# INTRODUCTION

At least 84% of the marine eco-regions have reported the presence of an invasive species (Molnar et al., 2008). These represent a major threat to local biodiversity and the economic activities that depend on it, like tourism or fisheries (Bax et al., 2003). Invasive species may also threaten native species through competition (DAVIS, 2003) or predation. By 2005, the economic cost of invasive species to the United States was estimated at $120 billion per year and nearly 42% of species that have been included in the Endangered or Threatened species list have been labeled as such due to presence of invasive species (Pimentel, Zuniga & Morrison, 2005). This highlights the importance of understanding, managing, and preventing ecological invasions.

Lionfish (*Pterois volitans/miles* complex) are an invasive species in the North-Western Atlantic and the Caribbean, likely introduced through liberation of aquarium-kept organisms (Betancur-R et al., 2011). They are the first marine vertebrates to establish in North Atlantic (Schofield, 2009, 2010) and Caribbean coasts (Sabido-Itza et al., 2016). Lionfish have been widely reported in coral reefs (Aguilar-Perera & Tuz-Sulub, 2010), but also in other habitats such as estuaries (Jud et al., 2011), mangroves (Barbour et al., 2010), areas with hard-bottoms (Muñoz, Currin & Whitfield, 2011), and mesophotic reefs (Andradi-Brown et al., 2017). Due to its threat to local biodiversity, the speed of their spread, and its difficulty of management, their presence in these waters has been labeled as a major marine invasion (Hixon et al., 2016).

A significant amount of research has been done to describe lionfish feeding ecology in North Carolina (Muñoz, Currin & Whitfield, 2011), the Bahamas (Morris & Akins, 2009; Cote et al., 2013), Northern Gulf of Mexico (Dahl & Patterson, 2014), Mexican Caribbean (Valdez-Moreno et al., 2012; Villaseñor-Derbez & Herrera-P’erez, 2014), Belize (Hackerott et al., 2017), and Costa Rica (Sandel et al., 2015). Their feeding behavior and high consumption rates can reduce recruitment (Albins & Hixon, 2008) and population sizes (Green et al., 2012) of native reef-fish species, and further the endangerment of critically endangered reef fish (Rocha et al., 2015). (However, see Hackerott et al. (2017) for a case where there was no evidence that lionfish affected the density, richness, or composition of prey fishes). Major efforts have also been made to understand the possible impacts of the invasion by keeping track of its range through time (Schofield, 2009, 2010) and predicting invasion ranges under climate change scenarios (Grieve, Curchitser & Rykaczewski, 2016). By combining information from these disciplines, researchers have been able to predict the trophic impacts of lionfish (Arias-Gonzalez et al., 2011), which can then be translated into ecosystem-level and economic impacts.

Seeking to reduce lionfish densities, governments and non-profit organizations have promoted removal programs and incentivized its consumption (Chin, Aiken & Buddo, 2016). In some cases, these have shown to significantly reduce -but not quite eliminate- lionfish abundances at local scales (Sandel et al., 2015, Chin, Aiken & Buddo (2016), de Leon et al. (2013)). The rapid recovery rates exhibited by lionfish (Barbour et al., 2011) and the persistent populations in mesophotic coral ecosystems (Andradi-Brown et al., 2017) -which can contribute with recruitment to shallow-water populations- make of complete eradication through fishing effort an unlikely solution. However, further incentivizing its consumption might create a demand big enough to promote and sustain a stable fishery (Chin, Aiken & Buddo, 2016), which can reduce local abundances and control -not eradicate- the invasion while providing alternative livelihoods.

The feasibility of lionfish removal programs has been extensively evaluated through field observations(Sandel et al., 2015, Chin, Aiken & Buddo (2016), de Leon et al. (2013); Usseglio et al., 2017) and empirical modeling (Barbour et al., 2011; Morris, Shertzer & Rice, 2011; Johnston & Purkis, 2015). The latter measure changes in biomass or density (Barbour et al., 2011; Johnston & Purkis, 2015) in response to increased mortality (*i.e.* lionfish removal). In this case, biomass represents the sum of all fish's individual weight. Total Weight (TW) can be estimated from Total Length (TL) observations using the allometric growth equation (). Parameters and for this equation exist for North Carolina (Barbour et al., 2011), Northern (Fogg et al., 2013) and Southern Gulf of Mexico (Aguilar-Perera & Quijano-Puerto, 2016), the Southern Mexican Caribbean (Sabido-Itza et al., 2016), Little Cayman (Edwards, Frazer & Jacoby, 2014), Jamaica (Chin, Aiken & Buddo, 2016), Bonaire (de Leon et al., 2013) and Costa Rica (Sandel et al., 2015), but remain unavailable for the central Mexican Caribbean. The weight-at-length of a species can vary across regions as a response to biotic (*e.g.* local food availability) and abiotic (*e.g.* water temperature) conditions (Johnson & Swenarton, 2016). Thus, when using biomass-informed models or estimating biomass from length observations, it is important to use site-specific parameters to obtain an accurate estimate. This is especially important when research involves identifying the total biomass available for harvest by fishers (Chin, Aiken & Buddo, 2016) or the efficacy of lionfish removals (Barbour et al., 2011; Morris, Shertzer & Rice, 2011; Johnston & Purkis, 2015).

Here, I provide the first allometric growth parameters for the invasive lionfish in the central Mexican Caribbean region. At the same time, I highlight the importance of using site-specific parameters by estimating biomass with parameters from other regions across the invasion range and comparing them to observed biomass. I also provide other 13 standardized parameters from eight studies through the invasion range, making them readily accessible for future research. Finally, I discuss the way in which allometric parameters are reported, and call for standardization to facilitate their use.

# MATERIALS AND METHODS

**Area of study.** The study took place off the coasts of Playa del Carmen, in the Mexican Caribbean (**Fig. 1**). The region represents the northernmost section of the Mesoamerican Barrier Reef System (Ruiz-Zarate & Arias-Gonzalez, 2004). Coral reefs and mangroves are locally important habitats that represent important sources of income in terms of extractive (*e.g.* recreational fishing) and non-extractive (*e.g.* SCUBA diving) activities related to tourism, the main source of income to the local economy (Murray, 2007).

In the region, the reef profile has been described by Arias-Gonzalez (1998), indicating that the reef lagoon extends about 500 m from the coast, until the reef crest is reached. The reef becomes deeper, leading to the reef front often found at 700 m from the coastline end extends for an additional 300 m. At approximately 1000 m away from shore and 30 - 40 m depth, the reef leads to a drop-off. Along a perpendicular profile to the coast, bands of reef are interrupted by sand patches at 8 - 12 m deep and 16-18 m deep. Along the coast, these reefs have been reported to be under significant anthropogenic pressure, likely causing a shift in structure and function (Bozec et al., 2008).

**Fish sampling.** Organisms sampled by Villaseñor-Derbez & Herrera-P’erez (2014) for stomach content analysis between May and August of 2010 were used in this work. A total of 33 SCUBA immersions were performed in 10 sampling sites along the coast (Fig. 1, Table 1). Sampling locations included wall and carpet reefs at depths between 5.7 m and 38.1 m. All observed organisms (n = 109) were collected using hand nets and numbered collection bottles. The use of hand nets prevents any weight loss due to bleeding(when collected with spears), and increases the range of sizes that can be sampled, providing a representative sample of the population. Information on depth and other comments were recorded in an underwater slide. Depth was recorded by dive gauges held by divers as safety procedures during the collections. Samples were frozen within 30 minutes of completing the dive and stored for posterior analysis in the lab. For every organism Total Length (TL; mm) and Total Weight (TW; gr) were recorded in the lab.

**Data analysis.** The weight at length relationship between the observed variables is described by the allometric growth function:

(1)

Where is the Total Weight (gr), is the observed Total Length (mm), is the ponderal index and is the scaling exponent or allometric parameter. When , it is said that the organism exhibits a perfect isometric growth. The dependent and independent variables were transformed via base-10 logarithms so that the equation is then:

(2)

To simplify this equation, we can re-write it as:

(3)

Where , , , and . Since , we will only use throughout the paper for simplicity. The coefficients ( and ) were estimated with an Ordinary Least Square Regression and heteroskedastic-robust standard errors. Both coefficients were tested against the null hypothesis of no change (*i.e.* and ). Additionally, the allometric parameter was tested against the null hypothesis of isometric growth (). Coefficients were tested with a two-tailed Student's t-test. The significance of the regression was corroborated with an F-test.

Other allometric growth parameters were obtained from peer-reviewed literature. Parameters were obtained for North Carolina (Barbour et al., 2011), Northern (Fogg et al., 2013) and Southern Gulf of Mexico (Aguilar-Perera & Quijano-Puerto, 2016), the Southern Mexican Caribbean (Sabido-Itza et al., 2016), Little Cayman (Edwards, Frazer & Jacoby, 2014), Jamaica (Chin, Aiken & Buddo, 2016), Bonaire (de Leon et al., 2013) and Costa Rica (Sandel et al., 2015). Additionally, parameters from Fishbase (Froese & Pauly, 2016) were also included. When available, information on sampling methods, gender differentiation, location, and depth ranges of each study was retrieved. Whenever gender was not specified, it was assumed that the results were presented for both genders pooled together. During the review process, it was noticed that some papers indistinctly use to report either the ponderal index in eq. 1 or the y-intercept () in eq. 3, which might sometimes be overlooked. Furthermore, some studies report their parameters as mm-to-gr conversions, but a rapid evaluation of such parameters indicates that they were estimated as cm-to-gr conversions. Here, all parameters are reported as TL(mm) to TW(gr) conversions. When required, values from other studies are transformed for consistency.

Since standard errors or confidence intervals were not provided for many studies, it was impossible to test for statistical differences between coefficients. Thus expected biomass for each organism was calculated with the growth parameters estimated in this study, as well as those retrieved from additional literature. All expected biomass values were divided by the observed biomass to obtain a ratio that allowed rapid identification of over- or underestimation with respect to the observed biomass. Therefore, values lower than 1 indicate that the parameters underestimate biomass, and values larger than 1 indicate overestimation. Median and mean expected to observed biomass ratios were calculated for each study. Difference in mean biomass ratios across studies were tested with an two-tailed, one-way Analysis of Variance (ANOVA).

All hypothesis testing was performed with a confidence level of in R version 3.4.0 (R Core Team, 2017). Data wrangling was performed with the tidyverse package (Wickham, 2017). Maps were created with a mix of functions from the sp (Pebesma & Bivand, 2005), rgdal (Bivand, Keitt & Rowlingson, 2017), tmap (Tennekes, 2017a), and tmaptools (Tennekes, 2017b) packages. Calculation of heteroskedastic-robust standard errors was done with the sandwich (Zeileis, 2004) and lmtest (Zeileis & Hothorn, 2002) packages. Models were manipulated with with the broom package (Robinson, 2017). Stargazer (Hlavac, 2015), knitr (Xie, 2017), and kableExtra (Zhu, 2017) were used to produce the tables, and RefManageR (McLean, 2014) was used to keep track of citations. The manuscript was written in r markdown (Allaire et al., 2017) and processed with the knitr package (Xie, 2017). Raw data and code used in this work is available at [github.com/jcvdav/lionfish\_biometry](file:///C:\Users\JC\Documents\GitHub\lionfish_biometry\github.com\jcvdav\lionfish_biometry).

# RESULTS

Organism TL ranged between 34 and 310 mm and TW between 0.3 and 397.7 gr. The smallest organism (TL = 34.00 mm) was also the lightest organism (TW = 0.30 gr). However, the largest organism (TL = 310.00 mm) was not the heaviest (TW = 303.70 gr), and the heaviest organism (TW = 397.70 gr) was 292.00 mm in total length. Kernell density plots (Fig. 2) show the distribution for TL and TW. Both measures were positively skewed, with skewness of 0.87 for TL and 2.25 for TW.

**Length-weight relationship.** The model adjusted to eq. 3 estimated the coefficient values at and . Thus, TW (gr) can be calculated from TL (mm) as a linear equation: or its exponential form: . The intercept () and slope were significantly different from zero ( and , respectively), rejecting the null hypothesis of no change. Additionally, the allometric factor () was significantly different from the value of isometric growth of (), indicating that lionfish present allometric growth. More information on model fit and confidence intervals for the estimated coefficients is presented in Table 2. The relationship between Total Length and Total Weight is presented in Figure 3.

**Comparison of allometric parameters.** From the eight peer-reviewed studies including information on growth parameters for *P. volitans* and Fishbase (Froese & Pauly, 2016), 13 parameters were identified (Table 3). Two studies (Fogg et al., 2013; Aguilar-Perera & Quijano-Puerto, 2016) reported gender-level and pooled parameters, while the rest of the studies always presented pooled results. The smallest coefficient of determination was presented by Chin, Aiken & Buddo (2016) with , while Sabido-Itza et al. (2016) reported the highest value at . These studies presented information for organisms obtained at depths between 0.5 and 57 m. Two studies (Aguilar-Perera & Quijano-Puerto, 2016; Chin, Aiken & Buddo, 2016) explicitly stated that their organisms were sampled with pole spears. Five studies (Barbour et al., 2011; Fogg et al., 2013; Edwards, Frazer & Jacoby, 2014; Sandel et al., 2015; Sabido-Itza et al., 2016) mentioned that some of their organisms were obtained with pole spears (or other type of harpoon). A single study (de Leon et al., 2013) did not specify how samples were obtained.

Parameters from models fit to males or females exclusively tend to have a higher steepness (*i.e.* higher allometric parameter), with mean standard deviation values of and males and females respectively, compared to parameters from models for pooled genders with a mean standard deviation value of . In the case of the ponderal index () and its transformed parameter (), values were higher for parameters for pooled genders. Figure 4 shows the predicted weights for organisms within the size range of these study using the 14 parameters previously described.

From all allometric parameters reviewed, those of Edwards, Frazer & Jacoby (2014) slightly underestimated the observed biomass, with median (0.96) and mean (0.98) bellow one. There is no significance in comparing the parameters estimated in this study since, by definition, they are the pair of parameters for which residual sum of squares was minimized with values of median =1.00 and mean =1.03. For all the other studies, the 95% confidence interval around the mean fell further away from a value of 1, indicating overestimation. Predicted to observed biomass ratios are presented in Figure 5.

# DISCUSSION

This study provides the first pair of allometric growth parameters specific to the Central Mexican Caribbean, complementing other studies performed in Mexican waters in the Alacranes Reef (Aguilar-Perera & Quijano-Puerto, 2016) and Xcalak National Park (Sabido-Itza et al., 2016). By using hand nets instead of spears, we are able to sample a wider range of sizes often ignored by pole spear samples, allowing us to include smaller organisms. Estimating parameters by including smaller organisms ensures better estimatio of weight for smaller sizes. This is especially important when biomass is calculated from visual census, where small organisms can be registered. Thus, this study is the first to provide allometric growth parameters for lionfish in the central region of the Mexican Caribbean and increases certainty in weight estimation of small organisms.

The parameters estimated in this study were within the range of studies in other regions (Barbour et al., 2011; de Leon et al., 2013; Fogg et al., 2013; Edwards, Frazer & Jacoby, 2014; Sandel et al., 2015; Aguilar-Perera & Quijano-Puerto, 2016; Chin, Aiken & Buddo, 2016; Sabido-Itza et al., 2016). As compared to other parameters, the ones presented here tend to provide lower weight estimates for a same length. Until about TL = 200 mm, there are no appreciable differences between the parameters described in this study and those for little Cayman (Edwards, Frazer & Jacoby, 2014) and Jamaica (Chin, Aiken & Buddo, 2016). Yet, for larger organisms (TL > 270 mm) parameters from Costa Rica (Sandel et al., 2015) and Bonaire (de Leon et al., 2013) provide similar estimates to those from this study. Conversely, these same studies tend to overestimate weight -as compared to the ones reported here- for smaller organisms, likely due to the lack of small organisms in the samples used to estimate their parameters. When ever possible, future works should consider the use of hand nets to obtain the samples not only for studies on weight-at-length, but also diet, behavior and life history, where length can be an important factor.

There are evident differences in weight-at-length between organisms from the Caribbean and Gulf of Mexico / North-Western Atlantic. Weight estimates with parameters from the Gulf of Mexico and North-Western Atlantic (Barbour et al., 2011; Fogg et al., 2013; Aguilar-Perera & Quijano-Puerto, 2016; Sabido-Itza et al., 2016) tend to be higher than those from the Caribbean (de Leon et al., 2013; Edwards, Frazer & Jacoby, 2014; Sandel et al., 2015; Chin, Aiken & Buddo, 2016), except for the ones from Xcalak National Park, Mexico (Sabido-Itza et al., 2016). This indicates that there are differences between lionfish across the invasion range, pattern that has also been reported for age and growth parameters (Fogg et al., 2015). These differences can have major implications in management, especially when estimating biomass available for harvest or predicting effects on local ecosystems, or evaluating the effectiveness of removal programs. Using site-specific values provides a more accurate estimate of fish biomass. Future research should try to use, to the extent possible, parameters calculated for their region, or use different parameters to provide upper and lower bounds in their results. At the same time, this highlights the need for more basic research. To have a better invasion management, we must perform research that can describe biologically important information of lionfish throughout its invasion range (Johnson & Swenarton, 2016).

While performing the literature review, it was often unclear if parameters were presented for eq.1 or eq. 3. Sometimes, they were even mislabeled and yielded senseless results when using the suggested conversion equation. On some others, parameters were said to be reported for mm to gr conversions, when they were actually reported as cm to gr conversions. Perhaps these minor discrepancies can be easily solved by any researcher, but why should they exist in the first place? It is important that we report our information in a standard way, making it readily available for other researchers and managers. In this particular case, I provide my humble opinion through 5 guidelines to report allometric parameters:

1. Be explicit in the methods section. What may seem obvious to you as an author -because you have been deeply immersed throughout the process- may not be clear to the reader. Specify any transformation performed on the data. When using log-transformations, mention the base used to transform. Do not assume that "data were log-transformed" means . These assumptions vary across disciplines and software and can be a source of confusion. For example, in biology we often assume "log-transformed" indicates the use of base 10, however in R the proper command is log10() and not log(), which uses base .
2. Use mm and gr to measure TL and TW, respectively. While conversion is always possible, we should aim at using standard units to report these parameters. If you prefer to use cm to gr conversions, that is certainly valid, but make sure to explicitly mention units when presenting the parameters.
3. Specify the equation for which parameters are presented by including an explicit example with the parameters substituted into it, as done by some of the papers reviewed (de Leon et al., 2013; Sandel et al., 2015; Chin, Aiken & Buddo, 2016; Sabido-Itza et al., 2016). If possible, present the parameters in their exponential (eq. 1) and linear (eq. 3) forms.
4. Report standard errors and/or confidence intervals around the obtained estimates. Given that small changes in , , and can result in important changes in estimated weight, it is important that we report uncertainty around each parameter and not just general model fit and significance. Reporting uncertainty around parameters allows researchers and managers to include upper and lower bounds in their predictions.
5. Make your data -and code- available. Even if this is not requited by the journal or publisher, you can use free cloud data storage services or third-party repositories to make your research accessible to others. Resources will always be limited and budget will never be enough. It is important that we take advantage of open science tools that promote the advancement of knowledge and foster collaboration. Ultimately, this promotes transparency allows replicability of research and advances science.

As final conclusion statements 1) A new pair of allometric growth parameters are provided for lionfish from the central Mexican Caribbean, where they exhibit different weight-at-length, as compared to other regions. 2) Given the differences in weight-at-length patterns through the invasion range, it is important to use site-specific parameters. 3) When possible, collect organisms with hand nets to obtain a representative sample of the population that includes small organisms. 4) Report allometric parameters (and other life history parameters) in a way that is readily available for non-experts, and make your research available to advance science.

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# REFERENCES

Aguilar-Perera, A. & Quijano-Puerto, L. 2016. Relations between fish length to weight, and otolith length and weight, of the lionfish pterois volitans in the parque nacional arrecife alacranes, southern gulf of mexico. *Rev. biol. mar. oceanogr.* 51(2):469–474. DOI: [10.4067/S0718-19572016000200025](https://doi.org/10.4067/S0718-19572016000200025).

Aguilar-Perera, A. & Tuz-Sulub, A. 2010. Non-native, invasive red lionfish (pterois volitans [linnaeus, 1758]: Scorpaenidae), is first recorded in the southern gulf of mexico, off the northern yucatan peninsula, mexico. *AI*. 5(Supplement 1):S9–S12. DOI: [10.3391/ai.2010.5.S1.003](https://doi.org/10.3391/ai.2010.5.S1.003).

Albins, M. & Hixon, M. 2008. Invasive indo-pacific lionfish pterois volitans reduce recruitment of atlantic coral-reef fishes. *Mar. Ecol. Prog. Ser.* 367:233–238. DOI: [10.3354/meps07620](https://doi.org/10.3354/meps07620).

Allaire, J., Cheng, J., Xie, Y., McPherson, J., Chang, W., Allen, J., Wickham, H., Atkins, A., et al. 2017. *Rmarkdown: Dynamic documents for r*. ed. (nos.). Available: <https://CRAN.R-project.org/package=rmarkdown>.

Andradi-Brown, D.A., Grey, R., Hendrix, A., Hitchner, D., Hunt, C.L., Gress, E., Madej, K., Parry, R.L., et al. 2017. Depth-dependent effects of culling-do mesophotic lionfish populations undermine current management? *R Soc Open Sci*. 4(5):170027. DOI: [10.1098/rsos.170027](https://doi.org/10.1098/rsos.170027).

Arias-Gonzalez, J.E. 1998. Trophic models of protected and unprotected coral reef ecosystems in the south of the mexican caribbean. *J Fish Biol*. 53(sa):236–255. DOI: [10.1111/j.1095-8649.1998.tb01030.x](https://doi.org/10.1111/j.1095-8649.1998.tb01030.x).

Arias-Gonzalez, J.E., Gonzalez-Gandara, C., Luis Cabrera, J. & Christensen, V. 2011. Predicted impact of the invasive lionfish pterois volitans on the food web of a caribbean coral reef. *Environ Res*. 111(7):917–925. DOI: [10.1016/j.envres.2011.07.008](https://doi.org/10.1016/j.envres.2011.07.008).

Barbour, A., Montgomery, M., Adamson, A., D?az-Ferguson, E. & Silliman, B. 2010. Mangrove use by the invasive lionfish pterois volitans. *Mar. Ecol. Prog. Ser.* 401:291–294. DOI: [10.3354/meps08373](https://doi.org/10.3354/meps08373).

Barbour, A.B., Allen, M.S., Frazer, T.K. & Sherman, K.D. 2011. Evaluating the potential efficacy of invasive lionfish (pterois volitans) removals. *PLoS ONE*. 6(5):e19666. DOI: [10.1371/journal.pone.0019666](https://doi.org/10.1371/journal.pone.0019666).

Bax, N., Williamson, A., Aguero, M., Gonzalez, E. & Geeves, W. 2003. Marine invasive alien species: A threat to global biodiversity. *Marine Policy*. 27(4):313–323. DOI: [10.1016/S0308-597X(03)00041-1](https://doi.org/10.1016/S0308-597X(03)00041-1).

Betancur-R, R., Hines, A., Acero, A., Orti, G., Wilbur, A. & Freshwater, D. 2011. Reconstructing the lionfish invasion: Insights into greater caribbean biogeography. *J Biogeography*. 38:1281–1293. DOI: [10.1111/j.1365-2699.2011.02496.x](https://doi.org/10.1111/j.1365-2699.2011.02496.x).

Bivand, R., Keitt, T. & Rowlingson, B. 2017. *Rgdal: Bindings for the geospatial data abstraction library*. ed. (nos.). Available: <https://CRAN.R-project.org/package=rgdal>.

Bozec, Y., Acosta-Gonz’alez, G., N’uñez-Lara, E. & Arias-Gonz’alez, J. 2008. Impacts of coastal development on ecosystem structure and function of yucatan coral reefs, mexico. In *Proceedings of the 11th international coral reef symposium*. ed. I.C.R.S. ICRF, Ed. (nos.). Ft. Lauderdale, Florida: 11th International Coral Reef Symposium.

Chin, D.A., Aiken, K.A. & Buddo, D. 2016. Lionfish population density in discovery bay, jamaica. *International Journal of Scientific & Engineering Research*. 7(12):1327–1331.

Cote, I., Green, S., Morris, J., Akins, J. & Steinke, D. 2013. Diet richness of invasive indo-pacific lionfish revealed by dna barcoding. *Mar. Ecol. Prog. Ser.* 472:249–256. DOI: [10.3354/meps09992](https://doi.org/10.3354/meps09992).

Dahl, K.A. & Patterson, W.F. 2014. Habitat-specific density and diet of rapidly expanding invasive red lionfish, pterois volitans, populations in the northern gulf of mexico. *PLoS ONE*. 9(8):e105852. DOI: [10.1371/journal.pone.0105852](https://doi.org/10.1371/journal.pone.0105852).

DAVIS, M.A. 2003. Biotic globalization: Does competition from introduced species threaten biodiversity? *Bioscience*. 53(5):481. DOI: [10.1641/0006-3568(2003)053[0481:BGDCFI]2.0.CO;2](https://doi.org/10.1641/0006-3568(2003)053%5b0481:BGDCFI%5d2.0.CO;2).

de Leon, R., Vane, K., Bertuol, P., Chamberland, V.C., Simal, F., Imms, E. & Vermeij, M.J.A. 2013. Effectiveness of lionfish removal efforts in the southern caribbean. *Endanger Species Res*. 22(2):175–182. DOI: [10.3354/esr00542](https://doi.org/10.3354/esr00542).

Edwards, M.A., Frazer, T.K. & Jacoby, C.A. 2014. Age and growth of invasive lionfish (pterois spp.) in the caribbean sea, with implications for management. *BMS*. 90(4):953–966. DOI: [10.5343/bms.2014.1022](https://doi.org/10.5343/bms.2014.1022).

Fogg, A.Q., Evans, J.T., Ingram JR, G.W., Peterson, M.S. & Brown-Peterson, N.J. 2015. Comparing age and growth patterns of invasive lionfish among three ecoregions of the northern gulf of mexico. In *Proceedings of the 68 th gulf and caribbean fisheries institute*. ed. G. GCFI & C.F. Institute, Eds. (nos.). Panama City: Gulf; Caribbean Fisheries Institute.

Fogg, A.Q., Hoffmayer, E.R., Driggers, W.B., Campbell, M.D., Pellegrin, G.J. & Stein, W. 2013. Distribution and length frequency of invasive lionfish (pterois sp.) in the northern gulf of mexico. *GCR*. 25. DOI: [10.18785/gcr.2501.08](https://doi.org/10.18785/gcr.2501.08).

Froese, R. & Pauly, D. 2016. Available: <http://www.fishbase.org/> [2016, December 15].

Green, S.J., Akins, J.L., Maljkovi’c, A. & Côt’e, I.M. 2012. Invasive lionfish drive atlantic coral reef fish declines. *PLoS ONE*. 7(3):e32596. DOI: [10.1371/journal.pone.0032596](https://doi.org/10.1371/journal.pone.0032596).

Grieve, B., Curchitser, E. & Rykaczewski, R. 2016. Range expansion of the invasive lionfish in the northwest atlantic with climate change. *Mar. Ecol. Prog. Ser.* 546:225–237. DOI: [10.3354/meps11638](https://doi.org/10.3354/meps11638).

Hackerott, S., Valdivia, A., Cox, C.E., Silbiger, N.J. & Bruno, J.F. 2017. Invasive lionfish had no measurable effect on prey fish community structure across the belizean barrier reef. *PeerJ*. 5:e3270. DOI: [10.7717/peerj.3270](https://doi.org/10.7717/peerj.3270).

Hixon, M., Green, S., Albins, M., Akins, J. & Morris, J. 2016. Lionfish: A major marine invasion. *Mar. Ecol. Prog. Ser.* 558:161–165. DOI: [10.3354/meps11909](https://doi.org/10.3354/meps11909).

Hlavac, M. 2015. *Stargazer: Well-formatted regression and summary statistics tables*. ed. (nos.). Cambridge, USA: Harvard University. Available: <http://CRAN.R-project.org/package=stargazer>.

Johnson, E.G. & Swenarton, M.K. 2016. Age, growth and population structure of invasive lionfish (pterois volitans/miles) in northeast florida using a length-based, age-structured population model. *PeerJ*. 4:e2730. DOI: [10.7717/peerj.2730](https://doi.org/10.7717/peerj.2730).

Johnston, M. & Purkis, S. 2015. A coordinated and sustained international strategy is required to turn the tide on the atlantic lionfish invasion. *Mar. Ecol. Prog. Ser.* 533:219–235. DOI: [10.3354/meps11399](https://doi.org/10.3354/meps11399).

Jud, Z., Layman, C., Lee, J. & Arrington, D. 2011. Recent invasion of a florida (usa) estuarine system by lionfish pterois volitans / p. miles . *Aquat. Biol.* 13(1):21–26. DOI: [10.3354/ab00351](https://doi.org/10.3354/ab00351).

McLean, M.W. 2014. *Straightforward bibliography management in r using the refmanager package*. ed. (nos.). Available: <http://arxiv.org/abs/1403.2036>.

Molnar, J.L., Gamboa, R.L., Revenga, C. & Spalding, M.D. 2008. Assessing the global threat of invasive species to marine biodiversity. *Frontiers in Ecology and the Environment*. 6(9):485–492. DOI: [10.1890/070064](https://doi.org/10.1890/070064).

Morris, J.A. & Akins, J.L. 2009. Feeding ecology of invasive lionfish (pterois volitans) in the bahamian archipelago. *Environ. Biol. Fishes*. 86(3):389–398. DOI: [10.1007/s10641-009-9538-8](https://doi.org/10.1007/s10641-009-9538-8).

Morris, J.A., Shertzer, K.W. & Rice, J.A. 2011. A stage-based matrix population model of invasive lionfish with implications for control. *Biol Invasions*. 13(1):7–12. DOI: [10.1007/s10530-010-9786-8](https://doi.org/10.1007/s10530-010-9786-8).

Muñoz, R., Currin, C. & Whitfield, P. 2011. Diet of invasive lionfish on hard bottom reefs of the southeast usa: Insights from stomach contents and stable isotopes. *Mar. Ecol. Prog. Ser.* 432:181–193. DOI: [10.3354/meps09154](https://doi.org/10.3354/meps09154).

Murray, G. 2007. Constructing paradise: The impacts of big tourism in the mexican coastal zone. *Coastal Management*. 35(2-3):339–355. DOI: [10.1080/08920750601169600](https://doi.org/10.1080/08920750601169600).

Pebesma, E.J. & Bivand, R.S. 2005. Classes and methods for spatial data in R. *R News*. 5(2):9–13. Available: <https://CRAN.R-project.org/doc/Rnews/>.

Pimentel, D., Zuniga, R. & Morrison, D. 2005. Update on the environmental and economic costs associated with alien-invasive species in the united states. *Ecological Economics*. 52(3):273–288. DOI: [10.1016/j.ecolecon.2004.10.002](https://doi.org/10.1016/j.ecolecon.2004.10.002).

R Core Team. 2017. *R: A language and environment for statistical computing*. ed. (nos.). Vienna, Austria: R Foundation for Statistical Computing. Available: <https://www.R-project.org/>.

Robinson, D. 2017. *Broom: Convert statistical analysis objects into tidy data frames*. ed. (nos.). Available: <https://CRAN.R-project.org/package=broom>.

Rocha, L.A., Rocha, C.R., Baldwin, C.C., Weigt, L.A. & McField, M. 2015. Invasive lionfish preying on critically endangered reef fish. *Coral Reefs*. 34(3):803–806. DOI: [10.1007/s00338-015-1293-z](https://doi.org/10.1007/s00338-015-1293-z).

Ruiz-Zarate, M. & Arias-Gonzalez, J. 2004. Spatial study of juvenile corals in the northern region of the mesoamerican barrier reef system (mbrs). *Coral Reefs*. (September, 9). DOI: [10.1007/s00338-004-0420-z](https://doi.org/10.1007/s00338-004-0420-z).

Sabido-Itza, M., Medina-Quej, A., De Jesus-Navarrete, A., Gomez-Poot, J. & Garcia-Rivas, M. 2016. Uso de la estructura de tallas como evidencia del establecimiento poblacional del pez le?n pterois volitans (scorpaeniformes: Scorpaenidae) en el sur del caribe mexicano. *RBT*. 64(1):353. DOI: [10.15517/rbt.v64i1.18943](https://doi.org/10.15517/rbt.v64i1.18943).

Sandel, V., Mart’inez-Fern’andez, D., Wangpraseurt, D. & Sierra, L. 2015. Ecology and management of the invasive lionfish pterois volitans/miles complex (perciformes: Scorpaenidae) in southern costa rica. *Rev Biol Trop*. 63(1):213–221. Available: <http://www.ncbi.nlm.nih.gov/pubmed/26299126> [2017, June 27].

Schofield, P. 2009. Geographic extent and chronology of the invasion of non-native lionfish (pterois volitans [linnaeus 1758] and p. miles [bennett 1828]) in the western north atlantic and caribbean sea. *AI*. 4(3):473–479. DOI: [10.3391/ai.2009.4.3.5](https://doi.org/10.3391/ai.2009.4.3.5).

Schofield, P. 2010. Update on geographic spread of invasive lionfishes (pterois volitans [linnaeus, 1758] and p. miles [bennett, 1828]) in the western north atlantic ocean, caribbean sea and gulf of mexico. *AI*. 5(Supplement 1):S117–S122. DOI: [10.3391/ai.2010.5.S1.024](https://doi.org/10.3391/ai.2010.5.S1.024).

Tennekes, M. 2017a. *Tmap: Thematic maps*. ed. (nos.). Available: <https://CRAN.R-project.org/package=tmap>.

Tennekes, M. 2017b. *Tmaptools: Thematic map tools*. ed. (nos.). Available: <https://CRAN.R-project.org/package=tmaptools>.

Usseglio, P., Selwyn, J.D., Downey-Wall, A.M. & Hogan, J.D. 2017. Effectiveness of removals of the invasive lionfish: How many dives are needed to deplete a reef? *PeerJ*. 5:e3043. DOI: [10.7717/peerj.3043](https://doi.org/10.7717/peerj.3043).

Valdez-Moreno, M., Quintal-Lizama, C., G’omez-Lozano, R. & Garc’ia-Rivas, M.D.C. 2012. Monitoring an alien invasion: DNA barcoding and the identification of lionfish and their prey on coral reefs of the mexican caribbean. *PLoS ONE*. 7(6):e36636. DOI: [10.1371/journal.pone.0036636](https://doi.org/10.1371/journal.pone.0036636).

Villaseñor-Derbez, J.C. & Herrera-P’erez, R. 2014. Brief description of prey selectivity and ontogenetic changes in the diet of the invasive lionfish pterois volitans (actinopterygii, scorpaenidae) in the mexican caribbean. *PANAMJAS*. 9(2):131–135.

Wickham, H. 2017. *Tidyverse: Easily install and load ’tidyverse’ packages*. ed. (nos.). Available: <https://CRAN.R-project.org/package=tidyverse>.

Xie, Y. 2017. *Knitr: A general-purpose package for dynamic report generation in r*. ed. (nos.). Available: <http://yihui.name/knitr/>.

Zeileis, A. 2004. Econometric computing with hc and hac covariance matrix estimators. *Journal of Statistical Software*. 11(10):1–17. Available: <http://www.jstatsoft.org/v11/i10/>.

Zeileis, A. & Hothorn, T. 2002. Diagnostic checking in regression relationships. *R News*. 2(3):7–10. Available: <https://CRAN.R-project.org/doc/Rnews/>.

Zhu, H. 2017. *KableExtra: Construct complex table with ’kable’ and pipe syntax*. ed. (nos.). Available: <https://CRAN.R-project.org/package=kableExtra>.

# TABLES

Table 1: Coordinates, minimum, maximum and mean depth (m), and number of samples for each location.

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| Location | Lat. | Long. | Min. Depth | Max. Depth | Mean Depth | n |
| Canones | 20.477 | -87.233 | 15.0 | 31.2 | 21.6 | 11 |
| Castillo | 20.496 | -87.220 | 12.5 | 30.5 | 27.5 | 18 |
| Cuevitas | 20.478 | -87.244 | 7.4 | 12.8 | 11.2 | 4 |
| Islas | 20.490 | -87.228 | 14.0 | 19.4 | 16.7 | 10 |
| Paamul | 20.513 | -87.192 | 9.9 | 22.7 | 15.5 | 31 |
| Paraiso | 20.484 | -87.226 | 9.4 | 38.1 | 17.7 | 16 |
| Pared | 20.502 | -87.212 | 12.1 | 21.0 | 16.3 | 12 |
| Pedregal | 20.507 | -87.204 | 14.4 | 14.9 | 14.7 | 3 |
| Santos | 20.493 | -87.222 | 5.7 | 26.6 | 16.2 | 2 |
| Tzimin-Ha | 20.393 | -87.307 | 21.2 | 24.6 | 22.9 | 2 |
| Total |  |  | 5.7 | 38.1 | 18.6 | 109 |

Table 2: Regression table for the linear model fit between log10-transformed Total Weight (dependent variable) and Total Length (independent variable). Numbers in parentheses next to coefficient estimates indicate heteroskedastic-robust standard errors. The asterisks (\*) indicate the statistical significance. 95% Confidence intervals are provided for each parameter.

|  |  |
| --- | --- |
|  | |
|  | *Dependent variable:* |
|  |  |
|  | log10(TW) |
|  | |
| c | -5.494 (0.083)\*\*\* |
| b | 3.235 (0.039)\*\*\* |
|  | |
| 95% CI for c | (-5.657--5.331) |
| 85% CI for b | (3.159-3.311) |
| F Statistic | 6928.67\*\*\* (df = 1; 107) |
| Observations | 109 |
| Adjusted R2 | 0.976 |
| Residual Std. Error | 0.096 (df = 107) |
|  | |
| *Note:* | \*p<0.1; \*\*p<0.05; \*\*\*p<0.01 |

Table 3: Summary of 13 allometric growth parameters available for lionfish in the invaded range from eight peer-reviewed papers, Fishbase (Froese & Pauly, 2016), and this study. All parameters have been adjusted to convert from millimeters to grams. n = Sample size, a = scaling parameter for eq. 1, c = y-intercept for eq. 3, b = exponent or slope for eq. 1 or eq. 3, respectively. The R2 column indicates reported model fit. mDepth and MDepth indicate minimum and maximum depths (m), respectively, at which organisms were sampled. The Spear column indicates if the study collected organisms with pole spears. An asterisk (\*) indicates that some portion of organisms were sampled with pole spears.

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Reference | Sex | a | c | b | R2 | n | mDepth | MDepth | Spear |
| 1 | Both | 472 | 0 | -5.5400 | 3.3000 | 0.95 | 5 | 20 | Yes |
| 1 | Female | 67 | 0 | -5.9300 | 3.4700 | 0.95 | 5 | 20 | Yes |
| 1 | Male | 59 | 0 | -5.3800 | 3.2300 | 0.95 | 5 | 20 | Yes |
| 2 | Both | 458 | 0 | -4.4400 | 2.8100 | - | - | - | Yes\* |
| 3 | Both | 419 | 0 | -4.5600 | 2.8500 | 0.8715 | 18.3 | 18.3 | Yes |
| 4 | Both | 774 | 0 | -4.5391 | 2.8900 | - | 27 | 45 | Yes\* |
| 5 | Both | 1450 | 0 | -4.6411 | 2.8900 | 0.96 | - | - | NA |
| 6 | Both | 582 | 0 | -5.8600 | 3.4349 | 0.99 | - | - | Yes\* |
| 6 | Male | 119 | 0 | -5.5700 | 3.3100 | 0.97 | - | - | Yes\* |
| 6 | Female | 115 | 0 | -5.1700 | 3.1437 | 0.94 | - | - | Yes\* |
| 7 | Both | 1887 | 0 | -5.5229 | 3.2400 | 0.97 | 15 | 30 | Yes\* |
| 8 | Both | 2143 | 0 | -5.2828 | 3.1832 | 0.9907 | 0.5 | 57 | Yes\* |
| 9 | Both | NA | 0 | -5.0293 | 3.0900 | - | - | - | NA |
| 10 | Both | 109 | 0 | -5.4941 | 3.2347 | 0.9766 | 5.7 | 38.1 | No |

Note: 1 = Aguilar-Perera & Quijano-Puerto (2016), 2 = Sandel et al. (2015), 3 = Chin, Aiken & Buddo (2016), 4 = Barbour et al. (2011), 5 = de Leon et al. (2013), 6 = Fogg et al. (2013), 7 = Edwards, Frazer & Jacoby (2014), 8 = Sabido-Itza et al. (2016), 9 = Froese & Pauly (2016), 10 = This study.