# Lab 2: Spectrophotometry

### Data

Create two datasets: 1-standard curve, 2-plant extract measurements. The columns represent the following:

BSA\_conc: concentrations of BSA protein in micrograms/ml, prepared during the lab.

BSAabsorption: absorbtion of BSA samples at 595 nm light, measured during the lab on spectrophotometer.

- p\_ratio: plant extract diluted samples, known dilution ratios, but unknown concentration.
- **p\_absorption**: absorption of plant extract samples.
- **p\_concentration**: concentration of proteins in the plant extract samples, set to NA as they are unknown so far, and will be calculated later.

##		BSA_conc	BSAabsorption
##	1	0	0.300
##	2	0	0.308
##	3	5	0.409
##	4	5	0.438
##	5	10	0.516
##	6	10	0.530
##	7	15	0.599
##	8	15	0.600
##	9	20	0.685
##	10	20	0.696
##	11	25	0.745
##	12	25	0.764

extract

##		p_ratio	<pre>p_absorption</pre>	<pre>p_concentration</pre>
##	1	1/10	1.249	NA
##	2	1/10	1.289	NA
##	3	1/50	0.654	NA
##	4	1/50	0.576	NA
##	5	1/100	0.493	NA
##	6	1/100	0.476	NA

## Standard Curve

Build a standard curve (linear regression) using BSA concentration and absorption values.

```
curve=lm(BSAabsorption~BSA_conc, data=std_data)
paste0("y = ",round(curve[[1]][1],4)," + ",round(curve[[1]][2],4),"*x", sep="") # linear equation expla
## [1] "y = 0.3256 + 0.0179*x"

r2=summary(curve)$r.squared
paste0("r2 = ", round(r2,3))
```

```
## [1] "r2 = 0.988"
```

#### Plant extract concentrations

Find concentrations of proteins in plant extract. By looking at the equation of our standard curve, we know the concentrations of BSA and their absorbance. Our goal is to find the concentrations of our plant extract (x values in the equation), based on the absorbance we measured (y values). Therefore, in the equation: y = 0.33 + 0.0179 \* x,

the known is y, and we want to solve for the unknown x (concentration of protein plant extract): x = (y - 0.33)/0.0179

extract\$p\_concentration=(extract\$p\_absorption-curve[[1]][1])/curve[[1]][2]
extract

```
##
     p_ratio p_absorption p_concentration
## 1
        1/10
                     1.249
                                  51.628062
## 2
                                  53.864483
        1/10
                     1.289
## 3
        1/50
                     0.654
                                  18.361289
## 4
        1/50
                     0.576
                                  14.000266
## 5
       1/100
                     0.493
                                   9.359691
## 6
       1/100
                     0.476
                                   8.409212
```

Convert dilutions of our samples back to the true concentration value. We know that we diluted sample 1/10 ten times, 1/50 fifty, and so on. To find the concentration of the original extract (the one we tested today, and saved for next time), we need to multiply each calculated concentration by the respective dilution factor (10, 50, 100).

The variability in the numbers found is a measurement error, and we can take a mean, and sd to quantify it. Note: at this step we exclude plant samples with the ratio of 1/10 as their values were too high to be used with our standard curve.

```
sample=extract$p_concentration[3:6]*c(50,50,100,100)
sample

## [1] 918.0644 700.0133 935.9691 840.9212
c(mean=mean(sample),sd=sd(sample))

## mean sd
```

#### Conclusion

## 848.7420 107.3865

We conclude that the concentration of proteins in the plant extract is about 848.7 micrograms/ml, which is the same as 0.8487 mg/ml.

#### Plotting the standard curve

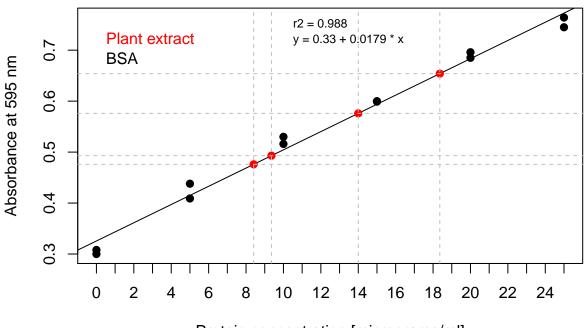
We can build a plot of a standard curve with plant extract data points on it.

Note: the line represents a simple least squares regression based on all BSA points; points of the unknown (plant extract in this case) will always be on the line because the we use the equation to find concentrations. Note: two points of plant extract are not on the plot as they were beyond the x range of the standard curve. We cannot assume that the relationship is linear outside of the tested range.

```
plot(std_data$BSAabsorption~std_data$BSA_conc,xlab="Protein concentration [micrograms/ml]",ylab = "Absorption"
#grid()+
```

```
abline(curve[[1]][1],curve[[1]][2])+
points(extract$p_absorption~extract$p_concentration,col="red",pch=19)+
text(x=0,y=0.72, labels = ("Plant extract"),col="red",pos=4)+
text(x=0,y=0.68, labels="BSA", col="Black",pos=4)+
text(10,0.75,labels=paste0("r2 = ", round(r2,3)),cex=0.75,pos=4)+
text(10,0.72,labels=c("y = 0.33 + 0.0179 * x"),cex=0.75,pos=4)+
abline(h=extract$p_absorption,v=extract$p_concentration,col="gray",lty="dashed")
```

# **Protein absorbance**



Protein concentration [micrograms/ml]

## integer(0)