

# SLIM Table Generator

## (tablegenerator.py)

### User Guide

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## Index

1. Introduction .....	1
2. Input files .....	2
3. Output files .....	2
4. Configuration file .....	3
5. Example case .....	7
5.1. Input files .....	7
5.2. Configuration file .....	8
5.3. Output tables .....	10
5.4. Output abundances list .....	11
Useful links .....	12
Credits .....	13
License .....	13

## 1. Introduction

The SLIM Table Generator is a Python 3 script, `tablegenerator.py`, that creates tables in LaTeX format (.tex) out of the results of the SLIM tool from the MADCUBA package.<sup>1</sup> It can create a table of the transition lines studied and a table of the abundances of the corresponding species with respect to a reference molecule.

The script requires the modules `os`, `re`, `sys`, `copy`, and `platform`, from the Python's standard library, and also the libraries `PyYAML`, `NumPy`, `Pandas`, and `RichValues`.<sup>2</sup> In order to run the code, it is necessary to specify some options in a configuration file. For running the code we have to write the following command line in the terminal, being in the folder of the file `tablegenerator.py`:

```
python3 tablegenerator.py <path> .
```

The argument `<path>` is the path of the configuration file. If we do not specify it, the

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1. <https://cab.inta-csic.es/madcuba/>

2. You can download this library from the following link:

<https://drive.google.com/drive/folders/1g5l3I2gowZ-s37z2w-eEDqUb12wj3dHN?usp=sharing>.

In order to install it, you have to copy the file `richvalues.py` to one of the Python system paths. To display the list of all these paths, you can run Python (for example, from the terminal, writing `python3`), and run the two following commands: `import sys` and `sys.path`. For example, for Mac, one of these paths would be: `/Users/<user>/Library/Python/<3.x>/lib/python/site-packages`, with `<user>` being your username and `<3.x>` your exact Python version.

script will search in the current folder (./) for the configuration file specified inside the script in the variable `config_file`, at the beginning of the code; we can use this variable to run the script in a programming environment like Spyder. If the script `tablegenerator.py` is not in the current folder, but in a folder with path `<folder>`, we should write `<folder>/tablegenerator.py` instead of `tablegenerator.py`. Moreover, if the script is included in a folder of executable files, the term `python3` can be removed from the command line, and the script will be run from any path.

## 2. Input files

Apart from the configuration file, we need at least two files in order to run this script. They are described below, using the same name used in the configuration file:

- **MADCUBA table.** Transition lines table in .csv format generated by MADCUBA. To generate it in SLIM, select one of the species in the SimFit tab to open the transition table window, check the Spectral, Fit, and Phys. options, and click the save icon to export the table in .csv format.
- **processing table.** Table in .csv format with information of the RMS noise and resolution of the spectra used in MADCUBA. Each row, which corresponds to one spectral range, must have the following columns: min. frequency (MHz) (minimum frequency), max. frequency (MHz) (maximum frequency), rms noise (mK) (RMS noise), and resolution (MHz) (frequency resolution). This file can be generated by the Python Reduction Pipeline for CLASS (script `processing.py`) once we have run the steps `--selection`, `--line_search`, and `--reduction`, adding the argument `--spectra_tables` (for example: `python3 processing.py config.yaml --spectra_table`). If there are multiple sources, this variable should be a list.
- **LaTeX template** (optional). Template with the structure of the tables in .tex format. Each of the two possible tables must contain only one row with a specific keyword; in the case of the transitions table, it is `--tablelines--`, and for the abundances table, it is `--tableabunds--`. It can be easily done with a graphical LaTeX editor like Lyx. If we do not supply this file, a generic simple template will be automatically generated according to the parameters set in the configuration file.
- **non-LTE lines table** (optional). In case that we want to manually set the parameters of the transitions of some species (usually because they are not studied with a local thermodynamical equilibrium (LTE) model), this table in .tex format should contain the information of those transitions. It can be easily done with a graphical LaTeX editor like Lyx. This is a nested variable that should contain another variable with the name of the table in .tex format, and then a list containing the MADCUBA names of the molecules included in the table.

## 3. Output files

The main output file that will be generated with this script is a .tex file containing two tables

(or only one of them, depending on what we write in the configuration file):

- **The transition table.** It contains information about the specified transitions in the configuration file. The possible columns for each row are:
  - The species name.
  - The quantum numbers of the transition.
  - The frequency of the transition, in MHz.
  - The integrated area of the line, in  $\text{mK} \cdot \text{km/s}$ .
  - The line width of the transition, in  $\text{km/s}$ .
  - The local sidereal rest (LSR) velocity of the line, in  $\text{km/s}$ .
  - The signal-to-noise (S/N) value for the integrated area of the line.
- **The abundances table.** It contains information about the abundance of each detected molecule. The possible columns for each row are:
  - The species name.
  - The temperature of the species (in case of LTE, excitation temperature).
  - The column density of the species.
  - The abundance of the species relative to the reference molecule.

Additionally, there is a plain text file in .csv format that can be created containing the abundances of the observed species, for each observed source. This file can be used as an input for the Molecular Abundances Bar Plotter (`abundancesbarplot.py`).

## 4. Configuration file

The configuration file is a plain text document with the YAML formatting style. This way, we can specify the options of the script `tablegenerator.py` by defining different variables that can be nested.

In order to define the values of the variables, we have to write, in a single line, the name of the variable followed by a colon (:), and its value, separated with a space. For the logical variables, the possible values are yes/no. There are variables whose value can be a list of elements, in which case they can be written between brackets and separated by commas or in single lines preceded by a hyphen (-). Similarly, in the case of nested variables, which work like Python dictionaries, next level variables must be written in a different line and with an indent. Text variables can be written between simple quotation marks ('), but this is optional. If the definition of a variable is not written,<sup>3</sup> its default value will be used (see Section 3).

Below is a list of all the variables with their meaning and possible values:

- **create lines table.** Logical variable that determines if the lines transitions table is created.
- **create abundances table.** Logical variable that determines if the abundances table is created.

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3. Here, 'not written' means that neither the variable name nor its value are written in the configuration file. If the variable name is written (and the colon) but its value is not, the corresponding Python variable will get the value `None`.

- **export abundances list.** Logical variable that allows to export a plain text file with the abundances of the studied species.
- **input files.** Nested variable for specifying the names of the input files.
  - **MADCUBA table.** Transition lines table in .csv format generated by MADCUBA. To generate it in SLIM, select one of the species in the SimFit tab to open the transition table window, check the Spectral, Fit, and Phys. options, and click the save icon to export the table in .csv format. If there are multiple sources, this variable should be a list.
  - **processing table.** Table in .csv format with information of the RMS noise and resolution of the spectra used in MADCUBA. This file can be generated by the Python Reduction Pipeline for CLASS (script processing.py) once we have run the steps --selection, --line\_search, and --reduction, adding the argument --spectra\_tables. If there are multiple sources, this variable should be a list.
  - **LaTeX template.** Template with the structure of the tables in .tex format. Each of the two possible tables must contain only one row with a specific keyword; in the case of the transitions table, it is --tablelines--, and for the abundances table, it is --tableabunds--. It can be easily done with a graphical LaTeX editor like Lyx. If this variable is not specified, a generic simple template will be automatically generated with the rows specified in the variables abundances table and lines table (see next pages).
  - **non-LTE lines table.** In case we want to manually specify the values of the transition table for one or more species (usually because they have been studied with a non-LTE model, like RADEX), we have to create that part of the table in a separated .tex file, which is the file this variable refers to. It can be easily done with a graphical LaTeX editor like Lyx.
- **output files.** Nested variable for specifying the names of the output files generated by the script.
  - **abundances list.** Plain text file in .csv format with the abundances of the studied species. If there are multiple sources, it should be a list. This file can be used as an input for the Molecular Abundances Bar Plotter (abundancesbarplot.py).
  - **LaTeX file.** Text file in .tex format containing the transitions and/or abundances tables.
- **molecules.** List of molecules studied with SLIM. Each of the elements of the list contains a variable whose name is the one that will appear on the table, and whose value is the name which appears in SLIM. For the name that will appear on the table, numbers will automatically be written as subscripts; if we want to write a superscript, we should precede the number by a hat symbol (^). If we want to specify more than one thermodynamical variants of the molecule, the element of the molecule list should be also a list containing the corresponding names (the final name and the SLIM name) for each variant.

If its value is set to 'auto', the script will search for all the transitions in the .csv

file generated by MADCUBA which were checked in the SLIM transition table window (in the .csv file this corresponds to the column CHECK). For the name that will appear on the table, it will be used the one which appears in SLIM (which in the .csv file corresponds to the column Formula) but formatted.

- **non-LTE molecules.** In case there are molecules that have been studied with a non-LTE model (like RADEX, which can be used through the RADEX Online Python Interface –script radexonline.py– or the RADEX Python Interface –script radex.py–), this nested variable specifies their parameters for the abundances table. Each next level variable name must be the name of the molecule as it appears on the molecule list (variable molecules). These variables are also nested, with the following variables:
  - **column density (/cm<sup>2</sup>).** Value and uncertainty of the column density of the molecule, in /cm<sup>2</sup>, written in a specific format. If the value has no uncertainty, just put the number. If it has uncertainty, you can join the central value and the uncertainty with +/-; for example: 5.2 +/- 0.4. If it has a lower and an upper uncertainty, you should write the lower uncertainty just after the central value preceded by -, and then write the upper uncertainty preceded by +; for example: 5.2-0.3+0.4. Finally, if you want to use scientific notation (exponential notation with decimal base), for the central value and the uncertainty (or uncertainties), you just have to write an e, separated by a space from the numbers and followed by the exponent argument; for example: 5.2-0.3+0.4 e-3. If there are multiple sources, this variable should be a list with a value for each source.
  - **kinetic temperature (K).** Value and uncertainty of the kinetic temperature of the molecule, in K. It should be specified in the same format as the column density. If there are multiple sources, it should be a list.
- **reference column density (/cm<sup>2</sup>).** Value and uncertainty of the column density of the reference molecule in the region of the observations, in /cm<sup>2</sup>. It is used to calculate the abundances of the observed species. If there are multiple sources, it should be a list.
- **lines (MHz).** Nested variable which contains a list of the transition lines that will appear on the transitions table. Each next level variable name is the one that will appear on the table. This variable is also nested, and contains another variables with the name of the molecule which appears in SLIM; in case there is more than one thermodynamical variant of the molecule, there should be a variable for each of them. Then, the value of these variables should be a list with the values of the studied transitions, in MHz. For the names that will appear on the table, numbers will automatically be written as subscripts; if we want to write a superscript, we should precede the number by a hat symbol (^).
- **lines margin (MHz).** Frequency range around the given value of the transition that will be used by the script to detect line, in MHz.
- **frequency decimals.** Number of decimals of the frequency shown in the transitions table.
- **S/N threshold.** Signal-to-noise value for line area used to determine if a line has been

properly detected or not.

- **recalculate area uncertainty.** Logical variable that determines if the line area uncertainty is recalculated instead of using the value given by MADCUBA; the expression used for the area uncertainty is  $\Delta A = \Delta T (\Delta v \delta v)^{1/2}$ , where  $\Delta T$  is the RMS noise level in the surroundings of the line,  $\Delta v$  is the velocity line width, and  $\delta v$  is the velocity resolution of the spectra which contains the line.
- **correct false detections.** Logical variable that determines if, in case a line is detected below the current signal-to-noise threshold, the upper limit (that is, the detected area) will be increased by the area uncertainty times the difference between the signal-to-noise threshold and the current signal-to-noise value.
- **abundances table.** List which defines the rows of the abundances table. Each of the elements is a list with two elements. The first one is a natural number which refers to the source of the observations (in case there is only one source, that number should always be 1); the order of the sources is the one that has been used in the variables input files and output files. The second element is a list with row names; possible names are species (species name) temperature (in K), density (column density, in  $/\text{cm}^2$ ), and abundance (abundance relative to  $\text{H}_2$ ). Then, the values for these rows will correspond to the data of the selected source. The rows defined in each element of the list abundances table will be concatenated in the same order in which they are written.
- **lines table.** List which defines the rows of the transitions table. Each of the elements is a list with two elements. The first one is a natural number which refers to the source of the observations (in case there is only one source, that number should always be 1); the order of the sources is the one that has been used in the variables input files and output files. The second element is a list with row names; possible names are species (species name) transition (transition quantum numbers), frequency (transition frequency, in MHz), intensity (line peak intensity, in K), area (line area, in  $\text{mK} \cdot \text{km/s}$ ), width (line width, in km/s), velocity (line radial velocity offset, in km/s), and sn\_area (signal-to-noise of the line area). Then, the values for these rows will correspond to the data of the selected source. The rows defined in each element of the list abundances table will be concatenated in the same order in which they are written.
- **tables scientific notation.** Nested variable which determines if scientific notation is used to represent the values of some rows of the tables. For each table, a next level variable named lines table or abundances table should be used, and it should be a list containing the names of the desired rows. Moreover, there is an additional next level variable:
  - **use crosses.** Logical variable that determines if crosses are used in scientific notation for the products; if its value is negative, dots will be used instead.
- **multiplying factors.** Nested variable that can be used to define factors that divide the values of each row, for each table (so, to recover the actual value on the final table, we should multiply by those factors). The next level variables are abundances table and lines-table. Each of these variables should be dictionaries (or nested variables) with the next level variable names being the names of the desired row and the values being the

desired factors.

## Default variable values

Below are the default values of the variables of `tablegenerator.py`, that is, the values that they will take if they are not declared in the configuration file.

```
create lines table: no
create abundances table: no
export abundances list: no
input files:
    MADCUBA table: ''
    processing table: ''
    LaTeX template: ''
    non-LTE lines table: {}
output files:
    abundances list: ''
    LaTeX file: 'tables.tex'
reference column density (/cm2): 1
lines (MHz): 'auto'
lines margin (MHz): 0.2
frequency decimals: 3
S/N threshold: 3
recalculate area uncertainty: yes
correct false detections: yes
multiplying factors:
    abundances table: {}
    lines table: {}
```

Note that this default values will only be used if neither the variable name nor its value are written in the configuration file. If the variable name is written (and the colon) but its value is not, the corresponding Python variable will get the value `None`.

## 5. Example case

Here we show a brief example to see how to use the script `tabbegenerator.py`. We will use the observations of the starless core L1517B, using molecular hydrogen ( $\text{H}_2$ ) as the reference molecular for calculating the abundances of the observed species.

### 5.1. Input files

For this example, we will use the following four files:

- **MADCUBA table.** Transition lines table in `.csv` format generated by MADCUBA.
- **processing table.** Table in `.csv` format with information of the RMS noise and resolution of the spectra used in MADCUBA, generated by the Python Reduction Pipeline for CLASS (script `processing.py`) with the argument `--spectra_tables`.
- **LaTeX template.** Template with the structure of the tables in `.tex` format, created with the LaTeX editor Lyx.
- **non-LTE lines table.** Table in `.tex` format that contains the values of the methanol transitions, which we did not study with an LTE model. It was created with the LaTeX editor Lyx.

## 5.2. Configuration file

Below is an example of configuration file which produces the two possible tables using data from two sources, and making use of the results of RADEX Online.

```
create lines table: yes
create abundances table: yes
export abundances list: yes
input files:
  MADCUBA table:
    - L1517B-madcuba.csv
    - L1517B0FF1-madcuba.csv
  processing table:
    - L1517B-processing.csv
    - L1517B0FF1-processing.csv
  LaTeX template: tables-template.tex
  non-LTE lines table:
    table-extra.tex: [CH3OH]
output files:
  abundances list:
    - L1517B-center-all.csv
    - L1517B-methanol-all.csv
  LaTeX file: tables.tex
molecules:
- - CH3OH A: CH3OH-A,vt=0-2
- - CH3OH E: CH3OH-E,vt=0-2
- CH3O: Methoxy
- CH3OCHO: CH3OCHO
- CH3OCH3: CH3OCH3
- CH3CHO: CH3CHO
- HCOOH: HCOOH
- c-C3H2O: Cyclopropenone
- CH2CO: CH2CO
- CCCO: CCCO
- HCCCHO: HCCCHO
- HCCNC: HCCNC
- CH2CHCN: C2H3CN
- CH3NC: CH3NC
- CH3CN: CH3CN
- HCCCN: HCCCN
non-LTE molecules:
  CH3OH A:
    column density (/cm2):
      - 4.30+/-0.12 e12
      - 4.96+/-0.13 e12
    kinetic temperature (K): [10, 10]
  CH3OH E:
    column density (/cm2):
      - 3.85+/-0.24 e12
      - 4.51+/-0.20 e12
    kinetic temperature (K): [10, 10]
reference column density (/cm2): [3.5e22, 0.957981e22]
lines (MHz):
  CH3OH:
    CH3OH-A,vt=0-2:
      - 96741.371
      - 95914.310
```



- 97582.798
- CH3OH-E, vt=0-2 :
  - 96739.358
  - 96744.545
  - 96755.501
- CH3O:
  - Methoxy:
    - 82455.980
    - 82458.252
    - 82471.825
    - 82524.180
- CCC0:
  - CCC0:
    - 96214.813
- HC00H:
  - HC00H:
    - 87926.863
- CH2CO:
  - CH2CO:
    - 81586.299
    - 100094.510
    - 80832.189
- CH3OCHO:
  - CH3OCHO:
    - 84454.754
- CH3OCH3:
  - CH3OCH3:
    - 82650.325
- CH3CHO:
  - CH3CHO:
    - 95963.465
- c-C3H2O:
  - Cyclopropenone:
    - 79483.520
- HCCCHO:
  - HCCCHO:
    - 83775.842
- CH3CN:
  - CH3CN:
    - 110383.500
    - 110381.372
    - 110374.989
- CH3NC:
  - CH3NC:
    - 100526.541
- CH2CHCN:
  - C2H3CN:
    - 84946.000
    - 83207.505
    - 87312.812
    - 94276.636
    - 96982.442
- HCCCN:
  - HCCCN:
    - 81881.468
    - 100076.392
- HCCNC:
  - HCCNC:
    -

```

- 79484.131
- 99354.250
lines margin (MHz): 0.2
abundances table:
- [1, ['species']]
- [1, ['temperature', 'density', 'abundance']]
- [2, ['temperature', 'density', 'abundance']]
lines table:
- [1, ['species', 'transition', 'frequency']]
- [1, ['area', 'width', 'velocity', 'sn_area']]
- [2, ['area', 'width', 'velocity', 'sn_area']]
tables scientific notation:
  lines table: []
  abundances table: ['density', 'abundance']
  use crosses: no
multiplying factors:
  abundances table: {}
  lines table: {}
S/N threshold: 3

```

### 5.3. Output tables

Tables 1 and 2 are the abundances and transitions tables, respectively, generated for the previous configuration file.

In order to copy the tables to a .tex file, you can copy the text between `\begin{table}` and `\end{table}` for the whole table including header and notes, or between `\begin{tabular}` and `\end{tabular}` for just the table frame, for each table.

**Table 1.** Abundances table generated for the previous example configuration file.

**Table.** Excitation/kinetic temperatures ( $T$ ), column densities ( $N_{\text{obs}}$ ), and abundances ( $\chi_{\text{obs}}$ ) of COMs and COM precursors toward the dust and methanol peaks in L1517B.

Molecule	$T$ (K)	Dust peak		$T$ (K)	Methanol peak	
		$N_{\text{obs}}$ ( $\text{cm}^{-2}$ )	$\chi_{\text{obs}}$		$N_{\text{obs}}$ ( $\text{cm}^{-2}$ )	$\chi_{\text{obs}}$
CH <sub>3</sub> OH A	10.0	$4.71^{+0.03}_{-0.04} \times 10^{12}$	$1.35^{+0.22}_{-0.17} \times 10^{-10}$	10.0	$(5.48 \pm 0.04) \times 10^{12}$	$5.7^{+0.6}_{-0.5} \times 10^{-10}$
CH <sub>3</sub> OH E	10.0	$4.6^{+3.3}_{-0.7} \times 10^{12}$	$1.4^{+0.9}_{-0.3} \times 10^{-10}$	10.0	$(5.6 \pm 0.3) \times 10^{12}$	$5.9^{+0.7}_{-0.6} \times 10^{-10}$
CH <sub>3</sub> OH	10.0	$9.3^{+3.3}_{-0.7} \times 10^{12}$	$2.82^{+0.91}_{-0.51} \times 10^{-10}$	10.0	$(1.11 \pm 0.03) \times 10^{13}$	$1.163^{+0.121}_{-0.113} \times 10^{-9}$
CH <sub>3</sub> O	$10 \pm 4$	$(2.8 \pm 0.9) \times 10^{11}$	$7.79^{+2.81}_{-2.41} \times 10^{-12}$	$7 \pm 4$	$(1.8 \pm 0.6) \times 10^{11}$	$1.91^{+0.69}_{-0.62} \times 10^{-11}$
CH <sub>3</sub> OCHO	7.7	$< 9 \times 10^{11}$	$< 3.5 \times 10^{-11}$	9.7	$< 1.0 \times 10^{12}$	$< 1.32 \times 10^{-10}$
CH <sub>3</sub> OCH <sub>3</sub>	7.7	$< 1.0 \times 10^{12}$	$< 4.0 \times 10^{-11}$	9.7	$< 5 \times 10^{11}$	$< 5.9 \times 10^{-11}$
CH <sub>3</sub> CHO	7.7	$< 2.5 \times 10^{11}$	$< 9.7 \times 10^{-12}$	9.7	$(2.1 \pm 0.4) \times 10^{11}$	$2.16^{+0.48}_{-0.43} \times 10^{-11}$
t-HCOOH	7.7	$< 3 \times 10^{12}$	$< 1.29 \times 10^{-10}$	9.7	$< 1.9 \times 10^{12}$	$< 2.6 \times 10^{-10}$
c-C <sub>3</sub> H <sub>2</sub> O	7.7	$< 1.5 \times 10^{10}$	$< 6.6 \times 10^{-13}$	9.7	$< 5 \times 10^{10}$	$< 6.7 \times 10^{-12}$
H <sub>2</sub> CCO	$7.7 \pm 1.0$	$(8.4 \pm 1.3) \times 10^{11}$	$2.38^{+0.54}_{-0.48} \times 10^{-11}$	$10 \pm 3$	$(7.2 \pm 1.9) \times 10^{11}$	$7.64^{+2.21}_{-2.16} \times 10^{-11}$
CCCO	7.7	$< 2.4 \times 10^{11}$	$< 9.7 \times 10^{-12}$	9.7	$< 1.2 \times 10^{12}$	$< 1.58 \times 10^{-10}$
HCCCHO	7.7	$< 1.6 \times 10^{11}$	$< 6.7 \times 10^{-12}$	9.7	$< 1.9 \times 10^{11}$	$< 2.5 \times 10^{-11}$
HCCNC	6.7	$(2.4 \pm 1.0) \times 10^{11}$	$6.7^{+3.4}_{-2.7} \times 10^{-12}$	5.3	$(1.9 \pm 0.8) \times 10^{11}$	$1.92^{+0.89}_{-0.81} \times 10^{-11}$
CH <sub>2</sub> CHCN	6.7	$< 4 \times 10^{10}$	$< 1.54 \times 10^{-12}$	5.3	$< 3 \times 10^{10}$	$< 4.2 \times 10^{-12}$
CH <sub>3</sub> NC	6.7	$(1.9 \pm 0.9) \times 10^{10}$	$5.20^{+2.93}_{-2.47} \times 10^{-13}$	5.3	$< 2.3 \times 10^{10}$	$< 3.0 \times 10^{-12}$
CH <sub>3</sub> CN	10.0	$(2.1 \pm 0.3) \times 10^{11}$	$6.07^{+1.28}_{-1.11} \times 10^{-12}$	10.0	$< 1.4 \times 10^{11}$	$< 1.89 \times 10^{-11}$
HCCCN	10.0	$(5.16 \pm 0.04) \times 10^{12}$	$1.470^{+0.263}_{-0.199} \times 10^{-10}$	10.0	$(3.72 \pm 0.09) \times 10^{12}$	$3.86^{+0.45}_{-0.34} \times 10^{-10}$

**Notes.** Temperatures ( $T$ ) refer to excitation temperatures ( $T_{\text{ex}}$ ) for all the species except for methanol (CH<sub>3</sub>OH), cyanoacetylene (HCCCN) and acetonitrile (CH<sub>3</sub>CN), where they refer to kinetic temperatures ( $T_{\text{kin}}$ ). We used MADCUBA to derive the molecular parameters from the observations except for methanol (CH<sub>3</sub>OH), cyanoacetylene (HCCCN) and acetonitrile (CH<sub>3</sub>CN), where we used RADEX. Molecular abundances were calculated using an H<sub>2</sub> column density of  $(3.5 \pm 0.5) \cdot 10^{22} / \text{cm}^2$  for the dust continuum peak and of  $(9.6 \pm 1.0) \cdot 10^{21} / \text{cm}^2$  for the position of the methanol peak. For the non-detections and also for some detections, we had to fix the excitation temperature ( $T_{\text{ex}}$ ) so that MADCUBA could fit the column density ( $N_{\text{obs}}$ ).

**Table 2.** Transition lines table generated for the previous example configuration file.**Table.** COMs and COM precursors transitions covered in our L1517B observations and their derived line parameters.

Species	Line	Frequency (MHz)	Dust peak			S/N <sup>c</sup>	Methanol peak			
			Area <sup>a</sup> (mK km s <sup>-1</sup> )	Linewidth (km s <sup>-1</sup> )	LSR velocity <sup>b</sup> (km s <sup>-1</sup> )		Area <sup>a</sup> (mK km s <sup>-1</sup> )	Linewidth (km s <sup>-1</sup> )	LSR velocity <sup>b</sup> (km s <sup>-1</sup> )	S/N <sup>c</sup>
CH <sub>3</sub> OH	2 <sub>0,2</sub> → 1 <sub>0,1</sub> A	96741.371	206.1 ± 1.9	0.278 ± 0.004	5.785 ± 0.002	109	241.1 ± 2.0	0.277 ± 0.005	5.788 ± 0.002	121
	2 <sub>1,2</sub> → 1 <sub>1,1</sub> A	95914.310	< 6	...	...	...	< 6	...	...	...
	2 <sub>1,1</sub> → 1 <sub>1,0</sub> A	97582.798	< 2.4	...	...	...	< 2.4	...	...	...
	2 <sub>1,2</sub> → 1 <sub>1,1</sub> E	96739.358	150.4 ± 1.9	0.262 ± 0.008	5.786 ± 0.003	80	181.6 ± 2.0	0.262 ± 0.003	5.785 ± 0.003	92
	2 <sub>0,2</sub> → 1 <sub>0,1</sub> E	96744.545	9.7 ± 1.9	0.244 ± 0.012	5.84 ± 0.04	5.1	11.2 ± 2.0	0.23 ± 0.08	5.78 ± 0.03	5.6
	2 <sub>1,1</sub> → 1 <sub>1,0</sub> E	96755.501	< 6	...	...	...	< 6	...	...	...
CH <sub>3</sub> O	1 <sub>0,0,6,1</sub> → 0 <sub>0,1,6,0</sub>	82455.980	< 5	...	...	...	< 4	...	...	...
	1 <sub>0,0,2,2</sub> → 0 <sub>0,1,2,1</sub>	82458.252	5.4 ± 1.2	0.41 ± 0.08	5.81 ± 0.03	4.5	4.2 ± 0.9	0.39 ± 0.08	5.82 ± 0.03	4.7
	1 <sub>0,1,2,2</sub> → 0 <sub>0,0,2,1</sub>	82471.825	5.4 ± 1.2	0.41 ± 0.08	5.81 ± 0.03	4.5	4.2 ± 0.9	0.39 ± 0.08	5.82 ± 0.03	4.7
	1 <sub>0,1,6,1</sub> → 0 <sub>0,0,6,0</sub>	82524.180	< 5	...	...	...	< 4	...	...	...
CCCO	10 → 9	96214.813	< 7	...	...	...	< 7	...	...	...
t-HCOOH	1 <sub>1,1</sub> → 0 <sub>0,0</sub>	87926.863	< 1.9	...	...	...	< 2.2	...	...	...
H <sub>2</sub> CCO	4 <sub>1,3</sub> → 3 <sub>1,2</sub>	81586.299	11.1 ± 1.3	0.53 ± 0.10	5.89 ± 0.04	8.5	12.6 ± 1.2	0.70 ± 0.25	5.95 ± 0.11	10
	5 <sub>1,5</sub> → 4 <sub>1,4</sub>	100094.510	9.9 ± 0.9	0.53 ± 0.10	5.89 ± 0.04	11	12.6 ± 1.2	0.70 ± 0.25	5.95 ± 0.11	10
	4 <sub>0,4</sub> → 3 <sub>0,3</sub>	80832.189	21 ± 4	0.53 ± 0.10	5.89 ± 0.04	5.2	17 ± 4	0.70 ± 0.25	5.95 ± 0.11	4.2
CH <sub>3</sub> OCHO	7 <sub>2,6,0</sub> → 6 <sub>2,5,0</sub>	84454.754	< 2.0	...	...	...	< 2.5	...	...	...
CH <sub>3</sub> OCH <sub>3</sub>	3 <sub>1,3,1</sub> → 2 <sub>0,2,1</sub>	82650.180	< 3	...	...	...	< 2.4	...	...	...
CH <sub>3</sub> CHO	5 <sub>1,4,0</sub> → 4 <sub>1,3,0</sub>	98900.944	< 3	...	...	...	4.0 ± 0.8	0.21 ± 0.05	5.765 ± 0.018	5.0
	5 <sub>0,5,0</sub> → 4 <sub>0,4,0</sub>	95963.459	< 7	...	...	...	< 7	...	...	...
	4 <sub>1,3,0</sub> → 3 <sub>1,2,0</sub>	79150.166	< 3	...	...	...	3.9 ± 0.7	0.21 ± 0.05	5.765 ± 0.018	5.6
	4 <sub>1,3,2</sub> → 3 <sub>1,2,2</sub>	79099.313	< 3	...	...	...	3.9 ± 0.7	0.21 ± 0.05	5.765 ± 0.018	5.6
	6 <sub>1,6</sub> → 5 <sub>1,5</sub>	79483.519	< 3	...	...	...	< 3	...	...	...
HCCCHO	9 <sub>0,9</sub> → 8 <sub>0,8</sub>	83775.816	< 2.0	...	...	...	< 2.5	...	...	...
CH <sub>3</sub> CN	6 <sub>0</sub> → 5 <sub>0</sub>	110383.500	6.4 ± 1.9	0.13 ± 0.13	5.902 ± 0.023	3.4	< 6	...	...	...
	6 <sub>1</sub> → 5 <sub>1</sub>	110381.372	11.8 ± 1.9	0.24 ± 0.08	5.81 ± 0.03	6.2	< 6	...	...	...
	6 <sub>2</sub> → 5 <sub>2</sub>	110374.989	< 10	...	...	...	< 6	...	...	...
CH <sub>3</sub> NC	5 <sub>0</sub> → 4 <sub>0</sub>	100526.541	4.2 ± 0.9	0.6 ± 0.3	< 6	4.7	< 2.2	...	...	...
CH <sub>2</sub> CHCN	9 <sub>0,9,0,8</sub> → 8 <sub>0,8,0,7</sub>	84945.988	< 2.0	...	...	...	< 2.5	...	...	...
	9 <sub>1,9,0,9</sub> → 8 <sub>1,8,0,8</sub>	83207.496	< 3	...	...	...	< 2.4	...	...	...
	9 <sub>1,8,0,9</sub> → 8 <sub>1,7,0,8</sub>	87312.806	< 1.9	...	...	...	< 2.2	...	...	...
	10 <sub>0,10,0,9</sub> → 9 <sub>0,9,0,8</sub>	94276.625	< 3	...	...	...	< 2.4	...	...	...
	10 <sub>1,9,0,10</sub> → 9 <sub>1,8,0,9</sub>	96982.430	< 4	...	...	...	< 4	...	...	...
HCCCN	9 → 8	81881.461	693.6 ± 1.0	0.395 ± 0.001	5.824 ± 0.001	694	346.7 ± 0.9	0.383 ± 0.002	5.816 ± 0.001	385
	11 → 10	100076.392	286.5 ± 0.7	0.318 ± 0.001	5.842 ± 0.001	409	96.3 ± 0.8	0.290 ± 0.003	5.837 ± 0.001	120
HCCNC	8 <sub>8</sub> → 7 <sub>8</sub>	79484.131	16.1 ± 1.1	0.31 ± 0.15	5.83 ± 0.07	15	6.9 ± 0.9	0.29 ± 0.16	5.86 ± 0.06	7.7
	10 <sub>10</sub> → 9 <sub>10</sub>	99354.250	6.8 ± 0.7	0.31 ± 0.15	5.83 ± 0.07	9.7	< 3	...	...	...

**Notes.** Line profiles were fitted using MADCUBA, except for methanol (CH<sub>3</sub>OH), cyanoacetylene (HCCCN) and acetonitrile (CH<sub>3</sub>CN), where we used CLASS. (a) Uncertainties in the line area are calculated as  $\Delta T (\Delta v \delta v)^{1/2}$ , with  $\Delta T$  the RMS noise level,  $\Delta v$  the line width, and  $\delta v$  the velocity resolution of the spectrum. Similarly, upper limits are calculated as  $3 \Delta T (\Delta v \delta v)^{1/2}$ . (b) LSR stands for *local sidereal rest*. (c) This refers to the signal to noise ratio in integrated intensity area. If a certain value has no uncertainty, this means that it had to be fixed so that MADCUBA could fit the line.

## 5.4. Output abundances list

Below is the content of one of the output abundances list files for the previous configuration file (L1517B-methanol-all.dat), that is, for one of the sources.

molecule	abundance
CH3OH	1.160-0.107+0.131 e-9
CH3O	1.91-0.62+0.68 e-11
CH3OCHO	< 1.33 e-10
CH3OCH3	< 6.3 e-11
CH3CHO	< 2.15-0.44+0.49 e-11
t-HCOOH	< 2.6 e-10
c-C3H2O	< 6.6 e-12
H2CCO	7.55-2.08+2.23 e-11
CCCO	< 1.59 e-10
HCCCHO	< 2.6 e-11
HCCNC	1.89-0.81+0.93 e-11

CH2CHCN	< 4.2 e-12
CH3NC	< 3.0 e-12
CH3CN	< 1.89 e-11
HCCCN	3.88-0.34+0.43 e-10

---

As you can see, the file content of the file is quite auto-explicative, and the values of the abundance are displayed in an easily readable format.

## Useful links

The following links may be of interest:

- **MADCUBA.**  
<https://cab.inta-csic.es/madcuba/>
- **Python.**  
<https://www.python.org/>
- **Python's standard library.**  
<https://docs.python.org/3/library/>
- **PyYAML.**  
<https://pyyaml.org/wiki/PyYAMLDocumentation/>
- **NumPy.**  
<https://numpy.org/>
- **Pandas.**  
<https://pandas.pydata.org/>
- **Lyx.**  
<https://www.lyx.org/>

## Credits

This software has been developed at the Center for Astrobiology (*Centro de Astrobiología*, CAB), in Madrid (Spain), within the subgroup of Astrochemistry and Chemical Complexity (Group of Interstellar and Circumstellar Medium, Department of Astrophysics).

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## License

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