

# Creating a Database of Amphibian Dermal Exposure Data

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## Introduction

The purpose of this script is to combine the data sets from Van Meter et al. (2014, 2015, 2016, 2018, 2019), Glinski et al. (2018a, b, 2019, 2020), and Henson-Ramsey (2008) to create a collated database of amphibian dermal exposure data.

Manuscript	Data Set (Original Source Link)	Data Set (Repo Link)	Additional Data Sets
Van Meter et al. 2014	good_data.csv	vm2014_data.csv	
Van Meter et al. 2015	good_data.csv	vm2014_data.csv	
Van Meter et al. 2016	RDATA.csv	vm2016_data.csv	
Van Meter et al. 2018	rvm2018.csv	rvm2018.csv	
Van Meter et al. 2019	rvm2019.csv	rvm2019.csv	
Glinski et al. 2018a (dehydration)	dehydration3.csv	dag2016_data_dehydration.csv	
Glinski et al. 2018b (metabolites)	exposure_experiment.csv	dag2016_data_metabolites_4merge.csv	
Glinski et al. 2019 (biomarkers)	exposure_mixtures3.csv	dag2018_data_biomarkers.csv	dag_biomarker.csv (dag_biomarker2.csv)
Glinski et al. 2020 (dermal routes)	Water_soil.csv	dag2019_dermal_routes.csv	Dermal_routes_weights.csv (weights)
Henson-Ramsey 2008	HensonRamseyetal2008_data.pdf	hrf2008_data.csv	

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## Computational environment

This repository can be found at: [https://github.com/puruckertom/amphib\\_dermal\\_collation](https://github.com/puruckertom/amphib_dermal_collation)

If you are on a Mac and get xquartz complaints (knitr), install from: <https://www.xquartz.org/>

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## Data from Relevant Studies

### Henson-Ramsey 2008

Henson-Ramsey 2008 tested the biological impact of exposure to malathion for tiger salamanders (*Ambystoma tigrinum*). Tiger salamanders were exposed to contaminated soils with 50 ug/cm<sup>2</sup> or 100 ug/cm<sup>2</sup> malathion and through ingestion of an earthworm exposed to contaminated soils with 200 ug/cm<sup>2</sup> malathion. For each exposure, the malathion application rate was sprayed onto the approximately 1200g of soil in the 1060cm<sup>2</sup> polyethylene cages.

Tissue concentrations were assessed for five treatment groups: unexposed, exposed to 50 ug/cm<sup>2</sup> contaminated soil for 1 day, exposed to 50 ug/cm<sup>2</sup> for 2 days, exposed to 50 ug/cm<sup>2</sup> contaminated soil for 2 days and fed a contaminated worm on the first exposure day, and exposed to 100 ug/cm<sup>2</sup> contaminated soil for 2 days and fed a contaminated worm on the first exposure day.

#### Data Set Dimensions, Column Names, and Summary:

```
## [1] 9 12
```

```
## [1] "chemical"      "species"      "tissue_conc_ugg" "sample_id"
## [5] "body_weight_g" "formulation"  "soil_type"      "application"
## [9] "app_rate_g_cm2" "exp_duration" "soil_conc_ugg"  "source"
```

```
##      chemical      species      tissue_conc_ugg  sample_id
## Length:9          Length:9      Min.   :0.050      Length:9
## Class :character  Class :character  1st Qu.:0.350      Class :character
## Mode  :character  Mode  :character  Median :1.420      Mode   :character
##                                     Mean  :1.186
##                                     3rd Qu.:1.470
##                                     Max.   :3.730
## body_weight_g      formulation  soil_type      application
## Min.   :20.89      Mode:logical  Mode:logical    Length:9
## 1st Qu.:44.15      NA's:9        NA's:9          Class :character
## Median :46.26                                     Mode  :character
## Mean   :43.73
## 3rd Qu.:48.93
## Max.   :50.92
## app_rate_g_cm2     exp_duration  soil_conc_ugg      source
## Min.   :5e-05      Min.   :24      Mode:logical    Length:9
## 1st Qu.:5e-05      1st Qu.:24      NA's:9          Class :character
## Median :5e-05      Median :48                                     Mode  :character
## Mean   :5e-05      Mean   :40
## 3rd Qu.:5e-05      3rd Qu.:48
## Max.   :5e-05      Max.   :48
```

#### Van Meter et al. 2014 and Van Meter et al. 2015

Van Meter et al. 2014 performed exposures for 5 pesticide active ingredients (imidacloprid, pendimethalin, atrazine, fipronil, tridimefon) and 7 species (Southern leopard frog (*Lithobates sphenoccephala*), Fowler's toad (*Anaxyrus fowleri*), gray treefrog (*Hyla versicolor*), Northern cricket frog (*Acris crepitans*), Eastern narrowmouth toad (*Gastrophryne carolinensis*), barking treefrog (*Hyla gratiosa*), and green treefrog (*Hyla cinerea*)). Whole body tissue concentrations were measured after an 8 hour exposure period to contaminated soil. Pesticides were applied at the maximum legally allowable application rates scaled down to the area of a 10-gallon aquarium (1225 cm<sup>2</sup>).

Van Meter et al. 2015 contrasted two pesticide exposure scenarios: direct exposure through aerial overspray and indirect exposure through soil. These scenarios tested the same 5 pesticide active ingredients and two of the species (barking treefrog (*Hyla gratiosa*) and green treefrog (*Hyla cinerea*)). Pesticides were applied at the maximum legally allowable application rates scaled down to the size of a 10-gallon aquarium, with the exception of pendimethalin which was applied at 30% of the permitted application rate. This was due to pendimethalin's insolubility in the limited solvent and the water volumes used in this study.

For our purposes, the Van Meter et al. 2015 essentially adds the aerial overspray exposures to the Van Meter et al. 2014 data set.

*Note: this file does include metabolites into the total for the parents*

#### Data Set Dimensions, Column Names, and Summary:

```
## [1] 474 23
```

```

## [1] "Species"      "Sample"      "Chemical"    "Instrument"
## [5] "good"         "Application" "app_rate_g_cm2" "TissueConc"
## [9] "SoilConc"     "logKow"      "BCF"         "bodyweight"
## [13] "initialweight" "Solat20C_mgL" "Solat20C_gL"  "molmass_gmol"
## [17] "Density_gcm3" "AppFactor"    "SA_cm2"       "VapPrs_mPa"
## [21] "Koc_gmL"      "HalfLife_day" "HabFac"

## Species      Sample      Chemical      Instrument
## Length:474   Length:474   Length:474   Length:474
## Class :character Class :character Class :character Class :character
## Mode :character Mode :character Mode :character Mode :character
##
##
##
##      good      Application      app_rate_g_cm2      TissueConc
## Min.      :1      Length:474      Min.      :0e+00      Min.      : 0.007484
## 1st Qu.:1      Class :character      1st Qu.:0e+00      1st Qu.: 0.246753
## Median :1      Mode :character      Median :0e+00      Median : 0.575811
## Mean      :1                      Mean      :1e-05      Mean      : 1.908242
## 3rd Qu.:1                      3rd Qu.:2e-05      3rd Qu.: 1.743142
## Max.      :1                      Max.      :2e-05      Max.      :23.441298
##
##                      NA's      :151
##      SoilConc      logKow      BCF      bodyweight
## Min.      : 0.00625      Min.      :0.570      Min.      : 0.0018      Min.      :0.5004
## 1st Qu.: 0.20866      1st Qu.:2.500      1st Qu.: 0.0755      1st Qu.:1.3162
## Median : 3.49248      Median :3.110      Median : 0.2069      Median :1.8550
## Mean      : 7.22468      Mean      :3.142      Mean      :11.3804      Mean      :1.8658
## 3rd Qu.:10.06719      3rd Qu.:4.000      3rd Qu.: 1.0828      3rd Qu.:2.3489
## Max.      :81.71115      Max.      :5.180      Max.      :396.8461      Max.      :3.9931
##
## initialweight      Solat20C_mgL      Solat20C_gL      molmass_gmol
## Min.      :0.5004      Min.      : 0.30      Min.      :0.00030      Min.      :215.7
## 1st Qu.:1.6614      1st Qu.: 3.78      1st Qu.:0.00378      1st Qu.:215.7
## Median :2.1766      Median : 30.00      Median :0.03000      Median :291.7
## Mean      :2.2307      Mean      :123.20      Mean      :0.12320      Mean      :299.5
## 3rd Qu.:2.7601      3rd Qu.:260.00      3rd Qu.:0.26000      3rd Qu.:291.7
## Max.      :5.5480      Max.      :510.00      Max.      :0.51000      Max.      :437.1
##
## Density_gcm3      AppFactor      SA_cm2      VapPrs_mPa
## Min.      :1.170      Min.      : 850      Min.      : 0.7915      Min.      :0.00020
## 1st Qu.:1.187      1st Qu.: 47011      1st Qu.: 1.5393      1st Qu.:0.00037
## Median :1.220      Median : 143055      Median : 1.7866      Median :0.02000
## Mean      :1.288      Mean      : 291904      Mean      : 3.0232      Mean      :0.34774
## 3rd Qu.:1.480      3rd Qu.: 348598      3rd Qu.: 2.0882      3rd Qu.:0.04000
## Max.      :1.543      Max.      :4490329      Max.      :23.3326      Max.      :4.00000
##
##                      NA's      :151
##      Koc_gmL      HalfLife_day      HabFac
## Min.      : 122      Min.      : 26.00      Length:474
## 1st Qu.: 122      1st Qu.: 26.00      Class :character
## Median : 520      Median : 80.00      Mode :character
## Mean      : 20406      Mean      : 70.85
## 3rd Qu.: 825      3rd Qu.: 84.00
## Max.      :243000      Max.      :125.00
##

```

## Van Meter et al. 2016

Van Meter et al. 2016 considered bioconcentration of 5 current-use pesticides (imidacloprid, atrazine, triadimefon, fipronil, and pedimethalin) in American toads (*Bufo americanus*) across soil types. Toads were exposed to one of two soil types with significantly different organic matter content (14.1% = high organic matter, 3.1% = low organic matter). Whole body tissue concentrations were measured after an 8 hour exposure period to contaminated soil. Pesticides were applied at the maximum legally allowable application rates scaled down to the area of six 0.94 L Pyrex glass bowls each with a 15 cm diameter.

*Note: this file does include metabolites into the total for the parents*

### Data Set Dimensions, Column Names, and Summary:

```
## [1] 264 11
```

```
## [1] "Day"      "Row"      "Column"   "Pesticide" "SoilType"
## [6] "BodyBurden" "Soil"     "Weight"   "Total"     "Formulation"
## [11] "Parent"
```

```
##      Day      Row      Column      Pesticide
## Min.   :0.000   Min.   :1.000   Length:264   Length:264
## 1st Qu.:2.000   1st Qu.:2.000   Class :character   Class :character
## Median :2.000   Median :4.000   Mode  :character   Mode  :character
## Mean   :2.326   Mean   :4.023
## 3rd Qu.:3.000   3rd Qu.:6.000
## Max.   :3.000   Max.   :7.000
##      SoilType      BodyBurden      Soil      Weight
## Length:264      Min.   :-0.0378   Min.   :-0.10518   Min.   : 6.964
## Class :character 1st Qu.: 0.0486   1st Qu.: 0.02086   1st Qu.:10.524
## Mode  :character Median : 0.1099   Median : 1.49572   Median :11.740
##                  Mean  : 0.4955   Mean  : 6.02720   Mean  :12.044
##                  3rd Qu.: 0.3650   3rd Qu.: 8.64289   3rd Qu.:13.440
##                  Max.   : 6.8744   Max.   :39.57404   Max.   :23.340
##      Total      Formulation      Parent
## Min.   :0.0000   Min.   :0.0000   Min.   :0.0000
## 1st Qu.:0.0000   1st Qu.:0.0000   1st Qu.:0.0000
## Median :0.0000   Median :0.0000   Median :1.0000
## Mean   :0.3636   Mean   :0.4091   Mean   :0.5909
## 3rd Qu.:1.0000   3rd Qu.:1.0000   3rd Qu.:1.0000
## Max.   :1.0000   Max.   :1.0000   Max.   :1.0000
```

## Van Meter et al. 2018 (Multiple Pesticides Study)

Van Meter et al. 2018 evaluated risks to amphibians after exposure to a single pesticide and pesticide mixtures. The five pesticides studied were three herbicides (atrazine, metolachlor, and 2,4-D), one insecticide (malathion), and one fungicide (propiconazole). Juvenile green frogs (*Lithobates clamitans*) were exposed to contaminated soils for 8 hours and metabolic analysis of amphibian livers was conducted to measure the effects. Pesticides were applied at the maximum legally allowable application rates individually and in mixtures of two or three pesticides within an herbicide or mixed pesticide group, scaled down to the area of six 0.94 L Pyrex glass bowls each with a 15 cm diameter.

Two data sets were generated from this study, one containing data for exposure to herbicides (single and mixed) and the other containing data for exposure to mixed pesticide treatments (herbicides, insecticide, fungicide).

### Herbicide Data Set Data Set Dimensions, Column Names, and Summary:

```
## [1] 126 12
```

```
## [1] "Group"      "ATZ"      "D"         "ME"         "AppRate"   "Weight"
## [7] "SA"         "Media"    "Pesticide" "Conc"       "soil"      "sample_id"
```

```

##      Group      ATZ      D      ME
## Length:126    Min.   :-1.0000    Min.   :-1.0000    Min.   :-1.0000
## Class :character 1st Qu.: -1.0000    1st Qu.: -1.0000    1st Qu.: -1.0000
## Mode  :character Median :  1.0000    Median :  1.0000    Median :  1.0000
##              Mean  :  0.1429    Mean  :  0.1429    Mean  :  0.1429
##              3rd Qu.:  1.0000    3rd Qu.:  1.0000    3rd Qu.:  1.0000
##              Max.   :  1.0000    Max.   :  1.0000    Max.   :  1.0000
##      AppRate      Weight      SA      Media
## Min.   :14.30    Min.   :0.9634    Min.   :1.107    Length:126
## 1st Qu.:23.60    1st Qu.:1.6929    1st Qu.:1.534    Class :character
## Median :37.90    Median :2.0637    Median :1.720    Mode  :character
## Mean   :39.31    Mean   :2.0892    Mean   :1.715
## 3rd Qu.:54.50    3rd Qu.:2.4927    3rd Qu.:1.919
## Max.   :68.80    Max.   :3.6843    Max.   :2.406
## Pesticide      Conc      soil      sample_id
## Length:126      Min.   : 0.00000    Min.   : 0.0000    Length:126
## Class :character 1st Qu.: 0.00000    1st Qu.: 0.0000    Class :character
## Mode  :character Median : 0.06614    Median : 0.3629    Mode  :character
##              Mean  :  5.87233    Mean   :10.7665
##              3rd Qu.:  4.96882    3rd Qu.:24.5367
##              Max.   :72.62672    Max.   :76.0357

```

# Mixed Pesticide Data Set Data Set Dimensions, Column Names, and Summary:

```
## [1] 72 11
```

```

## [1] "Group"      "ATZ"      "MA"      "PROP"      "Pesticide" "Media"
## [7] "Conc"      "Weight"    "SA"      "soil"      "sample_id"

```

```

##      Group      ATZ      MA      PROP
## Length:72    Min.   :-1.0000    Min.   :-1.0000    Min.   :-1.0000
## Class :character 1st Qu.: -1.0000    1st Qu.: -1.0000    1st Qu.: -1.0000
## Mode  :character Median :  1.0000    Median :  1.0000    Median :  1.0000
##              Mean  :  0.3333    Mean  :  0.3333    Mean  :  0.3333
##              3rd Qu.:  1.0000    3rd Qu.:  1.0000    3rd Qu.:  1.0000
##              Max.   :  1.0000    Max.   :  1.0000    Max.   :  1.0000
## Pesticide      Media      Conc      Weight
## Length:72      Length:72      Min.   : 0.00054    Min.   :1.188
## Class :character Class :character 1st Qu.: 0.32504    1st Qu.:1.786
## Mode  :character Mode  :character Median : 1.33986    Median :2.014
##              Mean  :  6.38788    Mean  :2.203
##              3rd Qu.:  6.65664    3rd Qu.:2.455
##              Max.   :71.52122    Max.   :4.014
##      SA      soil      sample_id
## Min.   :1.447    Min.   : 0.7354    Length:72
## 1st Qu.:1.833    1st Qu.: 2.2633    Class :character
## Median :1.965    Median :10.5771    Mode  :character
## Mean   :2.047    Mean   : 9.3745
## 3rd Qu.:2.203    3rd Qu.:13.9342
## Max.   :2.929    Max.   :25.7013

```

The herbicide and mixed pesticide data sets were cleaned prior and joined into a merged data set (referred to as Van Meter et al. 2018 Multiple Pesticides Study in subsequent steps). The single and mixed-pesticide treatments that were retained in the merged data set include atrazine, propiconazole, 2,4-D, malathion, and metolachlor. Original columns from the herbicide and mixed pesticide data sets were altered for standardization. These standardized columns will be used in future data cleaning steps in order to merge all data sets.

## Merged Data Set Data Set Dimensions, Column Names, and Summary:

```
## [1] 137 12

## [1] "app_rate_g_cm2" "body_weight_g" "chemical" "tissue_conc_ugg"
## [5] "sample_id" "source" "application" "exp_duration"
## [9] "formulation" "soil_conc_ugg" "soil_type" "species"

## app_rate_g_cm2 body_weight_g chemical tissue_conc_ugg
## Min. :2.600e-06 Min. :0.9634 Length:137 Min. : 0.00054
## 1st Qu.:1.430e-05 1st Qu.:1.7623 Class :character 1st Qu.: 0.27576
## Median :2.360e-05 Median :2.0136 Mode :character Median : 1.41009
## Mean :2.004e-05 Mean :2.1086 Mean : 7.36154
## 3rd Qu.:2.590e-05 3rd Qu.:2.3395 3rd Qu.: 9.95084
## Max. :3.090e-05 Max. :4.0141 Max. :72.62672
## sample_id source application exp_duration
## Length:137 Length:137 Length:137 Min. :8
## Class :character Class :character Class :character 1st Qu.:8
## Mode :character Mode :character Mode :character Median :8
## Mean :8
## 3rd Qu.:8
## Max. :8
## formulation soil_conc_ugg soil_type species
## Min. :0 Min. : 0.1639 Mode:logical Length:137
## 1st Qu.:0 1st Qu.: 1.7763 NA's:137 Class :character
## Median :0 Median :12.5989 Mode :character
## Mean :0 Mean :14.1337
## 3rd Qu.:0 3rd Qu.:23.3872
## Max. :0 Max. :76.0357
```

## Van Meter et al. 2019

Van Meter et al. 2019 looked at joint exposure of two common herbicides (atrazine, alachlor) and a fertilizer (urea) while quantifying juvenile corticosterone stress levels, acetylcholinesterase (AChE) activity, and pesticide bioaccumulation in Southern leopard frogs. Single agrochemical or tank mixtures were applied to terrestrial microcosms, and then individual juveniles were added to microcosms for an 8-h exposure.

## Data Set Dimensions, Column Names, and Summary:

```
## [1] 96 12

## [1] "app_rate_g_cm2" "application" "body_weight_g" "chemical"
## [5] "exp_duration" "formulation" "sample_id" "soil_conc_ugg"
## [9] "soil_type" "source" "species" "tissue_conc_ugg"

## app_rate_g_cm2 application body_weight_g chemical
## Min. :2.36e-05 Length:96 Min. : 1.398 Length:96
## 1st Qu.:2.36e-05 Class :character 1st Qu.: 2.084 Class :character
## Median :2.92e-05 Mode :character Median : 2.550 Mode :character
## Mean :2.92e-05 Mean : 2.986
## 3rd Qu.:3.48e-05 3rd Qu.: 3.126
## Max. :3.48e-05 Max. :20.235
## exp_duration formulation sample_id soil_conc_ugg
## Min. :8 Min. :0 Length:96 Mode:logical
## 1st Qu.:8 1st Qu.:0 Class :character NA's:96
## Median :8 Median :0 Mode :character
## Mean :8 Mean :0
```

```
## 3rd Qu.:8      3rd Qu.:0
## Max. :8      Max. :0
## soil_type      source      species      tissue_conc_ugg
## Length:96      Length:96      Length:96      Min. :0.08123
## Class :character Class :character Class :character 1st Qu.:0.40279
## Mode :character Mode :character Mode :character Median :0.94646
## Mean :1.28035
## 3rd Qu.:1.43585
## Max. :6.61248
```

## Van Meter et al. 2021

Van Meter et al. 2021

### Data Set Dimensions, Column Names, and Summary:

```
## [1] 96 12
```

```
## [1] "app_rate_g_cm2" "application" "body_weight_g" "chemical"
## [5] "exp_duration" "formulation" "sample_id" "soil_conc_ugg"
## [9] "soil_type" "source" "species" "tissue_conc_ugg"
```

```
## app_rate_g_cm2      application      body_weight_g      chemical
## Min. :2.36e-05      Length:48      Min. :2.091      Length:48
## 1st Qu.:2.36e-05      Class :character 1st Qu.:3.313      Class :character
## Median :2.92e-05      Mode :character Median :3.791      Mode :character
## Mean :2.92e-05      Mean :3.795
## 3rd Qu.:3.48e-05      3rd Qu.:4.342
## Max. :3.48e-05      Max. :5.589
## exp_duration      formulation      sample_id      soil_conc_ugg
## Min. :8      Min. :0      Length:48      Min. : 3.705
## 1st Qu.:8      1st Qu.:0      Class :character 1st Qu.: 7.427
## Median :8      Median :0      Mode :character Median :11.115
## Mean :8      Mean :0      Mean :11.525
## 3rd Qu.:8      3rd Qu.:0      3rd Qu.:12.994
## Max. :8      Max. :0      Max. :25.873
## soil_type      source      species      tissue_conc_ugg
## Length:48      Length:48      Length:48      Min. : 0.2155
## Class :character Class :character Class :character 1st Qu.: 1.1327
## Mode :character Mode :character Mode :character Median : 1.9263
## Mean : 3.2363
## 3rd Qu.: 3.9569
## Max. :13.9376
```

## Glinski et al. 2018a (Dehydration)

Glinski et al. 2018a studied how amphibian hydration status influences uptake of pesticides through dermal exposure. Amphibians (Southern leopard frogs (*Lithobates sphenoccephala*) and Fowler's toads (*Anaxyrus fowleri*)) were dehydrated for periods of 0, 2, 4, 6, 8, or 10 hours prior to exposure to pesticide-contaminated soils. Pesticides studied included atrazine, triadimefon, metolachlor, chlorothalonil, and imidacloprid. Soil and whole-body homogenates were measured after an 8 hour exposure period. Pesticides were applied at the maximum legally allowable application rates scaled down to the area of six 0.94 L Pyrex glass bowls each with a 15 cm diameter.

*Note: this file does not combine daughters with parents*

*Note: this file has body burdens and soil concentrations as separate rows*

### Data Set Dimensions, column Names, and Summary:

```
## [1] 1494      8
```

```
## [1] "time"      "parent"  "analyte" "matrix"  "species" "conc"    "ID"
## [8] "weight"
```

	time	parent	analyte	matrix
## Min.	: 0	Length:1494	Length:1494	Length:1494
## 1st Qu.:	2	Class :character	Class :character	Class :character
## Median :	5	Mode :character	Mode :character	Mode :character
## Mean :	5			
## 3rd Qu.:	8			
## Max. :	10			

	species	conc	ID	weight
## Length:1494	Min. :	0.00000	Length:1494	Min. :0.6821
## Class :character	1st Qu.:	0.02215	Class :character	1st Qu.:1.6108
## Mode :character	Median :	0.08482	Mode :character	Median :3.0890
##	Mean :	6.17646		Mean :3.0810
##	3rd Qu.:	2.60007		3rd Qu.:4.3124
##	Max. :	238.15019		Max. :7.2481

### Glinski et al. 2018b (Metabolites)

Glinski et al. 2018b assessed the potential metabolic activation of pesticides (atrazine, triadimefon, fopronil) in amphibians. This data set (1) contains *in vitro* and *in vivo* metabolic rate constants derived from toad (*Anaxyrus terrestris*) livers during experiments measuring the depletion of pesticides and the formation of their metabolites. Pesticides were applied at the maximum legally allowable application rates scaled down to the area of a 10-gallon aquarium (1225 cm<sup>2</sup>).

### Metabolites Data Set (1) Data Set Dimensions, Column Names, and Summary:

```
## [1] 352 6
```

	time	parent	analyte	matrix
## Min.	: 0.00	Length:352	Length:352	Length:352
## 1st Qu.:	2.00	Class :character	Class :character	Class :character
## Median :	12.00	Mode :character	Mode :character	Mode :character
## Mean :	16.41			
## 3rd Qu.:	24.00			
## Max. :	48.00			

	conc	replicate
## Min.	:-0.01244	Min. :1.00
## 1st Qu.:	0.01292	1st Qu.:1.75
## Median :	0.08373	Median :2.50
## Mean :	2.12963	Mean :2.50
## 3rd Qu.:	0.97824	3rd Qu.:3.25
## Max. :	32.47385	Max. :4.00

The *in vitro* derived constants were assessed for their precitability by exposing Fowler's toads (*Anaxyrus fowleri*) to contaminated soils at maximum application rate for 2, 4, 12, and 48 hours. This data set (merged) contains the data from the Fowler's toad experiment along with the tissue concentrations from data set 1; this data set (merged) is used in subsequent steps.

### Metabolites Data Set (merged) Data Set Dimensions, Column Names, and Summary:

```
## [1] 60 12
```



```
## [1] "exp_duration"      "chemical"           "tissue_conc_ugg" "sample_id"
## [5] "soil_type"          "app_rate_g_cm2"    "soil_conc_ugg"   "body_weight_g"
## [9] "formulation"        "species"            "application"      "source"

## exp_duration  chemical      tissue_conc_ugg  sample_id
## Min.      : 2      Length:60      Min.      :0.08328  Length:60
## 1st Qu.: 4      Class :character 1st Qu.:0.33733  Class :character
## Median :12      Mode  :character Median :0.86010  Mode  :character
## Mean      :18
## 3rd Qu.:24
## Max.      :48
## Max.      :7.62649
## soil_type      app_rate_g_cm2      soil_conc_ugg      body_weight_g
## Mode:logical   Min.      :1.100e-06  Min.      : 0.2858  Min.      :0.1879
## NA's:60        1st Qu.:1.100e-06  1st Qu.: 0.9041  1st Qu.:0.5925
##                Median :2.700e-06  Median : 4.8333  Median :0.7144
##                Mean      :9.237e-06  Mean      : 9.1511  Mean      :0.7350
##                3rd Qu.:2.290e-05  3rd Qu.:18.0554  3rd Qu.:0.8782
##                Max.      :2.290e-05  Max.      :32.4738  Max.      :1.4909
## formulation    species      application      source
## Min.      :0      Length:60      Length:60      Length:60
## 1st Qu.:0      Class :character  Class :character  Class :character
## Median :0      Mode  :character  Mode  :character  Mode  :character
## Mean      :0
## 3rd Qu.:0
## Max.      :0
```

## Glinski et al. 2019 (Biomarkers)

Glinski et al. 2019 exposed Southern leopard frogs (*Lithobates sphenoccephala*) to either the maximum or 1/10th maximum pesticide application rate to single, double, or triple pesticide mixtures of bifenthrin, metolachlor, and triadimefon to consider the typical co-application of pesticides during agricultural growing seasons. Tissue concentrations and metabolomic profiling of amphibian livers were studied after an 8 hour exposure period to pesticide-contaminated soil. Pesticides application rates were scaled down to the area of eight 0.94 L Pyrex glass bowls each with a 15 cm diameter.

## Data Set Dimensions, Column Names, and Summary:

```
## [1] 192 9

## [1] "group"      "met"          "tdt"          "bif"          "frog.weight"
## [6] "sample_id"  "pesticide"    "rate"         "conc"

## group      met      tdt      bif
## Length:192  Min.     :-1.0000  Min.     :-1.0000  Min.     :-1.0000
## Class :character 1st Qu.: -1.0000 1st Qu.: -1.0000 1st Qu.: -1.0000
## Mode :character Median : 1.0000 Median : 1.0000 Median : 1.0000
##                Mean      : 0.3333 Mean      : 0.3333 Mean      : 0.3333
##                3rd Qu.: 1.0000 3rd Qu.: 1.0000 3rd Qu.: 1.0000
##                Max.      : 1.0000 Max.      : 1.0000 Max.      : 1.0000
## frog.weight sample_id      pesticide      rate
## Min.      :1.012  Length:192  Length:192  Length:192
## 1st Qu.:2.745  Class :character  Class :character  Class :character
## Median :3.142  Mode  :character  Mode  :character  Mode  :character
## Mean      :3.299
## 3rd Qu.:3.789
## Max.      :6.739
## conc
## Min.      : 0.001061
## 1st Qu.: 0.069055
```

```
## Median : 0.212920
## Mean   : 0.801643
## 3rd Qu.: 0.521471
## Max.   :19.879783
```

**Glinski et al. 2020 (Dermal Routes)**

Glinski et al. 2020 assessed dermal uptake in amphibians from exposure to three pesticides (bifenthrin, chlorpyrifos, trifloxystrobin). Pesiticide body burdens and hepatic metabolome for Leopard frogs were measured for two routes of uptake: uptake from contaminated soils versus uptake from contaminated surface water. Pesticides were applied at 1 ppm, scaled down to the area of eight 0.94 L Pyrex glass bowls each with a 15 cm diameter.

**Data Set Dimensions, Column Names, and Summary:**

```
## [1] 192    5

## [1] "Sample.ID"      "Analyte"        "Media"          "Matrix"
## [5] "Concentration"

##   Sample.ID      Analyte      Media      Matrix
## Length:192      Length:192      Length:192      Length:192
## Class :character Class :character Class :character Class :character
## Mode  :character Mode  :character Mode  :character Mode  :character
##
##
##
## Concentration
## Min.      :0.00000
## 1st Qu.:0.01036
## Median :0.15326
## Mean  :0.33962
## 3rd Qu.:0.44162
## Max.    :3.40759
```

---

**Application Rates**

The table below concisely displays the pesticide applications rates (ug/cm<sup>2</sup>) used in each relevant study as well as the variables used to compute the application rates.

pesticide	app_rate_ug_cm2	applied_mL	container	area_cm2	total_area_cm2	density_g_cm3	pesticide_
<b>Van Meter et al. 2014/2015</b>							
atrazine	22.9000	75 MeOH	10-gal aquarium	1225	1225	1.1900	28.1
fipronil	1.1000	75 MeOH	10-gal aquarium	1225	1225	1.5515	1.3
imidacloprid	5.7000	75 MeOH	10-gal aquarium	1225	1225	1.6000	7.0
pendimethalin	19.8000	75 MeOH	10-gal aquarium	1225	1225	1.1700	24.3
triadimefon	2.7000	75 MeOH	10-gal aquarium	1225	1225	1.2200	3.3
<b>Van Meter et al. 2016</b>							
atrazine	22.9000	75 MeOH	.94 L bowl	225*6	1350	1.1900	30.9
fipronil	1.1000	75 MeOH	.94 L bowl	225*6	1350	1.5515	1.5
imidacloprid	5.7000	75 MeOH	.94 L bowl	225*6	1350	1.6000	7.7
pendimethalin	69.8000	75 MeOH	.94 L bowl	225*6	1350	1.1700	94.2
triadimefon	2.7000	75 MeOH	.94 L bowl	225*6	1350	1.2200	3.6
<b>Van Meter et al. 2018</b>							
atrazine	23.6000	50 MeOH	.94 L bowl	225*6	1350	1.1900	31.9
2,4-D	14.3000	50 MeOH	.94 L bowl	225*6	1350	1.5000	19.3
metolachlor	30.9000	50 MeOH	.94 L bowl	225*6	1350	1.1000	41.7
malathion	25.9000	50 MeOH	.94 L bowl	225*6	1350	1.2300	35.0
propiconazole	2.6000	50 MeOH	.94 L bowl	225*6	1350	1.3000	3.5
<b>Van Meter et al. 2019</b>							
alachlor	34.8000	75 MeOH	.94 L bowl	225*6	1350	1.1300	47.0
atrazine	23.6000	75 MeOH	.94 L bowl	225*6	1350	1.1900	31.9
<b>Van Meter et al. 2021</b>							
alachlor	34.8000	75 MeOH	.94 L bowl	225*6	1350	1.1300	47.0
atrazine	23.6000	75 MeOH	.94 L bowl	225*10	1354	1.1900	31.9
<b>Henson-Ramsey et al. 2008</b>							
malathion	50.0000	NA	cage	1060	NA	1.2300	NA
<b>Glinski et al. 2018a</b>							
atrazine	23.9500	75 MeOH	.94 L bowl	225*6	1350	1.1900	32.3
chlorothalonil	44.3000	75 MeOH	.94 L bowl	225*6	1350	1.8000	59.8
imidacloprid	5.3900	75 MeOH	.94 L bowl	225*6	1350	1.6000	7.3
metolachlor	31.0100	75 MeOH	.94 L bowl	225*6	1350	1.1000	41.9
triadimefon	2.9100	75 MeOH	.94 L bowl	225*6	1350	1.2200	3.9
<b>Glinski et al. 2018b</b>							
atrazine	22.9000	75 MeOH	10-gal aquarium	1225	1225	1.1900	28.1
fipronil	1.1000	75 MeOH	10-gal aquarium	1225	1225	1.5515	1.3
triadimefon	2.7000	75 MeOH	10-gal aquarium	1225	1225	1.2200	3.3
<b>Glinski et al. 2019</b>							
bifenthrin (max)	3.4500	75 MeOH	.94 L bowl	225*8	1800	1.3000	6.2
metolachlor (max)	30.6200	75 MeOH	.94 L bowl	225*8	1800	1.1000	55.1
triadimefon (max)	2.8700	75 MeOH	.94 L bowl	225*8	1800	1.2200	5.2
bifenthrin (1/10 max)	0.3450	75 MeOH	.94 L bowl	225*8	1800	1.3000	0.6
metolachlor (1/10 max)	3.0620	75 MeOH	.94 L bowl	225*8	1800	1.1000	5.5
triadimefon (1/10 max)	0.2870	75 MeOH	.94 L bowl	225*8	1800	1.2200	0.5
<b>Glinski et al. 2020</b>							
bifenthrin	0.2889	400ml of 1ppm pesticide in water	.94 L bowl	225*8	1800	1.3000	50ml/bowl
chlorpyrifos	0.3111	400ml of 1ppm pesticide in water	.94 L bowl	225*8	1800	1.4000	50ml/bowl
trifloxystrobin	0.3022	400ml of 1ppm pesticide in water	.94 L bowl	225*8	1800	1.3600	50ml/bowl

---

## Cleaning and Merging the Data Sets

Each data set was cleaned for merging. This consisted of dropping unneeded columns and standardizing column names of retained columns. Four columns were added to all data sets (soil type, formulation, exposure duration, and research study source).

Once each data set was cleaned, a local copy was saved and the data set was merged with the previously cleaned data sets.

The process of cleaning and merging each data set is briefly described below.

---

### Van Meter et al. 2014/2015

Metabolites and parents that do not include metabolites were dropped from the data set. This includes atrazine, deisopropyl atrazine, desethyl atrazine, fipronil, fipronil-sulfone, triadimefon, triadimenol.

```
# drop metabolites and parents that do not include metabolites
vm2015_chem_drop <- c("Atrazine", "Deisopropyl Atrazine", "Desethyl Atrazine", "Fipronil", "Fipronil-Sulfone", "Triadimefon", "Triadimenol")
chem_vector_drop <- which(vm2015$Chemical %in% vm2015_chem_drop)
vm2015_subset1 <- vm2015[-chem_vector_drop, ]
vm2015_subset2 <- droplevels(vm2015_subset1)
```

There were 278 observations with these chemicals. After dropping the 278 observations from the initial 474, the updated dimensions are:

```
## [1] 196 23
```

There were 15 unneeded columns dropped and 4 added for standarization.

```
# drop unneeded columns for merging
all_cols <- colnames(vm2015_subset2)
drop_cols <- c("Instrument", "good", "logKow", "BCF", "initialweight", "Solat20C_mgL", "Solat20C_gL", "molmass_gmol",
              "AppFactor", "SA_cm2", "VapPrs_mPa", "Koc_gmL", "HalfLife_day", "HabFac")
vm2015_subset3 <- vm2015_subset2[, !(names(vm2015_subset2) %in% drop_cols)]
colnames(vm2015_subset3)
```

```
## [1] "Species"      "Sample"      "Chemical"    "Application"
## [5] "app_rate_g_cm2" "TissueConc"  "SoilConc"    "bodyweight"
```

```
# add columns
soil_type <- c(rep("PLE", nrow(vm2015_subset3)))
formulation <- (rep(0, nrow(vm2015_subset3)))
exp_duration <- (rep(8, nrow(vm2015_subset3)))
source <- c(rep("rvm2015", nrow(vm2015_subset3)))
vm2015_subset4 <- cbind(vm2015_subset3, formulation, soil_type, exp_duration, source)
# standardize column names
colnames(vm2015_subset4)
```

```
## [1] "Species"      "Sample"      "Chemical"    "Application"
## [5] "app_rate_g_cm2" "TissueConc"  "SoilConc"    "bodyweight"
## [9] "formulation"   "soil_type"   "exp_duration" "source"
```

```

colnames(vm2015_subset4)[which(colnames(vm2015_subset4) == "Sample")] <- "sample_id"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4) == "Species")] <- "species"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4) == "Chemical")] <- "chemical"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4) == "Application")] <- "application"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4) == "TissueConc")] <- "tissue_conc_ugg"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4) == "SoilConc")] <- "soil_conc_ugg"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4) == "bodyweight")] <- "body_weight_g"
colnames(vm2015_subset4)

```

```

## [1] "species"      "sample_id"    "chemical"     "application"
## [5] "app_rate_g_cm2" "tissue_conc_ugg" "soil_conc_ugg" "body_weight_g"
## [9] "formulation"   "soil_type"    "exp_duration" "source"

```

```

# reorder vm2015 alphabetically
vm2015_merge <- vm2015_subset4[, order(names(vm2015_subset4))]

```

```

# write a local copy
vm2015_merge_filename <- paste(amphibdir_data_out, "vm2015_merge.csv", sep = "")
write.csv(vm2015_merge, file = vm2015_merge_filename)

```

The data set's dimensions are:

```
## [1] 196 12
```

## Van Meter et al. 2016

From the initial 11 columns, 4 columns were dropped and consolidated into 1, and 4 columns were added.

```

# add sample_id
vm2016$sample_id <- paste(vm2016$Day, vm2016$Row, vm2016$Column, sep = "_")
vm2016_subset2 <- subset(vm2016, select = c(-Day, -Row, -Column, -Total))
# add additional columns
species <- c(rep("American toad", nrow(vm2016_subset2)))
application <- c(rep("Indirect", nrow(vm2016_subset2)))
exp_duration <- (rep(8, nrow(vm2016_subset2)))
source <- c(rep("rvm2016", nrow(vm2016_subset2)))
vm2016_subset3 <- cbind(vm2016_subset2, species, application, exp_duration, source)

```

Application rates for several pesticides were inserted. There were 108 observations with decay products that were not sprayed; these observations were dropped so as to only include the parents in the cleaned data set. There were 60 observations with atrazine, fipronil, or triadimefon that were dropped because they do not include metabolites in total.

```

# assign values to application rate unique(vm2016_subset3$Pesticide)
vm2016_subset3$app_rate_g_cm2[vm2016_subset3$Pesticide == "ATZTOT"] <- 2.29e-05
vm2016_subset3$app_rate_g_cm2[vm2016_subset3$Pesticide == "Imid"] <- 5.7e-06
vm2016_subset3$app_rate_g_cm2[vm2016_subset3$Pesticide == "FipTOT"] <- 1.1e-06
vm2016_subset3$app_rate_g_cm2[vm2016_subset3$Pesticide == "TNDTOT"] <- 2.7e-06
vm2016_subset3$app_rate_g_cm2[vm2016_subset3$Pesticide == "Pendi"] <- 6.98e-05
# drop decay products that were not sprayed, keeping only parents
rows_to_drop <- which(vm2016_subset3$Parent == 0)
vm2016_subset4 <- vm2016_subset3[-rows_to_drop, ]
# drop ATZ, Fip, TDN since do not include metabolites in total
chems_to_drop <- c("ATZ", "Fip", "TDN")
vm2016_subset5 <- vm2016_subset4[!(vm2016_subset4$Pesticide %in% chems_to_drop), ]
# drop parent field
drop_cols <- c("Parent")
vm2016_subset6 <- vm2016_subset5[, !(names(vm2016_subset5) %in% drop_cols)]

```

Several column names were standardized and all columns were ordered for ease of merging with the combined data set.

```
# standardize column names
colnames(vm2016_subset6)

## [1] "Pesticide"      "SoilType"      "BodyBurden"    "Soil"
## [5] "Weight"        "Formulation"   "sample_id"     "species"
## [9] "application"    "exp_duration"  "source"        "app_rate_g_cm2"

colnames(vm2016_subset6)[which(colnames(vm2016_subset6) == "Pesticide")] <- "chemical"
colnames(vm2016_subset6)[which(colnames(vm2016_subset6) == "SoilType")] <- "soil_type"
colnames(vm2016_subset6)[which(colnames(vm2016_subset6) == "BodyBurden")] <- "tissue_conc_ugg"
colnames(vm2016_subset6)[which(colnames(vm2016_subset6) == "Soil")] <- "soil_conc_ugg"
colnames(vm2016_subset6)[which(colnames(vm2016_subset6) == "Weight")] <- "body_weight_g"
colnames(vm2016_subset6)[which(colnames(vm2016_subset6) == "Formulation")] <- "formulation"

# alter chemical name
vm2016_subset6$chemical <- as.character(vm2016_subset6$chemical)
vm2016_subset6$chemical[vm2016_subset6$chemical == "Imid"] <- "imidacloprid"

# reorder columns alphabetically to help with merge
colnames(vm2016_subset6)

## [1] "chemical"      "soil_type"     "tissue_conc_ugg" "soil_conc_ugg"
## [5] "body_weight_g" "formulation"   "sample_id"     "species"
## [9] "application"   "exp_duration"  "source"        "app_rate_g_cm2"

vm2016_merge <- vm2016_subset6[, order(names(vm2016_subset6))]
colnames(vm2016_merge)

## [1] "app_rate_g_cm2" "application"    "body_weight_g"  "chemical"
## [5] "exp_duration"   "formulation"    "sample_id"     "soil_conc_ugg"
## [9] "soil_type"      "source"         "species"        "tissue_conc_ugg"

# write a local copy
vm2016_merge_filename <- paste(amphibdir_data_out, "vm2016_merge.csv", sep = "")
write.csv(vm2016_merge, file = vm2016_merge_filename)
```

The updated dimensions are:

```
## [1] 96 12
```

The Van Meter et al. 2014/2015 and Van Meter et al. 2016 data sets were combined.

**The combined data set's updated dimensions are:**

```
## [1] 292 12
```

### Glinski et al. 2018a (Dehydration)

The metabolite products were dropped from the data set; 600 rows from the initial 1494 rows were retained.

```
# drop metabolite products
parent_keepers <- which(as.vector(dag2016_dehy0$parent) == as.vector(dag2016_dehy0$analyte))
dag2016_dehy1 <- dag2016_dehy0[parent_keepers, ]
```

Several column names were altered for standarization across the data set, and 7 columns were added for standarization.

```
## time is length of dehydration colnames(dag2016_dehy1)[which(colnames(dag2016_dehy1)=='time')]<-'exp_duration'

# standardize column names
colnames(dag2016_dehy1)[which(colnames(dag2016_dehy1) == "analyte")] <- "chemical"
colnames(dag2016_dehy1)[which(colnames(dag2016_dehy1) == "conc")] <- "tissue_conc_ugg"
colnames(dag2016_dehy1)[which(colnames(dag2016_dehy1) == "ID")] <- "sample_id"
colnames(dag2016_dehy1)[which(colnames(dag2016_dehy1) == "weight")] <- "body_weight_g"

# add additional columns
exp_duration <- c(rep(8, nrow(dag2016_dehy1)))
soil_type <- c(rep("PLE", nrow(dag2016_dehy1)))
application <- c(rep("Indirect", nrow(dag2016_dehy1)))
formulation <- (rep(0, nrow(dag2016_dehy1)))
app_rate_g_cm2 <- (rep(0, nrow(dag2016_dehy1)))
soil_conc_ugg <- (rep(0, nrow(dag2016_dehy1)))
source <- c(rep("dag_dehydration", nrow(dag2016_dehy1)))
dag2016_dehy2 <- cbind(dag2016_dehy1, formulation, soil_type, application, app_rate_g_cm2, exp_duration, soil_conc_ugg)
```

The updated dimensions are:

```
## [1] 600 15
```

Multiple soil concentration observations were given the same ID. Until a many-to-one merge of soil concentrations could be executed, 300 rows were temporarily dropped. There were also 3 columns dropped.

```
# drop the soil until we can do a many-to-one merge of soil concentrations drop decay products that were not sprayed
# only parents
rows_to_drop <- which(dag2016_dehy2$matrix == "soil")
dag2016_dehy3 <- dag2016_dehy2[-rows_to_drop, ]
# parent, time and matrix columns delete
drop_cols <- c("parent", "time", "matrix")
dag2016_dehy4 <- dag2016_dehy3[, !(names(dag2016_dehy3) %in% drop_cols)]
```

The updated dimensions are:

```
## [1] 300 12
```

The application rate values were inserted, the temporarily dropped soil concentrations were updated to the current data set, and the species names were standardized.

```
# fill in application rates unique(dag2016_dehy4$chemical)
update_atrazine <- which(dag2016_dehy4$chemical == "atrazine")
dag2016_dehy4$app_rate_g_cm2[update_atrazine] <- 2.395e-05 # atrazine g/cm2
update_chloro <- which(dag2016_dehy4$chemical == "chloro+d")
dag2016_dehy4$app_rate_g_cm2[update_chloro] <- 4.43e-05 # chloro g/cm2
update_metol <- which(dag2016_dehy4$chemical == "metol")
dag2016_dehy4$app_rate_g_cm2[update_metol] <- 3.101e-05 # metol g/cm2
update_tdn <- which(dag2016_dehy4$chemical == "tdn")
dag2016_dehy4$app_rate_g_cm2[update_tdn] <- 2.91e-06 # tdn g/cm2
update_imid <- which(dag2016_dehy4$chemical == "imid")
dag2016_dehy4$app_rate_g_cm2[update_imid] <- 5.39e-06 # imid g/cm2

# add back in soil concentrations (in already-made soil_conc_ugg column)
dag2016_soil <- dag2016_dehy2[rows_to_drop, ]
dag2016_dehy4$soil_conc_ugg <- dag2016_soil$tissue_conc_ugg
```

```
# rename species names, according to standardized names
dag2016_dehy4$species <- as.character(dag2016_dehy4$species)
dag2016_dehy4$species[dag2016_dehy4$species == "LF"] <- "Leopard frog"
dag2016_dehy4$species[dag2016_dehy4$species == "BA"] <- "Fowlers toad"
dag2016_dehy4$species <- as.factor(dag2016_dehy4$species)
```

The dimensions are:

```
## [1] 300 12
```

The Glinkski et al. 2018a (Dehydration) was combined with the previously merged data sets.

The combined data set's updated dimensions are:

```
## [1] 592 12
```

### Henson-Ramsey 2008

The Henson-Ramsey 2008 data set did not require any additional data cleaning. It was combined with the previously merged data sets.

The combined data set's updated dimensions are:

```
## [1] 601 12
```

### Glinski et al. 2018b (Metabolites)

Apart from standardizing the species name, the Glinski et al. 2018b (Metabolites) data set did not require any additional data cleaning. It was combined with the previously merged data sets.

```
# rename species names, according to standardized names
dag2016_metabolite_merge$species <- as.character(dag2016_metabolite_merge$species)
dag2016_metabolite_merge$species[dag2016_metabolite_merge$species == "Anaxyrus fowleri"] <- "Fowlers toad"
dag2016_metabolite_merge$species <- as.factor(dag2016_metabolite_merge$species)
```

The combined data set's updated dimensions are:

```
## [1] 661 12
```

### Glinski et al. 2019 (Biomarkers)

Five columns were dropped from the original biomarkers data set and the names of two columns were standardized.

```
# drop columns
drop_cols <- c("met", "tdt", "bif", "rate", "group")
dag_biomarker_subset <- dag_biomarker[, !(names(dag_biomarker) %in% drop_cols)]

# standardize column names
colnames(dag_biomarker_subset)[which(colnames(dag_biomarker_subset) == "conc")] <- "tissue_conc_ugg"
colnames(dag_biomarker_subset)[which(colnames(dag_biomarker_subset) == "frog.weight")] <- "body_weight_g"
```

The updated column names and dimensions are:

```
## [1] "body_weight_g" "sample_id" "pesticide" "tissue_conc_ugg"
```



```
## [1] 192 4
```

The application rates and soil concentrations were not included in the original biomarkers data set. Both are included in the following data set:

#### Data Set Dimensions, Column Names, and Summary:

```
## [1] 136 15
```

```
## [1] "frog.weight" "SAMPLE"      "Met"          "TDN"          "TDL"
## [6] "BIF"         "soil.weight" "Met.soil"     "TDN.soil"     "TDL.soil"
## [11] "BIF.soil"    "Rate"        "app.rate.met" "app.rate.tdn" "app.rate.bif"
```

```
## frog.weight      SAMPLE              Met              TDN
## Min.   :1.012    Length:136        Min.   : 0.0000    Min.   :0.00000
## 1st Qu.:2.749    Class :character   1st Qu.: 0.0000    1st Qu.:0.00000
## Median :3.164    Mode  :character   Median : 0.0000    Median :0.00000
## Mean   :3.302                                Mean  : 0.9123    Mean   :0.06927
## 3rd Qu.:3.762                                3rd Qu.: 0.4298    3rd Qu.:0.07447
## Max.   :6.784                                Max.   :19.8798    Max.   :0.55921
## TDL              BIF              soil.weight      Met.soil
## Min.   :0.00000    Min.   :0.0000    Min.   : 4.476    Min.   :0.000
## 1st Qu.:0.00000    1st Qu.:0.0000    1st Qu.: 6.731    1st Qu.:0.000
## Median :0.00000    Median :0.0000    Median : 7.772    Median :0.000
## Mean   :0.02259    Mean   :0.1276    Mean   : 8.043    Mean   :1.605
## 3rd Qu.:0.01770    3rd Qu.:0.1299    3rd Qu.: 9.050    3rd Qu.:2.265
## Max.   :0.30815    Max.   :1.0271    Max.   :13.571    Max.   :6.758
## TDN.soil          TDL.soil          BIF.soil          Rate
## Min.   :0.0000    Min.   :0.000000    Min.   :0.0000    Length:136
## 1st Qu.:0.0000    1st Qu.:0.000000    1st Qu.:0.0000    Class :character
## Median :0.0000    Median :0.000000    Median :0.0000    Mode  :character
## Mean   :0.7168    Mean   :0.010160    Mean   :0.7417
## 3rd Qu.:0.5312    3rd Qu.:0.007463    3rd Qu.:1.1472
## Max.   :3.6300    Max.   :0.061563    Max.   :5.2658
## app.rate.met      app.rate.tdn      app.rate.bif
## Min.   : 0.000    Min.   :0.0000    Min.   :0.0000
## 1st Qu.: 0.000    1st Qu.:0.0000    1st Qu.:0.0000
## Median : 0.000    Median :0.0000    Median :0.0000
## Mean   :14.263    Mean   :1.3389    Mean   :1.6070
## 3rd Qu.: 5.511    3rd Qu.:0.5173    3rd Qu.:0.6209
## Max.   :55.106    Max.   :5.1730    Max.   :6.2090
```

The application rates were converted from mg to g/cm<sup>2</sup>.

```
dag_biomarker2_update <- replace.value(dag_biomarker2, "app.rate.met", from = 55.106, to = 3.062e-05, verbose = TRUE)
dag_biomarker2_update <- replace.value(dag_biomarker2_update, "app.rate.met", from = 5.5106, to = 3.062e-06, verbose = TRUE)
dag_biomarker2_update <- replace.value(dag_biomarker2_update, "app.rate.tdn", from = 5.173, to = 2.87e-06, verbose = TRUE)
dag_biomarker2_update <- replace.value(dag_biomarker2_update, "app.rate.tdn", from = 0.5173, to = 2.87e-07, verbose = TRUE)
dag_biomarker2_update <- replace.value(dag_biomarker2_update, "app.rate.bif", from = 6.209, to = 3.45e-06, verbose = TRUE)
dag_biomarker2_update <- replace.value(dag_biomarker2_update, "app.rate.bif", from = 0.6209, to = 3.45e-07, verbose = TRUE)
```

A one-to-one merge was conducted based on the unique sample id for each measured pesticide (either bifenthrin, metolachlor, or triadimefon) to join the original biomarkers data set and the data set containing the application rates and soil concentrations. Vectors containing the application rates and soil concentrations were joined to the original data set.

```

# bif extraction
dag_biomarker_subset_bif <- dag_biomarker_subset[dag_biomarker_subset$pesticide == "bif", ]
dag_biomarker2_subset_bif <- dag_biomarker2_update[dag_biomarker2_update$BIF != 0, ]

dag_biomarker_bif_merge <- merge(x = dag_biomarker_subset_bif, y = dag_biomarker2_subset_bif, by.x = "sample_id",
  all.x = TRUE)

# met extraction
dag_biomarker_subset_met <- dag_biomarker_subset[dag_biomarker_subset$pesticide == "met", ]
dag_biomarker2_subset_met <- dag_biomarker2_update[dag_biomarker2_update$Met != 0, ]

dag_biomarker_met_merge <- merge(x = dag_biomarker_subset_met, y = dag_biomarker2_subset_met, by.x = "sample_id",
  all.x = TRUE)

# tdt extraction
dag_biomarker_subset_tdt <- dag_biomarker_subset[dag_biomarker_subset$pesticide == "tdt", ]
dag_biomarker2_subset_tdt <- dag_biomarker2_update[dag_biomarker2_update$TDN != 0, ]

dag_biomarker_tdt_merge <- merge(x = dag_biomarker_subset_tdt, y = dag_biomarker2_subset_tdt, by.x = "sample_id",
  all.x = TRUE)

# combine bif, met, and tdt
app_bind_bmt <- c(dag_biomarker_bif_merge[, "app.rate.bif"], dag_biomarker_met_merge[, "app.rate.met"], dag_biomarker_tdt_merge[, "app.rate.tdt"])
soil_bind_bmt <- c(dag_biomarker_bif_merge[, "BIF.soil"], dag_biomarker_met_merge[, "Met.soil"], dag_biomarker_tdt_merge[, "TDN.soil"])

# join app and soil vectors to data set
dag_biomarker_subset2 <- dag_biomarker_subset[order(dag_biomarker_subset[, 3]), ]
rownames(dag_biomarker_subset2) <- seq(length = nrow(dag_biomarker_subset2))

dag_biomarker_subset3 <- cbind(dag_biomarker_subset2, app_bind_bmt, soil_bind_bmt)

# standardize column names
colnames(dag_biomarker_subset3)[which(colnames(dag_biomarker_subset3) == "app_bind_bmt")] <- "app_rate_g_cm2"
colnames(dag_biomarker_subset3)[which(colnames(dag_biomarker_subset3) == "soil_bind_bmt")] <- "soil_conc_ugg"

```

The updated column names and dimensions are:

```

## [1] "body_weight_g"    "sample_id"        "pesticide"        "tissue_conc_ugg"
## [5] "app_rate_g_cm2"   "soil_conc_ugg"

## [1] 192    6

```

New columns were created for standarization, the columns were ordered alphabetically, and a local copy was stored.

```

# create new columns
application <- c(rep("soil", nrow(dag_biomarker_subset3)))
exp_duration <- c(rep(8, nrow(dag_biomarker_subset3)))
formulation <- c(rep(0, nrow(dag_biomarker_subset3)))
soil_type <- c(rep(NA, nrow(dag_biomarker_subset3)))
source <- c(rep("dag_biomarker", nrow(dag_biomarker_subset3)))
species <- c(rep("Leopard frog", nrow(dag_biomarker_subset3)))

# combine columns
dag_biomarker_subset4 <- cbind(dag_biomarker_subset3, application, exp_duration, formulation, soil_type, source, species)

```

```

# standardize pesticide column
dag_biomarker_subset4$pesticide <- as.character(dag_biomarker_subset4$pesticide)
dag_biomarker_subset4$pesticide[dag_biomarker_subset4$pesticide == "bif"] <- "Bifenthrin"
dag_biomarker_subset4$pesticide[dag_biomarker_subset4$pesticide == "met"] <- "Metolachlor"
dag_biomarker_subset4$pesticide[dag_biomarker_subset4$pesticide == "tdt"] <- "Triadimefon"

colnames(dag_biomarker_subset4)[which(colnames(dag_biomarker_subset4) == "pesticide")] <- "chemical"

# unite function for sample id and chemical
dag_biomarker_subset5 <- unite(data = dag_biomarker_subset4, col = "sample_id", "sample_id", "chemical", sep = " ")

# order columns in abc for merge
dag_biomarker_merge <- dag_biomarker_subset5[, order(names(dag_biomarker_subset5))]

```

The updated column names and dimensions are:

```

## [1] "app_rate_g_cm2" "application" "body_weight_g" "chemical"
## [5] "exp_duration" "formulation" "sample_id" "soil_conc_ugg"
## [9] "soil_type" "source" "species" "tissue_conc_ugg"

## [1] 192 12

```

The Glinski et al. 2019 (Biomarkers) was combined with the previously merged data sets.

The combined data set's updated dimensions are:

```
## [1] 853 12
```

#### Van Meter et al. 2018 (Multiple Pesticides Study)

The Van Meter et al. 2018 (Multiple Pesticides Study) data set did not require any additional data cleaning. It was combined with the previously merged data sets.

The combined data set's updated dimensions are:

```
## [1] 990 12
```

#### Glinski et al. 2020 (Dermal Routes)

The dermal routes data set did not include the body weights for the measured amphibians. These weights were included in a separate data set:

Data Set Dimensions, Column Names, and Summary:

```
## [1] 48 2
```

```
## [1] "Weight_g" "Sample"
```

```

##      Weight_g      Sample
## Min.   :0.9555 Length:48
## 1st Qu.:1.4204 Class :character
## Median :1.7817 Mode  :character
## Mean   :1.7784
## 3rd Qu.:2.1319
## Max.   :2.8197

```

A one-to-many merge was employed to merge the dermal routes data set and the weights data set based on the Sample ID. Only rows where the Matrix is “Amphibian” have a body weight; all other rows are NA.

```
# merge (one-to-many) dermal routes data with weights data, based on Sample ID
dermal_routes_subset2 <- dermal_routes[order(dermal_routes$Sample.ID), ]
weights_2 <- weights[order(weights$Sample), ]

dermal_routes_subset3 <- merge(dermal_routes_subset2, weights_2, by.x = "Sample.ID", by.y = "Sample", all.x = TRUE)
```

The updated dimensions are:

```
## [1] 192 6
```

The soil concentrations, where the Media and Matrix are both “Soil,” was subset from the data set to be used later in the data cleaning process. These soil concentrations (currently listed in the “Concentration” column) will be used for the soil\_conc\_ugg column in the cleaned data set.

```
# subset soil to be used later for soil concentration column (will use 'Concentration' column)
soil_subset <- dermal_routes_subset2[dermal_routes_subset2$Media == "Soil", ]
soil_subset2 <- soil_subset[soil_subset$Matrix == "Soil", ]
```

The dimensions of this soil subset are:

```
## [1] 48 5
```

Referring back to the main dermal routes data set: we are only interested in the pesticide exposures on amphibians while in soil. These rows were subset.

```
# want Media == soil because interested in dermal exposure in soil
dermal_routes_subset4 <- dermal_routes_subset3[dermal_routes_subset3$Media == "Soil", ]
# sum(dermal_routes_subset3$Media == 'Soil') # == 96 dim(dermal_routes_subset4) # == 96 x 6

# want Matrix == Amphibian because interested in amphib exposure
dermal_routes_subset5 <- dermal_routes_subset4[dermal_routes_subset4$Matrix == "Amphibian", ]
# sum(dermal_routes_subset4$Matrix == 'Amphibian') # == 48 dim(dermal_routes_subset5) # == 48 x 6
```

The updated dimensions are:

```
## [1] 48 6
```

The soil concentrations were appended to the main dermal routes data set.

```
# add in soil concentration column, previously subset order by Sample.ID, then by Analyte name to match up rows for
# sets
dermal_routes_subset6 <- dermal_routes_subset5[order(dermal_routes_subset5[, 1], dermal_routes_subset5[, 2]), ]
soil_subset3 <- soil_subset2[order(soil_subset2[, 1], soil_subset2[, 2]), ]

# dim(dermal_routes_subset6) # == 48 x 6 dim(soil_subset3) # == 48 x 5

dermal_routes_subset7 <- cbind(dermal_routes_subset6, soil_subset3$Concentration)
```

The updated dimensions are:

```
## [1] 48 7
```

The metabolites were dropped from the data set. Additionally, several new columns were created for standardization, existing columns were standardized according to the naming conventions of the collated data set, and unneeded columns were dropped. Columns were ordered alphabetically for ease of merging.

```
# drop metabolites
rows_to_drop <- c("4-OH", "CPO", "TFSa")
dermal_routes_subset8 <- dermal_routes_subset7[!(dermal_routes_subset7$Analyte %in% rows_to_drop), ]

# create new columns
app_rate_g_cm2 <- c(rep(NA, nrow(dermal_routes_subset8)))
application <- c(rep("soil", nrow(dermal_routes_subset8)))
exp_duration <- c(rep(8, nrow(dermal_routes_subset8)))
formulation <- c(rep(0, nrow(dermal_routes_subset8)))
soil_type <- c(rep("OLS", nrow(dermal_routes_subset8)))
source <- c(rep("dag_dermal_routes", nrow(dermal_routes_subset8)))
species <- c(rep("Leopard frog", nrow(dermal_routes_subset8)))

# alter existing column names
colnames(dermal_routes_subset8)
```

```
## [1] "Sample.ID"          "Analyte"
## [3] "Media"              "Matrix"
## [5] "Concentration"      "Weight_g"
## [7] "soil_subset3$Concentration"
```

```
colnames(dermal_routes_subset8)[which(colnames(dermal_routes_subset8) == "Analyte")] <- "chemical"
colnames(dermal_routes_subset8)[which(colnames(dermal_routes_subset8) == "Sample.ID")] <- "sample_id"
colnames(dermal_routes_subset8)[which(colnames(dermal_routes_subset8) == "Concentration")] <- "tissue_conc_ugg"
colnames(dermal_routes_subset8)[which(colnames(dermal_routes_subset8) == "soil_subset3$Concentration")] <- "soil_conc_ugg"
colnames(dermal_routes_subset8)[which(colnames(dermal_routes_subset8) == "Weight_g")] <- "body_weight_g"
```

```
# combine columns
dermal_routes_subset9 <- cbind(dermal_routes_subset8, app_rate_g_cm2, application, exp_duration, formulation, soil_conc_ugg, body_weight_g)
names(dermal_routes_subset9)
```

```
## [1] "sample_id"          "chemical"           "Media"              "Matrix"
## [5] "tissue_conc_ugg"    "body_weight_g"      "soil_conc_ugg"      "app_rate_g_cm2"
## [9] "application"        "exp_duration"       "formulation"         "soil_type"
## [13] "source"             "species"
```

```
# drop columns
cols_to_drop <- c("Matrix", "Media")
dermal_routes_subset10 <- dermal_routes_subset9[, !(names(dermal_routes_subset9) %in% cols_to_drop)]

# insert application rates
dermal_routes_subset10$chemical <- as.character(dermal_routes_subset10$chemical)
unique(dermal_routes_subset10$chemical)
```

```
## [1] "BIF" "CPF" "TFS"
```

```
dermal_routes_subset10$app_rate_g_cm2[dermal_routes_subset10$chemical == "BIF"] <- 2.8889e-07 #bifenthrin g/cm2
dermal_routes_subset10$app_rate_g_cm2[dermal_routes_subset10$chemical == "CPF"] <- 3.1111e-07 #chlorpyrifos g/cm2
dermal_routes_subset10$app_rate_g_cm2[dermal_routes_subset10$chemical == "TFS"] <- 3.0222e-07 #trifloxystrobin g/cm2

summary(dermal_routes_subset10$app_rate_g_cm2)
```

```
##      Min.   1st Qu.   Median     Mean   3rd Qu.    Max.
## 2.889e-07 2.889e-07 3.022e-07 3.007e-07 3.111e-07 3.111e-07
```

```
# order columns in abc for merge
dermal_routes_merge <- dermal_routes_subset10[, order(names(dermal_routes_subset10))]
```

The updated column names and dimensions are:

```
## [1] 24 12

## [1] "app_rate_g_cm2" "application" "body_weight_g" "chemical"
## [5] "exp_duration" "formulation" "sample_id" "soil_conc_ugg"
## [9] "soil_type" "source" "species" "tissue_conc_ugg"
```

A local copy was saved, and the data set was combined with the collated data set.

The combined data set's updated dimensions are:

```
## [1] 1014 12
```

## Final Product

Minor alterations were made to the final collated data set to standardize names of the application types and chemicals.

```
amphib_dermal_collated <- combined_data6
```

```
colnames(amphib_dermal_collated)
```

```
## [1] "app_rate_g_cm2" "application" "body_weight_g" "chemical"
## [5] "exp_duration" "formulation" "sample_id" "soil_conc_ugg"
## [9] "soil_type" "source" "species" "tissue_conc_ugg"
```

```
# check to see if everything ok
```

```
summary(amphib_dermal_collated$app_rate_g_cm2)
```

```
##      Min.   1st Qu.   Median     Mean   3rd Qu.    Max.
## 2.870e-07 2.790e-06 5.700e-06 1.616e-05 2.395e-05 6.980e-05
```

```
summary(amphib_dermal_collated$body_weight_g)
```

```
##      Min. 1st Qu.  Median     Mean 3rd Qu.    Max.
## 0.1879  1.3043  2.1247  3.3800  3.0412 50.9200
```

```
summary(amphib_dermal_collated$exp_duration)
```

```
##      Min. 1st Qu.  Median     Mean 3rd Qu.    Max.
## 2.000  8.000  8.000  8.876  8.000 48.000
```

```
summary(amphib_dermal_collated$soil_conc_ugg) # 206 NAs
```

```
##      Min. 1st Qu.  Median     Mean 3rd Qu.    Max.    NA's
## 0.1125  2.0326  5.5791 13.9733 16.7043 238.1502      9
```

```
summary(amphib_dermal_collated$tissue_conc_ugg)
```

```
##      Min.   1st Qu.    Median      Mean   3rd Qu.      Max.
## 0.00054 0.16908 0.52573 2.51415 2.06812 72.62672
```

```
# standardize application levels
amphib_dermal_collated$application <- tolower(amphib_dermal_collated$application)
amphib_dermal_collated$application <- as.factor(amphib_dermal_collated$application)

# standardize chemical levels
amphib_dermal_collated$chemical <- as.character(amphib_dermal_collated$chemical)

amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "fip"] <- "fipronil"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "BIF"] <- "bifenthrin"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "MET"] <- "metolachlor"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "MAT"] <- "malathion"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "ATZT"] <- "atrazine"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "PROPT"] <- "propiconazole"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "metol"] <- "metolachlor"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "tdn"] <- "triadimefon"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "imid"] <- "imidacloprid"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "chloro+d"] <- "chlorothalonil"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "CPF"] <- "chlorpyrifos"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "TFS"] <- "trifloxystrobin"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "FipTOT"] <- "fipronil"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "ATZTOT"] <- "atrazine"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "TNDTOT"] <- "triadimefon"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "Pendi"] <- "pendimethalin"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "Total Atrazine"] <- "atrazine"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "Total Fipronil"] <- "fipronil"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "Total Triadimefon"] <- "triadimefon"

amphib_dermal_collated$chemical <- tolower(amphib_dermal_collated$chemical)
amphib_dermal_collated$chemical <- as.factor(amphib_dermal_collated$chemical)

# because we combined the

# write out file
amphib_dermal_collated_filename <- paste(amphibdir_data_out, "amphib_dermal_collated.csv", sep = "")
write.csv(amphib_dermal_collated, file = amphib_dermal_collated_filename)
```

## Column Names

```
## [1] "app_rate_g_cm2" "application" "body_weight_g" "chemical"
## [5] "exp_duration" "formulation" "sample_id" "soil_conc_ugg"
## [9] "soil_type" "source" "species" "tissue_conc_ugg"
```

## Dimensions

```
## [1] 1014 12
```

## Variable Summaries

```
## app_rate_g_cm2 application body_weight_g chemical
```

```
## Min. :2.870e-07 indirect :396 Min. : 0.1879 triadimefon :223
## 1st Qu.:2.790e-06 overspray: 45 1st Qu.: 1.3043 atrazine :191
## Median :5.700e-06 soil :573 Median : 2.1247 metolachlor :154
## Mean :1.616e-05 Mean : 3.3800 imidacloprid: 78
## 3rd Qu.:2.395e-05 3rd Qu.: 3.0412 bifenthrin : 72
## Max. :6.980e-05 Max. :50.9200 fipronil : 71
## (Other) :225
## exp_duration formulation sample_id soil_conc_ugg
## Min. : 2.000 Min. :0.00000 Length:1014 Min. : 0.1125
## 1st Qu.: 8.000 1st Qu.:0.00000 Class :character 1st Qu.: 2.0326
## Median : 8.000 Median :0.00000 Mode :character Median : 5.5791
## Mean : 8.876 Mean :0.03582 Mean : 13.9733
## 3rd Qu.: 8.000 3rd Qu.:0.00000 3rd Qu.: 16.7043
## Max. :48.000 Max. :1.00000 Max. :238.1502
## NA's :9 NA's :9
## soil_type source species tissue_conc_ugg
## Length:1014 Length:1014 Length:1014 Min. : 0.00054
## Class :character Class :character Class :character 1st Qu.: 0.16908
## Mode :character Mode :character Mode :character Median : 0.52573
## Mean : 2.51415
## 3rd Qu.: 2.06812
## Max. :72.62672
##
```

---

## Session Information

```
## R version 4.0.2 (2020-06-22)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 10 x64 (build 18363)
##
## Matrix products: default
##
## locale:
## [1] LC_COLLATE=English_United States.1252
## [2] LC_CTYPE=English_United States.1252
## [3] LC_MONETARY=English_United States.1252
## [4] LC_NUMERIC=C
## [5] LC_TIME=English_United States.1252
##
## attached base packages:
## [1] stats graphics grDevices utils datasets methods base
##
## other attached packages:
## [1] tinytex_0.32 formatR_1.11 anchors_3.0-8 MASS_7.3-51.6
## [5] rgenoud_5.8-3.0 stringr_1.4.0 tidyr_1.1.2 dplyr_1.0.5
## [9] knitr_1.31 kableExtra_1.3.4 reshape2_1.4.4 gridExtra_2.3
## [13] ggplot2_3.3.3
##
## loaded via a namespace (and not attached):
## [1] Rcpp_1.0.6 pillar_1.6.0 compiler_4.0.2 plyr_1.8.6
## [5] tools_4.0.2 digest_0.6.27 viridisLite_0.3.0 evaluate_0.14
## [9] lifecycle_1.0.0 tibble_3.1.1 gtable_0.3.0 pkgconfig_2.0.3
## [13] rlang_0.4.10 rstudioapi_0.13 DBI_1.1.1 yaml_2.2.1
## [17] xfun_0.24 xml2_1.3.2 http_1.4.2 withr_2.4.1
## [21] systemfonts_1.0.2 generics_0.1.0 vctrs_0.3.7 webshot_0.5.2
## [25] grid_4.0.2 tidyselct_1.1.0 svglite_2.0.0 glue_1.4.1
```



```
## [29] R6_2.5.0          fansi_0.4.2      rmarkdown_2.6    purrr_0.3.4
## [33] magrittr_2.0.1    scales_1.1.1     ellipsis_0.3.1   htmltools_0.5.1.1
## [37] rvest_0.3.6       assertthat_0.2.1 colorspace_1.4-1 utf8_1.2.1
## [41] stringi_1.5.3     munsell_0.5.0    crayon_1.4.1
```