
## 3 Estrogen_Receptors.docx 2020-10-12 14:25:27
## 4 README.md            2019-01-25 14:19:39
## 5 analysis.Rmd         2020-10-12 14:25:32
## 6 citrate_v_time.png    2020-10-29 14:00:40
## 7 mice.csv             2020-10-12 14:29:31
## 8 miceps.Rproj          2019-01-25 14:19:39
## 9 proteins_v_time.png   2020-10-29 14:00:39
```

Figure 2.6: Output: System/User Information and File List

Together, `proj_check()`, `proj_analyze()`, and `proj_badges()` cover a significant portion of all six major components of reproducibility.

Features Allowing For Summary By Group

Users wanting to focus on groups of checks can do so using the `proj_check_some()` function. `proj_check_some()` leverages helper functions from `tidyselect` (Henry & Wickham (2020)) to allow users to call groups of similar checks together. `tidyselect` helpers tell *fertile* to call only the checks whose names meet certain conditions. The helper functions that can be passed to `proj_check_some()` are:

- `starts_with()`: Starts with a prefix.
- `ends_with()`: Ends with a suffix.
- `contains()`: Contains a literal string.
- `matches()`: Matches a regular expression.

For example, a variety of checks in *fertile* focus on making sure the project has a “tidy” structure—essentially that there are not files cluttered together all in one folder. Users interested in checking their tidyness can do so all at once using `proj_check_some()` as follows:

```
proj_check_some("project_miceps", contains("tidy"))
```

```
-- Running reproducibility checks ----- fertile 1.1.9003 --
```



```

v Checking for no *.R scripts at root level

v Checking for no *.rda files at root level

v Checking for no A/V files at root level

* Checking for no image files at root level

  Problem: Image files in root directory clutter project

  Solution: Move source files to img/ directory

  See for help: ?fs::file_move

# A tibble: 2 x 2
  culprit          expr
  <fs::path>      <glue>
1 project_miceps/citrate_v_t~ fs::file_move('project_miceps/citrate_v_time.png'~
2 project_miceps/proteins_v_~ fs::file_move('project_miceps/proteins_v_time.png~

* Checking for no raw data files at root level

  Problem: Raw data files in root directory clutter project

  Solution: Move raw data files to data-raw/ directory

  See for help: ?fs::file_move

# A tibble: 3 x 2
  culprit          expr
  <fs::path>      <glue>
1 project_miceps/Blot_data_u~ fs::file_move('project_miceps/Blot_data_updated.c~
2 project_miceps/CS_data_red~ fs::file_move('project_miceps/CS_data_redone.csv'~
3 project_miceps/mice.csv    fs::file_move('project_miceps/mice.csv', here::he~

v Checking for no source files at root level

-- Summary of fertile checks ----- fertile 1.1.9003 --

v Reproducibility checks passed: 4

* Reproducibility checks to work on: 2

* Checking for no image files at root level

  Problem: Image files in root directory clutter project

  Solution: Move source files to img/ directory

```

See for help: `?fs::file_move`

```
# A tibble: 2 x 2
  culprit          expr
  <fs::path>      <glue>
1 project_miceps/citrate_v_t~ fs::file_move('project_miceps/citrate_v_time.png'~
2 project_miceps/proteins_v_~ fs::file_move('project_miceps/proteins_v_time.png'~

* Checking for no raw data files at root level
```

Problem: Raw data files in root directory clutter project

Solution: Move raw data files to data-raw/ directory

See for help: `?fs::file_move`

```
# A tibble: 3 x 2
  culprit          expr
  <fs::path>      <glue>
1 project_miceps/Blot_data_u~ fs::file_move('project_miceps/Blot_data_updated.c~
2 project_miceps/CS_data_red~ fs::file_move('project_miceps/CS_data_redone.csv'~
3 project_miceps/mice.csv     fs::file_move('project_miceps/mice.csv', here::he~
```

Users might also attempt to call the two checks involving project roots together:

```
proj_check_some("project_miceps", ends_with("root"))
```

```
-- Running reproducibility checks ----- fertile 1.1.9003 --
v Checking for nested .Rproj files within project
v Checking for single .Rproj file at root level
-- Summary of fertile checks ----- fertile 1.1.9003 --
v Reproducibility checks passed: 2
```

Or, perhaps, they might want to run all the functions beginning with “has_only”:

```
proj_check_some("project_miceps", starts_with("has_only"))
```

```
-- Compiling... ----- fertile 1.1.9003 --
-- Rendering R scripts... ----- fertile 1.1.9003 --
-- Running reproducibility checks ----- fertile 1.1.9003 --
```

```

Joining, by = "path_abs"

* Checking for only portable paths

  Problem: Non-portable paths won't necessarily work for others

  Solution: Use relative paths.

  See for help: ?fs::path_rel

# A tibble: 1 x 2
  culprit                                expr
  <fs::path>                            <glue>
1 /Users/audreybertin/Documents/thesis/in~ fs::path_rel('/Users/audreybertin/Do~

* Checking to see if all files in directory are used in code

  Problem: You have files in your project directory which are not being used.

  Solution: Use or delete files.

  See for help: ?fs::file_delete

# A tibble: 2 x 1
  path_abs
  <chr>
1 /Users/audreybertin/Documents/thesis/index/project_miceps/Estrogen_Receptors.~
2 /Users/audreybertin/Documents/thesis/index/project_miceps/mice.csv

-- Summary of fertile checks ----- fertile 1.1.9003 --

v Reproducibility checks passed: 0

* Reproducibility checks to work on: 2

* Checking for only portable paths

  Problem: Non-portable paths won't necessarily work for others

  Solution: Use relative paths.

  See for help: ?fs::path_rel

# A tibble: 1 x 2
  culprit                                expr
  <fs::path>                            <glue>
1 /Users/audreybertin/Documents/thesis/in~ fs::path_rel('/Users/audreybertin/Do~

```

* Checking to see if all files in directory are used in code

Problem: You have files in your project directory which are not being used.

Solution: Use or delete files.

See for help: `?fs::file_delete`

```
# A tibble: 2 x 1
```

```
  path_abs
```

```
  <chr>
```

```
1 /Users/audreybertin/Documents/thesis/index/project_miceps/Estrogen_Receptors.~
```

```
2 /Users/audreybertin/Documents/thesis/index/project_miceps/mice.csv
```

Individual Checks

If users do not want to run functions in groups, and prefer to run them individually, that option is also provided to them. Every check that makes up `check()` and every subcomponent of `project_analyze()` can be run individually.

2.1.9 Educational Features

Simply noting and correcting issues with reproducibility is not enough to produce lasting change in the scientific community. Data analysts and software users must also be educated on why their choices were not reproducible so that they do not fall victim to those mistakes again in the future, but also so that they can share their knowledge and experience with others in the scientific community.

fertile prioritizes this idea of reproducibility education throughout many of its functionalities.

One of the major ways through which *fertile* educates its users is a system of command tracking and interactive messaging. As long as *fertile* is loaded in *R*, the package records when commands that have the potential to affect reproducibility are run in the console.

As soon as a dangerous function is called, *fertile* alerts the user of their mistake and provides suggestions for alternative solutions. This behavior, explained in more detail in Chapter 2.2.4, gives users immediate feedback on their behavior. In addition to assisting users in the moment, this method has also been shown to increase long-run retention of information when compared with feedback after the fact (Epstein et al. (2002)).

Users interested in looking back at their past choices and mistakes can do so as well. The `log_report()` function provides access to a log listing the commands with a link to reproducibility that have been called.

```
log_report()
```

Reading from `/Users/audreybertin/Documents/thesis/index/.fertile_log.csv`

```
# A tibble: 735 x 4
  path          path_abs          func      timestamp
  <chr>         <chr>          <chr>      <dtm>
1 package:remotes <NA>          base::re~ 2020-09-20 15:20:34
2 package:thesis~ <NA>          base::re~ 2020-09-20 15:20:34
3 package:thesis~ <NA>          base::li~ 2020-09-20 15:20:34
4 package:stringr <NA>          base::li~ 2020-09-21 13:28:29
5 package:fertile <NA>          base::li~ 2020-09-21 13:28:29
6 ~/Desktop/my_d~ ~/Desktop/my_data.csv  utils::r~ 2020-09-21 13:28:29
7 ../../../../Deskt~ /Users/audreybertin/Desktop/my~ utils::r~ 2020-09-21 13:28:29
8 package:purrr <NA>          base::li~ 2020-09-21 13:28:36
9 package:forcats <NA>          base::li~ 2020-09-21 13:28:36
10 project_miceps~ /Users/audreybertin/Documents/~ readr::r~ 2020-09-21 13:28:36
# ... with 725 more rows
```

Depending on how much history they want to keep track of, users have the option to clear the log and start from scratch via the function `log_clear()`.

In addition to this educational logging behavior, the reproducibility checks contained in the `check()` function also include educational messages. We consider several examples below:

- `has_no_randomness()`, when it fails, tells users that their code uses randomness and they should use the `set.seed()` function to control it.
- `has_only_used_files()`, when it fails, tells users that there are files present in the directory not being used for any purpose and provides a function (`fs::file_delete`) to use in order to remove them.
- `has_no_lint()`, when it fails, provides users a list of all of the ways in which their code fails to follow tidy style and points them to the exact lines and characters in the code where the mistakes occurred.
- `has_tidy_images`, when it fails, tells users that it has found files in the root directory which add clutter and recommends that they be moved to an `img/` directory.

Users are provided with an informative message about their issue but are not always provided a fully automated solution. This behavior encourages them to learn as they have to execute the suggested solution themselves.

However, even though **fertile** often requires users to take their own actions, that does not mean that the package requires users to be of high skill level to use. **fertile** is designed in such a way so its educational benefits can be achieved with relative simplicity and minimal effort so that even users brand new to R can gain knowledge and awareness of reproducibility from using it.

For example, the interactive messaging features require no effort beyond the loading of the package with `library(fertile)` to activate. And many of the educational benefits contained within the checks can be gained all at once as well, with a single

call of the `check()` function. Users looking for more customizability have the option to go into more detail with the more complex functions like `sandbox()`, but they are not necessary for users to gain benefit from the package.

2.2 How *fertile* Works

In the world of R packages, *fertile*'s behavior is rather unusual. Almost no other packages manipulate their users' R environments or file structure, instead remaining relatively self-contained. This unique behavior necessitates the use of several non-traditional techniques, described below.

2.2.1 Shims

Much of the functionality in *fertile* is achieved by writing *shims*. In their application to *fertile*, shims can be defined as functions that transparently intercept users' intended actions and slightly alter their execution.

fertile contains shims for a variety of common functions that may affect reproducibility, including those that read and write files, load libraries, and set random number generation seeds. A full list of shimmed functions, organized by their original package, is provided below:

Package	Function(s)
dplyr	tbl
readr	read_csv, read_csv2, read_delim, read_file, read_file_raw, read_fwf, read_lines, read_lines_raw, read_log, read_table, read_table2, read_tsv, write_csv
ggplot2	ggsave
stats	read.ftable
utils	read.csv, read.csv2, read.delim, read.delim2, read.DIF, read.fortran, read.fwf, read.table, write.csv
base	library, load, read.dcf, require, save, set.seed, setwd, source
readxl	read_excel
rjson	fromJSON
foreign	read.dta, read.mtp, read.spss, read.systat
sas7bdat	read.sas7bdat

Figure 2.7: List of Functions Shimmed by 'fertile'

When users perform actions that may threaten reproducibility, the package's shimmed functions intercept the user's commands and perform various logging and

checking tasks before executing the desired function.

This allows **fertile** to warn users when they make mistakes and also to keep track of past behavior via a log of previously entered commands.

The process for writing a shim is as follows:

1. Identify an R function that is likely to be involved in operations that may break reproducibility. Popular functions associated with only one package (e.g., `read_csv()` from **readr**) are ideal candidates.
2. Create a function in **fertile** with the same name that takes the same arguments (and always the dots `...`).
3. Write this new function so that it:
 - a) captures any arguments,
 - b) logs the name of the function called,
 - c) performs any checks on these arguments, and
 - d) calls the original function with the original arguments. Except where warranted, the execution looks the same to the user as if they were calling the original function.

Most shims, when written, are relatively simple. Several, such as that for `library()` are more complex, but many follow the same basic format that can be seen in this example for `read_csv()`:

```
fertile::read_csv

function(file, ...) {
  if (interactive_log_on()) {
    if (Sys.getenv("FERTILE_RENDER_MODE") == TRUE) {
      log_push("Seed Before", .Random.seed[2])
    }

    log_push(file, "readr::read_csv")

    check_path_safe(file, ... = "readr::read_csv")

    data <- readr::read_csv(file, ...)

    if (Sys.getenv("FERTILE_RENDER_MODE") == TRUE) {
      log_push("Seed After", .Random.seed[2])
    }

    return(data)
  }
}
<bytecode: 0x7f824046d608>
<environment: namespace:fertile>
```

This functionality all occurs without the knowledge of the user. Consider the example of `read_csv()`. `read_csv()` is a very popular function from the `readr` package for reading in data files. Users with both `readr` and `fertile` loaded, will experience the following. The user will call `read_csv()` as normal, thinking that they are calling `readr::read_csv()`. However, they will actually be calling `fertile::read_csv()`, a very similar function with the same name. `fertile::read_csv()` will then capture the file path the user provided and check whether it is reproducible. If it is deemed okay, the function will execute as intended and read in the data just as `readr::read_csv()` would. If it is deemed non-reproducible, the function will return an error telling the user to use an alternate file path. Either way, `fertile` will record that the user called `read_csv()` and note the path that was provided to it for future reference.

This behavior, however, is dependent `fertile` remaining at the top of the `search()` path so that its functions are called preferentially over the original functions that it has shimmed. In order to ensure that the `fertile` versions of functions (“shims”) always supersede (“mask”) their original namesakes when called, `fertile` uses its own shims of the `library` and `require` functions to manipulate the R search path so that it is always located in the first position.

In the `fertile` version of `library()`, `fertile` is detached from the search path, the requested package is loaded, and then `fertile` is reattached. This ensures that when a user executes a command, R will check `fertile` for a matching function before considering other packages.

2.2.2 Hidden Files

In order to store and analyze information about user behavior and code structure, `fertile` utilizes three different types of hidden files, two of which are `.csv` format and one of which is a text file. The hidden `.csv` files for `project_miceps` can be seen below:

```
fs::dir_ls("project_miceps", all = TRUE, glob = "*_log.csv")
```

project_miceps/.fertile_log.csv

project_miceps/.fertile_render_log.csv

The interactive log (`.fertile_log.csv`), accessible via `log_report()`, is created as soon as a user executes their first piece of code that could threaten reproducibility. This file tracks all of the shimmed functions executed by the user, either in the console or when running code chunks by hand (rather than knitting a file). It reports the function called, the relevant argument passed in (either a file path or R package name), and the time stamp of when the function was executed. Users can clear the data from this file and start fresh at any time with `log_clear()`.

The render log (`.fertile_render_log.csv`), accessible via `render_log_report()`, has a similar structure to the interactive log but is not under the control of the user. It tracks information about the code contained within `.R` and `.Rmd` files within the project a user is testing for reproducibility. A new render log file can be generated in one of three different ways:

1. The first time a user runs one of the major reproducibility checks from **fertile**, such as `proj_check()`, `proj_analyze()`, or one of the smaller checks within that requires access to the contents of code files.
2. Any time a check involving code is called in **fertile**, the package checks to see whether any code files have been updated since the last time a render log was generated. If so, a new render log is generated.
3. The user can generate a new file manually at any time by executing the `proj_render()` command.

The render log contains information used to run many of the checks in **fertile**. It captures the random number generator seed before and after executing code, notes which packages and files are accessed and the function with which they were called, and contains a timestamp of the last time the code was run by **fertile**. Users cannot erase it easily.

```
render_log_report("project_miceps")
```

Reading from `/Users/audreybertin/Documents/thesis/index/project_miceps/.fertile_rend`

A tibble: 21 x 4

	path <chr>	path_abs <chr>	func <chr>	timestamp <dtm>
1	Seed @ Start	<NA>	599	2021-02-19 21:57:39
2	package:dplyr	<NA>	base::~	2021-02-19 21:57:39
3	package:readr	<NA>	base::~	2021-02-19 21:57:39
4	package:tidyr	<NA>	base::~	2021-02-19 21:57:39
5	package:ggplot2	<NA>	base::~	2021-02-19 21:57:39
6	package:purrr	<NA>	base::~	2021-02-19 21:57:39
7	Seed Before	<NA>	599	2021-02-19 21:57:39
8	/Users/audreybertin/Do~	/Users/audreybertin/Docu~	readr::~	2021-02-19 21:57:40
9	Seed After	<NA>	613	2021-02-19 21:57:40
10	Seed Before	<NA>	613	2021-02-19 21:57:40

... with 11 more rows

These files, integral to many of the functionalities in **fertile**, are not visible when looking at the file system. Users can only access them with functions provided by **fertile**, and user permissions are quite restrictive. Except for removing the command history in the interactive log, users cannot modify their contents, and neither the render log nor the interactive log can be deleted without the user modifying their file system. This prevents the files from mistakenly being tampered with, potentially impairing their functionality, and ensures that **fertile** always retains accurate information of user behavior.

In addition to the two log files, **fertile** keeps a third hidden file to track project dependencies. Described in detail in the section on documentation, this file keeps

track of the software setup that a project is run under. A new version is generated every time **fertile** compiles the project code files or when users request to view the file (via a special access function). Like the log files, however, there is no simple way to manually modify or delete the file, ensuring that it does not accidentally get changed in a way that would damage reproducibility.

2.2.3 Environment Variables

The shims in **fertile** are designed to be able to write to both the interactive and render logs. That way, no matter whether a user calls `read_csv()` interactively or writes it in their code file, **fertile** will still take note of the fact that the action has happened.

Given the different purposes of each file, however, it is important that **fertile** be able to identify when a function execution should be saved to the interactive log versus when it should be saved to the render log.

This information is tracked via a logical environment variable: `FERTILE_RENDER_MODE`. When `FERTILE_RENDER_MODE` is `TRUE`, executed shims are saved to the render log. When it is `FALSE`, they are saved to the interactive log.

Since **fertile** is designed to always be capturing information about interactive user behavior, `FERTILE_RENDER_MODE` is `FALSE` by default.

It is only changed to `TRUE` when **fertile** is executing functions that involve the rendering of `.R` and `.Rmd` code files. At the start of all such functions, **fertile** sets the environment variable to `TRUE`, executes the majority of the function, and then sets it back to `FALSE` before exiting. This ensures that as soon as the function has finished running, all new commands get executed on the interactive log, rather on the render log which was just generated.

The example of `has_only_portable_paths()` illustrates this functionality. We see several clear steps to this function:

1. The environment variable is set to `TRUE`, so that **fertile** knows to write any information about shimmed functions to the render log.
2. If a render log does not yet exist or the project has been updated since the last time one was generated, then the project is rendered with `proj_render()`, generating a new render log.
3. This render log is read to find information about the file paths that were captured when executing the code files.
4. **fertile** checks to see if the paths are portable and outputs a list of the ones that are not, if any, in addition to some information about how to correct that.
5. The environment variable is set back to `FALSE` so that interactive behavior is once again captured.

```

has_only_portable_paths

function(path = ".") {
  Sys.setenv("FERTILE_RENDER_MODE" = TRUE)

  check_is_dir(path)

  if (!has_rendered(path)) {
    proj_render(path)
  }

  paths <- log_report(path) %>%
    dplyr::filter(!grepl("package:", path)) %>%
    dplyr::pull(path)

  good <- paths %>%
    is_path_portable()

  errors <- tibble(
    culprit = as_fs_path(paths[!good]),
    expr = glue("fs::path_rel('{culprit}')" )
  )

  Sys.setenv("FERTILE_RENDER_MODE" = FALSE)

  make_check(
    name = "Checking for only portable paths",
    state = all(good),
    problem = "Non-portable paths won't necessarily work for others",
    solution = "Use relative paths.",
    help = "?fs::path_rel",
    errors = errors
  )
}
<bytecode: 0x7f824688b5b8>
<environment: namespace:fertile>
attr("req_compilation")
[1] TRUE

```

2.2.4 The Dots (...)

fertile utilizes the dots (...), which allow a function to accept additional arguments beyond those pre-defined in the function, to facilitate much of its behavior. The two primary locations the dots are used are in shims, the file path messaging system, and the `proj_check_some()` function.

Many of the shimmed functions in *fertile* accept a large number of arguments. Consider `read_csv`. The function requires only one argument (`file`) but allows for up to 14.

```
readr::read_csv
```

```
function (file, col_names = TRUE, col_types = NULL, locale = default_locale(),
  na = c("", "NA"), quoted_na = TRUE, quote = "\"", comment = "",
  trim_ws = TRUE, skip = 0, n_max = Inf, guess_max = min(1000,
    n_max), progress = show_progress(), skip_empty_rows = TRUE)
{
  tokenizer <- tokenizer_csv(na = na, quoted_na = quoted_na,
    quote = quote, comment = comment, trim_ws = trim_ws,
    skip_empty_rows = skip_empty_rows)
  read_delimited(file, tokenizer, col_names = col_names, col_types = col_types,
    locale = locale, skip = skip, skip_empty_rows = skip_empty_rows,
    comment = comment, n_max = n_max, guess_max = guess_max,
    progress = progress)
}
<bytecode: 0x7f823cd95098>
<environment: namespace:readr>
```

When users call the shimmed version of `read_csv`, *fertile* does not need to process any arguments other than `file`, since that is the only piece of information directly relevant to reproducibility. Instead of defining all of the arguments once again, *fertile*'s `read_csv` is written to accept a file name and the dots, which then capture and save the additional input provided by the user. *fertile* checks the file path, and then uses the saved dots to then execute `readr::read_csv` the way that the user originally intended.

```
fertile::read_csv
```

```
function(file, ...) {
  if (interactive_log_on()) {
    if (Sys.getenv("FERTILE_RENDER_MODE") == TRUE) {
      log_push("Seed Before", .Random.seed[2])
    }

    log_push(file, "readr::read_csv")

    check_path_safe(file, ... = "readr::read_csv")

    data <- readr::read_csv(file, ...)

    if (Sys.getenv("FERTILE_RENDER_MODE") == TRUE) {
```

```

    log_push("Seed After", .Random.seed[2])
  }

  return(data)
}
}
<bytecode: 0x7f824046d608>
<environment: namespace:fertile>

```

The majority of shims have a similar structure. Since almost all shimmed functions in **fertile** take a large number of arguments, the **fertile** versions utilize the dots to simplify the process of capturing this user input and saving it for execution later.

Additionally, the dots play a big role in the `proj_check_some` function. Recall that `proj_check_some` allows users to run a selection of **fertile**'s checks by calling a `tidyselect` helper to pull out a subset of checks with names matching a certain definition.

This function, which accepts the arguments “(path,...),” operates by allowing the users to pass in their `tidyselect` call to the dots. The list of available checks are converted into the columns of a dataframe, then passed through `dplyr::select(...)`, where the dots contain the information about the user's `tidyselect` call. Then, all of the checks matching the `tidyselect` call are run on the provided directory path.

2.2.5 Parameterized Reports

The `proj_badges()` function builds an **html** document based on a collection of information calculated when rendering an R project, then displays that **html** document in the **Viewer** window of **RStudio**.

This **html** document is customized based on both the computer and the directory that `proj_badges()` was run on:

Mac and Windows users will receive different outputs, as will those who have the same type of computer but are using a different **R** version or operating system (e.g. macOS Catalina 10.15.7 vs macOS Mojave 10.14.6).

The output of `proj_badges()` is also dependent on the files included in the directory it analyzes. When run on files with different content, names, and/or metadata, the function can produce reports showing different sets of passed/failed badges, different explanations for why a project failed certain badges, and a different list summarizing the names/modification dates of the files used to generate the project summary.

This customization is achieved through the use of an **RMarkdown Parameterized Report**. Parameterized reports allow for the creation of customized outputs based on different sets of inputs (parameters), all using the same basic template document.

Within **fertile**, there is an **RMarkdown** template that contains a variety of defined parameters. These include, among other things:

1. Vectors containing the names of badges that were awarded and of those that

- were failed, `2.tibbles` containing information why the failed badges were not achieved (the names of the badge-specific tests that failed), and
- 2. A `tibble` containing information about the names and last-modified dates of files in the project directory of interest.

Although it always follows the same basic structure defined in the template, the appearance of the knit output document varies based on the value of these parameters.

This functionality, along with the other methods of shims, hidden files, environment variables, and the dots helps improve the user experience of *fertile*. These unconventional techniques allow for the reliable tracking of user behavior behind the scenes and provide functions, like `proj_check_some`, that are intuitive for the user to operate.

2.3 Summary

Recall the list of features necessary for an effective reproducibility tool that were defined at the end of Chapter 1. Many of the tools and teaching methods considered previously fail to meet these standards: they are often complicated and/or narrow in scope. *fertile*, on the other hand, has none of these problems. Rather, the package meets all of the defined conditions for effectiveness:

1) Be simple, with a small library of functions/tools that are straightforward to use.

fertile is very simple to use. Most of the package’s features can be achieved with just two functions: `proj_analyze()` and `proj_check()`. The the interactive warning features—which throw errors when users reference absolute paths or those leading outside of the project directory—are even more straightforward. As soon as *fertile* is loaded, they are automatically enabled and require no additional effort from the user. Additionally, the operation of the functions themselves is quite simple—most functions in the package require only one argument: the path to the directory that the user wants to run the function on.

2) Be accessible to a variety of users, with a relatively small learning curve.

fertile has very few barriers to entry. The package only requires that its users be familiar with installing and loading packages from GitHub, executing functions in the console, and creating R projects, all of which can be learned in a handful of quick web searches. Running `proj_check_some()` successfully would also require knowledge of the *tidyselect* helpers (`starts_with()`, `contains()`, etc.), but that function is not necessary for use in any way. Users who were unfamiliar with its behavior could simply choose just to run a handful of individual functions instead and achieve the same results.

**** 3) Be able to address a wide variety of aspects of reproducibility, rather than just one or two key issues.****

fertile contains functions that address all six primary components of reproducibility (see Fig. 2.2). Some components, which are more complicated and have a variety of smaller parts—such as documentation—have more functions associated with

them. Randomness, the simplest component, has just one. While there are a variety of different functions for each component, it is not necessary for them to all be run independently. If a user is interested in checking all six components simultaneously, running `proj_analyze()`, `proj_check()`, and `proj_dependency_report()` together will achieve this goal.

4) Have features specific to a particular coding language that can address that language’s unique challenges.

`fertile` contains features to address the package system in R.

R is an open-source software that relies on packages (collections of functions) to achieve much of its functionality. Users who are looking to expand the available functions can contribute software packages to the community for public use. These packages are typically hosted either on the Comprehensive R Archive Network (CRAN) or on GitHub. Once a package is available on either site, any R user can download it and install it on their local R version.

Most data analyses in R are dependent on a variety of packages. Depending on the complexity and topic of the analysis, the exact number may vary, but most analyses rely on at least a handful to operate successfully.

The challenge with this system is that R packages are updated constantly by their creators. Users who go a few weeks without updating their software might find that dozens of their packages have updates available. Due to the frequency of updates, it is a common occurrence for code that once worked to stop functioning due to a change in the functionality of one of the packages it is dependent on.

As a result of this, tracking dependencies in R is more important than in some other coding languages. `fertile` attempts to address this through its dependency-tracking features (`proj_pkg_script()` and `proj_dependency_report()`), which together help the user of a package record the exact package versions that their project is dependent on and simplify the process of installing these dependencies by identifying which came from GitHub and which came from CRAN and creating a script with which to install them.

Another R-specific feature that `fertile` addresses is randomness. Randomness is incredibly important in statistics. Many statistical methods rely on random sampling in some way, and as a result, R—due to its purpose as a statistics-specific coding language—contains a wide variety of functions that use random number generation. In their default states, these functions produce a different result each time, and as a result are an inherent threat to reproducibility. In order to account for this, `fertile` contains the `has_no_randomness()` function, which reads scripts to ensure that any randomness (if present), is controlled (made pseudo-random) and reproducible.

5) Be customizable, allowing users to choose for themselves which aspects of reproducibility they want to focus on.

As discussed in Chapter 2.1.8, `fertile` is highly customizable, providing users with a wide variety of options for how they can run the package’s reproducibility tests. The main reproducibility checks can be run all at once (using `proj_check()`), in groups (using `proj_check_some()`), or individually. Users who only want basic information can rely on only `proj_analyze()` and `proj_check()`, while those looking for more advanced reproducibility information can delve into the project dependency functions,

as well as `sandbox() + proj_render()`, to ensure that projects are completely self contained.

6) Be educational, teaching those that use it about why their projects are not reproducible and how to correct that in the future.

`fertile` contains many different educational features. All of the reproducibility checks, when they fail, produce an informative warning message detailing where the failure occurred and providing a solution for how to address it. Additionally, the interactive warning system plays a big role in reproducibility education. Users are stopped immediately any time they use a non-reproducible file path and told why their path is problematic. This helps educate users on the correct use of file paths, ensuring that such mistakes do not happen many times.

7) Be applicable to a wide variety of domains.

Due to its low learning curve, customizability, and all-component-encompassing nature, `fertile` has the potential to provide benefits to users in a wide variety of domains. We will consider these in detail in the next chapter.

Chapter 3

Incorporating **fertile** Into The Greater Data Science Community

Finding a solution to addressing reproducibility on a widespread scale is a challenging problem. Attempts to do so—in academic publishing, software, and data science education—have made some progress, but many solutions have significant flaws. Primarily, they either:

- A) Only address one small aspect of reproducibility—for example, software that focuses on version control or a set of journal guidelines requesting only that code and data be provided, but giving no further detail.

OR

- B) Are challenging, time consuming, and/or burdensome to implement—for example, extensive journal guidelines, complex software packages with confusing functions, or academic courses on reproducibility that are only accessible to masters' students and take time away from other topics.

fertile is an attempt to address reproducibility in a way that does not fall victim to either of these challenges. Rather than focus on one area of expertise, **fertile** contains features focused on each of the six major components of reproducibility. Its self-contained nature allows users to address all aspects of reproducibility in one package; users can achieve near- or complete reproducibility with just a single piece of software.

fertile also makes the processes of both achieving *and* checking reproducibility simple and fast. Those looking to check whether a project is reproducible can almost instantaneously receive a full report of where the project succeeds and where it fails, and those looking to improve their reproducibility can receive and act on **fertile**'s clear suggestions with minimal effort. Some of the package's features are enabled automatically and most others can be accessed with only a handful of functions, all of which are very simple in function.

Additionally, **fertile** does not just provide a report on reproducibility and leave it at that. Instead, it attempts to teach its users the concepts of reproducibility in the

same way that reproducibility-focused classes are meant to do. Users receive instant feedback when making mistakes and, when checking work after writing it, receive reports clearly indicating where issues were found, why they occurred, and how to correct them.

It is also highly customizable, allowing users to utilize the tool in the way that fits their needs best. Those who want to focus their reproducibility checking in a certain direction have that option and those who want widespread overviews can also have their needs meet. Users who are interested in going beyond the base functionality of `proj_analyze` and `proj_check` also have additional functions at their disposal that they can use to check reproducibility, file paths, file types, etc.

3.1 Potential Applications of **fertile**

These features make **fertile** an excellent tool for addressing the issue of scientific reproducibility on a widespread scale. **fertile** can provide a variety of benefits to users in all different application domains and with all different experiences. In this chapter, we consider the many potential uses of the package.

3.1.1 In Journal Review

As discussed in Chapter 1, Academic journals have a significant reproducibility problem. In an attempt to address this, many journals have instituted reproducibility policies for submitting researchers to follow. Although a variety of journals have these policies, particularly in the Statistical and Data Sciences, very few actually go through the process of verifying that the standards are met. Authors, finding it to be a complicated and challenging task, will often not take the necessary steps to make their work truly reproducible. And journals, given the amount of time and money required to verify submissions' reproducibility, will often give submitting authors the benefit of the doubt in assuming that their work is reproducible as long as some code and/or data has been provided. This results in the publishing of many articles that claim to be reproducible in theory, but do not meet such standards when tested in practice.

fertile could provide significant assistance with this process. Journals could integrate **fertile** into their article review workflow, ensuring that certain reproducibility checks were passed before an article could be accepted.

Depending on the level of detail with which the journal wanted to examine reproducibility, the integration could be done in a variety of ways. Here, we'll consider two:

1. Journal reviewers run **fertile** on every submitted R project.

Journals that desire a detailed summary of the reasons for reproducibility failure (such as information that one specific file was not commented enough)—and whose editors were willing to put in a little bit of time to collect this information—could choose to run **fertile** on all submitted papers that included R code.

They could require a list of checks to pass and provide a list of exceptions for cases in which checks could fail—for instance, a journal could state that even though they support good documentation, they do not require code to be fully commented.

Authors could run *fertile* on their work before submitting it to a journal to ensure that they passed the required list of checks and that any failures they saw were accounted for in the journal exceptions. Then journals, in order to ensure that authors followed the provided reproducibility guidelines, could run *fertile* on each submitted article as part of the review process. If the required checks were passed, the article could be accepted, but if they failed it would be rejected.

Although it would require some effort on the journal’s side, this would still be an incredibly fast process: as long as a journal required that all submissions in R be in the R Project format, one reviewer could load the submission, run *fertile*, and receive a summary of the submission’s reproducibility in a matter of minutes.

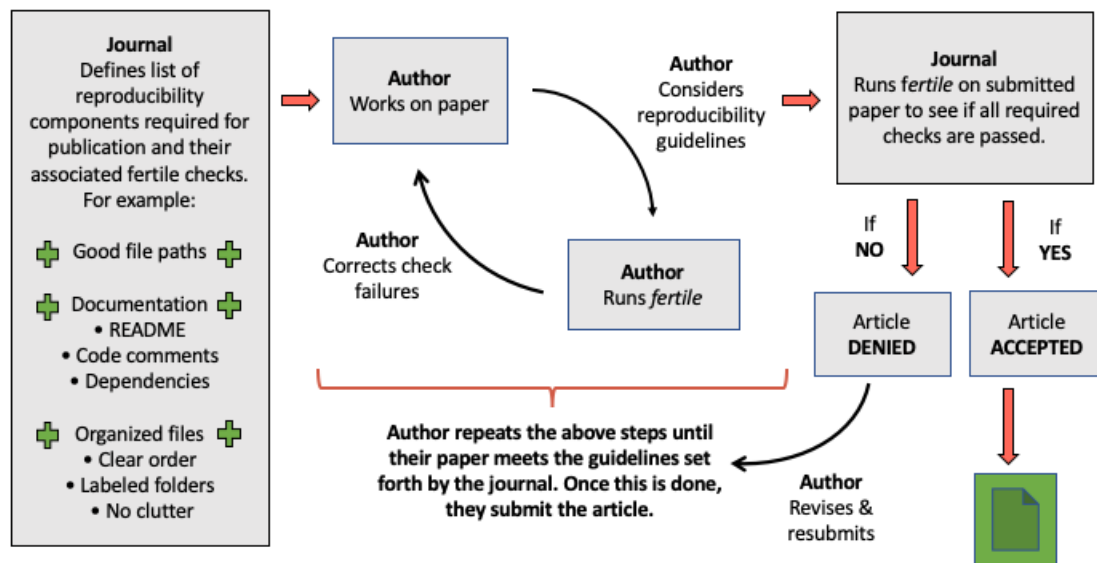


Figure 3.1: Potential ‘fertile’ Journal Review Process

2. Journal reviewers do not run *fertile* on each project, but instead require submitting authors to include a *fertile* summary sheet showing the reproducibility badges awarded to their work.

Some journals may desire a reproducibility summary, but not require details as specific as the exact reasons for failure. These journals could run a simplified approach to reproducibility review. Rather than spend the additional time to run *fertile* on every submission and review the reasons for check failure, the journals could instead require that authors include a coversheet—produced by the `proj_badges()` function—with their submission.

This cover sheet would show which of the six primary reproducibility components were met and which ones failed, a short summary of specific checks that were not

successful, and information about the cover sheet’s generation: who generated it, when, and with which files.

Journals could place an acceptance requirement that articles achieve a certain subset of badges—for instance, file paths, randomness, and documentation. Submitting authors would run *fertile* on their end to see which badges they passed. Once they met the requirements, they could run `proj_badges()` to generate an article coversheet. This coversheet would then be submitted alongside the article and considered as part of the review process. All that reviewers would need to do to ensure that reproducibility requirements are met would be to look at the cover sheet to see which badges the project achieved and check the cover sheet generation information to ensure that it was truly produced by the project that the author says it was produced by.

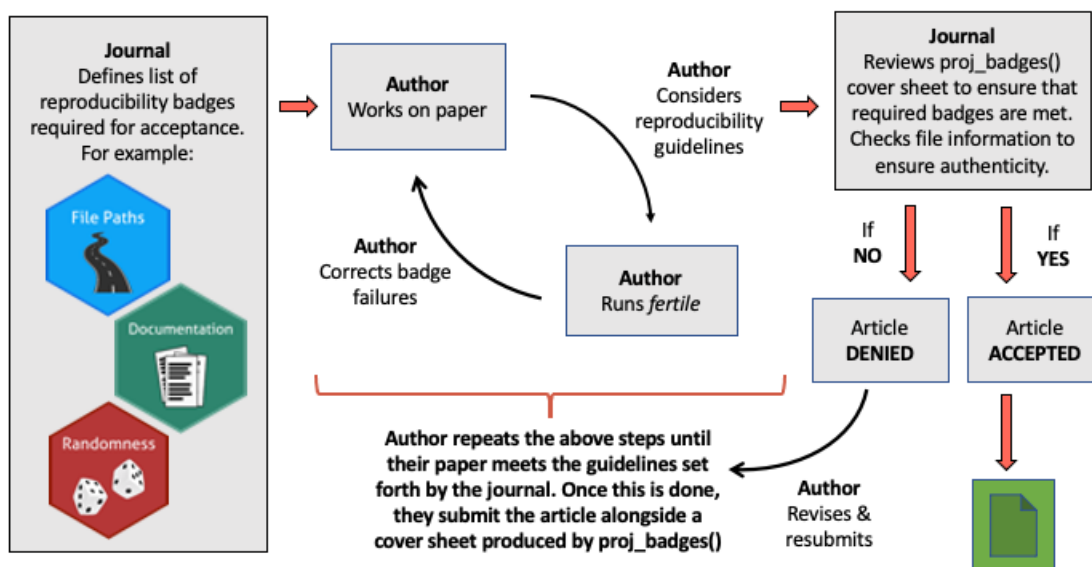


Figure 3.2: Another Potential ‘fertile’ Journal Review Process

Both of these processes would be much faster than that employed currently at the American Journal of Political Science, which goes through a thorough, multi-week-long reproducibility confirmation procedure for all submitted articles. Submitting researchers would know exactly which goals they were trying to achieve. They could download *fertile* on their own, run it on their project, check to see if their goals are met, and take the recommended steps to address failures if not. Then, upon submission, journals would only need to take minimal steps to ensure that those standards were met.

For journals that currently spend little effort on ensuring the reproducibility of submitted articles, *fertile* could provide significant benefit for minimal cost. Rather than spending money to hire a reproducibility editors, any journal board wanting to improve their publication’s reproducibility could instead choose to implement *fertile*—for free—as part of the review workflow.

Journals that already have designated reproducibility editors, such as *Biostatistics* and the *Journal of the American Statistical Association*, could also see benefits from *fertile* (Biostatistics (2020), Journal of the American Statistical Association Editors (2020)). The integration of *fertile* could speed up the review process, allowing the editors to review more papers for reproducibility adherence in a shorter time.

Although this would only address a small aspect of reproducibility—that involving data analysis projects written in R—it would provide a significant time- and money-saving impact for both authors and reviewers in that domain.

3.1.2 For Teaching Reproducibility

fertile could also be integrated into Statistical and Data Sciences coursework in order to educate students on topics of reproducibility.

Many of the existing programs to teach reproducibility are courses focused on replication studies, where students must take a published paper and replicate the steps within completely. This process, which includes requesting the necessary data and code files from the original author(s) and sometimes even expanding the existing analysis further, often requires that participants have knowledge of data analysis and the scientific research process to be successful. As a result, such courses focused on reproducibility tend to exist only at the graduate level.

5 Replication Paper Assignment

For graduate students enrolled in Gov2001, the main class assignment is to write a research paper that replicates and extends an existing scholarly work, while applying some advanced method to a substantive problem in some substantive field of study. Most undergraduate students enrolled in Gov1002 and all non-Harvard students enrolled in Stat E-200 complete a final exam instead of the replication paper (more information for these students in Section 10).

Your goal for this assignment is to produce a paper that is publishable in a scholarly journal — something we assume you have never done before and do not presently have any idea how to do. We will show you! Detailed information on the assignment can be found in an article I wrote about it called “Publication, Publication,” at [GaryKing/papers](http://garyking.org/papers) along with continuing updates. For initial versions of some papers from recent years, see the class Dataverse at [j.mp/G2001dv](https://dataverse.harvard.edu/dataset.xhtml?persistentId=doi:10.7927/H73T-6T90).

As this assignment involves carefully choreographed hand-offs and interactions that connects you to everyone else in class, please respect these deadlines. Everyone is depending on you. Here’s the list of requirements; the exact date and time of the deadline for each is given on the class web site, j.mp/G2001.

1. Identify your coauthors. All papers must be coauthored with one or two other members of this class. If you have problems, or would like suggestions, talk to the TFs.
2. Identify a scholarly article to replicate that meets our specific criteria. Upload to Canvas a PDF copy of the article, along with a brief paragraph (of less than about 200 words) explaining your choice. This paragraph must also list a classmate (outside your group) willing to testify that your article choice met all the criteria listed in “Publication, Publication”.
3. Turn in a draft of your paper with completed figures and tables, and a proposed outline of the paper, in a relatively polished form. This draft need not have much text yet (although the more you complete, the more useful comments you will get in return). Also turn in a *replication data file*, with all of the data and information necessary to replicate your results and reproduce your tables and figures. At the same time, we will assign your paper to several other students to replicate, and you will receive another group’s paper to replicate.
4. Replicate the other group’s proto-paper and write a memo to them (with a copy to us), pointing out ways to improve their paper and analysis. You will be evaluated based on how helpful, not how destructive, you are. The best comments are written so fellow students can hear and learn from them rather than trying to demonstrate how smart you are.
5. Turn in the final version of the paper. By the same deadline, you must also follow standard academic practice and create a permanent replication data archive by uploading all your data and code to the Gov2001 Dataverse ([j.mp/G2001dv](https://dataverse.harvard.edu/dataset.xhtml?persistentId=doi:10.7927/H73T-6T90)). If you would like an *extension* with this (and only this) deadline, you do not need to ask permission: We will accept papers for a week after the due date given on the class web site (although since you will have had more time, papers turned in after the original deadline will be graded according to proportionately higher standards).

Figure 3.3: The Replication Assignment From Harvard Professor Gary King’s Gov 2001 Graduate Course

Undergraduate students, therefore, do not get exposed to reproducibility very often. There are a few exceptions—for instance, introductory courses at Smith College and

Duke University that integrate RMarkdown to promote reproducible workflows—but overall, reproducibility is not covered in data science courses the undergraduate level.

fertile could help change this, allowing for many more colleges and universities to integrate reproducibility into their courses. The barriers to entry for using and benefitting from the package are very low, requiring only that participating students have:

- R and RStudio installed on their computer
- Knowledge of how to install a package from GitHub and load it into their environment
- Knowledge of how to create an R project
- Knowledge of how to run basic functions and input simple file paths

Though the process may entail some confusion and troubleshooting at first, even those brand new to R could succeed in overcoming these barriers in only a few days of class. As a result, *fertile* could provide professors with an opportunity to teach reproducibility concepts in introductory level courses.

fertile could easily integrate into coursework in a similar way to how RMarkdown was integrated at Smith College and Duke University. While there is not only one way to utilize the software in class, a potential use of *fertile* could look as follows:

At the beginning of their courses, the professor provides their students with a brief introduction to reproducibility, including its importance and a basic description of how it is achieved. Shortly after, they introduce R Projects and the *fertile* package, explaining that they are tools to help with reproducibility. Then, they institute a requirement for all submitted homework assignments in the course: students must create and submit their work in the R Project format, but prior to submission must run *fertile* on their project to ensure that it passes reproducibility standards. When reproducibility errors inevitably occur, they can be used as teaching moments: the professor can share the error, explain why it happened, walk through *fertile*'s response to it, and interactively work with students to illustrate how it can be fixed.

The integration of *fertile* in this way would be an excellent method to introduce students to reproducibility concepts early on in their data science education, but at a low cost to the professor.

There are a variety of benefits to introducing students to reproducibility sooner, rather than later—in graduate school or through independent research on the topic:

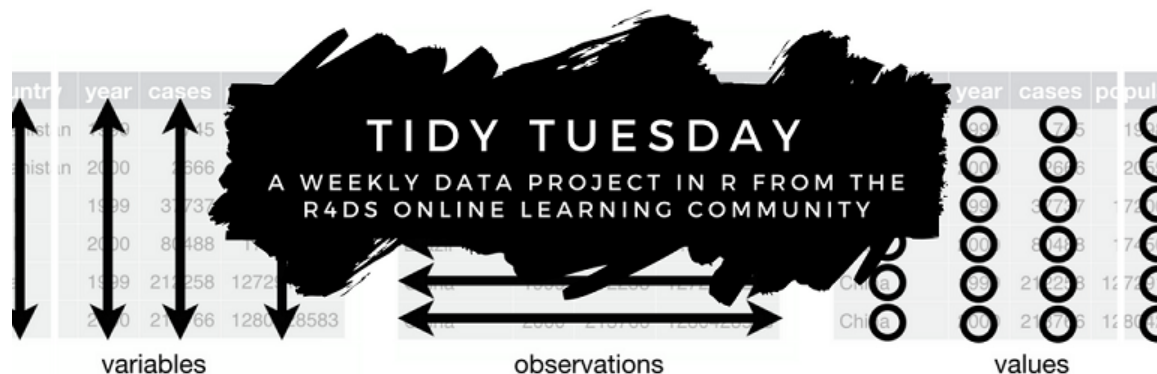
- Teaching reproducibility early on gives students important research tools and understanding before they conduct any of their own important analysis.
- Practicing before students are believed to be skilled and highly educated in data science gives them an opportunity to fail and learn without fear of judgement.
- Integrating concepts early helps ingrain them in the minds of students, ensuring that reproducibility begins to come naturally to them.

These students would then be prepared for entering the research world and contributing to data science work in a transparent and reproducible way.

3.1.3 In Other Areas

fertile can also provide benefits in other domains. Though not an exhausted list, some of the potential uses of the software are:

- *Private Companies:* Data analysis-focused companies could require their employees to use *fertile* to check the reproducibility of their projects before presenting them to clients. This would help such companies ensure that clients could trust the results that were being produced.
- *Conferences:* Similar to academic journals, conferences promoting open research could require that papers written in R pass a *fertile* check as a condition for acceptance. Even if there were an exception given for those using confidential/identifiable data, this would likely increase the overall reproducibility of conference papers significantly.
- *Informal Data Analysis:* A lot of content in the R community is created purely for fun and interest. Outside of work, many R users will create data visualizations or analyses for their own private blogs or their twitter. Sometimes, users will also participate in community events like Tidy Tuesday, a weekly social project where a data set is posted and users are asked to analyze it and create a visualization of their choice. Many people use these informal analyses as an opportunity for learning and discussion, often sharing them on social media to try and get feedback on their work. Ensuring that the work is reproducible would facilitate this process. Users could run their project files through *fertile* to check that they are reproducible and post a link to download them. This would then allow others to run the code on their own to understand how it works and more easily be able to make suggestions as to how to improve it!



fertile is incredibly versatile in its applicability. It can be used anywhere from informal data analysis projects to academic journal review.

3.2 Testing fertile in the Real World: Experimental Design and Analysis

3.2.1 Background

Many of the suggested uses for `fertile` in the previous section are hypothetical, included for the purpose of demonstrating the potential that the package could have. Without experimentation or testing, though, it is difficult to know just how successful `fertile` would be when employed for any given purpose.

In order to gain a better understanding of the practical effects of using `fertile` in a real life scenario, Professor Baumer and I decided to conduct an experiment (approved by Smith College IRB, Protocol #19-032). We were interested in testing whether the integration of `fertile` into an introductory data science course might have an effect on students' learning of reproducibility concepts.

After designing our experimental structure, working out technical mechanics, and creating the necessary materials, we chose to use the following two course sections for our study:

1. Introduction to Data Science (SDS 192), Fall 2020 Semester
2. Introduction to Data Science (SDS 192), January-Term 2021

There were several motives for selecting these specific courses:

- They were both being taught by Prof. Baumer, which made it easy to ensure that everything went smoothly by removing the need to interact with a 3rd party.
- They were two different sessions of the same course, with essentially identical material, ensuring that we could compare results from each section of the experiment.
- They involved introductory data science students, one of the groups that were of most interest as we wanted to ensure that `fertile` could provide benefit to users of all skill levels.
- They were somewhat larger than many other classes in the Statistical and Data Sciences department, providing an opportunity for more data collection.

3.2.2 Experimental Design

For the experimental structure, we chose to complete a randomized controlled trial (RCT) since RCT designs can be used to conclude causality and are designed in such a way as to limit bias in the results.

Our goal was to determine whether students who had `fertile` installed and loaded on their R Studio applications would learn more about reproducibility throughout the course than students who did not, or whether there was instead no difference or even a negative effect.

In order to measure this potential effect, we opted to use reproducibility “tests,” forms containing multiple choice and select-all-that-apply questions about different aspects of reproducibility that are focused on in `fertile`.

We used two different testing structures for our two experimental sections—partially due to challenges which led to our first experiment session starting on a slight delay, but also so that we could compare which structure worked more effectively. While the primary goal of the experiment was to determine the effect of *fertile*, a secondary interest was in the experimental method itself—a new method that we developed, which has implications beyond purely this case.

- Structure #1 (Used in Fall 2020): Students received a reproducibility test at the end of the semester to test their knowledge after the course. Final scores were compared between students who had *fertile* installed and loaded and those who did not.
- Structure #2 (Used in January Term 2021): Students received a reproducibility test both at the beginning of the course *and* at the end of the course in order to test their *change* in knowledge. *Differences in scores* were compared to see if any given group learned more, less, or the same.

Although the testing structures were different across the two different sections, the reproducibility tests were identical.

To reduce bias to a maximum degree, we opted to use blinding, in which participants were unaware of whether they had received the experimental or control condition—in this case, whether they had *fertile* installed and loaded while working on course assignments, or whether they didn't.

To achieve the desired blinding, we decided to use some unique features of R packages which allowed us to give all students in the course the same software, but have it behave differently when loaded, depending on which experimental group the student was a part of. This technique is described in the step by step experimental structure, explained below:

1. Students were given a form at the start of the semester asking whether they consented to participate in a study on data science education. Additionally, information was collected about the previous number of courses that student had taken in data/computer science fields, which we believed might impact their knowledge of reproducibility.

In order to successfully consent, students had to provide a unique computer identifier, collected through either the command `Sys.getenv("LOGNAME")` or `Sys.getenv("USERNAME")`, depending on the student's operating system. This identifier was collected in order to ensure that students could be individually identified and therefore assigned effectively to treatment or control groups.

2. To maintain student privacy, the unique identifiers were then transformed into hexadecimal strings via the `md5()` hashing function. While these hashed sequences have meaning in the students' computers and the course software program, to those without the original identifier, they appear only as a meaningless string of letters and numbers.
3. These hexadecimal strings, which correspond to unique students, were then randomly assigned into equally sized groups, one experimental group that receives the features of *fertile* and one group that receives a control.

4. The students in the course were then asked to download a package called `sds192` (the course number and prefix), which was created for the purpose of this trial. The package contained several templates and data sets that were necessary for use in the course, so all students in the class—regardless of their experimental status—needed to install it.

The `sds192` package operated by leveraging an `.onAttach()` function to scan the R environment of each student *each time the package is loaded*, collect their unique identifier—again, via `Sys.getenv("LOGNAME")` or `Sys.getenv("USERNAME")`—and run it through the same hashing algorithm as before. This function came pre-coded with the hashed identifiers of all of the students, grouped by whether they had been assigned to the experimental or the control group.

Each time a student's hashed identifier was collected (every time they loaded the `sds192` package for class), it was then compared with the provided groups to determine how the package should function for that individual.

The structure of this function can be seen below:

```
# .onAttach() from the sds192 package
# Run automatically any time the sds192 package is loaded

.onAttach <- function(libname, pkgname) {

  # The experimental group gets `fertile` loaded secretly
  if (is_experimental()) {
    suppressMessages(library(fertile))
  }
}

is_experimental <- function(logname = whoami::username()) {

  # Students are placed into experimental and control groups but remain anonymous.
  fertile_group <- c(
    "f7b0a9d5117b88cecec122f8ba0e52fb", "4d0295a810fb8491f91f914771572485",
    "36211a1f19f82ae07aed990b671c9b20", "b5d2b72b4f36f3afdce32a8409dc6ea0",
    "d498227fd9e6a4c42494bbebc42f6aa8", "191922566ef6a9910682ad9836b6d018",
    "b1b8278d6b7eecd2c595ab6138de17e0", "4894c3d932fa6cfc9ec59214c41f12c7",
    "2f1e21316352049069f8e4351d6cb88d", "73f4a06a26ad5342e30e2e7fdf92dba2",
    "9ff232c0e56f04be4ba2037aec6471b2", "abb2b711c8d45ffac12006945e10ed31",
    "3a49dd3c6e591933a0909734a94ca37c", "db1b731709316be69dae1d56382e4243",
    "8e76b368a5e999869c7ca8f9f1566cf8"
  )

  control_group <- c(
    "9aa36583f54766205850428e8f1a4c89", "fdc61d4d4c93473be5f485eea55140b3",
    "f03020938b31818063c79d2422755183", "d0a981421d0e378e26186f99c618c748",
    "7ec57b1f2bca9ac1e702fb68427b781b", "8857ca4d5e1fe92ae70b1ba95c0b7b8b",
```

```

    "e5e30623e9d09d29ded851b7fb40cb51", "592572bb9fce168f37117fd0d6e0e5ee",
    "c40b1786558a15f8dd151da163ebd0b4", "67b8da3952e07ba3f2c5c715c4042220",
    "f5ca3188ff14cbf43b6d4cd5b225376e", "b3b222f42beaae6ca2b309b94b1340be",
    "2dbac7345a71a13c434af882853e86bd", "3c5df10c572b5627418c034f65f52dee",
    "0ff07a4f6faaf3df57b348dad7bd22c9"
  )

  digest::digest(logname, algo = "md5") %in% fertile_group
}

```

5. The experimental group received the basic **sds192** package, which consisted of some data sets and R Markdown templates necessary for completing homework assignments and projects in the class, but also loaded **fertile** silently in the background. The package's proactive features were enabled, and therefore users would receive warning messages when they used absolute or non-portable paths or attempted to change their working directory. The control group received only the basic **sds192** package, including its data sets and R Markdown templates.
6. [January Term 2021 Only] Students then took a reproducibility test prior to fully starting the course, which tested their knowledge before completing the class.
7. The students then completed their coursework as normal, regularly using the **sds192** package for projects and homework assignments.
8. [Both Fall 2020 and January Term 2021] The last step was for students to take a reproducibility test at the end of the course, to measure their knowledge of the subject after having completed the coursework and used the **sds192** package.

3.2.3 Results

In order to understand the results, it is important to understand the structure of the experimental reproducibility “test” and the way in which each student’s answers were scored.

The test consisted of two primary sections:

1. A set of six questions on RStudio projects, two of which were select-all-that-apply, and four of which were multiple choice. These questions tested students knowledge of RStudio projects, their understanding of which file paths they could include in code submitted in project format, and their awareness of the dangers of changing working directories while coding.

What are the benefits of using RStudio Projects? Select all that apply. *

- ☐ They make it easy to share all of your code/files with other people
- ☐ They help keep all your files in one place on your computer, separated from other unrelated files
- ☐ They allow you to connect to version control tools like GitHub
- ☐ They speed up how fast your R session loads/saves
- ☐ They take up less space in your hard drive than when storing all of your files in one place

Figure 3.4: Sample Question From The Section On RStudio Projects

...

Assume that you have stored some data that you need to analyze in your homework in a file called `'my_data.csv'` on your computer's Desktop. In your R Markdown file, you use the following code to read in your data and it executes successfully. You submit both files to your instructor. Will the R Markdown file compile on your instructor's computer? *

```
```{r}
data <- read_csv("~/Desktop/my_data.csv")
```
```

- ☐ Yes
- ☐ No
- ☐ It depends
- ☐ I'm not sure

Figure 3.5: Another Sample Question From The Section On RStudio Projects

2. A set of three questions about file paths, all of which were select-all-that apply. These questions tested students' knowledge of the difference between absolute and relative paths and their understanding of which paths are reproducible.

Which of the following are absolute paths? Select all that apply *

- ☐ "C:\Documents\Datasets\SnowConeSales.csv" (Windows)
- ☐ "Datasets\SnowConeSales.csv" (Windows)
- ☐ "SDS192/mp2/contributions.csv" (Mac)
- ☐ "~/Desktop/bikes.csv" (Mac)
- ☐ "/Users/<yourname>/Documents/SDS192/mp2/contributions.csv" (Mac)

Figure 3.6: Sample Question From The Section On File Paths

For each test submission, every question was scored individually and then scores for each section (projects and paths) were calculated, as well as an overall total score.

The following method was used to compute scores:

- For each question, students would start at a score of zero. Based on their answers, points would then be added, subtracted, or stay constant.
- On select-all-that apply questions, the following scoring rules were used. This meant that students who answered some things correctly, but other things incorrectly, could still receive negative or neutral scores, based on the overall analysis of how correct they were:
 - Each correct box that was checked → +1 Point
 - Each incorrect box that was checked → -1 Point
 - Each box that was not checked but should have been checked → No change (0)
- On multiple choice questions, the scoring rules were slightly different. This was due to the inclusion of an option for “I’m not sure,” which was included as a way to discourage random guessing and ensure that students were answering based on their actual knowledge.
 - If the correct answer was selected → +1 Point
 - If the incorrect answer was selected → -1 Point
 - If the student expressed uncertainty (“I’m not sure”) → No change (0)

This method meant that individual questions could have negative, positive, or neutral scores, and total scores could vary quite dramatically, from large negative scores (<-10) for students who regularly answered incorrectly to large positive scores for those who often answered correctly (>10).

Fall 2020 Results

The data from Fall 2020, after being scored and cleaned up, looked as follows:

```
glimpse(fall2020)
```

```
Rows: 18
Columns: 15
$ User_Hash      <chr> "\"67b8da3952e07ba3f2c5c715c4042220\"", "\"f030209...
$ Group          <chr> "Control", "Control", "Fertile", "Fertile", "Ferti...
$ Previous_Classes <dbl> 0, 0, 2, 1, 2, 1, 6, 0, 1, 1, 1, 2, 1, 1, 5, 1, 3, 1
$ Q1_Points      <dbl> 3, 1, 1, 3, 3, 2, 3, 3, 3, 1, 1, 3, 1, 1, 2, 2, 3, 1
$ Q2_Points      <dbl> 0, -1, 0, -1, -1, 1, -1, 1, -1, 0, -1, -1, 0, -1, ...
$ Q3_Points      <dbl> 0, 1, 0, -1, -1, 0, -1, 1, -1, 0, -1, -1, 0, 1, 1,...
$ Q4_Points      <dbl> 1, 1, 1, 1, 1, 0, 1, -1, 1, 0, 1, -1, 0, 1, 1, -1,...
$ Q5_Points      <dbl> -1, 1, 0, 1, 1, 1, 0, 1, 1, 0, -1, 1, 0, -1, 1, -1...
$ Q6_Points      <dbl> 0, 1, 1, 1, 1, 1, 0, 0, 1, 0, 0, 0, 0, 1, 1, -1, 1, 0
$ Q7_Points      <dbl> 2, 2, 3, 0, -1, 3, 3, 3, -1, 2, 2, 2, 2, 2, 3, 1, ...
$ Q8_Points      <dbl> -1, 2, -2, 2, -2, 2, 2, 2, -2, 2, 1, 1, -1, 1, 2, ...
$ Q9_Points      <dbl> 1, 1, 0, 2, 2, 2, 2, 2, 1, -1, 1, -2, 0, 1, 1, 1, ...
$ Projects_Total  <dbl> 3, 4, 3, 4, 4, 5, 2, 5, 4, 1, -1, 1, 1, 2, 7, -2, ...
$ Paths_Total    <dbl> 2, 5, 1, 4, -1, 7, 7, 7, -2, 3, 4, 1, 1, 4, 6, 1, ...
$ Overall_Total   <dbl> 5, 9, 4, 8, 3, 12, 9, 12, 2, 4, 3, 2, 2, 6, 13, -1...
```

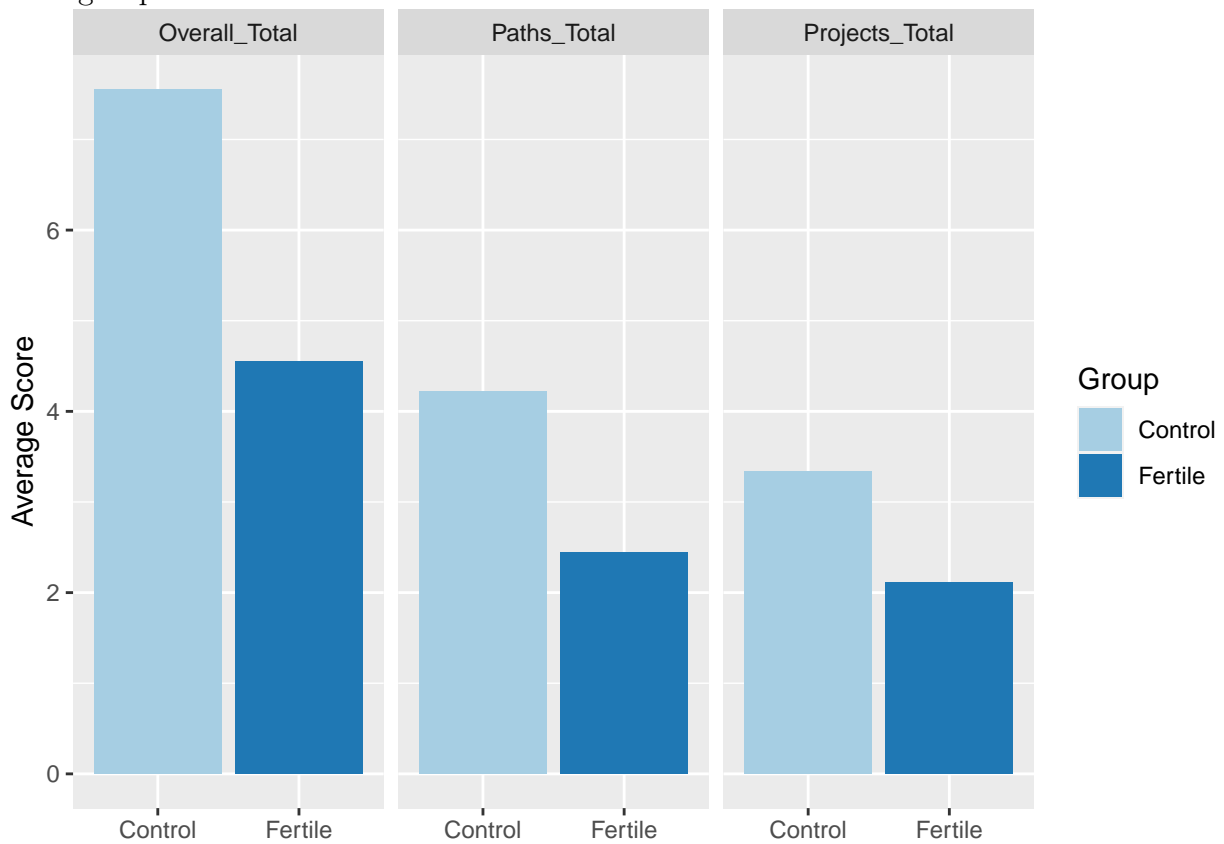
There were 18 students who completed the reproducibility test, split evenly into two groups of 9, and 15 variables per student. Since the Fall 2020 section of the experiment contained only one reproducibility test, each student only has one associated observation in the data.

1. **User_Hash**: Hashed identifiers, representing unique students.
2. **Group**: A categorical variable summarizing whether the student was in the experimental (**fertile**) or control group.
3. **Previous_Classes**: The number of previous data science / computer science courses the student had taken at the start of the semester.
4. **Qx_Points**: The number of points scored on each individual question.
5. **Projects_Total**: The total number of points scored on the six questions associated with R Projects.
6. **Paths_Total**: The total number of points scored on the three questions associated with file paths.
6. **Overall_Total**: The total combined score of all nine questions.

The score averages for both groups of students were as follows

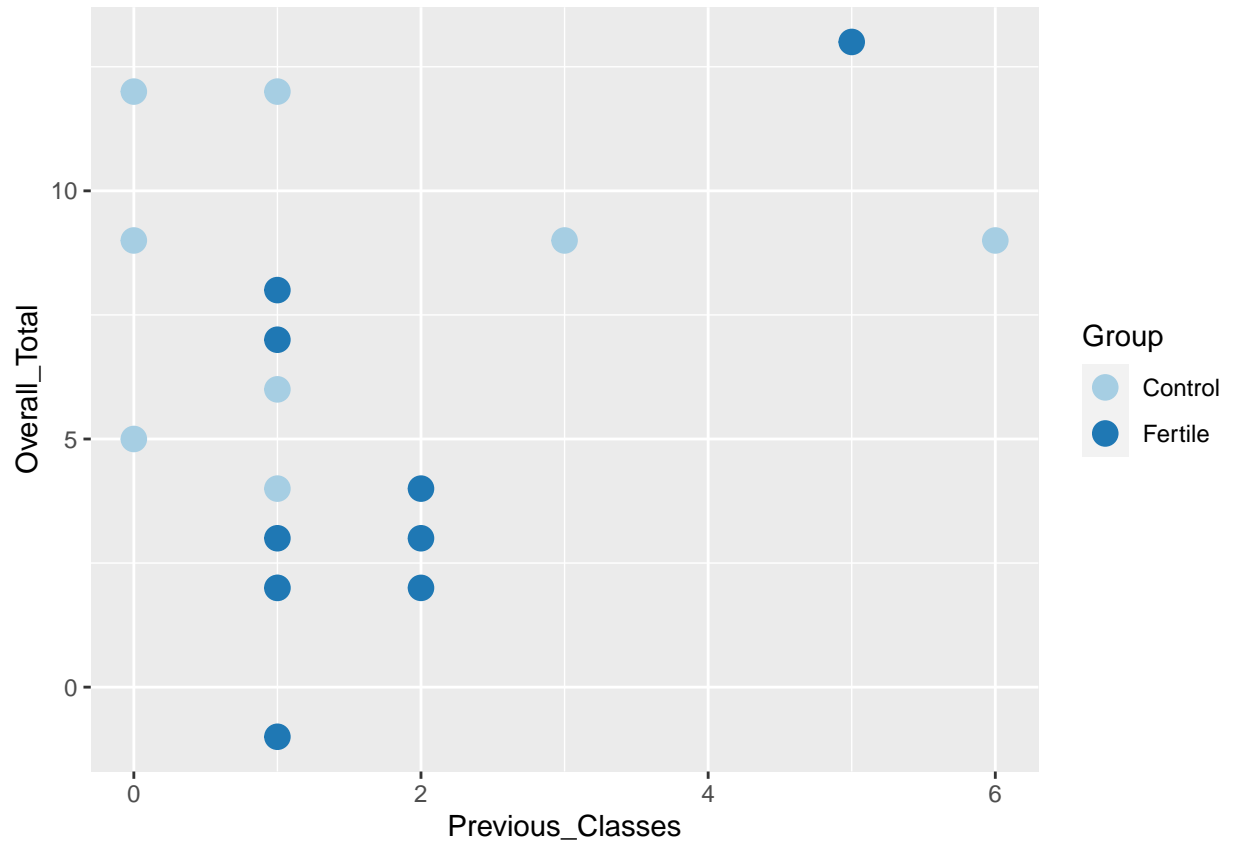
```
# A tibble: 2 x 4
  Group Projects_Avg Paths_Avg Overall_Avg
* <chr>      <dbl>      <dbl>      <dbl>
1 Control    3.33      4.22      7.56
2 Fertile    2.11      2.44      4.56
```

These numbers, combined with the visual below seem to indicate, at least in this small sample, that the *fertile* group actually performed slightly worse, overall, than the *control* group.



Looking at the overall total score (*Overall_Total*), this was true even when controlling for the number of relevant courses taken previously:

```
ggplot(fall2020, aes(x = Previous_Classes, y = Overall_Total, col = Group)) +  
  geom_point(size = 4) +  
  scale_color_brewer(palette = "Paired")
```

However, when analyzed mathematically, the results appear less negative. A t-test and analysis of variance (ANOVA) conducted on the simple regression model $Overall_{Total} \sim Group$ both indicate that, although there is a potential negative trend between the use of `fertile` and a student's score on the reproducibility test, there is not enough evidence to conclude that this relationship was *not* just the product of chance. The p-values of 0.117 are not below the threshold of 0.05 used to determine statistical significance.

Call:

```
lm(formula = Overall_Total ~ Group, data = fall2020)
```

Residuals:

| Min | 1Q | Median | 3Q | Max |
|--------|--------|--------|-------|-------|
| -5.556 | -2.556 | -1.056 | 2.194 | 8.444 |

Coefficients:

| | Estimate | Std. Error | t value | Pr(> t) |
|--------------|----------|------------|---------|--------------|
| (Intercept) | 7.556 | 1.281 | 5.896 | 2.25e-05 *** |
| GroupFertile | -3.000 | 1.812 | -1.655 | 0.117 |

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 3.844 on 16 degrees of freedom
 Multiple R-squared: 0.1462, Adjusted R-squared: 0.09288
 F-statistic: 2.741 on 1 and 16 DF, p-value: 0.1173

Analysis of Variance Table

Response: Overall_Total

| | Df | Sum Sq | Mean Sq | F value | Pr(>F) |
|-----------|----|--------|---------|---------|--------|
| Group | 1 | 40.50 | 40.500 | 2.7406 | 0.1173 |
| Residuals | 16 | 236.44 | 14.778 | | |

January Term 2021 Results

We see a similar story with the data from January Term 2021, though the structure of the data set is different.

The January Term data looks as follows:

```
Rows: 76
Columns: 17
$ User_Hash      <chr> "\"96eb0673c18b9fa79740686995b1d970\"", "\"980a97c...
$ Student_ID     <dbl> 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15,...
$ Group          <chr> "Control", "Control", "Fertile", "Fertile", "Ferti...
$ Previous_Classes <dbl> 1, 1, 2, 1, 1, 4, 3, 0, 0, 1, 1, 4, 3, 4, 2, 4, 4,...
$ Test           <fct> Pre, Pre, Pre, Pre, Pre, Pre, Pre, Pre, Pre, Pre, ...
$ Q1_Points      <dbl> 2, 2, 1, 2, 1, 3, 2, 2, 1, 2, 1, 2, 2, 1, 2, 1, 1,...
$ Q2_Points      <dbl> -1, 0, -1, -1, 1, 1, 1, 0, 0, -1, -1, 0, 1, -1, -1...
$ Q3_Points      <dbl> -1, 0, -1, 0, 0, 1, -1, 0, 0, -1, 0, 0, 1, -1, 0, ...
$ Q4_Points      <dbl> 1, 0, 1, 1, 0, 1, 1, 0, 1, 1, -1, 1, 1, 1, 1, 0, 1...
$ Q5_Points      <dbl> 1, 0, -1, 1, 0, 1, -1, 0, 0, -1, 1, 0, 1, 1, 0, 0,...
$ Q6_Points      <dbl> 1, 1, 1, 1, 1, 1, 0, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1,...
$ Q7_Points      <dbl> 2, -2, 0, 0, 3, 2, 3, 1, 2, 3, 1, 2, 3, 3, 2, 2, -...
$ Q8_Points      <dbl> 1, -3, -1, -2, 2, 1, 2, 0, 1, 2, 1, 1, 2, 2, 1, 1,...
$ Q9_Points      <dbl> 1, 2, 1, 1, 1, 1, -2, 0, -2, 2, -1, -2, 2, 2, -1, ...
$ Projects_Total <dbl> 3, 3, 0, 4, 3, 8, 2, 3, 3, 1, 1, 4, 7, 2, 3, 2, 6,...
$ Paths_Total    <dbl> 4, -3, 0, -1, 6, 4, 3, 1, 1, 7, 1, 1, 7, 7, 2, 4, ...
$ Overall_Total  <dbl> 7, 0, 0, 3, 9, 12, 5, 4, 4, 8, 2, 5, 14, 9, 5, 6, ...
```

It has many of the same features as the Fall 2020 data, though there are a few differences.

1. This data set contains data on 38 students, rather than 18.
2. Each student has two rows of data that correspond to them, rather than just one.
3. There are two new variables, `Student_ID` and `Test`. `Student_ID` is an index, used to keep track of which rows belong to the same student. This was created as an alternative to `User_Hash` as we found that users of Smith College-provided

laptop computers would occasionally have non-unique hashed IDs. `Test` has either the value “Pre” or “Post”, identifying which test the row corresponds to—the test at the beginning of the course, or the one at the end, respectively.

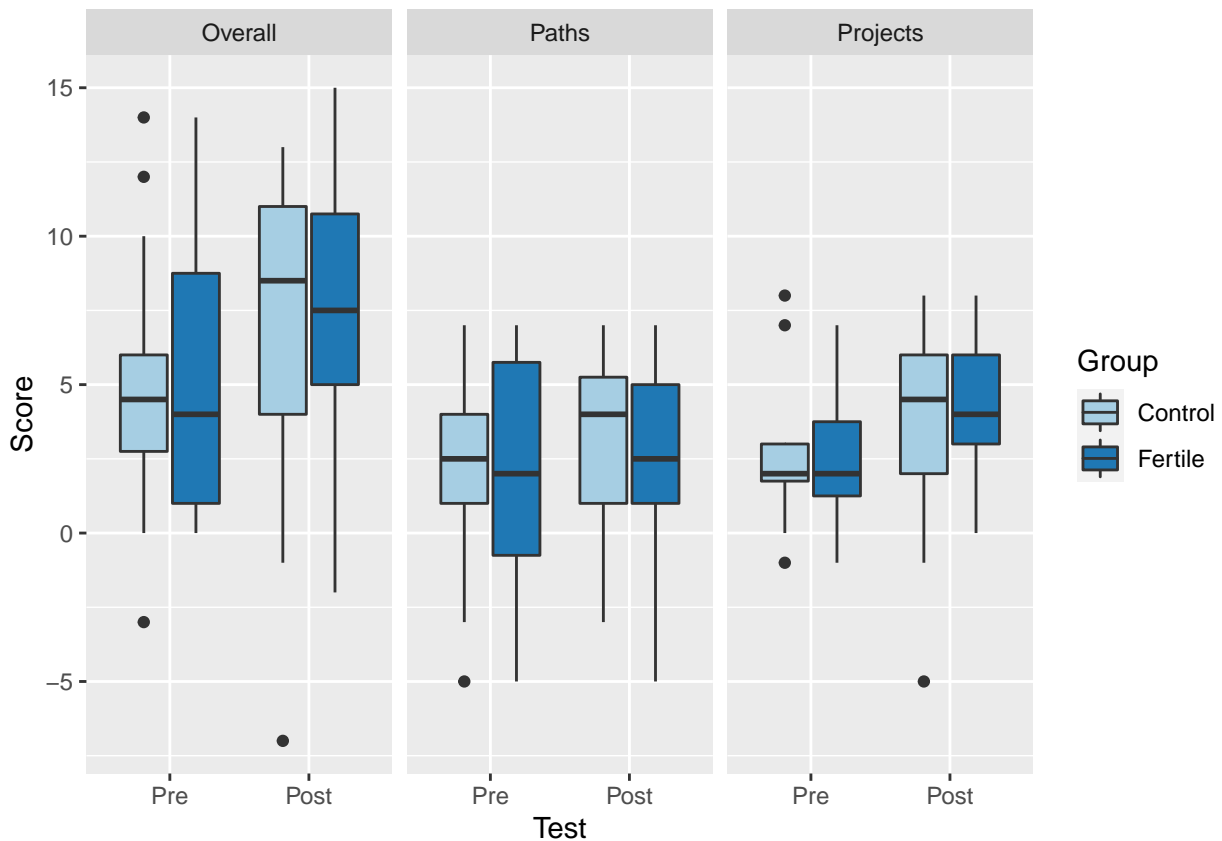
When calculating the difference in reproducibility test scores between the end (“Post”) and beginning (“Pre”) of the course, the story looks a little bit less negative than before, but once again there does not appear to be much strong evidence in favor of `fertile`.

For the questions about file paths, there appears to be no difference in the average score at the beginning and end of the course for either group. For the questions about R Projects, the `fertile` group had slightly higher average score differences but not by that much (1.92). For the overall total number of points, the score differences were very similar (2.15 for the `control` group and 2.22 for `fertile`).

```
# A tibble: 6 x 3
# Groups:   Group [2]
  Group Question_Group Avg_Score_Difference
  <chr>   <chr>                <dbl>
1 Control Paths                0
2 Control Projects            1.2
3 Control Total               2.15
4 Fertile Paths                0
5 Fertile Projects            1.89
6 Fertile Total               2.22
```

When considering the distribution of test scores, in addition to just the pure difference, we see a pattern where those students in the `fertile` group appeared to score worse than the `control` group, on average. Yet, when considered all together, the `fertile` group’s scores improved slightly more over the time of the course than the `control` group’s.

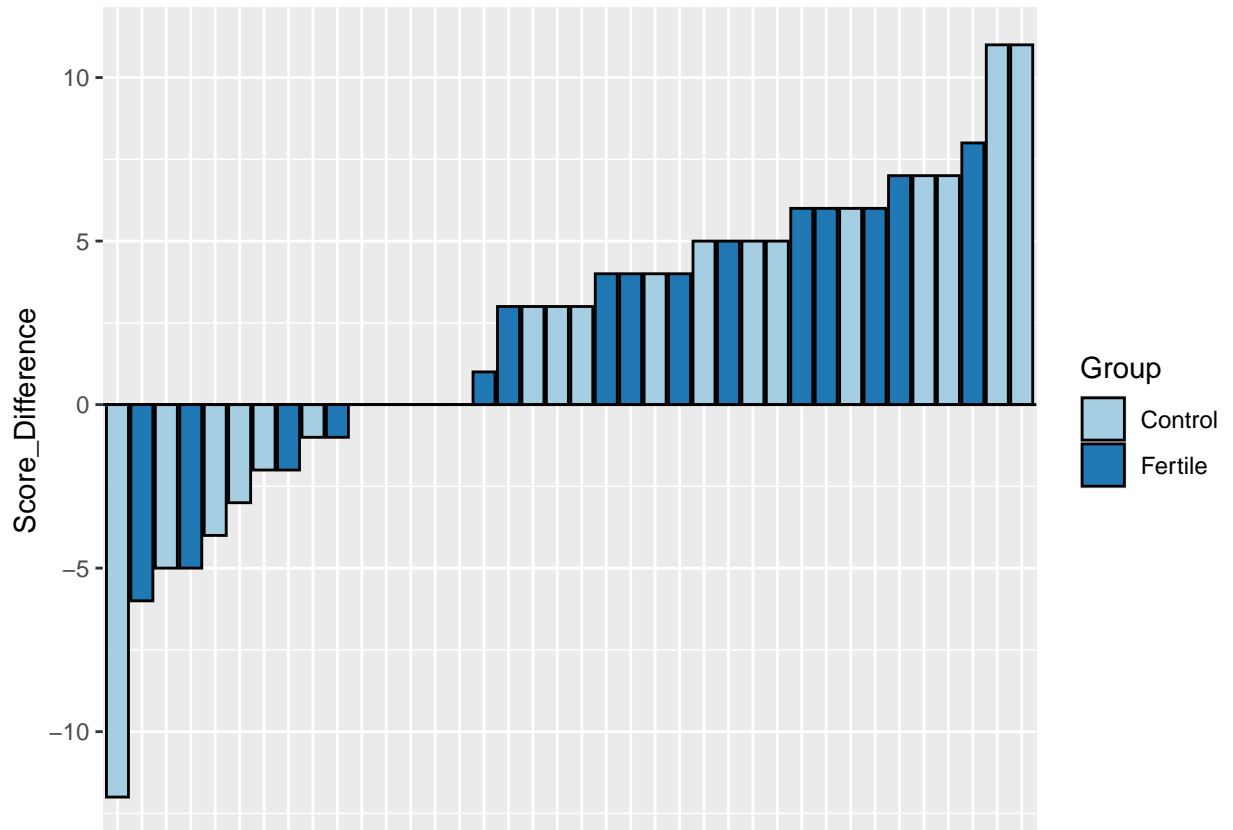
This evidence appears to strengthen the previous claim—using the Fall 2020 data—that the potential negative relationship between reproducibility test score and use of `fertile` is likely a product of random chance. In both experimental sections, Fall and January Term, students in the `fertile` group performed slightly worse than the `control` group on the reproducibility tests taken at the end of the course. However, the additional evidence from the January Term “Pre-Test” appears to indicate that the reason for that lower score might be just that they started from a lower point of knowledge at the beginning, rather than that `fertile` directly affected their scores.



We can also view the difference in scores on a more individual level. Below, we see the score differences (looking purely at `Overall_Total`, rather than just `Paths` or `Projects`) of all 38 students in the January Term study, ordered from lowest score differential (those who performed worse on their Post-Test than their pre-test), to highest.

As we can see, the majority of students improved on their reproducibility knowledge throughout the course. However, some students actually performed significantly worse over time. While students in both the `fertile` and `control` group had negative performances such as these, more of the highly-negative showings came from those in the `control` group. There is one outlier who appears to have performed significantly worse on their Post-Test than their Pre-Test. When looking at students who performed better over the course, the distribution is slightly more balanced. The two students who made the most improvement, however, were both in the `control` group.

Once again, there is not much evidence that `fertile` is providing a significant benefit, and the outlier on the left indicates that the reason for the slight edge of `fertile` in terms of average score differences might just be due to one student skewing the `control` group's averages.



It is also interesting to examine the relationship between the number of previous courses taken by students and their performance on the reproducibility tests.

Below, we see a graphical representation of the number of previous courses taken by each student and the difference between their overall total score on their Pre- and Post-Tests.

Although it is challenging to make any strong claims with so few data points, one interesting pattern emerges. For the **control** group, previous experience did not appear to be an indicator of stronger performance or overall learning throughout the course. In some cases, overall performance of students with several previous classes was worse on the Post-Tests than the Pre-Tests. However, with the **fertile** group, there does appear to be a slight positive relationship between the number of previous classes and the test scores. On average, the **fertile** group always performed better on their Post-Tests than on their Pre-Tests.

```
Warning in simpleLoess(y, x, w, span, degree = degree, parametric =
parametric, : pseudoinverse used at 2
```

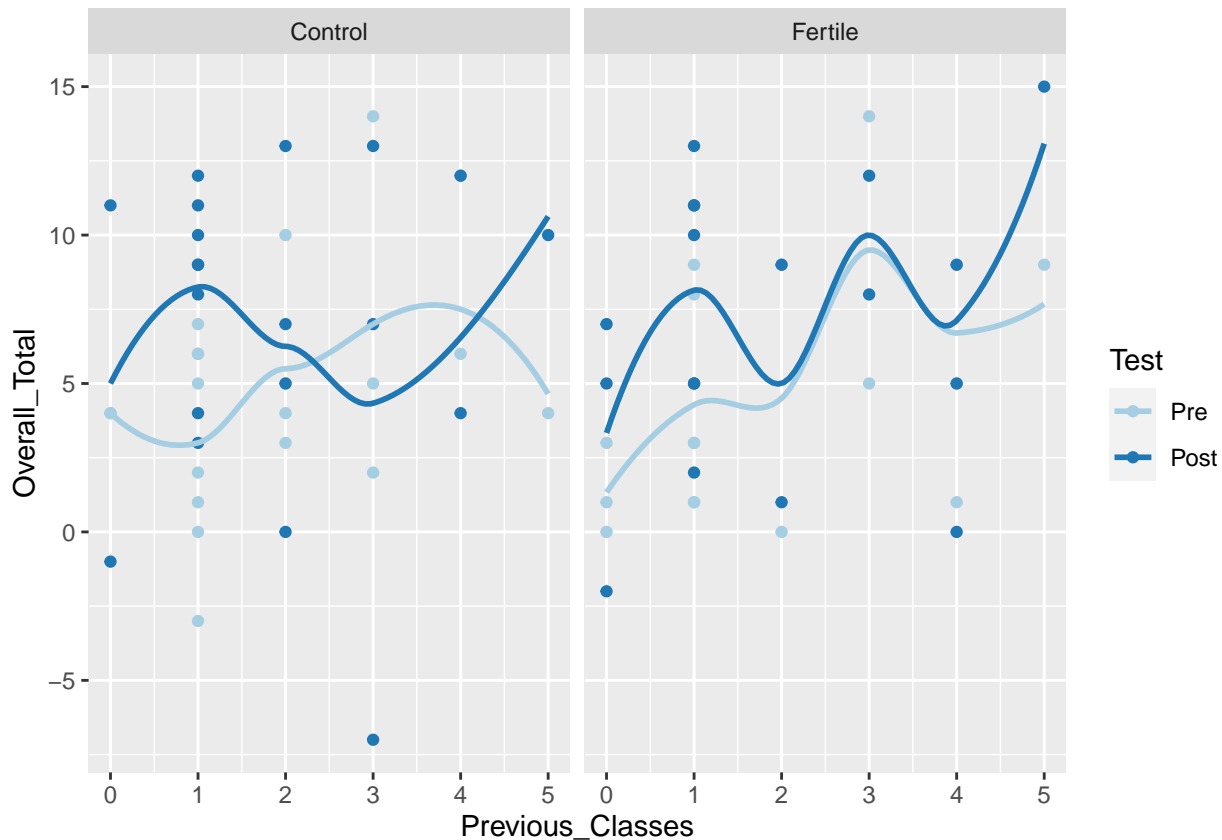
```
Warning in simpleLoess(y, x, w, span, degree = degree, parametric =
parametric, : neighborhood radius 1
```

```
Warning in simpleLoess(y, x, w, span, degree = degree, parametric =
parametric, : reciprocal condition number 0
```

```
Warning in simpleLoess(y, x, w, span, degree = degree, parametric =
parametric, : pseudoinverse used at 2
```

```
Warning in simpleLoess(y, x, w, span, degree = degree, parametric =
parametric, : neighborhood radius 1
```

```
Warning in simpleLoess(y, x, w, span, degree = degree, parametric =
parametric, : reciprocal condition number 0
```



Although, visually, it does not seem as though *fertile* provided a statistically significant benefit (or change at all) in student knowledge of reproducibility across the semester, statistical tests are necessary to solidify the analysis.

After constructing a simple regression model, predicting overall `Score_Difference` based on `Group` (*fertile* or *control*), and running a t-test and analysis of variance, we once again find that there is not enough evidence to argue that *fertile* provides any benefit in terms of performance on the reproducibility tests. This can be seen with the incredibly high p-values of over 0.96, indicating an approximately 96% probability that the slight positive relationship between being in the *fertile* group and the difference in reproducibility test scores across the semester was just due to chance.

Call:

```
lm(formula = Score_Difference ~ Group, data = jterm_model_data)
```

Residuals:

| Min | 1Q | Median | 3Q | Max |
|---------|--------|--------|-------|-------|
| -14.150 | -2.918 | 0.850 | 3.546 | 8.850 |

Coefficients:

| | Estimate | Std. Error | t value | Pr(> t) |
|--------------|----------|------------|---------|----------|
| (Intercept) | 2.15000 | 1.10658 | 1.943 | 0.0599 . |
| GroupFertile | 0.07222 | 1.60783 | 0.045 | 0.9644 |

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 4.949 on 36 degrees of freedom

Multiple R-squared: 5.604e-05, Adjusted R-squared: -0.02772

F-statistic: 0.002018 on 1 and 36 DF, p-value: 0.9644

Analysis of Variance Table

Response: Score_Difference

| | Df | Sum Sq | Mean Sq | F value | Pr(>F) |
|-----------|----|--------|---------|---------|--------|
| Group | 1 | 0.05 | 0.0494 | 0.002 | 0.9644 |
| Residuals | 36 | 881.66 | 24.4906 | | |

Adding in a second variable, the number of previous related courses taken by the student, we see the same story. Neither the student's experimental group, nor their number of previous classes have a statistically significant relationship with score difference, holding the other variable constant.

Call:

```
lm(formula = Score_Difference ~ Group + Previous_Classes, data = jterm_model_data)
```

Residuals:

| | Min | 1Q | Median | 3Q | Max |
|--|---------|--------|--------|-------|-------|
| | -13.395 | -2.130 | 1.043 | 2.996 | 8.232 |

Coefficients:

| | Estimate | Std. Error | t value | Pr(> t) |
|------------------|----------|------------|---------|----------|
| (Intercept) | 3.45431 | 1.52672 | 2.263 | 0.030 * |
| GroupFertile | 0.06459 | 1.59648 | 0.040 | 0.968 |
| Previous_Classes | -0.68648 | 0.55789 | -1.230 | 0.227 |

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 4.914 on 35 degrees of freedom

Multiple R-squared: 0.04152, Adjusted R-squared: -0.01325

F-statistic: 0.7581 on 2 and 35 DF, p-value: 0.4761

3.2.4 Limitations

Although the findings of the *fertile* experiment are interesting, they come with many caveats and should be treated with caution.

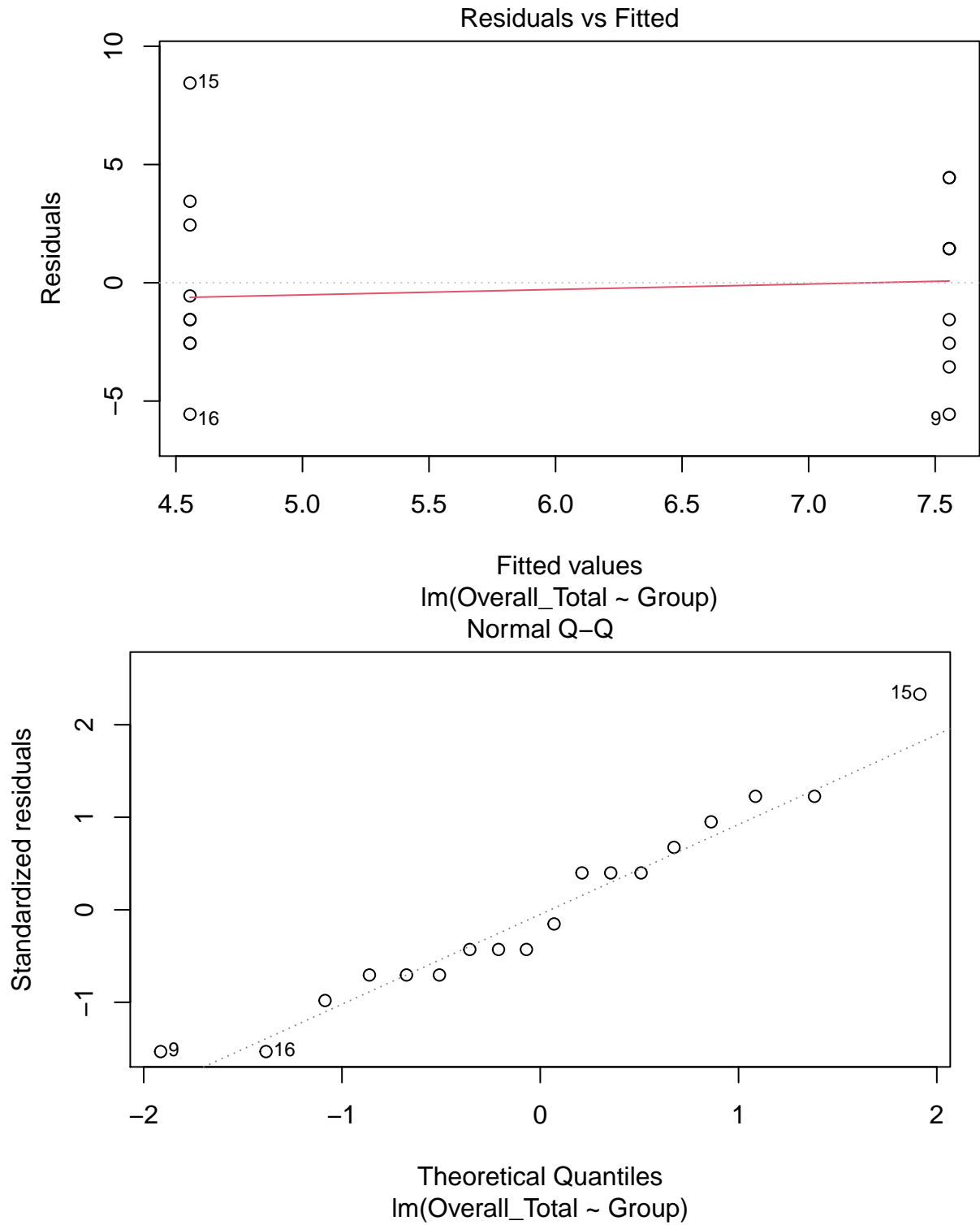
For one, the sample sizes of both experimental sections were very small. The first only contained data on 18 students, while the second on only 38. While sample sizes like these may provide some evidence of the existence of experimental relationships, conclusive statements should not be made without additional data.

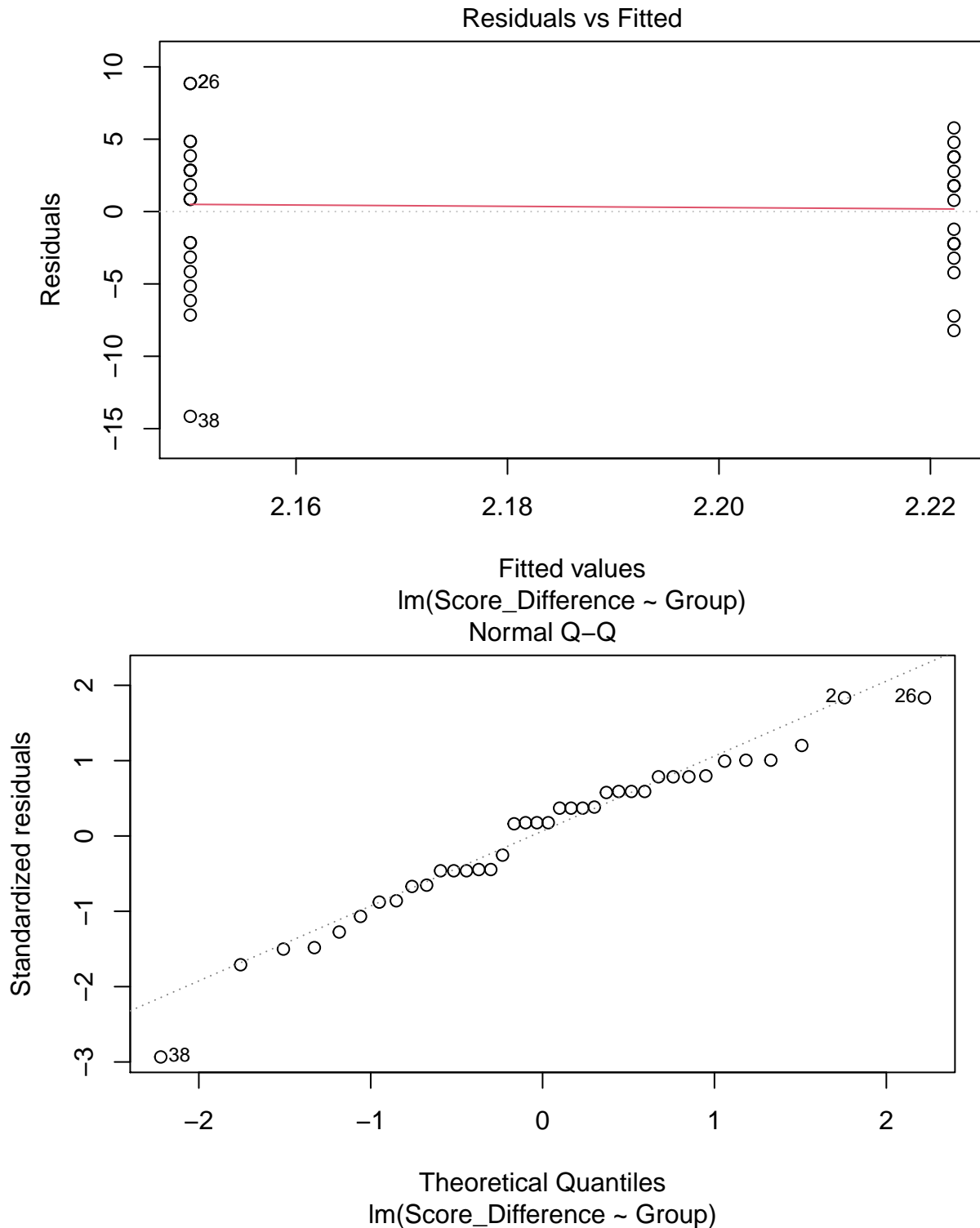
Second, the fact that the environment for the experiment was an active classroom came with several challenges. Although we wanted students in the experimental group to independently learn from *fertile* the classroom setting meant that students who had questions about some of the errors that appeared as a result of having *fertile* loaded would sometimes bring them to Professor Baumer. As the course instructor, he would then be impelled to help them, explaining concepts and clarifying the meaning of any errors, at the expense of any potential effect on the experimental results.

Additionally, students in a classroom have the unfortunate characteristic—in experimental terms—of not being fully independent from one another. Students regularly communicated with each other, asking and answering questions, on a course Slack channel and also worked in groups on many of the class projects. This meant that they could spread knowledge; experiences from the *fertile* group could be disseminated to the *control* group and vice versa. Any sort of cross-contamination could be a potential threat to the validity of the experimental results, as independence is a key assumption necessary for linear regression.

Although independence is at risk, many of the other assumptions for linear regression appear to be mostly met—with the caveat, of course, that there are not enough data points to make a definitive conclusion.

In the residuals versus fitted values and normal quantile-quantile plots for the Fall 2020 and January Term 2021 data, displayed below in that order, there do not appear to be any trends indicating an egregious lack of adherence to regression conditions.





Finally, there is the additional limitation that, although this experimental structure tests the pure educational power of *fertile*, it does not accurately represent the ways in which *fertile* would be used in practice.

Those who use R packages for their own benefit do so with specific intention and knowledge that they are doing so. In practice, unless attempting to do some sort of

software testing, no one will use a package that they do not know is loaded.

In order to create an effective experimental environment, however, concessions must be made. If **fertile** were tested in its natural use state, where students were aware that they were using it, it would introduce the possibility of significant bias in the results. Students in the **fertile** group, for example, might change their behavior, knowing that their software is designed to improve reproducibility, and pursue extra research into the topic, affecting their test performance.

We do not know, from this data, whether **fertile** has significant benefits in its standard practice. All we can conclude from this study is that we do not have evidence that the package's interactive reproducibility messaging provides learning benefits when used without students' knowledge.

3.2.5 Implications

Although the conclusions from this experiment are not particularly exciting, the implications of the study extend far beyond just this software.

... [Go into idea of software A/B testing]

3.2.6 Future Work

Potential sources:

<https://arxiv.org/abs/1401.3269>
<https://academic.oup.com/isp/article-abstract/17/4/392/2528285>
<https://berkeleysciencereview.com/2014/06/reproducible-collaborative-data-science/>
<https://guides.lib.uw.edu/research/reproducibility/teaching>
<https://escholarship.org/uc/item/90b2f5xh>

Conclusion

In the field of data science, research is considered fully *reproducible* when the requisite code and data files produce identical results when run by another analyst. Though there is no clear consensus on the exact standards necessary to achieve reproducibility, writing on the topic identifies several components of great importance:

1. The basic project components are made accessible to the public.
2. The file structure of project is well-organized.
3. The project is documented well.
4. File paths used in code are not system- or user-dependent.
5. Randomness is accounted for.
6. Code is readable and consistently styled.

Reproducibility is incredibly important to the scientific community, helping ensure the accuracy of the findings of studies and data analyses and simplifying the process of collaboration and knowledge sharing. When all of the files necessary for a study to be run are published simultaneously, it makes it much easier for others to understand the methods and ideas that were used and apply them to other work in similar areas, promoting the process of scientific advancement.

However, even though it has many benefits in the scientific community, reproducibility is currently facing a crisis. The vast majority of researchers across all scientific fields have been unable to reproduce another researcher's results, and many have been unable to reproduce even their own. In some fields, more than half of published articles have failed attempts at reproducibility.

Researchers across the sciences have recognized this problem and taken steps to address it. Data-intensive academic journals have put in place reproducibility guidelines requiring that submitted articles meet at least some of the standards listed above. Software developers have built tools to help data analysts make their projects more reproducible. These include a small library of R packages, which provide benefits to users of the popular statistical language R, as well as several continuous integration tools, which are much more broad in their application. Educators have also taken action on reproducibility, introducing courses and workshops focused on the topic at their universities.

However, many of these solutions have fallen short. Even with reproducibility guidelines, many journals still publish work that is not reproducible. Journal review takes significantly longer and requires many more resources when ensuring that reproducibility guidelines are fully met. Journals often cannot spare the additional time and money, leaving it up to submitting authors to ensure that their articles meet standards. Since the adherence to standards is not often checked, and achieving full reproducibility can sometimes be time consuming and challenging, many authors fail to take this step.

Software solutions have not fared much better. Many of the R specific solutions only address a single component of reproducibility—for example, version control—and leave out the rest, making it challenging for researchers to address their problems all at once. Additionally, a lot of the software is quite complex in its operation, containing barriers to entry that prevent less-experienced R users from trying them. The continuous integration tools do not face the same issues, but have their own problems. Due to their general nature, these tools do not have any way of addressing coding-language-specific reproducibility problems such as managing software dependencies.

Finally, educational attempts to address reproducibility are inaccessible and time consuming to implement. Many of the existing educational options require a great deal of research knowledge, and as a result are only available at the graduate level, leaving undergraduate students with little to no exposure on the subject. Additionally, many current versions of reproducibility education require a lot of time and focus in class. Professors wanting to share the topic of reproducibility with their students must do so while sacrificing a significant portion of time from other important topics.

Due to their many associated shortcomings, none of these solutions appear to have the potential to address the problem of reproducibility on a wider scale. That is where my work comes in. I have worked to develop *fertile*, a package built for R users that is focused on creating optimum conditions for reproducibility for data analysis projects written in R.

The package is designed to possess none of the challenges apparent in other solutions—to serve as a one-stop solution, where users can go to address all of their reproducibility needs at one time. Rather than being time consuming, complicated, inaccessible, or too focused on one issue, *fertile* is:

- 1) Simple, with a small library of functions/tools that are straightforward to use. Some of the package's functionalities, like the interactive file path warning system, require no effort from the user to enable, while most others can be run with only a handful of functions requiring one argument: a path to a project directory.
- 2) *Accessible to a variety of users, with a relatively small learning curve.* Users do not need a background in research, nor do they need an advanced knowledge of R programming, to run the majority of functions in *fertile*.
- 3) *Able to address a wide variety of aspects of reproducibility, rather than just one or two key issues.* *fertile* contains functions to address all six previously

defined reproducibility components, unlike most software which addresses only one or two.

- 4) *Language specific, possessing features that address some of the reproducibility challenges associated with R.* The `has_no_randomness()`, `proj_pkg_script()`, and `proj_dependency_report()` functions handle the R-specific issues of random number generation and package dependencies.
- 5) *Customizable, allowing users to choose for themselves which aspects of reproducibility they want to focus on.* There is no one way that users are forced to use **fertile**. Those who only want to focus on one aspect of reproducibility can do so, while those who want to address everything at once have that option as well.
- 6) *Educational, teaching those that use it about why their projects are not reproducible and how to correct that in the future.* Throughout the reproducibility checking process, **fertile** provides informative messages that explain to users the flaws in their code and show them ways to correct the issues.

Due to its many advantages compared with traditional reproducibility solutions, **fertile** has the potential to provide significant benefits in a variety of domains, including in the areas of journal review and education, where other solutions have not quite met the mark.

In the journal review process, **fertile** can be integrated to help ensure that submitted articles meet reproducibility standards. Since **fertile** is both free and publicly available, authors and journal reviewers could take advantage of it in their reproducibility-checking process. Journals might require that certain **fertile** checks be passed for articles to be approved, submitting authors could ensure that their projects pass on their end, and then all that would be required for journals to ensure reproducibility would be a quick run of **fertile**'s functions. This would greatly reduce the barriers on both the reviewing and authoring side of the issue and would vastly speed up the reproducibility-checking process, making it much easier to ensure that published articles are truly reproducible.

In the classroom, **fertile** could be used to extend reproducibility education to undergraduate students—even those at the introductory level. Due to its simplicity, the package would take minimal time and effort to integrate into courses, allowing professors to teach reproducibility without taking important time away from other key topics. If professors were to integrate **fertile** in to their courses, students would be able begin learning about reproducibility much earlier in their data science careers than would likely happen otherwise, increasing the chance that they prioritize reproducibility in their future work.

Academia and research are not the only applications, however. **fertile** could also assist the general world of data science. R bloggers and Tidy Tuesday participants could use the package to improve the quality of their code- and data- sharing, making it easier for others to learn from their work. Data journalists, in some situations, could do something similar, sharing the (reproducible) code and data behind their projects to remove the black-box nature that some such projects possess, building

a relationship of trust with their readers. And private companies doing consulting work could integrate **fertile** into their workflow as a way of increasing transparency between themselves and their clients.

Appendix A

The First Appendix

This first appendix includes all of the R chunks of code that were hidden throughout the document (using the `include = FALSE` chunk tag) to help with readability and/or setup.

In the main Rmd file

```
# This chunk ensures that the thesishdown package is
# installed and loaded. This thesishdown package includes
# the template files for the thesis.
if (!require(remotes)) {
  if (params$`Install needed packages for {thesishdown}`) {
    install.packages("remotes", repos = "https://cran.rstudio.com")
  } else {
    stop(
      paste('You need to run install.packages("remotes")',
            "first in the Console.")
    )
  }
}
if (!require(thesishdown)) {
  if (params$`Install needed packages for {thesishdown}`) {
    remotes::install_github("ismayc/thesishdown")
  } else {
    stop(
      paste(
        "You need to run",
        'remotes::install_github("ismayc/thesishdown")',
        "first in the Console."
      )
    )
  }
}
library(thesishdown)
```

```
# Set how wide the R output will go
```

Appendix B

The Second Appendix, for Fun

References

- American Economic Association. (2020). Data and code availability policy. Retrieved from <https://www.aeaweb.org/journals/data/data-code-policy>
- American Journal of Political Science. (2016, May). Guidelines for preparing replication files. Retrieved from https://ajps.org/wp-content/uploads/2018/05/ajps_replication-guidelines-2-1.pdf
- American Statistical Association. (2020). JASA acs reproducibility guide. Retrieved from <https://jasa-acsgithub.github.io/repro-guide/pages/author-guidelines>
- Baker, M. (2015). Over half of psychological studies fail reproducibility test. *Nature*. Retrieved from <https://www.nature.com/news/over-half-of-psychology-studies-fail-reproducibility-test-1.18248>
- Baker, M. (2016). 1,500 scientists lift the lid on reproducibility. *Nature*. Retrieved from <https://www.nature.com/news/1-500-scientists-lift-the-lid-on-reproducibility-1.19970>
- Baumer, B., Cetinkaya-Rundel, M., Bray, A., Loi, L., & Horton, N. J. (2014). R markdown: Integrating a reproducible analysis tool into introductory statistics. *arXiv Preprint arXiv:1402.1894*.
- Begley, C. G., & Ellis, L. M. (2012). Raise standards for preclinical cancer research. *Nature*, 483(7391), 531–533.
- Biostatistics. (2020). Information for authors. Retrieved from https://academic.oup.com/biostatistics/pages/General_Instructions
- Blischak, J., Carbonetto, P., & Stephens, M. (2019). Workflowr: A framework for reproducible and collaborative data science. Retrieved from <https://CRAN.R-project.org/package=workflowr>
- Bollen, K., Cacioppo, J. T., Kaplan, R. M., Krosnick, J. A., Olds, J. L., & Dean, H. (2015). Report of the subcommittee on replicability in science advisory committee to the nsf sbe directorate. Retrieved from https://www.nsf.gov/sbe/SBE_Spring_2015_AC_Meeting_Presentations/Bollen_Report_on_Replicability_SubcommitteeMay_2015.pdf
- Broman, K. (2019). Initial steps toward reproducible research: Organize your data and

- code. *Sitewide ATOM*. Retrieved from <https://kbroman.org/steps2rr/pages/organize.html>
- Cambridge University Press. (2020). Experimental results - transparency and openness policy. Retrieved from <https://www.cambridge.org/core/journals/experimental-results/information/transparency-and-openness-policy>
- Claerbout, J. F., & Karrenbach, M. (1992). Electronic documents give reproducible research a new meaning. In *SEG technical program expanded abstracts 1992* (pp. 601–604). Society of Exploration Geophysicists.
- Cooper, N., Hsing, P.-Y., Croucher, M., Graham, L., James, T., Krystalli, A., & Michonneau, F. (2017). A guide to reproducible code in ecology and evolution. *British Ecological Society*. Retrieved from <https://www.britishecologicalsociety.org/wp-content/uploads/2017/12/guide-to-reproducible-code.pdf>
- Eisner, D. A. (2018). Reproducibility of science: Fraud, impact factors and carelessness. *Journal of Molecular and Cellular Cardiology*, 114, 364–368. <http://doi.org/https://doi.org/10.1016/j.yjmcc.2017.10.009>
- Epstein, M. L., Lazarus, A. D., Calvano, T. B., Matthews, K. A., Hendel, R. A., Epstein, B. B., & Brosvic, G. M. (2002). Immediate feedback assessment technique promotes learning and corrects inaccurate first responses. *The Psychological Record*, 52(2), 187–201.
- Fidler, F., & Wilcox, J. (2018). Reproducibility of scientific results. In E. N. Zalta (Ed.), *The stanford encyclopedia of philosophy* (Winter 2018). <https://plato.stanford.edu/archives/win2018/entries/scientific-reproducibility/>; Metaphysics Research Lab, Stanford University.
- FitzJohn, R., Ashton, R., Hill, A., Eden, M., Hinsley, W., Russell, E., & Thompson, J. (2020). Orderly: Lightweight reproducible reporting. Retrieved from <https://CRAN.R-project.org/package=orderly>
- Gancarz, M. (2003). *Linux and the unix philosophy* (2nd ed.). Woburn, MA: Digital Press.
- Goodman, S. N., Fanelli, D., & Ioannidis, J. P. A. (2016). What does research reproducibility mean? *Science Translational Medicine*, 8(341), 1–6. <http://doi.org/10.1126/scitranslmed.aaf5027>
- Gosselin, R.-D. (2020). Statistical analysis must improve to address the reproducibility crisis: The access to transparent statistics (acts) call to action. *BioEssays*, 42(1), 1900189. <http://doi.org/10.1002/bies.201900189>
- Hardwicke, T. E., Mathur, M. B., MacDonald, K., Nilsonne, G., Banks, G. C., Kidwell, M. C., ... others. (2018). Data availability, reusability, and analytic reproducibility: Evaluating the impact of a mandatory open data policy at the journal cognition. *Royal Society Open Science*, 5(8), 180448. Retrieved from

- <https://royalsocietypublishing.org/doi/full/10.1098/rsos.180448>
- Henry, L., & Wickham, H. (2020). Tidysselect: Select from a set of strings. Retrieved from <https://CRAN.R-project.org/package=tidysselect>
- Hermans, F., & Aldewereld, M. (2017). Programming is writing is programming. In *Companion to the first international conference on the art, science and engineering of programming* (pp. 1–8).
- Hillenbrand, S. (2014). Reproducible and collaborative: Teaching the data science life. Berkeley Science Review. Retrieved from <https://berkeleysciencereview.com/2014/06/reproducible-collaborative-data-science/>
- Horton, N. J., Baumer, B. S., & Wickham, H. (2014). Teaching precursors to data science in introductory and second courses in statistics. *arXiv Preprint arXiv:1401.3269*.
- Hrynaskiewicz, I. (2020). Publishers' responsibilities in promoting data quality and reproducibility. *Handbook of Experimental Pharmacology*, 257, 319–348. http://doi.org/https://doi.org/10.1007/164_2019_290
- Jacoby, W. G., Lafferty-Hess, S., & Christian, T.-M. (2017). Should journals be responsible for reproducibility? Inside Higher Ed. Retrieved from <https://www.insidehighered.com/blogs/rethinking-research/should-journals-be-responsible-reproducibility>
- Janz, N. (2016). Bringing the gold standard into the classroom: Replication in university teaching. *International Studies Perspectives*, 17(4), 392–407.
- Journal of Computational and Graphical Statistics. (2020). Instructions for authors. Retrieved from <https://www.tandfonline.com/action/authorSubmission?show=instructions&journalCode=ucgs20>
- Journal of Statistical Software. (2020). Instructions for authors. Retrieved from <https://www.jstatsoft.org/pages/view/authors#review-process>.
- Journal of the American Statistical Association Editors. (2020). JASA editors talk reproducibility. Retrieved from <https://www.amstat.org/ASA/Publications/Q-and-As/JASA-Editors-Talk-Reproducibility.aspx>
- Kitzes, J., Turek, D., & Deniz, F. (2017). *The practice of reproducible research: Case studies and lessons from the data-intensive sciences*. Berkeley, CA: University of California Press. Retrieved from <https://www.practicereproducibleresearch.org>
- Leopold, S. S. (2015). Editorial: Increased manuscript submissions prompt journals to make hard choices. *Clinical Orthopaedics and Related Research*, 473(3), 753–755. <http://doi.org/10.1007/s11999-014-4129-1>
- Lupia, A., & Elman, C. (2014). Openness in political science: Data access and research

- transparency. *PS, Political Science & Politics*, 47(1), 19.
- Martinez, C., Hollister, J., Marwick, B., Szöcs, E., Zeitlin, S., Kinoshita, B. P., ... Meinke, B. (2018). Reproducibility in Science: A Guide to enhancing reproducibility in scientific results and writing. Retrieved from <http://ropensci.github.io/reproducibility-guide/>
- Marwick, B. (2019). Rrtools: Creates a reproducible research compendium. Retrieved from <https://github.com/benmarwick/rrtools>
- Marwick, B., Boettiger, C., & Mullen, L. (2018). Packaging data analytical work reproducibly using R (and friends). *The American Statistician*, 72(1), 80–88. <http://doi.org/doi.org/10.1080/00031305.2017.1375986>
- McArthur, S. L. (2019). Repeatability, reproducibility, and replicability: Tackling the 3R challenge in biointerface science and engineering. *Biointerphases*, 14(2), 1–2. <http://doi.org/10.1116/1.5093621>
- McIntire, E. J. B., & Chubaty, A. M. (2020). Reproducible: A set of tools that enhance reproducibility beyond package management. Retrieved from <https://CRAN.R-project.org/package=reproducible>
- National Institutes of Health. (2014). Principles and guidelines for reporting preclinical research. Retrieved from <https://www.nih.gov/research-training/rigor-reproducibility/principles-guidelines-reporting-preclinical-research>
- OpenSci, R. (2020). Drake: A pipeline toolkit for reproducible computation at scale. Retrieved from <https://cran.r-project.org/package=drake>
- Oracle Corporation. (2019). Wercker. Retrieved from <https://github.com/wercker/wercker>
- R-Core-Team. (2020). Writing r extensions. *R Foundation for Statistical Computing*. Retrieved from <http://cran.stat.unipd.it/doc/manuals/r-release/R-exts.pdf>
- R Journal Editors. (2020). Instructions for authors. Retrieved from <https://journal.r-project.org/share/author-guide.pdf>
- Ross, N., DeCicco, L., & Randhawa, N. (2018). Checkers: Automated checking of best practices for research compendia. Retrieved from <https://github.com/ropenscilabs/checkers/blob/master/DESCRIPTIONr>
- Stodden, V., Seiler, J., & Ma, Z. (2018a). An empirical analysis of journal policy effectiveness for computational reproducibility. *Proceedings of the National Academy of Sciences*, 115(11), 2584–2589. Retrieved from <https://www.pnas.org/content/115/11/2584>
- Stodden, V., Seiler, J., & Ma, Z. (2018b). An empirical analysis of journal policy

- effectiveness for computational reproducibility. *Proceedings of the National Academy of Sciences*, 115(11), 2584–2589. <http://doi.org/10.1073/pnas.1708290115>
- The American Statistician. (2020). Instructions for authors. Retrieved from <https://www.tandfonline.com/action/authorSubmission?show=instructions&journalCode=utas20>
- Ushey, K., & RStudio. (2020). Renv: Project environments. Retrieved from <https://cran.r-project.org/web/packages/renv/index.html>
- Wallach, J. D., Boyack, K. W., & Ioannidis, J. P. A. (2018). Reproducible research practices, transparency, and open access data in the biomedical literature, 2015-2017. *PLOS Biology*, 16(11), 1–20. <http://doi.org/10.1371/journal.pbio.2006930>
- Wickham, H. (2015). *R packages* (1st ed.). Sebastopol, CA: O'Reilly Media, Inc.
- Wilson, G., Aruliah, D. A., Brown, C. T., Hong, N. P. C., Davis, M., Guy, R. T., ... others. (2014). Best practices for scientific computing. *PLOS Biology*, 12(1), e1001745.
- Wilson, G., Bryan, J., Cranston, K., Kitzes, J., Nederbragt, L., & Teal, T. K. (2017). Good enough practices in scientific computing. *PLOS Biology*, 13(6), e1005510. Retrieved from <https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1005510>
- Woolston, C. (2020). TOP factor rates journals on transparency, openness. Nature Index. Retrieved from <https://www.natureindex.com/news-blog/top-factor-rates-journals-on-transparency-openness>
- Yu, B., & Hu, X. (2019). Toward training and assessing reproducible data analysis in data science education. *Data Intelligence*, 1(4), 381–392.