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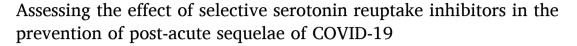
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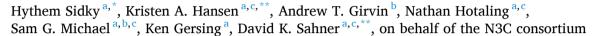
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### Research article





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#### ABSTRACT

*Background:* Post-acute sequelae of COVID-19 (PASC) produce significant morbidity, prompting evaluation of interventions that might lower risk. Selective serotonin reuptake inhibitors (SSRIs) potentially could modulate risk of PASC via their central, hypothesized immunomodulatory, and/or antiplatelet properties although clinical trial data are lacking.

Materials and Methods: This retrospective study was conducted leveraging real-world clinical data within the National COVID Cohort Collaborative (N3C) to evaluate whether SSRIs with agonist activity at the sigma-1 receptor (S1R) lower the risk of PASC, since agonism at this receptor may serve as a mechanism by which SSRIs attenuate an inflammatory response. Additionally, determine whether the potential benefit could be traced to S1R agonism. Presumed PASC was defined based on a computable PASC phenotype trained on the U09.9 ICD-10 diagnosis code.

*Results*: Of the 17,908 patients identified, 1521 were exposed at baseline to a S1R agonist SSRI, 1803 to a non-S1R agonist SSRI, and 14,584 to neither. Using inverse probability weighting and Poisson regression, relative risk (RR) of PASC was assessed.

A 29% reduction in the RR of PASC (0.704 [95% CI, 0.58–0.85];  $P=4\times10^{-4}$ ) was seen among patients who received an S1R agonist SSRI compared to SSRI unexposed patients and a 21% reduction in the RR of PASC was seen among those receiving an SSRI without S1R agonist activity (0.79 [95% CI, 0.67 - 0.93]; P=0.005). Thus, SSRIs with and without reported agonist activity at the S1R were associated with a significant decrease in the risk of PASC.

#### 1. Introduction

# 1.1. Post-acute sequelae of SARS-CoV-2

Post-Acute Sequelae of SARS-CoV-2 Infection (PASC) or "Long COVID" has been estimated by the World Health Organization to afflict between  $\sim\!10\text{--}20\%$  of patients with COVID-19, although estimates vary considerably based on methodology. [1] At least one relatively large EHR-based study suggested that approximately one-third of patients may have symptoms compatible with PASC 3–6 months after diagnosis of COVID-19, with a higher risk among females and those with more severe disease. [2] Symptoms of PASC are generally nonspecific across

multiple organ systems including respiratory, neurological and gastro-intestinal symptoms. [3–5] These symptoms may substantially impair function, persist for an extended duration, and fluctuate over time. [2,

The World Health Organization (WHO) has developed a case definition of "post COVID-19 condition" using Delphi methodology with input from researchers, patients, and others across all WHO regions, suggesting that the syndrome is usually diagnosed several months after the onset of acute symptoms of COVID-19 based on new-onset or lingering symptoms (e.g., fatigue, dyspnea, cognitive dysfunction) of at least two months duration which cannot be explained by an alternative etiology. At present, there is no specific recommended treatment for

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PASC other than supportive care and rehabilitation, [7] although some preliminary observational and cross-sectional data suggest that SARS-CoV-2 vaccination, either before or shortly after infection, may attenuate the risk of PASC. [8–10].

### 1.2. Pathogenesis of PASC

The characterization of PASC, its potentially variable phenotypes and possible pathophysiological underpinnings, remains at a relatively early stage, although insights have been provided by a number of studies. Hypotheses that have been adduced have focused primarily on immune dysregulation (discussed below), but other speculative explanations have also been put forth, including redox imbalance and/or mitochondrial dysfunction, [11–13] persistence of virus in sanctuary sites, [14,15] and formation of fibrinolysis-resistant amyloid fibrin microclots. [16].

Most work attempting to clarify the pathogenesis of PASC has centered on a potentially aberrant immune response primarily because little to no evidence exists of productive viral replication in the vast majority of patients. Although viral RNA has been detected in various tissues at autopsy after extended follow-up [17] and shedding of virus in the stool may persist after respiratory shedding has ceased, prolonged shedding of virus over many months is unusual, with notable exceptions including reports in immunocompromised patients. [18–20] Swank and colleagues [15] customized an existing assay in order to detect spike and/or nucleocapsid antigen in the blood of 31 patients with PASC and found evidence for prolonged antigenemia, but, importantly, discordance was noted between long-term expression of spike protein (60% of patients) and nucleocapsid antigen (a single patient). To our knowledge, there have been no reports of virus cultured from the blood of any patient with PASC.

Markers of immune activation and inflammation among patients with PASC were evaluated by Peluso et al. [21] Trends in these data revealed that patients with PASC were more likely to be female (61.6%) and to have a history of autoimmune disease (11%) versus those who had recovered from COVID-19 without PASC (2.1%). Significantly higher levels of TNF-α and IP-10, and a trend toward higher levels of IL-6, were seen during early recovery among those patients who developed PASC. At the late recovery time point, IL-6 levels were significantly higher among patients with PASC. In another study, compared with healthy controls patients with PASC had higher plasma levels of CCL5/RANTES, IL-2, IL-4, CCL3, IL-6, IL-10, IFN-γ, and VEGF; decreased T regulatory cells and B cell elevations were also seen. [22].

Su and colleagues [23] extensively characterized a primary cohort of over 200 SARS-CoV-2-infected inpatients and outpatients followed longitudinally for up to 2–3 months versus matched controls. In addition, a separate cohort of 100 COVID-19 inpatients and outpatients was asked to return at 60 or 90 days of follow-up and 33 COVID-19 patients from a third cohort also provided plasma samples to validate some of the findings. Significant associations were variably seen between certain late PASC manifestations and autoantibodies during acute infection directed at IFN $\alpha$ -2, U1-snRNP, and LL/SS-B, but given the number of comparisons and limited sample size, it is difficult to draw definitive conclusions from this observation.

Of note, 44% of patients in the study by Su and colleagues had autoantibodies at the extended follow-up visit but 56% of patients had class-switched autoantibodies during the acute infection, suggesting that antibodies antedated COVID-19 infection. The vast majority of patients with autoantibodies had no prior diagnosis of an auto-immune condition. Patients with elevated plasma IFN- $\gamma$  during acute infection had elevated autoantibodies directed at IFN $\alpha$ -2 at extended follow-up, and those with increased plasma levels of the chemokine CXCL9 exhibited elevated autoantibody to P1 at delayed follow-up (P < 0.001). There was an anticorrelation between virus-specific antibody levels and autoantibodies (antinuclear and anti-IFN $\alpha$ -2). CMV-specific CD8 + T cells were seen at late follow-up in patients with gastrointestinal PASC

symptoms, leading to the hypothesis that bystander activation of these cells may be seen in GI PASC. Several immune phenotypes at delayed follow-up were variably associated with different PASC symptoms, including myeloid-derived suppressor cells, memory-like NK cells, and innate immune activation, whereas activated Treg cells during acute infection seemed to augur PASC.

Vijayakumar et al. identified an immunoproteomic signature in patients with respiratory disease after acute COVID-19, particularly using BAL (airway) samples. An increase in activated CD8 + T cells in bronchoalveolar lavage fluid was associated with a reduction in forced vital capacity (FVC). [24] Chemokines that recruit T cells and NK cells were also highly associated with epithelial damage markers. In addition, increased memory B cells in airways was linked to reduction in FVC or TCLO, or imaging abnormalities.

In aggregate, the data above suggest that immune dysregulation, associated with increased systemic and/or pulmonary levels of chemokines, pro-inflammatory cytokines, and specific lymphocyte phenotypes, may be responsible for the pathogenesis of PASC, although further study is needed, particularly of larger cohorts of patients followed prospectively, with intensive evaluation of both plasma and cell-based biomarkers. Immune dysregulation may be associated with autoantibodies, and the potential role of cross-reactive antibodies directed at host targets cannot be excluded, but early published data suggest that these autoantibodies may antedate COVID-19, possibly identifying a subset of patients predisposed to develop PASC.

### 1.3. SSRIs as immunomodulatory agents in COVID-19

SSRIs may harbor potential as immunomodulators. Abundant in vitro, animal model, and, to some extent, clinical data suggest SSRIs have immunomodulatory properties, primarily immunosuppressive activity. [25-30] In interpreting these data, attention must be paid, however, to dose, concentration, activation status of lymphocytes, experimental design and the underlying disease/model. Doses and concentrations in preclinical studies may be higher than those in humans. For example, concentrations at which lymphocyte proliferation is inhibited by SSRIs are typically  $\geq 1 \mu M$ , higher than those typically achieved in the clinic. Sertraline, however, at concentrations of 0.01 and 1  $\mu M$  decreases the IFN/IL-10 ratio in the supernatant of mitogen-stimulated whole blood. [26] SSRIs deplete platelet serotonin and, in mice, platelet-derived serotonin appears to promote neutrophil endothelial adhesion and inflammation-triggered extravasation of neutrophils into tissue [31], suggesting a possible mechanism for SSRI-mediated anti-inflammatory activity. Several other mechanisms have been proposed to explain the immunomodulatory effects of SSRIs. Inhibition of the serotonin transporter protein, resulting in increased extracellular concentrations of 5-HT, was thought to be responsible for the observed effects of SSRIs on lymphocytes, but multiple lines of experimental evidence contravene this hypothesis. [26] The impact of SSRIs on several signal transduction pathways has been posited as the basis for diminished lymphocyte proliferation and apoptosis; these effects include activation of PKA through induction of cAMP, inhibition of the translocation of PKC to the cell surface, and reduction in calcium influx. [26] However, reduced lymphocyte proliferation and apoptosis are generally not expected at typical clinical concentrations of commonly used SSRIs. Finally, a recent study [32] provided data that has led to the hypothesis that type I interferon-mediated depletion of serotonin through several mechanisms, including impaired gut absorption of its precursor (tryptophan), may be responsible for some manifestations of Long COVID through reduced vagal signaling, resulting in impaired cognition (e.g., brain fog, which is typical of Long COVID).

Recently, attention has focused on the observation that several SSRIs bind with moderate to high affinity to the sigma-1 receptor (SIR) [33], and it has been suggested that ligands for this receptor may modulate the immune system. The S1R is a ubiquitous endoplasmic reticulum (ER)-resident chaperone protein that associates with IRE1 in the context

of ER stress, leading to regulation of the production of inflammatory cytokines. IRE1 is necessary for cytokine production, presumably through XBP1-mediated transactivation of IL-6 and TNF-α. Fluvoxamine, which exhibits the highest affinity for the S1R among antidepressants and exerts agonistic activity, has been shown to produce anti-inflammatory effects in a variety of cell models. [34] Rosen et al. [35] showed that S1R KO mice experienced increased mortality in two sub-lethal models of sepsis, and that WT mice treated with fluvoxamine were protected from mortality and had lower IL-6 levels in a lethal model of sepsis. Data suggest that, at least for some SSRIs, typical clinical concentrations may result in appreciable binding at the S1R (see Table S1 in Supplementary Appendix). In particular, we have calculated approximate ratios of free (unbound) typical clinical concentrations to the dissociation constant at the sigma-1 receptor of 2-8.5 for fluvoxamine, 0.2 - 0.6 for citalogram, 0.2 to 0.4 for escitalogram, 0.1 to 0.5 for fluoxetine and 0.1 or less for sertraline. Of note, fluoxemine, fluoxetine, and escitalopram are agonists at the S1R whereas sertraline is an antagonist; paroxetine is not expected to bind appreciably to the S1R and citalopram is not believed to be an agonist at the SIR. [33,36,37].

Whatever the mechanistic basis might be, some clinical data suggest that pro-inflammatory cytokine levels in the clinic may be affected by SSRIs. In a large meta-analysis of largely uncontrolled data from longitudinal studies of depressed patients treated pharmacologically, IL-6 levels fell significantly irrespective of treatment response. [38] This analysis was not stratified by type of antidepressant treatment, but, in another meta-analysis of pharmacologic treatment for depression that partially overlapped with the study above, changes in cytokine levels were also assessed by drug class. [39] Overall, treatment reduced levels of IL-1\u03b3. Selective serotonin reuptake inhibitors were also found to potentially affect levels of IL-6 and TNF- $\alpha$ , but other antidepressants in this meta-analysis did not appear to reduce measured cytokine levels. In a small randomized controlled study, decreases in IL-1 $\beta$  were seen among depressed patients treated with fluoxetine or acupuncture vs. placebo, [40] but only 68/95 subjects contributed baseline samples, and cytokine data were available at follow-up for only 72 subjects. There was no apparent impact of fluoxetine on TNF- $\alpha$  or Th2 cytokine levels. Other data suggest that other forms of treatment for depression may influence cytokine levels. [41,42] Thus, the vast majority of the available clinical data suggesting SSRIs may reduce cytokine levels derive from uncontrolled studies, and the paucity of prospective randomized controlled data in which cytokine data have been consistently measured make it difficult to determine if this effect is specific to SSRIs in the context of treatment for depression.

In view of the earlier described findings suggesting SSRIs may exert immunomodulatory effects, possibly through sigma-1 receptor (S1R) binding, we hypothesized that chronically administered SSRIs might dampen a pro-inflammatory immune response during and after SARS-CoV-2 infection and thereby reduce the risk of PASC. As noted above, some data suggest that ongoing immune dysregulation may be responsible for the pathogenesis of PASC. We therefore evaluated, in this retrospective study relying upon a dataset including de-identified EHR and PPRL-linked ancillary data, the risk of PASC among recipients of commonly prescribed SSRIs. In particular, we separately studied the effects of SSRIs with (a) agonist activity at the S1R or (b) no agonist activity at the S1R (i.e., no meaningful binding at the S1R or antagonist activity at that receptor) in an effort to discern whether SIR agonism played a role in any observed beneficial effect.

# 2. Materials and Methods

# 2.1. Setting and population

We used data from the National COVID Cohort Collaborative (N3C), a centralized real-world repository of de-identified electronic health records supported by the National Institutes of Health (NIH). N3C includes detailed information on clinical encounters including procedures,

diagnoses, ordered and administered medications, demographic data, vitals, and lab orders and results. Records in N3C are aggregated across participating clinical organizations in the United States, known as data partners, harmonized using the Observational Medical Outcomes Partnership (OMOP) data model, and subjected to quality review and checks. This research was possible because of the patients whose information is included within the data and the organizations (htt ps://ncats.nih.gov/n3c/resources/data-contribution/data-transfer-ag reement-signatories) and scientists who have contributed to the ongoing development of this community resource. The N3C data transfer to NCATS is performed under a Johns Hopkins University Reliance Protocol #IRB00249128 or individual site agreements with the NIH. The N3C Data Enclave is managed under the authority of the NIH; information can be found at https://ncats.nih.goc/n3c/resources. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health or the N3C program. Use of N3C data for this study does not involve human subjects (45 CFR 46.102) as determined by the NIH Office of IRB

We included adult patients (> 18 years) with either RT-PCR or antigen (AG) confirmed SARS-CoV-2 infection or a recorded U07.1 diagnosis for COVID-19. Due to the delayed availability of the U09.9 diagnosis code for PASC on which our computable phenotype was trained (see below), only patients with an index date, defined as the earliest record of infection or diagnosis, on or after October 1, 2021 through April 7, 2022 were included. This would mitigate the potential lack of generalizability of the algorithm over time and also limit the analysis to the inclusion of subjects with either Delta or Omicron infection. Patients were required to have at least one interaction with the participating health care system prior to and following the index diagnosis of COVID-19. Since collection of vaccination data in the EHR may have been incomplete, the analysis was restricted to participating N3C sites with vaccination rates comparable to those reported by the CDC for the corresponding geographic region, as vaccination may affect the risk of PASC and was considered a covariate. To allow enough time for collection of data relevant to a PASC diagnosis, we included only patients with at least 90 days between their index diagnosis date and the data extraction date. We excluded patients with missing gender data and those who died during COVID-19-related hospitalization or within 45 days of their index date.

#### 2.2. Exposures

Three groups were defined: (1) patients with documented baseline exposure to least one SSRI with agonist activity at the S1R (fluvoxamine, fluoxetine, escitalopram) [33], (2) patients with documented baseline exposure to at least one SSRI without agonist activity at S1R (sertraline, which is a SIR antagonist; paroxetine, which does not appreciably bind to the S1R; and citalopram, which did not show agonist activity at the sigma-1 receptor in nerve growth factor-induced neurite outgrowth in PC12 cells) [33,36,37], and (3) control patients with no documented baseline exposure to either (1) or (2). Baseline use of a drug was defined as recorded at or prior to the COVID-19 index diagnosis date. Records in N3C date back to January 1, 2018 for patients who had been followed for the longest period of time. Patients with documented baseline exposure to both (1) and (2) were excluded. Treatment comparisons were performed between patients in the exposure group (1) versus controls and the exposure group (2) versus controls. To further interrogate the effect of S1R agonism as a mechanism of action, we conducted a secondary analysis with non-S1R agonist SSRI exposure as a comparator to S1R agonist SSRI exposure.

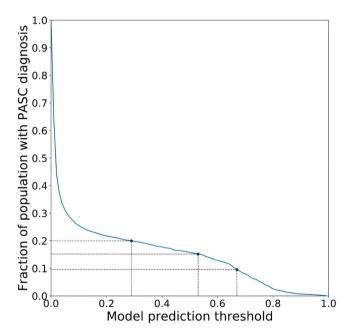
#### 2.3. Outcomes

The primary outcome consisted of a presumptive diagnosis of PASC using an XGBoost-based machine learning algorithm trained on patients

assigned the U09.9 (PASC) diagnosis code. [43] This surrogate was chosen in an effort to more comprehensively capture patients with Long COVID because the U09.9 code was established in October of 2021 and was not rapidly and fully embraced by all participating healthcare organizations. Neither vaccination nor the SSRIs under study were included as features in this predictive model. The prediction threshold for the model was chosen to be 0.53 (see Fig. 1), which corresponds to 15% of the total patient population receiving a PASC diagnosis. Rates of PASC reported in the literature have varied widely, but the likely rate is generally believed to reside within a range of approximately 10–20%. [1] In view of this, a supplementary analysis was performed using a lower and higher score threshold yielding a rate of 10% and 20% respectively.

## 2.4. Statistical analysis

For the primary analyses we compared the risk of PASC among patients receiving a S1R agonist SSRI (fluvoxamine, fluoxetine, or escitalopram) at baseline to patients not receiving any SSRI and we also compared the risk of PASC among patients receiving an SSRI with no S1R agonism (sertraline, paroxetine, or citalopram) at baseline to patients not receiving any SSRI. Relative risks (RRs) and 95% CIs were estimated using weighted Poisson regression with robust standard errors. Inverse probability weighting (IPW) as implemented in the R WeightIt package, version 0.9.0, was used to adjust for confounding. IPW as a method to adjust for baseline confounding is a common and effective method to elucidate causal relationships between exposures and outcomes [44]. The inverse propensity scores are used as weights in the final Poisson model between exposure and outcome. The individual exposure propensity scores were calculated using multivariate logistic regression and included a variety of potential baseline confounders: age, sex, body mass index (BMI), race and ethnicity, baseline comorbidities (myocardial infarction, congestive heart failure, cerebrovascular disease, dementia, chronic lung disease, uncomplicated type II diabetes, complicated type II diabetes, kidney disease, liver disease, HIV infection, and cancer), record of post-infection hospitalization, use of approved immunomodulatory therapy for acute COVID-19 (dexamethasone,



**Fig. 1.** Fraction of patient population receiving a PASC diagnosis as a function of predictive model probability threshold. A cutoff of 0.53 was chosen for the main analysis which corresponds to 15% of the patient population. Cutoffs of 0.29 and 0.67 are also shown, corresponding to 10% and 20% of the population respectively.

baricitinib, tocilizumab), baseline immunotherapy treatment (etanercept, infliximab, adalimumab, certolizumab pegol, golimumab, sarilumab, tocilizumab, baricitinib, tofacitinib, or upadacitinib), baseline exposure to other ligands exhibiting S1R binding with moderate to high affinity defined as a dissociation constant of less than 250 nanomolar (trifluoperazine, pimozide, fluphenazine, chlorpromazine, perphenazine, haloperidol, pentazocine, progesterone, methamphetamine, hydroxychloroguine, dextromethorphan, clemastine, hydroxyzine, tamoxifen), any baseline SNRI medication (desvenlafaxine, duloxetine, levomilnacipran, venlafaxine), baseline bupropion exposure, any baseline tricyclic antidepressant medication (clomipramine, amoxapine, amitriptyline, desipramine, nortriptyline), any baseline benzodiazepine medication (alprazolam, chlordiazepoxide, diazepam, lorazepam), any baseline antipsychotic medication (risperidone, quetiapine, ziprasidone, aripiprazole, aripiprazole lauroxil, clozapine), and any record of COVID-19 vaccination. For BMI, which was the only variable with missing data, a missing data indicator was created. The estimated probabilities were used to calculate stabilized inverse probability weights and used to weight each patient's contribution to the Poisson regression model. Covariate balance before and after inverse probability weighting was assessed by calculating absolute standardized mean differences (SMDs) and comparing the distributions of propensity scores for exposed and unexposed groups, which were generated using the R Cobalt package, version 4.2.2.

We performed additional analyses using the methods detailed above for the primary analysis. To isolate and determine the significance of S1R agonism in reducing the risk of long-COVID, we used non-S1R agonist SSRIs as a comparator to S1R agonist SSRIs. The effect of both SSRI classes across sex and COVID-19 vaccination status (record of any COVID-19 vaccination vs. unvaccinated) subgroups was also assessed. As a sensitivity analysis, we repeated our primary analysis with lower and higher probability thresholds for the PASC computable phenotype ML model of 0.67 and 0.29, which correspond to 10% and 20% of the population respectively, to establish the robustness of the results to the long-COVID diagnosis assignments. Finally, to assess the impact of increased specificity to recent ongoing SSRI treatment prior to infection, we limited the exposure definition to include only patients with a record of exposure within 180 days of the index COVID-19 diagnosis date.

Data extraction was carried out in the N3C enclave using Spark SQL 3.2.1 and Python 3.6.7, and all statistical analysis was done using R version 3.5.

#### 3. Results

There were 17 908 eligible patients identified, with a total of 1521 patients exposed to S1R agonist SSRIs at baseline, 1803 exposed to non-S1R agonist SSRIs, and 14 584 unexposed patients (Fig. 2). Both the S1R and non-S1R agonist SSRI exposed patients were more female compared with unexposed patients (1180 [78%] and 1327 [74%] vs 8943 [61] and more likely to be White non-Hispanic (1177 [77%] and 1367 [76%] vs 9860 [68%]). Exposed patients generally had more comorbidities (congestive heart failure, 141 [9.3%] and 217 [12%] vs 1181 [8.1%]; cerebrovascular disease, 99 [6.5%] and 138 [7.7%] vs 641 [4.4%]; chronic lung disease, 434 [29%] and 512 [28%] vs 2967 [20%]; uncomplicated type II diabetes, 301 [20%] and 455 [25%] vs 2793 [19%]; cancer, 168 [11%] and 248 [14%] vs 1621 [11%]); exposed patients were also more likely to have been exposed to other high S1R affinity ligands at baseline (402 [26%] and 495 [27%] vs 1931 [13%]), and more likely to have received other classes of psychiatric medication at baseline (bupropion, 289 [19%] and 250 [14%] vs 586 [4.0%]; benzodiazepines, 514 [34%] and 574 [32%] vs 2113 [14%]; tricyclic antidepressants, 99 [6.5%] and 116 [6.4%] vs 392 [2.7%]; antipsychotics, 135 [8.9%] and 139 [7.7%] vs 298 [2.0%]; SNRIs, 189 [12%] and 230 [13%] vs 987 [6.8%]). Exposed patients were more likely to have received a COVID-19 vaccine (1026 [67%] and 1215 [67%] vs 8744 [60%]). COVID-19 related hospitalization was similar across all three

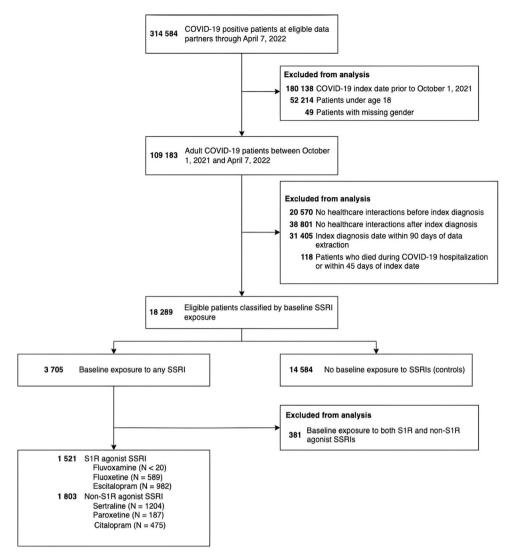


Fig. 2. Flow diagram of patient selection. Note that SSRI exposure is non-exclusive and small numbers (<20) were obscured to protect patient identities.

groups (208 [14%] and 270 [15%] vs 2308 [16%]) as was baseline immunotherapy (24 [1.6%] and 35 [1.9%] vs 244 [1.7%]) and COVID-19 immunotherapy (24 [1.6%] and 46 [2.6%] vs 372 [2.6%]) but unexposed patients, unadjusted, were more likely to develop long-COVID (175 [12%] and 215 [12%] vs 2352 [16%]). Complete cohort characteristics are shown in Table 1.

After applying IPW, patient characteristics between exposed and unexposed groups for both S1R agonist and non-S1R agonist SSRIs analyses were adequately balanced, with absolute standardized mean differences (SMDs) under 0.1 for almost all covariates. Fig. 3 shows the SMDs for treated and control groups along with the propensity score distributions before and after weighting. Covariate balances for secondary and subgroup analyses were also adequate and are presented in the Supplement (eFigure 1 to eFigure 11).

In the weighted analysis, there was a 29% reduction in the RR of long-COVID (0.704 [95% CI, 0.58–0.85];  $P=4\times 10^{-4}$ ) among patients with baseline S1R agonist SSRI exposure compared to unexposed patients. For patients with baseline non-S1R agonist SSRI exposure compared to unexposed patients, there was a 21% reduction in RR (0.79 [95% CI, 0.67 - 0.93]; P=0.005). When considering non-S1R agonist SSRIs as a comparator, no statistically significant difference in RR (0.96 [95% CI, 0.80 - 1.16]; P=0.7) was observed between S1R agonist SSRI recipients and non-S1R agonist SSRIs. Additional analysis showed that the significance of RR reduction among both genders for S1R agonist

SSRI exposure was maintained (male: 0.64 [95% CI, 0.44 - 0.93], P=0.02; female: 0.73 [95% CI, 0.60 - 0.90], P=0.003) but not for non-S1R agonist SSRIs (male: 0.61 [95% CI, 0.43 - 0.87], P=0.006; female: 0.92 [95% CI, 0.77 - 1.1], P=0.4). Subgroup analysis on vaccinated and non-vaccinated patients with S1R agonist SSRI exposure revealed a significant RR reduction for vaccinated patients (0.73 [95% CI, 0.57 - 0.94]; P=0.01) but not among unvaccinated patients (0.75 [95% CI, 0.55 - 1.03]; P=0.1) and similarly for vaccinated (0.76 [95% CI, 0.60 – 0.97]; P=0.03) and unvaccinated patients (0.87 [95% CI, 0.68 - 1.1]; P=0.2) with non-S1R agonist SSRI exposure.

The results of our analyses proved to be robust to the ML model threshold for Long-COVID assignment. At a threshold of 0.29 where 20% of the patient population received a long-COVID diagnosis, we still observed a 19% reduction in the RR of long-COVID (0.80 [95% CI, 0.68 - 0.94]; P=0.007) among S1R agonist SSRI exposed patients and a 18% reduction in RR of long-COVID (0.82 [95% CI, 0.72 - 0.94]; P=0.004) among non-S1R agonist SSRI exposed patients, both compared to unexposed patients. At a threshold of 0.67 where 10% of the patient population received a long-COVID diagnosis, RR significance is lost for both S1R agonist exposure (0.80 [95% CI, 0.64 - 1.0]; P=0.06) and non-S1R agonist exposure versus control (0.84 [95% CI, 0.69 – 1.03]; P=0.1). Furthermore, the second sensitivity analysis limiting the SSRI exposure window to 180 days prior to the COVID-19 index date yielded a lower RR (0.67 [95% CI, 0.54 - 0.83];  $P=2\times10^{-4}$ ) compared to the

**Table 1**Cohort characteristics by SSRI exposure group. Note that small numbers (<20) were obscured to protect patient identities.

	•		
Characteristic	S1R agonist	Non-S1R agonist	Unexposed
	SSRI exposure	SSRI exposure	$N = 14584^{a}$
	$N = 1521^{a}$	$N = 1803^{a}$	
Sex			
Male	341 (22%)	476 (26%)	5641 (39%)
Female		1327 (74%)	8943 (61%)
Age, median (years)	1180 (78%)	1327 (74%)	8943 (01%)
• •	006 (540/)	000 (400/)	7006 (400/)
Age (years)	826 (54%)	883 (49%)	7006 (48%)
18-49	194 (13%)	311 (17%)	2332 (16%)
≥ 70	286 (19%)	341 (19%)	2765 (19%)
50-59	215 (14%)	268 (15%)	2481 (17%)
60-69			
BMI, median (kg/m²)	166 (11%)	195 (11%)	1957 (13%)
BMI (kg/m <sup>2</sup> )	382 (25%)	424 (24%)	2518 (17%)
< 25	339 (22%)	393 (22%)	3595 (25%)
≥ 40	373 (25%)	454 (25%)	3457 (24%)
25-29	249 (16%)	322 (18%)	2250 (15%)
30-34	12 (0.8%)	15 (0.8%)	807 (5.5%)
35-39.9			
Missing	1177 (77%)	1367 (76%)	9860 (68%)
Race and ethnicity	25 (1.6%)	24 (1.3%)	309 (2.1%)
White Non-Hispanic	208 (14%)	289 (16%)	3094 (21%)
Asian Non-Hispanic	62 (4.1%)	88 (4.9%)	899 (6.2%)
Black or African	49 (3.2%)	35 (1.9%)	422 (2.9%)
American Non-			
Hispanic			
Hispanic or Latino Any	54 (3.6%)	102 (5.7%)	630 (4.3%)
Race			
Other Non-Hispanic /	141 (9.3%)	217 (12%)	1181 (8.1%)
Unknown			
Myocardial infarction	99 (6.5%)	138 (7.7%)	641 (4.4%)
Congestive heart	42 (2.8%)	70 (3.9%)	245 (1.7%)
failure			
Cerebrovascular	434 (29%)	512 (28%)	2967 (20%)
disease			
Dementia	301 (20%)	455 (25%)	2793 (19%)
Chronic lung disease	171 (11%)	281 (16%)	1564 (11%)
Type II diabetes,	176 (12%)	270 (15%)	1567 (11%)
uncomplicated			
Type II diabetes,	165 (11%)	179 (9.9%)	1009 (6.9%)
complicated			
Kidney disease	9 (0.6%)	13 (0.7%)	82 (0.6%)
Liver disease	168 (11%)	248 (14%)	1621 (11%)
HIV infection	208 (14%)	270 (15%)	2308 (16%)
Cancer	289 (19%)	250 (14%)	586 (4.0%)
COVID-19 related	514 (34%)	574 (32%)	2113 (14%)
hospitalization			
Baseline bupropion use	99 (6.5%)	116 (6.4%)	392 (2.7%)
Baseline	135 (8.9%)	139 (7.7%)	298 (2.0%)
benzodiazepine use			
Baseline tricyclic	189 (12%)	230 (13%)	987 (6.8%)
antidepressant use		• •	
Baseline antipsychotic	24 (1.6%)	35 (1.9%)	244 (1.7%)
use	,	,	
Baseline SNRI use	402 (26%)	495 (27%)	1931 (13%)
Baseline	24 (1.6%)	46 (2.6%)	372 (2.6%)
immunotherapy			
treatment			
Baseline moderate to	1026 (67%)	1215 (67%)	8744 (60%)
high affinity S1R	(3, 70)	(=/ ////	2 (0070)
ligand use			

<sup>&</sup>lt;sup>a</sup> Statistics presented: n (%); median (IQR).

primary analysis for the S1R agonist exposure. For the corresponding non-S1R agonist exposure analysis, the RR (0.78 [95% CI, 0.62 - 0.99]; P=0.04) was similar to the primary analysis. Fig. 4 summarizes the RR estimates for all these analyses.

### 4. Discussion

In this multicenter retrospective study, we observed a statistically significant 29% reduction in the RR of long-COVID among patients

receiving baseline S1R agonist SSRIs when compared with controls. A similar effect size of 21% reduction in the RR of long-COVID was observed for patients receiving non-S1R agonist SSRIs compared to controls. Furthermore, when comparing patients receiving baseline S1R agonist SSRIs to those receiving non-S1R agonist SSRIs, no significant difference was observed. Our findings suggest that use of SSRIs, initiated prior to diagnosis of COVID-19, may be effective in reducing the risk of long-COVID compared with controls. We did not, however, find conclusive evidence to attribute this effect to SSRI-related S1R-agonism. We speculate that part of the reason why a statistically significant difference was observed for both S1R agonist SSRIs and non-S1R agonist SSRIs versus controls may reside in protective effects unrelated to S1R agonism. We did not see a higher risk of PASC among the group of patients receiving paroxetine, sertraline or citalogram even though sertraline is an S1R antagonist. We hypothesize that this is related to the fact that typical free circulating concentrations of sertraline relative to its dissociation constant are low (see eTable 1 in the Supplementary Appendix).

As discussed in the introduction, the pathogenesis of PASC has not yet been firmly established, but immune dysregulation with an elevation in proinflammatory cytokine levels may play a role in the condition. We speculate that SSRIs may attenuate such an aberrant response through immunomodulation. In addition, SSRIs exert antiplatelet activity, which might, hypothetically, provide benefit in PASC, as microclots may contribute to the pathology of PASC. [16].

Several antidepressant drugs, such as fluoxetine, have been shown to have antiviral properties. The in vitro antiviral effect of fluoxetine has been tentatively ascribed to a reduction in intra-lysosomal ceramide related to inhibition of acid sphingomyelinase, but fluoxetine exhibits only low micromolar antiviral potency [34] raising doubt that it would show evidence of in vivo antiviral efficacy at clinically achieved concentrations. In a prospective randomized double-blind controlled study of COVID-19 outpatients, fluvoxamine exerted no antiviral effect. [45].

Retrospective studies evaluating the potential clinical benefit of SSRIs in hospitalized patients with acute COVID-19 have yielded conflicting findings, ranging from protective effects, such as decreased risk of intubation or death, to absence of any discernible benefit. [46-48] Similarly, several, [45,49] though not all [50-52] prospective, randomized, controlled investigations into the potential therapeutic effects of fluvoxamine in outpatients with acute COVID-19, have suggested evidence of benefit. A number of other investigators have been interested in the evaluating whether SSRIs prevent COVID-19, exert a treatment effect in acute infection or lower early mortality after acute COVID-19 [53-57], and a recent uncontrolled study including 95 patients with PASC treated with SSRI therapy has been published [58], but to our knowledge, ours is the first study to suggest a diminished risk of the development of Long COVID in patients receiving SSRIs at baseline (i.e., prior to SARS-CoV-2 infection). Although an immunological basis for this observation has been postulated, we acknowledge the possibility that the findings in the current study may reflect non-immunological effects of SSRIs. Future prospective studies of SSRIs in patients with PASC or at high risk for PASC that also include immunologic biomarkers seem warranted.

This study has several limitations. First, as a retrospective study using IPW, residual confounding and confounding by indication are two relevant concerns. Residual confounding is a known concern in observational research, but we verify that all known confounders in the weighted population were balanced. For confounding by indication, we minimize this through the inclusion of other medications that share indications with SSRIs. Second, the ICD diagnosis code for PASC (U09.9) is of recent vintage, so it is likely that many subjects in N3C without that code may have had PASC. We restricted our analysis to the time during which the code has been available but, since the U09.9 code was not immediately and widely adopted at all sites, we relied on ML predictions of the PASC phenotype to identify patients more likely to have the condition, including those without a U09.9 code. Using the model's

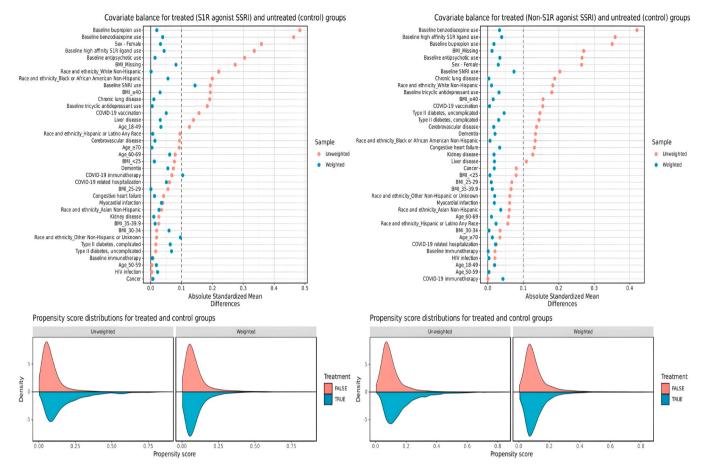


Fig. 3. Covariate balance and propensity score distributions before and after weighting for (left) S1R agonist SSRI exposed group vs control and (right) non-S1R agonist SSRI exposed group vs control.

predictions requires the selection of a probability threshold beyond which patients are labeled with the outcome. We conducted a sensitivity analysis which established a degree of robustness to our choice of prediction threshold, although at the lowest computable phenotype prevalence, significance was lost for S1R agonist SSRIs. Additionally, the ML model relies exclusively on structured data, whereas symptoms and signs of PASC may not be fully captured in structured data fields of deidentified EHRs. Although the ML model did not include the medications investigated here as predictors, it did include several of the comorbidities we included as covariates (diabetes, chronic kidney disease, congestive heart failure, and chronic lung disease). We do not consider this to be a major concern because IPW balances these characteristics in the exposed and unexposed groups, and all relevant comorbidities are more prevalent in the exposed population (see Table 1).

Another limitation imposed by the ML model is the inability to conduct a sensitivity analysis on patient healthcare interaction. Since N3C contains de-identified EHR records, it is not possible to distinguish between previously healthy patients with a genuine lack of baseline medical history and patients who are new to a healthcare organization for whom prior medical history was not collected. The same applies to interactions with a healthcare organization after the index date: there are many possible reasons why a patient may have no further interactions. Our analyses were therefore limited to patients with some record of interaction with the healthcare organization prior to and after their index diagnosis. Yet since the ML model has a 90-day blackout window around the COVID-19 index date, we could not assess how a lack of baseline medical history and loss to follow-up affected the analysis. We were further unable to study the competing risk of death within 45 days of the COVID-19 index date. However, patients who died represented a very small fraction of the total population (118/109 183).

Third, data on vaccination, which may affect the risk of PASC, are potentially incompletely captured in the N3C database, though we did mitigate this by selecting sites with vaccination rates comparable to those of CDC records. Fourth, our primary analyses did not consider timing, duration, and dosage of the medications at baseline. However, these medications are typically administered chronically and therefore patient exposure will typically be for an extended period. We further conducted a sensitivity analysis by restricting treatment to patients who were exposed to the SSRIs within 180 days prior to their COVID-19 index diagnosis date. This analysis is more rigorous in capturing active SSRI use prior to infection and the resulting estimates remained significant. Fifth, by exclusively relying on de-identified EHR records and selecting for patients with any baseline medical history, the data may not be representative of the general population. Patients with limited access to healthcare or those who seek care at small practices or community hospitals are likely underrepresented. Finally, our results are applicable only to the Delta and Omicron variant given the period of study. Future use of adapted machine learning models to reliably identify additional PASC patients prior to the advent of the U09.9 code may be useful to increase the sizes of the populations in analyses, enabling a finer analysis of the contributions of individual SSRI agents and the influence of specific variants on the risk of PASC. Similarly, repeated analyses in the future, including patients infected with the most recent variants of SARS-CoV-2 will be of interest.

In conclusion, baseline exposure to either S1R agonist or non-S1R agonist SSRIs was associated with a significant reduction in relative risk of long-COVID. Our findings can be used to guide additional research into mechanisms of action of SSRIs on COVID-19. Future prospective, randomized, double-blind, placebo-controlled clinical studies of SSRIs as a potential means of preventing Long COVID appear

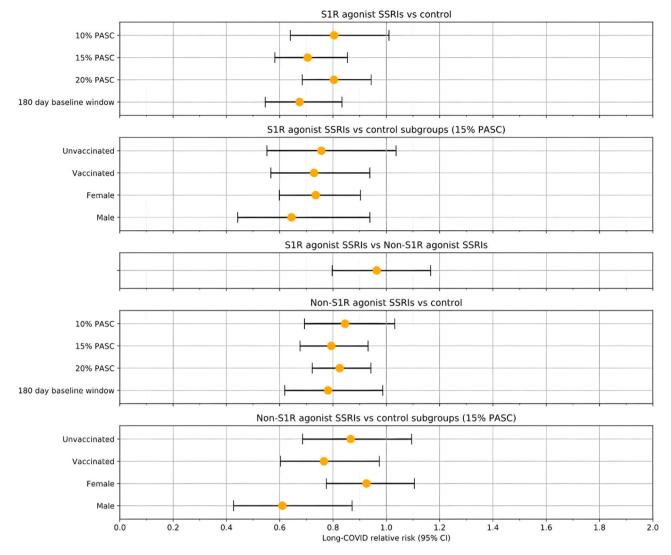


Fig. 4. Associations between baseline S1R agonist and non-S1R agonist SSRI exposure and long-COVID for different patient groups and model specifications.

warranted.

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#### **Declaration of Competing Interest**

The authors hereby declare no conflicting interests pertaining to the material in this manuscript.

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Authorship was determined using ICMJE recommendations.

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- Design: HS, DKS.
- Analysis: HS, ATG, KH.
- Manuscript drafting: HS, DKS, KH.
- Critical revision of manuscript: DKS, KH, HS, ATG, NH, SGM, MGK, KG.

### The N3C Consortium

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#### Data sharing statement

The analyses described in this publication were conducted with data or tools accessed through the NCATS N3C Data Enclave covid.cd2h.org/enclave and supported by CD2H - The National COVID Cohort Collaborative (N3C) IDeA CTR Collaboration 3U24TR002306–04S2 NCATS U24 TR002306. This research was possible because of the patients whose information is included within the data from participating organizations (covid.cd2h.org/dtas) and the organizations and scientists (covid.cd2h.org/duas) who have contributed to the on-going development of this community resource. Enclave data is protected, and can be accessed for COVID-related research with an approved (1) IRB protocol and (2) Data Use Request (DUR). Enclave and data access instructions can be found at <a href="https://covid.cd2h.org/for-researchers">https://covid.cd2h.org/for-researchers</a>; all code used to produce the analyses in this manuscript is available within the N3C Enclave to users with valid login credentials to support reproducibility.

### Data Partners with Released Data

The following institutions whose data is released or pending:

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#### Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.csbj.2023.12.045.

#### References

- Coronavirus disease (COVID-19): Post COVID-19 condition. https://www.who.int/ news-room/questions-and-answers/item/coronavirus-disease-(covid-19)-postcovid-19-condition.
- [2] Taquet M, et al. Incidence, co-occurrence, and evolution of long-COVID features: a 6-month retrospective cohort study of 273,618 survivors of COVID-19. PLoS Med 2021;18:e1003773.
- [3] Nalbandian A, et al. Post-acute COVID-19 syndrome. Nat Med 2021;27:601-15.
- [4] Huang, Y. et al. COVID Symptoms, Symptom Clusters, and Predictors for Becoming a Long-Hauler: Looking for Clarity in the Haze of the Pandemic. 2021.03.03.21252086 Preprint at https://doi.org/10.1101/2021.03.03.21252086 (2021).
- [5] Michelen M, et al. Characterising long COVID: a living systematic review. BMJ Glob Health 2021;6:e005427.
- [6] Canas, L.S. et al. Profiling post-COVID syndrome across different variants of SARS-CoV-2. 2022.07.28.22278159 Preprint at https://doi.org/10.1101/2022.07.28.22278159 (2022).
- [7] Chaplin S. Summary of joint guideline on the management of long COVID. Prescriber 2021;32:33–5
- [8] Simon, M.A., Luginbuhl, R.D. & Parker, R. Reduced Incidence of Long-COVID Symptoms Related to Administration of COVID-19 Vaccines Both Before COVID-19 Diagnosis and Up to 12 Weeks After. 2021.11.17.21263608 Preprint at https://doi. org/10.1101/2021.11.17.21263608 (2021).
- [9] Kuodi, P. et al. Association between vaccination status and reported incidence of post-acute COVID-19 symptoms in Israel: a cross-sectional study of patients tested between March 2020 and November 2021. 2022.01.05.22268800 Preprint at http s://doi.org/10.1101/2022.01.05.22268800 (2022).
- [10] Mahase E. Covid-19: one in eight adults develops long covid symptoms, study suggests. BMJ 2022;378:o1946.
- [11] Komaroff AL, Lipkin WI. Insights from myalgic encephalomyelitis/chronic fatigue syndrome may help unravel the pathogenesis of postacute COVID-19 syndrome. Trends Mol Med 2021:27:895–906.
- [12] Paul BD, Lemle MD, Komaroff AL, Snyder SH. Redox imbalance links COVID-19 and myalgic encephalomyelitis/chronic fatigue syndrome. Proc Natl Acad Sci USA 2021:118:e2024358118.
- [13] Wood E, Hall KH, Tate W. Role of mitochondria, oxidative stress and the response to antioxidants in myalgic encephalomyelitis/chronic fatigue syndrome: a possible approach to SARS-CoV-2 'long-haulers'? Chronic Dis Transl Med 2020;7:14–26.
- [14] Jacobs JJL. Persistent SARS-2 infections contribute to long COVID-19. Med Hypotheses 2021;149:110538.
- [15] Swank, Z., Senussi, Y., Alter, G. & Walt, D.R. Persistent circulating SARS-CoV-2 spike is associated with post-acute COVID-19 sequelae. 2022.06.14.22276401 Preprint at https://doi.org/10.1101/2022.06.14.22276401 (2022).
- [16] Kell DB, Laubscher GJ, Pretorius E. A central role for amyloid fibrin microclots in long COVID/PASC: origins and therapeutic implications. Biochem J 2022;479: 537-59
- [17] Chertow, D. et al. SARS-CoV-2 infection and persistence throughout the human body and brain. (2021) doi:10.21203/rs.3.rs-1139035/v1.
- [18] Leung WF, et al. COVID-19 in an immunocompromised host: persistent shedding of viable SARS-CoV-2 and emergence of multiple mutations: a case report. Int J Infect Dis 2022;114:178–82.
- [19] Zhang Y, et al. Prevalence and persistent shedding of fecal SARS-CoV-2 RNA in patients with COVID-19 infection: a systematic review and meta-analysis. Clin Transl Gastroenterol 2021;12:e00343.
- [20] Zhang L, et al. Reverse-transcribed SARS-CoV-2 RNA can integrate into the genome of cultured human cells and can be expressed in patient-derived tissues. Proc Natl Acad Sci USA 2021;118:e2105968118.
- [21] Peluso MJ, et al. Markers of immune activation and inflammation in individuals with postacute sequelae of severe acute respiratory syndrome coronavirus 2 infection. J Infect Dis 2021;224:1839–48.
- [22] Patterson BK, et al. Immune-based prediction of COVID-19 severity and chronicity decoded using machine learning. Front Immunol 2021;12.
- [23] Su Y, et al. Multiple early factors anticipate post-acute COVID-19 sequelae. e20 Cell 2022;185:881–95. e20.
- [24] Vijayakumar B, et al. Immuno-proteomic profiling reveals aberrant immune cell regulation in the airways of individuals with ongoing post-COVID-19 respiratory disease. e5 Immunity 2022;55:542–56. e5.
- [25] Di Rosso ME, Palumbo ML, Genaro AM. Immunomodulatory effects of fluoxetine: a new potential pharmacological action for a classic antidepressant drug? Pharmacol Res 2016;109:101–7.
- [26] Gobin V, Van Steendam K, Denys D, Deforce D. Selective serotonin reuptake inhibitors as a novel class of immunosuppressants. Int Immunopharmacol 2014;20: 149. E6
- [27] Szałach ŁP, Lisowska KA, Cubała WJ. The influence of antidepressants on the immune system. Arch Immunol Ther Exp] 2019;67:143–51.
- [28] Kostadinov I, et al. Study on anti-inflammatory and immunomodulatory effects of fluoxetine in rat models of inflammation. Eur J Inflamm 2015;13:173–82.
- [29] Taler M, et al. Immunomodulatory effect of selective serotonin reuptake inhibitors (SSRIs) on human T lymphocyte function and gene expression. Eur Neuropsychopharmacol J Eur Coll Neuropsychopharmacol 2007;17:774–80.
- [30] Taler M, Gil-Ad I, Korob I, Weizman A. The immunomodulatory effect of the antidepressant sertraline in an experimental autoimmune encephalomyelitis mouse model of multiple sclerosis. Neuroimmunomodulation 2011;18:117–22.
- [31] Duerschmied Daniel, Suidan Georgette L, Demers Melanie, Herr Nadine, Carbo Carla, Brill Alexander, et al. Platelet serotonin promotes the recruitment of

- neutrophils to sites of acute inflammation in mice. Blood 2013;121(6):1008–15. https://doi.org/10.1182/blood-2012-06-437392.
- [32] Wong Andrea C, Devason Ashwarya S, Umana Iboro C, Cox Timothy O, Dohnalová Lenka, Litichevskiy Lev, et al. Serotonin reduction in post-acute sequelae of viral infection. Cell 2023;186(22). https://doi.org/10.1016/j. cell.2023.09.013.
- [33] Hashimoto K. Repurposing of CNS drugs to treat COVID-19 infection: targeting the sigma-1 receptor. Eur Arch Psychiatry Clin Neurosci 2021;271:249–58.
- [34] Müller JK, Müller WE. Psychotropic drugs: therapeutic perspectives against Covid-19. Psychopharmakotherapie 2021:102–10.
- [35] Rosen DA, et al. Modulation of the sigma-1 receptor–IRE1 pathway is beneficial in preclinical models of inflammation and sepsis. Sci Transl Med 2019;11:eaau5266.
- [36] Hashimoto K. Overview of the potential use of fluvoxamine for COVID-19 and long COVID. Discov Ment Health 2023;3:9. https://doi.org/10.1007/s44192-023-00036-3.
- [37] Ishima T, Fujita Y, Hashimoto K. Interactions of new antidepressants with sigma-1 receptor chaperones and their potentiation of neurite outgrowth in PC12 cells. Eur J Pharmacol 2014;727:167–73.
- [38] Strawbridge R, et al. Inflammation and clinical response to treatment in depression: a meta-analysis. J Eur Coll Neuropsychopharmacol 2015;25:1532–43.
- [39] Hannestad J, DellaGioia N, Bloch M. The effect of antidepressant medication treatment on serum levels of inflammatory cytokines: a meta-analysis. Neuropsychopharmacol Publ Am Coll Neuropsychopharmacol 2011;36:2452–9.
- [40] Song C, Halbreich U, Han C, Leonard BE, Luo H. Imbalance between pro- and antiinflammatory cytokines, and between Th1 and Th2 cytokines in depressed patients: the effect of electroacupuncture or fluoxetine treatment. Pharmacopsychiatry 2009;42:182–8.
- [41] Dahl J, et al. The plasma levels of various cytokines are increased during ongoing depression and are reduced to normal levels after recovery. Psycchoneuroendocrinology 2014;45:77–86.
- [42] Brunoni AR, et al. Cytokines plasma levels during antidepressant treatment with sertraline and transcranial direct current stimulation (tDCS): results from a factorial, randomized, controlled trial. Psychopharmacology 2014;231:1315–23.
- [43] Pfaff ER, et al. Identifying who has long COVID in the USA: a machine learning approach using N3C data. Lancet Digit Health 2022;4:e532–41.
- [44] Stürmer T, Wyss R, Glynn RJ, Brookhart MA. Propensity scores for confounder adjustment when assessing the effects of medical interventions using nonexperimental study designs. J Intern Med 2014;275(6):570–80. https://doi. org/10.1111/joim.12197.
- [45] Reis G, et al. Effect of early treatment with fluvoxamine on risk of emergency care and hospitalisation among patients with COVID-19: the TOGETHER randomised, platform clinical trial. Lancet Glob Health 2022;10:e42–51.
- [46] Hoertel N, et al. Association between antidepressant use and reduced risk of intubation or death in hospitalized patients with COVID-19: results from an observational study. Mol Psychiatry 2021;26:5199–212.

- [47] Oskotsky T, et al. Mortality risk among patients with COVID-19 prescribed selective serotonin reuptake inhibitor antidepressants. JAMA Netw Open 2021;4: e2133090.
- [48] Brennan BP, Schnabel J, Pope HG, Hudson JI. Antidepressant use and risk of intubation or death in hospitalized patients with COVID-19: a retrospective cohort study of clinical effectiveness. Front Psychiatry 2022;13.
- [49] Lenze EJ, et al. Fluvoxamine vs placebo and clinical deterioration in outpatients with symptomatic COVID-19: a randomized clinical trial. JAMA 2020;324: 2292–300
- [50] Bramante CT, et al. Randomized trial of metformin, ivermectin, and fluvoxamine for Covid-19. New Engl J Med 2022;387:599–610.
- [51] McCarthy, M.W. et al. Fluvoxamine for Outpatient Treatment of COVID-19: A Decentralized, Placebo-controlled, Randomized, Platform Clinical Trial. 2022.10.17.22281178 Preprint at https://doi.org/10.1101/2022.10.17.22281178 (2022)
- [52] Naggie, Susanna. Effect of higher-dose fluvoxamine vs placebo on time to sustained recovery in outpatients with mild to moderate COVID-19: A randomized clinical trial, 2023. https://doi.org/10.1101/2023.09.12.23295424.
- [53] Foletto Vitória Segabinazzi, da Rosa Taciéli Fagundes, Serafin Marissa Bolson, Hörner Rosmari. Selective serotonin reuptake inhibitor (SSRI) antidepressants reduce COVID-19 infection: prospects for use. Eur J Clin Pharmacol 2022;78(10): 1601–11. https://doi.org/10.1007/s00228-022-03372-5.
- [54] Glebov Oleg O, Mueller Christoph, Stewart Robert, Aarsland Dag, Perera Gayan. Antidepressant drug prescription and incidence of COVID-19 in mental health outpatients: a retrospective cohort study. BMC Med 2023;21(1). https://doi.org/ 10.1186/x12916-023-02877-9
- [55] Stauning Marius Ahm, Gür Dogukan Jesper, Torp-Pedersen Christian, Tingleff Jens. Covid-19 mortality among selective serotonin reuptake inhibitor users—results from a nationwide cohort. Clin Microbiol Infect 2023;29(8):1075–82. https://doi. org/10.1016/j.cmi.2023.04.028.
- [56] Visos-Varela Irene, Zapata-Cachafeiro Maruxa, Piñeiro-Lamas María, Carracedo-Martínez Eduardo, Saez Marc, Herdeiro María Teresa, et al. Repurposing selective serotonin reuptake inhibitors for severity of COVID-19: a population-based study. Eur Neuropsychopharmacol 2023;71:96–108. https://doi.org/10.1016/j.euroneuro.2023.03.011.
- [57] Tsiakalos Aristotelis, Ziakas Panayiotis D, Polyzou Eleni, Schinas Georgios, Akinosoglou Karolina. Early fluvoxamine reduces the risk for clinical deterioration in symptomatic outpatients with Covid-19: a real-world, retrospective, before–after analysis. Microorganisms 2023;11(8):2073. https://doi.org/10.3390/ microorganisms11082073.
- [58] Rus Carla P, de Vries Bert E, de Vries Ingmar E, Nutma Idelette, Kooij JJ. Treatment of 95 post-covid patients with ssris. Sci Rep 2023;13(1). https://doi.org/10.1038/ s41598-023-45072-9.