- Compounds with therapeutic potential against novel respiratory 2019 coronavirus
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AAC Accepted Manuscript Posted Online 9 March 2020 Antimicrob. Agents Chemother. doi:10.1128/AAC.00399-20

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fighting SARS-CoV-2 infections.

Abstract

Currently, the expansion of the novel human respiratory coronavirus (known as: SARS-CoV-2, COVID-2019, or 2019-nCoV) has stressed the need for therapeutic alternatives to alleviate and stop this new epidemic. The previous epidemics of high-morbidity human coronaviruses, such as the acute respiratory syndrome coronavirus (SARS-CoV) in 2003, and the Middle East respiratory syndrome corona virus (MERS-CoV) in 2012, prompted the characterization of compounds that could be potentially active against the currently emerging novel coronavirus SARS-CoV-2. The most promising compound is remdesivir (GS-5734), a nucleotide analog prodrug currently in clinical trials for treating Ebola virus infections. Remdesivir inhibited the replication of SARS-CoV and MERS-CoV in tissue cultures, and it displayed efficacy in nonhuman animal models. In addition, a combination of the human immunodeficiency virus type 1 (HIV-1) protease inhibitors, lopinavir/ritonavir, and interferon beta (LPV/RTV-INFb) were shown to be effective in patients infected with SARS-CoV. LPV/RTV-INFb also improved clinical parameters in marmosets and mice infected with MERS-CoV. Remarkably, the therapeutic efficacy of remdesivir appeared to be superior to that of LPV/RTV-INFb against MERS-CoV in a transgenic humanized mice model. The relatively high mortality rates associated with these three novel human coronavirus infections, SARS-CoV, MERS-CoV, and SARS-CoV-2, has suggested that pro-inflammatory responses might play a role in the pathogenesis. It remains unknown whether the generated inflammatory state should be targeted. Therapeutics that target the coronavirus alone might not be able to reverse highly pathogenic infections. This

minireview aimed to provide a summary of therapeutic compounds that showed potential in

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38 of China (1). In the early stages of this pneumonia, patients developed severe acute respiratory infection symptoms, and some patients rapidly developed acute respiratory distress syndrome 39 40 (2). Real time RT-PCR and deep sequencing analysis from lower respiratory tract samples identified a novel human coronavirus, now called SARS-CoV-2 (3-5). By the end of January, 41 42 2020, nearly 50,000 confirmed cases were reported in China, and the first confirmed cases were 43 reported in Thailand, Nepal, Republic of Korea, USA, Singapore, France, Viet Nam, Canada, Australia, Malaysia, Germany, UAE, Finland, Italy, Cambodia, Sri Lanka, the Russian Federation, 44 45 Spain, Sweden, India, and the Philippines. Among the patients with confirmed cases, most were aged 30–80 years and had mild infections (80%). The fatality rate was around 2% (6). 46 47 Coronaviruses can cause different types of infections in diverse animals. In humans, they mainly 48 49 produce respiratory tract infections, as observed with SARS-CoV and MERS-Cov (7, 8). 50 Sequencing and phylogenetic analyses have shown that the novel SARSCoV-2 virus is closely 51 related to a group of human SARS-like coronaviruses and bat SARS-related coronaviruses (9-52 11). The origin of SARSCoV-2 remains unclear; it is unknown how it was first transmitted to 53 humans. The high prevalence of SARS-related coronaviruses in bats has suggested that a bat 54 coronavirus might have jumped into a civet or some other mammal, and from there to humans, which started the former 2003 SARS epidemic. Initial confirmed cases of SARSCoV-2 were 55 associated with Huanan seafood and live animal markets. However, no animal source has been 56

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On December 30, 2019, a cluster of 27 pneumonia cases (including 7 severe cases) of unknown

origin emerged in Wuhan (Hubei, China) and were reported to the National Health Commission

identified to date, and spillover events may continue to occur. Although bats might be the

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source of SARSCoV-2, it is critical to identify the intermediate species to stop the current spread and to prevent future human SARS-related coronavirus epidemics. A key question is whether the current SARSCoV-2 epidemic is similar to other SARS outbreaks or whether it shows different features. The epidemiological and clinical characteristics of SARSCoV-2 indicate that this new outbreak is different from the 2003-SARS. SARSCoV-2 displays higher transmissibility and lower mortality compared to the 2003-SARS (1, 3, 4). SARSCoV-2 has shown efficient intra-familial spread (4). The asymptomatic period of SARSCoV-2 infections oscillates between 2 and 14 days, and some individuals probably transmit the virus without developing any disease symptoms. It remains to be elucidated whether this virus replicates more readily in the upper airway than SARS-CoV and MEERS-CoV and whether it is similar to other human coronaviruses (HCoV) that cause colds, but not pneumonia. It will be necessary to identify molecular determinants that mediate transmission from animal to human, and from human to human. Of note, in the novel SARS-CoV-2, the nucleotide sequence of the external ectodomain in the spike protein receptor-binding domain is different from that of the 2003 SARS-CoV. When individual bat coronavirus spike genes were introduced into SARS-CoV infectious clones, the SARS-CoV/bat-CoV spike viruses could bind to the human, bat, or civet angiotensin converting enzyme 2 (ACE2) cellular receptor (12). Understanding the interaction between this novel SARS-CoV-2 spike protein and the host ACE2 receptor might reveal how this virus overcame the species barrier between animals and humans. As discussed below, this

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information might promote the design of effective antivirals.

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To predict new zoonotic coronavirus jumps across species and to understand the rate of virus spread among people, it is crucial to determine whether SARSCoV-2 is mutating to improve its binding to human receptors for infection. As an RNA virus, SARS-CoV-2 has intrinsic genetic variability, which results in a high mutation rate. Moreover, coronaviruses have the largest genomes (\sim 30 kb) among RNA viruses. However, part of their sequence encodes a proofreading 3' exonuclease that can increase replication fidelity (13). It has been suggested that any adaptation in the SARS-CoV-2 sequence that might make it more efficient at transmitting from person to person might also increase its virulence (14). However, this mechanism could lead to a genetic bottleneck, known as Muller's ratchet, which could significantly decrease viral fitness, (15). Muller's ratchet predicts that, when mutation rates are high and a significant proportion of mutations are deleterious, a type of irreversible ratchet mechanism will gradually reduce the mean fitness of small populations of asexual organisms. Because genetic bottlenecks for RNA viruses often occur during respiratory droplet transmissions, the SARS-CoV-2 is expected to become less virulent through human to human transmissions (16). From the public health perspective, we urgently need to develop an effective vaccine and antiviral therapeutics to stop the SARS-CoV-2 epidemic. Moreover, social and economic issues generated by this epidemic also call for rapid interventions. This review focuses on the potential of repurposing preexisting compounds that might provide new opportunities for treating people infected with SARS-CoV-2. Previous work with SARS-CoV and MERS-CoV has

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provided an opportunity to accelerate the identification of meaningful therapies for fighting the

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novel SARS-CoV-2 epidemic. Nevertheless, we must be aware that, currently, no compound that targets SARS-CoV or MERS-CoV has moved beyond phase 1 trials. The most promising antiviral for fighting SARS-CoV-2 is remdesivir (GS-5734). Remdesivir is an adenosine nucleotide analogue prodrug with broad-spectrum antiviral activity against filoviruses, paramyxoviruses, pneumoviruses, and pathogenic coronaviruses, like SARS-CoV and MERS-CoV (17). Pharmacokinetic studies have been completed and clinical trials are ongoing for testing remdesivir efficacy in treating Ebola virus (18). Previous studies have indicated that nucleotide analogues generally show low efficacy against coronaviruses, due to the virus exonuclease proofreading enzyme. Nevertheless, remdesivir was effective against SARS-CoV, MERS-CoV, and bat-CoV strains (17). In tissue cultures, remdesivir displayed half-maximum effective concentrations (EC50s) of 0.069 for SARS-CoV and 0.074 µM for MERS-CoV. Of note, tissue culture studies have shown that remdesivir is also active in the submicromolar EC50 range against a number of highly divergent coronaviruses, including the endemic human CoVs, OC43 (HCoV-OC43) and 229E (HCoV-229E). Thus, remdesivir has broad-spectrum anti-CoV activity (19). In a mouse model of SARS-CoV pathogenesis, prophylactic and early therapeutic administration of remdesivir significantly reduced the lung viral load. Viral titers were reduced by >2 orders of magnitude on day 4 or 5 post infection. Remdesivir improved the clinical signs of disease and respiratory function compared to untreated control animals (17). Comparable

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mouse transgenic model. In that model, a humanized MERS-CoV receptor (dipeptidyl peptidase

results were obtained with MERS-CoV in prophylactic studies carried out with a MERS-CoV

4, hDPP4) was expressed and carboxylesterase 1c (Ces1c) was deleted to improve the

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demonstrated by propagating the virus in tissue culture. After 23 passages in the presence of drug, two mutations were identified (F276L and V553L) in the viral RNA-dependent RNA polymerase gene. These mutations increased the replication capacity of the virus in the presence of remdesivir (21). However, these amino acid changes decreased the viral fitness and attenuated SARS-CoV pathogenesis in mice (21). The efficacy of prophylactic and therapeutic remdesivir treatment was recently tested in a nonhuman primate (rhesus macaque) model of MERS-CoV infection (22). When prophylactic remdesivir treatment was initiated 24 h prior to inoculation, MERS-CoV was prevented from inducing clinical disease and inhibited from replicating in respiratory tissues, which prevented the formation of lung lesions. Similar results were obtained when therapeutic remdesivir treatment was initiated at 12 h after virus inoculation (22). Human safety data are available for remdesivir (18); thus, human trials can be initiated for testing the efficacy of this compound against novel coronaviruses. Therapies that are approved by the Food and Drug Administration (FDA) have been evaluated for antiviral activity against SARS-CoV and MERS-CoV. For example, lopinavir (LPV), a human immunodeficiency virus 1 (HIV-1) protease inhibitor, was combined with ritonavir (RTV) to increase the LPV half-life. LPV/RTV was shown to be effective against SARS-CoV in patients and in tissue culture. The estimated EC50 in fetal rhesus kidney-4 cells was 4 μg/ml (23). LPV/RTV also reduced weight loss, clinical scores, viral titers, and disease progression in marmosets

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pharmacokinetics of nucleotide prodrugs (20). Remdesivir specificity for coronavirus was

infected with MERS-CoV (24). Nevertheless, the antiviral activity of LPV against MERS-CoV in

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of 8 µM was reported in Huh7 cells (26). Clinical observations in animals and humans showed that MERS-CoV infections were mediated by both virus replication and host inflammatory responses. Those findings led to explorations of combination therapies that included types I and II interferons (IFN I and II). Interferon beta (IFNb) displayed the best efficacy, with EC50s of 1.37-17 IU/ml, for reducing MERS-CoV replication in tissue culture (25, 27). Similar to LPV/RTV, clinical improvements with IFNb were observed in common marmosets infected with MERS-CoV (24). In the Kingdom of South Arabia, an ongoing randomized control trial (MIRACLE Trial) was initiated to determine whether the combination of LPV/RTV and IFNb could improve clinical outcomes in MERS-CoV infections (28). Importantly, another controlled trial was launched in China to test the efficacy of LPV/RTV and IFN α -2b in hospitalized patients with SARS-CoV-2 infections (ChiCTR2000029308). The prophylactic and therapeutic properties of remdesivir and LPV/RTV-IFNb were compared in a humanized transgenic mouse MERS-CoV infection model (29). Remdesivir improved pulmonary function, reduced lung viral loads, and ameliorated severe lung pathology. In contrast, prophylactic LPV/RTV-IFNb only slightly reduced viral loads and did not impact other disease parameters, and therapeutic LPV/RTV-IFNb improved pulmonary function, but did not reduce virus replication or severe lung pathology (29). Overall, these results indicated that remdesivir showed more potential than LPV/RTV-IFNb for treating MERS-CoV infections.

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tissue culture remains controversial. No optimal EC50 was found in Vero cells (25), but an EC50

Ribavirin, a guanosine analogue, is an antiviral compound used to treat several virus infections, including respiratory syncytial virus, hepatitis C virus, and some viral hemorrhagic fevers. In most cases, ribavirin is combined with IFN. Ribavirin was first marketed in 1980 for the treatment of respiratory syncytial virus in children. Although promising results were obtained with ribavirin and IFNα-2b in a MERS-CoV rhesus macaque model (30), data have been conflicting on patients with MERS-CoV infections that were treated with a combination of ribavirin and IFN (either $\alpha 2a$ or $\beta 1$) (31). However, ribavirin reduces hemoglobin concentrations, an undesirable side effect in patients with respiratory disorders. This feature reduces its potential as an antiviral against SARS-CoV-2.

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Work with influenza virus has shown that monoclonal and polyclonal antibodies can be useful prophylactic and therapeutic tools. Several antibodies have been shown to bind influenza virus hemagglutinin and inhibit virus replication (12). For example, human immunoglobulin G1 (IgG1) monoclonal antibody (MHAA4549A) binds to a highly conserved epitope on the stalk of influenza A hemagglutinin. In a phase 2 human influenza A virus challenge study, MHAA4549A significantly reduced the clinical symptoms and viral burden relative to placebo (32). Another example is VIS410, a monoclonal antibody engineered to target all known influenza A strains. A phase 2a trial showed that VIS410 had some clinical benefits (33). Current development efforts in monoclonal and polyclonal antibodies against coronaviruses are mainly targeting MERS-CoV. In a phase 1 clinical trial, a human polyclonal antibody, SAB-301, which is generated in transchromosomic cattle, was found to be safe and well tolerated in healthy participants. (34). However, therapeutic treatment with human monoclonal antibodies did not protect against the Downloaded from http://aac.asm.org/ on March 28, 2020 by guest

severe disease or the loss of lung function induced by MERS-CoV in animal models (20). The lack of viral sequence homology among different human coronaviruses suggests that current investigational antibody-based therapeutics will not be effective against novel virus variants. Nevertheless, immune-based therapies should be not discarded, when considering future treatments for novel coronaviruses.

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Another potential treatment option could be the use of novel coronavirus sera prepared from the blood of patients in convalescence (convalescent sera). Passive immunization is well established for viral infection prophylaxis. Polyclonal antibody products have been licensed that target cytomegalovirus, hepatitis B virus, and varicella-zoster virus. A meta-analysis of reports on the 1918 influenza A (H1N1) epidemic concluded that early administration of convalescent blood products reduced the absolute risk of pneumonia-related death from 37% to 16% (35). Nevertheless, the appropriate titer of convalescent sera antibody that is required for therapeutic efficacy against SARS-CoV-2 remains to be determined. Moreover, additional studies performed with influenza virus have produced controversial results regarding the clinical benefit of administering high titers of anti-influenza immunoglobulins (36). Finally, it remains unclear whether a sufficient pool of potential donors is feasible. Work carried out with MERS-CoV showed that sera from patients recovering from infections did not contain sufficient antibody titers for therapeutic use (37).

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Another interesting therapeutic alternative that was previously explored with influenza virus is to target cellular components involved in the host inflammatory response to the infection. For

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example, the activation of the inflammatory response to an infection can induce a cytokine outburst that results in an acute lung injury. An example of a therapy for this type of infection has been to target the cellular toll-like receptor 4 (TLR4) with specific antibodies. TLR4 is a transmembrane protein that belongs to the pattern recognition receptor (PRR) family. The prototype pathogen-associated molecular pattern (PAMP) that TLR4 recognizes is the gramnegative bacteria, endotoxin, lipopolysaccharide (LPS). TLR4 has been implicated in the pathology associated with other infections and with tissue damage caused by non-infectious insults. TLR4 activation leads to the NF-kB intracellular signaling pathway and inflammatory cytokine production, which activate the innate immune system. Interestingly, TLR4-null mice were highly resistant to infection by the mouse-adapted influenza A virus (38). Thus, protection against influenza infections was achieved by targeting TLR4 with small molecule antagonists, like TAK-242, or with anti-TLR4-specific antibodies (39, 40). Indeed, targeting a cellular protein would overcome the drawbacks associated with virus or coronavirus genetic heterogeneity. The high mortally rates observed in some emerging respiratory diseases induced by viruses like MERS-CoV, SARS-CoV, and novel influenza A strains (H5N1) has given rise to the hypothesis that the pro-inflammatory response might be involved in the disease pathogenesis. Consequently, immunosuppressants (e.g., corticosteroids) might be used as an adjunct for treating severe forms of the disease. However, the therapeutic use of immunosuppressants is not free of controversy. To date, no conclusive results have been found for the effects of immunosuppressants in severe influenza virus infections (12). Furthermore, the use of

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corticosteroids to treat influenza virus has been associated with an increased risk of

superinfection, prolonged viral replication, and an increased risk of death (41). In contrast, corticosteroid treatment for MERS-CoV infections was not significantly associated with mortality, although a delay in MERS-CoV RNA clearance was observed (42). Further studies should be performed to clarify the potential clinical benefit of prescribing immunosuppressants for coronavirus infections.

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To end this minireview, we will discuss an interesting potential antiviral strategy. The spike protein of SARS-CoV mediates viral entry into target cells. Intriguingly, the cleavage and activation of the SARS-CoV spike protein by a host cell protease is essential for infectious viral entry (43). This host protease could be a type II transmembrane serine protease, TMPRSS2, which was shown to cleave and activate SARS-CoV spike protein in cell cultures. Therefore, TMPRSS2 is a potential a target for antiviral interventions. For example, the serine protease inhibitor, camostat mesylate, inhibits the enzymatic activity of TMPRSS2 (44). Recently, K11777, a cysteine protease inhibitor, was shown in tissue cultures to inhibit SARS-CoV and MERS-CoV replication in the sub-nanomolar range (45). Future tissue culture and animal model studies should be conducted to clarify the potential antiviral activity of targeting TMPRSS2.

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By the end of February 2020, two months after the first cases of SARS-CoV-2 were reported in China, several hundreds of new infection cases had been registered, mainly in other Asian regions and Europe. This news has strongly suggested that we are in the thick of a SARS-CoV-2 pandemic. Social alarm and health authorities have called for the development of therapeutic alternatives for fighting the current, and possibly new, coronavirus epidemics. Animal models

and clinical studies are urgently needed for evaluating the effectiveness and safety of promising
antiviral compounds that target the virus and/or the immunopathology involved in the host
responses. The identification and characterization of novel compounds and therapeutic
alternatives will be required to better control this probable pandemic outbreak.
Funding. This work was supported by the Spanish Ministry of Science and Innovation [SAF2016-
75277-R].
Conflicts of interest. The author declares no conflict of interest.

References

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- WHO. 2020. Novel coronavirus (2019-nCoV) situation reports. World Health 266 1. Organisation. https://www.who.int/emergencies/diseases/novel-coronavirus-267 2019/situation-reports/. 268
- Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, 269 2. 270 Wei Y, Wu W, Xie X, Yin W, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao 271 Z, Jin Q, Wang J, Cao B. 2020. Clinical features of patients infected with 2019 novel 272 coronavirus in Wuhan, China. Lancet 395:497-506.
- 273 Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, Qiu Y, Wang J, Liu Y, Wei Y, Xia J, Yu T, 274 Zhang X, Zhang L. 2020. Epidemiological and clinical characteristics of 99 cases of 2019 275 novel coronavirus pneumonia in Wuhan, China: a descriptive study. Lancet 395:507-513.
- 276 Chan JFW, Yuan S, Kok KH, To KKW, Chu H, Yang J, Xing F, Liu J, Yip CCY, Poon RWS, Tsoi 277 HW, Lo SKF, Chan KH, Poon VKM, Chan WM, Ip JD, Cai JP, Cheng VCC, Chen H, Hui CKM, 278 Yuen KY. 2020. A familial cluster of pneumonia associated with the 2019 novel 279 coronavirus indicating person-to-person transmission: a study of a family cluster. Lancet 280 395:514-523.
- 281 5. Gorbalenya AE. 2020. Severe acute respiratory syndrome-related coronavirus – The 282 species and its viruses, a statement of the Coronavirus Study Group. bioRxiv 283 2020.02.07.937862.
- 284 6. Team TNCPERE. 2020. The Epidemiological Characteristics of an Outbreak of 2019 Novel Coronavirus Diseases (COVID-19) — China, 2020. China CDC Wkly 2. 285
- 286 7. Zaki AM, Van Boheemen S, Bestebroer TM, Osterhaus ADME, Fouchier RAM. 2012. 287 Isolation of a novel coronavirus from a man with pneumonia in Saudi Arabia. N Engl J 288 Med 367:1814-1820.
- Drosten C, Günther S, Preiser W, Van der Werf S, Brodt HR, Becker S, Rabenau H, Panning 289 8. 290 M, Kolesnikova L, Fouchier RAM, Berger A, Burguière AM, Cinatl J, Eickmann M, Escriou 291 N, Grywna K, Kramme S, Manuguerra JC, Müller S, Rickerts V, Stürmer M, Vieth S, Klenk HD, Osterhaus ADME, Schmitz H, Doerr HW. 2003. Identification of a novel coronavirus in 292 293 patients with severe acute respiratory syndrome. N Engl J Med 348:1967–1976.
- 294 9. Zhou P, Yang X-L, Wang X-G, Hu B, Zhang L, Zhang W, Si H-R, Zhu Y, Li B, Huang C-L, Chen 295 H-D, Chen J, Luo Y, Guo H, Jiang R-D, Liu M-Q, Chen Y, Shen X-R, Wang X, Zheng X-S, Zhao 296 K, Chen Q-J, Deng F, Liu L-L, Yan B, Zhan F-X, Wang Y-Y, Xiao G-F, Shi Z-L. 2020. A 297 pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature 298 Feb 3[Online ahead of print].
- Lu R, Zhao X, Li J, Niu P, Yang B, Wu H, Wang W, Song H, Huang B, Zhu N, Bi Y, Ma X, Zhan 299 10. 300 F, Wang L, Hu T, Zhou H, Hu Z, Zhou W, Zhao L, Chen J, Meng Y, Wang J, Lin Y, Yuan J, Xie 301 Z, Ma J, Liu WJ, Wang D, Xu W, Holmes EC, Gao GF, Wu G, Chen W, Shi W, Tan W. 2020. 302 Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for 303 virus origins and receptor binding. Lancet 395:565-574.
- 304 Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, Zhao X, Huang B, Shi W, Lu R, Niu P, Zhan F, 305 Ma X, Wang D, Xu W, Wu G, Gao GF, Tan W. 2020. A Novel Coronavirus from Patients with Pneumonia in China, 2019. N Engl J Med 382:727-733. 306
- 12. Beigel JH, Nam HH, Adams PL, Krafft A, Ince WL, El-Kamary SS, Sims AC. 2019. Advances 307 308 in respiratory virus therapeutics – A meeting report from the 6th isirv Antiviral Group

- 309 conference. Antiviral Res 167:45-67.
- Bradwell K, Combe M, Domingo-Calap P, Sanjuán R. 2013. Correlation between mutation 310 13. rate and genome size in riboviruses: Mutation rate of bacteriophage QB. Genetics 311 312 195:243-251.
- 14. Wang C, Horby PW, Hayden FG, Gao GF. 2020. A novel coronavirus outbreak of global 313 health concern. Lancet 395:470-473. 314
- Chao L. 1990. Fitness of RNA virus decreased by Muller's ratchet. Nature 348:454-455. 315 15.
- 316 Duarte E, Clarke D, Moya A, Domingo E, Holland J. 1992. Rapid fitness losses in
- 317 mammalian RNA virus clones due to Muller's ratchet. Proc Natl Acad Sci U S A 89:6015-6019. 318
- 319 17. Sheahan TP, Sims AC, Graham RL, Menachery VD, Gralinski LE, Case JB, Leist SR, Pyrc K, 320 Feng JY, Trantcheva I, Bannister R, Park Y, Babusis D, Clarke MO, MacKman RL, Spahn JE, 321 Palmiotti CA, Siegel D, Ray AS, Cihlar T, Jordan R, Denison MR, Baric RS. 2017. Broad-322 spectrum antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses. Sci Transl 323 Med 9.
- 324 18. Mulangu S, Dodd LE, Davey RT, Mbaya OT, Proschan M, Mukadi D, Manzo ML, Nzolo D, Oloma AT, Ibanda A, Ali R, Coulibaly S, Levine AC, Grais R, Diaz J, Clifford Lane H, 325 Muyembe-Tamfum JJ, Sivahera B, Camara M, Kojan R, Walker R, Dighero-Kemp B, Cao H, 326 327 Mukumbayi P, Mbala-Kingebeni P, Ahuka S, Albert S, Bonnett T, Crozier I, Duvenhage M, Proffitt C, Teitelbaum M, Moench T, Aboulhab J, Barrett K, Cahill K, Cone K, Eckes R, 328 329 Hensley L, Herpin B, Higgs E, Ledgerwood J, Pierson J, Smolskis M, Sow Y, Tierney J, 330 Sivapalasingam S, Holman W, Gettinger N, Vallée D, Nordwall J. 2019. A randomized, 331 controlled trial of Ebola virus disease therapeutics. N Engl J Med 381:2293-2303.

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- 19. Brown AJ, Won JJ, Graham RL, Dinnon KH, Sims AC, Feng JY, Cihlar T, Denison MR, Baric 332 333 RS, Sheahan TP. 2019. Broad spectrum antiviral remdesivir inhibits human endemic and 334 zoonotic deltacoronaviruses with a highly divergent RNA dependent RNA polymerase. 335 Antiviral Res 169:104541.
- Cockrell AS, Yount BL, Scobey T, Jensen K, Douglas M, Beall A, Tang XC, Marasco WA, 336 20. 337 Heise MT, Baric RS. 2016. A mouse model for MERS coronavirus-induced acute 338 respiratory distress syndrome. Nat Microbiol 2:16226.
- 339 21. Agostini ML, Andres EL, Sims AC, Graham RL, Sheahan TP, Lu X, Smith EC, Case JB, Feng JY, Jordan R, Ray AS, Cihlar T, Siegel D, Mackman RL, Clarke MO, Baric RS, Denison MR. 340 341 2018. Coronavirus susceptibility to the antiviral remdesivir (GS-5734) is mediated by the 342 viral polymerase and the proofreading exoribonuclease. MBio 9:e00221-18.
- 343 22. de Wit E, Feldmann F, Cronin J, Jordan R, Okumura A, Thomas T, Scott D, Cihlar T, 344 Feldmann H. 2020. Prophylactic and therapeutic remdesivir (GS-5734) treatment in the 345 rhesus macaque model of MERS-CoV infection. Proc Natl Acad Sci 201922083.
- Chu CM, Cheng VCC, Hung IFN, Wong MML, Chan KH, Chan KS, Kao RYT, Poon LLM, Wong 346 23. 347 CLP, Guan Y, Peiris JSM, Yuen KY. 2004. Role of lopinavir/ritonavir in the treatment of 348 SARS: Initial virological and clinical findings. Thorax 59:252–256.
- 349 24. Chan JFW, Yao Y, Yeung ML, Deng W, Bao L, Jia L, Li F, Xiao C, Gao H, Yu P, Cai JP, Chu H, Zhou J, Chen H, Qin C, Yuen KY. 2015. No TitleTreatment With Lopinavir/Ritonavir or 350 Interferon-β1b Improves Outcome of MERS-CoV Infection in a Nonhuman Primate Model 351 352 of Common Marmoset. J Infect Dis 212:1904-1913.

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- Chan JFW, Chan KH, Kao RYT, To KKW, Zheng BJ, Li CPY, Li PTW, Dai J, Mok FKY, Chen H, 353 25. 354 Hayden FG, Yuen KY. 2013. Broad-spectrum antivirals for the emerging Middle East respiratory syndrome coronavirus. J Infect 67:606-616. 355
- De Wilde AH, Jochmans D, Posthuma CC, Zevenhoven-Dobbe JC, Van Nieuwkoop S, 356 26. Bestebroer TM, Van Den Hoogen BG, Neyts J, Snijder EJ. 2014. Screening of an FDA-357 approved compound library identifies four small-molecule inhibitors of Middle East 358 359 respiratory syndrome coronavirus replication in cell culture. Antimicrob Agents 360 Chemother 58:4875-4884.
- 361 Hart BJ, Dyall J, Postnikova E, Zhou H, Kindrachuk J, Johnson RF, Olinger GG, Frieman MB, 362 Holbrook MR, Jahrling PB, Hensley L. 2014. Interferon-β and mycophenolic acid are 363 potent inhibitors of middle east respiratory syndrome coronavirus in cell-based assays. J 364 Gen Virol 95:571-577.
- 365 28. Arabi YM, Alothman A, Balkhy HH, Al-Dawood A, AlJohani S, Al Harbi S, Kojan S, Al Jeraisy 366 M, Deeb AM, Assiri AM, Al-Hameed F, AlSaedi A, Mandourah Y, Almekhlafi GA, Sherbeeni 367 NM, Elzein FE, Memon J, Taha Y, Almotairi A, Maghrabi KA, Qushmaq I, Al Bshabshe A, Kharaba A, Shalhoub S, Jose J, Fowler RA, Hayden FG, Hussein MA, Martin GS, Schoenfeld 368 DA, Walmsley SL, Carson S, Harbi S Al, Jeraisy M Al, Muhaidib M Al, Musharaf S, Anizi H 369 Al, Dael R, AlMazroa M, Asiri A, Memish ZA, Ghazal SS, Alfaraj SH, Harthy A Al, Sulaiman 370 371 M Al, Mady A, Ahmad A, Ghaleb A Almekhlafi, Muhammed R, Samirrai S Al, Awad S, Cabal RC, Onazi B Al, Aljuhani M, Vince M, Enani M Al, Algurashi A, Alenezi F, Alkhani N, 372 Thagafi A, Oraabi O Al, Rifai J, Elsamadisi P, Medhat SH, Basher SAB, Abduldhaher M, 373 374 Bajhamoum W, Alahsa SS, Bashir S, Al-Dossary I, Al-Muhainy Dammam B, Khobar SS Al, 375 Alshahrani MS, Al Jabri A, Farid M, Alaidarous A, Alseraihi W, Shahada H, Taif JS. 2018. Treatment of Middle East Respiratory Syndrome with a combination of lopinavir-376 377 ritonavir and interferon-β1b (MIRACLE trial): Study protocol for a randomized controlled 378
- 379 29. Sheahan TP, Sims AC, Leist SR, Schäfer A, Won J, Brown AJ, Montgomery SA, Hogg A, Babusis D, Clarke MO, Spahn JE, Bauer L, Sellers S, Porter D, Feng JY, Cihlar T, Jordan R, 380 381 Denison MR, Baric RS. 2020. Comparative therapeutic efficacy of remdesivir and 382 combination lopinavir, ritonavir, and interferon beta against MERS-CoV. Nat Commun 383
- 30. Falzarano D, De Wit E, Rasmussen AL, Feldmann F, Okumura A, Scott DP, Brining D, 384 385 Bushmaker T, Martellaro C, Baseler L, Benecke AG, Katze MG, Munster VJ, Feldmann H. 386 2013. Treatment with interferon-α2b and ribavirin improves outcome in MERS-CoV-387 infected rhesus macagues. Nat Med 19:1313–1317.
- 388 31. Arabi, YM, Shalhoub, S, Omari, AA, Mandourah, Y, Al-Hameed, F, Sindi A, Alraddadi, B, 389 Motairi, AA, Khatib, KA, Mommin, AA, Qushmaq, IA, Mady A, Solaiman, O, Aithan, AA, Balkhy, HH, Al-Raddadi, R, Rajab, A, Mekhlafi G, Harthy, AA, Kharaba, A, Al-Jabbary, A, 390 391 Pinto, R, Sadat, M, Mutairi, HA Q, EA, Jose, J, Deeb, AM, Merson, L, Hayden, FG, Fowler, 392 R, Aldawood A. 2017. Effect of ribavirin and interferon on the outcome of critically ill 393 patients with MERS. Am J Respir Crit Care Med 195:A6067.
- 394 32. McBride JM, Lim JJ, Burgess T, Deng R, Derby MA, Maia M, Horn P, Siddiqui O, Sheinson D, Chen-Harris H, Newton EM, Fillos D, Nazzal D, Rosenberger CM, Ohlson MB, Lambkin-395 396 Williams R, Fathi H, Harris JM, Tavela JA. 2017. Phase 2 randomized trial of the safety and

- 397 efficacy of MHAA4549A, a broadly neutralizing monoclonal antibody, in a human influenza a virus challenge model. Antimicrob Agents Chemother 61: e01154-17. 398
- Hershberger E, Sloan S, Narayan K, Hay CA, Smith P, Engler F, Jeeninga R, Smits S, Trevejo 399 33. J, Shriver Z, Oldach D. 2019. Safety and efficacy of monoclonal antibody VIS410 in adults 400 401 with uncomplicated influenza A infection: Results from a randomized, double-blind, 402 phase-2, placebo-controlled study. EBioMedicine 40:574-582.
- 403 34. Beigel JH, Voell J, Kumar P, Raviprakash K, Wu H, Jiao JA, Sullivan E, Luke T, Davey RT. 404 2018. Safety and tolerability of a novel, polyclonal human anti-MERS coronavirus 405 antibody produced from transchromosomic cattle: a phase 1 randomised, double-blind, single-dose-escalation study. Lancet Infect Dis 18:410-418. 406
- 407 35. Luke TC, Kilbane EM, Jackson JL, Hoffman SL. 2006. Meta-analysis: Convalescent blood products for Spanish influenza pneumonia: A future H5N1 treatment? Ann Intern Med. 408 409 American College of Physicians 145:599-609.
- Hung IFN, To KKW, Lee CK, Lee KL, Yan WW, Chan K, Chan WM, Ngai CW, Law KI, Chow 410 36. 411 FL, Liu R, Lai KY, Candy CC, Liu SH, Chan KH, Lin CK, Yuen KY. 2013. Hyperimmune IV 412 immunoglobulin treatment: A multicenter double-blind randomized controlled trial for patients with severe 2009 influenza A(H1N1) infection. Chest 144:464-473. 413
- 414 37. Arabi Y, Balkhy H, Hajeer AH, Bouchama A, Hayden FG, Al-Omari A, Al-Hameed FM, Taha 415 Y, Shindo N, Whitehead J, Merson L, AlJohani S, Al-Khairy K, Carson G, Luke TC, Hensley L, Al-Dawood A, Al-Qahtani S, Modjarrad K, Sadat M, Rohde G, Leport C, Fowler R. 2015. 416 417 Feasibility, safety, clinical, and laboratory effects of convalescent plasma therapy for 418 patients with Middle East respiratory syndrome coronavirus infection: a study protocol. 419 Springerplus 4:1-8.

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- 420 38. Shirey KA, Lai W, Scott AJ, Lipsky M, Mistry P, Pletneva LM, Karp CL, McAlees J, Gioannini 421 TL, Weiss J, Chen WH, Ernst RK, Rossignol DP, Gusovsky F, Blanco JCG, Vogel SN. 2013. 422 The TLR4 antagonist Eritoran protects mice from lethal influenza infection. Nature 423 497:498-502.
- 424 Perrin-Cocon L, Aublin-Gex A, Sestito SE, Shirey KA, Patel MC, André P, Blanco JC, Vogel 425 SN, Peri F, Lotteau V. 2017. TLR4 antagonist FP7 inhibits LPS-induced cytokine production 426 and glycolytic reprogramming in dendritic cells, and protects mice from lethal influenza 427 infection. Sci Rep 7:40791.
- 40. Piao W, Shirey KA, Ru LW, Lai W, Szmacinski H, Snyder GA, Sundberg EJ, Lakowicz JR, 428 429 Vogel SN, Toshchakov VY. 2015. A Decoy Peptide that Disrupts TIRAP Recruitment to 430 TLRs Is Protective in a Murine Model of Influenza. Cell Rep 11:1941–1952.
- 431 41. Rodrigo C, Leonardi-Bee J, Nguyen-Van-Tam J, Lim WS. 2016. Corticosteroids as 432 adjunctive therapy in the treatment of influenza. Cochrane Database Syst Rev. John 433 Wiley and Sons Ltd 3:CD010406.
- Arabi YM, Mandourah Y, Al-Hameed F, Sindi AA, Almekhlafi GA, Hussein MA, Jose J, Pinto 434 42. 435 R, Al-Omari A, Kharaba A, Almotairi A, Al Khatib K, Alraddadi B, Shalhoub S, Abdulmomen 436 A, Qushmaq I, Mady A, Mady O, Al-Aithan AM, Al-Raddadi R, Ragab A, Balkhy HH, Balkhy 437 A, Deeb AM, Al Mutairi H, Al-Dawood A, Merson L, Hayden FG, Fowler RA. 2018. Corticosteroid therapy for critically ill patients with middle east respiratory syndrome. 438
- 439 Am J Respir Crit Care Med 197:757–767.
- 440 43. Glowacka I, Bertram S, Muller MA, Allen P, Soilleux E, Pfefferle S, Steffen I, Tsegaye TS,

- 441 He Y, Gnirss K, Niemeyer D, Schneider H, Drosten C, Pohlmann S. 2011. Evidence that 442 TMPRSS2 Activates the Severe Acute Respiratory Syndrome Coronavirus Spike Protein for 443 Membrane Fusion and Reduces Viral Control by the Humoral Immune Response. J Virol 444 85:4122-4134.
- 445 44. Kawase M, Shirato K, van der Hoek L, Taguchi F, Matsuyama S. 2012. Simultaneous 446 Treatment of Human Bronchial Epithelial Cells with Serine and Cysteine Protease Inhibitors Prevents Severe Acute Respiratory Syndrome Coronavirus Entry. J Virol 447 448 86:6537-6545.
- 449 Zhou Y, Vedantham P, Lu K, Agudelo J, Carrion R, Nunneley JW, Barnard D, Pöhlmann S, McKerrow JH, Renslo AR, Simmons G. 2015. Protease inhibitors targeting coronavirus 450 and filovirus entry. Antiviral Res 116:76-84. 451

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