Class 8: Mini Project

Andrew Sue

The goal of this mini-project is for you to explore a complete analysis using the unsupervised learning techniques covered in class. You'll extend what you've learned by combining PCA as a preprocessing step to clustering using data that consist of measurements of cell nuclei of human breast masses. This expands on our RNA-Seq analysis from last day.

```
# Save your input data file into your Project directory
fna.data <- read.csv("WisconsinCancer.csv", row.names=1)

# Complete the following code to input the data and store as wisc.df
wisc.df <- fna.data</pre>
```

Q1. How many observations/samples/patients/rows?

There are 569 individuals in this dataset.

Q2. What is in the \$diagnosis column? How many of each type?

There are 357, 212.

```
table(wisc.df$diagnosis)
```

```
B M
357 212
```

Q3. How many variables/features in the data are suffixed with _mean?

```
#gives you column indexes
grep("_mean",colnames(wisc.df))
```

[1] 2 3 4 5 6 7 8 9 10 11

```
#gives you values (in this case column name)
grep("_mean",colnames(wisc.df), value=TRUE)
[1] "radius_mean"
            "texture_mean"
                      "perimeter_mean"
[4] "area_mean"
            "smoothness_mean"
                      "compactness_mean"
[7] "concavity_mean"
            "concave.points_mean"
                      "symmetry_mean"
[10] "fractal_dimension_mean"
#Gives you the number of values as it counts
length(grep("_mean",colnames(wisc.df), value=TRUE))
Γ1 10
 Q. How many variables/dimensions have we?
ncol(wisc.df)
[1] 31
Save the diagnosis for reference later.
#Factors useful for categorical data and provides levels to data.
diagnosis <- as.factor(wisc.df$diagnosis)</pre>
diagnosis
```

Levels: B M

Remove or exclude this column from any of our analysis

```
wisc.data <- wisc.df[,-1]
head(wisc.data)</pre>
```

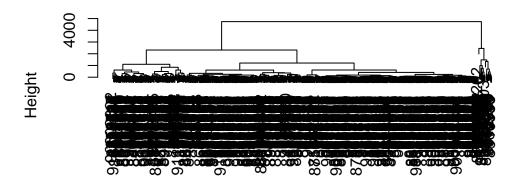
	radius_mean text	ure_mean	perimet	er_mean	area_mean	smoothn	ess_mean
842302	17.99	10.38		122.80	1001.0	1	0.11840
842517	20.57	17.77		132.90	1326.0	1	0.08474
84300903	19.69	21.25		130.00	1203.0	1	0.10960
84348301	11.42	20.38		77.58	386.1		0.14250
84358402	20.29	14.34		135.10	1297.0	1	0.10030
843786	12.45	15.70		82.57	477.1		0.12780
	compactness_mean	concavit	ty_mean	concave.	points_me	an symme	try_mean
842302	0.27760		0.3001		0.147	10	0.2419
842517	0.07864		0.0869		0.070	17	0.1812
84300903	0.15990		0.1974		0.127	90	0.2069
84348301	0.28390		0.2414		0.105	20	0.2597
84358402	0.13280		0.1980		0.104	30	0.1809
843786	0.17000		0.1578		0.080	89	0.2087
	fractal_dimensio	n_mean ra	adius_se	texture	e_se perim	eter_se	area_se
842302	0	.07871	1.0950	0.9	9053	8.589	153.40
842517	0	.05667	0.5435	0.7	7339	3.398	74.08
84300903	0	.05999	0.7456	0.7	7869	4.585	94.03
84348301	0	.09744	0.4956	1.1	L560	3.445	27.23
84358402	0	.05883	0.7572	0.7	7813	5.438	94.44
843786	0	.07613	0.3345	0.8	3902	2.217	27.19
	smoothness_se co	mpactness	s_se con	cavity_s	se concave	.points_	se
842302	0.006399	0.04	1904	0.0537	73	0.015	87
842517	0.005225	0.01	L308	0.0186	30	0.013	340
84300903	0.006150	0.04	1006	0.0383	32	0.020	58
84348301	0.009110	0.07	7458	0.0566	31	0.018	67
84358402	0.011490	0.02	2461	0.0568	38	0.018	85
843786	0.007510	0.03	3345	0.0367	72	0.011	.37
	symmetry_se frac	tal_dimer	nsion_se	radius_	_worst tex	ture_wor	st
842302	0.03003	(0.006193		25.38	17.	33
842517	0.01389	(0.003532		24.99	23.	41
84300903	0.02250	(0.004571		23.57	25.	53

84348301 0.05963	0.009208		14.91		26.50
84358402 0.01756	0.005115		22.54		16.67
843786 0.02165	0.0	0.005082		17	23.75
perimeter_worst	area_worst	smoothness_worst compactn		compactne	ess_worst
842302 184.60	2019.0		0.1622		0.6656
842517 158.80	1956.0		0.1238		0.1866
84300903 152.50	1709.0		0.1444		0.4245
84348301 98.87	567.7		0.2098		0.8663
84358402 152.20	1575.0	0.1374		0.2050	
843786 103.40	741.6	0.1791			0.5249
concavity_worst	concave.po	ints_worst	symmetr	ry_worst	
842302 0.7119)	0.2654		0.4601	
842517 0.2416	;	0.1860		0.2750	
84300903 0.4504	=	0.2430		0.3613	
84348301 0.6869)	0.2575		0.6638	
84358402 0.4000)	0.1625		0.2364	
843786 0.5355	,	0.1741		0.3985	
fractal_dimensi	on_worst				
842302	0.11890				
842517	0.08902				
84300903	0.08758				
84348301	0.17300				
84358402	0.07678				
843786	0.12440				

Lets try clustering this data:

```
wisc.hc<-hclust(dist(wisc.data))
plot(wisc.hc)</pre>
```

Cluster Dendrogram



dist(wisc.data)
hclust (*, "complete")

#Principal Component Analysis

Let's try PCA on this data. Before doing any analysis like this we should check if our input data needs to be scaled first? Scaling in data for PCA is important as PCA looks at variance, thus a category with the largest variance within it will dominate PCA.

Check column means and standard deviations
colMeans(wisc.data)

perimeter_mean	texture_mean	radius_mean
9.196903e+01	1.928965e+01	1.412729e+01
${\tt compactness_mean}$	${\tt smoothness_mean}$	area_mean
1.043410e-01	9.636028e-02	6.548891e+02
${ t symmetry_mean}$	concave.points_mean	concavity_mean
1.811619e-01	4.891915e-02	8.879932e-02
texture_se	radius_se	$fractal_dimension_mean$
1.216853e+00	4.051721e-01	6.279761e-02
smoothness_se	area_se	perimeter_se
7.040979e-03	4.033708e+01	2.866059e+00
concave.points_se	concavity_se	compactness_se
1.179614e-02	3.189372e-02	2.547814e-02
radius_worst	fractal_dimension_se	symmetry_se

```
2.054230e-02
                                3.794904e-03
                                                         1.626919e+01
       texture_worst
                             perimeter_worst
                                                           area_worst
        2.567722e+01
                                1.072612e+02
                                                         8.805831e+02
    smoothness_worst
                           compactness_worst
                                                      concavity_worst
        1.323686e-01
                                2.542650e-01
                                                         2.721885e-01
concave.points_worst
                              symmetry_worst fractal_dimension_worst
        1.146062e-01
                                2.900756e-01
                                                         8.394582e-02
```

apply(wisc.data,2,sd)

perimeter_mean	texture_mean	radius_mean
2.429898e+01	4.301036e+00	3.524049e+00
compactness_mean	smoothness_mean	area_mean
5.281276e-02	1.406413e-02	3.519141e+02
symmetry_mean	concave.points_mean	concavity_mean
2.741428e-02	3.880284e-02	7.971981e-02
texture_se	radius_se	<pre>fractal_dimension_mean</pre>
5.516484e-01	2.773127e-01	7.060363e-03
smoothness_se	area_se	perimeter_se
3.002518e-03	4.549101e+01	2.021855e+00
concave.points_se	concavity_se	compactness_se
6.170285e-03	3.018606e-02	1.790818e-02
radius_worst	${\tt fractal_dimension_se}$	symmetry_se
4.833242e+00	2.646071e-03	8.266372e-03
area_worst	perimeter_worst	texture_worst
5.693570e+02	3.360254e+01	6.146258e+00
concavity_worst	${\tt compactness_worst}$	smoothness_worst
2.086243e-01	1.573365e-01	2.283243e-02
${\tt fractal_dimension_worst}$	symmetry_worst	concave.points_worst
1.806127e-02	6.186747e-02	6.573234e-02

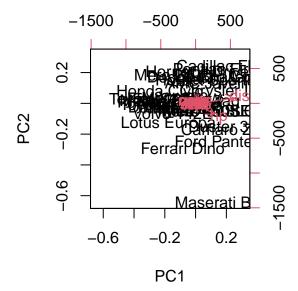
Use mtcars dataframe as an example of analysis with small dataset.

```
pc <- prcomp(mtcars)
summary(pc)</pre>
```

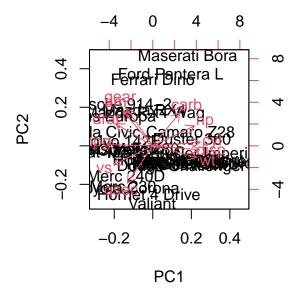
Importance of components:

PC1 PC2 PC3 PC4 PC5 PC6 PC7
Standard deviation 136.533 38.14808 3.07102 1.30665 0.90649 0.66354 0.3086
Proportion of Variance 0.927 0.07237 0.00047 0.00008 0.00004 0.00002 0.0000

biplot(pc)



pc.scale <- prcomp(mtcars,scale =TRUE)
biplot(pc.scale)</pre>



Q. Do we need to scale out cancer data set?

Yes we do! If you look at the standard deviation between the variance is too large to compare.

```
wisc.pr <- prcomp(wisc.data, scale=TRUE)</pre>
```

How well do the PCs capture the variance?

```
summary(wisc.pr)
```

Importance of components:

```
PC1
                                  PC2
                                          PC3
                                                  PC4
                                                           PC5
                                                                   PC6
                                                                           PC7
Standard deviation
                       3.6444 2.3857 1.67867 1.40735 1.28403 1.09880 0.82172
Proportion of Variance 0.4427 0.1897 0.09393 0.06602 0.05496 0.04025 0.02251
Cumulative Proportion 0.4427 0.6324 0.72636 0.79239 0.84734 0.88759 0.91010
                           PC8
                                   PC9
                                          PC10
                                                 PC11
                                                          PC12
                                                                  PC13
                                                                          PC14
Standard deviation
                       0.69037 0.6457 0.59219 0.5421 0.51104 0.49128 0.39624
Proportion of Variance 0.01589 0.0139 0.01169 0.0098 0.00871 0.00805 0.00523
Cumulative Proportion
                       0.92598 \ 0.9399 \ 0.95157 \ 0.9614 \ 0.97007 \ 0.97812 \ 0.98335
                                                            PC19
                           PC15
                                   PC16
                                           PC17
                                                    PC18
                                                                    PC20
                                                                           PC21
Standard deviation
                       0.30681 0.28260 0.24372 0.22939 0.22244 0.17652 0.1731
Proportion of Variance 0.00314 0.00266 0.00198 0.00175 0.00165 0.00104 0.0010
Cumulative Proportion 0.98649 0.98915 0.99113 0.99288 0.99453 0.99557 0.9966
```

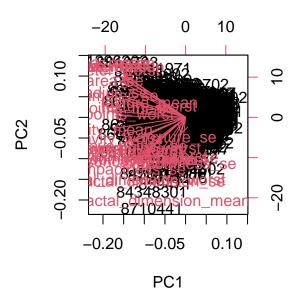
PC22 PC23 PC24 PC25 PC26 PC27 PC28 Standard deviation 0.16565 0.15602 0.1344 0.12442 0.09043 0.08307 0.03987 Proportion of Variance 0.00091 0.00081 0.0006 0.00052 0.00027 0.00023 0.00005 Cumulative Proportion 0.99749 0.99830 0.9989 0.99942 0.99969 0.99992 0.99997 PC29 PC30 Standard deviation 0.02736 0.01153 Proportion of Variance 0.00002 0.00000 Cumulative Proportion 1.00000 1.00000

- Q4. From your results, what proportion of the original variance is captured by the first principal components (PC1)?
- 44.27% of original variance is calculated in PC1.
 - Q5. How many principal components (PCs) are required to describe at least 70% of the original variance in the data?

The first 3 PCs are required to get 70% of the variance.

Q6. How many principal components (PCs) are required to describe at least 90% of the original variance in the data?

You need 7 PCs to get 90% of the variance.



Q7. What stands out to you about this plot? Is it easy or difficult to understand? Why?

This plot is unreadable and not useful as everything is on top of each other. This is because it is graphing all of the categories rather than the most principal components.

Our main PC score plot (a.k.a PC plot, PC1 vs PC2, ordination plot).

```
attributes(wisc.pr)
```

\$names

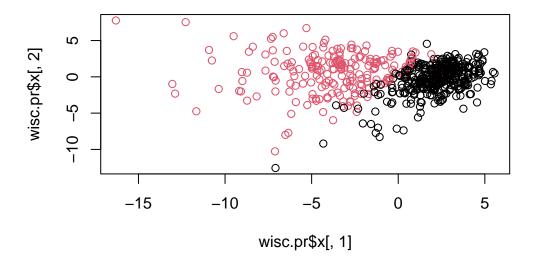
[1] "sdev" "rotation" "center" "scale" "x"

\$class

[1] "prcomp"

We need to build our own plot here:

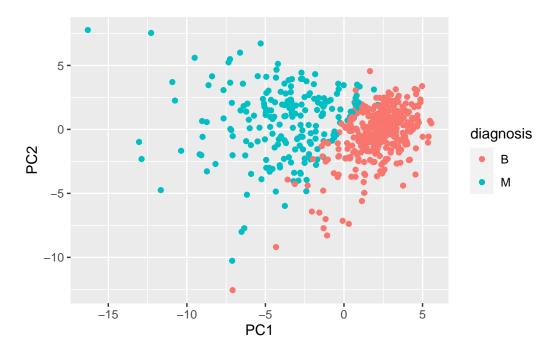
```
plot(wisc.pr$x[,1], wisc.pr$x[,2],col=diagnosis)
```



wisc.pr\$x

Make a nice ggplot version of plot

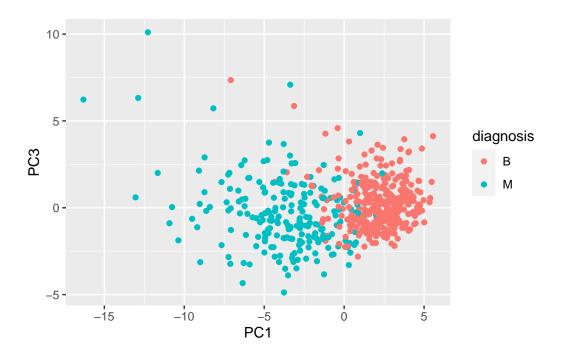
```
pc <-as.data.frame(wisc.pr$x)
library(ggplot2)
ggplot(pc) +
   aes(PC1, PC2, col=diagnosis) +
   geom_point()</pre>
```



Q8. Generate a similar plot for principal components 1 and 3. What do you notice about these plots?

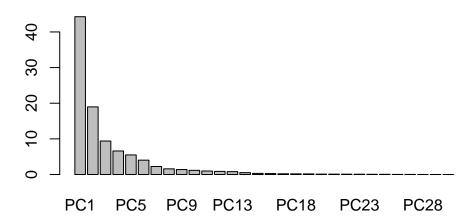
While they are similar, there is slightly less variance within the PC1 and PC3 than the previous. Which makes sense since PC3 has less influence than PC2.

```
pc <-as.data.frame(wisc.pr$x)
library(ggplot2)
ggplot(pc) +
   aes(PC1, PC3, col=diagnosis) +
   geom_point()</pre>
```



Make scree plot of variance

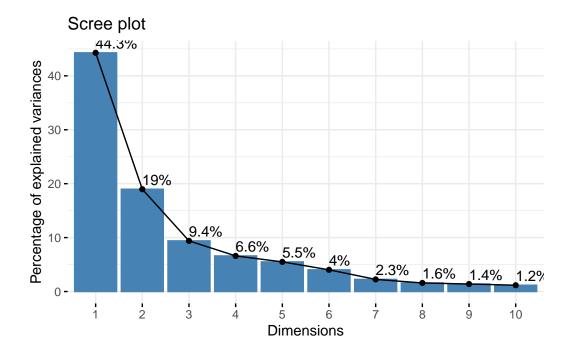
```
wisc.scree<-summary(wisc.pr)
# wisc.scree
barplot(wisc.scree$importance[2,]*100) #graphing only proportion variance</pre>
```



#factoextra package automatically calculates variance within it (doing what you did above library(factoextra)

 ${\tt Welcome!\ Want\ to\ learn\ more?\ See\ two\ factoextra-related\ books\ at\ https://goo.gl/ve3WBa}$

fviz_eig(wisc.pr, addlabels = TRUE)



Q9. For the first principal component, what is the component of the loading vector (i.e. wisc.pr\$rotation[,1]) for the feature concave.points_mean? This tells us how much this original feature contributes to the first PC.

It is contributing 1.6% to the original PC1. (Category/Dimension 8)

Hierarchical clustering

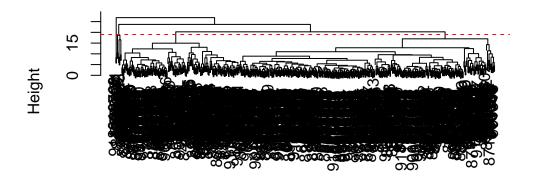
```
data.scaled <-scale(wisc.data)
data.dist <-dist(data.scaled)
wisc.hclust<- hclust(data.dist,method ="complete")</pre>
```

Q10. Using the plot() and abline() functions, what is the height at which the clustering model has 4 clusters?

```
Height = 19
```

```
plot(wisc.hclust)
abline(h=19,col="red", lty=2)
```

Cluster Dendrogram



data.dist hclust (*, "complete")

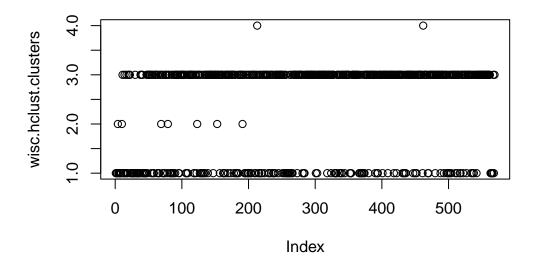
This exercise will help you determine if, in this case, hierarchical clustering provides a promising new feature.

Use cutree() to cut the tree so that it has 4 clusters. Assign the output to the variable wisc.hclust.clusters.

```
wisc.hclust.clusters <- cutree(wisc.hclust, 4)
table(wisc.hclust.clusters, diagnosis)</pre>
```

```
\begin{array}{c|cccc} & \text{diagnosis} \\ \text{wisc.hclust.clusters} & \text{B} & \text{M} \\ & 1 & 12 & 165 \\ & 2 & 2 & 5 \\ & 3 & 343 & 40 \\ & 4 & 0 & 2 \\ \end{array}
```

```
plot(wisc.hclust.clusters)
```

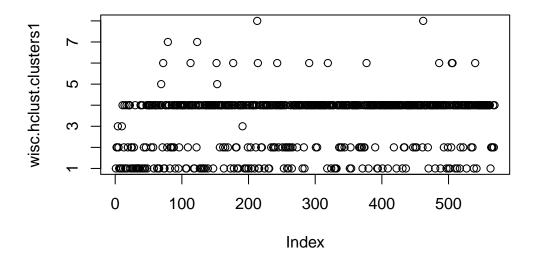


Q11. OPTIONAL: Can you find a better cluster vs diagnoses match by cutting into a different number of clusters between 2 and 10? How do you judge the quality of your result in each case?

In this case, it would be visually inspecting the tree and/or calling the predicted diagnosis to the actual to see if you are become farther due to the cuts. using the complete method, cutting at 4 seems to make the most sense.

```
wisc.hclust.clusters1 <- cutree(wisc.hclust,8)
table(wisc.hclust.clusters1, diagnosis)</pre>
```

```
diagnosis
wisc.hclust.clusters1
                            В
                                 М
                           12
                                86
                        2
                            0
                                79
                        3
                            0
                                 3
                        4
                          331
                                39
                        5
                            2
                                 0
                           12
                        6
                                 1
                        7
                            0
                                 2
                            0
                                 2
                        8
```



Q12. Which method gives your favorite results for the same data.dist dataset? Explain your reasoning.

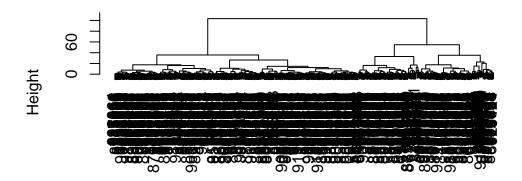
ward.D2 is better as it clustered better and minimized the variance between better.

Using 3 PCs

We start with using 3 PCs $\,$

```
wisc.pr.hclust<- hclust(dist(wisc.pr$x[,1:3]), method="ward.D2")
plot(wisc.pr.hclust)</pre>
```

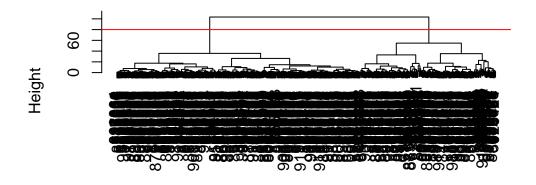
Cluster Dendrogram



dist(wisc.pr\$x[, 1:3]) hclust (*, "ward.D2")

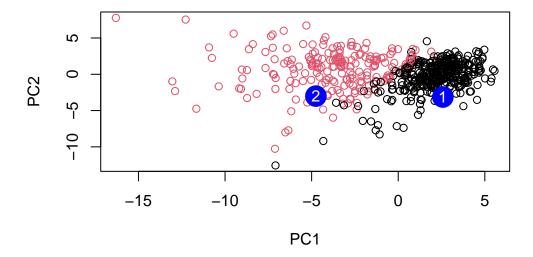
plot(wisc.pr.hclust)
abline(h=80, col="red") #abline just draws a line across

Cluster Dendrogram



dist(wisc.pr\$x[, 1:3]) hclust (*, "ward.D2")

```
grps<- cutree(wisc.pr.hclust, h=80)</pre>
  table(grps)
grps
     2
 1
203 366
  table(grps, diagnosis)
   diagnosis
      В
grps
          М
  1 24 179
  2 333 33
  #url <- "new_samples.csv"</pre>
  url <- "https://tinyurl.com/new-samples-CSV"</pre>
  new <- read.csv(url)</pre>
  npc <- predict(wisc.pr, newdata=new)</pre>
  npc
          PC1
                    PC2
                               PC3
                                         PC4
                                                   PC5
                                                              PC6
                                                                         PC7
[1,] 2.576616 -3.135913 1.3990492 -0.7631950 2.781648 -0.8150185 -0.3959098
[2,] -4.754928 -3.009033 -0.1660946 -0.6052952 -1.140698 -1.2189945 0.8193031
                                                  PC12
           PC8
                     PC9
                               PC10
                                        PC11
                                                            PC13
[1,] -0.2307350 0.1029569 -0.9272861 0.3411457 0.375921 0.1610764 1.187882
[2,] -0.3307423 0.5281896 -0.4855301 0.7173233 -1.185917 0.5893856 0.303029
                    PC16
                                PC17
                                           PC18
                                                       PC19
[1,] 0.3216974 -0.1743616 -0.07875393 -0.11207028 -0.08802955 -0.2495216
PC22
                                PC23
                                          PC24
                                                      PC25
          PC21
[1.] 0.1228233 0.09358453 0.08347651 0.1223396 0.02124121 0.078884581
[2,] -0.1224776 0.01732146 0.06316631 -0.2338618 -0.20755948 -0.009833238
            PC27
                        PC28
                                    PC29
                                                 PC30
[1,] 0.220199544 -0.02946023 -0.015620933 0.005269029
[2,] -0.001134152  0.09638361  0.002795349 -0.019015820
  plot(wisc.pr$x[,1:2], col=diagnosis)
  points(npc[,1], npc[,2], col="blue", pch=16, cex=3)
```



Q16. Which of these new patients should we prioritize for follow up based on your results?

We would prioritize patient 2 given that is falls within the malignant side.