# **MALARIA DETECTION**



ANGELA TSENG x DEEP LEARNING

## **Executive Summary**

Problem: Malaria remains a global health challenge, with over 229M cases and 400,000 deaths in 2019.

Solution: Early and accurate diagnosis is vital to treatment. I developed a deep learning solution to automate blood cell image analysis for malaria in order to accelerate manual microscopy.

**Key Findings:** I evaluated base plus four model variants. **Model 3** emerged as the optimal solution, delivering **98.6% test accuracy with the highest recall for parasitized cells (99%) and a lightweight architecture.** It catches virtually all infections with minimal false negatives, a priority for patient safety.

**Recommendation**: **Adopt Model 3 in the malaria detection workflow** to minimize missed infections and enable fast, cost-effective deployment. Labs and clinics can dramatically speed up diagnosis, achieve higher accuracy, and mass deploy malaria screening in resource-constrained regions.

**Expected Benefits**: Implementing Model 3 could reduce image review from minutes to seconds, **enabling higher throughput and faster treatment decisions**. Eliminating inter-observer variability will **improve reliability, increase early malaria detections, save expert labor, and improve patient outcomes**, aligning both public health and business efficiency goals.

### **Problem Definition**



# Malaria is a contagious disease

Caused by Plasmodium parasites transmitted to humans through infected mosquitoes bites



# Half of the world's population is at risk

Especially in sub-Saharan Africa. Over 229 million cases and 400,000 deaths reported in 2019



# Children under 5 are the most vulnerable

Accounted for 67% of malaria deaths worldwide in 2019



# Traditional diagnosis ismanual & slow

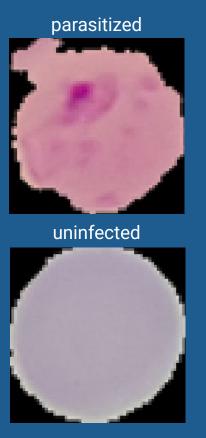
Delays and inter-observer variability lead to missed diagnoses and continued high mortality

### Problem to Solve

**Project Goal:** Build an automated malaria detection **deep learning** model to classify red blood cell images as parasitized vs uninfected and exceed expert-level accuracy and speed.

**Target Outcome**: A model that reduces diagnostic time, improves detection sensitivity, and flags infected cells with high confidence. Success means **fewer cases of malaria go undetected**.

Rationale: Even a small improvement in speed or sensitivity can translate into millions of lives saved and more efficient use of limited resources.



Dataset with **24,958 train** and **2,600 test** images

### Solution Approach

Exploratory Data
Analysis

Data
Preprocessing

Model Building

Choosing the
Best Model

Conclusion &
Recommendati
on

- Load the data
- Data overview
- Visualize data
- Summarize key findings
- Convert RGB to HSV images
- Gaussian blurring
- Split the data
- One hot encoding

- Build base model
- Check model performance
- Tune model with 4 variations
- Define success metrics
- Compare model performance
- Choose the best model
- Final solution design & business recommendations
- Refine insights
- Understand risks and challenges

# Key Findings: Exploratory Data Analysis

**Examine dataset shape** 

24,958 train and 2,600 test images; size 64×64 pixels with 3 color channels. Labels are binary (parasitized vs uninfected)

**Check for class imbalance** 

No significant imbalance (parasitized v uninfected are roughly 50/50)

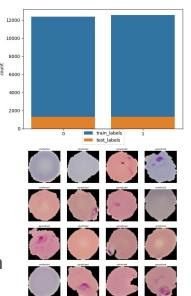
Normalize & visualize images

Pixel values scaled to 0–1 for faster, more stable model training

**Observe visual distinctions** 

Shapes and colors don't signal infection.

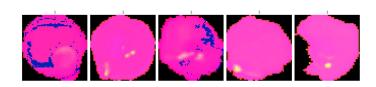
Parasitized cells have **darker spots within** 



# **Key Findings: Data Preprocessing**

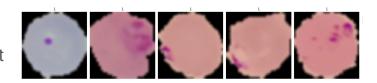
**Convert RGB to HSV images** 

Significant color separation and intensity (blue & yellow spots) inside infected cells



**Gaussian blurring** 

Reduce noise while retaining parasite blobs-no new insight



**Split the data** 

Partition training data into train and validation sets (80/20 split)

One hot encoding

Encode the class labels for binary, 0/1 encoding

Takeaway: Visually examining the images reveals good data quality and clear distinguishing features conducive for model training

# Key Findings: Model Building

**Base Model** 

A simple CNN with 3 conv layers (each followed by max pooling) and 2 dense layers. This is my baseline performance.

Model 1

Increase depth - 4 conv layers and 3 dense layers - to explore if a deeper network improves accuracy.

Model 2

Improve activation & regularization - 3 conv layers, each with BatchNorm, LeakyRelu, max pooling, and a dense layer (BN+LR).

Model 3

Augment data with optimized architecture - 3 conv layers with more filters and kernel size (with max pooling) and a smaller dense layer.

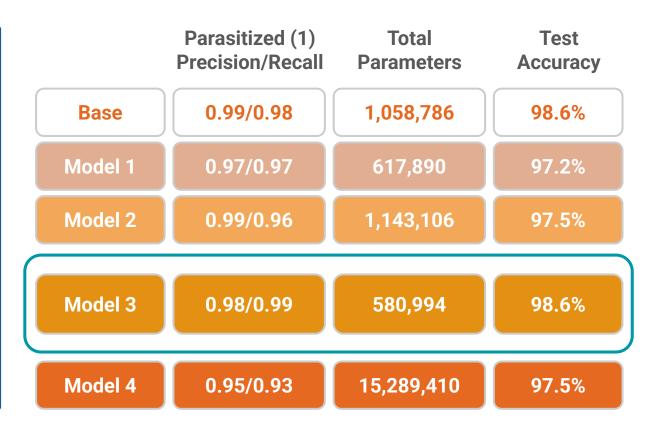
Model 4

Transfer learning approach - extract and flatten pretrained VGG16 network and add 3 dense layers with BatchNorm to classify malaria.

# Key Findings: Choosing the Best Model

#### **Decision Criteria**

- Recall on parasitized cell is the most critical metric
- Missing an infection (false negative) is a worse outcome than a false alarm
- Model parameter counts (indicate size) affect deployment feasibility
- Test accuracy



### Model 3: Model Selection Rationale

#### 1. Aggressive data augmentation

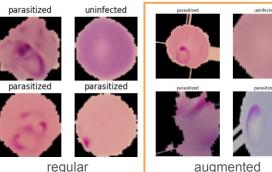
- Flipping, rotating, and zooming focus model on intrinsic cell features; improve rotation- and translation-invariance
- Benefits outweigh feature loss

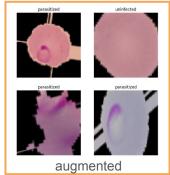
### 2. Compact architecture with 45% less parameters

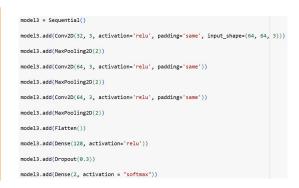
- Increase layer filters to 64
- Larger kernel 3x3 for richer feature extraction
- Reduce dense layer to 128 neurons to avoid overfitting

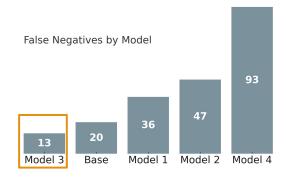
### 3. Recall maximization for infected cells

- Model catches almost every infected cell, resulting in very few false negatives
- Missing an infection could lead to late treatment and mortality









### **Business Recommendation**

#### **Data Science & IT**

Prepare model for deployment (set up servers or embed in device). Establish a pipeline of new blood smear images.



#### Management

Allocate budget and resources for necessary equipment and training. Monitor program implementation and impact.

#### **Lab Operations**

Develop new procedures to integrate AI detection with human review. Train technicians to use the new system.



#### **Partners and Regulators**

Align community partners on Al detection benefits. Engage medical device regulators to ensure policy compliance

# **Cost and Benefit Analysis**

	COST	BENEFIT
Development & Integration	One-time cost. Integrate Model 3 into user-friendly applications for technicians, and to set up necessary infrastructure.	Improved Speed & Scale - Model 3 can be deployed in multiple locations on standard hardware without expensive GPU servers.
Hardware & Equipment	Fixed cost. Digital microscopy equipment and inexpensive computing devices. Can repurpose existing lab infrastructure without expansion.	Enhanced Accuracy & Consistency - Process bulk image volume without additional cost. Images can be reused for further analysis.
Training & Adoption	Upfront cost. Technicians will need training to learn the new system. Anticipate initial slowdown and resistance to change.	Lab Efficiency & Labor Savings - Remove mundane tasks and free up skilled labor for patient care and confirming false positives.
Maintenance & Updates	Ongoing cost. Model retraining needed if new data emerges - when parasite mutates, adapt to new hospital systems.	<b>Strategic Advantage</b> - Easy to retrain Model 3 for sustained accuracy and maintain leading edge of delivering high-quality medical service.
False Positive Follow up	Time for an expert to quickly double-check false alarms either with a human review or a secondary test.	Save Lives - Rare false positives with Model 3 and faster/cheaper to double check result than missing an infection that becomes fatal.

# Implementation Risk and Challenges

- **Data Shift Risk:** Images from different
- hospitals, microscopes, staining techniques may differ widely to degrade performance
  - **Computing Constraints:** Limited compute
- resources (power & device) or internet connectivity in remote areas
  - **Ongoing Maintenance:** Software bug or
- hardware failure could disrupt diagnoses without a support or fallback process
  - **User Adoption & Training:** Technicians may
- show reluctance to change routine if tool is not user-friendly and benefits are unclear

- False Positives Burden: Redesign workflow
- so that images flagged are efficiently reviewed and cleared with minimal effort
  - False Sense of Security: Over-reliance on Al
- so technicians skip further review such as spot-checking and additional testing
  - Regulatory and Ethical Risks: Maintain
- transparency, data privacy, and document results to avoid oversight and liability risks

### **Next Steps**

Pilot Deployment: Limited-scope Model 3 deployment in a controlled setting. Run model in parallel with standard diagnostics as a proof-of-concept to uncover and address real-world issues before scaling up.

Gather Additional Data & Retraining: Expand training dataset to fine-tune Model 3 to adapt to local specificities. Plan for periodic retraining and evaluation if new data indicates changes.

**Develop Deployment Infrastructure**: Cloud-based API or model running on local device. Integrate software with existing lab information systems and workflow.

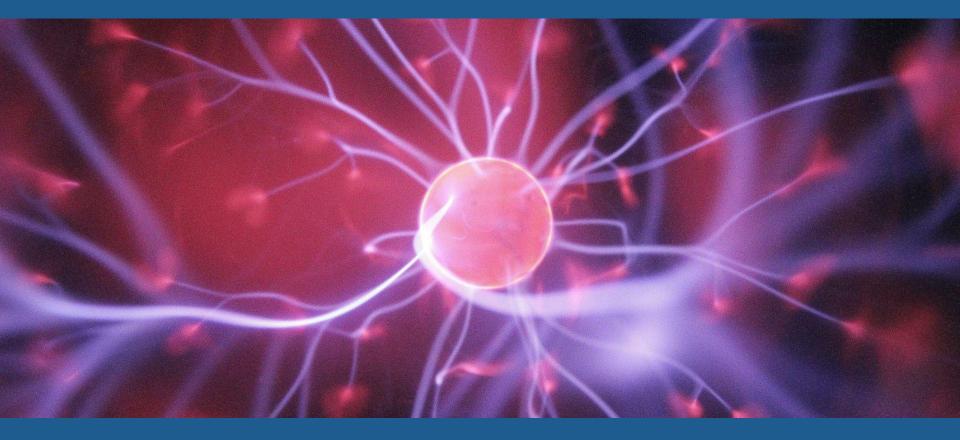
**User Training & Documentation:** Include a quick-start guide, result interpretation, and procedure for handling positive cases. Conduct training at pilot site to get technicians comfortable with the tool.

Establish Feedback Loop: Dashboard to track key metrics for troubleshooting and model improvement.

**Iterate and Scale Up**: Deploy model to other labs and regions in a phased manner.

Communicate Success: Secure ongoing support, raise team motivation, and attract partnerships.

# **THANK YOU**

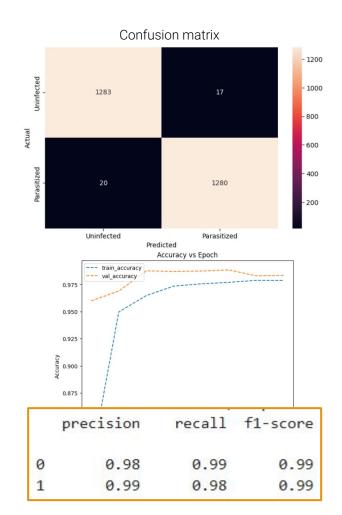


**QUESTIONS?** 

## Appendix: Base Model

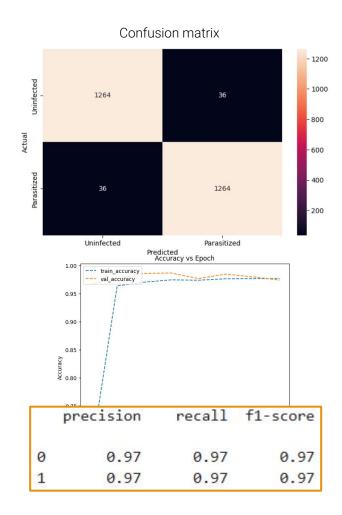
Observations: 98.6% test accuracy; high recall; simple design, some inefficiencies - low filter/small kernel/large dense layer

```
model = Sequential()
model.add(Conv2D(filters = 32, kernel size = 2, padding = "same", activation = "relu", input shape = (64, 64, 3)))
model.add(MaxPooling2D(pool size = 2))
model.add(Dropout(0.2))
model.add(Conv2D(filters = 32, kernel size = 2, padding = "same", activation = "relu"))
model.add(MaxPooling2D(pool size = 2))
model.add(Dropout(0.2))
model.add(Conv2D(filters = 32, kernel size = 2, padding = "same", activation = "relu"))
model.add(MaxPooling2D(pool size = 2))
model.add(Dropout(0.2))
model.add(Flatten())
model.add(Dense(512, activation = "relu"))
model.add(Dropout(0.4))
model.add(Dense(2, activation = "softmax"))
```



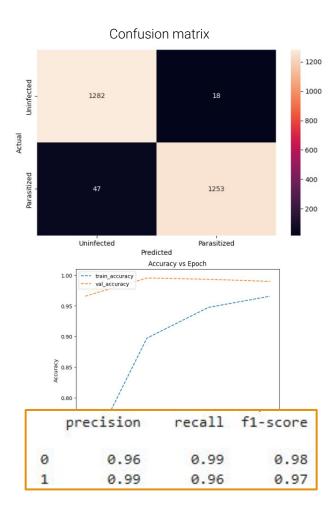
**Observations: 97.2% test accuracy, lower recall;** deeper not wider; more aligned train/val accuracy; sub optimized new layers/dropout

```
model1 = Sequential()
model1.add(Conv2D(filters = 32, kernel size = 2, padding = "same", activation = "relu", input shape = (64, 64, 3)))
model1.add(MaxPooling2D(pool_size = 2))
model1.add(Dropout(0.2))
model1.add(Conv2D(filters = 32, kernel_size = 2, padding = "same", activation = "relu"))
model1.add(MaxPooling2D(pool_size = 2))
model1.add(Dropout(0.2))
model1.add(Conv2D(filters = 32, kernel_size = 2, padding = "same", activation = "relu"))
model1.add(MaxPooling2D(pool_size = 2))
model1.add(Dropout(0.2))
model1.add(Conv2D(filters = 64, kernel size = 3, padding = "same", activation = "relu"))
model1.add(MaxPooling2D(pool_size = 2))
model1.add(Dropout(0.3))
model1.add(Flatten())
model1.add(Dense(512, activation = "relu"))
model1.add(Dropout(0.4))
model1.add(Dense(128, activation="relu"))
model1.add(Dropout(0.3))
model1.add(Dense(2, activation = "softmax"))
```



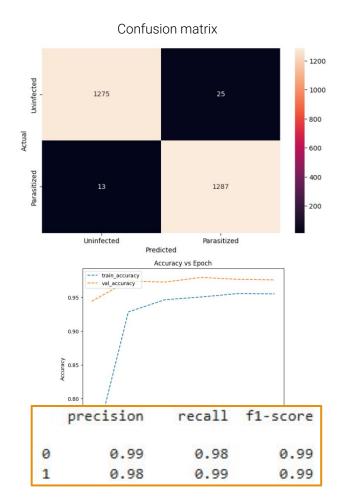
# **Observations: 97.5% test accuracy, lower class I recall**; high Dropout and BatchNorm seem to over regularize

```
model2 = Sequential()
model2.add(Conv2D(32, (3, 3), input shape = (64, 64, 3), padding = 'same'))
model2.add(BatchNormalization())
                                                                model2.add(Conv2D(128, kernel_size = 3, padding = "same"))
model2.add(LeakyReLU())
                                                                model2.add(BatchNormalization())
model2.add(MaxPooling2D(pool size=2))
                                                                model2.add(LeakyReLU())
model2.add(Dropout(0.2))
                                                                model2.add(MaxPooling2D(2))
                                                                model2.add(Dropout(0.4))
model2.add(Conv2D(64, kernel size = 3, padding = "same"))
                                                                model2.add(Flatten())
model2.add(BatchNormalization())
                                                                model2.add(Dense(128, activation = "relu"))
model2.add(LeakyReLU())
                                                                model1.add(BatchNormalization())
model2.add(MaxPooling2D(2))
                                                                model2.add(LeakyReLU())
model2.add(Dropout(0.3))
                                                                model2.add(Dropout(0.3))
model2.add(Conv2D(128, kernel_size = 3, padding = "same"))
                                                                model2.add(Dense(2, activation = "softmax"))
model2.add(BatchNormalization())
                                                                adam = optimizers.Adam(learning rate = 0.0005)
```



**Observations: 98.6% test accuracy, highest Class I recall.** 45% less parameters than base model. Efficient and elegant architecture

```
model3 = Sequential()
model3.add(Conv2D(32, 3, activation='relu', padding='same', input shape=(64, 64, 3)))
model3.add(MaxPooling2D(2))
model3.add(Conv2D(64, 3, activation='relu', padding='same'))
model3.add(MaxPooling2D(2))
model3.add(Conv2D(64, 3, activation='relu', padding='same'))
model3.add(MaxPooling2D(2))
model3.add(Flatten())
model3.add(Dense(128, activation='relu'))
model3.add(Dropout(0.3))
model3.add(Dense(2, activation = "softmax"))
```



Observations: 97.5% test accuracy, lowest recall; over rely on Image -Net; over regularized model too heavy for data size; 14x params

```
transfer layer = vgg.get layer('block5 pool')
vgg.trainable = False
x = Flatten()(transfer layer.output)
x = Dense(256, activation = 'relu')(x)
x = Dense(128, activation = 'relu')(x)
x = Dropout(0.3)(x)
x = Dense(128, activation = 'relu')(x)
x = BatchNormalization()(x)
pred = Dense(2, activation = 'softmax')(x)
model4 = Model(vgg.input, pred)
```

#### Confusion matrix 1000 1239 800 1207 Uninfected Parasitized Predicted Accuracy vs Epoch --- train accuracy val\_accuracy 0.94 0.93 recall f1-score precision 0.93 0.95 0.94

0.93

0.94

0.95