Effect of water, sanitation, and hygiene interventions on enteropathogen detection in the environment: an individual data meta-analysis

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## Abstract

**Background:** Drinking water, sanitation, and hygiene (WASH) improvements are considered cornerstones to reduce diarrheal disease in low-income countries. However, recent trials have found no or mixed effects of household- and community-level WASH interventions on child health. Measuring pathogens in the environment can help investigate if limited health effects occur because WASH interventions do not sufficiently reduce environment contamination. **Methods:** We conducted a systematic review and individual participant data meta-analysis to assess the effects of WASH interventions on enteropathogens and microbial source tracking (MST) markers in environmental samples. We used covariate-adjusted regression models with robust standard errors to estimate intervention effects and pooled results across studies. **Findings:** We identified and received data from five randomized or quasi-experimental studies. Environmental sampling was primarily focused on sanitation interventions. Most individual studies found no effects on pathogens or MST markers in environmental samples, including drinking water, hands, soil and flies. We found a small but significant reduction in the prevalence of any pathogen in any sample type, with a pooled prevalence ratio of 0.94 (95% CI: 0.89, 0.99). There was no overall effect on MST markers, and no consistent differences in intervention effects by season, animal presence, urbanicity, study design, or intervention uptake. **Interpretation:** Few trials have measured pathogens or host-specific fecal markers in the environment. The small effect of sanitation interventions on pathogens in the environment in this study is consistent with the lack of health impact in sanitation trials. **Funding:** The Bill & Melinda Gates Foundation.

## Research in context

**Evidence before this study.** Children in areas with poor drinking water, sanitation, and hygiene conditions (WASH) experience increased diarrheal disease and reduced growth. Recent rigorous intervention studies on WASH improvements have shown mixed effects on reducing diarrheal disease in children and no improvements in child growth. Quantifying the effect of WASH improvements on enteric pathogens in environmental samples may help elucidate if interventions successfully interrupt the causal pathway between poor WASH, environmental exposure to fecal pathogens, and child health. Most previous studies and meta-analyses on the effect of WASH interventions on fecal contamination in the environment have focused on fecal indicator bacteria (FIB). Studies have shown FIB reductions in water and on hands due to water treatment and handwashing, respectively, but no effects from sanitation. However, limitations of FIB as predictors of enteropathogens and disease risk limit the interpretation of these findings. We conducted a systematic review and individual participant data meta-analysis of WASH intervention studies that measured in the environment specific enteropathogens or microbial source tracking (MST) markers, which indicate the human or animal host of fecal microorganisms. We tested if interventions reduced the prevalence and abundance of these targets in drinking water, hand rinse, soil, and fly samples.

**Added value of this study.** We obtained data from 5 out of 6 eligible intervention studies identified in our systematic review that measured enteropathogens and MST markers in environmental samples. Most individual studies indicated a protective effect of interventions on the prevalence of individual pathogens and MST markers, but most estimates were not statistically significant due to small sample sizes and rare detection of some of the targets. The individual participant data meta-analysis design of our study allowed us to detect a small but significant reduction in the prevalence of any type of pathogen in any type of sample by pooling across all studies. There was no overall intervention effect on the prevalence of MST markers. This study takes advantage of recently used methods to enumerate enteropathogens and host-specific fecal markers in a range of environmental samples, including understudied reservoirs such as soil. We provide the first synthesis of evidence of the effect of WASH interventions on these important targets in the domestic environment.

**Implications of all the available science.** The environmental sampling in the studies in our review were mostly focused on sanitation interventions. The small reduction we observed in pathogen prevalence in the environment when pooled across all studies may explain the small effect the interventions had on child health. These findings also support previous findings of no effect from sanitation interventions on fecal indicator bacteria in the environment, further demonstrating the insufficiency of non-piped sanitation solutions in reducing fecal contamination in the environment and the importance of animal sources of feces. Possibly, high coverage of more intensive WASH interventions like safely managed water and sanitation are needed to reduce environmental contamination enough to improve child health. We note that only a small number of trials met our inclusion criteria and only a subset of households were environmentally sampled in each study. Pathogen targets and diagnostic methods varied by study. Future research would benefit from environmental sampling following implementation of a more diverse set of WASH interventions using a standardized set of laboratory methods to enumerate a common range of pathogen targets.

## Introduction

Water, sanitation and hygiene (WASH) improvements are often assumed to reduce childhood enteric infections, subsequent diarrheal disease and growth faltering by reducing exposure to fecal-orally transmitted pathogens in the environment. Until recently, trials of WASH interventions have primarily focused on documenting health outcomes such as caregiver-reported diarrhea without quantifying intermediate outcomes along the causal chain, such as detection of pathogens in environmental samples and in human biological specimens. Without measuring these causal intermediates, trials are limited to a “black box” understanding, where underlying mechanisms of interventions are unknown and investigators can only speculate about reasons for intervention success or failure. Inspecting the causal chain is especially important given the small or null effects on child diarrhea and growth reported in recent well-conducted trials of WASH interventions.1–3

Pathogens are transmitted from the feces of infected individuals to new hosts through interconnected environmental pathways. Studies assessing the effect of WASH interventions on environmental contamination have primarily focused on drinking water (and, to a smaller extent, hands and food) while other pathways such as soil and surfaces in the domestic environment have received less attention.4 Household water treatment and handwashing have been associated with reduced fecal contamination of drinking water5 and hands,6,7 respectively while sanitation interventions have had limited impact on environmental fecal contamination, including drinking water, hands, objects, surfaces, soil and flies.4

These studies have mostly relied on measuring indicator organisms such as *E. coli* in the environment as a proxy for pathogens, and while *E. coli* has been shown to correlate with risk of diarrhea,8 fecal indicator bacteria (FIB) like *E. coli* correlate poorly with pathogens in the environment10 because both humans and animals shed FIB8 and FIB can originate from non-fecal sources.9 Recent applications of in nucleic-acid molecular methods to environmental sampling in low-income settings now allow for detection of a range of enteropathogens in human biological specimens and environmental samples,11,12 as well as distinction between human vs. animal fecal sources through microbial source tracking (MST).13 Using a systematic review and individual participant data (IPD) meta-analysis where we combined sample-level observations of environmental sampling, we aimed to assess the effect of WASH interventions on detection of specific enteropathogens and human- and animal-specific MST markers in the domestic environment.

## Methods

### Search methods

We conducted a systematic literature search to identify WASH intervention studies in low-income countries that have measured pathogens and/or MST markers in environmental samples as well as at least one of the following health outcomes in children: caregiver-reported diarrhea, growth, or pathogen detection in stool. We focused on studies with environmental as well as child health measurements in order to assess: (1) the impact of WASH interventions on environmental contamination with human enteropathogens and fecal markers, (2) associations between measures of environmental contamination and child health outcomes, and (3) examine the hypothesized causal pathway from WASH intervention to child health mediated through reduced environmental contamination. In this paper, we present findings for the first aim. The analyses for the latter two aims are presented in a companion paper [Mertens et al. 2021 in prep].

We searched the PubMed, Embase, CAB Direct Global Health, Agricultural & Environmental Science Database, Web Of Science, and Scopus databases. Search terms are listed in Supplementary Table S1 and our PubMed search string is listed in Supplementary Table S2. We only included studies published after 2000 to capture more recently developed advanced pathogen and MST detection methods. We limited our search to studies published in English. We included studies meeting the following inclusion criteria: 1) prospective studies with a water, sanitation, or hygiene intervention and concurrent control (i.e., randomized controlled trial, matched cohort, controlled before-and-after study), consistent with prior WHO burden of disease reviews ,14,15 2) measured pathogens and/or MST markers in environmental samples, and 3) measured child anthropometry, diarrheal disease, or pathogen-specific infections for use in a companion manuscript [Mertens et al. 2021 in prep]. We excluded studies that only measured fecal indicator bacteria such as coliforms or *E. coli*. One reviewer (AM) screened the abstracts of studies according to our inclusion/exclusion criteria, and two independent reviewers (AM, RT) examined the full texts of short-listed articles.

### Data Collection and Analysis

For each study eligible for inclusion in the IPD meta-analysis, we contacted the corresponding authors to request individual data on the presence and abundance of pathogen and MST markers in environmental samples, child health outcomes, and potentially confounding baseline characteristics, including socioeconomic and demographic indicators. The IPD approach using individual environmental and covariate data offers advantages over a traditional meta-analysis by allowing harmonized statistical analysis across different studies and by allowing for conducting and pooling novel subgroup analyses. All personal identifiers were removed from the data, including GPS locations, and sampling dates were coarsened to a monthly resolution prior to being shared. If the corresponding author was unwilling to share individual data, that study was excluded from our analysis.

Our primary outcomes were the detection of any enteropathogen and any MST markers in any type of environmental sample. We analyzed prevalence separately for each sample type (e.g., water, hands, soil, flies) and also as a composite measure indicating detection of a given target (any pathogen or MST marker) in any sample type collected from the same compound during the same sampling round. Secondary outcomes included the prevalence of specific pathogen types (any viruses, any bacteria, any protozoa, any helminths), the prevalence of MST markers from specific host types (general, human, animal), and the prevalence and abundance of individual enteropathogens and MST markers.

We compared each outcome between the WASH intervention and control arms of the included studies. We estimated prevalence ratios using modified Poisson regressions.16 For abundance outcomes, we used linear regressions to estimate differences in log-transformed gene copies and negative binomial regressions to estimate count differences in soil-transmitted helminth (STH) egg counts. Because of repeated sampling or clustered designs in some studies, we used the Huber Sandwich Estimator to calculate robust standard errors.17 For abundance measures, we imputed values for samples below the limit of quantification (LOQ) and the limit of detection (LOD). For samples below the LOD, we used half the LOD and for samples below the LOQ, we used the midpoint between the LOD and LOQ for the imputation. To avoid analyzing targets where most of the data were imputed, we only included targets where at least 50% of samples were within the quantifiable range in our analysis.

All analyses were adjusted for potential confounders. While estimated intervention effects from randomized controlled trials should be unconfounded, covariate adjustment may increase statistical efficiency, and improve exchangeability with matched cohorts and non-randomized trials.18 Covariates were prescreened using likelihood ratio tests, and only variables associated with the outcome with a p-value < 0.2 were included in the model for each outcome. We included the following variables in the prescreening set if they were measured within an included study: number of people in the household, age and education of primary caregiver in the household, asset-based household wealth, number of rooms, construction materials (walls, floor, roof), access to electricity, land ownership and if anyone in the household works in agriculture. For rare binary outcomes, we only included one independent variable (including the treatment arm) per 10 positive samples, or per 10 negative samples if <50% of samples were negative. We therefore did not estimate prevalence ratios for any targets with fewer than 10 positive or negative values for a given sample type, or with fewer than two positive or negative values per study arm for a given sample type. Targets with <10 negatives were also excluded from the composite “any pathogen” and “any MST marker” variables to avoid generating sparse cells; targets with <10 positives were included in the composite variables.

Given the heterogeneity in study settings (e.g., local WASH conditions, climate, urbanization, population density, region-specific infectious disease patterns, intervention designs), we reported individual study-specific estimates for all analyses. For targets where data were available from four or more studies, we tested for heterogeneity in estimates using Cochran’s Q-test.19 If there was no significant heterogeneity (p-value>0.2), we pooled estimates using fixed-effects models. If there was evidence for heterogeneity but there was qualitative support for combining studies, we pooled estimates using random-effects models.

We conducted subgroup analyses by season (dry vs. wet), animal ownership (at least one vs. no animal owned) and pathogens with zoonotic vs. no zoonotic transmission. The wet season for each study was defined as the 6 months of highest average rainfall, obtained from <https://www.weather-atlas.com/>.20 The pathogens we considered zoonotic were *Campylobacter, Salmonella, Yersinia enterocolitica, C. difficile, Cryptosporidium, Giardia* and *Ascaris*.21 Studies included detected virulence genes associated with specific *E. coli* pathotypes (EAEC, EPEC/EHEC, STEC, EIEC, ETEC). Among these, we classified STEC and EPEC (due to atypical EPEC) as zoonotic.21. We assessed effect modification by examining the p-values on the interaction terms between the treatment and the indicator variable for the subgroup in the regression models; a p-value <0.2 was considered evidence of effect modification. There was no heterogeneity in study setting within any individual study as each study was conducted either in a primarily rural or primarily urban setting. We therefore explored heterogeneity by study setting by pooling estimates separately for rural vs. urban studies and comparing the pooled estimates with Wald tests. We also separately pooled estimates from randomized and quasi-experimental studies and from studies with high vs. low intervention uptake and compared pooled estimates with Wald tests.

All analyses were conducted in R 4.0, and analysis scripts are publicly available (<https://github.com/amertens/wash-ipd>). The systematic review search strategies and the analysis plan were pre-registered on Open Science Framework (<https://osf.io/8sgzn/>). Our PRISMA checklist can be found in Supplementary Table S3.

## Results

### Search results and data acquisition

The systematic review was conducted on 1/19/2021 and returned 3,376 results after removing duplicates. Of these, 3,253 were excluded by abstract screening, and of the 125 short-listed studies, eight met the inclusion criteria after full-text screening. The eight publications reported findings from six unique intervention studies: WASH Benefits Bangladesh and Kenya trials,22 the Maputo Sanitation (MapSan) study in Mozambique,23 the Gram Vikas study in India,24 the Odisha Total Sanitation Campaign trial in India,25 and the CHoBI7 trial in Bangladesh25 (Table 1). Data were obtained from all studies except the CHoBI7 trial.

Seven unique studies on pathogens or MST markers in the environment were nested within the five intervention studies with available data. Three separate publications reported environmental results from the WASH Benefits Bangladesh study .26–28 These publications focused on samples collected from different subsets of trial participants at different times; therefore, we report results from these three studies separately. We also received additional unpublished data from the MapSan study, which we include along with the published results from this study.29 For the Odisha Total Sanitation Campaign trial, only village-level source water quality data were shared.

### Characteristics of included studies

The WASH Benefits trials were cluster-randomized, multi-armed factorial designed trials of water, hygiene, sanitation, and nutrition interventions in rural Kenya and Bangladesh, but pathogens and MST markers in environmental samples were only measured in the control and sanitation arms. The sanitation intervention included the construction of a double-pit pour-flush improved latrine and provision of a child potty and sani-scoop for feces removal. The MapSan study was a controlled before-and-after study of a decentralized sanitation intervention in urban Mozambique. The intervention entailed the construction of pour-flush toilets that drain to septic tanks, shared by a minimum of 12 people. The intervention delivery was not randomized, but control sites were matched to intervention sites on compound size and time of enrollment. The Gram Vikas study was a matched cohort evaluating the effect of a piped water and sanitation intervention provided by the Gram Vikas non-governmental organization in rural India. Gram Vikas built a water tank and piped distribution system, and provided materials for the construction of pour-flush toilets in each household. After each household in the village completed latrine construction, the water system was turned on for the whole village. The control villages were matched to intervention villages on pre-intervention characteristics. The Total Sanitation Campaign study in Odisha, India, was a cluster-randomized trial assessing the Government of India’s Total Sanitation Campaign, which included promoting the construction of a pour-flush latrine with a single pit and Y-joint for a future second pit, which was subsidized post hoc at the household level by government funding.

Intervention uptake was high in most studies; 97% of intervention compounds in WASH Benefits Bangladesh had a latrine with a functional water seal compared to 31% of controls, 78% percent of intervention compounds in WASH Benefits Kenya had improved latrines compared to 20% of controls, 85% of intervention compounds in Gram Vikas had improved latrines compared to 18% of controls, and 86% percent of intervention compounds in MapSan had clean latrines compared to 45% of controls. Odisha had the lowest uptake, with 38% percent of intervention compounds having functional latrines at endline compared to 10% of controls.

### Sample types and targets in included studies

Samples were collected from 4 months26 to 6-10 years24 after intervention delivery, with most studies collecting samples at 1-2 years post intervention (Table 1). Types of samples collected included source and stored drinking water, child and mother hand rinses (or swabs?), soil from the courtyard, household and latrine areas, food, and flies caught in the compound’s latrine and kitchen areas. The number of environmental samples in individual studies varied from 6030 to 345231. The pooled dataset across all studies included 12,199 samples, with a total of 41,692 observations for pathogen/MST marker prevalence.

The studies measured a range of bacterial, viral, protozoan and helminthic pathogens, including pathogenic *E. coli, V. cholerae, Shigella, Campylobacter, Salmonella, Yersinia, C. difficile*, rotavirus, norovirus, sapovirus, adenovirus, astrovirus, enterovirus, *Cryptosporidium, Giardia, Entamoeba histolytica, Ascaris lumbricoides and Trichuris trichiura* (Table S4). The MST markers included general (GenBac3, BacUni), human (HumM2, HF183, BacHum, *M. smithii*), animal (BacCan, BacCow), ruminant (BacR) and avian (GFD) fecal markers (Tables S5). Most studies used quantitative polymerase chain reaction (qPCR) or reverse-transcriptase (RT)-qPCR to quantify these targets (Table 1). One study used slide agglutination serotyping to detect *V. cholerae* and *Shigella*.31 One study detected *Cryptosporidium* oocysts and *Giardia* cysts using direct fluorescent antibody (DFA) microscopy.30 Two studies used microscopy to enumerate STH eggs.

Many targets had low or no variation, with 26/275 target-sample combinations having no variation in prevalence (all samples negative), and 62/275 of combinations having too little variation to estimate a prevalence ratio (<10 positive or negative samples). Among these sparse combinations, most (88.7%) had too few positive samples, and 11.3% had too few negative samples. Specifically, the BacCow animal fecal marker in Odagiri et al. 2016, the GenBac3 general fecal marker in Boehm et al. 2016, and the human Bacteroides marker in Holcomb et al. 2020 had close to 100% prevalence. We excluded these three targets from the aggregated “any MST marker” variable to allow estimation of prevalence ratios. Overall, 187/275 sample-target combinations had sufficient variability to be individually included in our meta-analysis. Among these, the prevalence of pathogens ranged from 2.3% for *Giardia* in mothers’ hand rinses27 to 61.7% for *Ascaris* in soil32. The prevalence of MST markers ranged from 2.4% for HumM2 on child hands26 to 96.7% for BacCow on mothers’ hands.27

### Intervention effects on the prevalence of any enteropathogen and any MST marker

The interventions generally decreased the prevalence of pathogens and MST markers but the confidence intervals for prevalence ratios often crossed the null, with 73.3% (137/187) of study-specific intervention effects being protective but non-significant and 6.4% (12/187) being protective and significant. Interventions decreased the prevalence of any pathogen in any sample type in all specific studies except for Boehm et al. 2016 but the decrease was only significant in Fuhrmeister et al. 2020 with an adjusted PR of 0.92 (95% CI: 0.86, 0.98). The interventions had no significant effects on the prevalence of any pathogen among individual sample types (Figure 1). Overall, study-specific estimates were largely homogeneous, with no significant Cochran’s Q-tests of homogeneity. Therefore, we pooled estimates using fixed-effects models. When pooled across all studies, there was a small reduction in the prevalence of any pathogen detected in any sample type, with an adjusted pooled PR of 0.94 (95% CI: 0.89, 0.99) (Figure 1). Interventions had no effect on the prevalence of any MST marker in any sample type (adjusted pooled PR= 0.99 (95% CI: 0.95, 1.04)) or within specific sample types.

### Intervention effects on the prevalence of pathogen types and individual pathogens

Interventions reduced the prevalence of any bacterial pathogens in any sample type, with 11 out of 13 study- and sample-specific prevalence ratios indicating a protective effect (Figure 2), though the effect was only significant in any sample type in Fuhrmeister et al. 2020, with an adjusted PR of 0.92 (95% CI: 0.86, 0.99). Pooled across all studies, interventions reduced the prevalence of any bacteria in any sample type, with an adjusted pooled PR of 0.91 (95% CI: 0.85, 0.97). Interventions only reduced STH in soil (adjusted PR: 0.60 (95% CI: 0.42, 0.85)) and in any sample type (adjusted PR: 0.64 (95% CI: 0.45, 0.93), Figure 2) in Holcomb et al. 2020. Interventions did not significantly reduce the presence of viruses or protozoa in any sample type or within specific sample types, though point estimates from individual studies were protective except for viruses in child and mother’s hand rinses (Figure 2). Among specific pathogens, interventions reduced the prevalence of pathogenic *E. coli* in any sample type in Fuhrmeister et al. 2020 (adjusted PR: 0.92 (95% CI: 0.86, 0.99)) and of adenovirus (adjusted PR: 0.20 (95% CI: 0.06, 0.63)) and *Shigella* (adjusted PR: 0.32 (95% CI: 0.11, 0.93)) in Holcomb et al. 2020 (Figure S1). These reductions were driven by non-significant reductions in all sample types in Fuhrmeister et al. 2020 and by significant reductions in soil samples in Holcomb et al. 2020.

### Intervention effects on the prevalence of MST marker types and individual markers

Among specific types of MST markers (general, human, animal), effects of interventions were inconsistent and largely null, with 30 out of 54 study-specific prevalence ratios indicating a protective effect (Figure 3). The only significant estimates were observed for any animal MST marker in stored water in Boehm et al. 2016 (adjusted PR=0.67 (95% CI: 0.49, 0.93)), driven primarily by a significant effect of the sanitation intervention on the ruminant (BacR) marker (adjusted PR: 0.60 (95% CI: 0.41, 0.88), Figure S2). There were no other intervention effects on individual MST markers (Figure S2).

### Intervention effects on the abundance of individual pathogens and MST markers

Of all samples, 21.4% had abundances quantified, including STH egg counts in Steinbaum et al. 2019 and Kwong et al. 2021 and gene copies of enteropathogens and MST targets in Boehm et al. 2016, Fuhrmeister et al. 2020, and Holcomb et al. 2020. Of these, 18.7% were below the specific study reported limit of detection, 22.5% were below the study-reported limit of quantification, and 58.7% were in the study-reported range of quantification. Of targets enumerated within specific sample types, only 21.1% had >50% of samples within the range of quantification and were therefore included in our analysis. Table 2 shows the mean log10-transformed abundances stratified by arm as well as covariate-adjusted differences in abundances between the intervention and control arms for these combinations. The interventions had significant effects on the abundance of three MST targets, but not on STH egg counts. The abundance of the general *Bacteroidales* fecal marker (GenBac3) in household soil samples in Boehm et al. 2016 was lower in the intervention arm, with an adjusted log10-transformed difference of -0.20 (95% CI: -0.37, -0.02). The abundance of the BacCow animal marker was lower in mothers’ hand rinses in the intervention arm in Fuhrmeister et al. 2020, with an adjusted log10-transformed difference of -0.28 (95% CI: -0.49, -0.07). However, the sanitation intervention in Holcomb et al. 2020 significantly increased the abundance of human-specific *Bacteroides* in flies caught in latrines (adjusted log10-transformed difference: 0.70 (95% CI: 0.11, 1.28); Table 2). There were not sufficient studies with abundance data to pool estimates.

### Subgroup and adjusted analyses

The wet season was May-October in Bangladesh and India and November-April in Mozambique. Western Kenya has two distinct periods of high rain, so the wet season was defined as March-May and October-December for WASH Benefits Kenya. Interventions had significantly different effects by season in several study-specific sample types, but the direction of the effects was inconsistent (Figure S3). Interventions decreased the prevalence of any MST marker in child hand rinses and in soil in Boehm et al. 2016, the prevalence of any MST marker in stored water in Fuhrmeister et al. 2020, and the prevalence of any pathogen in soil in Kwong et al. 2021, but only during the wet season. Conversely, interventions increased the prevalence of any pathogen in stored water in Reese et al. 2017 and in stored water (marginally significant) and courtyard soil in Fuhrmeister et al. 2020 in the wet season but had no effect during the dry season.

There was no significant effect of any interventions on any pathogen or any MST marker prevalence when households were stratified by animal presence, but there was one significant interaction in Holcomb et al. 2020, the only urban study, with a non-significant protective effect of the intervention in homes without animals, and a non-significant harmful effect of the intervention in homes with animals (Figure S4). There were no differences in intervention effects on pathogens with possible zoonotic transmission versus pathogens with only human hosts (Figure S5). Samples from compounds with animals did not have a significantly higher prevalence of zoonotic pathogens (35.8%) than households without animals (32.3%). There were also no significant differences in pooled estimates between the one urban study (Holcomb et al. 2020) and the four rural studies (Wald-test p-value for any pathogen in any sample type: 0.75), between randomized trials and quasi-experimental studies (Wald-test p-value: 0.46), or between the four studies with higher intervention uptake compared to the Odisha Total Sanitation Campaign trial (Wald-test p-value: 0.59). Unadjusted estimates did not vary greatly from adjusted estimates (Tables S6-S7, Figures S6-S8).

## Discussion

We obtained individual participant data from five different WASH intervention studies for an IPD analysis reported in 7 individual papers (ref). Despite differences in study settings and intervention designs across studies, results were relatively consistent, with no statistically significant heterogeneity in any of the pooled estimates. There were no studies examining the impact of hygiene or water interventions alone on enteropathogens or MST markers in the environment; Reese et al. 2017 evaluated the effect of a combined water and sanitation intervention on *V. cholerae* and *Shigella* prevalence while the other four studies focused on sanitation interventions. Study-specific intervention effects on the prevalence of pathogens and MST markers were primarily null, though 73.3% of the point estimates were protective for the sample-target combinations included in our analysis. Most study-specific estimates had small sample sizes and rare outcomes due to very low/high prevalence of the targets. When pooled across studies, there was an overall small but significant reduction in pathogen prevalence in any environmental sample among intervention recipients compared to controls but no effect on MST marker prevalence. There were also no overall effects on the abundance of individual pathogens and MST markers but most samples fell outside the quantifiable range and were excluded from analysis. Quantifying the abundance of pathogens or MST markers was useful for assessing intervention effects when a given target was ubiquitously detected, leading to insufficient variation in the binary prevalence variable. For example, we could not estimate a prevalence ratio for the general GenBac3 fecal marker in Boehm et al. 2016 because close to 100% of all samples from both arms had detectable GenBac3, but the intervention decreased the abundance of GenBac3.

The methods used to detect and quantify specific pathogens or MST markers are typically more expensive and complex than measuring fecal indicator bacteria, contributing to the small number of eligible studies to be included in our meta-analysis and a small number of samples analyzed in some of the studies. Across the included studies, the percent of study households that contributed environmental samples pathogens or MST marker analysis ranged from 9%26 to 35%31. DNANucleic-acid(NA)-based diagnostics allowing for the detection of specific enteropathogen genomes overcome the limitations of relying on fecal indictor bacteria but the trade-off can be decreased power to detect intervention effects due to smaller sample sizes. The IPD meta-analysis approach allowed us to detect small intervention effects on pathogen prevalence in the environment that the individual studies were not powered to detect. Advances in technology that reduce the costs of NA-based diagnostics or increase funding for environmental testing of enteropathogens within WASH trials, may more precisely estimate the impact of WASH interventions on environmental contamination.

Domestic animals can contribute to fecal contamination in the environment,33 and have been hypothesized to partly explain why sanitation interventions focused on isolating human fecal matter have achieved limited improvement in child health outcomes.34 MST markers allow differentiating the effect of sanitation improvements on fecal markers from human vs. animal sources. In our analysis, there were no reductions in human-specific MST markers from sanitation improvements, while we observed reduced prevalence of ruminant fecal markers in stored water and reduced abundance of animal fecal markers on mothers’ hands in two studies nested within the WASH Benefits Bangladesh trial. Notably, the sanitation intervention in this trial included a scoop for disposal of child and animal feces and may thus have reduced animal fecal contamination in environmental samples.

Unlike conventional meta-analyses that statistically pool reported summary estimates from different studies, IPD analysis pools raw data from individual studies and then estimates the effect of interest.35 The IPD approach has several advantages over pooling independently derived effect estimates from different studies. While effect estimates are often reported differently across studies (e.g., odds ratio vs. relative risk), individual participant data allow direct derivation of the effect estimate of interest, regardless of how it was reported in the original studies. The statistical analysis approach can also be standardized across studies, including adjusting for baseline factors consistently and using consistent statistical models. One limitation of the IPD approach arises from the heterogeneity of study data. Studies in our review measured different targets using different methods, which limits the comparability. Future research would benefit from systematic and standardized assessment of a broad panel of important enteropathogens, using molecular methods sensitive enough to permit simultaneous detection in both environmental and clinical samples.11 Additionally, not every covariate was measured in every study, and they were measured differently across studies (Table S8). However, because of the randomized or quasi-experimental nature of all the included studies, and the overlap between unadjusted and adjusted estimates, we do not believe there is substantial residual confounding biasing the results (Tables S6-S7). Definitions of effect modifiers also varied by study, as seasonal rainfall patterns vary by location and studies measured different types of animals in the compound as well as different types and numbers of zoonotic and non-zoonotic pathogens. To limit the number of comparisons, we did not evaluate effect modification by different types and numbers of animals, or different wet season definitions. The effect modification analyses involve small sample sizes after stratification, so they may be sensitive to the definitions of the effect modifiers. For example, we defined the wet season as the six rainiest months and found a significant reduction in the prevalence of any STH eggs in soil in the wet season while Kwong et al. 2021 defined the wet season as the five rainiest months in the original publication and found no evidence of effect modification.

We did not correct for multiple comparisons, and pooled estimates would no longer be significant after correction. While individual studies were likely underpowered, the consistency of the protective point estimates of intervention effects across studies, and the small but significant reduction in the prevalence of any pathogen in any sample type in the pooled analysis, indicate that there was a small effect of sanitation interventions on reducing the prevalence of pathogens, but not MST markers, in the environment, regardless of the study setting or the specifics of the sanitation improvements. The small reduction in pathogen prevalence in the environment and lack of effect on MST markers is consistent with the null findings of the parent sanitation studies on child diarrhea,, except for the WASH Benefits Bangladesh trial that found a significant reduction in diarrhea prevalence in the sanitation arm compared to controls.

These findings are also consistent with previous studies that found no effect of sanitation interventions on fecal indicator bacteria in the environment,4 suggesting that null effects are not solely due to limitations of indicator bacteria but rather indicate the insufficiency of basic sanitation solutions in reducing fecal contamination in the environment. Future studies should investigate alternative sanitation modalities that can more effectively interrupt environmental pathogen transmission. Future WASH intervention studies should also assess the effect of water treatment and hygiene interventions across a range of pathogens and MST markers in drinking water, on hands and in food.

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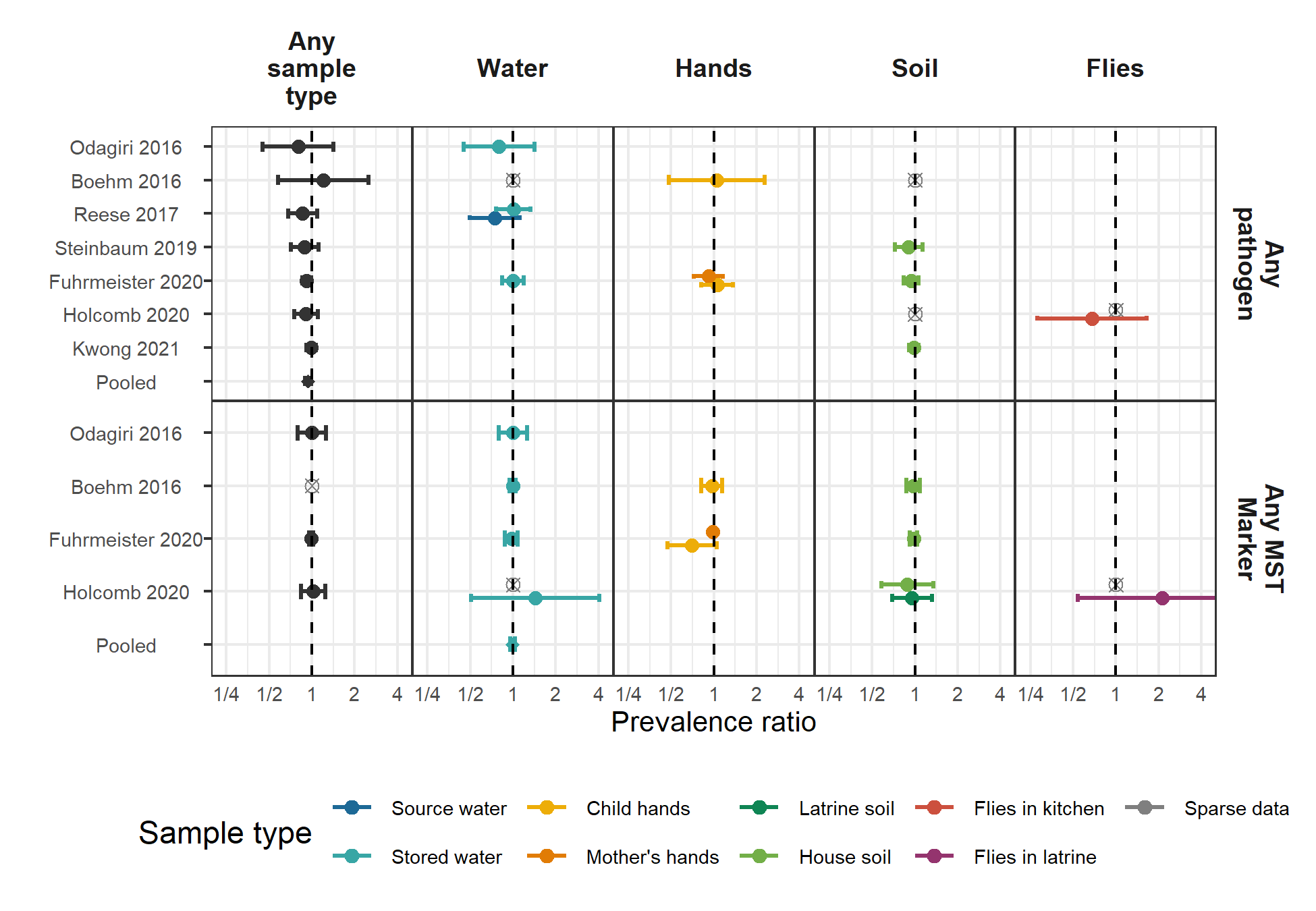
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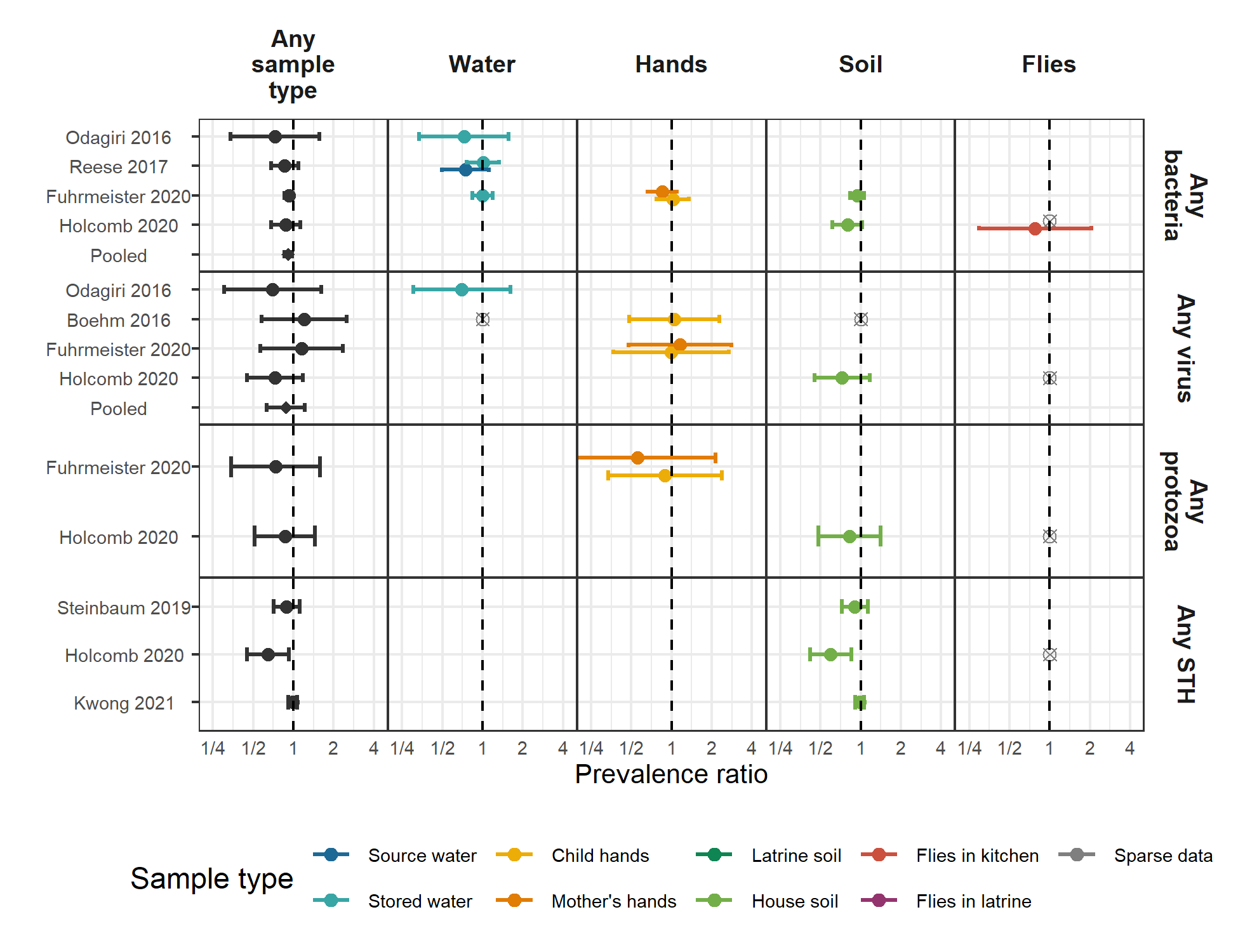
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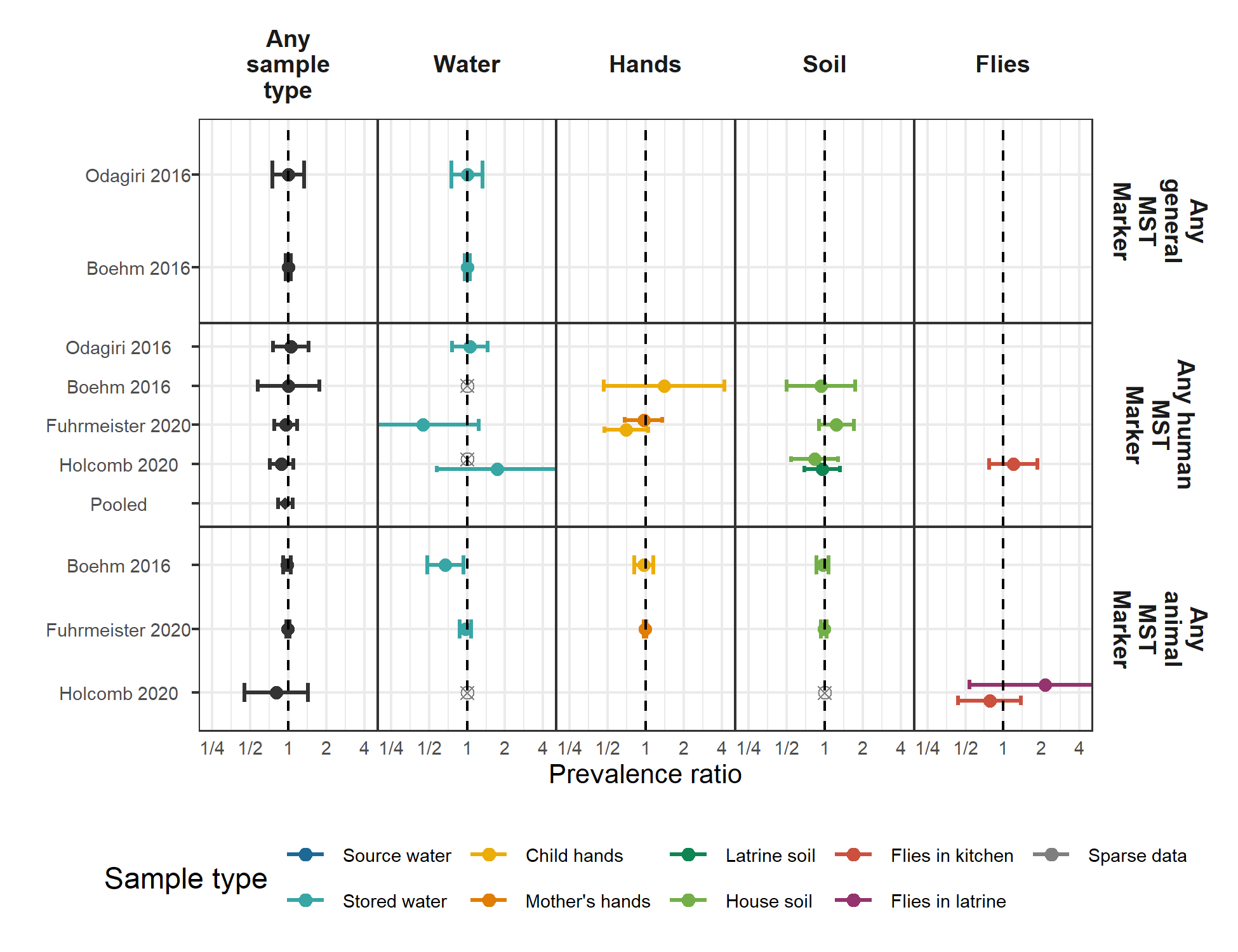
## Figures



**Figure 1.** Forest plots of intervention effects on the prevalence of any enteropathogen and any MST markers in different types of environmental samples. Pooled estimates are presented when there are four or more study-specific estimates for a specific sample type and target combination and are denoted with diamond-shaped points. Grey crossed points denote data that were too sparse to estimate a prevalence ratio (i.e., <10 positive or negative observations). Samples of the same type from different locations (source vs. stored water, flies in kitchen vs. latrine, soil from courtyard vs. latrine) or different individuals (child vs. mother’s hands) are plotted separately. All estimates are adjusted for potential confounders.



**Figure 2** Forest plots of intervention effects on the prevalence of any virus, any bacteria, any protozoa and any STH in different types of environmental samples. Pooled estimates are presented when there are four or more study-specific estimates for a specific sample type and target combination and are denoted with diamond-shaped points. Grey crossed points denote data that were too sparse to estimate a prevalence ratio (i.e., <10 positive or negative observations). Samples of the same type from different locations (source vs. stored water, flies in kitchen vs. latrine, soil from courtyard vs. latrine) or different individuals (child vs. mother’s hands) are plotted separately. All estimates are adjusted for potential confounders.



**Figure 3.** Forest plots of intervention effects on the prevalence of any general, human and animal MST markers in different types of environmental samples. Pooled estimates are presented when there are four or more study-specific estimates for a specific sample type and target combination and are denoted with diamond-shaped points. Grey crossed points denote data that were too sparse to estimate a prevalence ratio (i.e., <10 positive or negative observations). Samples of the same type from different locations (source vs. stored water, flies in kitchen vs. latrine, soil from courtyard vs. latrine) or different individuals (child vs. mother’s hands) are plotted separately. All estimates are adjusted for potential confounders.

## Tables

### Table 1. Characteristics of included publications. qPCR indicates either qPCR or RT-qPCR

| **Parent study** | **Study design** | **Intervention** | **Time between intervention and environmental sampling** | **Location** | **Author/ year** | **Sample types** | **Targets** | **Analytic method** | **Number of samples** |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| WASH Benefits Bangladesh | Cluster-randomized trial | Latrine upgrades, child potties, scoops for feces disposal | 4 months | Rural Bangladesh | Boehm et al. 2016 | Stored drinking water, child hands, soil | Rotavirus, General, human, avian and ruminant fecal markers | qPCR | 1,497 |
| - | - | - | 16-35 months | - | Fuhrmeister et al. 2020 | Stored drinking water, child and mother hands, soil | Pathogenic E. coli, norovirus, Giardia | qPCR | 2,606 |
| - | - | - | ~2 years | - | Kwong et al. 2021 | Courtyard soil | Soil-transmitted helminths | Microscopy | 1,396 |
| WASH Benefits Kenya | Cluster-randomized trial | Latrine upgrades, child potties, scoops for feces disposal | ~2 years | Rural Kenya | Steinbaum et al. 2019 | Courtyard soil | Soil-transmitted helminths | Microscopy | 2,107 |
| MapSan | Controlled before-and-after study | Latrine upgrades | ~1 year | Urban Mozambique | Holcomb et al. 2020 | Source and stored water, household and latrine soil, food, flies caught in latrine and kitchen | Panel of 17 enteropathogens, human and avian fecal markers | qPCR | 1,081 |
| Gram Vikas | Matched cohort study | Latrine upgrades, piped water | ~6-10 years | Rural India | Reese et al. 2017 | Source and stored water | V. cholerae, Shigella | Slide agglutination serotyping | 3,452 |
| Total Sanitation Campaign | Cluster-randomized trial | Latrine upgrades | ~1 year | Rural India | Odagiri et al. 2016 | Source water | V. cholerae, rotavirus, adenovirus,general, human, and animal fecal markers | qPCR, microscopy | 60 |

### 

### Table 2. Mean (SD) abundances by study arm and adjusted abundance differences between intervention and control arms. Means are log10 transformed concentrations for MST markers, and are mean egg counts for soil transmitted helminths (*Ascaris* and *Trichuris*).

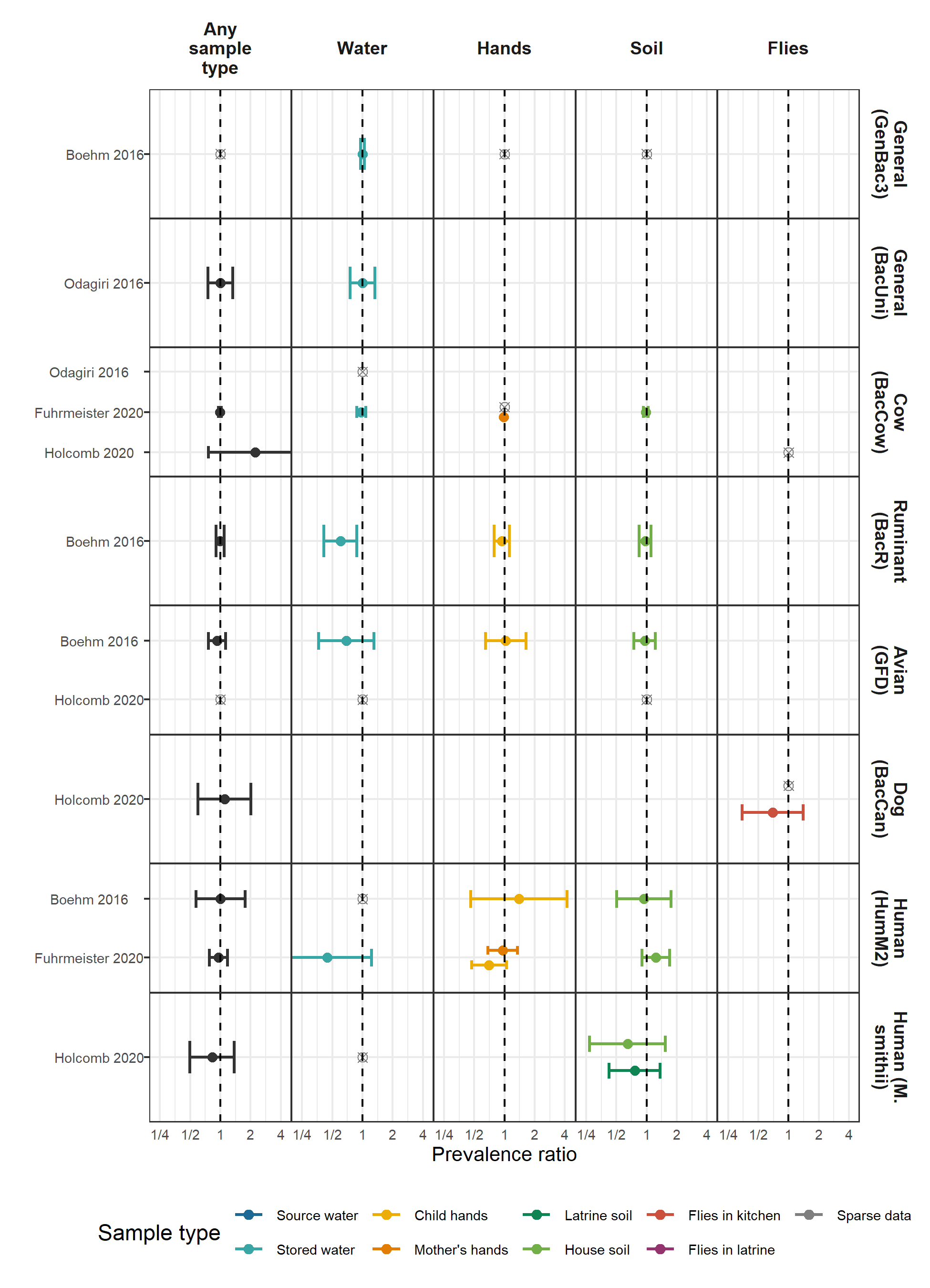
| **Study** | **Sample** | **Target** | **N** | **% in ROQ** | **Control mean (SD)** | **Intervention mean (SD)** | **Difference (95% CI)** | **P value** |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Boehm 2016 | Stored water | General (GenBac3) | 479 | 83.9 | 1.8 (1.1) | 1.7 (1) | -0.09 (-0.29 0.1) | 0.35 |
| - | Child hands | General (GenBac3) | 493 | 98.0 | 5.5 (0.9) | 5.5 (1.1) | 0.04 (-0.17 0.25) | 0.74 |
| - | House soil | General (GenBac3) | 483 | 100.0 | 6.6 (0.9) | 6.4 (0.9) | -0.2 (-0.37 -0.02) | 0.03 |
| Fuhrmeister 2020 | Child hands | Cow (BacCow) | 365 | 75.9 | 3.6 (1.4) | 3.4 (1.4) | -0.17 (-0.47 0.12) | 0.25 |
| - | Mother's hands | Cow (BacCow) | 725 | 66.5 | 3.3 (1.4) | 3 (1.5) | -0.28 (-0.49 -0.07) | 0.01 |
| Holcomb 2020 | Latrine soil | Human (M. smithii) | 113 | 51.3 | 6.7 (0.6) | 6.5 (0.5) | -0.13 (-0.34 0.07) | 0.19 |
| - | Flies in kitchen | Human (Bacteroides) | 113 | 77.0 | 3.8 (1.2) | 3.9 (0.9) | 0.23 (-0.16 0.62) | 0.26 |
| - | Flies in latrine | Human (Bacteroides) | 60 | 78.3 | 3.9 (1.3) | 4.6 (0.7) | 0.7 (0.11 1.28) | 0.02 |
| Steinbaum 2019 | House soil | Ascaris | 2,101 | 100.0 | 2.2 (18.8) | 1.4 (9.3) | 0.65 (0.33 1.28)a | 0.21 |
| - | - | Trichuris | 2,102 | 100.0 | 0.2 (1.8) | 0.2 (1) | 0.73 (0.36 1.48)a | 0.38 |
| Kwong 2021 | House soil | Ascaris | 1,423 | 100.0 | 2.3 (6.7) | 2.2 (6.9) | 0.96 (0.68 1.37)a | 0.84 |
| - | - | Trichuris | 1,423 | 100.0 | 1.6 (5) | 2 (5) | 1.21 (0.86 1.71)a | 0.27 |

ROQ: Range of quantification; SD: Standard deviation; CI: Confidence interval.

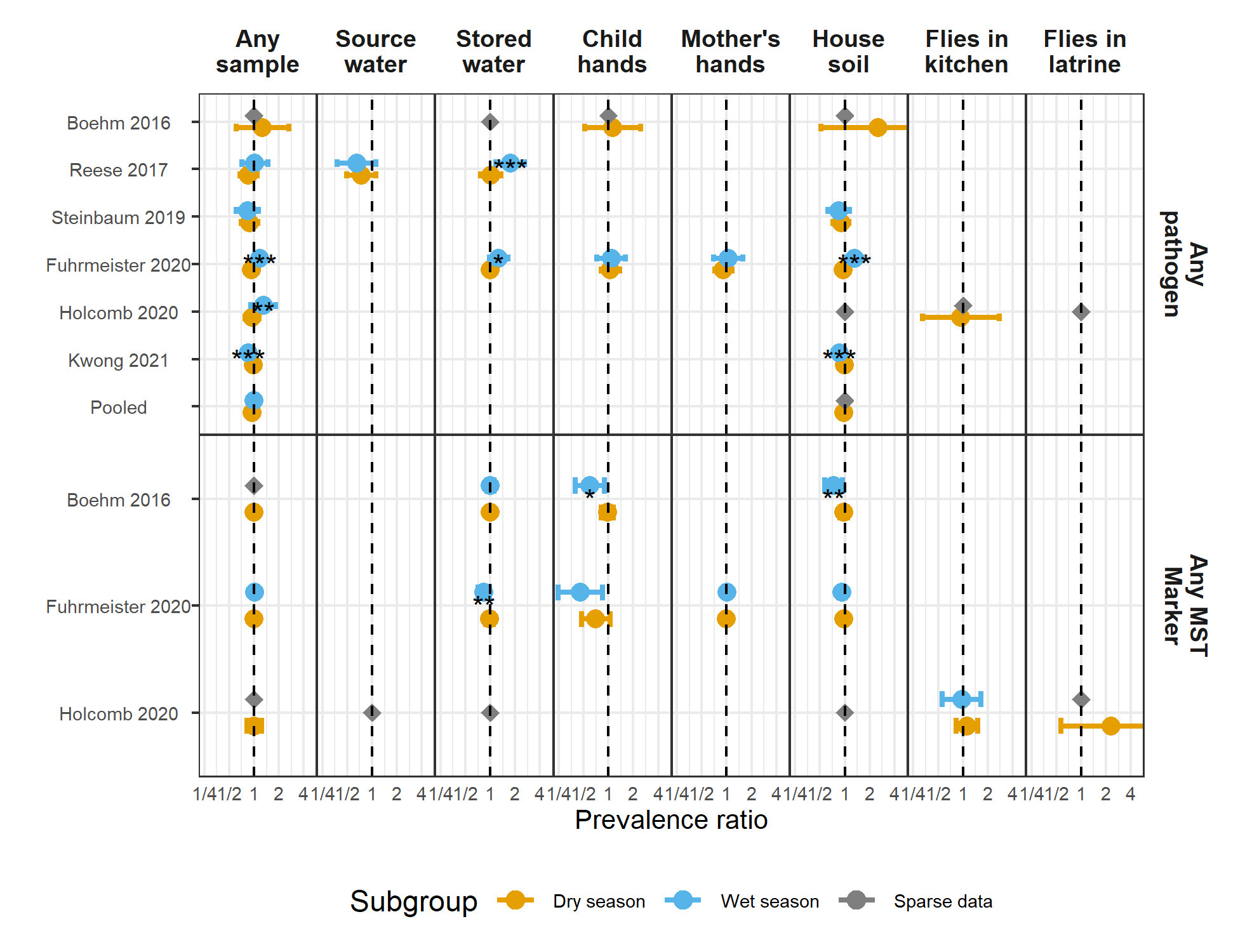
a Marks estimates from negative binomial models.

## Supplementary Figures

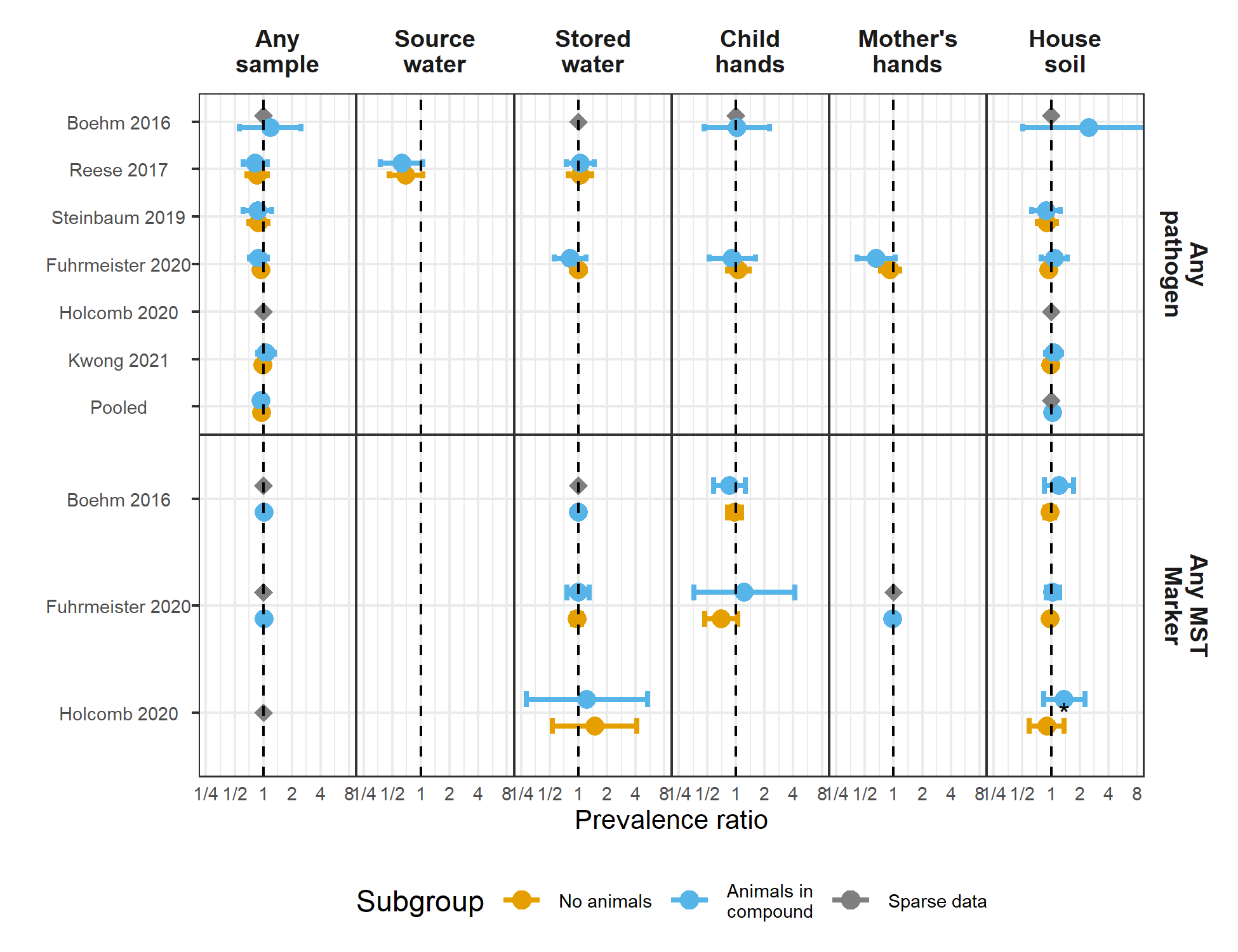
**Figure S1.** Forest plots of intervention effects on the prevalence of specific pathogens.



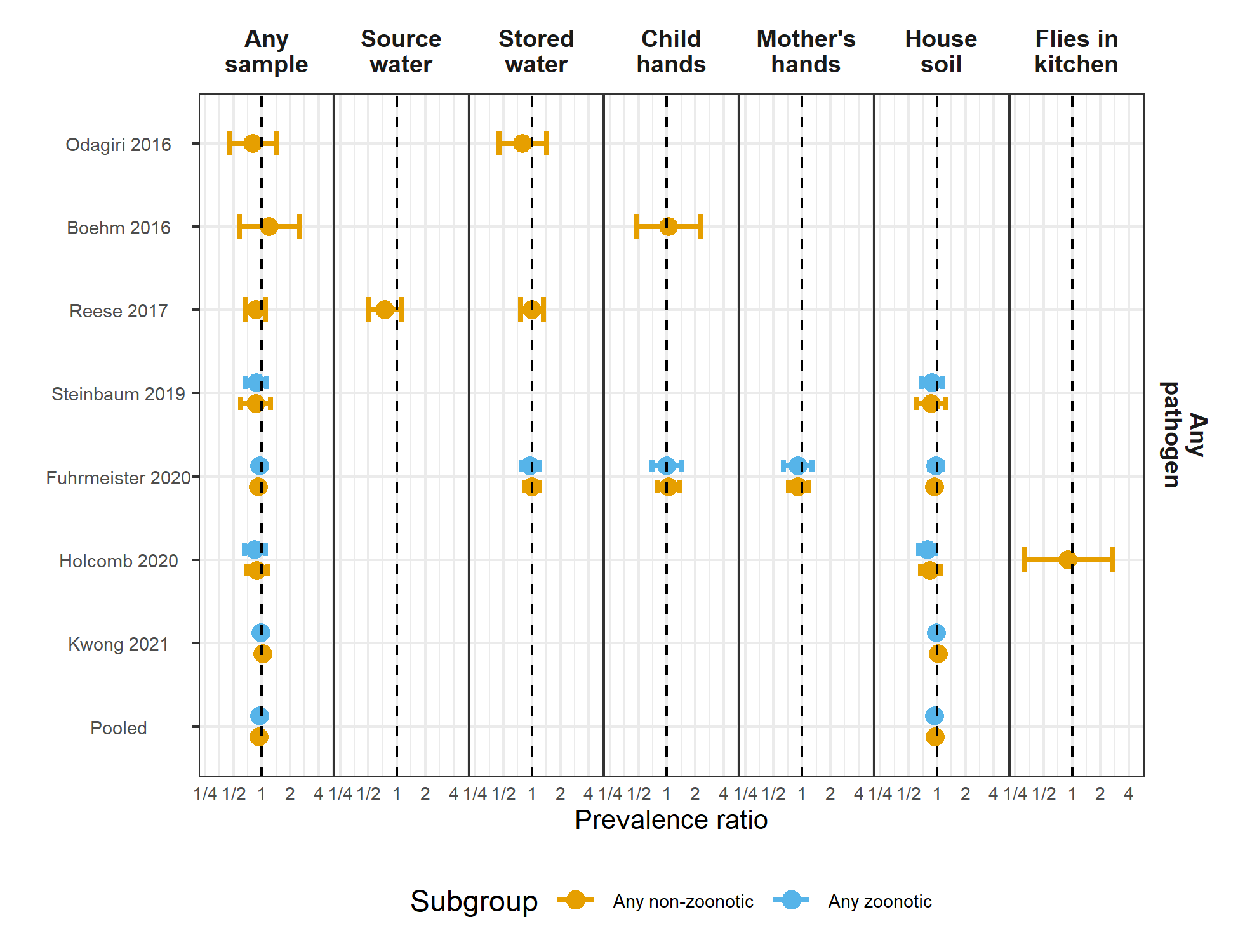
**Figure S2.** Forest plots of intervention effects on the prevalence of specific MST markers.



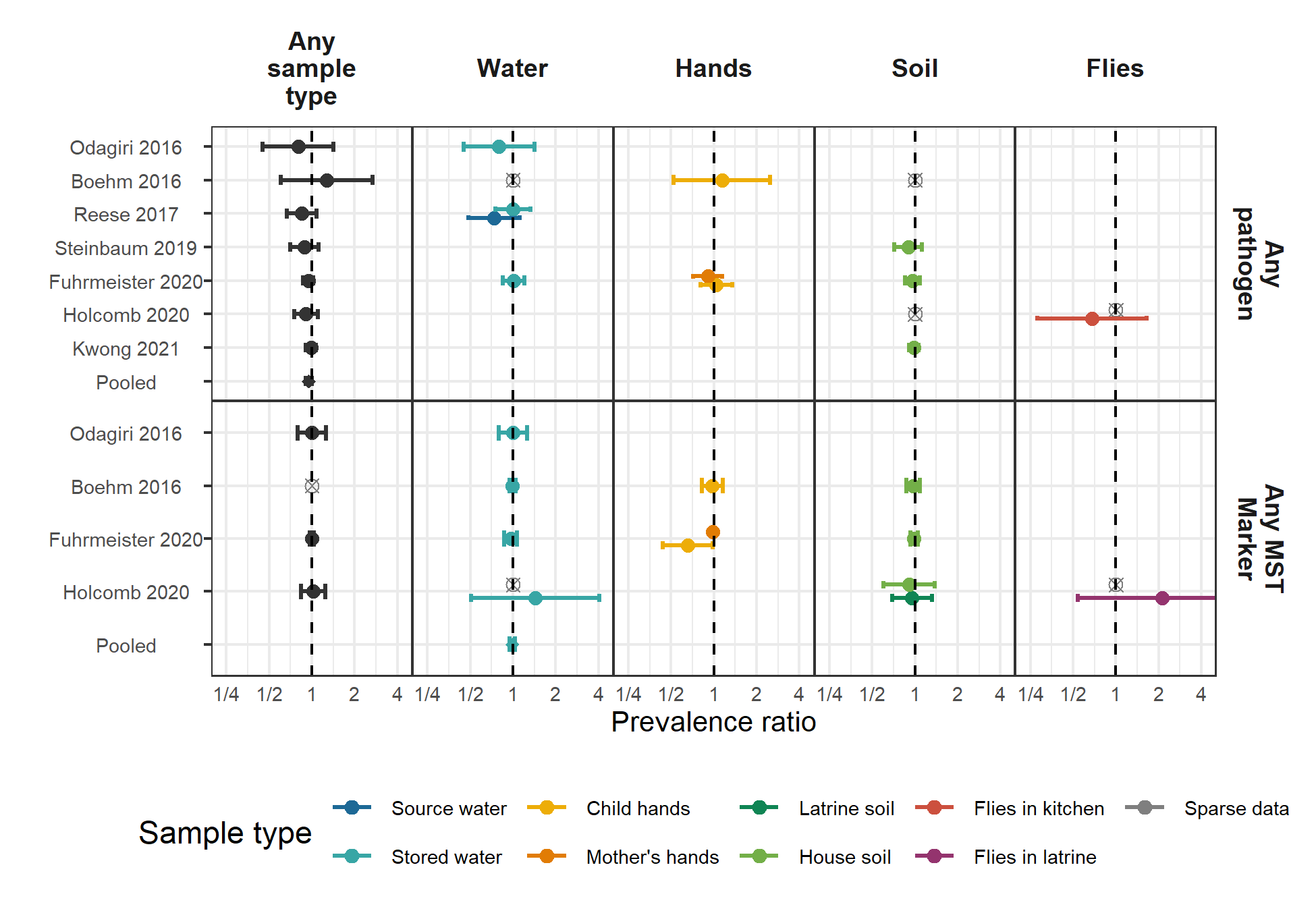
**Figure S3.** Forest plots of intervention effects on any enteropathogen, and any MST markers in different types of environmental samples, stratified by whether the sample was collected during the wet versus dry season (defined by the 6 months of highest average rainfall). Significant effect modification, as determined by the p-values on the regression model interaction term, is marked with asterisks (P < 0.05 = “\*”, P < 0.01 = “\*\*”, P < 0.001 = “\*\*\*”).



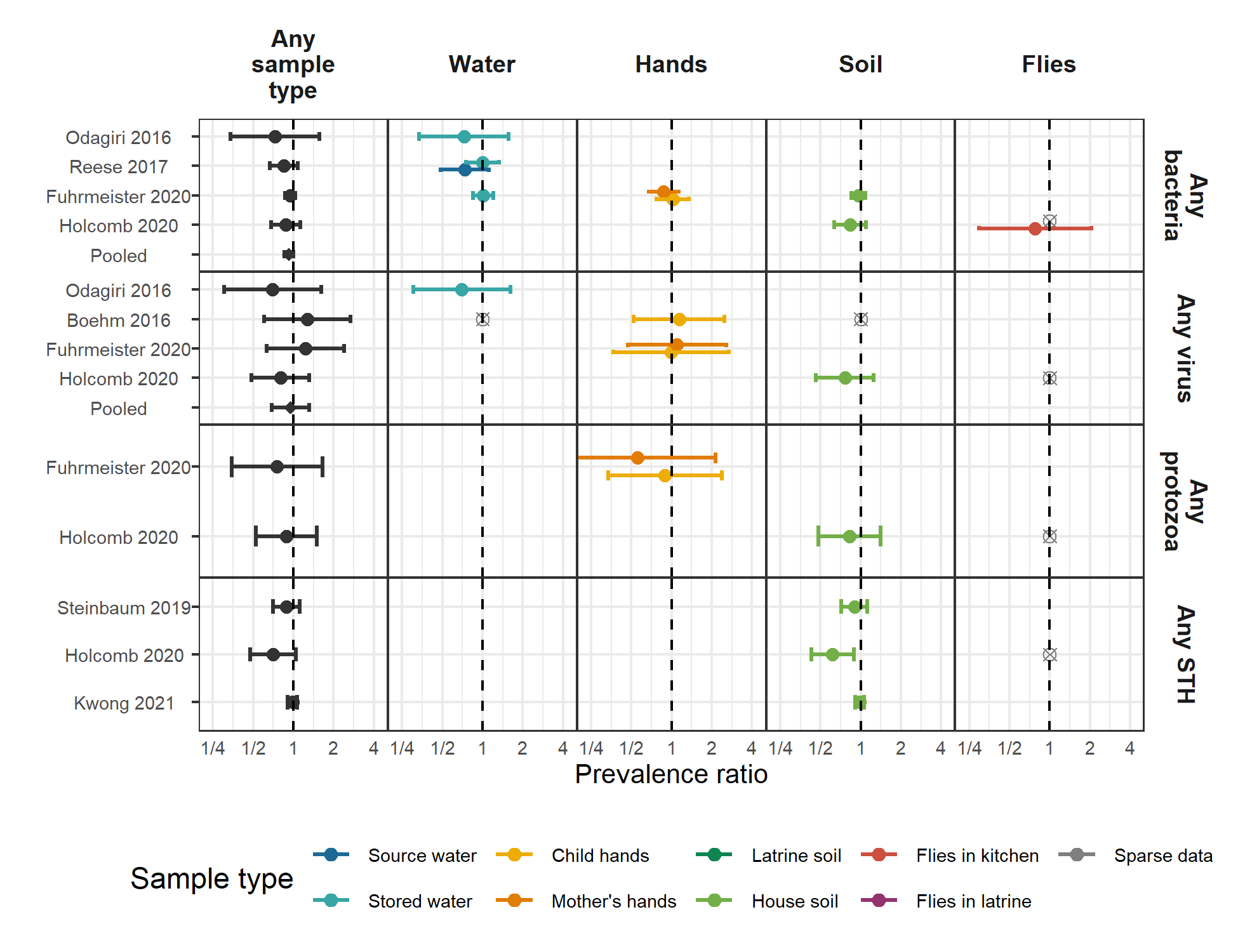
**Figure S4.** Forest plots of intervention effects on any enteropathogen, and any MST markers in different types of environmental samples, stratified by whether any animals were present in the compound. Significant effect modification, as determined by the p-values on the regression model interaction term, is marked with asterisks (P < 0.05 = “\*”, P < 0.01 = “\*\*”, P < 0.001 = “\*\*\*”).



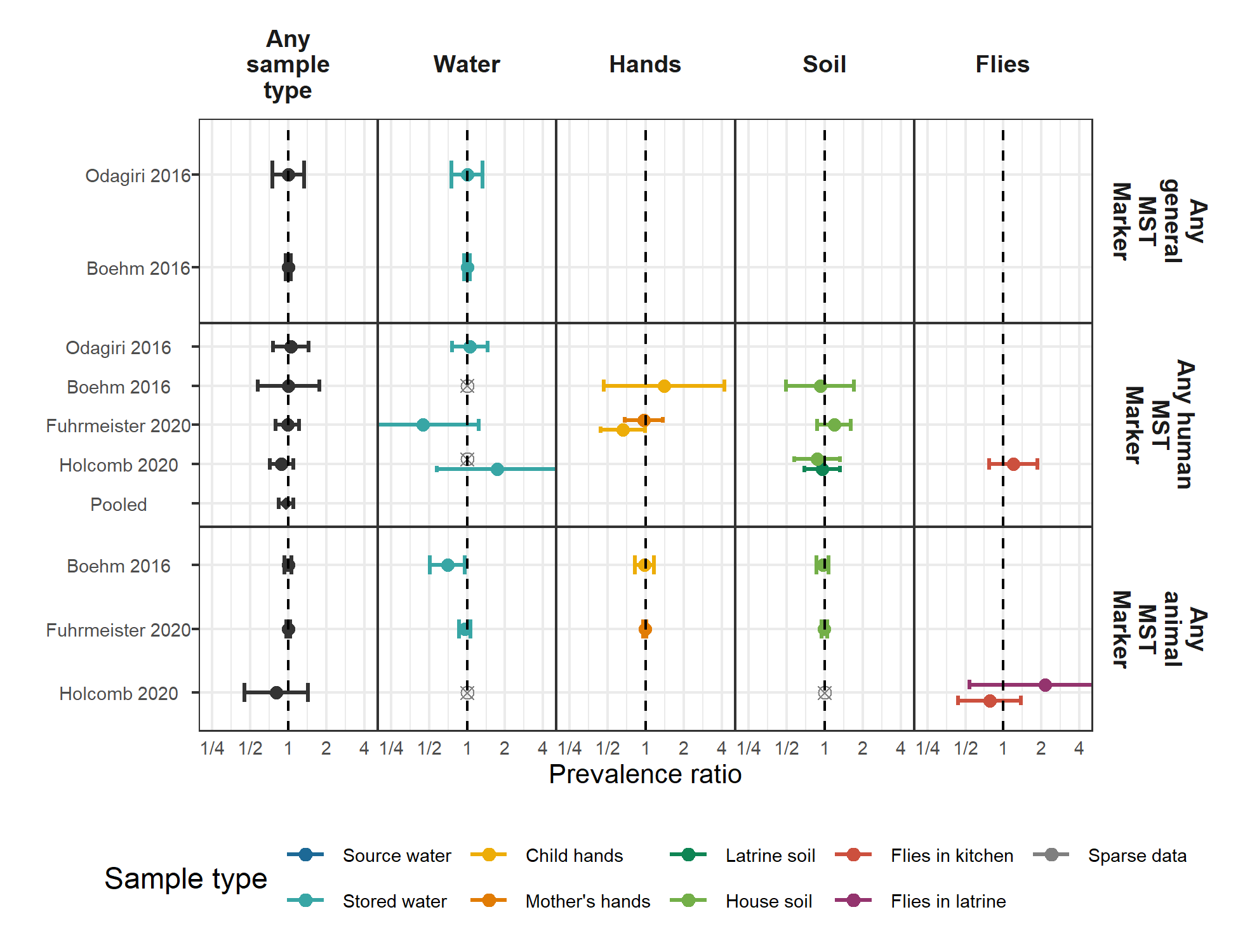
**Figure S5.** Forest plots of intervention effects on any enteropathogen in any type of environmental samples, stratified by whether the pathogen is zoonotically transmitted.



**Figure S6.** Forest plots of unadjusted intervention effects on the prevalence of any enteropathogen, and any MST markers in different types of environmental samples.



**Figure S7.** Forest plots of unadjusted intervention effects on the prevalence of any virus, any bacteria, any protozoa and any STH in different types of environmental samples.



**Figure S8.** Forest plots of unadjusted intervention effects on the prevalence of any general, human and animal MST markers in different types of environmental samples.

**Supplementary Tables**

**Table S1. Systematic review search terms**

Search terms were combined with “OR” within columns and with “AND” across columns.

| **Study design** | **WASH** | **Environmental markers** | **Child health** |
| --- | --- | --- | --- |
| matched, trial, RCT, experiment, intervention, randomized, randomised, quasi-randomized, quasi-randomised, quasi-experimental, pseudo-randomized, pseudo-randomised, non-randomized controlled trials | Water, Sanitation, Hygiene, Handwashing, WSH, Sanitation, Water Supply, Sanitary Drainage, Toilet Facilities, Drinking Water, Hand Hygiene, Water Purification, Waste Water, disinfection | molecular source tracking, microbial source tracking, microbial transmission, diarrheal pathogen, diarrheal pathogens, diarrhoeal pathogen, diarrhoeal pathogens, fecal-oral, faecal-oral, entericpathogen, entericpathogens, ruminant, avian, Feces, Faeces, Fecal, Faecal, Fecally, Faecally | Entericinfection, Soil-transmitted helminth, Protozoan, Seroconversion, Fecal microbiology, Faecal microbiology, Fecal biomarker, Faecal biomarker, Intestinal Diseases, Parasitic, Seroconversion, Enteritis, Helminthiasis, Helminthiases, Intestinal infection, Viral infection, Bacterial infection, Parasite infection, Parasitic infection, Helminth infection, Fecal sampling, Faecal sampling, Stool sampling, Stool collection, Diarrhea, Dysentery, Child growth faltering, Growth faltering, Child development, Length-for-age, Height-for-age, Weight-for-age, Head circumference, Waist circumference, Stunting, Stunted, Wasting, Wasted, Linear growth, Anthropometric measurement, Malnutrition, Undernourished, Undernutrition, Underweight, Growth Disorders, Childnutrition disorder, Wasting syndrome, Thinness, Growth velocity |

**Table S2. Pubmed search string**

[MH] are mesh headers and [TW] are text words.

|  |
| --- |
| ((matched [tw]) OR (trial [tw]) OR (RCT [tw]) OR (experiment [tw]) OR (intervention [tw]) OR (randomized [tw]) OR (randomised [tw]) OR (quasi-randomized [tw]) OR (quasi-randomised [tw]) OR (quasi-experimental [tw]) OR (pseudo-randomized [tw]) OR (pseudo-randomised [tw]) OR (“non-randomized controlled trials as topic” [mh])) AND ((Water [tw]) OR (Sanitation [tw]) OR (Hygiene [tw]) OR (Handwashing [tw]) OR (WSH [tw]) OR (“Sanitation” [mh]) OR (“Water Supply” [mh]) OR (“Drainage, Sanitary” [mh]) OR (Sanitary Drainage [tw]) OR (“Toilet Facilities” [mh]) OR (“Drinking Water” [mh]) OR (“Hand Hygiene” [mh]) OR (“Water Purification” [mh]) OR (“Waste Water” [mh]) OR (disinfect\* [tw])) AND ((molecular source tracking [tw]) OR (microbial source tracking [tw]) OR (microbial transmission [tw]) OR (diarrheal pathogen [tw]) OR (diarrheal pathogens [tw]) OR (diarrhoeal pathogen [tw]) OR (diarrhoeal pathogens [tw]) OR (fecal-oral [tw]) OR (faecal-oral [tw]) OR (enteric pathogen [tw]) OR (enteric pathogens [tw]) OR (ruminant\* [tw]) OR (avian\* [tw]) OR (“Feces” [mh]) OR (Feces [tw]) OR (Faeces [tw]) OR (Fecal [tw]) OR (Faecal [tw]) OR (Fecally [tw]) OR (Faecally [tw])) AND (((Enteric infection\* [tw]) OR (Soil-transmitted helminth\* [tw]) OR (Protozoan\* [tw]) OR (Seroconversion [tw]) OR (Fecal microbio\* [tw]) OR (Faecal microbio\* [tw]) OR (Fecal biomarker\* [tw]) OR (Faecal biomarker\* [tw]) OR (“Intestinal Diseases, Parasitic/epidemiology” [mh]) OR (“Seroconversion” [mh]) OR (Seroconversion [tw]) OR (“Enteritis/epidemiology” [mh]) OR (“Helminthiasis/complications” [mh]) OR (Helminthiasis [tw]) OR (Helminthiases)OR (“Helminthiasis/epidemiology” [mh]) OR (“Helminthiasis/prevention and control” [mh]) OR (Intestinal infection\* [tw]) OR (Viral infection\* [tw]) OR (Bacterial infection\* [tw]) OR (Parasite infection\* [tw]) OR (Parasitic infection\* [tw]) OR (Helminth infection\* [tw]) OR (Fecal sampling [tw]) OR (Faecal sampling [tw]) OR (Bacterial infection\* [tw]) OR (Parasite infection\* [tw]) OR (Parasitic infection\* [tw]) OR (Helminth infection\* [tw]) OR (Fecal sampling [tw]) OR (Faecal sampling [tw]) OR (Stool sampling [tw]) OR (Stool collection [tw])) OR ((Diarrh\* [tw]) OR (Dysentery [tw]) OR (“Diarrhea/epidemiology” [mh]) OR (“Diarrhea/etiology” [mh]) OR (“Diarrhea/prevention and control” [mh]) OR (“Diarrhea, Infantile” [mh]) OR (“Dysentery” [mh])) OR (Child growth faltering [tw]) OR (Growth faltering [tw])OR (Child development [tw]) OR (Length-for-age [tw]) OR (Height-for-age [tw]) OR (Weight-for-age [tw]) OR (Head circumference [tw]) OR (Waist circumference [tw]) OR (Stunt\* [tw]) OR (Wasting [tw]) OR (Wasted [tw]) OR (Linear growth [tw]) OR (Anthropometric measurement\* [tw]) OR (Maln\* [tw]) OR (Undernourish\* [tw]) OR (Undernutrition [tw]) OR (Underweight [tw]) OR (“Growth Disorders” [mh]) OR (Growth Disorders [tw]) OR (“Child nutrition disorders” [mh]) OR (Child nutrition disorder\* [tw]) OR (“Malnutrition” [mh]) OR (“Wasting Syndrome” [mh]) OR (Wasting syndrome [tw]) OR (“Thinness” [mh]) OR (Thinness [tw]) OR (Growth velocity [tw])) |

**Table S3. PRISMA Checklist**

(See separate attachment)

**Table S4. Prevalence of pathogens by sample type tested in each study**

| **Study** | **Sample** | **Target** | **Percent positive (n/N)** | **PR (95% CI)** |
| --- | --- | --- | --- | --- |
| Odagiri 2016 | Stored water | V. cholerae | 31.7% (19/60) | 0.73 (0.34, 1.57) |
| - | - | Adenovirus | 8.3% (5/60) | - |
| - | - | Rotavirus | 23.3% (14/60) | 0.75 (0.29, 1.93) |
| Boehm 2016 | Stored water | Rotavirus | 0.6% (3/493) | - |
| - | Child hands | Rotavirus | 6.1% (30/493) | 1.05 (0.48, 2.27) |
| - | House soil | Rotavirus | 1.4% (7/496) | - |
| Reese 2017 | Source water | Shigella | 10.7% (161/1499) | 0.73 (0.46, 1.15) |
| - | - | V. cholerae | 13% (36/276) | 0.93 (0.46, 1.85) |
| - | Stored water | Shigella | 10.1% (190/1874) | 1.08 (0.77, 1.51) |
| - | - | V. cholerae | 23.7% (100/422) | 1.03 (0.66, 1.6) |
| Steinbaum 2019 | House soil | Ascaris | 13% (273/2107) | 0.89 (0.69, 1.15) |
| - | - | Trichuris | 6.9% (146/2107) | 0.85 (0.6, 1.22) |
| Fuhrmeister 2020 | Stored water | Pathogenic E. coli | 38.6% (286/741) | 1 (0.84, 1.19) |
| - | Child hands | Pathogenic E. coli | 34% (127/373) | 1.02 (0.78, 1.35) |
| - | - | Giardia | 4.8% (15/311) | 0.89 (0.33, 2.38) |
| - | - | Norovirus | 4.2% (14/337) | 0.99 (0.37, 2.69) |
| - | Mother's hands | Pathogenic E. coli | 24% (177/737) | 0.85 (0.67, 1.09) |
| - | - | Giardia | 2.3% (14/602) | 0.56 (0.14, 2.13) |
| - | - | Norovirus | 3.1% (21/684) | 1.16 (0.48, 2.81) |
| - | House soil | Pathogenic E. coli | 61.3% (453/739) | 0.94 (0.83, 1.06) |
| Holcomb 2020 | House soil | C. difficile | 14.8% (13/88) | 0.9 (0.32, 2.48) |
| - | - | Campylobacter | 6.8% (6/88) | - |
| - | - | Pathogenic E. coli | 56.8% (50/88) | 0.72 (0.49, 1.07) |
| - | - | Salmonella | 6.8% (6/88) | - |
| - | - | Shigella | 21.6% (19/88) | 0.28 (0.1, 0.78) |
| - | - | V. cholerae | 0% (0/88) | - |
| - | - | Yersinia | 4.5% (4/88) | - |
| - | - | Ascaris | 60.2% (53/88) | 0.64 (0.43, 0.94) |
| - | - | Trichuris | 17% (15/88) | 0.92 (0.36, 2.33) |
| - | - | Cryptosporidium | 8% (7/88) | - |
| - | - | Entamoeba histolytica | 1.1% (1/88) | - |
| - | - | Giardia | 31.8% (28/88) | 0.6 (0.31, 1.15) |
| - | - | Adenovirus | 20.5% (18/88) | 0.21 (0.06, 0.68) |
| - | - | Astrovirus | 29.5% (26/88) | 1.04 (0.52, 2.07) |
| - | - | Norovirus | 2.3% (2/88) | - |
| - | - | Rotavirus | 4.5% (4/88) | - |
| - | - | Sapovirus | 0% (0/88) | - |
| - | Flies in kitchen | Campylobacter | 2.1% (1/48) | - |
| - | - | Pathogenic E. coli | 25% (12/48) | 0.78 (0.25, 2.47) |
| - | - | Shigella | 2.1% (1/48) | - |
| - | - | V. cholerae | 4.2% (2/48) | - |
| - | - | Ascaris | 0% (0/48) | - |
| - | - | Trichuris | 4.2% (2/48) | - |
| - | - | Giardia | 2.1% (1/48) | - |
| - | - | Adenovirus | 0% (0/48) | - |
| - | - | Astrovirus | 0% (0/48) | - |
| - | - | Norovirus | 0% (0/48) | - |
| - | - | Pan enterovirus | 0% (0/48) | - |
| - | - | Rotavirus | 0% (0/48) | - |
| - | - | Sapovirus | 0% (0/48) | - |
| - | Flies in latrine | Campylobacter | 0% (0/38) | - |
| - | - | Pathogenic E. coli | 36.8% (14/38) | - |
| - | - | Shigella | 2.6% (1/38) | - |
| - | - | V. cholerae | 0% (0/38) | - |
| - | - | Ascaris | 0% (0/38) | - |
| - | - | Trichuris | 2.6% (1/38) | - |
| - | - | Giardia | 7.9% (3/38) | - |
| - | - | Adenovirus | 10.5% (4/38) | - |
| - | - | Astrovirus | 0% (0/38) | - |
| - | - | Norovirus | 5.3% (2/38) | - |
| - | - | Pan enterovirus | 0% (0/38) | - |
| - | - | Rotavirus | 2.6% (1/38) | - |
| - | - | Sapovirus | 0% (0/38) | - |
| Kwong 2021 | House soil | Ascaris | 62.3% (886/1423) | 0.97 (0.87, 1.08) |
| - | - | Trichuris | 56.1% (798/1423) | 1.02 (0.91, 1.15) |

**Table S5. Prevalence of microbial source tracking markers by sample type tested in each study**

| **Study** | **Sample** | **Target** | **Percent positive (n/N)** | **PR (95% CI)** |
| --- | --- | --- | --- | --- |
| Odagiri 2016 | Stored water | Animal (BacCow) | 91.7% (55/60) | - |
| - | - | General (BacUni) | 76.7% (46/60) | 1 (0.75, 1.33) |
| - | - | Human (BacHum) | 71.7% (43/60) | - |
| Boehm 2016 | Stored water | Avian (GFD) | 9.3% (46/493) | 0.69 (0.37, 1.3) |
| - | - | Ruminant (BacR) | 21.9% (108/493) | 0.6 (0.41, 0.88) |
| - | - | General (GenBac3) | 93.5% (461/493) | 1 (0.96, 1.05) |
| - | - | Human (HumM2) | 0% (0/493) | - |
| - | Child hands | Avian (GFD) | 16.2% (80/493) | 1.03 (0.65, 1.64) |
| - | - | Ruminant (BacR) | 54.2% (267/493) | 0.94 (0.79, 1.12) |
| - | - | General (GenBac3) | 98.6% (486/493) | - |
| - | - | Human (HumM2) | 2.4% (12/493) | 1.39 (0.46, 4.2) |
| - | House soil | Avian (GFD) | 33.3% (165/496) | 0.96 (0.74, 1.23) |
| - | - | Ruminant (BacR) | 66.7% (331/496) | 0.97 (0.85, 1.11) |
| - | - | General (GenBac3) | 100% (496/496) | - |
| - | - | Human (HumM2) | 8.9% (44/496) | 0.94 (0.5, 1.75) |
| Fuhrmeister 2020 | Stored water | Animal (BacCow) | 68.5% (482/704) | 0.97 (0.88, 1.08) |
| - | - | Human (HumM2) | 2.6% (17/651) | 0.44 (0.16, 1.23) |
| - | - | Non-zoonotic E. coli | 19.7% (146/741) | 0.89 (0.66, 1.18) |
| - | - | Zoonotic E. coli | 27.9% (207/741) | 0.96 (0.77, 1.21) |
| - | Child hands | Animal (BacCow) | 97.5% (356/365) | - |
| - | - | Human (HumM2) | 21.9% (74/338) | 0.7 (0.47, 1.05) |
| - | - | Non-zoonotic E. coli | 18% (67/373) | 0.88 (0.57, 1.35) |
| - | - | Zoonotic E. coli | 24.9% (93/373) | 1.03 (0.71, 1.48) |
| - | Mother's hands | Animal (BacCow) | 96.7% (702/726) | 0.98 (0.96, 1.01) |
| - | - | Human (HumM2) | 18.1% (118/651) | 0.96 (0.68, 1.35) |
| - | - | Non-zoonotic E. coli | 12.2% (90/737) | 0.74 (0.51, 1.06) |
| - | - | Zoonotic E. coli | 15.7% (116/737) | 0.9 (0.63, 1.3) |
| - | House soil | Animal (BacCow) | 90.6% (572/631) | 0.99 (0.93, 1.04) |
| - | - | Human (HumM2) | 20.1% (127/631) | 1.24 (0.91, 1.7) |
| - | - | Non-zoonotic E. coli | 28.1% (208/739) | 0.8 (0.64, 0.98) |
| - | - | Zoonotic E. coli | 50.2% (371/739) | 0.97 (0.83, 1.13) |
| Holcomb 2020 | Source water | Avian (GFD) | 0% (0/41) | - |
| - | - | Human (HF183) | 2.4% (1/41) | - |
| - | - | Human (M. smithii) | 0% (0/41) | - |
| - | Stored water | Avian (GFD) | 1.1% (1/94) | - |
| - | - | Human (HF183) | 14.9% (14/94) | - |
| - | - | Human (M. smithii) | 0% (0/94) | - |
| - | Latrine soil | Avian (GFD) | 3.3% (2/60) | - |
| - | - | Human (HF183) | 50% (30/60) | - |
| - | - | Human (M. smithii) | 45% (27/60) | 0.76 (0.42, 1.37) |
| - | House soil | Avian (GFD) | 3.6% (3/83) | - |
| - | - | Human (HF183) | 42.2% (35/83) | - |
| - | - | Human (M. smithii) | 24.1% (20/83) | 0.65 (0.27, 1.54) |
| - | - | Non-zoonotic E. coli | 54.5% (48/88) | 0.75 (0.5, 1.11) |
| - | - | Zoonotic E. coli | 18.2% (16/88) | 0.35 (0.12, 1.01) |
| - | Flies in kitchen | Animal (BacCow) | 14.6% (7/48) | - |
| - | - | Dog (BacCan) | 35.4% (17/48) | 0.69 (0.34, 1.39) |
| - | - | Human (BacHum) | 68.8% (33/48) | - |
| - | Flies in latrine | Animal (BacCow) | 10.5% (4/38) | - |
| - | - | Dog (BacCan) | 23.7% (9/38) | - |
| - | - | Human (BacHum) | 76.3% (29/38) | - |

**Table S6.**

Unadjusted and adjusted results by study, sample type, and aggregated variables for pathogen targets (any pathogen, any bacteria, any viruses, any protozoa, any STH).

| **Study** | **Target** | **Sample** | **Positive, Intervention** | **Negative, Intervention** | **Positive, Control** | **Negative, Control** | **Total observations** | **Unadjusted Prevalence Ratio** | **Unadjusted p-value** | **Adjusted Prevalence Ratio** | **Adjusted p-value** |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Holcomb 2020 | Any pathogen | Any sample | 44 | 19 | 63 | 19 | 145 | PR=0.91 (95% CI: 0.75, 1.1) | 0.33 | PR=0.91 (95% CI: 0.75, 1.1) | 0.33 |
| Fuhrmeister 2020 | Any pathogen | Any sample | 236 | 59 | 261 | 39 | 595 | PR=0.94 (95% CI: 0.87, 1.02) | 0.17 | PR=0.92 (95% CI: 0.86, 0.98) | 0.01 |
| Steinbaum 2019 | Any pathogen | Any sample | 205 | 968 | 169 | 700 | 2,042 | PR=0.88 (95% CI: 0.7, 1.11) | 0.29 | PR=0.89 (95% CI: 0.71, 1.11) | 0.31 |
| Reese 2017 | Any pathogen | Any sample | 185 | 792 | 238 | 825 | 2,040 | PR=0.85 (95% CI: 0.66, 1.08) | 0.18 | PR=0.86 (95% CI: 0.68, 1.09) | 0.21 |
| Boehm 2016 | Any pathogen | Any sample | 18 | 225 | 15 | 226 | 484 | PR=1.27 (95% CI: 0.6, 2.68) | 0.53 | PR=1.21 (95% CI: 0.58, 2.51) | 0.62 |
| Odagiri 2016 | Any pathogen | Any sample | 12 | 18 | 15 | 15 | 60 | PR=0.8 (95% CI: 0.45, 1.42) | 0.45 |  |  |
| Reese 2017 | Any pathogen | Source water | 68 | 588 | 122 | 747 | 1,525 | PR=0.74 (95% CI: 0.49, 1.12) | 0.15 | PR=0.74 (95% CI: 0.5, 1.12) | 0.16 |
| Fuhrmeister 2020 | Any pathogen | Stored water | 138 | 218 | 148 | 237 | 741 | PR=1.01 (95% CI: 0.85, 1.2) | 0.93 | PR=1 (95% CI: 0.84, 1.19) | 1 |
| Reese 2017 | Any pathogen | Stored water | 134 | 786 | 147 | 860 | 1,927 | PR=1 (95% CI: 0.75, 1.32) | 0.99 | PR=1.01 (95% CI: 0.77, 1.34) | 0.94 |
| Boehm 2016 | Any pathogen | Stored water | 2 | 243 | 1 | 245 | 491 | Not estimated |  | Not estimated |  |
| Odagiri 2016 | Any pathogen | Stored water | 12 | 18 | 15 | 15 | 60 | PR=0.8 (95% CI: 0.45, 1.42) | 0.45 |  |  |
| Fuhrmeister 2020 | Any pathogen | Child hands | 75 | 113 | 72 | 116 | 376 | PR=1.04 (95% CI: 0.8, 1.35) | 0.76 | PR=1.05 (95% CI: 0.81, 1.37) | 0.69 |
| Boehm 2016 | Any pathogen | Child hands | 15 | 227 | 14 | 224 | 480 | PR=1.14 (95% CI: 0.52, 2.48) | 0.75 | PR=1.05 (95% CI: 0.48, 2.27) | 0.91 |
| Fuhrmeister 2020 | Any pathogen | Mother's hands | 96 | 266 | 110 | 267 | 739 | PR=0.91 (95% CI: 0.72, 1.15) | 0.43 | PR=0.92 (95% CI: 0.72, 1.16) | 0.47 |
| Kwong 2021 | Any pathogen | House soil | 363 | 125 | 687 | 221 | 1,396 | PR=0.98 (95% CI: 0.91, 1.06) | 0.67 | PR=0.98 (95% CI: 0.91, 1.06) | 0.68 |
| Holcomb 2020 | Any pathogen | House soil | 37 | 6 | 43 | 2 | 88 | Not estimated |  | Not estimated |  |
| Fuhrmeister 2020 | Any pathogen | House soil | 209 | 141 | 224 | 136 | 710 | PR=0.96 (95% CI: 0.86, 1.08) | 0.53 | PR=0.94 (95% CI: 0.83, 1.06) | 0.32 |
| Steinbaum 2019 | Any pathogen | House soil | 208 | 988 | 169 | 718 | 2,083 | PR=0.9 (95% CI: 0.72, 1.13) | 0.35 | PR=0.9 (95% CI: 0.72, 1.13) | 0.37 |
| Boehm 2016 | Any pathogen | House soil | 5 | 242 | 2 | 247 | 496 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any pathogen | Flies in latrine | 1 | 3 | 17 | 17 | 38 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any pathogen | Flies in kitchen | 7 | 20 | 8 | 13 | 48 | PR=0.68 (95% CI: 0.28, 1.65) | 0.4 | PR=0.68 (95% CI: 0.28, 1.65) | 0.4 |
| Holcomb 2020 | Any bacteria | Any sample | 35 | 28 | 52 | 30 | 145 | PR=0.88 (95% CI: 0.68, 1.13) | 0.31 | PR=0.88 (95% CI: 0.68, 1.13) | 0.31 |
| Fuhrmeister 2020 | Any bacteria | Any sample | 233 | 62 | 257 | 43 | 595 | PR=0.94 (95% CI: 0.86, 1.03) | 0.18 | PR=0.92 (95% CI: 0.86, 0.99) | 0.03 |
| Reese 2017 | Any bacteria | Any sample | 185 | 792 | 238 | 825 | 2,040 | PR=0.85 (95% CI: 0.66, 1.08) | 0.18 | PR=0.86 (95% CI: 0.68, 1.09) | 0.21 |
| Odagiri 2016 | Any bacteria | Any sample | 8 | 22 | 11 | 19 | 60 | PR=0.73 (95% CI: 0.34, 1.57) | 0.42 |  |  |
| Reese 2017 | Any bacteria | Source water | 68 | 588 | 122 | 747 | 1,525 | PR=0.74 (95% CI: 0.49, 1.12) | 0.15 | PR=0.74 (95% CI: 0.5, 1.12) | 0.16 |
| Fuhrmeister 2020 | Any bacteria | Stored water | 138 | 218 | 148 | 237 | 741 | PR=1.01 (95% CI: 0.85, 1.2) | 0.93 | PR=1 (95% CI: 0.84, 1.19) | 1 |
| Reese 2017 | Any bacteria | Stored water | 134 | 786 | 147 | 860 | 1,927 | PR=1 (95% CI: 0.75, 1.32) | 0.99 | PR=1.01 (95% CI: 0.77, 1.34) | 0.94 |
| Odagiri 2016 | Any bacteria | Stored water | 8 | 22 | 11 | 19 | 60 | PR=0.73 (95% CI: 0.34, 1.57) | 0.42 |  |  |
| Fuhrmeister 2020 | Any bacteria | Child hands | 64 | 122 | 63 | 124 | 373 | PR=1.02 (95% CI: 0.78, 1.35) | 0.88 | PR=1.02 (95% CI: 0.78, 1.35) | 0.88 |
| Fuhrmeister 2020 | Any bacteria | Mother's hands | 81 | 281 | 96 | 279 | 737 | PR=0.87 (95% CI: 0.68, 1.13) | 0.3 | PR=0.85 (95% CI: 0.67, 1.09) | 0.2 |
| Holcomb 2020 | Any bacteria | House soil | 28 | 15 | 35 | 10 | 88 | PR=0.84 (95% CI: 0.64, 1.1) | 0.2 | PR=0.8 (95% CI: 0.61, 1.03) | 0.09 |
| Fuhrmeister 2020 | Any bacteria | House soil | 209 | 141 | 224 | 136 | 710 | PR=0.96 (95% CI: 0.86, 1.08) | 0.53 | PR=0.94 (95% CI: 0.83, 1.06) | 0.32 |
| Holcomb 2020 | Any bacteria | Flies in latrine | 1 | 3 | 14 | 20 | 38 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any bacteria | Flies in kitchen | 7 | 20 | 7 | 14 | 48 | PR=0.78 (95% CI: 0.29, 2.06) | 0.61 | PR=0.78 (95% CI: 0.29, 2.06) | 0.61 |
| Holcomb 2020 | Any virus | Any sample | 16 | 47 | 26 | 56 | 145 | PR=0.8 (95% CI: 0.49, 1.32) | 0.38 | PR=0.73 (95% CI: 0.45, 1.18) | 0.2 |
| Fuhrmeister 2020 | Any virus | Any sample | 16 | 223 | 13 | 223 | 475 | PR=1.23 (95% CI: 0.63, 2.4) | 0.54 | PR=1.15 (95% CI: 0.56, 2.35) | 0.7 |
| Boehm 2016 | Any virus | Any sample | 18 | 225 | 15 | 226 | 484 | PR=1.27 (95% CI: 0.6, 2.68) | 0.53 | PR=1.21 (95% CI: 0.58, 2.51) | 0.62 |
| Odagiri 2016 | Any virus | Any sample | 7 | 23 | 10 | 20 | 60 | PR=0.7 (95% CI: 0.3, 1.62) | 0.4 |  |  |
| Boehm 2016 | Any virus | Stored water | 2 | 243 | 1 | 245 | 491 | Not estimated |  | Not estimated |  |
| Odagiri 2016 | Any virus | Stored water | 7 | 23 | 10 | 20 | 60 | PR=0.7 (95% CI: 0.3, 1.62) | 0.4 |  |  |
| Fuhrmeister 2020 | Any virus | Child hands | 7 | 162 | 7 | 161 | 337 | PR=0.99 (95% CI: 0.37, 2.69) | 0.99 | PR=0.99 (95% CI: 0.37, 2.69) | 0.99 |
| Boehm 2016 | Any virus | Child hands | 15 | 227 | 14 | 224 | 480 | PR=1.14 (95% CI: 0.52, 2.48) | 0.75 | PR=1.05 (95% CI: 0.48, 2.27) | 0.91 |
| Fuhrmeister 2020 | Any virus | Mother's hands | 11 | 320 | 9 | 318 | 658 | PR=1.1 (95% CI: 0.47, 2.57) | 0.83 | PR=1.16 (95% CI: 0.48, 2.81) | 0.75 |
| Holcomb 2020 | Any virus | House soil | 16 | 27 | 22 | 23 | 88 | PR=0.76 (95% CI: 0.46, 1.25) | 0.28 | PR=0.73 (95% CI: 0.45, 1.17) | 0.19 |
| Boehm 2016 | Any virus | House soil | 5 | 242 | 2 | 247 | 496 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any virus | Flies in latrine | 0 | 4 | 5 | 29 | 38 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any virus | Flies in kitchen | 0 | 27 | 0 | 21 | 48 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any protozoa | Any sample | 15 | 48 | 22 | 60 | 145 | PR=0.89 (95% CI: 0.53, 1.5) | 0.66 | PR=0.86 (95% CI: 0.51, 1.45) | 0.58 |
| Fuhrmeister 2020 | Any protozoa | Any sample | 11 | 222 | 15 | 215 | 463 | PR=0.75 (95% CI: 0.35, 1.65) | 0.48 | PR=0.74 (95% CI: 0.34, 1.59) | 0.44 |
| Fuhrmeister 2020 | Any protozoa | Child hands | 7 | 147 | 8 | 149 | 311 | PR=0.89 (95% CI: 0.33, 2.38) | 0.82 | PR=0.89 (95% CI: 0.33, 2.38) | 0.82 |
| Fuhrmeister 2020 | Any protozoa | Mother's hands | 5 | 296 | 9 | 292 | 602 | PR=0.56 (95% CI: 0.14, 2.13) | 0.39 | PR=0.56 (95% CI: 0.14, 2.13) | 0.39 |
| Holcomb 2020 | Any protozoa | House soil | 15 | 28 | 19 | 26 | 88 | PR=0.83 (95% CI: 0.48, 1.42) | 0.49 | PR=0.83 (95% CI: 0.48, 1.42) | 0.49 |
| Holcomb 2020 | Any protozoa | Flies in latrine | 0 | 4 | 3 | 31 | 38 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any protozoa | Flies in kitchen | 0 | 27 | 1 | 20 | 48 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any STH | Any sample | 20 | 43 | 37 | 45 | 145 | PR=0.7 (95% CI: 0.48, 1.04) | 0.08 | PR=0.64 (95% CI: 0.45, 0.93) | 0.02 |
| Steinbaum 2019 | Any STH | Any sample | 205 | 968 | 169 | 700 | 2,042 | PR=0.88 (95% CI: 0.7, 1.11) | 0.29 | PR=0.89 (95% CI: 0.71, 1.11) | 0.31 |
| Kwong 2021 | Any STH | House soil | 363 | 125 | 687 | 221 | 1,396 | PR=0.98 (95% CI: 0.91, 1.06) | 0.67 | PR=0.98 (95% CI: 0.91, 1.06) | 0.68 |
| Holcomb 2020 | Any STH | House soil | 20 | 23 | 34 | 11 | 88 | PR=0.62 (95% CI: 0.43, 0.89) | 0.01 | PR=0.6 (95% CI: 0.42, 0.85) | 0 |
| Steinbaum 2019 | Any STH | House soil | 208 | 988 | 169 | 718 | 2,083 | PR=0.9 (95% CI: 0.72, 1.13) | 0.35 | PR=0.9 (95% CI: 0.72, 1.13) | 0.37 |
| Holcomb 2020 | Any STH | Flies in latrine | 0 | 4 | 1 | 33 | 38 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any STH | Flies in kitchen | 0 | 27 | 2 | 19 | 48 | Not estimated |  | Not estimated |  |

**Table S7.**

Unadjusted and adjusted results by study, sample type, and aggregated variables for MST targets (any MST, any general MST, any human MST, any animal MST).

| **Study** | **Target** | **Sample** | **Positive, Intervention** | **Negative, Intervention** | **Positive, Control** | **Negative, Control** | **Total observations** | **Unadjusted Prevalence Ratio** | **Unadjusted p-value** | **Adjusted Prevalence Ratio** | **Adjusted p-value** |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Holcomb 2020 | Any MST Marker | Any sample | 34 | 7 | 39 | 9 | 89 | PR=1.02 (95% CI: 0.84, 1.24) | 0.84 | PR=1.02 (95% CI: 0.84, 1.24) | 0.84 |
| Fuhrmeister 2020 | Any MST Marker | Any sample | 282 | 13 | 290 | 10 | 595 | PR=1 (95% CI: 0.97, 1.03) | 0.97 | PR=0.99 (95% CI: 0.96, 1.02) | 0.49 |
| Boehm 2016 | Any MST Marker | Any sample | 246 | 2 | 244 | 5 | 497 | Not estimated |  | Not estimated |  |
| Odagiri 2016 | Any MST Marker | Any sample | 25 | 5 | 25 | 5 | 60 | PR=1 (95% CI: 0.79, 1.26) | 1 |  |  |
| Holcomb 2020 | Any MST Marker | Source water | 1 | 21 | 0 | 19 | 41 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any MST Marker | Stored water | 9 | 39 | 6 | 40 | 94 | PR=1.44 (95% CI: 0.51, 4.08) | 0.5 | PR=1.44 (95% CI: 0.51, 4.08) | 0.5 |
| Fuhrmeister 2020 | Any MST Marker | Stored water | 230 | 119 | 256 | 119 | 724 | PR=0.97 (95% CI: 0.87, 1.07) | 0.52 | PR=0.98 (95% CI: 0.88, 1.08) | 0.65 |
| Boehm 2016 | Any MST Marker | Stored water | 228 | 16 | 230 | 15 | 489 | PR=1 (95% CI: 0.95, 1.04) | 0.85 | PR=1 (95% CI: 0.95, 1.04) | 0.91 |
| Odagiri 2016 | Any MST Marker | Stored water | 25 | 5 | 25 | 5 | 60 | PR=1 (95% CI: 0.79, 1.26) | 1 |  |  |
| Fuhrmeister 2020 | Any MST Marker | Child hands | 30 | 142 | 44 | 122 | 338 | PR=0.66 (95% CI: 0.44, 0.99) | 0.04 | PR=0.7 (95% CI: 0.47, 1.04) | 0.08 |
| Boehm 2016 | Any MST Marker | Child hands | 141 | 101 | 143 | 95 | 480 | PR=0.98 (95% CI: 0.82, 1.16) | 0.78 | PR=0.97 (95% CI: 0.82, 1.15) | 0.71 |
| Fuhrmeister 2020 | Any MST Marker | Mother's hands | 346 | 14 | 359 | 9 | 728 | PR=0.99 (95% CI: 0.96, 1.01) | 0.26 | PR=0.99 (95% CI: 0.96, 1.01) | 0.29 |
| Holcomb 2020 | Any MST Marker | House soil | 21 | 18 | 26 | 18 | 83 | PR=0.91 (95% CI: 0.6, 1.38) | 0.66 | PR=0.88 (95% CI: 0.58, 1.35) | 0.57 |
| Fuhrmeister 2020 | Any MST Marker | House soil | 274 | 38 | 284 | 33 | 629 | PR=0.99 (95% CI: 0.93, 1.05) | 0.7 | PR=0.98 (95% CI: 0.92, 1.04) | 0.49 |
| Boehm 2016 | Any MST Marker | House soil | 180 | 67 | 187 | 62 | 496 | PR=0.97 (95% CI: 0.87, 1.08) | 0.59 | PR=0.97 (95% CI: 0.87, 1.08) | 0.58 |
| Holcomb 2020 | Any MST Marker | Latrine soil | 21 | 9 | 22 | 8 | 60 | PR=0.95 (95% CI: 0.69, 1.32) | 0.78 | PR=0.95 (95% CI: 0.69, 1.32) | 0.78 |
| Holcomb 2020 | Any MST Marker | Flies in latrine | 2 | 2 | 8 | 26 | 38 | PR=2.12 (95% CI: 0.54, 8.4) | 0.28 | PR=2.12 (95% CI: 0.54, 8.4) | 0.28 |
| Holcomb 2020 | Any MST Marker | Flies in kitchen | 23 | 4 | 16 | 5 | 48 | Not estimated |  | Not estimated |  |
| Boehm 2016 | Any general MST Marker | Any sample | 230 | 16 | 229 | 16 | 491 | PR=1 (95% CI: 0.95, 1.05) | 0.99 | PR=1 (95% CI: 0.96, 1.05) | 0.92 |
| Odagiri 2016 | Any general MST Marker | Any sample | 23 | 7 | 23 | 7 | 60 | PR=1 (95% CI: 0.75, 1.33) | 1 |  |  |
| Boehm 2016 | Any general MST Marker | Stored water | 230 | 16 | 229 | 16 | 491 | PR=1 (95% CI: 0.95, 1.05) | 0.99 | PR=1 (95% CI: 0.96, 1.05) | 0.92 |
| Odagiri 2016 | Any general MST Marker | Stored water | 23 | 7 | 23 | 7 | 60 | PR=1 (95% CI: 0.75, 1.33) | 1 |  |  |
| Holcomb 2020 | Any human MST Marker | Any sample | 30 | 9 | 33 | 5 | 77 | PR=0.89 (95% CI: 0.71, 1.1) | 0.27 | PR=0.89 (95% CI: 0.71, 1.1) | 0.27 |
| Fuhrmeister 2020 | Any human MST Marker | Any sample | 109 | 183 | 119 | 181 | 592 | PR=0.99 (95% CI: 0.8, 1.22) | 0.91 | PR=0.96 (95% CI: 0.77, 1.18) | 0.67 |
| Boehm 2016 | Any human MST Marker | Any sample | 26 | 222 | 26 | 223 | 497 | PR=1 (95% CI: 0.57, 1.75) | 0.99 | PR=1 (95% CI: 0.57, 1.76) | 0.99 |
| Odagiri 2016 | Any human MST Marker | Any sample | 22 | 8 | 21 | 9 | 60 | PR=1.05 (95% CI: 0.76, 1.45) | 0.78 |  |  |
| Holcomb 2020 | Any human MST Marker | Source water | 1 | 21 | 0 | 19 | 41 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any human MST Marker | Stored water | 9 | 39 | 5 | 41 | 94 | PR=1.72 (95% CI: 0.57, 5.18) | 0.33 | PR=1.72 (95% CI: 0.57, 5.18) | 0.33 |
| Fuhrmeister 2020 | Any human MST Marker | Stored water | 5 | 310 | 12 | 324 | 651 | PR=0.44 (95% CI: 0.16, 1.23) | 0.12 | PR=0.44 (95% CI: 0.16, 1.23) | 0.12 |
| Boehm 2016 | Any human MST Marker | Stored water | 0 | 245 | 0 | 246 | 491 | Not estimated |  | Not estimated |  |
| Odagiri 2016 | Any human MST Marker | Stored water | 22 | 8 | 21 | 9 | 60 | PR=1.05 (95% CI: 0.76, 1.45) | 0.78 |  |  |
| Fuhrmeister 2020 | Any human MST Marker | Child hands | 30 | 142 | 44 | 122 | 338 | PR=0.66 (95% CI: 0.44, 0.99) | 0.04 | PR=0.7 (95% CI: 0.47, 1.04) | 0.08 |
| Boehm 2016 | Any human MST Marker | Child hands | 7 | 240 | 5 | 241 | 493 | PR=1.39 (95% CI: 0.46, 4.2) | 0.56 | PR=1.39 (95% CI: 0.46, 4.2) | 0.56 |
| Fuhrmeister 2020 | Any human MST Marker | Mother's hands | 58 | 268 | 60 | 265 | 651 | PR=0.96 (95% CI: 0.68, 1.37) | 0.84 | PR=0.96 (95% CI: 0.68, 1.35) | 0.82 |
| Holcomb 2020 | Any human MST Marker | House soil | 20 | 19 | 26 | 18 | 83 | PR=0.87 (95% CI: 0.57, 1.32) | 0.5 | PR=0.83 (95% CI: 0.55, 1.28) | 0.4 |
| Fuhrmeister 2020 | Any human MST Marker | House soil | 68 | 243 | 59 | 261 | 631 | PR=1.19 (95% CI: 0.87, 1.61) | 0.28 | PR=1.24 (95% CI: 0.91, 1.7) | 0.18 |
| Boehm 2016 | Any human MST Marker | House soil | 21 | 226 | 23 | 226 | 496 | PR=0.92 (95% CI: 0.5, 1.71) | 0.79 | PR=0.94 (95% CI: 0.5, 1.75) | 0.84 |
| Holcomb 2020 | Any human MST Marker | Latrine soil | 21 | 9 | 22 | 8 | 60 | PR=0.95 (95% CI: 0.69, 1.32) | 0.78 | PR=0.95 (95% CI: 0.69, 1.32) | 0.78 |
| Holcomb 2020 | Any human MST Marker | Flies in kitchen | 20 | 7 | 13 | 8 | 48 | PR=1.2 (95% CI: 0.77, 1.86) | 0.42 | PR=1.2 (95% CI: 0.77, 1.86) | 0.42 |
| Holcomb 2020 | Any animal MST Marker | Any sample | 13 | 28 | 19 | 29 | 89 | PR=0.8 (95% CI: 0.45, 1.42) | 0.45 | PR=0.8 (95% CI: 0.45, 1.43) | 0.46 |
| Fuhrmeister 2020 | Any animal MST Marker | Any sample | 281 | 13 | 289 | 11 | 594 | PR=1 (95% CI: 0.96, 1.03) | 0.86 | PR=0.99 (95% CI: 0.96, 1.03) | 0.66 |
| Boehm 2016 | Any animal MST Marker | Any sample | 214 | 29 | 216 | 25 | 484 | PR=0.99 (95% CI: 0.93, 1.06) | 0.88 | PR=0.98 (95% CI: 0.92, 1.04) | 0.48 |
| Holcomb 2020 | Any animal MST Marker | Source water | 0 | 22 | 0 | 19 | 41 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any animal MST Marker | Stored water | 0 | 48 | 1 | 45 | 94 | Not estimated |  | Not estimated |  |
| Fuhrmeister 2020 | Any animal MST Marker | Stored water | 229 | 113 | 253 | 109 | 704 | PR=0.96 (95% CI: 0.86, 1.07) | 0.43 | PR=0.97 (95% CI: 0.88, 1.08) | 0.6 |
| Boehm 2016 | Any animal MST Marker | Stored water | 55 | 185 | 80 | 158 | 478 | PR=0.7 (95% CI: 0.51, 0.96) | 0.03 | PR=0.67 (95% CI: 0.49, 0.93) | 0.02 |
| Boehm 2016 | Any animal MST Marker | Child hands | 140 | 102 | 142 | 96 | 480 | PR=0.98 (95% CI: 0.82, 1.16) | 0.78 | PR=0.96 (95% CI: 0.81, 1.14) | 0.67 |
| Fuhrmeister 2020 | Any animal MST Marker | Mother's hands | 344 | 15 | 358 | 9 | 726 | PR=0.98 (95% CI: 0.96, 1.01) | 0.17 | PR=0.98 (95% CI: 0.96, 1.01) | 0.19 |
| Holcomb 2020 | Any animal MST Marker | House soil | 2 | 37 | 1 | 43 | 83 | Not estimated |  | Not estimated |  |
| Fuhrmeister 2020 | Any animal MST Marker | House soil | 272 | 30 | 278 | 27 | 607 | PR=0.99 (95% CI: 0.94, 1.05) | 0.82 | PR=0.99 (95% CI: 0.93, 1.04) | 0.59 |
| Boehm 2016 | Any animal MST Marker | House soil | 178 | 69 | 186 | 63 | 496 | PR=0.96 (95% CI: 0.86, 1.08) | 0.53 | PR=0.96 (95% CI: 0.86, 1.08) | 0.51 |
| Holcomb 2020 | Any animal MST Marker | Latrine soil | 2 | 28 | 0 | 30 | 60 | Not estimated |  | Not estimated |  |
| Holcomb 2020 | Any animal MST Marker | Flies in latrine | 2 | 2 | 8 | 26 | 38 | PR=2.12 (95% CI: 0.54, 8.4) | 0.28 | PR=2.12 (95% CI: 0.54, 8.4) | 0.28 |
| Holcomb 2020 | Any animal MST Marker | Flies in kitchen | 10 | 17 | 10 | 11 | 48 | PR=0.78 (95% CI: 0.44, 1.38) | 0.39 | PR=0.78 (95% CI: 0.44, 1.38) | 0.39 |

**Table S8.**

Baseline covariates by study. Note that Odigari et al. 2016 is not included as data shared from this study were from village water sources and did not have associated covariates from individual households; therefore all estimates for this study are unadjusted.

| **.** | **Boehm 2016** | **Reese 2017** | **Steinbaum 2019** | **Fuhrmeister 2020** | **Holcomb 2020** | **Kwong 2021** |
| --- | --- | --- | --- | --- | --- | --- |
| Household wealth |  |  |  |  |  |  |
| 1 | 6197 (27.3%) | 4539 (19.7%) | 6964 (27.8%) | 11343 (24.1%) | 399 (3.9%) | 4086 (24.3%) |
| 2 | 5954 (26.2%) | 4454 (19.3%) | 7007 (28.0%) | 11196 (23.8%) | 336 (3.3%) | 4476 (26.6%) |
| 3 | 5615 (24.7%) | 4591 (19.9%) | 5738 (22.9%) | 11771 (25.0%) | 543 (5.3%) | 4278 (25.5%) |
| 4 | 4974 (21.9%) | 4714 (20.5%) | 5299 (21.2%) | 12778 (27.1%) | 449 (4.4%) | 3966 (23.6%) |
| Missing | 0 (0%) | 4728 (20.5%) | 24 (0.1%) | 0 (0%) | 8448 (83.0%) | 0 (0%) |
| Number of people in the household |  |  |  |  |  |  |
| <5 | 12397 (54.5%) | 1451 (6.3%) | 7240 (28.9%) | 25590 (54.3%) | 581 (5.7%) | 9426 (56.1%) |
| 5-8 | 9125 (40.1%) | 17171 (74.6%) | 13630 (54.5%) | 18449 (39.2%) | 841 (8.3%) | 6360 (37.8%) |
| >8 | 1218 (5.4%) | 4404 (19.1%) | 2950 (11.8%) | 3049 (6.5%) | 305 (3.0%) | 1020 (6.1%) |
| Missing | 0 (0%) | 0 (0%) | 1212 (4.8%) | 0 (0%) | 8448 (83.0%) | 0 (0%) |
| Number of rooms in the household |  |  |  |  |  |  |
| 1-2 | 0 (0%) | 0 (0%) | 0 (0%) | 0 (0%) | 810 (8.0%) | 0 (0%) |
| >3 | 0 (0%) | 0 (0%) | 0 (0%) | 0 (0%) | 917 (9.0%) | 0 (0%) |
| Missing | 22740 (100%) | 23026 (100%) | 25032 (100%) | 47088 (100%) | 8448 (83.0%) | 16806 (100%) |
| Improved wall |  |  |  |  |  |  |
| 0 | 3588 (15.8%) | 0 (0%) | 23976 (95.8%) | 15951 (33.9%) | 336 (3.3%) | 4428 (26.3%) |
| 1 | 19152 (84.2%) | 0 (0%) | 1056 (4.2%) | 31137 (66.1%) | 1391 (13.7%) | 12378 (73.7%) |
| Missing | 0 (0%) | 23026 (100%) | 0 (0%) | 0 (0%) | 8448 (83.0%) | 0 (0%) |
| Improved floor |  |  |  |  |  |  |
| 0 | 21083 (92.7%) | 0 (0%) | 23732 (94.8%) | 41960 (89.1%) | 41 (0.4%) | 15084 (89.8%) |
| 1 | 1657 (7.3%) | 0 (0%) | 1300 (5.2%) | 5128 (10.9%) | 1686 (16.6%) | 1722 (10.2%) |
| Missing | 0 (0%) | 23026 (100%) | 0 (0%) | 0 (0%) | 8448 (83.0%) | 0 (0%) |
| Improved roof |  |  |  |  |  |  |
| 0 | 368 (1.6%) | 0 (0%) | 8164 (32.6%) | 514 (1.1%) | 0 (0%) | 276 (1.6%) |
| 1 | 22372 (98.4%) | 0 (0%) | 16868 (67.4%) | 46574 (98.9%) | 0 (0%) | 16530 (98.4%) |
| Missing | 0 (0%) | 23026 (100%) | 0 (0%) | 0 (0%) | 10175 (100%) | 0 (0%) |
| Electricity |  |  |  |  |  |  |
| 0 | 10690 (47.0%) | 3780 (16.4%) | 23266 (92.9%) | 18953 (40.3%) | 82 (0.8%) | 7020 (41.8%) |
| 1 | 12050 (53.0%) | 18812 (81.7%) | 1742 (7.0%) | 28135 (59.7%) | 1645 (16.2%) | 9786 (58.2%) |
| Missing | 0 (0%) | 434 (1.9%) | 24 (0.1%) | 0 (0%) | 8448 (83.0%) | 0 (0%) |
| Father in agriculture |  |  |  |  |  |  |
| 0 | 15188 (66.8%) | 10778 (46.8%) | 0 (0%) | 32589 (69.2%) | 0 (0%) | 11466 (68.2%) |
| 1 | 7552 (33.2%) | 9489 (41.2%) | 0 (0%) | 14499 (30.8%) | 0 (0%) | 5340 (31.8%) |
| Missing | 0 (0%) | 2759 (12.0%) | 25032 (100%) | 0 (0%) | 10175 (100%) | 0 (0%) |
| Land owned |  |  |  |  |  |  |
| 0 | 0 (0%) | 8718 (37.9%) | 0 (0%) | 0 (0%) | 0 (0%) | 0 (0%) |
| 1 | 0 (0%) | 11486 (49.9%) | 0 (0%) | 0 (0%) | 0 (0%) | 0 (0%) |
| Missing | 22740 (100%) | 2822 (12.3%) | 25032 (100%) | 47088 (100%) | 10175 (100%) | 16806 (100%) |
| Acres of land owned |  |  |  |  |  |  |
| Mean (SD) | 0.111 (0.129) | NA (NA) | NA (NA) | 0.154 (0.199) | NA (NA) | 0.142 (0.211) |
| Median [Min, Max] | 0.0700 [0.0100, 1.23] | NA [NA, NA] | NA [NA, NA] | 0.0800 [0.0100, 2.10] | NA [NA, NA] | 0.0800 [0.0100, 3.15] |
| Missing | 598 (2.6%) | 23026 (100%) | 25032 (100%) | 1792 (3.8%) | 10175 (100%) | 744 (4.4%) |
| Maternal education |  |  |  |  |  |  |
| No education | 3904 (17.2%) | 0 (0%) | 0 (0%) | 6652 (14.1%) | 72 (0.7%) | 2484 (14.8%) |
| Incomplete Primary | 0 (0%) | 9113 (39.6%) | 12892 (51.5%) | 0 (0%) | 749 (7.4%) | 0 (0%) |
| Primary | 8228 (36.2%) | 2401 (10.4%) | 6028 (24.1%) | 14206 (30.2%) | 313 (3.1%) | 5424 (32.3%) |
| Secondary | 10608 (46.6%) | 6205 (26.9%) | 5940 (23.7%) | 26230 (55.7%) | 547 (5.4%) | 8898 (52.9%) |
| More than secondary | 0 (0%) | 685 (3.0%) | 0 (0%) | 0 (0%) | 0 (0%) | 0 (0%) |
| Missing | 0 (0%) | 4622 (20.1%) | 172 (0.7%) | 0 (0%) | 8494 (83.5%) | 0 (0%) |
| Maternal age |  |  |  |  |  |  |
| Mean (SD) | 23.7 (5.18) | NA (NA) | 26.4 (6.33) | 23.7 (5.07) | NA (NA) | 23.9 (5.03) |
| Median [Min, Max] | 23.0 [15.0, 42.0] | NA [NA, NA] | 25.5 [14.9, 47.9] | 23.0 [15.0, 41.0] | NA [NA, NA] | 24.0 [15.0, 43.0] |
| Missing | 0 (0%) | 23026 (100%) | 280 (1.1%) | 0 (0%) | 10175 (100%) | 24 (0.1%) |