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**Cross-species integration of single cell RNA-seq data from the primary motor cortex between human and mouse using SATURN**

To perform SATURN, I used the datasets we selected at the beginning of the internship. No big modifications of the `anndata` objects are required, only a filtering on the cells and the genes, adding a species column and a cell type column in the `.obs` dataframe plus putting the names of the genes as the `.var_names` of the `anndata` object. This part on the names is not specified but since the protein embeddings files are from *Ensembl*, the gene names required seems to be HGNC names at least for the human.

I runned several integrations with SATURN, mostly always with the default parameters (HVG = 8000, macrogenes = 2000). I tried first only for the human and mouse again, with the whole dataset (fig. 1), only *GABAergic* (fig. 2), only *Glutamatergic* (fig. 3) and only *Non-Neuronal* (fig. 4). These integrations might need more training, it should be possible to pass in the arguments a file with the *centroids*<sup>1</sup>. Unfortunately, this file is not generated automatically and when I tried, I had a memory issue<sup>2</sup>, even with 150G on the GPU node. By doing that, the *centroids of the macrogenes* should not be calculated from scratch again but improved after each training.

Finally, I tried to perform the integration with the fly dataset as well, unfortunately there is a problem with the gene names<sup>3</sup>. This part of the scripts should be the part that take the gene names from the `anndata` object and match them with the ones in the proteome embeddings files. The index was first of type *CategoricalIndex* that I change to `str` and was instead of type *object*, but it should not be the problem since the mouse dataset also have an index of type *CategoricalIndex* that does not cause any problem. The gene names annotation is from *FlyBase* which seems to be the one use by *Ensembl*. Sadly, I did not have the time to investigate this problem.

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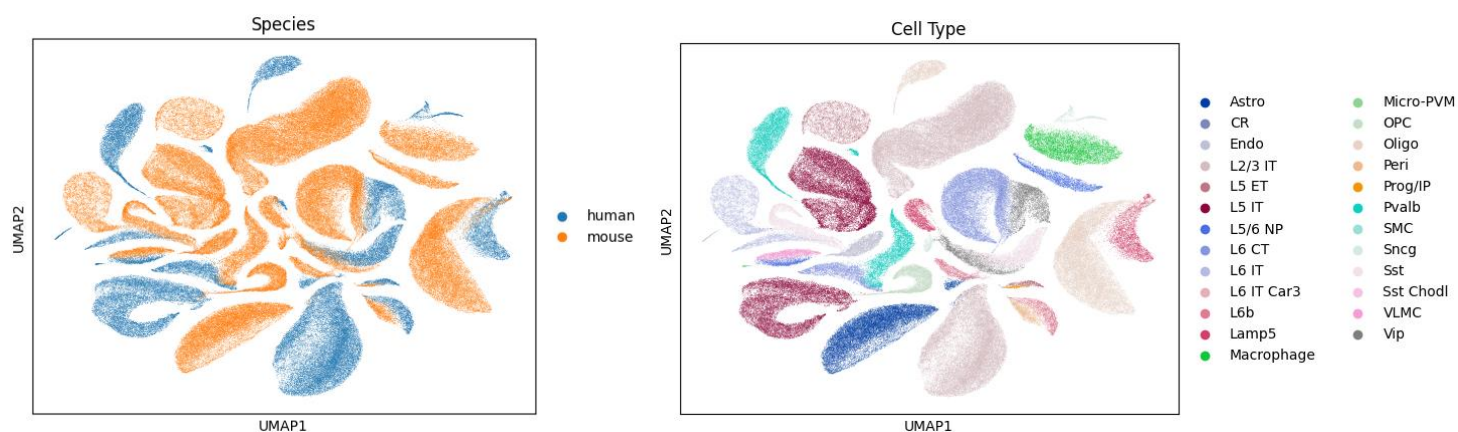
<sup>1</sup> With the command `--centroids_init_path` that either indicates the path of the file or where it should be save when done for the first time

<sup>2</sup> File `"/nfs/research/irene/anaelle/Scripts/SATURN/model/saturn_model.py"`, line 105, in `forward`

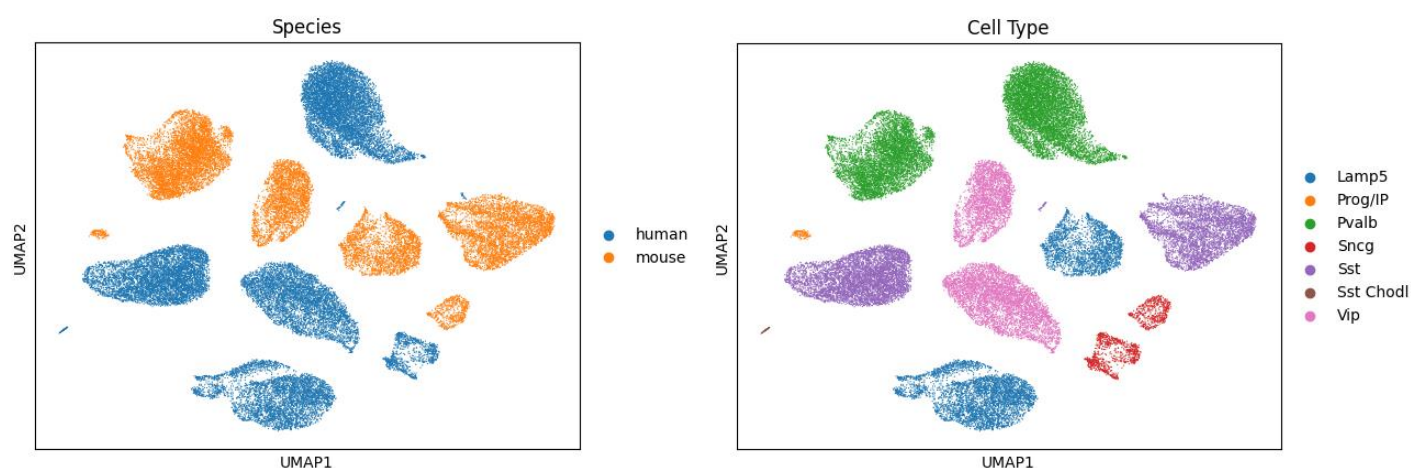
```
expr = torch.zeros(batch_size, self.num_genes).to(inp.device)
RuntimeError: CUDA out of memory. Tried to allocate 166.00 MiB (GPU 0; 31.75 GiB total capacity; 305.90 MiB already allocated; 26.69 MiB free; 330.00 MiB reserved in total by PyTorch) If reserved memory is >> allocated memory try setting max_split_size_mb to avoid fragmentation. See documentation for Memory Management and PYTORCH_CUDA_ALLOC_CONF
```

<sup>3</sup> File `"/nfs/research/irene/anaelle/Scripts/SATURN/data/gene_embeddings.py"`, line 110, in `<setcomp>`

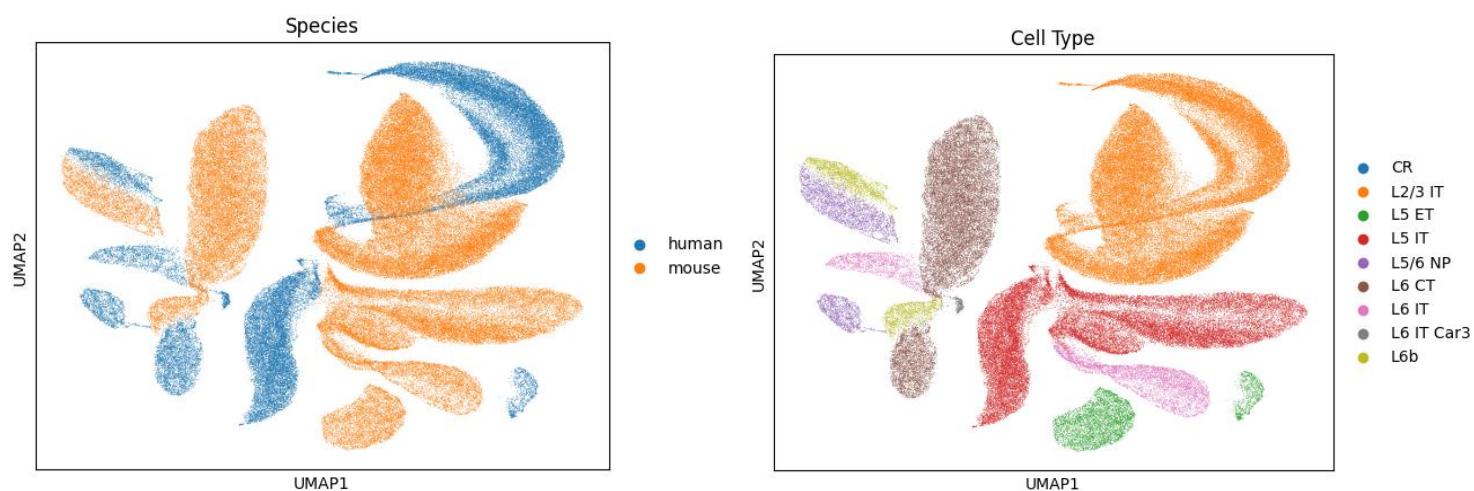
```
genes_to_use = {gene for gene in adata.var_names if gene.lower() in
genes_with_embeddings}
AttributeError: 'float' object has no attribute 'lower'
```



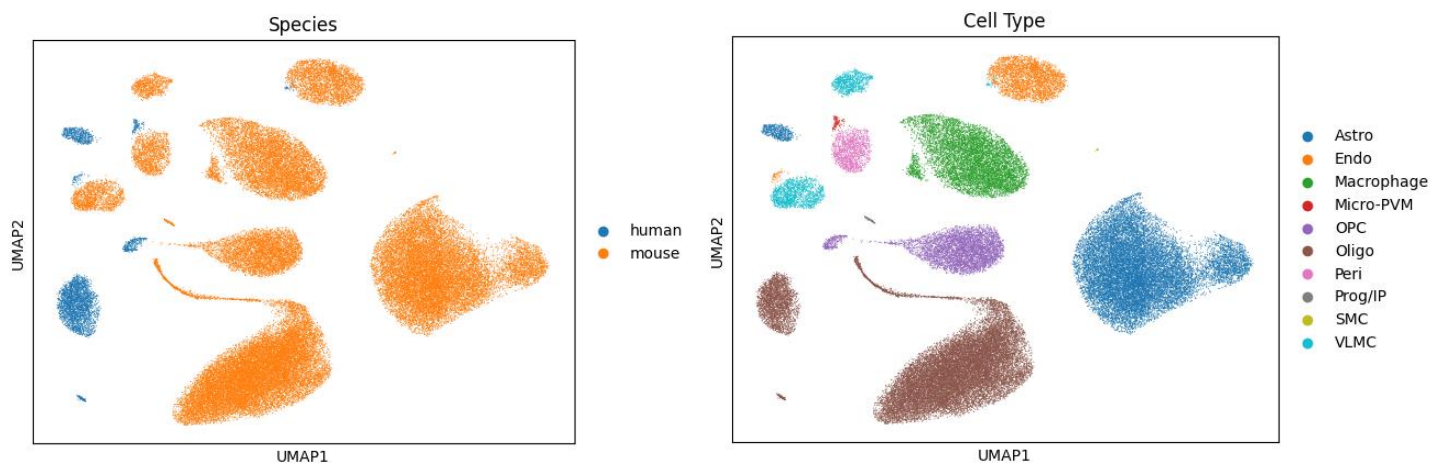
**Figure 1:** UMAP plots after the first training on human and mouse data, using the whole datasets. First plot is colored by species and second plot is colored by cell types.



**Figure 2:** UMAP plots after the second training on human and mouse, only using GABAergic cells. First plot is colored by species and second plot by cell types.



**Figure 3:** UMAP plot of the third training on human and mouse, only using Glutamatergic cells. First plot is colored by species and second plot by cell types.



**Figure 4:** UMAP plot of the fourth training on human and mouse, only using Non-Neuronal cells. First plot is colored by species and second plot by cell types.