**Abstract**

The “paradox of the great speciators” has puzzled evolutionary biologists for over half a century. A great speciator requires excellent dispersal ability to explain their occurrence on multiple islands, but reduced dispersal ability to explain their high number of subspecies. A rapid reduction in dispersal ability is often invoked to solve this apparent paradox, but a proximate mechanistic is lacking. Here we explore the role of six genes linked to migration and animal personality differences (*DRD4*, *SERT*, *ADCYAP1*, *CLOCK*, *CREB1* and *NPAS2*) in 20 south Pacific populations of silvereye (*Zosterops lateralis*) that range from highly sedentary to partially migratory, to determine if genetic variation is associated with dispersal propensity. We detected associations in three of the six genes: i) In a partial migrant population, migrant individuals had longer microsatellite alleles at the *CLOCK* gene compared to resident individuals from the same population; ii) *CREB1* displayed longer average microsatellite allele lengths in recently colonised island populations (< 200 years), compared to evolutionarily older populations. Bayesian linear and broken stick regression models supported a reduction in *CREB1* length with time since colonisation and decreasing dispersal propensity; iii) *DRD4* shows differences in polymorphisms reflecting the pattern found in *CREB1* but further work is needed to confirm. *ADCYAP1*, *SERT* and *NPAS2* were variable but that variation was not associated with dispersal propensity. This work shows that the “paradox of the great speciators” can be understood from a genetic perspective, paving the way for a resolution of the paradox.

Keywords: dispersal, diversification, *Zosterops lateralis*, candidate gene, great speciator, *CLOCK*, *CREB1*, *ADCYAP1*, *NPAS2*, *DRD4*

**Introduction**

Dispersal ability often determines the extent of species ranges and patterns of population divergence and speciation (Birand et al., 2012; Waters et al., 2020)⁠⁠. Highly dispersive taxa tend to have larger geographic ranges (Lester et al., 2007; Sheard et al., 2020)⁠ and more opportunities to exchange genes among populations in different parts of their range, inhibiting diversification (Birand et al., 2012; Gillespie et al., 2012; Lack, 1969; Mayr, 1942, but see Cardillo et al., 2003). In addition, the spatial scale of speciation is linked to variation in gene flow across a range of taxa (Kisel & Barraclough, 2010)⁠⁠, and reduced gene flow during early stages of divergence is central to many speciation models (Coyne & Orr, 1998; Grant & Grant, 2008; Hedrick PW, 1983; Mayr & Diamond, 2001; Price, 2008; Slatkin, 1987)⁠⁠.

Great speciators are those bird species that inhabit multiple islands reached by overwater dispersal events, suggesting excellent colonisation ability, yet display multiple morphological subspecies (Diamond et al., 1976; Manthey et al., 2020; Mayr, 1942; Moyle et al., 2009; Pedersen et al., 2018)⁠. To explain the observed distribution and diversity, a primary explanation relies on strong selection acting against dispersal on each of the newly established island populations. Selection may reflect the energetic cost of this behaviour (McNab 2002; Bonte et al., 2012) or the idea that once most islands are filled, the risks of overwater dispersal are higher than the demographic pay‐off. However, it is still unclear how these relatively rapid, post-colonisation changes in dispersal propensity could arise.

Variation in dispersal propensity is largely determined by differences in a combination of morphological, physiological and behavioural traits (Matthysen, 2012)⁠⁠. However, among the great speciators, it is thought that the shift from dispersive to sedentary forms primarily involves behavioural rather than morphological or physiological changes, at least initially (Diamond 1981). This is termed “behavioural flightlessness”, a reluctance to disperse (especially across water), despite the maintenance of normal wings and flight. In consequence, focusing solely on morphological proxies for dispersal ability that may take much longer to transpire, such as a more rounded wing (as indicated by a lower hand-wing index, Lockwood et al., 1998; Kipp et al., 1959) or a shift to a more graviportal body plan (reduced flight muscles and longer legs) (Wright et al. 2016) misses the part of the process that may be behaviourally driven.

A useful but relatively unexplored framework to understand behavioural flightlessness and subsequent divergence considers the links between dispersive behaviours and personality traits (Ingley & Johnson, 2014)⁠. Migration (seasonal movements) or dispersal (single movement to a new area) behaviours and personality traits are linked to some extent – migratory or highly dispersive taxa sometimes show increased levels of boldness, aggression and exploration, and lower levels of sociability. For example, migratory individuals within a species tend to be bolder than non-migratory ones (Chapman et al., 2011)⁠⁠; highly aggressive Western bluebird (*Sialia mexicana*) individuals were found to be more dispersive than their less aggressive counterparts (Duckworth & Badyaev, 2007; Duckworth & Kruuk, 2009)⁠⁠; European Stonechats (*Saxicola rubicola*) migrant males were more territorial than resident males (Marasco et al. 2011); and dispersive great tit (*Parus major*)individuals had a greater exploration rate than non-dispersive individuals (Korsten et al., 2013)⁠.

Migratory and dispersive behaviours have a high narrow sense heritability (h2=0.456, Dochtermann et al., 2019) suggesting a relevant genetic influence to these behavioural phenotypes. In fact, genetic variation for migration, dispersive behaviour and personality traits are well documented (Bubac et al., 2020), including the identification of a number of candidate genes underlying personality traits such as tendency for boldness or explorative behaviour (Canestrelli et al., 2016; Fidler et al., 2007; Ruegg et al., 2014; Steinmeyer et al., 2009)⁠⁠. Changes in genetic variation at these candidate genes may catalyse population-level shifts in dispersive behaviour.

To date, six candidate genes are mainly thought to play a role in contributing to a migratory phenotype, dispersive behaviour and personality traits in at least some bird species (Bubac et al. 2020; Table S1). In four of these genes, “adenylate cyclase activating polypeptide 1 gene” (*ADCYAP1*); the polyglutamine repeat region of the “circadian locomotor output cycles kaput gene” (*CLOCK*; Johnsen et al., 2007)⁠⁠; “neuronal PAS domain protein 2” (*NPAS2*); and “cAMP responsive element binding protein 1” (*CREB1*)), microsatellite allele length variation is associated with migratory-related traits across a variety of avian taxa (Table S1). The other two genes(“dopamine receptor D4” (*DRD4*) and “serotonin transporter” (*SERT*)) show associations between single nucleotide polymorphisms (SNPs) and differences in avian personality and migratory propensity (Table S1)⁠.

The Zosteropidae family (white-eyes, yuhinas and allies) consists of 142 species (Clements Checklist v2021), many of which are highly dispersive as evidenced by colonisation of numerous oceanic islands throughout the Indian and Pacific oceans, along with the broad continental distributions of some species (Clegg et al., 2002; Linck et al., 2016; Mees, 1969). This family shows one of highest per-lineage diversification rates for vertebrates (Moyle et al., 2009) and divergence can occur even across minor geographic barriers (e.g. water gaps of just 2 km) (Bertrand et al., 2014; Cowles & Uy, 2019; Manthey et al., 2020; Mees, 1969; Moyle et al., 2009)⁠⁠. A particularly interesting species within this family is the silvereye (*Zosterops lateralis*) as it can be considered a great speciator having multiple subspecies (at least 17 morphological subspecies; Clements Checklist v2021) that are distributed across a very wide natural distribution - including the Australian mainland and Tasmania, the North and South Islands of Aotearoa New Zealand, outlying oceanic islands of Australian and Aotearoa New Zealand, and the archipelagos of New Caledonia, Vanuatu, and Fiji (Figure 1). Silvereyes also display a variety of gene flow potentials: the Tasmanian subspecies (*Z. l. lateralis*) is a partial migrant (Mees, 1969)⁠; those in the central Vanuatu archipelago display high levels of outgoing gene flow, while more peripheral populations have high levels of incoming gene flow (Clegg & Phillimore, 2010)⁠⁠; and others, such as on Heron Island and Lord Howe Island, are sedentary and genetically isolated (Sendell-Price et al., 2020)⁠.

Here, we assess variation in six personality-related candidate genes in the silvereye examining different levels: individuals, populations and subspecies. Our aim is to determine whether there are signatures consistent with a genetic switch that would explain rapid shifts in dispersal, leading to reduced gene flow and subsequent increased opportunity for divergence. We ask i) if candidate gene variation follows patterns of neutral genomic divergence, that reflect drift and gene flow processes only; ii) if candidate gene variation is correlated with dispersal propensity among a range of dispersive and non-dispersive populations of silvereye, considering time since colonisation; and iii) if candidate gene variation is correlated with individual migratory status in the partially migratory Tasmanian population.

**Methods**

*Sampling and DNA Extraction*

Silvereye blood samples were collected between 1996 and 2016 from 20 sites in Tasmanian Melanesia (New Caledonia and Vanuatu) referred to henceforth as SM, and eastern Australia and Aotearoa New Zealand, including outlying islands, henceforth ANZO (Figure 1; Table S2). The sampling included putative Tasmanian winter migrant individuals captured in the Australian mainland states of Queensland and New South Wales. Individuals were assigned as Tasmanian migrants based on plumage differences: resident silvereyes (subspecies *Z. l. cornwalli*) exhibit bright yellow throat and vent plumage, while migrant individuals (subspecies *Z. l. lateralis*)exhibit white-to-pale-yellow throat and vent plumage (Fig. SX; Higgins et al., 2006). Birds were caught in mist-nets or traps and blood samples were taken via venipuncture of the brachial wing vein and stored in 90% ethanol or lysis buffer (10mM EDTA pH 8.0, 10mM TRIS-HCl pH 8.0, 20mM NaCl, 1% SDS, Seutin et al., 1991)⁠.

*Candidate microsatellite genotyping*

We extracted DNA from 477 samples using a standard chelex protocol (Walsh et al., 1991)⁠ and amplified four microsatellite candidate genes (NPAS2, CREB1, CLOCK and ADCYAP1) with the primers from Steinmeyer et al., (2009) using a touchdown polymerase chain reaction (PCR) protocol and fluorescent labelling (VIC and FAM) using M13 tags (Schuelke 2000). Fluorescent labelling with VIC and FAM was done in multiplex PCR reactions containing two loci each (VIC: CLOCK and ADCYAP1; FAM: NPAS2 and CREB1); final reaction volume of 3 µL contained 10ng DNA, 1 µl Type-it Master Mix (Qiagen), 0.17 µM of either FAM or VIC, and 0.01 µM forward M13-labelled primer and 0.05 µM reverse primer for each locus).

Thermal cycling consisted of an initial denaturation step of 95˚C for 15 min, eight cycles of 94˚C for 30 s, an annealing temperature of 60˚C, reduced by 1˚C each cycle, for 90 s and a 72˚C extension step for 60 s, then 25 cycles of 94˚C for 30 s, 52˚C for 90 s, and a 72˚C for 60 s, and a final extension of 60˚C for 30 min. VIC and FAM multiplexes were pooled for each sample and allele sizes for the candidate gene microsatellites were determined in relation to LIZ600 size standard on an ABI 3730xl DNA Analyser and scored using the Microsatellite Plugin within Geneious 2020.1 (http://www.geneious.com, Kearse et al., 2012)⁠⁠. We tested for the presence of null alleles, deviations from Hardy-Weinberg Equilibrium and linkage disequilibrium (LD) using the R package *genepop* version 1.1.7 (Rousset, 2008)⁠⁠. No locus showed evidence of LD (all p-values = 1).

*DRD4 and SERT sequencing*

We targeted one region of *DRD4*, encompassing intron 2 and exon 3, and part of intron 3 (1480 bp). For *SERT*, we amplified part of the promoter region (508 bp). Each 25 µl PCR reaction was comprised of approximately 10ng DNA template, 1.0 unit BioTaq (Bioline USA Inc.), 1 x BioTaq reaction buffer, and 0.5µM each forward and reverse primer, 200µM dNTPs, 1.5mM MgCl2, and Milli Q water to reach 25 µl. The PCR reaction profiles for both *DRD4* and *SERT* fragments consisted of an initial denaturation step of 94˚C for 3 min, followed by 10 cycles of 94˚C for 3 min, a touchdown step at 65˚C for 30 sec, and an extension step of 72˚C for 2 min. This was followed by 25 cycles of 94˚C for 30 sec, 55 ˚C annealing step for 30 sec, an extension step of 72˚C for 2 min, and a final extension step of 72˚C for 10 min. We purified PCR products using Acroprep 96 filter plates (Pall Corporation) following the manufacturer's protocol, then Sanger sequenced with forward and reverse primers on an ABI 3730xl DNA Analyser (Genetics Analysis Service, Otago University, NZ). We aligned the sequences using MUSCLE (Edgar, 2004)⁠⁠ in Geneious version 2020.1 (http://www.geneious.com, Kearse et al., 2012)⁠, and identified SNPs and insertions/deletions (indels). For *DRD4*, we aligned sequences to the great tit *DRD4* gene sequence (Genbank accession no.: DQ006801.1; Fidler et al., 2007)⁠. We called SNPs with a minimum minor allele frequency (MAF) of 0.05.

*Population genetic structure*

A subset of whole genome sequences available from Estandía et al. (in prep) were used to examine population structure patterns in NGSadmix (Skotte et al., 2013)⁠, a module implemented in ANGSD (Korneliussen et al., 2014)⁠. These whole genome sequences covered 336 of 422 individuals included in the present study. Note that not all individuals were screened for all candidate genes. We generated a BEAGLE file containing genotype likelihoods and created a subset of 10,000 SNPs picked at random after applying filtering for a MAF of 0.05. We ran NGSadmix with a range of genetic clusters (*k*), from 2 to the 20. We selected the best *k* for our dataset based on the mean estimate likelihoods, which indicated *k* = 2 as the optimal number of clusters, corresponding to ANZO and SM groupings. We re-ran NGSAdmix in each of the main cluster to explore potential substructuring. We visualised clustering patterns *k* = 2 to *k* = 5 using a custom R script (available at: [https://github.com/andreaestandia/1.0\_silvereye-candidate-genes](https://github.com/andreaestandia/0.0_silvereye-candidate-genes)/notebooks/1.0\_plots\_fig2\_popstructure.Rmd).

We generated a covariance matrix in PCAngsd (Meisner & Albrechtsen, 2018)⁠ using the BEAGLE file. Because not all individuals were screened for all candidate genes (e.g. 375 individuals for *NPAS2* but 258 for *CLOCK*) and that not all individuals in the candidate gene and whole-genome dataset coincide, we produced a population-level covariance matrix.

We calculated contemporary rates of gene flow among *Z. lateralis* populations for each of the two clusters (ANZO and SM) separately using BA3-SNP (Mussmann et al., 2019), an extension of BayesAss (Wilson et al., 2003) that allows SNPs as input. As input, we used the same file as for the population structure analysis (10,000 independent SNPs). This software employs a Bayesian approach with Markov chain Monte-Carlo (MCMC) sampling to estimate migration rates. We adjusted delta values for migration rates (m), allele frequencies (a) and inbreeding coefficients (f) to ensure that parameter space sampling acceptance rate was between 20% and 60% (Wilson & Rannala, 2003)⁠⁠. We ran the program for one million iterations, discarding the first 10% as burn-in. We estimated the 95% credible sets by calculating the mean±1.96\*Standard deviation (SD) as recommended by Rannala (2007).⁠

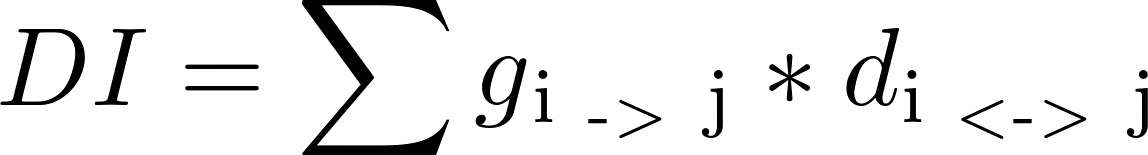
To assess model convergence, we compared results from 10 replicate runs each with a different random starting seed. We considered runs to have converged on a similar solution if gene flow estimates were within 0.005 (0.5%) across runs. Additionally, we inspected the likelihood trace files in Tracer v 1.7 (Rambaut et al., 2018)⁠ to determine that the log of the posterior probability values were consistent and to ensure that the Effective Sample Size (ESS) values were greater than 200. The run with the highest log posterior probability value was considered the best run to obtain parameter estimates.

*Candidate gene association analysis*

For all candidate genes we assessed variation among populations using the R package *brms* (Bürkner, 2017)⁠. Mean microsatellite length was set as the response variable and population was set as a categorical explanatory variable. In the case of *SERT* and *DRD4* the response variable was coded as 0 or 1 representing whether the individual carried the most frequent nucleotide or the variant and we used a bernoulli family for binary outcomes implemented in *brms.* For the two microsatellite candidate genes that showed obvious within and between population variation in length (*ADCYAP1* and *CREB1*) we tested whether population age and dispersal propensity could explain this variation by running Bayesian linear mixed models in *brms*. *CREB1* showed a clear distinction in allele lengths between a grouping consisting of the Australian mainland, Tasmania and recently colonised islands versus island populations thousands of years and older. Because of this structure (that does not completely align with ANZO and SM neutral structure groupings), we also applied a broken stick regression model for *CREB1* only to test whether including a single change point would improve predictive performance over an intercept-only and a linear model using *mcp* (multiple point change) (Lindeløv, 2020)⁠⁠. *NPAS2* and *CLOCK* displayed little among population variation in average allele length, hence we did not apply Bayesian linear mixed models to the whole population set. However, *CLOCK* showed variation within the partial migrant population of Tasmania so we explored whether individuals that migrated to the mainland had different *CLOCK* lengths than those that stayed over winter in Tasmania. The Tasmanian sample was restricted to 14 winter-caught birds (non-migrants) as the resident summer population includes a mix of migrants and non-migrants that cannot be phenotypically distinguished in Tasmania during the months where the migration is not occurring. The ‘migrant’ group consisted of those caught at Australian mainland sites in winter that were phenotypically identified as Tasmanian migrants (26 individuals).

For Bayesian linear mixed models (*CREB1* and *ADCYAP1*) and broken-stick regression models (*CREB1*), the following population-level (fixed) parameters used were:

(a) Dispersal Index (*DI*): the sum of each outgoing gene flow estimate (*g*) from island *i* into island *j* (where 95% credible interval did not overlap with zero), multiplied by the geographic distance between the islands (d) (Equation 1). The latter helps to account for differences in geographic opportunity for dispersal; for example, a geographically isolated island population that has moderate outgoing gene flow to few far islands would score higher than a centrally located island population with moderate outgoing gene flow to many close islands. *DI* was scaled from 0 (non-dispersive) to 1 (maximally dispersive).



Equation 1

(b) Age (Table S3): population ages for Aotearoa New Zealand, Chatham Island and Norfolk Island are known from historical records (c. 190 years; Clegg et al., 2002; Mees, 1969); for Heron Island, the population age was set as a maximum of 4000 years based on length of time the island has been vegetated (Clegg et al., 2008)⁠⁠; and molecular estimates for the remaining ancient populations were taken from a time-calibrated gene tree (Black, 2010)⁠⁠.

The intercept-only model represents the starting hypothesis that mean *CREB1* or *ADCYAP1* length differs between populations (which was set as group-level parameter or random effect) but does not change as a function of increasing dispersal propensity or time since colonisation. We controlled for population structure using the PCAngsd population-levelcovariancematrix. As *mcp* does not permit incorporation of distance matrices, we used a categorical variable that classified each individual according to its membership in one of the two population genetic clusters (ANZO or SM) identified from the top-level NGSadmix analysis.

For both *brms* ( a) Bayesian linear mixed model: Differences in length among populations for each candidate microsatellites; b) Bayesian linear mixed models: i) migration: *CLOCK*, and ii) population age and *DI*: *CREB1* and *ADCYAP1*) and *mcp* (*CREB1* only), average lengths were modelled with a Gaussian distribution and application of the default link function. For *DRD4* and *SERT* we used a bernoulli distribution to test for associations between presence/absence of the most common variant.

We set weakly informative priors for the intercepts of the segments before and after the change point (*mcp*). The same prior was applied for both *CREB1* length intercepts due to the lack of *a priori* information about a positive or negative correlation of mean *CREB1* length with population age and dispersal index. Priors for both intercepts were centred at 550 bp with a SD of 20 (N(550, 20)), based on known *CREB1* mean lengths in other passerines and how much it varies (Steinmeyer et al., 2009)⁠. For the *brms* mixed model, we set weakly informative priors with a normal distribution centred in 0 with a standard deviation of 20 for the *DI* and population age coefficients. This prior does not assume either an increase or decrease in bp but incorporates the prior information that changes will not be greater than 20 bp. We performed prior predictive checks, where data was generated according to the specified prior predictive distributions in order to assess their suitability (Gabry et al., 2019)⁠⁠.

For both the *brms* and *mcp* analyses we used MCMC with four chains of 4000 iterations each, including a warm-up of 400 iterations. We evaluated convergence via visual inspection of the MCMC trace plots, checking that the ESS>200 and the R values for each parameter (R = 1 at convergence). To evaluate model performance we compared our fitted models with an intercept-only model using leave-one-out cross-validation (LOO), a robust, fully Bayesian model selection approach (Vehtari et al., 2017)⁠.⁠

**Results**

*Population genetic structure and gene flow*

NGSadmix analysis of WGS data supported two main genetic clusters (*k* = 2): Cluster 1 comprised Australia, Aotearoa New Zealand and outlying island populations (ANZO), and Cluster 2 comprised Vanuatu and New Caledonia populations in Tasmanian Melanesia (SM) (Figure 2; Table S4). Other values of *k* also had high likelihoods (Table S?); *k* = 3 indicated sub-structuring within ANZO due to separation of Heron Island and Lord Howe Island from other populations (Fig. S1; see also *k*=2 in the ANZO analysis); *k* = 4 indicated sub-structuring within the SM cluster, primarily separating New Caledonia from Vanuatu populations, with the Tasmanian Vanuatu island of Tanna showing some affiliation with New Caledonia (Fig. S1, see also *k*=2 in the SM analysis). These population genetic patterns were consistent with those that emerged from the covariance matrix (Table S5, Fig. S2).

The ten independent BayesAss runs conducted to quantify the degree and direction of migration rates converged on the pattern of gene flow (Table S6). Of 42 pairwise comparisons within the ANZO cluster, and 132 in the SM cluster, eight and six respectively had estimates for which the credibility intervals did not overlap with zero. Within ANZO, this was primarily seen in relatively high outgoing gene flow estimates from Tasmania and Aotearoa New Zealand, and within SM, moderate outgoing levels from central islands of Pentecost and Malekula.

*Variation in candidate microsatellites, DRD4 and SERT*

Mean allele lengths for all microsatellite candidate loci are shown in Figure 2 for *CREB1* in particular and Figure 3 for all of them. *CREB1* showed longer mean lengths for the Australian mainland, Tasmania (migrants and non-migrants), and the recently colonised populations of Aotearoa New Zealand, Chatham Island, and Norfolk Island. Heron Island, Lord Howe Island and all Melanesian populations displayed shorter allele lengths on average.

*CLOCK* was monotypic in the majority of populations (Fig 3B). Compared to Tasmanian sedentary silvereyes, Tasmanian migrants showed longer allele lengths (Fig. 4; Fig S). Migrant individuals had long *CLOCK* variants (allele lengths of 289 and 291) not observed in any winter-caught Tasmanian birds (i.e. residents), or any other population except for a Heron Island sample. *NPAS2* showed some variation across populations but similar mean values for populations in the ANZO cluster. However, most SM populations were not variable at this locus, with the exception of peripherally located islands of Gaua, Efate and Tanna in Vanuatu. Variation between and within populations for *ADCYAP1* was evident for all populations.

We identified ten *DRD4* SNPs, seven of which were non-synonymous and three were synonymous. Five of them showed significant differences across populations but only SNP83 (a non-synonymous substitution) displayed consistent differences between ANZO and SM clusters (Table S11). SM individuals exclusively carried adenines, resulting in production of lysines, while those from Aotearoa New Zealand and Chatham Island only guanines producing argines. Tasmanian residents and Tasmanian migrants had both nucleotides represented (Fig. S3). We found one SNP and an INDEL in *SERT* but none were significantly different in frequency among populations.

*Candidate gene association tests*

Tasmanian migrants showed an increase of 1bp relative to non-migrants (Estimate=-1.06, 95% credibility intervals =-1.96, -0.12, Table SX).

Mean *CREB1* variation was better explained by a single change point model than the intercept-only or a linear regression model (Tables S11-S12). Time since colonisation and *DI* failed to explain the variability in *ADCYAP1* (Table S11).

The single change point model indicated that mean *CREB1* length decreased six base pairs with increasing population age (Intercept 1 = 552.294, Intercept 2 = 557.12; Table S12; Figure 5A), however the timing of this change had high uncertainty. The posterior probability density of change point ranged between 200 and 4000 years ago when we see the step reduction in allele length in the Heron Island population (maximum age 4000 years, Clegg et al. 2008), a reduction that is observed in other older populations (Lord Howe Island and Tasmanian Melanesian populations). The exception is Tasmania, an evolutionarily old population with long average *CREB1* length. *CREB1* decreased five base pairs with decreasing *DI* (Intercept 1 = 551.22, Intercept 2 = 556.22; Table S13; Figure 5B)*.* The posterior probability density of change point for *DI* ranged from 0.05 (Chatham Island) to 0.003 (Efate, an island in Tasmanian Vanuatu that shows low levels of outgoing gene flow into nearby islands).

**Discussion**

Rapid changes in dispersal could help to explain why taxa present on many islands show high levels of phenotypic diversity – the “paradox of the great speciators”. Quantifying variation in six candidate genes thought to underlie dispersal and migratory behaviour in populations of Australian mainland and island-colonising silvereyes has revealed three key results that hint at a role for genetic changes as a mechanism underlying this paradox. First, at the population level, we found that *CREB1* allele length variation was associated with time since colonisation and dispersal propensity, with recently colonised populations and those with high dispersal tendencies carrying longer alleles. Second, at the individual level, we found allele length variation in *CLOCK* associated with migratory status when comparing Tasmanian residents and Tasmanian migrants. Third, more limited data from a single *DRD4* SNP was fixed for different alleles in recently colonised versus evolutionarily older island populations. Together, these genes are likely to provide useful signatures of behavioural shifts in dispersal propensity in other silvereye populations, and possibly closely related species, though whether any act as a genetic switch remains unclear.

In the silvereye populations studied here, *CREB1* showed a disjunct pattern of allele sizes, being longer in the mainland, the partial migrant Tasmanian population, and all of the recently colonised populations when compared to island populations colonised thousands, or hundreds of thousands of years ago. This cannot be explained by population genetic groupings, as both the Heron Island population (up to 4000 years old) and Lord Howe Island population (over 100 thousand years old) (Clegg et al., 2002; Sendell-Price et al., 2020)⁠ had shorter *CREB1* allele lengths, yet clearly fell in the ANZO population genetic cluster that includes all of the long-allele-length populations. Also, the Heron Island samples were monomorphic for a unique (551bp) allele, and we cannot rule out that it may be fixed entirely due drift in this small population (McCallum et al. 2000). However, at the broad scale, the most dispersive populations, or those the result of evolutionarily recent colonisation events, carry longer allele lengths, compared to more sedentary, and more ancient island populations. This is the opposite pattern to that found by Chakarov et al. (2013) where non-dispersive individuals of common buzzard (*Buteo buteo*) carried longer *CREB1* allele lengths. *CREB1* allele lengths have also been shown to correlate with other life-history traits that also depend on circadian and circannual clocks, like dispersal ability e.g. incubation duration (Bourret & Garant, 2015)⁠, male moult speed (Bazzi et al., 2017)⁠. The broad scale patterns, and suggested association with dispersal, were not reflected within each of the longer- and shorter-allele length groups. For instance, in the short allele length group, Lord Howe Island, Gaua and Espiritu Santo displayed the longest lengths, but are not the most dispersive populations within that grouping; likewise for Norfolk and Chatham Islands in the long allele length group.

While the obvious length variation pattern seen at *CREB1* appears to have biological relevance for dispersal propensity in our study system, a mechanistic resolution of the “paradox of the great speciators” also requires dispersal shifts to be rapid (Diamond et al. 1976). We were unable to pinpoint when the *CREB1* allele size reduction occurs because of a lack of island populations of intermediate ages. At best we can say that for silvereyes, it takes more than 63 to 95 generations (South Island Aotearoa New Zealand, 190 years since colonisation, generation time of 2 to 3 years) and less than one to two thousand generations (Heron Island, 3,000 to 4,000 years old) (Clegg et al., 2008)⁠. Given the rapid phenotypic evolution observed in multiple silvereye populations post-colonisation (Clegg et al. 2002, Sendell-Price et al. 2020) and modelling that shows that the large Heron Island form likely evolved in far less time (some hundreds of generations) than its maximum age, ...

Variation at a second gene, *CLOCK*, was associated with migratory status of individuals, with migrant individuals having longer *CLOCK* allele variants, including long variants absent in individuals from non-migrant individuals from Tasmania, Australian mainland and all island populations. Previous studies in migratory birds focusing on *CLOCK* variation show mixed results. Longer allele lengths have been found to be positively correlated with migratory propensity (Peterson et al., 2013)⁠, variation in the phenology of migratory species (Bazzi et al., 2015; Caprioli et al., 2012; Liedvogel et al., 2009)⁠, but in some cases allele lengths are negatively correlated with migration date (Ralston et al., 2019)⁠ or not correlated at all (Contina et al., 2018; Mueller et al., 2011; Parody-Merino et al., 2019).

*CLOCK* plays a key role in regulating the circadian oscillator gene complex (Panda et al., 2002; Yu & Hardin, 2006)⁠, and is associated with variation in the phenology of photoperiodic traits (e.g. migratory behaviour) (Table S1). Photoperiodic stimulation experiments on caged Tasmanian silvereyes resulted in migratory restlessness being triggered in only migratory caged birds but not in resident ones, supporting a genetic link with migratory behaviour (Chan, 1994). In consequence, photoperiod changes could be the dispersal trigger with the onset of shorter autumnal days. The Tasmanian population is one of the few silvereye migrant population where changes in day length during the non-breeding, winter period is substantially greater than changes experienced further north and closer to the equator. Another population where these conditions are met is the one in Aotearoa New Zealand. In fact, even though records are few, large flocks of silvereyes seem to have migrated from the South to the North Island (Dennison et al. 1981), providing further support for photoperiod changes leading to migration. Some overwater dispersal events might be a direct consequence of migration. For example, the South Island of Aotearoa New Zealand population was founded by a small subset of migratory Tasmanian silvereyes. Flocks from this population kept colonising Pacific islands in a stepping-stone manner (Sendell-Price et al. 2021) suggesting widespread high dispersal ability. Given that migrants carry longer *CLOCK* alleles but we do not see them in the recently colonised populations, the high dispersal ability seen in those populations might be underlied by a set of genes that can be related to migration, but we have not included in this study. This question should be further explored in future work.

*DRD4* is one of the most well studied candidate genes related to exploratory and risk-taking behaviour (Bubac et al. 2020) – both of which could feasibly have links to dispersal propensity. We found a single polymorphism within *DRD4* (SNP83) that showed fixed differences between two young (more dispersive) and seven old (more sedentary) island populations. The Tasmanian population was polymorphic (G and A represented), the recently established populations of Chatham Island and South Island Aotearoa New Zealand were fixed for A, and Melanesian populations fixed for G. SNP83 is located in intron 2 and corresponds to base pair 9,423 on the *DRD4* orthologue of the great tit (Fidler et al., 2007). It has not been noted previously as having any phenotypic associations. Previous work assessing *DRD4* variation and personality in the great tit focused on associations between exploratory behaviour and variation at ‘SNP830’, revealing large effects in certain populations but not in others (Fidler et al., 2007; Korsten et al., 2013; Riyahi et al., 2017)⁠, however SNP830 was not variable in our dataset. Our data adds to the evidence of the role that variability in the DRD4 gene plays in a suite of behavioural phenotypes, however the extent of importance of SNP83 will require screening of more individuals and populations in silvereyes and other species.

Even though *NPAS2, ACDYAP1* and *SERT* showed some variability between populations it was not related to dispersal ability. Given the limitations of studying a handful of candidate genes for explaining complex behavioural phenotypes, failure to detect associations is not entirely unexpected. In birds, different associations (negative, positive or no correlation) in different populations, species and candidate genes are often reported (Table S1). Furthermore, few studies have taken into account candidate gene-environment interactions (but see Bourret & Garant, 2015; Liedvogel et al., 2009; Liedvogel & Sheldon, 2010)⁠⁠ or methylation patterns, that have been found to explain diverse complex behaviours in birds (Saino et al., 2019)⁠. The lack of replicability even within the same species could also be a product of sampling in different locations with a low number of individuals leading to a lack of statistical power, choosing different variables and proxies to measure migration and dispersiveness, using different methodologies used to analyse the data, publication bias or a combination of all of these (Yang et al. 2022). Thus, the lack of association in *NPAS2*, *ADCYAP1* and *SERT* does not rule out their potential role in dispersal behaviour as other factors might be masking their effects.

*Maintenance of high dispersal propensity in a continental island population*

Despite being an old insular form (>200K split from Australian mainland subspecies (Black, 2010)⁠), the Tasmanian silvereye (*Z. l. lateralis*) has maintained high dispersal propensity: it is a partial winter migrant to mainland Australia, it was the original source population for the historical sequential colonisation of Aotearoa New Zealand and outlying islands (Clegg et al., 2002; Mees, 1969)⁠; and as shown here, displays high levels of gene flow with Australian mainland subspecies. It also maintains the longer *CREB1* average allele lengths, and putative Tasmanian migrants caught on the mainland have shown unusually long *CLOCK* mean allele lengths. The maintenance of high dispersal potential, and its partial migrant status, is most likely explained by its geography and history of connectivity with the mainland. This continental island has repeatedly been connected to the Australian mainland during glacial periods and it is currently separated by a very shallow sea (average depth: 60 m and shortest distance to the Australian mainland: 200 km; Blom & Alsop, 1988)⁠. Over 50 islands of varying sizes can be found between the Australian mainland and Tasmania, which can act as migration stopovers, and facilitate connectivity.

*The future of dispersal genomics*

A candidate gene approach to understanding the “paradox of the great speciators” relies on knowledge of those genes in multiple systems. As discussed earlier, this approach has many limitations and often shows conflicting results. Alternative hypothesis-free approaches, like genome-wide association studies (GWAS), partially overcome the obstacles imposed by the incomplete understanding of the mechanisms underlying complex behaviours. GWAS requires phenotypes, in this case dispersal propensity, to be characterised at the individual level. Tasmanian migrants were relatively easy to identify because of their different morphological differences with the mainland subspecies. However, assigning a dispersal score to individuals where morphology is not easily distinguishable becomes challenging and it can only be properly studied using expensive tracking devices.

Genomics alone is unlikely to solve the complexity of the migratory phenotype. Recent studies have begun to incorporate transcriptome analysis in an effort to discover genetic determinants of migratory behaviour in birds, with the discovery of novel genes and chromosomal regions linked to migration strategies (Franchini et al., 2017; Frias-Soler et al., 2020; Horton et al., 2019; Lundberg et al., 2017)⁠.

Even though our results suggest that migrants have genetic differences to non-migrant individuals, and that more dispersive populations have different *CREB1* profiles to non-dispersive ones, a more thorough sampling is necessary to explore whether standing genetic variation within a population can provide the raw material for natural selection to act upon shifting a population to complete sedentariness.

**Conclusion**

In this study we assessed variation in six personality-related candidate genes in silvereye populations to examine whether signatures consistent with a genetic switch can explain rapid shifts in dispersal, leading to reduced gene flow and ultimately divergence in this great speciator. We find variation for all candidate microsatellite loci with the most clear associations found in *CREB1* which showed longer average allele lengths for mainland Australia, Tasmania and recently colonised islands, versus shorter average allele lengths in all island populations over 4000 years old. This pattern did not align with the population groupings of ANZO and Tasmanian Melanesia. We find strong support for the idea that more dispersive populations carry longer *CREB1* alleles, but length decreases with time and limited gene flow, suggesting that selection could be acting against dispersal ability following island colonisation. Although we also find large variation across populations in *ADCYAP1* our model failed to explain the variability based on time since colonisation or dispersal propensity. At the individual-level, partial migrants showed longer *CLOCK* lengths than non-partial migrant individuals.

Need a better closing sentence here – I’ll think about it….

*Figure captions*

Figure 1. Location of silvereye populations sampled for candidate gene variation. (A) Australian, Aotearoa New Zealand and outlying island (ANZO) cluster, showing location of inset (B) New Caledonia and Vanuatu (Tasmanian Melanesia – SM) populations. [deleteIn blue, populations that belong to the ANZO cluster. In grey, SM populations.delete]

Figure 2. (A) NGSadmix plot (*k* = 2) evidencing a split between ANZO and the SM populations. (B) The average *CREB1* gene allele length per population indicates a reduction in length for Heron and Lord Howe Islands, and all populations in the SM cluster. *CREB1* was not screened for Ambae and Pentecost samples.

Figure 3. Mean allele lengths (± ?) for the four microsatellite candidate genes. *CREB1* is longer in the source populations (Australia and Tasmania), and recently colonised populations (Aotearoa New Zealand, Chatham Islands and Norfolk Island) while shorter in older populations (Heron Island, Lord Howe Island, and SM populations). Migrant individuals from partial migrant populations have longer allele lengths at *CLOCK. ADCYAP1* shows extensive variation, including shorter lengths in Lord Howe Island and Malekula and longer lengths in Aotearoa New Zealand, Ouvea and Gaua. *NPAS2* shows little variation, with only Efate, Tanna and Gaua showing longer average lengths while Lord Howe Island shows the converse pattern. Blue and grey dots correspond to two population genetic clusters identified from admixture analysis.

Figure 4. Mean *CLOCK* length for Australia, Tasmania and putative Tasmanian migrants (Tasmanian silvereyes caught in winter on the Australian mainland). Only winter caught birds are included in this plot. Putative Tasmanian migrants show longer mean allele length. Mean values (± ? error) are shown in black, with coloured dots indicating….

Figure 5. Broken-stick regression model for CREB1 allele length variation. ANZO population data points (blue squares) and SM population data points (blue circles); 100 posterior draws (grey lines); change point posterior distributions (blue density curves); and 89% prediction intervals (grey dashed lines). (A) Relationship with population age, showing a higher mean length in recently colonised populations and their Tasmanian source population. [delete – too much here, save for discussion The position of Tasmanian samples is indicated, being the only old population with longer *CREB1* allele lengthsdue to its status as source population and high levels of gene flow with the mainland.delete] Change point posteriors range from ~200 years ago to ~4000 years ago. (B) Relationship with dispersal index (DI). The highest DI corresponds to Tasmania followed by Aotearoa New Zealand and Chatham Islands. Norfolk Island is among the young populations with no outgoing gene flow but long mean *CREB1* length. The change point posterior is centred between the DI values for Chatham Islands and Efate.

*References*

Bazzi, G., Ambrosini, R., Caprioli, M., Costanzo, A., Liechti, F., Gatti, E., Gianfranceschi, L., Podofillini, S., Romano, A., Romano, M., Scandolara, C., Saino, N., & Rubolini, D. (2015). Clock gene polymorphism and scheduling of migration: A geolocator study of the barn swallow Hirundo rustica. *Scientific Reports*, *5*(July), 1–7. https://doi.org/10.1038/srep12443

Bazzi, G., Podofillini, S., Gatti, E., Gianfranceschi, L., Cecere, J. G., Spina, F., Saino, N., & Rubolini, D. (2017). Candidate genes have sex-specific effects on timing of spring migration and moult speed in a long-distance migratory bird. *Current Zoology*, *63*(5), 479–486. https://doi.org/10.1093/cz/zow103

Bertrand, J. A. M., Bourgeois, Y. X. C., Delahaie, B., Duval, T., García-Jiménez, R., Cornuault, J., Heeb, P., Milá, B., Pujol, B., & Thébaud, C. (2014). Extremely reduced dispersal and gene flow in an island bird. *Heredity*, *112*(2), 190–196. https://doi.org/10.1038/hdy.2013.91

Birand, A., Vose, A., & Gavrilets, S. (2012). Patterns of species ranges, speciation, and extinction. *American Naturalist*, *179*(1), 1–21. https://doi.org/10.1086/663202

Black, R. A. (2010). *Phylogenetic and phenotypic divergence of an insular radiation of birds*. Imperial College London.

Blom, W. M., & Alsop, D. B. (1988). Carbonate mud sedimentation on a temperate shelf: Bass Basin, southeastern Australia. *Sedimentary Geology*, *60*(1–4), 269–280. https://doi.org/10.1016/0037-0738(88)90124-8

Bourret, A., & Garant, D. (2015). Candidate gene-environment interactions and their relationships with timing of breeding in a wild bird population. *Ecology and Evolution*, *5*(17), 3628–3641. https://doi.org/10.1002/ece3.1630

Bürkner, P. C. (2017). brms: An R package for Bayesian multilevel models using Stan. *Journal of Statistical Software*, *80*(1). https://doi.org/10.18637/jss.v080.i01

Cai, T., Cibois, A., Alström, P., Moyle, R. G., Kennedy, J. D., Shao, S., Zhang, R., Irestedt, M., Ericson, P. G. P., Gelang, M., Qu, Y., Lei, F., & Fjeldså, J. (2019). Near-complete phylogeny and taxonomic revision of the world’s babblers (Aves: Passeriformes). *Molecular Phylogenetics and Evolution*, *130*(May 2018), 346–356. https://doi.org/10.1016/j.ympev.2018.10.010

Canestrelli, D., Bisconti, R., & Carere, C. (2016). Bolder Takes All? The Behavioral Dimension of Biogeography. *Trends in Ecology and Evolution*, *31*(1), 35–43. https://doi.org/10.1016/j.tree.2015.11.004

Caprioli, M., Ambrosini, R., Boncoraglio, G., Gatti, E., Romano, A., Romano, M., Rubolini, D., Gianfranceschi, L., & Saino, N. (2012). Clock gene variation is associated with breeding phenology and maybe under directional selection in the migratory barn swallow. *PLoS ONE*, *7*(4). https://doi.org/10.1371/journal.pone.0035140

Cardillo, M., Huxtable, J. S., & Bromham, L. (2003). Geographic range size, life history and rates of diversification in Australian mammals. *Journal of Evolutionary Biology*, *16*(2), 282–288. https://doi.org/10.1046/j.1420-9101.2003.00513.x

Chakarov, N., Jonker, R. M., Boerner, M., Hoffman, J. I., & Krüger, O. (2013). Variation at phenological candidate genes correlates with timing of dispersal and plumage morph in a sedentary bird of prey. *Molecular Ecology*, *22*(21), 5430–5440. https://doi.org/10.1111/mec.12493

Chan, K. (1994). Nocturnal Activity of Caged Resident and Migrant Silvereyes (Zosteropidae: Aves). *Ethology*, *96*(4), 313–321. https://doi.org/10.1111/j.1439-0310.1994.tb01019.x

Chan, K. (2005). Partial migration in the silvereye (Aves Zosteropidae): Pattern, synthesis, and theories. *Ethology Ecology and Evolution*, *17*(4), 349–363. https://doi.org/10.1080/08927014.2005.9522589

Chapman, B. B., Hulthén, K., Blomqvist, D. R., Hansson, L. A., Nilsson, J. Å., Brodersen, J., Anders Nilsson, P., Skov, C., & Brönmark, C. (2011). To boldly go: Individual differences in boldness influence migratory tendency. *Ecology Letters*, *14*(9), 871–876. https://doi.org/10.1111/j.1461-0248.2011.01648.x

Clegg, S. M., Degnan, S. M., Kikkawa, J., Moritz, C., Estoup, A., & Owens, I. P. F. (2002). Genetic consequences of sequential founder events by an island-colonizing bird. *Proceedings of the National Academy of Sciences*, *99*(12), 8127–8132. https://doi.org/10.1073/pnas.102583399

Clegg, S. M., Degnan, S. M., Moritz, C., Estoup, A., Kikkawa, J., & Owens, I. P. F. (2002). Microevolution in island forms: The roles of drift and directional selection in morphological divergence of a passerine bird. *Evolution*, *56*(10), 2090–2099. https://doi.org/10.1111/j.0014-3820.2002.tb00134.x

Clegg, S. M., Frentiu, F. D., Kikkawa, J., Tavecchia, G., & Owens, I. P. F. (2008). 4000 Years of phenotypic change in an island bird: Heterogeneity of selection over three microevolutionary timescales. *Evolution*, *62*(9), 2393–2410. https://doi.org/10.1111/j.1558-5646.2008.00437.x

Clegg, S. M., & Phillimore, A. B. (2010). The influence of gene flow and drift on genetic and phenotypic divergence in two species of Zosterops in Vanuatu. *Philosophical Transactions of the Royal Society B: Biological Sciences*, *365*(1543), 1077–1092. https://doi.org/10.1098/rstb.2009.0281

Contina, A., Bridge, E. S., Ross, J. D., Shipley, J. R., & Kelly, J. F. (2018). Examination of clock and Adcyap1 gene variation in a neotropical migratory passerine. *PLoS ONE*, *13*(1), 1–13. https://doi.org/10.1371/journal.pone.0190859

Cowles, S. A., & Uy, J. A. C. (2019). Rapid, complete reproductive isolation in two closely related Zosterops White-eye bird species despite broadly overlapping ranges. *Evolution*, *73*(8), 1647–1662. https://doi.org/10.1111/evo.13797

Coyne, J. A., & Orr, A. H. (1998). The evolutionary genetics of speciation. *Philosophical Transactions of the Royal Society B: Biological Sciences*, *353*(1366), 287–305. https://doi.org/10.1098/rstb.1998.0210

Diamond, J. M. (1981). Flighlessness and the fear of flying in island species. *Nature*, *293*, 507–508.

Diamond, J. M., Gilpin, M. E., & Mayr, E. (1976). Species-distance relation for birds of the Solomon Archipelago, and the paradox of the great speciators. *Proceedings of the National Academy of Sciences of the United States of America*, *73*(6), 2160–2164. https://doi.org/10.1073/pnas.73.6.2160

Duckworth, R. A., & Badyaev, A. V. (2007). Coupling of dispersal and aggression facilitates the rapid range expansion of a passerine bird. *Proceedings of the National Academy of Sciences of the United States of America*, *104*(38), 15017–15022. https://doi.org/10.1073/pnas.0706174104

Duckworth, R. A., & Kruuk, L. E. B. (2009). Evolution of genetic integration between dispersal and colonization ability in a bird. *Evolution*, *63*(4), 968–977. https://doi.org/10.1111/j.1558-5646.2009.00625.x

Edgar, R. C. (2004). MUSCLE: Multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research*, *32*(5), 1792–1797. https://doi.org/10.1093/nar/gkh340

Fidler, A. E., Van Oers, K., Drent, P. J., Kuhn, S., Mueller, J. C., & Kempenaers, B. (2007). Drd4 gene polymorphisms are associated with personality variation in a passerine bird. *Proceedings of the Royal Society B: Biological Sciences*, *274*(1619), 1685–1691. https://doi.org/10.1098/rspb.2007.0337

Franchini, P., Irisarri, I., Fudickar, A., Schmidt, A., Meyer, A., Wikelski, M., & Partecke, J. (2017). Animal tracking meets migration genomics: transcriptomic analysis of a partially migratory bird species. *Molecular Ecology*, *26*(12), 3204–3216. https://doi.org/10.1111/mec.14108

Frias-Soler, R. C., Pildaín, L. V., Pârâu, L. G., Wink, M., & Bairlein, F. (2020). Transcriptome signatures in the brain of a migratory songbird. *Comparative Biochemistry and Physiology - Part D: Genomics and Proteomics*, *34*(February), 100681. https://doi.org/10.1016/j.cbd.2020.100681

Gabry, J., Simpson, D., Vehtari, A., Betancourt, M., & Gelman, A. (2019). Visualization in Bayesian workflow. *Journal of the Royal Statistical Society. Series A: Statistics in Society*, *182*(2), 389–402. https://doi.org/10.1111/rssa.12378

Gillespie, R. G., Baldwin, B. G., Waters, J. M., Fraser, C. I., Nikula, R., & Roderick, G. K. (2012). Long-distance dispersal: A framework for hypothesis testing. *Trends in Ecology and Evolution*, *27*(1), 47–56. https://doi.org/10.1016/j.tree.2011.08.009

Grant, B. R., & Grant, P. R. (2008). Fission and fusion of Darwin’s finches populations. *Philosophical Transactions of the Royal Society B: Biological Sciences*, *363*(1505), 2821–2829. https://doi.org/10.1098/rstb.2008.0051

Gu, Z., Pan, S., Lin, Z., Hu, L., Dai, X., Chang, J., Xue, Y., Su, H., Long, J., Sun, M., Ganusevich, S., Sokolov, V., Sokolov, A., Pokrovsky, I., Ji, F., Bruford, M. W., Dixon, A., & Zhan, X. (2021). Climate-driven flyway changes and memory-based long-distance migration. *Nature*, *591*(7849), 259–264. https://doi.org/10.1038/s41586-021-03265-0

Gwee, C. Y., Garg, K. M., Chattopadhyay, B., Sadanandan, K. R., Prawiradilaga, D. M., Irestedt, M., Lei, F., Bloch, L. M., Lee, J. G. H., Irham, M., Haryoko, T., Soh, M. C. K., Peh, K. S. H., Rowe, K. M. C., Ferasyi, T. R., Wu, S., Wogan, G. O. U., Bowie, R. C. K., & Rheindt, F. E. (2020). Phylogenomics of white-eyes, a ‘great speciator’’, reveals indonesian archipelago as the center of lineage diversity.’ *Elife*, *9*, 1–20. https://doi.org/10.7554/ELIFE.62765

Hedrick PW. (1983). *Genetics of populations*. Jones & Barlett Publishers.

Higgins, P. J., Peter, J. M., & Cowling, S. J. (2006). Vol. 7, Boatbill to Starlings. In *Handbook of Australian, New Zealand and Antarctic birds*. Oxford University Press.

Holtmann, B., Grosser, S., Lagisz, M., Johnson, S. L., Santos, E. S. A., Lara, C. E., Robertson, B. C., & Nakagawa, S. (2016). Population differentiation and behavioural association of the two “personality” genes DRD4 and SERT in dunnocks (Prunella modularis). *Molecular Ecology*, *25*(3), 706–722. https://doi.org/10.1111/mec.13514

Horton, W. J., Jensen, M., Sebastian, A., Praul, C. A., Albert, I., & Bartell, P. A. (2019). Transcriptome Analyses of Heart and Liver Reveal Novel Pathways for Regulating Songbird Migration. *Scientific Reports*, *9*(1), 1–12. https://doi.org/10.1038/s41598-019-41252-8

Ingley, S. J., & Johnson, J. B. (2014). Animal personality as a driver of reproductive isolation. *Trends in Ecology and Evolution*, *29*(7), 369–371. https://doi.org/10.1016/j.tree.2014.04.008

Johnsen, A., Fidler, A. E., Kuhn, S., Carter, K. L., Hoffmann, A., Barr, I. R., Biard, C., Charmantier, A., Eens, M., Korsten, P., Siitari, H., Tomiuk, J., & Kempenaers, B. (2007). Avian Clock gene polymorphism: Evidence for a latitudinal cline in allele frequencies. *Molecular Ecology*, *16*(22), 4867–4880. https://doi.org/10.1111/j.1365-294X.2007.03552.x

Kearse, M., Moir, R., Wilson, A., Stones-Havas, S., Cheung, M., Sturrock, S., Buxton, S., Cooper, A., Markowitz, S., Duran, C., Thierer, T., Ashton, B., Meintjes, P., & Drummond, A. (2012). Geneious Basic: An integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics*, *28*(12), 1647–1649. https://doi.org/10.1093/bioinformatics/bts199

Kisel, Y., & Barraclough, T. G. (2010). Speciation Has a Spatial Scale That Depends on Levels of Gene Flow. *The American Naturalist*, *175*(3), 316–334. https://doi.org/10.1086/650369

Korneliussen, T. S., Albrechtsen, A., & Nielsen, R. (2014). ANGSD: Analysis of Next Generation Sequencing Data. *BMC Bioinformatics*, *15*(1), 1–13. https://doi.org/10.1186/s12859-014-0356-4

Korsten, P., Van Overveld, T., Adriaensen, F., & Matthysen, E. (2013). Genetic integration of local dispersal and exploratory behaviour in a wild bird. *Nature Communications*, *4*(May 2013), 1–7. https://doi.org/10.1038/ncomms3362

Lack, D. (1969). The numbers of bird species on islands. *Bird Study*, *16*(4), 193–209. https://doi.org/10.1080/00063656909476244

Lester, S. E., Ruttenberg, B. I., Gaines, S. D., & Kinlan, B. P. (2007). The relationship between dispersal ability and geographic range size. *Ecology Letters*, *10*(8), 745–758. https://doi.org/10.1111/j.1461-0248.2007.01070.x

Liedvogel, M., & Sheldon, B. C. (2010). Low variability and absence of phenotypic correlates of Clock gene variation in a great tit Parus major population. *Journal of Avian Biology*, *41*(5), 543–550. https://doi.org/10.1111/j.1600-048X.2010.05055.x

Liedvogel, M., Szulkin, M., Knowles, S. C. L., Wood, M. J., & Sheldon, B. C. (2009). Phenotypic correlates of clock gene variation in a wild blue tit population: Evidence for a role in seasonal timing of reproduction. *Molecular Ecology*, *18*(11), 2444–2456. https://doi.org/10.1111/j.1365-294X.2009.04204.x

Linck, E., Bridge, E. S., Duckles, J. M., Navarro-Sigüenza, A. G., & Rohwer, S. (2016). Assessing migration patterns in Passerina ciris using the world’s bird collections as an aggregated resource. *PeerJ*, *2016*(4), 1–16. https://doi.org/10.7717/peerj.1871

Lindeløv, J. K. (2020). mcp: An R Package for Regression With Multiple Change Points. *Journal of Statistical Software*. https://doi.org/10.31219/osf.io/fzqxv

Lundberg, M., Liedvogel, M., Larson, K., Sigeman, H., Grahn, M., Wright, A., Åkesson, S., & Bensch, S. (2017). Genetic differences between willow warbler migratory phenotypes are few and cluster in large haplotype blocks. *Evolution Letters*, *1*(3), 155–168. https://doi.org/10.1002/evl3.15

Manthey, J. D., Oliveros, C. H., Andersen, M. J., Filardi, C. E., & Moyle, R. G. (2020). Gene flow and rapid differentiation characterize a rapid insular radiation in the southwest Pacific (Aves: Zosterops). *Evolution*. https://doi.org/10.1111/evo.14043

Matthysen, E. (2012). Multicausality of dispersal: a review. In *Dispersal ecology and evolution* (pp. 3–18). Oxford University Press.

Mayr, E. (1942). *Systematics and the origin of species*. Columbia University Press.

Mayr, E., & Diamond, J. (2001). *The Birds of Northern Melanesia: Speciation, Ecology, and Biogeography*. Oxford University Press.

Mees, G. (1969). A systematic review of the Indo-Australian Zosteropidae. In *Zoologische Verhandelingen* (102nd ed., pp. 1–390).

Mees, G. (1974). The migration of the Tasmanian race of the Silvereye. *Australian Bird Bander*, *12*, 51–54.

Mees, G. F. (1969). A Systematic Review of the Indo-Australian Zosterospidae (Part III). In *Rijksmuseum van Natuurlijke Historie, Leiden* (Vol. 102).

Meisner, J., & Albrechtsen, A. (2018). Inferring Population Structure and Admixture Proportions in Low-Depth NGS Data. *Genetics*, *210*(October), 719–731.

Moyle, R. G., Filardi, C. E., Smith, C. E., & Diamond, J. (2009). Explosive Pleistocene diversification and hemispheric expansion of a “great speciator.” *Proceedings of the National Academy of Sciences*, *106*(6), 1863–1868. https://doi.org/10.1073/pnas.0809861105

Mueller, J. C., Edelaar, P., Carrete, M., Serrano, D., Potti, J., Blas, J., Dingemanse, N. J., Kempenaers, B., & Tella, J. L. (2014). Behaviour-related DRD4 polymorphisms in invasive bird populations. *Molecular Ecology*, *23*(11), 2876–2885. https://doi.org/10.1111/mec.12763

Mueller, Jakob C., Pulido, F., & Kempenaers, B. (2011). Identification of a gene associated with avian migratory behaviour. *Proceedings of the Royal Society B: Biological Sciences*, *278*(1719), 2848–2856. https://doi.org/10.1098/rspb.2010.2567

Mussmann, S. M., Douglas, M. R., Chafin, T. K., & Douglas, M. E. (2019). BA3-SNPs: Contemporary migration reconfigured in BayesAss for next-generation sequence data. *Methods in Ecology and Evolution*, *10*(10), 1808–1813. https://doi.org/10.1111/2041-210X.13252

Oliveros, C. H., Andersen, M. J., & Moyle, R. G. (2021). A phylogeny of white-eyes based on ultraconserved elements. *Molecular Phylogenetics and Evolution*, 107273. https://doi.org/10.1016/j.ympev.2021.107273

Panda, S., Hogenesch, J. B., & Kay, S. A. (2002). Circadian rhythms from flies to human. *Nature*, *417*(6886), 329–335. https://doi.org/10.1038/417329a

Parody-Merino, Á. M., Battley, P. F., Conklin, J. R., & Fidler, A. E. (2019). No evidence for an association between Clock gene allelic variation and migration timing in a long-distance migratory shorebird (Limosa lapponica baueri). *Oecologia*, *191*(4), 843–859. https://doi.org/10.1007/s00442-019-04524-8

Pedersen, M. P., Irestedt, M., Joseph, L., Rahbek, C., & Jønsson, K. A. (2018). Phylogeography of a ‘great speciator’’ (Aves: Edolisoma tenuirostre) reveals complex dispersal and diversification dynamics across the Indo-Pacific.’ *J. Biogeogr.*, *45*(4), 826–837. https://doi.org/10.1111/jbi.13182

Peterson, M. P., Abolins-Abols, M., Atwell, J. W., Rice, R. J., Milá, B., & Ketterson, E. D. (2013). Variation in candidate genes CLOCK and ADCYAP1 does not consistently predict differences in migratory behavior in the songbird genus Junco. *F1000Research*, *2*, 1–16. https://doi.org/10.12688/f1000research.2-115.v1

Phillimore, A. B., Owens, I. P. F., Black, R. A., Chittock, J., Burke, T., & Clegg, S. M. (2008). Complex patterns of genetic and phenotypic divergence in an island bird and the consequences for delimiting conservation units. *Molecular Ecology*, *17*(12), 2839–2853. https://doi.org/10.1111/j.1365-294X.2008.03794.x

Price, T. (2008). *Speciation in Birds*. Roberts & Company Publishers.

Ralston, J., Lorenc, L., Montes, M., DeLuca, W. V., Kirchman, J. J., Woodworth, B. K., Mackenzie, S. A., Newman, A., Cooke, H. A., Freeman, N. E., Sutton, A. O., Tauzer, L., & Norris, D. R. (2019). Length polymorphisms at two candidate genes explain variation of migratory behaviors in blackpoll warblers (Setophaga striata). *Ecology and Evolution*, *9*(15), 8840–8855. https://doi.org/10.1002/ece3.5436

Rambaut, A., Drummond, A. J., Xie, D., Baele, G., & Suchard, M. A. (2018). Posterior Summarization in Bayesian Phylogenetics Using Tracer 1.7. *Systematic Biology*, *67*(5), 901–904. https://doi.org/10.1093/sysbio/syy032

Rannala, B. (2007). *BayesAss edition 3.0 user’s manual*. University of California, Davis.

Riyahi, S., Björklund, M., Mateos-Gonzalez, F., & Senar, J. C. (2017). Personality and urbanization: behavioural traits and DRD4 SNP830 polymorphisms in great tits in Barcelona city. *Journal of Ethology*, *35*(1), 101–108. https://doi.org/10.1007/s10164-016-0496-2

Rousset, F. (2008). GENEPOP’007: A complete re-implementation of the GENEPOP software for Windows and Linux. *Molecular Ecology Resources*, *8*(1), 103–106. https://doi.org/10.1111/j.1471-8286.2007.01931.x

Ruegg, K., Anderson, E. C., Boone, J., Pouls, J., & Smith, T. B. (2014). A role for migration-linked genes and genomic islands in divergence of a songbird. *Molecular Ecology*, *23*(19), 4757–4769. https://doi.org/10.1111/mec.12842

Saino, N., Albetti, B., Ambrosini, R., Caprioli, M., Costanzo, A., Mariani, J., Parolini, M., Romano, A., Rubolini, D., Formenti, G., Gianfranceschi, L., & Bollati, V. (2019). Inter-generational resemblance of methylation levels at circadian genes and associations with phenology in the barn swallow. *Scientific Reports*, *9*(1), 1–16. https://doi.org/10.1038/s41598-019-42798-3

Sendell-Price, A. T., Ruegg, K. C., Anderson, E. C., Quilodrán, C. S., van Doren, B. M., Underwood, V. L., Coulson, T., & Clegg, S. M. (2020). The genomic landscape of divergence across the speciation continuum in Island-colonising silvereyes (Zosterops lateralis). *G3: Genes, Genomes, Genetics*, *10*(9), 3147–3163. https://doi.org/10.1534/g3.120.401352

Sendell-Price, A. T., Ruegg, K. C., Robertson, B. C., & Clegg, S. M. (2021). An island-hopping bird reveals how founder events shape genome-wide divergence. *Molecular Ecology*, *30*(11), 2495–2510. https://doi.org/10.1111/mec.15898

Seutin, G., White, B. N., & Boag, P. T. (1991). Preservation of avian blood and tissue samples for DNA analyses. *Canadian Journal of Zoology*, *69*(1), 82–90. https://doi.org/10.1139/z91-013

Sheard, C., Neate-Clegg, M. H. C., Alioravainen, N., Jones, S. E. I., Vincent, C., MacGregor, H. E. A., Bregman, T. P., Claramunt, S., & Tobias, J. A. (2020). Ecological drivers of global gradients in avian dispersal inferred from wing morphology. *Nature Communications*, *11*(1). https://doi.org/10.1038/s41467-020-16313-6

Skotte, L., Korneliussen, T. S., & Albrechtsen, A. (2013). Estimating individual admixture proportions from next generation sequencing data. *Genetics*, *195*(3), 693–702. https://doi.org/10.1534/genetics.113.154138

Slatkin, M. (1987). Gene Flow and the Geographic Structure of Natural Populations. *Science*, *3576198*(787), 236. https://doi.org/10.4269/ajtmh.2010.09-0588

Steinmeyer, C., Mueller, J. C., & Kempenaers, B. (2009). Search for informative polymorphisms in candidate genes: Clock genes and circadian behaviour in blue tits. *Genetica*, *136*(1), 109–117. https://doi.org/10.1007/s10709-008-9318-y

Toews, D. P. L., Taylor, S. A., Streby, H. M., Kramer, G. R., & Lovette, I. J. (2019). Selection on VPS13A linked to migration in a songbird. *Proceedings of the National Academy of Sciences of the United States of America*, *116*(37), 18272–18274. https://doi.org/10.1073/pnas.1909186116

Vehtari, A., Gelman, A., & Gabry, J. (2017). Practical Bayesian model evaluation using leave-one-out cross-validation and WAIC. *Statistics and Computing*, *27*(5), 1413–1432. https://doi.org/10.1007/s11222-016-9696-4

Walsh, P. S., Metzger, D. A., & Higuchi, R. (1991). Chelex 100 as a medium for simple extraction of DNA for PCR-based typing from forensic material. *Biotechniques*, *10.4*, 506–513.

Warren, B. H. (2003). *Phylogeography and evolution of species-rich bird lineages of the western Indian Ocean islands*. University of East Anglia.

Waters, J. M., Emerson, B. C., Arribas, P., & Mcculloch, G. A. (2020). Dispersal Reduction: Causes , Genomic Mechanisms , and Evolutionary Consequences. *Trends in Ecology & Evolution*, 1–11. https://doi.org/10.1016/j.tree.2020.01.012

Wilson, G. A., & Rannala, B. (2003). Bayesian inference of recent migration rates using multilocus genotypes. *Genetics*, *163*(3), 1177–1191.

Yu, W., & Hardin, P. E. (2006). Circadian oscillators of Drosophila and mammals. *Journal of Cell Science*, *119*(23), 4793–4795. https://doi.org/10.1242/jcs.03174