# **i**CLIPit

# Version

1.0

# **Authors**

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# Description

Code in python and test data for RNA iCLIP analysis. This repository contains 4 scripts that you can run:

- iclipBarplot.py
- iclipBoxplot.py
- iclipFdr.py
- iclipRealFreq.py
- iclipZscores.py

Example data and results are in ToyExample folder.

# Installation

Clone git repository by following command git clone https://github.com/anastazie/iCLIP

# **Dependencies**

```
To use iCLIPit you need to have following packages: numPy, pandas
You can install them on terminal by running following commands:
sudo pip install numpy
sudo pip install pandas
sudo pip install python-dateutil --upgrade
sudo pip install pytz --upgrade
```

# Getting help

In order to get full list of parameters for function type: python <function\_name> -h

# Data preprocessing

#### SAM files

This package use only positions in sam file. Before running scripts extract first 4 columns without header using following command: grep -v "^@" test.sam |cut -f1,2,3,4 > test1\_sub.sam

#### GTF files

Preprocess GFT file by removing header using following command: grep -v "^##" test.gtf > test\_nohead.gtf

# Get real frequencies

### Description

Compute number of mapped reads starting at each genome position

#### **Parameters**

-f File containing first 4 columns of .sam files without header

### Output

CSV file real freq.csv containing two columns:

#### Command

python iclipRealFreq.py -f test1\_sub.sam

# Compute FDR (False Discovery Rate) threshold

#### Description

Compute FDR threshold using following formula: (mean number of positions with n reads across i iterations of randomized datasets - standard deviation of positions with n reads across i iterations of randomized datasets) / number of positions with n reads in real dataset

#### **Parameters**

- -f File containing first 4 columns of .sam files without header
- -g Corresponding annotation file in GTF format
- -r Range of nucleotides in one direction from the position to calculate number of reads starting in the range: position +- range number, default value: 15
- -i Number of iterations to generate randomized datasets from input file, default value: 100

### Output

CSV file containing two columns:

#### Command

```
python iclipFdr.py -f test1_sub.sam -g test_nohead.gtf -r 20 -i 10
```

# Compute z-scores for k-mers

### Description

Compute k-mer z-scores using following formula: (number k-mer in real dataset - mean number of k-mers across i iterations of randomized datasets)/standard deviation of k-mers number across i iterations of randomized datasets

Only positions (both in real and randomized datasets) above specified threshold are considered for this computation.

#### **Parameters**

- -f File containing first containing two columns:
- -fa Fasta file containing reference nucleotide sequence in fa.tab format with two columns:
- -g Corresponding annotation file in GTF format
- -t FDR threshold value
- -r Range of nucleotides in one direction from the position to calculate number of reads starting in the range: position +- range\_number, default value: 15
- -i Number of iterations to generate randomized datasets from input file, default value: 100
- -k k-mer type: pentamer, hexamer, etc.
- -l Length of sequence to output into seq.txt file: position +- length, default value: 10

### Output

Two files are produced:

- $\_$ seq.txt contains sequence for positions above threshold +- l
- \_zscores.csv contains three columns:

#### Command

```
iclipZscores.py -f test1_sub.sam -fa test.fa.tab -g test_nohead.gtf -t 100 -r 15 -i 10 -k 4 -1 5
```

# Create barplot with read fraction per each gene

# Description

Create barplot by computing reads fraction started at each gene

#### **Parameters**

- -f File containing first containing two columns:
- -g Corresponding annotation file in GTF format
- -n number of genes, default value: all genes detected in GTF file
- -t plot title, default value: "Barplot"

## Output

PNG file with barplot, x - gene name, y - reads fraction, %

#### Command

iclipBarplot.py -f test1\_sub\_real\_freq.csv -g test\_nohead.gtf -n 5 -t testBar

# Get boxplot with reads fraction per each gene interval

### Description

Create boxplot by computing dividing each gene into p parts and counting number or reads starting in each interval

#### **Parameters**

- -f File containing first containing two columns:
- -g Corresponding annotation file in GTF format
- -n number of genes, default value: all genes detected in GTF file
- -t plot title, default value: "Boxplot"
- -p number of parts, into which all genes will be divide, default value: 100

### Output

PNG file containing boxplot per each gene (distribution of reads number in ech interval)

### Command

iclipBoxplot.py -f test1\_sub\_real\_freq.csv -g test\_nohead.gtf -n 5 -t testBox -p 100

# References

- 1. Wagnon, Jacy L., et al. "CELF4 regulates translation and local abundance of a vast set of mRNAs, including genes associated with regulation of synaptic function." PLoS genetics 8.11 (2012): e1003067.
- 2. König, Julian, et al. "iCLIP reveals the function of hnRNP particles in splicing at individual nucleotide resolution." Nature structural & molecular biology 17.7 (2010): 909-915.
- 3. Wang, Zhen, et al. "iCLIP predicts the dual splicing effects of TIA-RNA interactions." PLoS biology  $8.10\ (2010)$ : e1000530.