

FULL ARTICLE

Two optical coherence tomography systems detect topical gold nanoshells in hair follicles, sweat ducts and measure epidermis

Mette Mogensen¹  | Sophie Bojesen^{1*}  | Niels M. Israelsen² | Michael Maria² | Mikkel Jensen² | Adrian Podoleanu³ | Ole Bang^{2,4} | Merete Haedersdal¹

¹Department of Dermatology, Bispebjerg Hospital, University of Copenhagen, Copenhagen, Denmark

²DTU Fotonik, Department of Photonics Engineering, Technical University of Denmark, Kongens Lyngby, Denmark

³School of Physical Sciences, Ingram Building, University of Kent, Canterbury, UK

⁴NKT Photonics A/S, Birkerød, Denmark

***Correspondence**

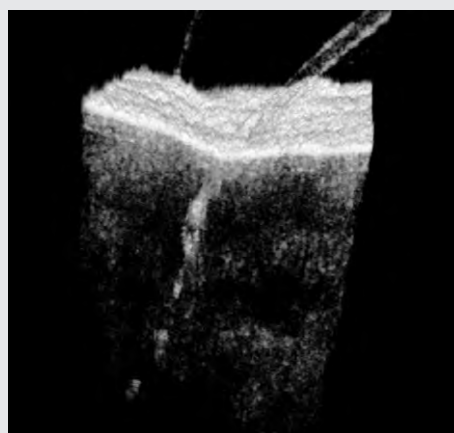
Sophie Bojesen, Department of Dermatology, Bispebjerg Hospital, University of Copenhagen, Nielsine Nielsens Vej 9, DK-2400 Copenhagen NV, Denmark.

Email: sophiebojesen@gmail.com

Funding information

European Union's Horizon 2020 grant GALAHAD, Grant/Award Numbers: 732613, 732613; Innovation Fund Denmark, ShapeOCT, Grant/Award Numbers: No. 4107-00011A, grant No. 4107-00011A, grant No. 4107-00011A, grant No. 4107-00011A, grant No. 4107-00011A, grant No. 4107-00011A; NIHR Biomedical Research Centre (BRC) at Moorfields Eye Hospital NHS Foundation Trust, UCL Institute of Ophthalmology; Royal Society Wolfson Research Merit Award; UBAPHODESA Marie Curie European Industrial Doctorate

Optical coherence tomography (OCT) is an established imaging technology for in vivo skin investigation. Topical application of gold nanoshells (GNS) provides contrast enhancement in OCT by generating a strong hyperreflective signal from hair follicles and sweat glands, which are the natural skin openings. This study explores the utility of 150 nm diameter GNS as contrast agent for OCT imaging. GNS was massaged into skin and examined in four skin areas of 11 healthy volunteers. A commercial OCT system and a prototype with 3 μ m resolution (UHR-OCT) were employed to detect potential benefits of increased resolution and variability in intensity generated by the GNS. In both OCT-systems GNS enhanced contrast from hair follicles and sweat ducts. Highest average penetration depth of GNS was in armpit $0.64 \text{ mm} \pm \text{SD } 0.17$, maximum penetration depth was 1.20 mm in hair follicles and 15 to $40 \mu\text{m}$ in sweat ducts. Pixel intensity generated from GNS in hair follicles was significantly higher in UHR-OCT images ($P = .002$) and epidermal thickness significantly lower 0.14 vs 0.16 mm ($P = .027$). This study suggests that GNSs are interesting candidates for increasing sensitivity in OCT diagnosis of hair and sweat gland disorders and demonstrates that choice of OCT systems influences results.



KEYWORDS

Au, contrast agent, contrast enhancement, epidermal thickness, hair follicle, nanoshell, optical coherence tomography, sweat duct

1 | INTRODUCTION

Optical coherence tomography (OCT) is an optical imaging technology that enables real-time, high-resolution, cross-sectional and *en face* investigation of skin by detecting reflected broad-spectrum near-infrared light from tissue. OCT provides micron-scale spatial resolution and millimeter-scale

depth of penetration [1]. Several commercial OCT systems with handheld probes targeted for Dermatology are now available [2].

The ability of OCT to achieve high diagnostic accuracy in skin diseases is hampered by the fact that not all diseases show sufficient contrast to be discriminated from normal skin. The challenge in realizing contrast enhancement in

OCT imaging is to achieve signal from exogenous contrast agents that can overcome the intrinsic signal of the skin itself, allowing segregation of normal skin structures from skin pathology [3]. Our clinical experience, supported by animal studies [4, 5], demonstrates that application of a contrast agent, such as gold nanoshells (GNSs), leads to a strong hyperreflective signal in OCT images deriving from the natural skin openings. This property of GNS can be used to generate enhanced contrast around hair follicles and sweat glands and potentially improve the diagnostic accuracy in some skin diseases.

The adnexal structures of skin consist of hairs, hair follicles, sebaceous glands, eccrine glands and sweat ducts delivering the sweat to the skin surface. The hair follicles are pathways from the upper skin layers to lower skin layers [6, 7] and therefore contrast agents can be delivered through the pilous route.

The diagnostic accuracy of OCT in Dermatology has been demonstrated as moderate to high [8, 9]. Several skin diseases have been studied clinically with OCT [10–16] including disorders involving adnexal structures: sweating [17], dyshidrotic skin conditions [18], follicular keratosis [19], abnormal hair growth [20], hair transplantation [21] and acne [22–24]. Identification and measurement of hair follicle calibers, size/condition of hair follicles and sweat gland structures can be useful in monitoring these diseases. The efficacy of OCT has already been established for investigation and measurement of wounds [25], hairs and nails [10, 26].

Since OCT systems can be quite different in regards to light source, focus point and data processing, the resolution and penetration depth vary among different systems. For example, data on thickness of keratinocyte carcinomas do not seem to correlate between OCT systems when compared clinically [27]. Several studies have compared different OCT systems used in dermatology. One study compared OCT imaging of actinic keratosis and basal cell carcinoma (BCC) in a clinical setting of 29 patients using 3 different OCT systems: VivoSight, by Michelson Diagnostics Ltd, Maidstone, UK; Callisto by Thorlabs Inc, and Skintell by Agfa Healthcare NV, Belgium. A correlation between the first 2 devices regarding BCC tumor thickness was established; however, the BCC thickness did not correlate to histology in either. The latter OCT system Skintell showed no correlation with VivoSight and Callisto. Reassuringly, all OCT systems differentiated skin cancer from normal skin [27]. However, the discrepancy highlights the need to compare OCT systems head-to-head clinically.

The natural first steps of introducing topical GNS in the dermatological clinic is to describe their distribution in normal skin and in adnexal structures, imaged by more than 1 OCT system and subsequently to apply GNS to diagnostic studies of skin disease.

Aim of study: To describe and explore the utility of GNS as contrast agent for OCT imaging in normal healthy human skin with special regard to distribution in hair follicles and sweat ducts. Four anatomical areas of the skin were scanned: cheek, palm of the hand, armpit and chest. To assess the versatility of GNS and potential differences in OCT signal and OCT intensity, the healthy volunteers were examined by 2 different OCT systems: A commercial VivoSight OCT (C-OCT) that has a 5 to 7.5 μm resolution (in tissue) and a prototype ultrahigh OCT (UHR-OCT) system with a 2.2 to 6 μm resolution (in tissue). Furthermore, we compared the epidermal thickness (ET) measurements performed using the 2 OCT systems.

2 | METHODS AND MATERIALS

2.1 | Study design

A clinical prospective study, consecutively including healthy volunteers from the Department of Dermatology at Bispebjerg Hospital, University of Copenhagen, Denmark was established in accordance with Helsinki II Declarations. The research protocol was approved by the Ethics Committee of The Capital Region of Denmark: no. H-16039077.

2.2 | Volunteers

Healthy volunteers were included from our department, and from social media advertisement, approved by the Ethics Committee. Inclusion criteria were age above 18 and normal healthy skin. Exclusion criteria consisted of pregnancy or lactation, active skin disease or systemic disease requiring ongoing medical therapy. Volunteers were instructed to report any discomfort or skin irritation during and after the study.

2.3 | OCT systems

The commercial OCT system is a VivoSight Dx OCT system (referred to here as C-OCT, Michelson Diagnostics Ltd., Maidstone, UK). The C-OCT is a multi-beam swept source frequency domain system with a tunable diode laser with a peak power of 15 mW at $\lambda = 1305$ nm. Maximal field of image is 6×6 mm. It offers a lateral optical resolution of <7.5 μm and an axial resolution of 5 μm in skin (in tissue). The penetration depth in skin varies around 1 to 2 mm and is limited by scattering effects. For each skin area of 4×4 mm, a cross-sectional multi-slice scan modality consisting of 250 B-scans was set up.

The UHR-OCT is a prototype of a novel OCT system assembled at the Technical University of Denmark and NKT Photonics (Birkerød, Denmark) with technical input from the Applied Optics Group, University of Kent, Canterbury, UK. The system uses a Supercontinuum light source combined with a broadband filter. The combination provides an

output light beam with an average power of 5 mW on the skin and a wavelength range from 1000 to 1500 nm. The system delivers an axial resolution of 2.2 μm and a lateral resolution of 5 to 6 μm (in tissue). A multi-slice scan modality of 1024 B-scans per 3×3 mm was set up for each skin area. Both OCT systems have the size of modern ultrasonography systems and handheld probes; however, the UHR-OCT system probe needs stabilization by both hands of the investigator.

2.4 | GNS contrast agent

GNSs utilized have a 120-nm-diameter silica core and 15-nm-thick gold shell lining coated with polyethylene glycol to prevent particles from aggregating. The polydispersity index is 0.10. These particular 150 nm GNSs were selected because they have been demonstrated to penetrate into hair follicles easily after vibratory massage. The GNSs are certified as medical device CE 612960 and approved for clinical use and topical treatment of acne vulgaris combined with selective photothermolysis [28]. *in vivo* studies have shown that these GNSs are not retained in swine skin 1 month after vibratory massage into the skin and that uptake in other organs is negligible since it is 74 times lower than the established safety threshold [28]. The GNS is supplied in glass vials in a 25% solution in quantities of 1 mL (Sebacia Micro-particles, Sebacia Inc., Duluth, Georgia) suspended in a combination of ethanol (54%), polysorbate 80 (1%) and diisopropyl adipate (20%).

GNSs come in a liquid dark blue solution applied to skin using a dispenser. Skin areas of 5×5 cm were cleansed and 0.50 mL of GNS suspension was massaged into each of 4 skin areas. Two stages of 30 seconds of massage with an oscillating handheld massage device (Sebacia Massager, Sebacia Inc., Duluth, Georgia) was applied to each of 4 test areas.

2.5 | Clinical setting

GNS was applied to cheek, palm of the hand, armpit and the front of the chest on each volunteer. Hairs were removed with a no-touch nontraumatic razor prior to OCT scanning. OCT images were acquired before and after GNS application. After application of GNS, skin was wiped with wet gauze. Volunteers were scanned 2 to 10 minutes after vibrational massage of GNS into the skin. In 1 volunteer, GNS was applied to lower arm and C-OCT images were acquired after 20 minute, 3 and 8 hours. Images of untreated skin and images of skin containing GNS contrast agent were recorded using both OCT systems. GNS penetration depth and ET were assessed and compared in chest and armpit skin only. All 4 skin areas were qualitatively assessed regarding skin structure and localization of GNS in skin.

2.6 | Comparison of pixel values

In order to estimate the intensity of gold particles in C-OCT and UHR-OCT images from chest and armpit in all volunteers, the 8-bit pixel values (256 gray scale steps spanning between black and white) were counted before and after GNS application. In each of the B-scans evaluated, an area of 41×41 pixels was analyzed. The area was chosen so it, for both OCT systems, was small enough to contain only the hair follicle and thereby also excluding the hyperreflective signal of the surface. Histograms (Figure 1E) were built as follows: first, a region of interest (ROI) in representative B-scans (showing hair follicles with and without GNS) was identified by Mette Mogensen (MeM). Next, histograms for each ROI were made after which these were summed in head groups of hair follicles with/without GNS for C-OCT/UHR-OCT.

2.7 | Measurement of ET and penetration depth of GNS

In obtaining the ET, a graphical approach was applied. For both OCT devices, surface recognition was performed visually in OCT images using the integrated VivoSight software module and ImageJ (open source Java image processing program, NIH, 2017 online edition) "Wand tracing tool" for C-OCT and UHR-OCT images, respectively. By visual recognition of the skin surface in OCT images, an identical line was used as an approximate delineation of the dermoepidermal junction (DEJ) visually assessed and positioned by MeM. The axial distance between the surface line and the visually positioned ghost surface tracing DEJ constituted a final measure of the ET. Measurements were performed in the central part of the OCT image.

Penetration depths of GNS were extracted after identification of ROIs assessed by MeM. The numbers on penetration depth describe the largest depth the GNS had traveled in a hair follicle. The GNSs were recognized as a hyperreflective signal similar to the surface signal.

2.8 | Statistics

Descriptive statistics were used and presented as means and medians with minimum and maximum ranges. Absolute and relative frequencies mean and SD for continuous measurements were calculated for each parameter. Statistics are done using MATLAB 2017a (MathWorks 2017 online version, Natick, Massachusetts). Image data were analyzed with Wilcoxon paired test, as number of volunteers were deemed too small for assuming normal distribution. The Wilcoxon paired test was performed on the 8-bit pixel values and the pairs consist of 1 image from each system. The Wilcoxon test was performed with MATLAB *signrank*. Multiple regression analysis was performed on ET and GNS penetration depth data. A *P* value $< .05$ was considered statistically significant. Bland-Altman plots were applied to detect a

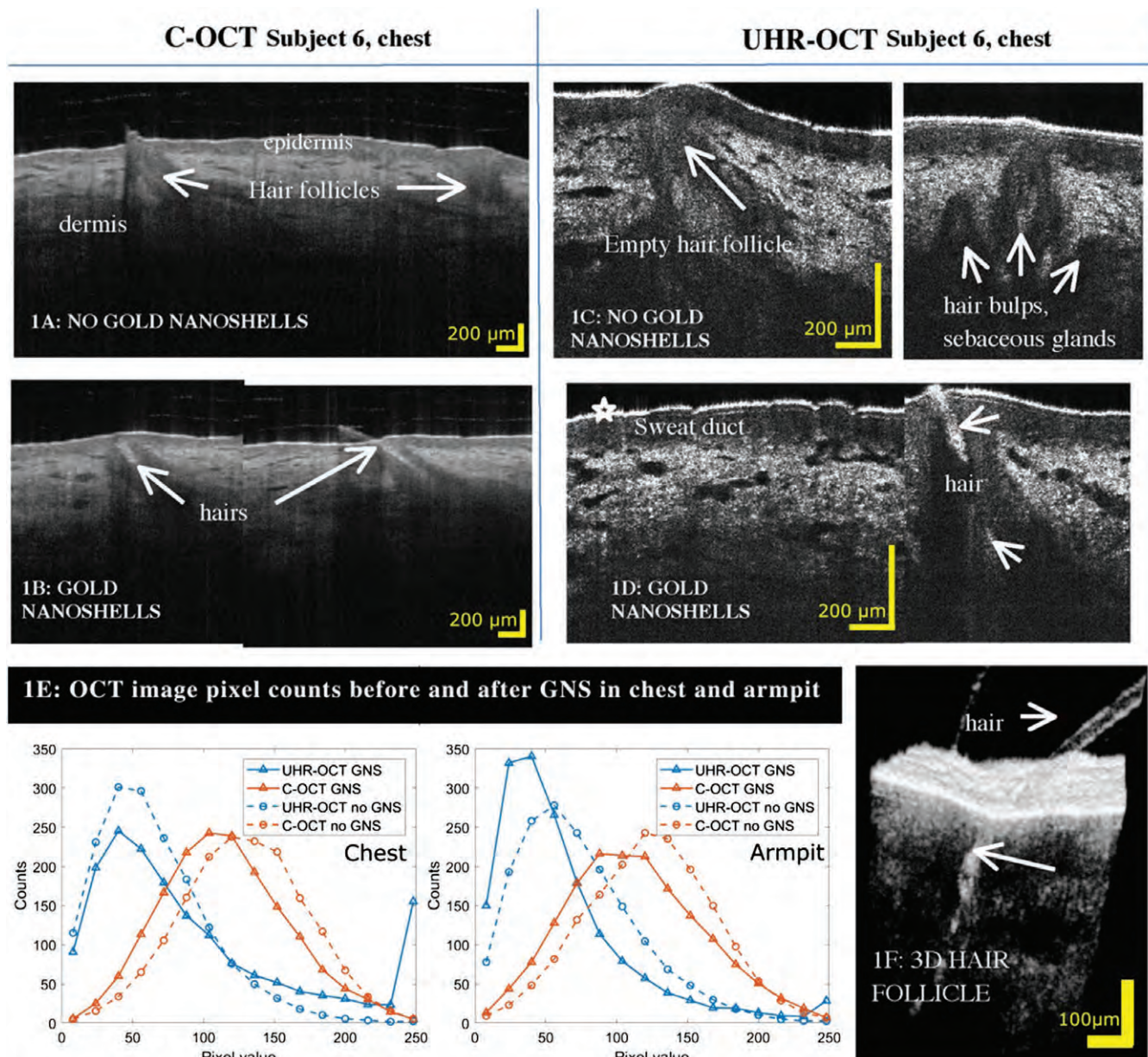


FIGURE 1 Two OCT systems: skin, hairs and pixel values in a hair follicle with and without 150 nm GNSs. OCT images from C-OCT and UHR-OCT. Images captured from chest skin of volunteers 6. The hair follicle without contrast agent is an oblique dark-gray elongated structure penetrating epidermis from dermis. The hair straw is not visible without contrast agent. After application of GNS, the hair straw is enhanced and appears as a bright slender structure inside the hair follicle. In UHR-OCT, the sweat ducts are also vaguely visualized as a grainy white speckle inside epidermis (white asterisk), epidermis is more homogenous dark without contrast agent. A 3D image of a hair on the cheek from the same volunteers show affinity of the gold to the hair straw outside the skin and the gold lining the hair follicle inside the skin. Histograms of pixel values in chest and armpit skin: Without GNS (dashed circles), the histograms for the 2 systems are similar. Application of GNS increased pixel intensity significantly, especially around hair follicles and with a higher intensity in the UHR-OCT system compared to C-OCT. No increase in the highest pixel values was detected in C-OCT after application of GNS, but rather a shift of the statistics toward lower pixel values (1E). This inclination indicates that a normalization of the pixel value distribution is performed according to the absolute signal values detected

potential systematic difference between OCT measurements of ET.

3 | RESULTS

A total of 11 volunteers were enrolled: 64% females, mean age of 37.6 years with a median Fitzpatrick skin type of 2 (Table 1). GNS did not irritate the skin nor leave

permanent discoloration and was easily removed from skin with lukewarm tap water.

3.1 | OCT images before and after application of GNSs

Before GNS: As seen in Figure 1A,C, both OCT systems depict the distinct layering of epidermis, dermis and hairs. In OCT images, epidermis is represented by an upper dark gray band, dermis below is lighter and less compact. Blood

TABLE 1 Data from healthy volunteers

Healthy volunteer no.	Age	Fitz-Patrick skin type	Sex (female/male)	Penetration depth GNSs, chest (mm)	Penetration depth GNSs, armpit (mm)	ET, chest. UHR-OCT (mm)	ET, chest. C-OCT (mm)	ET, armpit UHR-OCT (mm)	ET, armpit C-OCT (mm)
1	33	3	f	0.66	0.60	0.09	0.10	0.12	0.17
2	30	2	m	0.60	0.51	0.08	0.10	0.13	0.16
3	45	3	f	0.38	0.76	0.08	0.10	0.12	0.17
4	27	2	f	0.45	0.52	0.07	0.11	0.14	0.12
5	43	2	f	0.55	0.71	0.08	0.10	0.13	0.12
6	30	2	m	0.68	0.64	0.10	0.13	0.13	0.19
7	72	2	f	0.79	0.94	0.09	0.11	0.13	0.16
8	51	2	f	0.30	0.53	0.11	0.15	0.19	0.17
9	28	2	m	0.41	0.60	0.08	0.11	0.15	0.16
10	28	3	m	0.67	0.70	0.09	0.11	0.17	0.17
11	27	2	f	0.82	0.49	0.09	0.08	0.16	0.22
Results: mean and (95% CI)	Mean = 37.6	Median = 2	Female = 64%	0.57 SD 0.17	0.64 SD 0.14	0.09 mm (0.07, 0.11)	0.11 mm (0.07, 0.15)	0.14 mm (0.10, 0.18)	0.16 mm (0.11, 0.22)
P values						Chest	0.039	Armpit	0.027

Data from all 11 volunteers included. Penetration depth of GNS into skin is in bold. Descriptive statistics of ET included all 11 volunteers, but volunteer 1 was excluded from the measurement of GNS penetration depth as she had 2 mL of gold contrast agent applied to skin instead of 0.5 mL.

vessels are identified as black oval to round structures, in C-OCT images, blood vessels can be discriminated from lymphatic vessels by means of the integrated speckle variance OCT software (Dynamic OCT). Lymphatic vessels do not exhibit the high flow found in blood vessels and do not display a dynamic OCT signal. Hair follicles are identified as an oblique to vertical dark slender structure spanning through epidermis. Sometimes a hair is easily identified inside. In terminal hairs, such as beard hair in men and coarse hair in women, the bulge region is too deep to be imaged by OCT. The smaller vellus hairs present all over the body except from soles of hand and feet and mucosa are characterized by a dark bulge region in dermis next to sebaceous glands (Figure 1). The sweat ducts from the eccrine glands in the armpit, chest and face are not visible (Figure 2B,D) without GNS contrast agent, whereas sweat ducts are clearly visualized in the palm of the hand as distinct white hyperreflective spiral-shaped structures inside the stratum corneum part of the epidermis (Figure 3D). The eccrine glands themselves cannot be visualized, only the sweat ducts releasing the sweat produced onto the skin surface. ET in axillary skin tends to vary in thickness due to thickening around the terminal hair follicles (Figure 2F). Epidermis is more homogeneously dark without contrast agent in UHR-OCT images (Figure 2B,D,F).

After topical GNS: Dilated serpiginous blood vessels characteristics for armpit skin were detected by both OCT systems and looked the same with and without GNS (Figure 2,A(C)). Application of GNS improved contrast to normal skin in sweat glands of armpit skin and in hair follicles on the chest and armpits (Figure 2E). In general, GNS line the inside of hair follicles and occasionally cover the hair straw, generating an enhanced signal from the opening of the hair follicle (Figures 1B,D,E and 3A,C). Average

penetration depths into hair follicles are 0.57 ± 0.17 mm in chest and 0.64 ± 0.14 mm in armpit skin. Maximum penetration depth was 1.20 mm in hair follicles and only 15 to 40 μ m in the sweat ducts of the axillary epidermis (Figure 1, Table 1). Penetration depth of GNS is deeper in armpit skin hair follicles compared to follicles on the chest (Figure 4A) and is not significantly correlated to age and skin type, but to female sex ($P = .029$).

In axillary skin, the sweat ducts were only visualized after application of gold as tiny white vertical strands in epidermis (Figure 2E). The sweat ducts on the chest were barely visible (Figure 1, asterisk). The hair follicles that contain GNS exhibit various morphologies: bright cap on top or bright highly reflective dots from inside the hair follicle. In Figure 3, C-OCT images from all 4 anatomical regions demonstrate the deposition of gold indicated by white arrows. On the palm, there is a thin bright upper band caused by GNS reflection, as it tends to coat the skin surface and thereby increase the entrance signal. As a consequence, GNS can impair OCT image quality (Figure 3D), if not sufficiently wiped off.

Some typical bright spiral-shaped eccrine ducts of the palm are obscured by dark shadows after gold application (Figure 3).

All hairs are hyperreflective outside the hair follicle (Figure 1), but black hairs are also hyperreflective inside the hair follicle (Figure S1, Supporting Information). There was no difference in intensity or overall skin morphology after topical application and massage of the GNS into lower arm of subject 4 after 3 and 8 hours. Gold was not detected in hair follicles at 8 hours, but at 20 minutes and 3 hours (Figure S2). We did not find any contrast difference at the DEJ 3 hours or more after application of GNS (data not shown).

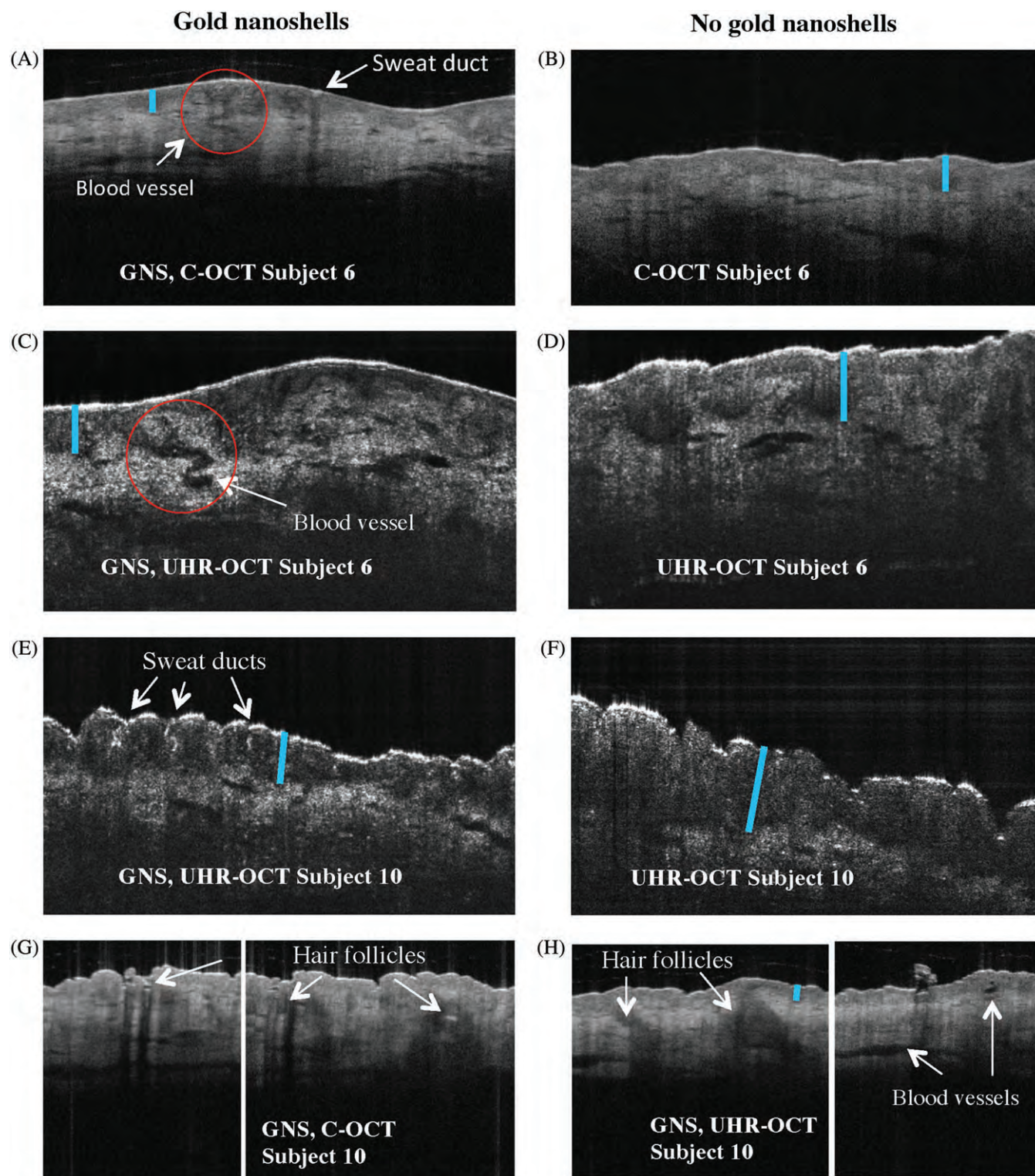


FIGURE 2 OCT images from both systems; armpit of subjects 6 and 10. All ETs are marked by blue bar. (A) C-OCT system demonstrates a dark-gray epidermal band with fluctuating thickness. Dilated serpiginous vessels characteristics for armpit skin marked by rings. No GNS is identified in sweat ducts or in hair follicles. Contrast between epidermis and dermis is slightly enhanced. (B) C-OCT system. No GNSs were applied. GNS is not identified neither in sweat ducts nor in hair follicles. (C) UHR-OCT, similar to (A) serpiginous dilated vessels is marked by a circle. (D) UHR-OCT, no GNS is applied. Contrast between epidermis and dermis is less visible than in (C). (E) UHR-OCT, sweat ducts stand out as thin white hyperreflective vertical strings spanning from top of the skin into epidermis. (F) No increased signal identified neither in sweat ducts nor in hair follicles. (G) C-OCT system. GNS is identified in 3 hair follicles but not in sweat ducts. The hair follicle far right contains a bright grain of GNS identified by arrow. (H) C-OCT system, the black round area is a very superficial blood vessel

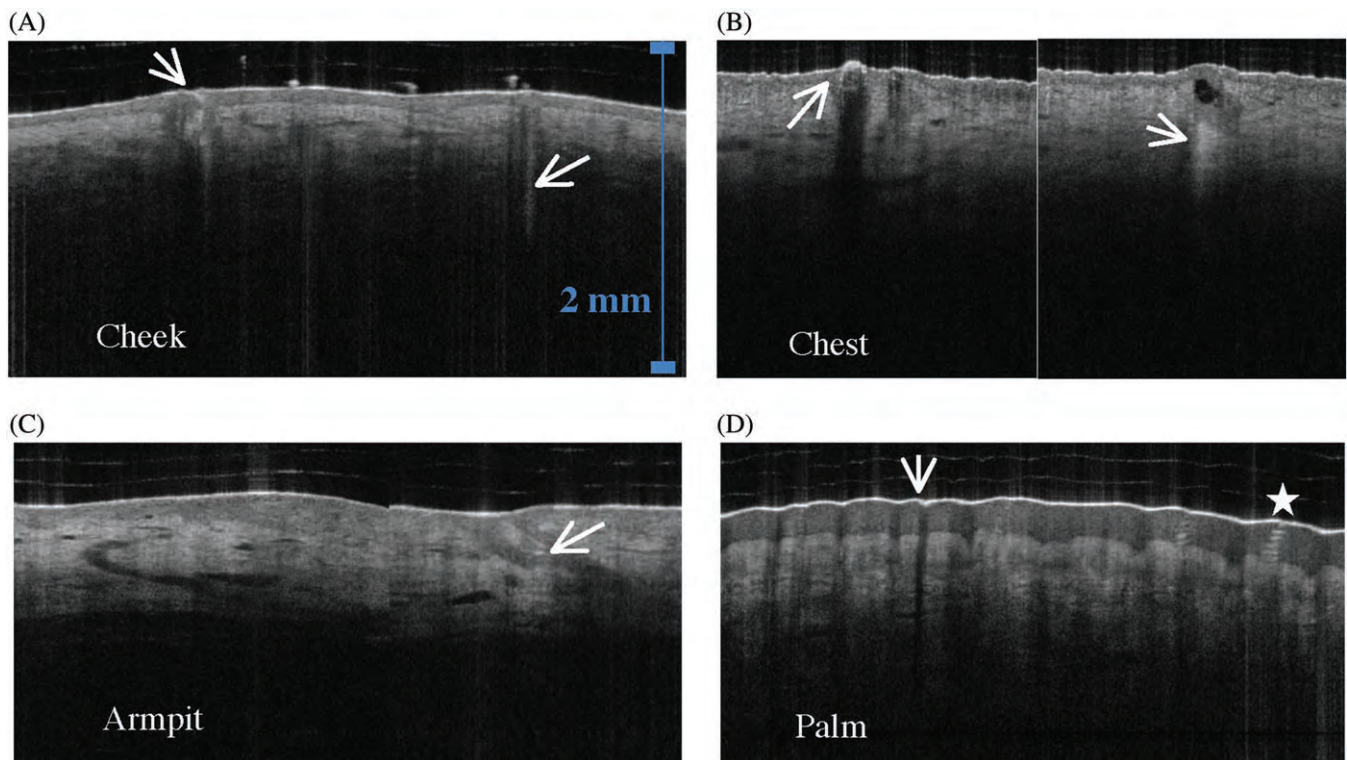


FIGURE 3 C-OCT images of 4 anatomical skin regions in healthy volunteers after topical delivery of the contrast agent: GNS. C-OCT images of anatomical skin regions included in the study after topical delivery of GNS. Deposition of GNS is marked by white arrows: In (A) and (C), the GNSs are deposited inside the hair follicle, in (B), the hair follicles containing GNSs have various morphologies: bright white cap on top of the skin and bright hazy highly refractive signals from inside hair follicle. On the palm, (D), a bright band on top of the skin is caused by surface accumulation of GNS. The typical bright spiral shaped eccrine ducts (white asterisk) are obscured by dark vertical shadows caused by gold deposition in the opening of the sweat duct

3.2 | Epidermal thickness

The ET appeared thinner in UHR-OCT images than in C-OCT images from both armpit and chest skin (Figure 4A). A statistically significant difference between ET measurements performed by the 2 OCT systems was demonstrated (chest $P < .039$ and armpit $P < .027$). Bland-Altman plots were applied to detect a potential systematic difference between ET measurements using UHR-OCT and C-OCT systems. The plots shown in Figure 4B,C demonstrate that ET is slightly thinner in UHR-OCT, but OCT measurements of epidermis correlate and are not skewed in the respect that no OCT system differs more around certain values. ET is not significantly correlated to age, sex or skin type (data not shown).

3.3 | Differences between UHR-OCT and C-OCT

The histograms in Figure 1 show OCT image pixel values of a hair follicle imaged by C-OCT and UHR-OCT with and without gold. Figure 1D shows the histograms for chest skin and Figure 1E shows the histograms for axil skin. Without gold (dashed circles), the histograms look similar. After GNS application, the pixel intensity was higher around hair follicles in UHR-OCT images compared to pixel values in C-OCT images $P = .002$ chest and $P = .002$ armpit and it was further demonstrated that ET was statistically significantly thinner when measured with UHR-OCT (Table 1; Figure 4A).

4 | DISCUSSION

In OCT images, the innate contrast of hair follicle and sweat gland is apparently low in some skin regions such as cheek, chest and armpit. Utilization of topically applied physical contrast-enhancing agents such as GNS may improve sensitivity of OCT skin investigation. Application of GNS as contrast agent increased pixel intensity significantly, especially around hair follicles and with a higher intensity in the UHR-OCT system compared to C-OCT.

Efficacy and limitations of OCT diagnosis of skin disease has previously been established in studies of skin appendages, such as hairs and nails [10, 18–21, 23]. However, various OCT systems with different axial and lateral resolution, different power on the skin and different imaging post-processing software have been applied in these studies. It is therefore imperative to compare commercially available and prototype OCT systems head-to-head, since the data retrieved from different OCT systems are not always comparable; that is, in this study, we find that ET and pixel intensity generated from GNS differs between 2 OCT-systems.

The statistically significant difference between ET measurements from 2 different OCT systems found here is clinically relevant, since many skin diseases are characterized by thickened epidermis. The statistically significant difference between GNS penetration in men and women is ascribed to the regular shaving of armpits performed by all women, except

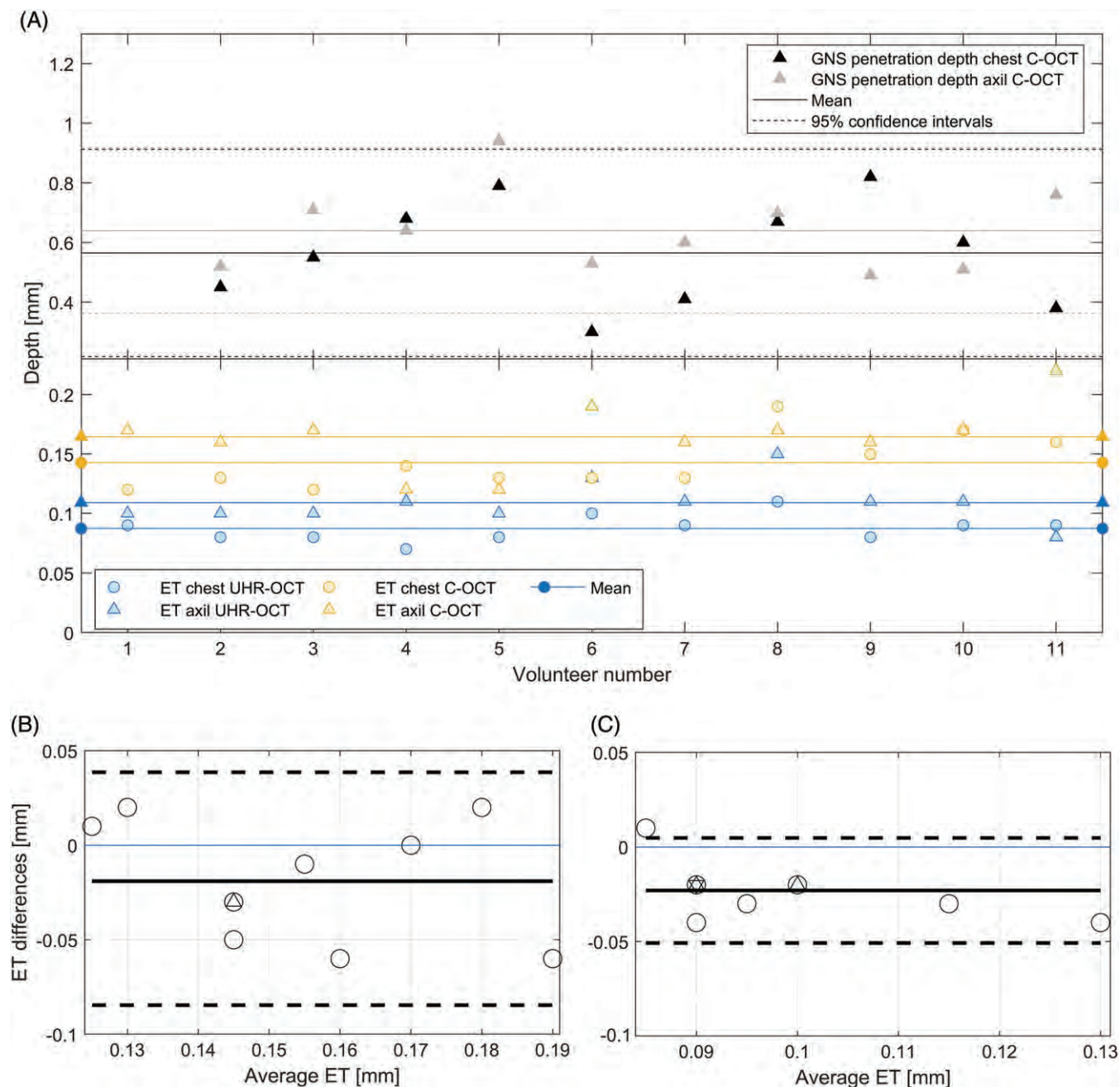


FIGURE 4 ET and penetration depth of GNS in OCT images from armpit and chest. Top: Penetration of GNS into skin measured in OCT images from the armpit and chest of healthy volunteers 2 to 11 (black and gray color). GNS penetrate deeper in armpit, also termed axilla. Bottom: Means of ET (yellow and blue) measured in OCT images from volunteers 1 to 11 in both skin areas. (B) and (C) are Bland-Altman plots demonstrating no systematic skewedness of ET measurements when UHR-OCT is compared to C-OCT in chest and armpit, respectively

one, included in the study. Measured ET values correlated within the 2 systems, but were significantly thicker in C-OCT. We explain these differences as due to a higher resolution of the UHR-OCT system that enhances the features of the DEJ, thereby facilitating the visual segregating of epidermis from dermis. Further, ET was not measured in the exact same area, but over a width of 2 mm by UHR-OCT and 1.5 mm by C-OCT, which explains part of the variation, also axillary ET tends to vary in thickness due to thickening around the terminal hair follicles (Figure 2F). Future OCT studies must take into account that resolution does affect visual segregation of epidermis from dermis and thereby ET measurements. We

specifically looked at chest and armpit OCT images for quantitative analysis because hair follicles stand out well in OCT chest images, and hairs are not as abundant as in the cheek region. The armpit images were chosen due to the abundance of sweat ducts. The palm also has numerous sweat ducts but surprisingly, in this study, GNS did not reflect infrared light inside palmar sweat glands, which is ascribed to a phenomenon caused by the funnel-shaped opening.

This is the first time that topically applied GNS has been investigated with OCT in vivo on human skin, but previous in vivo animal studies have been performed: a rabbit skin study [5] applied silica—GNS with 150 nm silica core size

and 25 nm lining. An increased intensity of the OCT signal in areas containing GNS was detected and contrast was enhanced in hair follicles and sweat glands. The effects lasted up to 24 hours. The presence of GNS in skin was confirmed by electron microscopy. Another OCT study performed on pig skin [4] also demonstrated an increased contrast in OCT images after a single application of GNS with 75 nm silica core and 25 nm gold lining. Multiple applications of gold did not increase the contrast. In the referenced animal studies, penetration of GNS into the dermis was observed. We did not detect GNS in dermis and we suggest that the epilation process must have caused epidermal damage in the animal studies, thus allowing GNS to enter dermis. None of the referenced studies reported their hair removal methods. We only detected gold accumulation through natural skin openings, with an affinity to hair follicles. Furthermore, we used a massage device on intact human skin. We did not find any difference in contrast intensity at the DEJ 3 or more hours after application of GNS. Our findings are supported by a review of literature reporting that GNS larger than 100 nm do not penetrate the intact skin barrier [29]. The handheld massage device applied to intact skin in this study thus gently forced GNS into hair follicles and sweat ducts. We could have chosen particles with even smaller diameter than 150 nm, since the pig skin study showed good penetration of 125 nm GNS into deeper layers, but so did the rabbit skin study that used 200 nm diameter. Therefore, in this study, we chose a GNS diameter in between.

An alternative to physical contrast agents would be optical-clearing agents (OCAs) that reduce the scattering of tissue and make tissue more transparent. OCA can either be physical (eg, massage, ultrasound and lasers), chemical (eg, glycerol and chemical peelers) or combined [30]. Some *ex vivo* studies have shown increased OCT signals from dermis after topical glycerol application [31] also combined with ultrasound [32]. Glycerol is one of the most common OCAs applied to skin samples, but its optical-clearing efficacy is not very high during topical treatment of skin *in vivo* due to low penetration. Dermal injection of glycerol has been reported to cause necrosis of skin [30]. This emphasizes how imperative it is to investigate the safety of OCAs and contrast agents *in vivo*.

GNSs are potential nanotechnology-based drug delivery systems that can accommodate 1 or more active drugs of nano-size to be dispersed, absorbed, conjugated or encapsulated and thereby targeted to be released in, for example, skin cancer or specific organs. Several experimental studies have implemented GNS in drug delivery, for example, by attaching GNS to anticancer antibodies [33–35]. The increased intensity of GNS in OCT images makes it a potential candidate for use as a carrier in targeted drug delivery systems monitored by OCT.

Conclusively, GNSs provide more intensity in UHR-OCT images, making it easier to discriminate sweat ducts compared to the C-OCT system. Hair follicles are easily

delineated by GNS in both OCT systems. The drawbacks of the UHR-OCT system were the narrow skin area scanned (2 mm) and the importance of manually stabilizing the probe without being able to see if it moves during scanning. The advantage of the C-OCT also the camera in the probe, which secures an exact scan of the area of interest. This clinical study provides novel data on GNS used as a physical contrast agent; it is easy to use in a clinical setting and can be topically applied to skin generating increased OCT signals from natural skin openings. This study suggests that GNSs are interesting candidates for increasing sensitivity in OCT diagnosis of hair and sweat gland disorders because GNS primarily enhances OCT contrast in natural skin openings. Hence, the future clinical impact is considerable and our study can be considered a first step, performed on normal skin, and a natural next step would be to perform clinical studies in patients with hair follicle and sweat gland diseases. This should encourage future trials exploring not only diagnostic accuracy of GNS in OCT imaging but also look into GNS targeted drug delivery monitored by OCT.

5 | CONCLUSION

Application of 150 nm GNS improves contrast to normal skin in sweat glands in armpit skin and in hair follicles in general. GNSs are highly scattering, independent of where they are located in skin, apart from the sweat duct openings of the palm. OCT detects signal from GNS presence in hair follicles, sweat ducts and demonstrate that GNS presents adhesion abilities on skin surface. Even though GNS was massaged into skin it was not detected by OCT beyond the depth of 1.2 mm into hair follicles and only 15 to 40 μm into sweat ducts of the axillary epidermis. The GNS penetrated deeper in armpit hair follicles than on chest, and deeper in female armpits. ET thickness was lower and pixel intensity higher in UHR-OCT system compared to the commercial system—however, both systems detected a significant enhancement of the OCT signal after application of GNS. GNS was not detected in OCT images after 8 hours and did not irritate the skin nor leave discoloration.

ACKNOWLEDGMENTS

We would like to acknowledge the support from Innovation Fund Denmark through the ShapeOCT grant No. 4107-00 011A. O. Bang and M. Maria acknowledge support from European Union's Horizon 2020 grant GALAHAD (732613). Additionally, M. Maria, A. Podoleanu and O. Bang acknowledge the UBAPHODESA Marie Curie European Industrial Doctorate. A. Podoleanu is also supported by the Royal Society Wolfson Research Merit Award and the NIHR Biomedical Research Centre (BRC) at Moorfields Eye Hospital NHS Foundation Trust, UCL Institute of Ophthalmology.

Conflict of interest

None declared.

AUTHOR BIOGRAPHIES

Please see Supporting Information online.

ORCID

Mette Mogensen  <http://orcid.org/0000-0001-6479-6034>

Sophie Bojesen  <http://orcid.org/0000-0002-7493-9665>

REFERENCES

- [1] D. Huang, E. A. Swanson, C. P. Lin, J. S. Schuman, W. G. Stinson, W. Chang, M. R. Hee, T. Flotte, K. Gregory, C. A. Puliafito, J. G. Fujimoto, *Science* **1991**, 254, 1178.
- [2] J. Welzel, S. Schuh, *J. Dtsch. Dermatol. Ges.* **2017**, 15, 999.
- [3] O. Liba, E. D. SoRelle, D. Sen, A. de la Zerda, *Sci. Rep.* **2016**, 6, 23337.
- [4] M. Kirillin, M. Shirmanova, M. Sirotkina, M. Bugrova, B. Khlebtsov, E. Zagaynova, *J. Biomed. Opt.* **2009**, 14, 021017.
- [5] E. V. Zagaynova, M. V. Shirmanova, M. Y. Kirillin, B. N. Khlebtsov, A. G. Orlova, I. V. Balalaeva, M. A. Sirotkina, M. L. Bugrova, P. D. Agrba, V. A. Kamensky, *Phys. Med. Biol.* **2008**, 53, 4995.
- [6] L. V. Roque, I. S. Dias, N. Cruz, A. Rebelo, A. Roberto, P. Rijo, C. P. Reis, *Skin Pharmacol. Physiol.* **2017**, 30, 197.
- [7] R. Su, W. Fan, Q. Yu, X. Dong, J. Qi, Q. Zhu, W. Zhao, W. Wu, Z. Chen, Y. Li, Y. Lu, *Oncotarget* **2017**, 8, 38214.
- [8] M. Mogensen, T. M. Joergensen, B. M. Nurnberg, H. A. Morsy, J. B. Thomsen, L. Thrane, G. B. Jemec, *Dermatol. Surg.* **2009**, 35, 965.
- [9] J. Olsen, L. Themstrup, C. N. De, M. Mogensen, G. Pellacani, G. B. Jemec, *Photodiagnosis Photodyn. Ther.* **2016**, 16, 44.
- [10] M. Mogensen, J. B. Thomsen, L. T. Skovgaard, G. B. Jemec, *Br. J. Dermatol.* **2007**, 157, 894.
- [11] M. Mogensen, H. A. Morsy, B. M. Nurnberg, G. B. Jemec, *J. Eur. Acad. Dermatol. Venereol.* **2008**, 22, 1458.
- [12] M. Mogensen, L. Thrane, T. M. Joergensen, P. E. Andersen, G. B. Jemec, *J. Biophotonics* **2009**, 2, 442.
- [13] H. C. Ring, L. Themstrup, C. A. Banzhaf, G. B. Jemec, M. Mogensen, *JAMA Dermatol* **2016**, 152, 1142.
- [14] H. C. Ring, A. A. Hussain, G. B. Jemec, R. Gniadecki, L. M. Gjerdrum, M. Mogensen, *J. Eur. Acad. Dermatol. Venereol.* **2016**, 30, 1228.
- [15] L. Themstrup, C. Banzhaf, M. Mogensen, G. B. Jemec, *Dermatology* **2012**, 225, 242.
- [16] S. Schuh, R. Kaestle, E. C. Sattler, J. Welzel, *J. Eur. Acad. Dermatol. Venereol.* **2016**, 30, 1321.
- [17] M. Ohmi, *Laser Ther* **2016**, 25, 251.
- [18] M. Reinholz, G. G. Gauglitz, K. Giehl, M. Braun-Falco, H. Schwaiger, J. Schaubert, T. Ruzicka, M. Berneburg, B. T. von, *J. Eur. Acad. Dermatol. Venereol.* **2016**, 30, 677.
- [19] S. Nguyen, C. Chiaverini, N. Cardot-Leccia, C. Queille-Roussel, K. Roussel, J. P. Lacour, P. Bahadoran, *J. Eur. Acad. Dermatol. Venereol.* **2016**, 30, 861.
- [20] J. Lindner, K. Hillmann, U. Blume-Peytavi, J. Lademann, A. Lux, A. Stroux, A. Schneider, B. N. Garcia, *Br. J. Dermatol.* **2012**, 167, 1272.
- [21] K. Schicho, R. Seemann, M. Binder, M. Figl, *Dermatol. Surg.* **2015**, 41, 358.
- [22] U. Baran, Y. Li, W. J. Choi, G. Kalkan, R. K. Wang, *Lasers Surg. Med.* **2015**, 47, 231.
- [23] M. Manfredini, M. Greco, F. Farnetani, S. Ciardo, C. N. De, V. D. Mandel, M. Starace, G. Pellacani, *J. Eur. Acad. Dermatol. Venereol.* **2017**, 31, 1541.
- [24] O. Markowitz, S. Utz, *J. Clin. Aesthet. Dermatol.* **2015**, 8, 48.
- [25] M. G. Ahlstrom, L. M. R. Gjerdrum, H. F. Larsen, C. Fuchs, A. L. Sorensen, J. L. Forman, M. S. Agren, M. Mogensen, *Skin Res. Technol.* **2018**, 24, 65.

- [26] T. Gambichler, A. Pljakic, L. Schmitz, *Clin. Cosmet. Investig. Dermatol.* **2015**, 8, 345.
- [27] S. Schuh, R. Kaestle, E. Sattler, J. Welzel, *Skin Res. Technol.* **2016**, 22, 395.
- [28] D. Y. Paithankar, F. H. Sakamoto, W. A. Farinelli, G. Kositratna, R. D. Blomgren, T. J. Meyer, L. J. Faupel, A. N. Kauvar, J. R. Lloyd, W. L. Cheung, W. D. Owczarek, A. M. Suwalska, K. B. Kochanska, A. K. Nawrocka, E. B. Paluchowska, K. M. Podolec, M. M. Pirowska, A. B. Wojas-Pelc, R. R. Anderson, *J. Invest. Dermatol.* **2015**, 135, 1727.
- [29] F. F. Larese, M. Mauro, G. Adami, M. Bovenzi, M. Crosera, *Regul. Toxicol. Pharmacol.* **2015**, 72, 310.
- [30] D. Zhu, K. V. Larin, Q. Luo, V. V. Tuchin, *Laser Photon. Rev.* **2013**, 7, 732.
- [31] T. Son, B. Jung, *Skin Res. Technol.* **2015**, 21, 327.
- [32] H. Zhong, Z. Guo, H. Wei, C. Zeng, H. Xiong, Y. He, S. Liu, *J. Biomed. Opt.* **2010**, 15, 036012.
- [33] L. Ding, R. Sun, X. Zhang, *Oncotarget* **2017**, 8, 21200.
- [34] M. H. Akhter, M. Rizwanullah, J. Ahmad, M. J. Ahsan, M. A. Mujtaba, S. Amin, *Artif. Cells Nanomed. Biotechnol.* **2018**, 46, 873.
- [35] M. C. Pierce, D. J. Javier, R. Richards-Kortum, *Int. J. Cancer* **2008**, 123, 1979.

SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

FIGURE S1 C-OCT images illustrating hyperreflectivity from black hairs vs hyperreflective GNS. OCT images of a dark hair in the beard region shows that black hairs can be traced into the dermis and appear as thin hyperreflective rods inside dermis. After topical GNS application, the gold particles tend to form dots and caps inside of dermis. Hence, the morphology of black hairs can be differentiated from GNS lining the hair follicle and hair itself

FIGURE S2 Consecutive C-OCT images over time after application of 150 nm GNSs at timepoint: 20 minute, 4 and 8 hours. Three C-OCT images of a small light brown dermal nevus on the lower left arm of volunteer 4. (A) 20 minute after application of GNS, a clinical photo of the nevus is inserted. The nevus is well-circumscribed oval dark gray, marked by “N.” (B) At 4 hours, the outline of the nevus and the penetration depth is unchanged. GNSs are detected in an adjacent hair follicle (arrow). (C) GNSs are located in the middle of the nevus, very superficially after 8 hours (arrow). Due to strong reflectance from GNS at the nevus surface, a shadow is created below the GNS contrast agent. In (A-C), a strong entrance signal is detected on top of the skin, even though GNS has been wiped off.

How to cite this article: Mogensen M, Bojesen S, Israelsen NM, et al. Two optical coherence tomography systems detect topical gold nanoshells in hair follicles, sweat ducts and measure epidermis. *J. Biophotonics*. 2018;e201700348. <https://doi.org/10.1002/jbio.201700348>