

Moderation

There aren't many options for latent variable interaction in R - at least not readily available. The two most common ones, LMS and product indicator approach, can be done in R with existing packages. Besides those, the authors of the book chapter have created multiple functions for other forms of latent variable interactions.

This script covers LMS and product indicator approaches.

LMS approach

<https://cran.r-project.org/web/packages/nlsem/nlsem.pdf> (<https://cran.r-project.org/web/packages/nlsem/nlsem.pdf>)

NLSEM can be used to perform latent variable interactions in models with **latent variables and latent interactions only**. The model cannot have observed variables as outcomes, latent variables with fewer than 3 indicators, and entering constraints is less straightforward than in *Mplus* or *lavaan*.

For this reason, we're using a different dataset to show this package.

```
library(nlsem)
```

```
## Warning: package 'nlsem' was built under R version 4.1.3
```

```
## Loading required package: orthopolynom
```

```
## Warning: package 'orthopolynom' was built under R version 4.1.3
```

```
## Loading required package: polynom
```

```
dataset <- read.csv("sample_timms2015.csv", header = TRUE)
```

The package cannot read variable names. You have to have your variables in order from x to y variables and with names x1, x2, x3, ... y1, y2, ..., and so on. So we're changing the variable names.

```
colnames(dataset) <- c(
    "x1",
    "x2",
    "x3",
    "x4",
    "x5",
    "x6",
    "y1",
    "y2",
    "y3"
)
```

There are a couple of ways of specifying models with NLSEM, one of them is to specify a lavaan type of model, and then use the `lav2nlsem()` function turn it into an object that can be read by the NLSEM package.

You also need to name the latent variables `xi1`, `xi2`, ..., for the exogenous latent variables (as in LISREL notation), and `eta1`, `eta2`, ..., for the endogenous latent variables.

```
lav_model <- 'xi1 =~ x1 + x2 + x3
             xi2 =~ x4 + x5 + x6
             eta1 =~ y1 + y2 + y3
             eta1 ~ xi1 + xi2 + xi1:xi2'

model <- lav2nlsem(lav_model)
model
```

```
## Model of class singleClass
##
## Number of latent endogenous variables: 1 (with 3 indicators)
## Number of latent exogenous variables: 2 (with 6 indicators)
##
## Structural model:
## -----
## eta1 = xi1 + xi2
## eta1 = xi1:xi2
## -----
##
## Measurement model:
## -----
## xi1 = x1 + x2 + x3
## xi2 = x4 + x5 + x6
## eta1 = y1 + y2 + y3
## -----
```

According to the NLSEM documentation, we then fit the model along with starting values, which we compute using the `runif()` function. Then we fit the model using the `em()` function.

```
# fit model
set.seed(123)
start <- runif(count_free_parameters(model))
start
```

```
## [1] 0.28757752 0.78830514 0.40897692 0.88301740 0.94046728 0.04555650
## [7] 0.52810549 0.89241904 0.55143501 0.45661474 0.95683335 0.45333416
## [13] 0.67757064 0.57263340 0.10292468 0.89982497 0.24608773 0.04205953
## [19] 0.32792072 0.95450365 0.88953932 0.69280341 0.64050681 0.99426978
## [25] 0.65570580 0.70853047 0.54406602 0.59414202 0.28915974 0.14711365
## [31] 0.96302423
```

If you run this code, you will notice that there are lots of errors during model fitting and that the process takes a **very** long time. This model can be fitted properly in Mplus, so here's a plug to go to Mplus if you can.

```
model_fit <- em(model = model,
               data = dataset,
               start = start,
               m = 16,
               max.iter = 500
               )
```

```
## Warning in value[[3L]](cond): Starting parameters for Phi are not positive
## definite. Identity matrix was used instead.
```

```
## Warning in em(model = model, data = dataset, start = start, m = 16, max.iter =
## 500): Loglikelihood should be increasing.
```

```
summary(model_fit)
```

```
## Warning in calc_standard_error(object$neg.hessian): Standard errors for some
## coefficients could not be computed.
```

```
##
## Summary for model of class singleClass
##
## Estimates:
##           Estimate Std. Error z value Pr(>|z|)
## Lambda.x2  1.2408417   0.052727  23.533 < 2e-16 ***
## Lambda.x3  1.3268918   0.051046  25.994 < 2e-16 ***
## Lambda.x11 0.8768280   0.022218  39.464 < 2e-16 ***
## Lambda.x12 0.8786082   0.022584  38.904 < 2e-16 ***
## Lambda.y2  1.0307247   0.024733  41.674 < 2e-16 ***
## Lambda.y3  1.0197537   0.024809  41.103 < 2e-16 ***
## Gamma1     0.5625107   0.113827   4.942 7.74e-07 ***
## Gamma2     0.0315354   0.074615   0.423 0.67256
## Theta.d1   0.1667143   0.011792  14.138 < 2e-16 ***
## Theta.d8   0.1644617   0.011630  14.141 < 2e-16 ***
## Theta.d15  0.1016765   0.007191  14.139 < 2e-16 ***
## Theta.d22  0.0827390   0.010568   7.830 4.90e-15 ***
## Theta.d29  0.0941960   0.009541   9.873 < 2e-16 ***
## Theta.d36  0.0987931   0.009892   9.988 < 2e-16 ***
## Theta.e1   0.1361468   0.011821  11.517 < 2e-16 ***
## Theta.e5   0.0580989   0.008392   6.923 4.42e-12 ***
## Theta.e9   0.0635175   0.008457   7.510 5.90e-14 ***
## Psi        0.7589051   0.066262  11.453 < 2e-16 ***
## Phi1       0.3344837   0.019148  17.468 < 2e-16 ***
## Phi2       0.3836239   0.032333  11.865 < 2e-16 ***
## Phi4       0.8423361   0.025777  32.678 < 2e-16 ***
## nu.x2      0.0017431      NaN      NaN      NaN
## nu.x3      0.0016258   0.004880   0.333 0.73901
## nu.x5     -0.0001687      NaN      NaN      NaN
## nu.x6     -0.0001684      NaN      NaN      NaN
## nu.y2     -0.0206417   0.020872  -0.989 0.32268
## nu.y3     -0.0089141   0.009646  -0.924 0.35541
## alpha     -0.1176024   0.055974  -2.101 0.03564 *
## tau1      0.0181249   0.006627   2.735 0.00624 **
## tau2      0.0194061   0.032983   0.588 0.55629
## Omega3    0.2536833   0.080090   3.167 0.00154 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Number of iterations: 101
## Final loglikelihood: -2168.723
```

Product indicator approach

As mentioned in lecture, if LMS is not possible, the product indicator approach is the next best option. the *semTools* package can do this in combination with lavaan.

semTools: <https://cran.r-project.org/web/packages/semTools/semTools.pdf> (<https://cran.r-project.org/web/packages/semTools/semTools.pdf>)

Load packages

```
library(semTools)
```

```
## Loading required package: lavaan
```

```
## This is lavaan 0.6-9  
## lavaan is FREE software! Please report any bugs.
```

```
##
```

```
## #####
```

```
## This is semTools 0.5-5
```

```
## All users of R (or SEM) are invited to submit functions or ideas for functions.
```

```
## #####
```

```
library(lavaan)
```

Load data and give variable names

```
narr_data <- read.table("PCL_MR.dat")
```

```
colnames(narr_data) <- c(  
  "PANASNEG",  
  "neurot",  
  "AIMN",  
  "ces",  
  "ptgi",  
  "closure",  
  "pcl"  
)
```

Create a new dataframe with new variables for the product of the indicators of the X and Z variables. To do this, we use the `indProd()` function of *semTools*.

```

prod_data <- indProd(
  data = narr_data, #create a new data.frame with the interaction indicators
  var1 = 1:3, #interaction indicators from the first latent
  var2 = 4:6, #interaction indicators from the second latent
  match = T, #use match-paired approach
  meanC = T, # mean centering the main effect indicator before making the products
  #residualC = T, #residual centering the products by the main effect indicators
  #doubleMC = T #centering the resulting products
)

head(prod_data[, (ncol(prod_data)-2):ncol(prod_data)]) #check the last three columns of the new data.frame

```

```

##      PANASNEG.ces neurot.ptgi AIMN.closure
## 1    -0.6368835    -1.438418     3.1226613
## 2     4.3838250    12.211928     5.9789113
## 3    -4.8706055    -8.966489    -5.7041317
## 4   -10.0010635   -11.455919    -0.2276461
## 5    -4.7256009     2.390108    -2.0786194
## 6     3.3995299    -5.275317    15.0419748

```

Specify a lavaan model

We'll now specify a lavaan model by creating a third latent variable for the interaction between X and Z. The indicators are the product terms created in the previous step.

```

product_model <- '
  NA_var =~ PANASNEG + neurot + AIMN
  Narr =~ ces + ptgi + closure
  NAxNarr =~ PANASNEG.ces + neurot.ptgi + AIMN.closure
  NAxNarr ~~ 0*NA_var + 0*Narr

  # Lines below necessary for model identification
  pcl ~ NA_var + Narr + NAxNarr # No correlation between int and xi1 xi2
  NA_var + Narr + NAxNarr ~ 0*1 # Intercept of latent variables fixed to 0
'

```

Fit the model

```

product_fit <- sem(product_model,
  data = prod_data,
  estimator = "MLR",
  meanstructure = TRUE)

```

Request the output

Note that, although the parameter estimates are not identical to Mplus's LMS approach, they have the same pattern and direction.

```
summary(product_fit,  
        standardized = TRUE,  
        rsquare = TRUE  
)
```

```

## lavaan 0.6-9 ended normally after 126 iterations
##
##   Estimator                      ML
##   Optimization method          NLMINB
##   Number of model parameters    33
##
##   Number of observations        488
##
## Model Test User Model:
##
##               Standard      Robust
##   Test Statistic      153.435    110.239
##   Degrees of freedom      32      32
##   P-value (Chi-square)    0.000    0.000
##   Scaling correction factor      1.392
##   Yuan-Bentler correction (Mplus variant)
##
## Parameter Estimates:
##
##   Standard errors          Sandwich
##   Information bread        Observed
##   Observed information based on      Hessian
##
## Latent Variables:
##
##               Estimate   Std.Err   z-value   P(>|z|)   Std.lv   Std.all
##   NA_var =~
##     PANASNEG      1.000
##     neurot        1.152    0.094    12.263    0.000    5.370    0.861
##     AIMN          1.245    0.099    12.534    0.000    5.805    0.805
##   Narr =~
##     ces           1.000
##     ptgi          0.915    0.064    14.361    0.000    0.905    0.634
##     closure       1.353    0.150     9.015    0.000    1.338    0.505
##   NAXNarr =~
##     PANASNEG.ces  1.000
##     neurot.ptgi   0.396    0.224     1.764    0.078    2.425    0.270
##     AIMN.closure  1.246    0.834     1.494    0.135    7.643    0.375
##
## Regressions:
##
##               Estimate   Std.Err   z-value   P(>|z|)   Std.lv   Std.all
##   pcl ~
##     NA_var        0.736    0.136     5.401    0.000    3.431    0.221
##     Narr          9.089    0.883    10.291    0.000    8.992    0.579
##     NAXNarr       0.431    0.239     1.806    0.071    2.646    0.170
##
## Covariances:
##
##               Estimate   Std.Err   z-value   P(>|z|)   Std.lv   Std.all
##   NA_var ~~
##     NAXNarr       0.000
##   Narr ~~
##     NAXNarr       0.000
##   NA_var ~~

```



```

##      Narr      1.133      0.276      4.099      0.000      0.246      0.246
##
## Intercepts:
##      Estimate Std.Err  z-value  P(>|z|)  Std.lv  Std.all
##      NA_var      0.000
##      Narr      0.000
##      NAXNarr      0.000
##      .PANASNEG    22.284    0.348    63.962    0.000    22.284    2.895
##      .neurot     30.020    0.282   106.347    0.000    30.020    4.814
##      .AIMN       27.838    0.326    85.331    0.000    27.838    3.863
##      .ces        2.805    0.049    56.958    0.000     2.805    2.578
##      .ptgi       2.990    0.065    46.248    0.000     2.990    2.094
##      .closure     4.810    0.120    40.087    0.000     4.810    1.815
##      .PANASNEG.ces -0.000    0.391   -0.000    1.000    -0.000   -0.000
##      .neurot.ptgi  0.000    0.406     0.000    1.000     0.000     0.000
##      .AIMN.closure 0.000    0.922     0.000    1.000     0.000     0.000
##      .pcl       32.635    0.702    46.494    0.000    32.635    2.103
##
## Variances:
##      Estimate Std.Err  z-value  P(>|z|)  Std.lv  Std.all
##      .PANASNEG    37.510    3.146    11.923    0.000    37.510    0.633
##      .neurot     10.048    1.717     5.853    0.000    10.048    0.258
##      .AIMN       18.244    2.387     7.643    0.000    18.244    0.351
##      .ces         0.205    0.061     3.363    0.001     0.205    0.173
##      .ptgi       1.221    0.094    13.000    0.000     1.221    0.599
##      .closure     5.235    0.348    15.062    0.000     5.235    0.745
##      .PANASNEG.ces 37.001   22.945     1.613    0.107    37.001    0.496
##      .neurot.ptgi 74.618    7.909     9.434    0.000    74.618    0.927
##      .AIMN.closure 356.134   45.572     7.815    0.000   356.134    0.859
##      .pcl       126.101   14.338     8.795    0.000   126.101    0.523
##      NA_var      21.721    3.188     6.813    0.000     1.000    1.000
##      Narr         0.979    0.087    11.273    0.000     1.000    1.000
##      NAXNarr     37.602   23.779     1.581    0.114     1.000    1.000
##
## R-Square:
##      Estimate
##      PANASNEG      0.367
##      neurot        0.742
##      AIMN           0.649
##      ces            0.827
##      ptgi           0.401
##      closure        0.255
##      PANASNEG.ces   0.504
##      neurot.ptgi    0.073
##      AIMN.closure    0.141
##      pcl            0.477

```

Probe the interaction

```
probe2way <- probe2WayMC(product_fit, # fitted lavaan model
  nameX = c("NA_var", "Narr", "NAxNarr"), # X variables
  nameY = "pcl", # Y variable
  modVar = "Narr", # moderator
  valProbe = c(-1, 0, 1)) # levels of the Z value at which to probe the
interaction

probe2way
```

```
## $SimpleIntcept
##   Narr    est    se      z pvalue
## 1   -1 23.546 0.993 23.721      0
## 2    0 32.635 0.702 46.494      0
## 3    1 41.723 1.249 33.407      0
##
## $SimpleSlope
##   Narr    est    se      z pvalue
## 1   -1 0.305 0.245 1.244 0.214
## 2    0 0.736 0.136 5.401 0.000
## 3    1 1.168 0.302 3.864 0.000
```

Using factor scores from Mplus

Finally, we can use Mplus factor scores to plot interactions in R. We'll use the same packages and approaches presented in lecture, which cover both spotlight (pick-a-point) and floodlight (Johnson-Neyman) approaches.

We create the factor scores in Mplus by fitting an LMS model, then request the saved factor scores. We can use the names in the Mplus output to determine the names of the columns in the dataset.

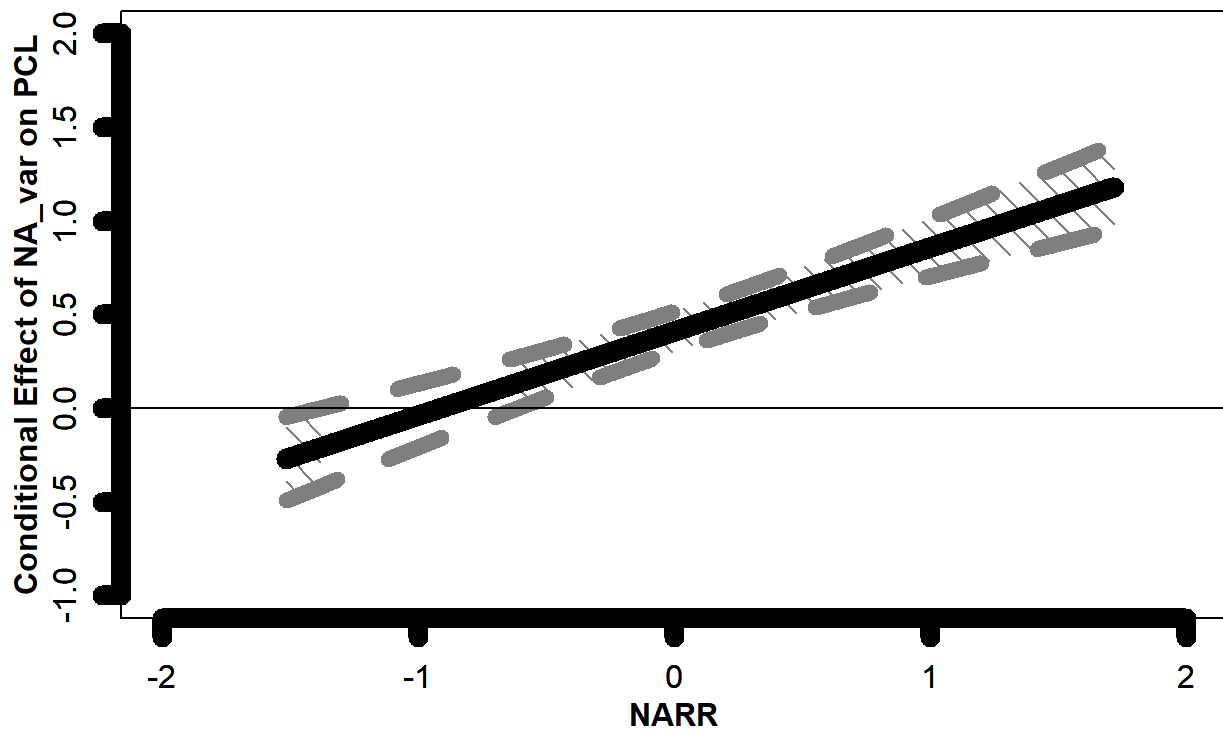
```
narr_scores <- read.csv("pcl_factorscores.csv", header = TRUE)
```

With *probemod* package

```
#install.packages("probemod")
library(probemod)
```

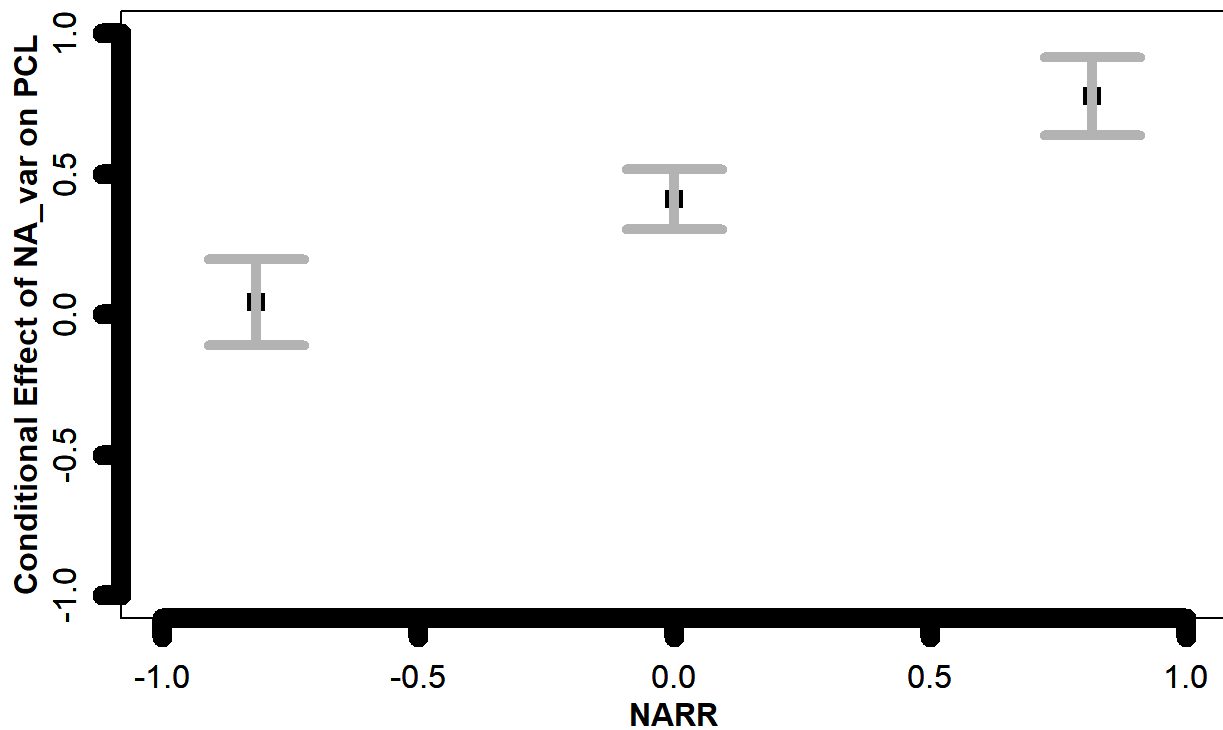
```
mod <- lm(PCL ~ NA_var + NARR + NA_var*NARR,
  data = narr_scores)
```

```
plotmod <- jn(mod, dv = "PCL", iv = "NA_var", mod = "NARR")
plot(plotmod)
```



```
## Values of NARR indicated by the shaded region
##           x           y           se           t           p           llci
## Lower Bound: -1.3801842 -0.2087620 0.1062463 -1.964888 0.05 -4.175240e-01
## Upper Bound: -0.6127909 0.1356036 0.0690134 1.964888 0.05 2.498002e-16
##           ulci
## Lower Bound: -2.498002e-16
## Upper Bound: 2.712071e-01
```

```
spot1 <- pickapoint(mod, dv = "PCL", iv = "NA_var", mod = "NARR", method = "meansd")
plot(spot1)
```



```
spot1
```

```
## Call:
## pickapoint(model = mod, dv = "PCL", iv = "NA_var", mod = "NARR",
##   method = "meansd")
## Conditional effects of NA_var on PCL at values of NARR
##      NARR      Effect      SE      t      p      llci
## -0.8163866685 0.04424052 0.07771446 0.5692701 5.694375e-01 -0.1084597
##  0.0002008197 0.41068186 0.05386281 7.6245898 1.309415e-13 0.3048475
##  0.8167883079 0.77712320 0.07102181 10.9420365 4.797968e-25 0.6375733
##      ulci
##  0.1969407
##  0.5165162
##  0.9166731
##
## Values for quantitative moderators are the mean and plus/minus one SD from the mean
## Values for dichotomous moderators are the two values of the moderator
```

With *interactions* package

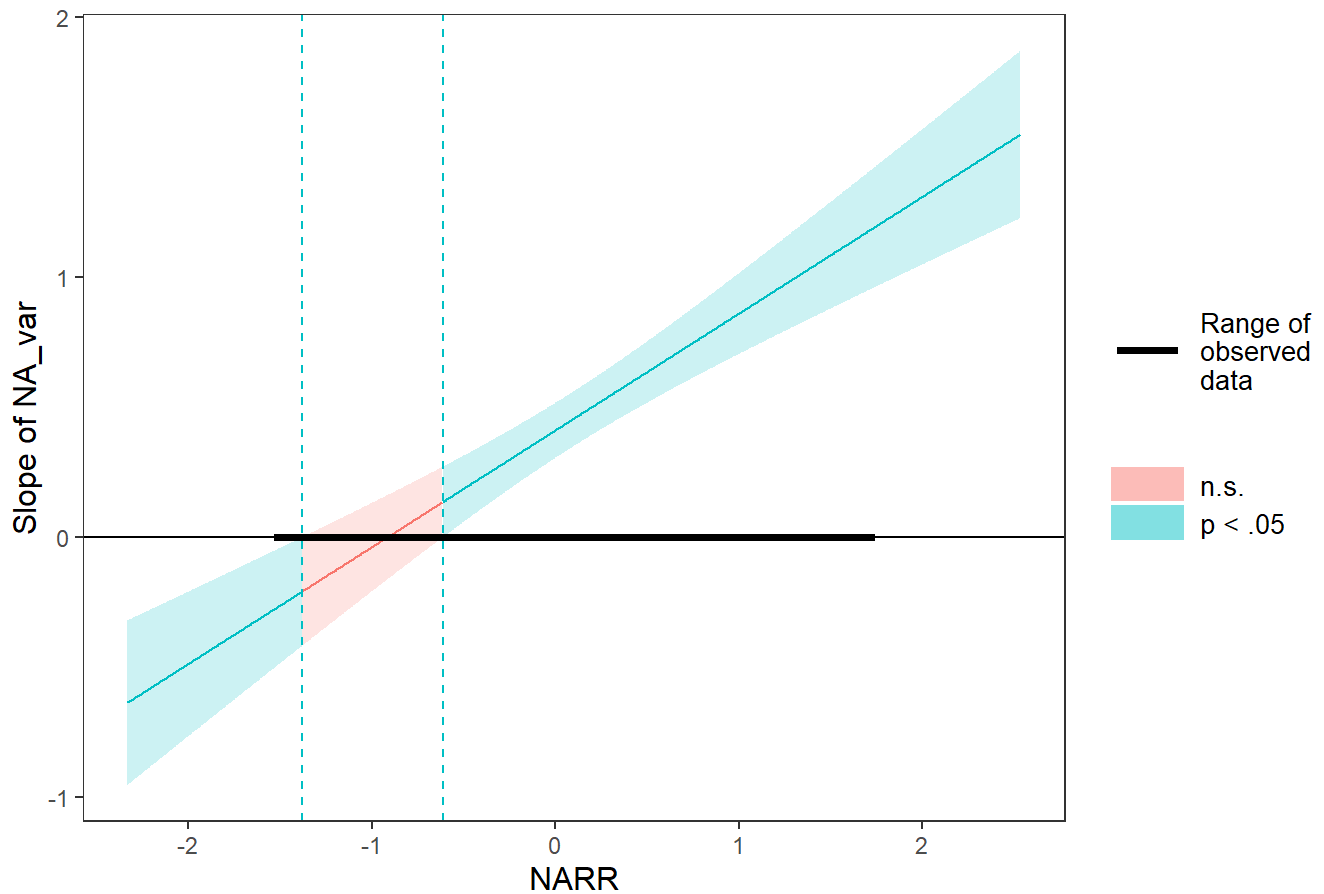
```
#install.packages("interactions")
library(interactions)
```

```
## Warning: package 'interactions' was built under R version 4.1.3
```

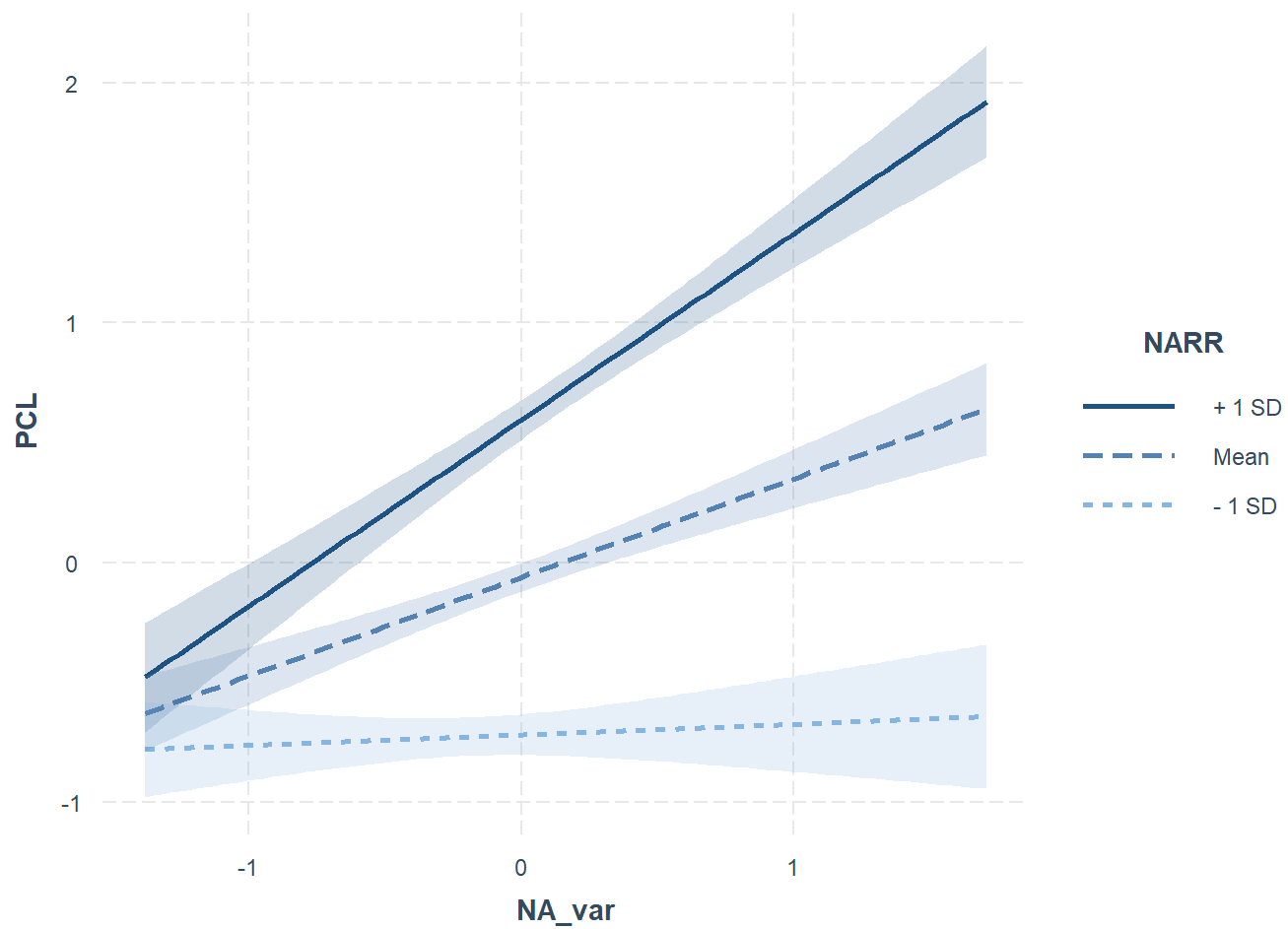
```
johnson_neyman(mod, pred = NA_var, modx = NARR)
```

```
## JOHNSON-NEYMAN INTERVAL
##
## When NARR is OUTSIDE the interval [-1.38, -0.61], the slope of NA_var is p
## < .05.
##
## Note: The range of observed values of NARR is [-1.52, 1.72]
```

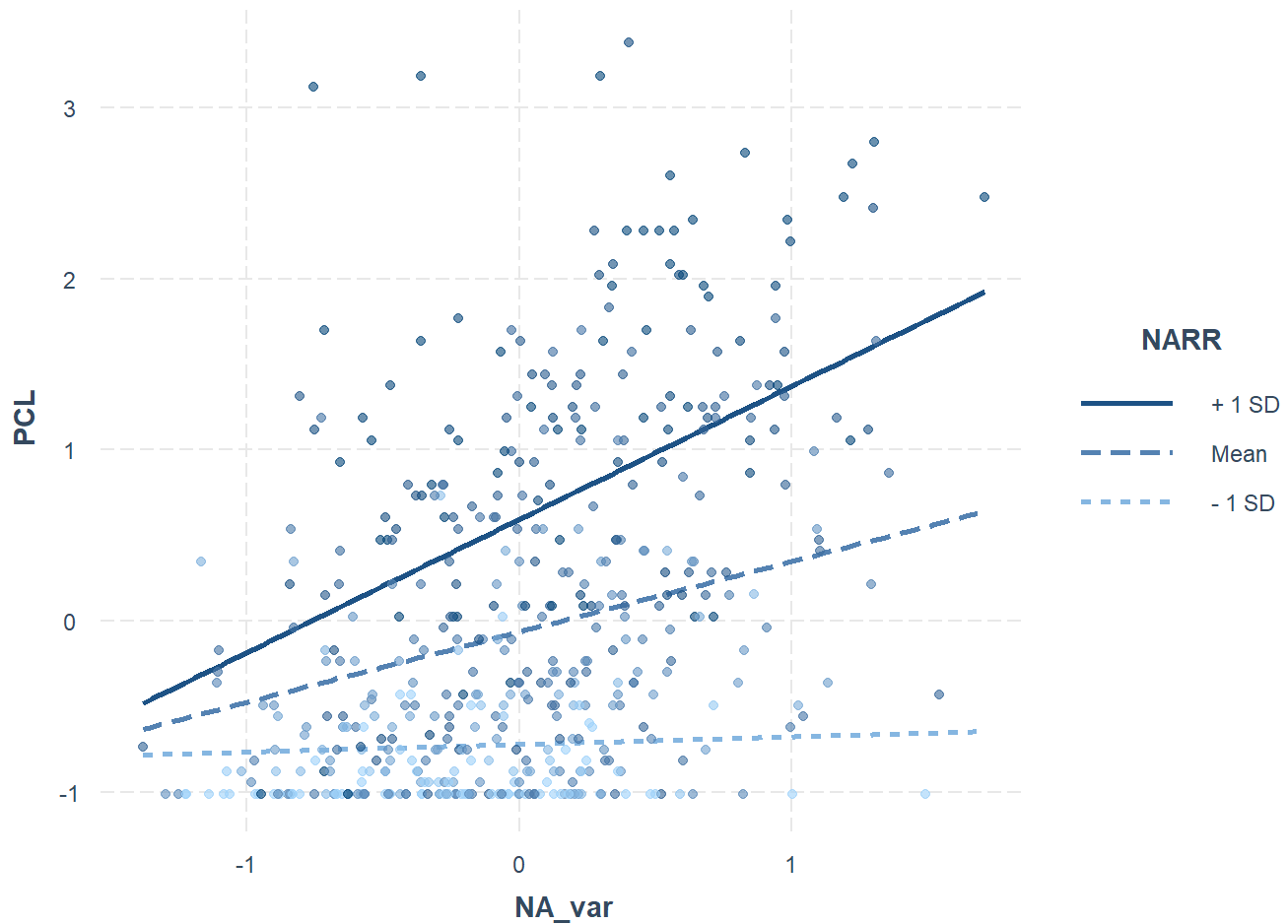
Johnson-Neyman plot



```
interact_plot(mod, pred = NA_var, modx = NARR, interval = TRUE)
```



```
interact_plot(mod, pred = NA_var, modx = NARR, plot.points = TRUE)
```



With *sandwich* package

```
library(sandwich)
```

```
## Warning: package 'sandwich' was built under R version 4.1.3
```

```
sim_slopes(mod, pred = NA_var, modx = NARR, jnplot = TRUE)
```

```
## JOHNSON-NEYMAN INTERVAL
```

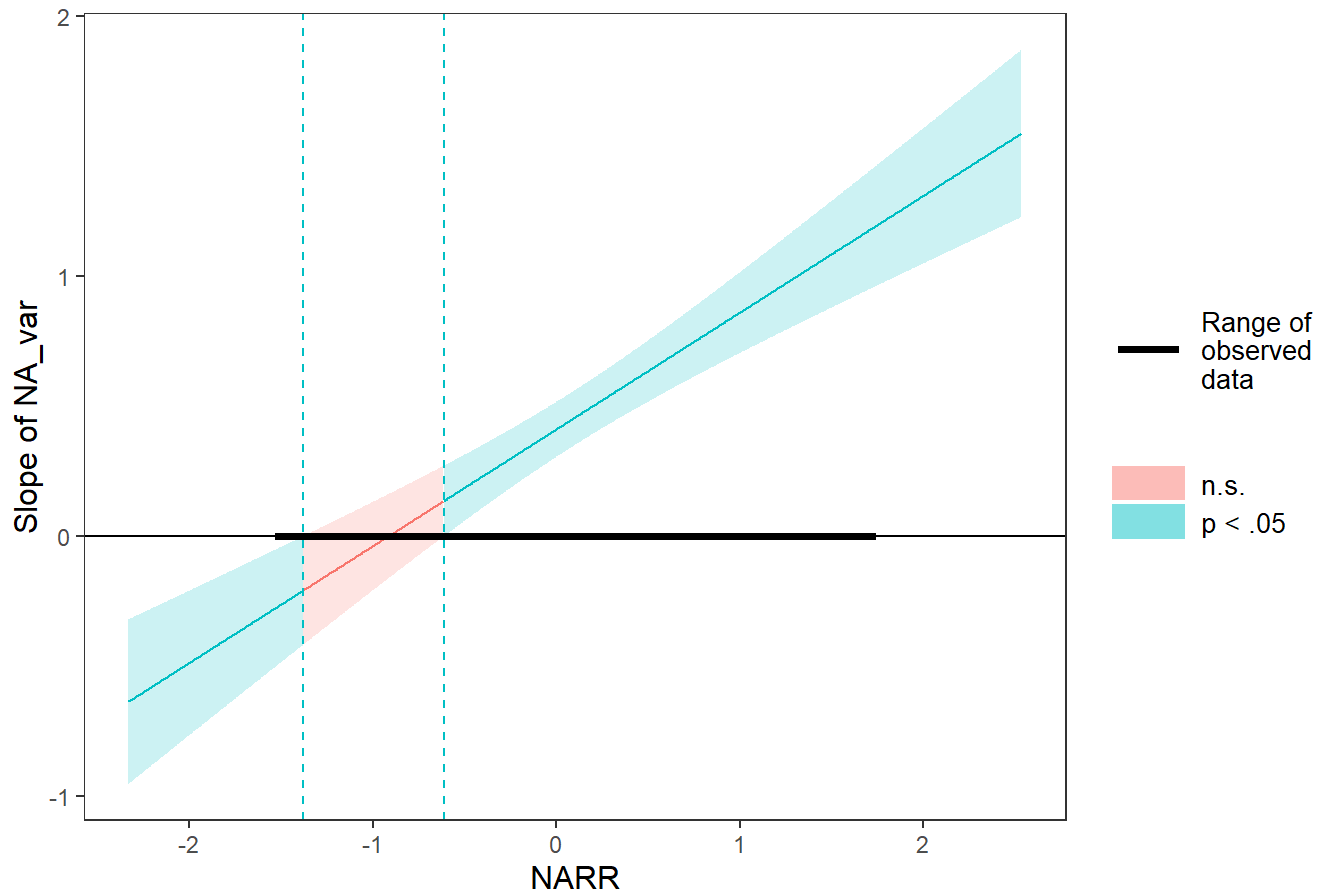
```
##
```

```
## When NARR is OUTSIDE the interval [-1.38, -0.61], the slope of NA_var is p  
## < .05.
```

```
##
```

```
## Note: The range of observed values of NARR is [-1.52, 1.72]
```

Johnson-Neyman plot



```
## SIMPLE SLOPES ANALYSIS
##
## Slope of NA_var when NARR = -0.816386685 (- 1 SD):
##
##   Est.   S.E.   t val.    p
## -----
##   0.04   0.08    0.57    0.57
##
## Slope of NA_var when NARR =  0.0002008197 (Mean):
##
##   Est.   S.E.   t val.    p
## -----
##   0.41   0.05    7.62    0.00
##
## Slope of NA_var when NARR =  0.8167883079 (+ 1 SD):
##
##   Est.   S.E.   t val.    p
## -----
##   0.78   0.07   10.94    0.00
```