

Creating a Database of Amphibian Dermal Exposure Data

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Introduction

The purpose of this script is to combine the data sets from Van Meter et al. (2014, 2015, 2016, 2018), Glinski et al. (2018a, b, 2019, 2020), and Henson-Ramsey (2008) to create a collated database of amphibian dermal exposure data.

Manuscript	Data Set (Original Source Link)	Data Set (Repo Link)	Additional Data Sets
Van Meter et al. 2014	good_data.csv	vm2014_data.csv	
Van Meter et al. 2015	good_data.csv	vm2014_data.csv	
Van Meter et al. 2016	RDATA.csv	vm2016_data.csv	
Van Meter et al. 2018	vm2017_merge.csv	vm2017_merge.csv	
Glinski et al. 2018a (dehydration)	dehydration3.csv	dag2016_data_dehydration.csv	
Glinski et al. 2018b (metabolites)	exposure_experiment.csv	dag2016_data_metabolites_4merge.csv	
Glinski et al. 2019 (biomarkers)	exposure_mixtures3.csv	dag2018_data_biomarkers_biomarker.csv	(dag_biomarker2.csv)
Glinski et al. 2020 (dermal routes)	Water_soil.csv	dag2019_dermal_routes.csv	Dermal_routes_weights.csv (weights)
Henson-Ramsey 2008	HensonRamseyetal2008_data.csv	h2008_data.csv	

Computational environment

This repository can be found at: https://github.com/puruckertom/amphib_dermal_collation

If you are on a Mac and get xquartz complaints (knitr), install from: <https://www.xquartz.org/>

Data from Relevant Studies

Van Meter et al. 2014 and Van Meter et al. 2015

Van Meter et al. 2014 performed exposures for 5 pesticide active ingredients (imidacloprid, pendimethalin, atrazine, fipronil, tridimefon) and 7 species (Southern leopard frog (*Lithobates sphenoccephala*), Fowler's toad (*Anaxyrus fowleri*), gray treefrog (*Hyla versicolor*), Northern cricket frog (*Acris crepitans*), Eastern narrowmouth toad (*Gastrophryne carolinensis*), barking treefrog (*Hyla gratiosa*) and green treefrog (*Hyla cinerea*)). Whole body tissue concentrations were measured after an 8 hour exposure period to contaminated soil. Pesticides were applied at the maximum legally allowable application rates scaled down to the area of a 10-gallon aquarium (1225 cm²).

Van Meter et al. 2015 contrasted two pesticide exposure scenarios: direct exposure through aerial overspray and indirect exposure through soil. These scenarios tested the same 5 pesticide active ingredients and two of the species (barking treefrog (*Hyla gratiosa*) and green treefrog (*Hyla cinerea*)). Pesticides were applied at the maximum legally allowable application rates scaled down to the size of a 10-gallon aquarium, with the exception of pendimethalin which was applied at 30% of the permitted application rate. This was due to pendimethalin's insolubility in the limited solvent and the water volumes used in this study.

For our purposes, the Van Meter et al. 2015 essentially adds the aerial overspray exposures to the Van Meter et al. 2014 data set.

Note: this file does include metabolites into the total for the parents

Data Set Dimensions, Column Names, and Summary:

```
## [1] 474 23
```

```
## [1] "Species"      "Sample"        "Chemical"      "Instrument"
## [5] "good"         "Application"   "app_rate_g_cm2" "TissueConc"
## [9] "SoilConc"     "logKow"        "BCF"           "bodyweight"
## [13] "initialweight" "Solat20C_mgL"  "Solat20C_gL"   "molmass_gmol"
## [17] "Density_gcm3" "AppFactor"     "SA_cm2"        "VapPrs_mPa"
## [21] "Koc_gmL"      "HalfLife_day"  "HabFac"
```

```
## Species      Sample      Chemical      Instrument
## Length:474    Length:474    Length:474    Length:474
## Class :character Class :character Class :character Class :character
## Mode :character Mode :character Mode :character Mode :character
```

```
##
```

```
##
```

```
##
```

```
##
```

```
##      good      Application      app_rate_g_cm2      TissueConc
## Min.      :1      Length:474      Min.      :0e+00      Min.      : 0.007484
## 1st Qu.:1      Class :character      1st Qu.:0e+00      1st Qu.: 0.246753
## Median :1      Mode :character      Median :0e+00      Median : 0.575811
## Mean      :1                                     Mean      :1e-05      Mean      : 1.908242
## 3rd Qu.:1                                     3rd Qu.:2e-05      3rd Qu.: 1.743142
## Max.      :1                                     Max.      :2e-05      Max.      :23.441298
##                                     NA's      :151
```

```
##      SoilConc      logKow      BCF      bodyweight
## Min.      : 0.00625      Min.      :0.570      Min.      : 0.0018      Min.      :0.5004
## 1st Qu.: 0.20866      1st Qu.:2.500      1st Qu.: 0.0755      1st Qu.:1.3162
## Median : 3.49248      Median :3.110      Median : 0.2069      Median :1.8550
```

```

## Mean : 7.22468 Mean :3.142 Mean : 11.3804 Mean :1.8658
## 3rd Qu.:10.06719 3rd Qu.:4.000 3rd Qu.: 1.0828 3rd Qu.:2.3489
## Max. :81.71115 Max. :5.180 Max. :396.8461 Max. :3.9931
##
## initialweight Solat20C_mgL Solat20C_gL molmass_gmol
## Min. :0.5004 Min. : 0.30 Min. :0.00030 Min. :215.7
## 1st Qu.:1.6614 1st Qu.: 3.78 1st Qu.:0.00378 1st Qu.:215.7
## Median :2.1766 Median : 30.00 Median :0.03000 Median :291.7
## Mean :2.2307 Mean :123.20 Mean :0.12320 Mean :299.5
## 3rd Qu.:2.7601 3rd Qu.:260.00 3rd Qu.:0.26000 3rd Qu.:291.7
## Max. :5.5480 Max. :510.00 Max. :0.51000 Max. :437.1
##
## Density_gcm3 AppFactor SA_cm2 VapPrs_mPa
## Min. :1.170 Min. : 850 Min. : 0.7915 Min. :0.00020
## 1st Qu.:1.187 1st Qu.: 47011 1st Qu.: 1.5393 1st Qu.:0.00037
## Median :1.220 Median : 143055 Median : 1.7866 Median :0.02000
## Mean :1.288 Mean : 291904 Mean : 3.0232 Mean :0.34774
## 3rd Qu.:1.480 3rd Qu.: 348598 3rd Qu.: 2.0882 3rd Qu.:0.04000
## Max. :1.543 Max. :4490329 Max. :23.3326 Max. :4.00000
## NA's :151
## Koc_gmL HalfLife_day HabFac
## Min. : 122 Min. : 26.00 Length:474
## 1st Qu.: 122 1st Qu.: 26.00 Class :character
## Median : 520 Median : 80.00 Mode :character
## Mean : 20406 Mean : 70.85
## 3rd Qu.: 825 3rd Qu.: 84.00
## Max. :243000 Max. :125.00
##

```

Van Meter et al. 2016

Van Meter et al. 2016 considered bioconcentration of 5 current-use pesticides (imidacloprid, atrazine, triadimefon, fipronil, and pedimethalin) in American toads (*Bufo americanus*) across soil types. Toads were exposed to one of two soil types with significantly different organic matter content (14.1% = high organic matter, 3.1% = low organic matter). Whole body tissue concentrations were measured after an 8 hour exposure period to contaminated soil. Pesticides were applied at the maximum legally allowable application rates scaled down to the area of six 0.94 L Pyrex glass bowls each with a 15 cm diameter.

Note: this file does include metabolites into the total for the parents

Data Set Dimensions, Column Names, and Summary:

```

## [1] 264 11

## [1] "Day" "Row" "Column" "Pesticide" "SoilType"
## [6] "BodyBurden" "Soil" "Weight" "Total" "Formulation"
## [11] "Parent"

## Day Row Column Pesticide
## Min. :0.000 Min. :1.000 Length:264 Length:264
## 1st Qu.:2.000 1st Qu.:2.000 Class :character Class :character
## Median :2.000 Median :4.000 Mode :character Mode :character
## Mean :2.326 Mean :4.023
## 3rd Qu.:3.000 3rd Qu.:6.000

```

```
## Max. :3.000 Max. :7.000
## SoilType BodyBurden Soil Weight
## Length:264 Min. :-0.0378 Min. :-0.10518 Min. : 6.964
## Class :character 1st Qu.: 0.0486 1st Qu.: 0.02086 1st Qu.:10.524
## Mode :character Median : 0.1099 Median : 1.49572 Median :11.740
## Mean : 0.4955 Mean : 6.02720 Mean :12.044
## 3rd Qu.: 0.3650 3rd Qu.: 8.64289 3rd Qu.:13.440
## Max. : 6.8744 Max. :39.57404 Max. :23.340
## Total Formulation Parent
## Min. :0.0000 Min. :0.0000 Min. :0.0000
## 1st Qu.:0.0000 1st Qu.:0.0000 1st Qu.:0.0000
## Median :0.0000 Median :0.0000 Median :1.0000
## Mean :0.3636 Mean :0.4091 Mean :0.5909
## 3rd Qu.:1.0000 3rd Qu.:1.0000 3rd Qu.:1.0000
## Max. :1.0000 Max. :1.0000 Max. :1.0000
```

Glinski et al. 2018a (Dehydration)

Glinski et al. 2018a studied how amphibian hydration status influences uptake of pesticides through dermal exposure. Amphibians (Southern leopard frogs (*Lithobates sphenoccephala*) and Fowler's toads (*Anaxyrus fowleri*)) were dehydrated for periods of 0, 2, 4, 6, 8, or 10 hours prior to exposure to pesticide-contaminated soils. Pesticides studied included atrazine, triadimefon, metolachlor, chlorothalonil, and imidacloprid. Soil and whole-body homogenates were measured after an 8 hour exposure period. Pesticides were applied at the maximum legally allowable application rates scaled down to the area of six 0.94 L Pyrex glass bowls each with a 15 cm diameter.

Note: this file does not combine daughters with parents

Note: this file has body burdens and soil concentrations as separate rows

Data Set Dimensions, column Names, and Summary:

```
## [1] 1494 8
```

```
## [1] "time" "parent" "analyte" "matrix" "species" "conc" "ID"
## [8] "weight"
```

```
## time parent analyte matrix
## Min. : 0 Length:1494 Length:1494 Length:1494
## 1st Qu.: 2 Class :character Class :character Class :character
## Median : 5 Mode :character Mode :character Mode :character
## Mean : 5
## 3rd Qu.: 8
## Max. :10
## species conc ID weight
## Length:1494 Min. : 0.00000 Length:1494 Min. :0.6821
## Class :character 1st Qu.: 0.02215 Class :character 1st Qu.:1.6108
## Mode :character Median : 0.08482 Mode :character Median :3.0890
## Mean : 6.17646 Mean :3.0810
## 3rd Qu.: 2.60007 3rd Qu.:4.3124
## Max. :238.15019 Max. :7.2481
```

Henson-Ramsey 2008

Henson-Ramsey 2008 tested the biological impact of exposure to malathion for tiger salamanders (*Ambystoma tigrinum*). Tiger salamanders were exposed to contaminated soils with 50 ug/cm² or 100 ug/cm² malathion and through ingestion of an earthworm exposed to contaminated soils with 200 ug/cm² malathion. For each exposure, the malathion application rate was sprayed onto the approximately 1200g of soil in the 1060cm² polyethylene cages. Tissue concentrations were assessed for five treatment groups: unexposed, exposed to 50 ug/cm² contaminated soil for 1 day, exposed to 50 ug/cm² for 2 days, exposed to 50 ug/cm² contaminated soil for 2 days and fed a contaminated worm on the first exposure day, and exposed to 100 ug/cm² contaminated soil for 2 days and fed a contaminated worm on the first exposure day.

Data Set Dimensions, Column Names, and Summary:

```
## [1] 9 12

## [1] "chemical"      "species"      "tissue_conc_ugg" "sample_id"
## [5] "body_weight_g" "formulation"  "soil_type"      "application"
## [9] "app_rate_g_cm2" "exp_duration" "soil_conc_ugg"  "source"

##      chemical      species      tissue_conc_ugg sample_id
## Length:9          Length:9      Min.   :0.050   Length:9
## Class :character  Class :character  1st Qu.:0.350   Class :character
## Mode  :character  Mode  :character  Median :1.420   Mode  :character
##                                     Mean  :1.186
##                                     3rd Qu.:1.470
##                                     Max.   :3.730
## body_weight_g  formulation  soil_type  application
## Min.   :20.89  Mode:logical  Mode:logical  Length:9
## 1st Qu.:44.15  NA's:9         NA's:9         Class :character
## Median :46.26                                     Mode  :character
## Mean   :43.73
## 3rd Qu.:48.93
## Max.   :50.92
## app_rate_g_cm2  exp_duration  soil_conc_ugg  source
## Min.   :5e-05  Min.   :24     Mode:logical  Length:9
## 1st Qu.:5e-05  1st Qu.:24     NA's:9         Class :character
## Median :5e-05  Median :48                                     Mode  :character
## Mean   :5e-05  Mean   :40
## 3rd Qu.:5e-05  3rd Qu.:48
## Max.   :5e-05  Max.   :48
```

Glinski et al. 2018b (Metabolites)

Glinski et al. 2018b assessed the potential metabolic activation of pesticides (atrazine, triadimefon, fopronil) in amphibians. This data set (1) contains *in vitro* and *in vivo* metabolic rate constants derived from toad (*Anaxyrus terrestris*) livers during experiments measuring the depletion of pesticides and the formation of their metabolites. Pesticides were applied at the maximum legally allowable application rates scaled down to the area of a 10-gallon aquarium (1225 cm²).

Metabolites Data Set (1) Data Set Dimensions, Column Names, and Summary:

```
## [1] 352 6
```

```
## [1] "time"      "parent"      "analyte"      "matrix"      "conc"      "replicate"

##      time      parent      analyte      matrix
## Min.   : 0.00   Length:352   Length:352   Length:352
## 1st Qu.: 2.00   Class :character Class :character Class :character
## Median :12.00   Mode  :character Mode  :character Mode  :character
## Mean    :16.41
## 3rd Qu.:24.00
## Max.    :48.00
##      conc      replicate
## Min.   :-0.01244 Min.    :1.00
## 1st Qu.: 0.01292 1st Qu.:1.75
## Median : 0.08373 Median :2.50
## Mean    : 2.12963 Mean    :2.50
## 3rd Qu.: 0.97824 3rd Qu.:3.25
## Max.    :32.47385 Max.    :4.00
```

The *in vitro* derived constants were assessed for their precitability by exposing Fowler's toads (*Anaxyrus fowleri*) to contaminated soils at maximum application rate for 2, 4, 12, and 48 hours. This data set (merged) contains the data from the Fowler's toad experiment along with the tissue concentrations from data set 1; this data set (merged) is used in subsequent steps.

Metabolites Data Set (merged) Data Set Dimensions, Column Names, and Summary:

```
## [1] 60 12
```

```
## [1] "exp_duration" "chemical"      "tissue_conc_ugg" "sample_id"
## [5] "soil_type"    "app_rate_g_cm2" "soil_conc_ugg"  "body_weight_g"
## [9] "formulation"  "species"        "application"    "source"

## exp_duration chemical      tissue_conc_ugg sample_id
## Min.   : 2   Length:60      Min.    :0.08328 Length:60
## 1st Qu.: 4   Class :character 1st Qu.:0.33733 Class :character
## Median :12   Mode  :character Median :0.86010 Mode  :character
## Mean    :18
## 3rd Qu.:24
## Max.    :48
## Max.    :7.62649
## soil_type app_rate_g_cm2 soil_conc_ugg body_weight_g
## Mode:logical Min.    :1.100e-06 Mode:logical Min.    :0.1879
## NA's:60      1st Qu.:1.100e-06 NA's:60      1st Qu.:0.5925
##              Median :2.700e-06              Median :0.7144
##              Mean    :9.237e-06              Mean    :0.7350
##              3rd Qu.:2.290e-05              3rd Qu.:0.8782
##              Max.    :2.290e-05              Max.    :1.4909
## formulation species      application source
## Min.   :0   Length:60      Length:60      Length:60
## 1st Qu.:0   Class :character Class :character Class :character
## Median :0   Mode  :character Mode  :character Mode  :character
## Mean    :0
## 3rd Qu.:0
## Max.    :0
```

Glinski et al. 2019 (Biomarkers)

Glinski et al. 2019 exposed Southern leopard frogs (*Lithobates sphenoccephala*) to either the maximum or 1/10th maximum pesticide application rate to single, double, or triple pesticide mixtures of bifenthrin, metolachlor, and triadimefon to consider the typical co-application of pesticides during agricultural growing seasons. Tissue concentrations and metabolomic profiling of amphibian livers were studied after an 8 hour exposure period to pesticide-contaminated soil. Pesticides application rates were scaled down to the area of eight 0.94 L Pyrex glass bowls each with a 15 cm diameter.

Data Set Dimensions, Column Names, and Summary:

```
## [1] 192 9
```

```
## [1] "group"      "met"        "tdt"        "bif"        "frog.weight"
## [6] "sample_id"  "pesticide"  "rate"       "conc"       "
```

```
##      group          met          tdt          bif
## Length:192      Min.   :-1.0000      Min.   :-1.0000      Min.   :-1.0000
## Class :character 1st Qu.: -1.0000      1st Qu.: -1.0000      1st Qu.: -1.0000
## Mode  :character Median :  1.0000      Median :  1.0000      Median :  1.0000
##              Mean  :  0.3333      Mean  :  0.3333      Mean  :  0.3333
##              3rd Qu.:  1.0000      3rd Qu.:  1.0000      3rd Qu.:  1.0000
##              Max.   :  1.0000      Max.   :  1.0000      Max.   :  1.0000
## frog.weight sample_id pesticide      rate
## Min.   :1.012 Length:192      Length:192      Length:192
## 1st Qu.:2.745 Class :character  Class :character  Class :character
## Median :3.142 Mode  :character  Mode  :character  Mode  :character
## Mean   :3.299
## 3rd Qu.:3.789
## Max.   :6.739
##      conc
## Min.   : 0.001061
## 1st Qu.: 0.069055
## Median : 0.212920
## Mean   : 0.801643
## 3rd Qu.: 0.521471
## Max.   :19.879783
```

Van Meter et al. 2018 (Multiple Pesticides Study)

Van Meter et al. 2018 evaluated risks to amphibians after exposure to a single pesticide and pesticide mixtures. The five pesticides studied were three herbicides (atrazine, metolachlor, and 2,4-D), one insecticide (malathion), and one fungicide (propiconazole). Juvenile green frogs (*Lithobates clamitans*) were exposed to contaminated soils for 8 hours and metabolic analysis of amphibian livers was conducted to measure the effects. Pesticides were applied at the maximum legally allowable application rates individually and in mixtures of two or three pesticides within an herbicide or mixed pesticide group, scaled down to the area of six 0.94 L Pyrex glass bowls each with a 15 cm diameter.

Two data sets were generated from this study, one containing data for exposure to herbicides (single and mixed) and the other containing data for exposure to mixed pesticide treatments (herbicides, insecticide, fungicide).

Herbicide Data Set Data Set Dimensions, Column Names, and Summary:

[1] 378 10

```
## [1] "Group"      "ATZ"         "D"           "ME"          "AppRate"     "Weight"
## [7] "SA"          "Media"       "Pesticide"   "Conc"
```

```
##      Group          ATZ              D              ME
## Length:378      Min.   :-1.0000   Min.   :-1.0000   Min.   :-1.0000
## Class :character 1st Qu.:-1.0000   1st Qu.:-1.0000   1st Qu.:-1.0000
## Mode  :character Median : 1.0000   Median : 1.0000   Median : 1.0000
##              Mean  : 0.1429   Mean  : 0.1429   Mean  : 0.1429
##              3rd Qu.: 1.0000   3rd Qu.: 1.0000   3rd Qu.: 1.0000
##              Max.   : 1.0000   Max.   : 1.0000   Max.   : 1.0000
##      AppRate      Weight          SA              Media
## Min.   :14.30   Min.   :0.9634   Min.   :1.107   Length:378
## 1st Qu.:23.60   1st Qu.:1.6929   1st Qu.:1.534   Class :character
## Median :37.90   Median :2.0637   Median :1.720   Mode  :character
## Mean   :39.31   Mean   :2.0892   Mean   :1.715
## 3rd Qu.:54.50   3rd Qu.:2.4927   3rd Qu.:1.919
## Max.   :68.80   Max.   :3.6843   Max.   :2.406
##      Pesticide      Conc
## Length:378      Min.   : 0.00000
## Class :character 1st Qu.: 0.00000
## Mode  :character Median : 0.06358
##              Mean   : 5.64721
##              3rd Qu.: 1.46036
##              Max.   :76.03573
```

Mixed Pesticide Data Set Data Set Dimensions, Column Names, and Summary:

[1] 216 9

```
## [1] "Group"      "ATZ"         "MA"          "PROP"        "Pesticide"   "Media"
## [7] "Conc"       "Weight"      "SA"
```

```
##      Group          ATZ              MA              PROP
## Length:216      Min.   :-1.0000   Min.   :-1.0000   Min.   :-1.0000
## Class :character 1st Qu.:-1.0000   1st Qu.:-1.0000   1st Qu.:-1.0000
## Mode  :character Median : 1.0000   Median : 1.0000   Median : 1.0000
##              Mean  : 0.3333   Mean  : 0.3333   Mean  : 0.3333
##              3rd Qu.: 1.0000   3rd Qu.: 1.0000   3rd Qu.: 1.0000
##              Max.   : 1.0000   Max.   : 1.0000   Max.   : 1.0000
##      Pesticide      Media          Conc              Weight
## Length:216      Length:216      Min.   : 0.00024   Min.   :1.188
## Class :character Class :character 1st Qu.: 0.32682   1st Qu.:1.786
## Mode  :character Mode  :character Median : 1.61181   Median :2.014
##              Mean   : 5.46049   Mean   :2.203
##              3rd Qu.: 9.99874   3rd Qu.:2.455
##              Max.   :71.52122   Max.   :4.014
##      SA
## Min.   :1.447
```



```
## 1st Qu.:1.833
## Median :1.965
## Mean :2.047
## 3rd Qu.:2.203
## Max. :2.929
```

The herbicide and mixed pesticide data sets were cleaned prior and joined into a merged data set (referred to as Van Meter et al. 2018 Multiple Pesticides Study in subsequent steps). The single and mixed-pesticide treatments that were retained in the merged data set include atrazine, propiconazole, 2,4-D, malathion, and metolachlor. Original columns from the herbicide and mixed pesticide data sets were altered for standardization. These standardized columns will be used in future data cleaning steps in order to merge all data sets.

Merged Data Set Data Set Dimensions, Column Names, and Summary:

```
## [1] 137 12

## [1] "app_rate_g_cm2" "body_weight_g" "chemical" "tissue_conc_ugg"
## [5] "sample_id" "source" "application" "exp_duration"
## [9] "formulation" "soil_conc_ugg" "soil_type" "species"

## app_rate_g_cm2 body_weight_g chemical tissue_conc_ugg
## Min. :2.600e-06 Min. :0.9634 Length:137 Min. : 0.00054
## 1st Qu.:1.430e-05 1st Qu.:1.7623 Class :character 1st Qu.: 0.27576
## Median :2.360e-05 Median :2.0136 Mode :character Median : 1.41009
## Mean :2.004e-05 Mean :2.1086 Mean : 7.36154
## 3rd Qu.:2.590e-05 3rd Qu.:2.3395 3rd Qu.: 9.95084
## Max. :3.090e-05 Max. :4.0141 Max. :72.62672
## sample_id source application exp_duration
## Length:137 Length:137 Length:137 Min. :8
## Class :character Class :character Class :character 1st Qu.:8
## Mode :character Mode :character Mode :character Median :8
## Mean :8
## 3rd Qu.:8
## Max. :8
## formulation soil_conc_ugg soil_type species
## Min. :0 Mode:logical Mode:logical Length:137
## 1st Qu.:0 NA's:137 NA's:137 Class :character
## Median :0 Mode :character
## Mean :0
## 3rd Qu.:0
## Max. :0
```

Glinski et al. 2020 (Dermal Routes)

Glinski et al. 2020 assessed dermal uptake in amphibians from exposure to three pesticides (bifenthrin, chlorpyrifos, trifloxystrobin). Pesticide body burdens and hepatic metabolome for Leopard frogs were measured for two routes of uptake: uptake from contaminated soils versus uptake from contaminated surface water. Pesticides were applied at 1 ppm, scaled down to the area of eight 0.94 L Pyrex glass bowls each with a 15 cm diameter.

Data Set Dimensions, Column Names, and Summary:

```
## [1] 192 5

## [1] "Sample.ID" "Analyte" "Media" "Matrix"
## [5] "Concentration"

## Sample.ID Analyte Media Matrix
## Length:192 Length:192 Length:192 Length:192
## Class :character Class :character Class :character Class :character
## Mode :character Mode :character Mode :character Mode :character
##
##
##
## Concentration
## Min. :0.00000
## 1st Qu.:0.01036
## Median :0.15326
## Mean :0.33962
## 3rd Qu.:0.44162
## Max. :3.40759
```

Application Rates

The table below concisely displays the pesticide applications rates (ug/cm²) used in each relevant study as well as the variables used to compute the application rates.

Cleaning and Merging the Data Sets

Each data set was cleaned for merging. This consisted of dropping unneeded columns and standardizing column names of retained columns. Four columns were added to all data sets (soil type, formulation, exposure duration, and research study source).

Once each data set was cleaned, a local copy was saved and the data set was merged with the previously cleaned data sets.

The process of cleaning and merging each data set is briefly described below.

Van Meter et al. 2014/2015

Metabolites and parents that do not include metabolites were dropped from the data set. This includes atrazine, deisopropyl atrazine, desethyl atrazine, fipronil, fipronil-sulfone, triadimefon, triadimenol.

```
# drop metabolites and parents that do not include metabolites
vm2015_chem_drop <- c("Atrazine", "Deisopropyl Atrazine", "Desethyl Atrazine", "Fipronil", "Fipronil-Sulfone")
chem_vector_drop <- which(vm2015$Chemical %in% vm2015_chem_drop)
vm2015_subset1 <- vm2015[-chem_vector_drop,]
vm2015_subset2 <- droplevels(vm2015_subset1)
```

pesticide	app_rate_ug_cm2	applied_mL	container	area_cm2	total
Van Meter et al. 2014/2015					
atrazine	22.9000	75 MeOH	10-gal aquarium	1225	
fipronil	1.1000	75 MeOH	10-gal aquarium	1225	
imidacloprid	5.7000	75 MeOH	10-gal aquarium	1225	
pendimethalin	19.8000	75 MeOH	10-gal aquarium	1225	
triadimefon	2.7000	75 MeOH	10-gal aquarium	1225	
Van Meter et al. 2016					
atrazine	22.9000	75 MeOH	.94 L bowl	225*6	
fipronil	1.1000	75 MeOH	.94 L bowl	225*6	
imidacloprid	5.7000	75 MeOH	.94 L bowl	225*6	
pendimethalin	69.8000	75 MeOH	.94 L bowl	225*6	
triadimefon	2.7000	75 MeOH	.94 L bowl	225*6	
Van Meter et al. 2018					
atrazine	23.6000	50 MeOH	.94 L bowl	225*6	
2,4-D	14.3000	50 MeOH	.94 L bowl	225*6	
metolachlor	30.9000	50 MeOH	.94 L bowl	225*6	
malathion	25.9000	50 MeOH	.94 L bowl	225*6	
propiconazole	2.6000	50 MeOH	.94 L bowl	225*6	
Henson-Ramsey et al. 2008					
malathion	50.0000	NA	cage	1060	
Glinski et al. 2018a					
atrazine	23.9500	75 MeOH	.94 L bowl	225*6	
chlorothalonil	44.3000	75 MeOH	.94 L bowl	225*6	
imidacloprid	5.3900	75 MeOH	.94 L bowl	225*6	
metolachlor	31.0100	75 MeOH	.94 L bowl	225*6	
triadimefon	2.9100	75 MeOH	.94 L bowl	225*6	
Glinski et al. 2018b					
atrazine	22.9000	75 MeOH	10-gal aquarium	1225	
fipronil	1.1000	75 MeOH	10-gal aquarium	1225	
triadimefon	2.7000	75 MeOH	10-gal aquarium	1225	
Glinski et al. 2019					
bifenthrin (max)	3.4500	75 MeOH	.94 L bowl	225*8	
metolachlor (max)	30.6200	75 MeOH	.94 L bowl	225*8	
triadimefon (max)	2.8700	75 MeOH	.94 L bowl	225*8	
bifenthrin (1/10 max)	0.3450	75 MeOH	.94 L bowl	225*8	
metolachlor (1/10 max)	3.0620	75 MeOH	.94 L bowl	225*8	
triadimefon (1/10 max)	0.2870	75 MeOH	.94 L bowl	225*8	
Glinski et al. 2020					
bifenthrin	0.2889	400ml of 1ppm pesticide in water	.94 L bowl	225*8	
chlorpyrifos	0.3111	400ml of 1ppm pesticide in water	.94 L bowl	225*8	
trifloxystrobin	0.3022	400ml of 1ppm pesticide in water	.94 L bowl	225*8	

There were 278 observations with these chemicals. After dropping the 278 observations from the initial 474, the updated dimensions are:

```
## [1] 196 23
```

There were 15 unneeded columns dropped and 4 added for standarization.

```
# drop unneeded columns for merging
all_cols <- colnames(vm2015_subset2)
drop_cols <- c("Instrument", "good", "logKow", "BCF", "initialweight",
              "Solat20C_mgL", "Solat20C_gL", "molmass_gmol", "Density_gcm3", "AppFactor", "SA_cm2", "VapPr",
              "Koc_gmL", "HalfLife_day", "HabFac")
vm2015_subset3 <- vm2015_subset2[,!(names(vm2015_subset2) %in% drop_cols)]
colnames(vm2015_subset3)
```

```
## [1] "Species"      "Sample"      "Chemical"    "Application"
## [5] "app_rate_g_cm2" "TissueConc"  "SoilConc"    "bodyweight"
```

```
# add columns
soil_type <- c(rep("PLE", nrow(vm2015_subset3)))
formulation <- (rep(0, nrow(vm2015_subset3)))
exp_duration <- (rep(8, nrow(vm2015_subset3)))
source <- c(rep("rvm2015", nrow(vm2015_subset3)))
vm2015_subset4 <- cbind(vm2015_subset3, formulation, soil_type, exp_duration, source)
# standardize column names
colnames(vm2015_subset4)
```

```
## [1] "Species"      "Sample"      "Chemical"    "Application"
## [5] "app_rate_g_cm2" "TissueConc"  "SoilConc"    "bodyweight"
## [9] "formulation"   "soil_type"   "exp_duration" "source"
```

```
colnames(vm2015_subset4)[which(colnames(vm2015_subset4)=="Sample")]<-"sample_id"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4)=="Species")]<-"species"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4)=="Chemical")]<-"chemical"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4)=="Application")]<-"application"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4)=="TissueConc")]<-"tissue_conc_ugg"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4)=="SoilConc")]<-"soil_conc_ugg"
colnames(vm2015_subset4)[which(colnames(vm2015_subset4)=="bodyweight")]<-"body_weight_g"
colnames(vm2015_subset4)
```

```
## [1] "species"      "sample_id"   "chemical"    "application"
## [5] "app_rate_g_cm2" "tissue_conc_ugg" "soil_conc_ugg" "body_weight_g"
## [9] "formulation"   "soil_type"   "exp_duration" "source"
```

```
# reorder vm2015 alphabetically
vm2015_merge <- vm2015_subset4[,order(names(vm2015_subset4))]

# write a local copy
vm2015_merge_filename <- paste(amphibdir_data_out, "vm2015_merge.csv", sep="")
write.csv(vm2015_merge, file=vm2015_merge_filename)
```

The data set's dimensions are:

```
## [1] 196 12
```

Van Meter et al. 2016

From the initial 11 columns, 4 columns were dropped and consolidated into 1, and 4 columns were added.

```
# add sample_id
vm2016$sample_id <- paste(vm2016$Day, vm2016$Row, vm2016$Column, sep="_")
vm2016_subset2 <- subset(vm2016, select=c(-Day, -Row, -Column, -Total))
# add additional columns
species <- c(rep("American toad", nrow(vm2016_subset2)))
application <- c(rep("Indirect", nrow(vm2016_subset2)))
exp_duration <- (rep(8, nrow(vm2016_subset2)))
source <- c(rep("rvm2016", nrow(vm2016_subset2)))
vm2016_subset3 <- cbind(vm2016_subset2, species, application, exp_duration, source)
```

Application rates for several pesticides were inserted. There were 108 observations with decay products that were not sprayed; these observations were dropped so as to only include the parents in the cleaned data set. There were 60 observations with atrazine, fipronil, or triadimefon that were dropped because they do not include metabolites in total.

```
# assign values to application rate
#unique(vm2016_subset3$Pesticide)
vm2016_subset3$app_rate_g_cm2[vm2016_subset3$Pesticide=="ATZTOT"] <- 22.9e-6
vm2016_subset3$app_rate_g_cm2[vm2016_subset3$Pesticide=="Imid"] <- 5.7e-6
vm2016_subset3$app_rate_g_cm2[vm2016_subset3$Pesticide=="FipTOT"] <- 1.1e-6
vm2016_subset3$app_rate_g_cm2[vm2016_subset3$Pesticide=="TNDTOT"] <- 2.7e-6
vm2016_subset3$app_rate_g_cm2[vm2016_subset3$Pesticide=="Pendi"] <- 69.8e-6
# drop decay products that were not sprayed, keeping only parents
rows_to_drop <- which(vm2016_subset3$Parent == 0)
vm2016_subset4 <- vm2016_subset3[-rows_to_drop,]
# drop ATZ, Fip, TDN since do not include metabolites in total
chems_to_drop <- c("ATZ", "Fip", "TDN")
vm2016_subset5 <- vm2016_subset4[!(vm2016_subset4$Pesticide %in% chems_to_drop),]
# drop parent field
drop_cols <- c("Parent")
vm2016_subset6 <- vm2016_subset5[,!(names(vm2016_subset5) %in% drop_cols)]
```

Several column names were standardized and all columns were ordered for ease of merging with the combined data set.

```
# standardize column names
colnames(vm2016_subset6)
```

```
## [1] "Pesticide"      "SoilType"      "BodyBurden"    "Soil"
## [5] "Weight"        "Formulation"   "sample_id"     "species"
## [9] "application"   "exp_duration"  "source"        "app_rate_g_cm2"
```

```
colnames(vm2016_subset6)[which(colnames(vm2016_subset6)=="Pesticide")]<-"chemical"
colnames(vm2016_subset6)[which(colnames(vm2016_subset6)=="SoilType")]<-"soil_type"
colnames(vm2016_subset6)[which(colnames(vm2016_subset6)=="BodyBurden")]<-"tissue_conc_ugg"
colnames(vm2016_subset6)[which(colnames(vm2016_subset6)=="Soil")]<-"soil_conc_ugg"
colnames(vm2016_subset6)[which(colnames(vm2016_subset6)=="Weight")]<-"body_weight_g"
colnames(vm2016_subset6)[which(colnames(vm2016_subset6)=="Formulation")]<-"formulation"
```

```

# alter chemical name
vm2016_subset6$chemical <- as.character(vm2016_subset6$chemical)
vm2016_subset6$chemical[vm2016_subset6$chemical=="Imid"] <- "imidacloprid"

# reorder columns alphabetically to help with merge
colnames(vm2016_subset6)

## [1] "chemical"      "soil_type"      "tissue_conc_ugg" "soil_conc_ugg"
## [5] "body_weight_g" "formulation"    "sample_id"      "species"
## [9] "application"   "exp_duration"   "source"          "app_rate_g_cm2"

vm2016_merge <- vm2016_subset6[,order(names(vm2016_subset6))]
colnames(vm2016_merge)

## [1] "app_rate_g_cm2" "application"    "body_weight_g"  "chemical"
## [5] "exp_duration"   "formulation"    "sample_id"      "soil_conc_ugg"
## [9] "soil_type"      "source"         "species"        "tissue_conc_ugg"

# write a local copy
vm2016_merge_filename <- paste(amphibdir_data_out,"vm2016_merge.csv", sep="")
write.csv(vm2016_merge, file=vm2016_merge_filename)

```

The updated dimensions are:

```
## [1] 96 12
```

The Van Meter et al. 2014/2015 and Van Meter et al. 2016 data sets were combined.

The combined data set's updated dimensions are:

```
## [1] 292 12
```

Glinski et al. 2018a (Dehydration)

The metabolite products were dropped from the data set; 600 rows from the initial 1494 rows were retained.

```

# drop metabolite products
parent_keepers <- which(as.vector(dag2016_dehy0$parent) == as.vector(dag2016_dehy0$analyte))
dag2016_dehy1 <- dag2016_dehy0[parent_keepers,]

```

Several column names were altered for standarization across the data set, and 7 columns were added for standarization.

```

## time is length of dehydration
#colnames(dag2016_dehy1)[which(colnames(dag2016_dehy1)=="time")]<-"exp_duration"

# standardize column names
colnames(dag2016_dehy1)[which(colnames(dag2016_dehy1)=="analyte")]<-"chemical"
colnames(dag2016_dehy1)[which(colnames(dag2016_dehy1)=="conc")]<-"tissue_conc_ugg"
colnames(dag2016_dehy1)[which(colnames(dag2016_dehy1)=="ID")]<-"sample_id"

```

```
colnames(dag2016_dehy1)[which(colnames(dag2016_dehy1)=="weight")]<-"body_weight_g"

# add additional columns
exp_duration <- c(rep(8,nrow(dag2016_dehy1)))
soil_type <- c(rep("PLE",nrow(dag2016_dehy1)))
application <- c(rep("Indirect",nrow(dag2016_dehy1)))
formulation <- (rep(0,nrow(dag2016_dehy1)))
app_rate_g_cm2 <- (rep(0,nrow(dag2016_dehy1)))
soil_conc_ugg <- (rep(0,nrow(dag2016_dehy1)))
source <- c(rep("dag_dehydration",nrow(dag2016_dehy1)))
dag2016_dehy2 <- cbind(dag2016_dehy1, formulation, soil_type, application,
                      app_rate_g_cm2, exp_duration, soil_conc_ugg, source)
```

The updated dimensions are:

```
## [1] 600 15
```

Multiple soil concentration observations were given the same ID. Until a many-to-one merge of soil concentrations could be executed, 300 rows were temporarily dropped. There were also 3 columns dropped.

```
# drop the soil until we can do a many-to-one merge of soil concentrations
# drop decay products that were not sprayed, keeping only parents
rows_to_drop <- which(dag2016_dehy2$matrix == 'soil')
dag2016_dehy3 <- dag2016_dehy2[-rows_to_drop,]
# parent, time and matrix columns delete
drop_cols <- c("parent","time","matrix")
dag2016_dehy4 <- dag2016_dehy3[,!(names(dag2016_dehy3) %in% drop_cols)]
```

The updated dimensions are:

```
## [1] 300 12
```

The application rate values were inserted, the temporarily dropped soil concentrations were updated to the current data set, and the species names were standardized.

```
# fill in application rates
#unique(dag2016_dehy4$chemical)
update_atrazine <- which(dag2016_dehy4$chemical == 'atrazine')
dag2016_dehy4$app_rate_g_cm2[update_atrazine] <- 0.00002395 # atrazine g/cm2
update_chloro <- which(dag2016_dehy4$chemical == 'chloro+d')
dag2016_dehy4$app_rate_g_cm2[update_chloro] <- 0.0000443 # chloro g/cm2
update_metol <- which(dag2016_dehy4$chemical == 'metol')
dag2016_dehy4$app_rate_g_cm2[update_metol] <- 0.00003101 # metol g/cm2
update_tdn <- which(dag2016_dehy4$chemical == 'tdn')
dag2016_dehy4$app_rate_g_cm2[update_tdn] <- 0.00000291 # tdn g/cm2
update_imid <- which(dag2016_dehy4$chemical == 'imid')
dag2016_dehy4$app_rate_g_cm2[update_imid] <- 0.00000539 # imid g/cm2

# add back in soil concentrations (in already-made soil_conc_ugg column)
dag2016_soil <- dag2016_dehy2[rows_to_drop,]
dag2016_dehy4$soil_conc_ugg <- dag2016_soil$tissue_conc_ugg
```

```
# rename species names, according to standardized names
dag2016_dehy4$species <- as.character(dag2016_dehy4$species)
dag2016_dehy4$species[dag2016_dehy4$species == "LF"] <- "Leopard frog"
dag2016_dehy4$species[dag2016_dehy4$species == "BA"] <- "Fowlers toad"
dag2016_dehy4$species <- as.factor(dag2016_dehy4$species)
```

The dimensions are:

```
## [1] 300 12
```

The Glinkski et al. 2018a (Dehydration) was combined with the previously merged data sets.

The combined data set's updated dimensions are:

```
## [1] 592 12
```

Henson-Ramsey 2008

The Henson-Ramsey 2008 data set did not require any additional data cleaning. It was combined with the previously merged data sets.

The combined data set's updated dimensions are:

```
## [1] 601 12
```

Glinski et al. 2018b (Metabolites)

Apart from standardizing the species name, the Glinski et al. 2018b (Metabolites) data set did not require any additional data cleaning. It was combined with the previously merged data sets.

```
# rename species names, according to standardized names
dag2016_metabolite_merge$species <- as.character(dag2016_metabolite_merge$species)
dag2016_metabolite_merge$species[dag2016_metabolite_merge$species == "Anaxyrus fowleri"] <- "Fowlers toad"
dag2016_metabolite_merge$species <- as.factor(dag2016_metabolite_merge$species)
```

The combined data set's updated dimensions are:

```
## [1] 661 12
```

Glinski et al. 2019 (Biomarkers)

Five columns were dropped from the original biomarkers data set and the names of two columns were standardized.

```
# drop columns
drop_cols <- c("met", "tdt", "bif", "rate", "group")
dag_biomarker_subset <- dag_biomarker[, !(names(dag_biomarker) %in% drop_cols)]

# standardize column names
colnames(dag_biomarker_subset)[which(colnames(dag_biomarker_subset)=="conc")]<-"tissue_conc_ugg"
colnames(dag_biomarker_subset)[which(colnames(dag_biomarker_subset)=="frog.weight")]<-"body_weight_g"
```


The updated column names and dimensions are:

```
## [1] "body_weight_g"      "sample_id"           "pesticide"           "tissue_conc_ugg"

## [1] 192      4
```

The application rates and soil concentrations were not included in the original biomarkers data set. Both are included in the following data set:

Data Set Dimensions, Column Names, and Summary:

```
## [1] 136  15

## [1] "frog.weight" "SAMPLE"      "Met"         "TDN"         "TDL"
## [6] "BIF"         "soil.weight" "Met.soil"    "TDN.soil"    "TDL.soil"
## [11] "BIF.soil"    "Rate"        "app.rate.met" "app.rate.tdn" "app.rate.bif"
```

##	frog.weight	SAMPLE	Met	TDN
##	Min. :1.012	Length:136	Min. : 0.0000	Min. :0.00000
##	1st Qu.:2.749	Class :character	1st Qu.: 0.0000	1st Qu.:0.00000
##	Median :3.164	Mode :character	Median : 0.0000	Median :0.00000
##	Mean :3.302		Mean : 0.9123	Mean :0.06927
##	3rd Qu.:3.762		3rd Qu.: 0.4298	3rd Qu.:0.07447
##	Max. :6.784		Max. :19.8798	Max. :0.55921
##	TDL	BIF	soil.weight	Met.soil
##	Min. :0.00000	Min. :0.0000	Min. : 4.476	Min. :0.000
##	1st Qu.:0.00000	1st Qu.:0.0000	1st Qu.: 6.731	1st Qu.:0.000
##	Median :0.00000	Median :0.0000	Median : 7.772	Median :0.000
##	Mean :0.02259	Mean :0.1276	Mean : 8.043	Mean :1.605
##	3rd Qu.:0.01770	3rd Qu.:0.1299	3rd Qu.: 9.050	3rd Qu.:2.265
##	Max. :0.30815	Max. :1.0271	Max. :13.571	Max. :6.758
##	TDN.soil	TDL.soil	BIF.soil	Rate
##	Min. :0.0000	Min. :0.000000	Min. :0.0000	Length:136
##	1st Qu.:0.0000	1st Qu.:0.000000	1st Qu.:0.0000	Class :character
##	Median :0.0000	Median :0.000000	Median :0.0000	Mode :character
##	Mean :0.7168	Mean :0.010160	Mean :0.7417	
##	3rd Qu.:0.5312	3rd Qu.:0.007463	3rd Qu.:1.1472	
##	Max. :3.6300	Max. :0.061563	Max. :5.2658	
##	app.rate.met	app.rate.tdn	app.rate.bif	
##	Min. : 0.000	Min. :0.0000	Min. :0.0000	
##	1st Qu.: 0.000	1st Qu.:0.0000	1st Qu.:0.0000	
##	Median : 0.000	Median :0.0000	Median :0.0000	
##	Mean :14.263	Mean :1.3389	Mean :1.6070	
##	3rd Qu.: 5.511	3rd Qu.:0.5173	3rd Qu.:0.6209	
##	Max. :55.106	Max. :5.1730	Max. :6.2090	

The application rates were converted from mg to g/cm².

```
dag_biomarker2_update <- replace.value(dag_biomarker2, "app.rate.met", from=55.106, to=3.062e-5, verbose=TRUE)
dag_biomarker2_update <- replace.value(dag_biomarker2_update, "app.rate.met", from=5.5106, to=3.062e-6, verbose=TRUE)
dag_biomarker2_update <- replace.value(dag_biomarker2_update, "app.rate.tdn", from=5.173, to=2.87e-6, verbose=TRUE)
dag_biomarker2_update <- replace.value(dag_biomarker2_update, "app.rate.tdn", from=.5173, to=2.87e-7, verbose=TRUE)
dag_biomarker2_update <- replace.value(dag_biomarker2_update, "app.rate.bif", from=6.209, to=3.45e-6, verbose=TRUE)
dag_biomarker2_update <- replace.value(dag_biomarker2_update, "app.rate.bif", from=.6209, to=3.45e-7, verbose=TRUE)
```

A one-to-one merge was conducted based on the unique sample id for each measured pesticide (either bifenthrin, metolachlor, or triadimefon) to join the original biomarkers data set and the data set containing the application rates and soil concentrations. Vectors containing the application rates and soil concentrations were joined to the original data set.

```
# bif extraction
dag_biomarker_subset_bif <- dag_biomarker_subset[dag_biomarker_subset$pesticide == "bif", ]
dag_biomarker2_subset_bif <- dag_biomarker2_update[dag_biomarker2_update$BIF != 0, ]

dag_biomarker_bif_merge <- merge(x = dag_biomarker_subset_bif, y = dag_biomarker2_subset_bif,
                                by.x = "sample_id", by.y = "SAMPLE", all.x = TRUE)

# met extraction
dag_biomarker_subset_met <- dag_biomarker_subset[dag_biomarker_subset$pesticide == "met", ]
dag_biomarker2_subset_met <- dag_biomarker2_update[dag_biomarker2_update$Met != 0, ]

dag_biomarker_met_merge <- merge(x = dag_biomarker_subset_met, y = dag_biomarker2_subset_met,
                                by.x = "sample_id", by.y = "SAMPLE", all.x = TRUE)

# tdt extraction
dag_biomarker_subset_tdt <- dag_biomarker_subset[dag_biomarker_subset$pesticide == "tdt", ]
dag_biomarker2_subset_tdt <- dag_biomarker2_update[dag_biomarker2_update$TDN != 0, ]

dag_biomarker_tdt_merge <- merge(x = dag_biomarker_subset_tdt, y = dag_biomarker2_subset_tdt,
                                by.x = "sample_id", by.y = "SAMPLE", all.x = TRUE)

# combine bif, met, and tdt
app_bind_bmt <- c(dag_biomarker_bif_merge[, "app.rate.bif"],
                  dag_biomarker_met_merge[, "app.rate.met"], dag_biomarker_tdt_merge[, "app.rate.tdn"])

soil_bind_bmt <- c(dag_biomarker_bif_merge[, "BIF.soil"],
                  dag_biomarker_met_merge[, "Met.soil"], dag_biomarker_tdt_merge[, "TDN.soil"])

# join app and soil vectors to data set
dag_biomarker_subset2 <- dag_biomarker_subset[order(dag_biomarker_subset[, 3]),]
rownames(dag_biomarker_subset2) <- seq(length=nrow(dag_biomarker_subset2))

dag_biomarker_subset3 <- cbind(dag_biomarker_subset2, app_bind_bmt, soil_bind_bmt)

# standardize column names
colnames(dag_biomarker_subset3)[which(colnames(dag_biomarker_subset3)=="app_bind_bmt")]<-"app_rate_g_cm2"
colnames(dag_biomarker_subset3)[which(colnames(dag_biomarker_subset3)=="soil_bind_bmt")]<-"soil_conc_ugg"
```

The updated column names and dimensions are:

```
## [1] "body_weight_g" "sample_id" "pesticide" "tissue_conc_ugg"
## [5] "app_rate_g_cm2" "soil_conc_ugg"

## [1] 192 6
```

New columns were created for standarization, the columns were ordered alphabetically, and a local copy was stored.

```

# create new columns
application <- c(rep("soil", nrow(dag_biomarker_subset3)))
exp_duration <- c(rep(8, nrow(dag_biomarker_subset3)))
formulation <- c(rep(0, nrow(dag_biomarker_subset3)))
soil_type <- c(rep(NA, nrow(dag_biomarker_subset3)))
source <- c(rep("dag_biomarker", nrow(dag_biomarker_subset3)))
species <- c(rep("Leopard frog", nrow(dag_biomarker_subset3)))

# combine columns
dag_biomarker_subset4 <- cbind(dag_biomarker_subset3, application, exp_duration,
                              formulation, soil_type, source, species)

# standardize pesticide column
dag_biomarker_subset4$pesticide <- as.character(dag_biomarker_subset4$pesticide)
dag_biomarker_subset4$pesticide[dag_biomarker_subset4$pesticide == "bif"] <- "Bifenthrin"
dag_biomarker_subset4$pesticide[dag_biomarker_subset4$pesticide == "met"] <- "Metolachlor"
dag_biomarker_subset4$pesticide[dag_biomarker_subset4$pesticide == "tdt"] <- "Triadimefon"

colnames(dag_biomarker_subset4)[which(colnames(dag_biomarker_subset4)=="pesticide")]<-"chemical"

# unite function for sample id and chemical
dag_biomarker_subset5 <- unite(data = dag_biomarker_subset4, col = "sample_id", "sample_id", "chemical")

# order columns in abc for merge
dag_biomarker_merge <- dag_biomarker_subset5[ ,order(names(dag_biomarker_subset5))]

```

The updated column names and dimensions are:

```

## [1] "app_rate_g_cm2" "application" "body_weight_g" "chemical"
## [5] "exp_duration" "formulation" "sample_id" "soil_conc_ugg"
## [9] "soil_type" "source" "species" "tissue_conc_ugg"

## [1] 192 12

```

The Glinski et al. 2019 (Biomarkers) was combined with the previously merged data sets.

The combined data set's updated dimensions are:

```
## [1] 853 12
```

Van Meter et al. 2018 (Multiple Pesticides Study)

The Van Meter et al. 2018 (Multiple Pesticides Study) data set did not require any additional data cleaning. It was combined with the previously merged data sets.

The combined data set's updated dimensions are:

```
## [1] 990 12
```

Glinski et al. 2020 (Dermal Routes)

The dermal routes data set did not include the body weights for the measured amphibians. These weights were included in a separate data set:

Data Set Dimensions, Column Names, and Summary:

```
## [1] 48 2
```

```
## [1] "Weight_g" "Sample"
```

```
##      Weight_g      Sample
## Min.      :0.9555  Length:48
## 1st Qu.:1.4204   Class :character
## Median :1.7817   Mode  :character
## Mean      :1.7784
## 3rd Qu.:2.1319
## Max.      :2.8197
```

A one-to-many merge was employed to merge the dermal routes data set and the weights data set based on the Sample ID. Only rows where the Matrix is “Amphibian” have a body weight; all other rows are NA.

```
# merge (one-to-many) dermal routes data with weights data, based on Sample ID
dermal_routes_subset2 <- dermal_routes[order(dermal_routes$Sample.ID), ]
weights_2 <- weights[order(weights$Sample),]

dermal_routes_subset3 <- merge(dermal_routes_subset2, weights_2,
                              by.x = "Sample.ID", by.y = "Sample", all.x = TRUE, all.y = TRUE)
```

The updated dimensions are:

```
## [1] 192 6
```

The soil concentrations, where the Media and Matrix are both “Soil,” was subset from the data set to be used later in the data cleaning process. These soil concentrations (currently listed in the “Concentration” column) will be used for the soil_conc_ugg column in the cleaned data set.

```
# subset soil to be used later for soil concentration column (will use "Concentration" column)
soil_subset <- dermal_routes_subset2[dermal_routes_subset2$Media == "Soil", ]
soil_subset2 <- soil_subset[soil_subset$Matrix == "Soil",]
```

The dimensions of this soil subset are:

```
## [1] 48 5
```

Referring back to the main dermal routes data set: we are only interested in the pesticide exposures on amphibians while in soil. These rows were subset.

```

# want Media == soil because interested in dermal exposure in soil
dermal_routes_subset4 <- dermal_routes_subset3[dermal_routes_subset3$Media == "Soil",]
#sum(dermal_routes_subset3$Media == "Soil") # == 96
#dim(dermal_routes_subset4) # == 96 x 6

# want Matrix == Amphibian because interested in amphib exposure
dermal_routes_subset5 <- dermal_routes_subset4[dermal_routes_subset4$Matrix == "Amphibian", ]
#sum(dermal_routes_subset4$Matrix == "Amphibian") # == 48
#dim(dermal_routes_subset5) # == 48 x 6

```

The updated dimensions are:

```
## [1] 48 6
```

The soil concentrations were appended to the main dermal routes data set.

```

# add in soil concentration column, previously subset
# order by Sample.ID, then by Analyte name to match up rows for the two data sets
dermal_routes_subset6 <- dermal_routes_subset5[order(dermal_routes_subset5[,1],
                                                    dermal_routes_subset5[,2]),]
soil_subset3 <- soil_subset2[order(soil_subset2[,1], soil_subset2[,2]),]

#dim(dermal_routes_subset6) # == 48 x 6
#dim(soil_subset3) # == 48 x 5

dermal_routes_subset7 <- cbind(dermal_routes_subset6, soil_subset3$Concentration)

```

The updated dimensions are:

```
## [1] 48 7
```

The metabolites were dropped from the data set. Additionally, several new columns were created for standardization, existing columns were standardized according to the naming conventions of the collated data set, and unneeded columns were dropped. Columns were ordered alphabetically for ease of merging.

```

# drop metabolites
rows_to_drop <- c("4-OH", "CPO", "TFSa")
dermal_routes_subset8 <- dermal_routes_subset7[!(dermal_routes_subset7$Analyte %in% rows_to_drop),]

# create new columns
app_rate_g_cm2 <- c(rep(NA, nrow(dermal_routes_subset8)))
application <- c(rep("soil", nrow(dermal_routes_subset8)))
exp_duration <- c(rep(8, nrow(dermal_routes_subset8)))
formulation <- c(rep(0, nrow(dermal_routes_subset8)))
soil_type <- c(rep("OLS", nrow(dermal_routes_subset8)))
source <- c(rep("dag_dermal_routes", nrow(dermal_routes_subset8)))
species <- c(rep("Leopard frog", nrow(dermal_routes_subset8)))

# alter existing column names
colnames(dermal_routes_subset8)

```

```
## [1] "Sample.ID"          "Analyte"
## [3] "Media"                "Matrix"
## [5] "Concentration"        "Weight_g"
## [7] "soil_subset3$Concentration"

colnames(dermal_routes_subset8)[which(colnames(dermal_routes_subset8)=="Analyte")]<-"chemical"
colnames(dermal_routes_subset8)[which(colnames(dermal_routes_subset8)=="Sample.ID")]<-"sample_id"
colnames(dermal_routes_subset8)[which(colnames(dermal_routes_subset8)=="Concentration")]<-"tissue_conc_ug"
colnames(dermal_routes_subset8)[which(colnames(dermal_routes_subset8)=="soil_subset3$Concentration")]<-"soil_conc_ug"
colnames(dermal_routes_subset8)[which(colnames(dermal_routes_subset8)=="Weight_g")]<-"body_weight_g"

# combine columns
dermal_routes_subset9 <- cbind(dermal_routes_subset8, app_rate_g_cm2, application, exp_duration,
                              formulation, soil_type, source, species)
names(dermal_routes_subset9)

## [1] "sample_id"          "chemical"          "Media"            "Matrix"
## [5] "tissue_conc_ug"     "body_weight_g"     "soil_conc_ug"     "app_rate_g_cm2"
## [9] "application"        "exp_duration"      "formulation"       "soil_type"
## [13] "source"             "species"

# drop columns
cols_to_drop <- c("Matrix", "Media")
dermal_routes_subset10 <- dermal_routes_subset9[, !(names(dermal_routes_subset9) %in% cols_to_drop)]

# insert application rates
dermal_routes_subset10$chemical <- as.character(dermal_routes_subset10$chemical)
unique(dermal_routes_subset10$chemical)

## [1] "BIF" "CPF" "TFS"

dermal_routes_subset10$app_rate_g_cm2[dermal_routes_subset10$chemical == "BIF"] <- 2.8889e-7 #bifenthrin
dermal_routes_subset10$app_rate_g_cm2[dermal_routes_subset10$chemical == "CPF"] <- 3.1111e-7 #chlorpyrif
dermal_routes_subset10$app_rate_g_cm2[dermal_routes_subset10$chemical == "TFS"] <- 3.0222e-7 #trifloxyst

summary(dermal_routes_subset10$app_rate_g_cm2)

##      Min.   1st Qu.   Median     Mean   3rd Qu.    Max.
## 2.889e-07 2.889e-07 3.022e-07 3.007e-07 3.111e-07 3.111e-07

# order columns in abc for merge
dermal_routes_merge <- dermal_routes_subset10[, order(names(dermal_routes_subset10))]
```

The updated column names and dimensions are:

```
## [1] 24 12

## [1] "app_rate_g_cm2" "application"    "body_weight_g"  "chemical"
## [5] "exp_duration"   "formulation"    "sample_id"      "soil_conc_ug"
## [9] "soil_type"      "source"         "species"        "tissue_conc_ug"
```

A local copy was saved, and the data set was combined with the collated data set.

The combined data set's updated dimensions are:

```
## [1] 1014 12
```

Final Product

Minor alterations were made to the final collated data set to standardize names of the application types and chemicals.

```
amphib_dermal_collated <- combined_data6
```

```
colnames(amphib_dermal_collated)
```

```
## [1] "app_rate_g_cm2" "application" "body_weight_g" "chemical"
## [5] "exp_duration" "formulation" "sample_id" "soil_conc_ugg"
## [9] "soil_type" "source" "species" "tissue_conc_ugg"
```

```
# check to see if everything ok
```

```
summary(amphib_dermal_collated$app_rate_g_cm2)
```

```
##      Min.   1st Qu.   Median     Mean   3rd Qu.     Max.
## 2.870e-07 2.790e-06 5.700e-06 1.616e-05 2.395e-05 6.980e-05
```

```
summary(amphib_dermal_collated$body_weight_g)
```

```
##      Min. 1st Qu.  Median     Mean 3rd Qu.     Max.
## 0.1879 1.3043 2.1247 3.3800 3.0412 50.9200
```

```
summary(amphib_dermal_collated$exp_duration)
```

```
##      Min. 1st Qu.  Median     Mean 3rd Qu.     Max.
## 2.000 8.000 8.000 8.876 8.000 48.000
```

```
summary(amphib_dermal_collated$soil_conc_ugg) # 206 NAs
```

```
##      Min. 1st Qu.  Median     Mean 3rd Qu.     Max.   NA's
## 0.1125 2.0709 5.2459 14.3042 15.3781 238.1502    206
```

```
summary(amphib_dermal_collated$tissue_conc_ugg)
```

```
##      Min. 1st Qu.  Median     Mean 3rd Qu.     Max.
## 0.00054 0.16908 0.52573 2.51415 2.06812 72.62672
```

```

# standardize application levels
amphib_dermal_collated$application <- tolower(amphib_dermal_collated$application)
amphib_dermal_collated$application <- as.factor(amphib_dermal_collated$application)

# standardize chemical levels
amphib_dermal_collated$chemical <- as.character(amphib_dermal_collated$chemical)

amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "fip"] <- "fipronil"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "BIF"] <- "bifenthrin"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "MET"] <- "metolachlor"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "MAT"] <- "malathion"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "ATZT"] <- "atrazine"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "PROPT"] <- "propiconazole"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "metol"] <- "metolachlor"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "tdn"] <- "triadimefon"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "imid"] <- "imidacloprid"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "chloro+d"] <- "chlorothalonil"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "CPF"] <- "chlorpyrifos"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "TFS"] <- "trifloxystrobin"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "FipTOT"] <- "fipronil"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "ATZTOT"] <- "atrazine"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "TNDTOT"] <- "triadimefon"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "Pendi"] <- "pendimethalin"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "Total Atrazine"] <- "atrazine"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "Total Fipronil"] <- "fipronil"
amphib_dermal_collated$chemical[amphib_dermal_collated$chemical == "Total Triadimefon"] <- "triadimefon"

amphib_dermal_collated$chemical <- tolower(amphib_dermal_collated$chemical)
amphib_dermal_collated$chemical <- as.factor(amphib_dermal_collated$chemical)

# write out file
amphib_dermal_collated_filename <- paste(amphibdir_data_out, "amphib_dermal_collated.csv", sep="")
write.csv(amphib_dermal_collated, file=amphib_dermal_collated_filename)

```

Column Names

```

## [1] "app_rate_g_cm2" "application" "body_weight_g" "chemical"
## [5] "exp_duration" "formulation" "sample_id" "soil_conc_ugg"
## [9] "soil_type" "source" "species" "tissue_conc_ugg"

```

Dimensions

```
## [1] 1014 12
```

Variable Summaries

```

## app_rate_g_cm2 application body_weight_g chemical
## Min. :2.870e-07 indirect :396 Min. : 0.1879 triadimefon :223
## 1st Qu.:2.790e-06 overspray: 45 1st Qu.: 1.3043 atrazine :191
## Median :5.700e-06 soil :573 Median : 2.1247 metolachlor :154

```



```
## Mean      :1.616e-05      Mean      : 3.3800      imidacloprid: 78
## 3rd Qu.:2.395e-05      3rd Qu.: 3.0412      bifenthrin  : 72
## Max.      :6.980e-05      Max.      :50.9200      fipronil    : 71
##                                     (Other)    :225
## exp_duration      formulation      sample_id      soil_conc_ugg
## Min.      : 2.000      Min.      :0.00000      Length:1014      Min.      : 0.1125
## 1st Qu.: 8.000      1st Qu.:0.00000      Class :character      1st Qu.: 2.0709
## Median : 8.000      Median :0.00000      Mode  :character      Median : 5.2459
## Mean      : 8.876      Mean      :0.03582      Mean      : 14.3042
## 3rd Qu.: 8.000      3rd Qu.:0.00000      3rd Qu.: 15.3781
## Max.      :48.000      Max.      :1.00000      Max.      :238.1502
##                                     NA's      :9      NA's      :206
## soil_type      source      species      tissue_conc_ugg
## Length:1014      Length:1014      Length:1014      Min.      : 0.00054
## Class :character      Class :character      Class :character      1st Qu.: 0.16908
## Mode  :character      Mode  :character      Mode  :character      Median : 0.52573
##                                     Mean      : 2.51415
##                                     3rd Qu.: 2.06812
##                                     Max.      :72.62672
##
```

Session Information

```
## R version 4.0.2 (2020-06-22)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 10 x64 (build 18363)
##
## Matrix products: default
##
## locale:
## [1] LC_COLLATE=English_United States.1252
## [2] LC_CTYPE=English_United States.1252
## [3] LC_MONETARY=English_United States.1252
## [4] LC_NUMERIC=C
## [5] LC_TIME=English_United States.1252
##
## attached base packages:
## [1] stats      graphics  grDevices  utils      datasets  methods    base
##
## other attached packages:
## [1] tinytex_0.32      anchors_3.0-8      MASS_7.3-51.6      rgenoud_5.8-3.0
## [5] stringr_1.4.0      tidyr_1.1.2        dplyr_1.0.5        knitr_1.31
## [9] kableExtra_1.3.4  reshape2_1.4.4      gridExtra_2.3      ggplot2_3.3.3
##
## loaded via a namespace (and not attached):
## [1] Rcpp_1.0.6      pillar_1.6.0      compiler_4.0.2      plyr_1.8.6
## [5] tools_4.0.2      digest_0.6.27      viridisLite_0.3.0  evaluate_0.14
## [9] lifecycle_1.0.0  tibble_3.1.1      gtable_0.3.0        pkgconfig_2.0.3
## [13] rlang_0.4.10     rstudioapi_0.13    DBI_1.1.1           yaml_2.2.1
## [17] xfun_0.24        xml2_1.3.2         httr_1.4.2          withr_2.4.1
## [21] systemfonts_1.0.2 generics_0.1.0      vctrs_0.3.7         webshot_0.5.2
```

## [25]	grid_4.0.2	tidyselect_1.1.0	svglite_2.0.0	glue_1.4.1
## [29]	R6_2.5.0	fansi_0.4.2	rmarkdown_2.6	purrr_0.3.4
## [33]	magrittr_2.0.1	scales_1.1.1	ellipsis_0.3.1	htmltools_0.5.1.1
## [37]	rvest_0.3.6	assertthat_0.2.1	colorspace_1.4-1	utf8_1.2.1
## [41]	stringi_1.5.3	munsell_0.5.0	crayon_1.4.1	