

Rscript04: BB-score analaysis and microbial top winners

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1 Load data

1.1 Load packages

```
rm(list=ls())
library(readxl)
library(ggplot2)
library(stringr)
library(dplyr)
library(ggrepel)
library(rstatix)
library(ggfortify)
library(cowplot)
library(tidyr)
library(scales)
library(ggbreak)
library(moments)
library(ggExtra)
library(purrr)
library(ggtext)
library(MASS)
library(tibble)
library(cooccur)
library(CooccurrenceAffinity)
library(pheatmap)
library(viridis)
library(ComplexHeatmap)
library(circlize)
library(grid)
library(stringr)
library(qgraph)
library(igraph)

cbp1 <- c("red", "#E69F00", "#0072B2", "#009E73", "#56B4E9", "#FOE442", "#100000")
```

1.2 Load rating-data

Loaded dataframe contains Elo-rating estimations (Category="Classic") BB-score estimations (Category="Corrected").

```
load(file = "../Scripts/Elo_MAPdata_from_Script02.RData")
str(df_MAP_Elo)
```

```
## 'data.frame': 1816294 obs. of 5 variables:
## $ player_id : chr "90_1;96_1;97_1;98_1;99_1" "90_1;96_1;97_1;98_1;99_1" "90_1;96_1;97_1;98_1;99_1"
## $ biome     : chr "animal.3" "plant.2" "freshwater.3" "freshwater.2" ...
## $ Category   : chr "Corrected" "Classic" "Corrected" "Classic" ...
## $ mean_rating: num 802 1000 794 1000 ...
## $ sd_rating  : num 8.089 0.028 6.7699 0.0505 0.0473 ...
```

1.3 Load taxonomy

```
# 1) Read the raw file (no header). Replace "myfile.tsv" with your actual filename.
df_raw <- read.delim("../data/otus.info",
                      header    = T,
                      sep       = "\t",
                      stringsAsFactors = FALSE,
                      quote     = "") # turn off any quoting behavior
str(df_raw)

## 'data.frame': 597954 obs. of 19 variables:
## $ OTU      : chr "90_17776;96_71281;97_92606;98_125911;99_193128" "90_17776;96_71281;97_92606;...
## $ Tax      : chr "Archaea" "Archaea" "Archaea" "Archaea" ...
## $ SpeciesRep : chr "" "" "" ...
## $ SeqCount : int 1 1 1 1 1 1 1 1 1 ...
## $ GoldCount : int 0 0 0 0 0 0 0 0 0 ...
## $ GenomeCount : int 0 0 0 0 0 0 0 0 0 ...
## $ TypeStrains : chr "" "" "" ...
## $ Strains   : chr "" "" "" ...
## $ Genomes   : chr "" "" "" ...
## $ GoldSeqs  : chr "" "" "" ...
## $ Aliases   : chr "" "" "" ...
## $ GoldHit   : chr "NR_074195:1..1470" "NR_074195:1..1470" "NR_074195:1..1470" "NR_074195:1..1470" ...
## $ GoldID    : int 725 725 725 725 725 569 569 569 569 ...
## $ GoldScore : num 0.754 0.754 0.754 0.754 0.754 ...
## $ RepSpecies : chr "" "" "" ...
## $ Taxaname  : chr "Archaea" "Archaea" "Archaea" "Archaea" ...
## $ OrigTax   : chr "Archaea" "Archaea" "Archaea" "Archaea" ...
## $ RepSequenceID: chr "KC471280:1..1464" "KC471280:1..1464" "KC471280:1..1464" "KC471280:1..1464" ...
## $ RepSequence : chr "TACCCGTTATCCTGCAGGAGGTGCTATCAGAATTGACTTAAGCTAGTTCTGGGGCTTCGGAAAGC"

#set taxonomy
taxonomy.table<- df_raw[,c("OTU", "Tax", "RepSequence")]

taxonomy.table <- df_raw %>%
  dplyr::select(OTU, Tax, RepSequenceID) %>%
  tidyr::separate(
    col    = Tax,
    into   = c("Domain", "Phylum", "Class", "Order", "Family", "Genus", "Species"),
    sep    = ";",
    fill   = "right")

str(taxonomy.table)

## 'data.frame': 597954 obs. of 9 variables:
## $ OTU      : chr "90_17776;96_71281;97_92606;98_125911;99_193128" "90_17776;96_71281;97_92606;...
## $ Domain   : chr "Archaea" "Archaea" "Archaea" "Archaea" ...
## $ Phylum   : chr NA NA NA NA ...
## $ Class    : chr NA NA NA NA ...
## $ Order    : chr NA NA NA NA ...
## $ Family   : chr NA NA NA NA ...
## $ Genus    : chr NA NA NA NA ...
```

```
## $ Species      : chr  NA NA NA NA ...
## $ RepSequenceID: chr  "KC471280:1..1464" "KC471280:1..1464" "KC471280:1..1464" ...
```

Cheching results

```
length(unique(df_MAP_Elo$player_id))
```

```
## [1] 124772
```

1.4 Load metadata

```
df.elo.diversity<-read_xlsx("../data/Supplementary_tables_ms.xlsx",
                           sheet="Table S2")
```

```
tibble(df.elo.diversity)
```

```
## # A tibble: 24 x 10
##   #> MAP_biomes Life.Style    MAP.biomesFullName n.samples.initial
##   #> <dbl> <chr>          <chr>                  <dbl>
## 1 1     airborne.1 Free-living    airborne            1595
## 2 2     animal.1   Host-associated animal-urogenital 29317
## 3 3     animal.2   Host-associated animal-proximalgut 180797
## 4 4     animal.3   Host-associated animal-distalgut 125143
## 5 5     animal.4   Host-associated animal-oral       37433
## 6 6     animal.5   Host-associated animal-skin      40488
## 7 7     animal.6   Host-associated animal-respiratory 10360
## 8 8     freshwater.1 Free-living    freshwater-sediments 19946
## 9 9     freshwater.2 Free-living    freshwater-water   46327
## 10 10    freshwater.3 Free-living    freshwater-biofilm 5067
## # i 14 more rows
## # i 5 more variables: n.samples.1000reads.3OTUS <dbl>, gamma.diversity <dbl>,
## #   Elo.coef <dbl>, Elo.coef.error <dbl>, Elo.pvalue <dbl>
```

```
df_MAP_Elo<-merge(df_MAP_Elo,df.elo.diversity[,c(2,3,4)],by.x="biome",by.y="MAP_biomes")
```

2 Microbial performance description

2.1 Classifying OTUs accoding performance

We operationally classified top performers as those OTUs which performance is greater than the biome species pool. We consider the top 5% as top performers for overall winners description and top 1% for a shorted list of winners.

```
df_MAP_Elo.corr <- df_MAP_Elo %>%
  filter(Category == "Corrected") %>%
  group_by(biome) %>%
  mutate(
    # --- normalize only by the mean (centered) ---
```

```

mean_rating_centered = mean_rating - median(mean_rating, na.rm = TRUE),

# --- empirical (5%) threshold + flag (on centered values) ---
thresh_95_emp_centered = stats::quantile(mean_rating_centered, 0.95),
exceed_95_emp_centered = mean_rating_centered > thresh_95_emp_centered,

# --- empirical (1%) threshold + flag (on centered values) ---
thresh_99_emp_centered = stats::quantile(mean_rating_centered, 0.99),
exceed_99_emp_centered = mean_rating_centered > thresh_99_emp_centered
) %>%
ungroup()

```

2.2 Species performance estimations based on BB-scores

For identification of successful taxa, we focused on OTUs in the top 5% of BB-scores in one or more biomes. For the quantification of the performance of these OTUs across the other biomes, we calculated average BB-scores for all biomes where they did not fall within the top 5% of BB-scores.

```

# Step 1: Filter only player_ids
df_Elo_corr<-df_MAP_Elo.corr #Just to keep the original
players_top <- df_Elo_corr%>%
  filter(exceed_95_emp_centered== TRUE) %>% # 5% top winners
  pull(player_id) %>%
  unique()

# Step 2: Filter df to include only those player_ids
filtered_df <- df_Elo_corr %>%
  filter(player_id %in% players_top)

# Step 4: Count and calculate percentages per player_id
df_top<- filtered_df %>%
  dplyr::select(player_id, biome, mean_rating_centered,exceed_95_emp_centered)

summary.tmp95<-df_top

#Step 5: Estimations
RP_summary_otus <- summary.tmp95 %>%
  group_by(player_id) %>%
  summarise(
    # In how many are greater than threshold
    count_BBgreater2 = sum(exceed_95_emp_centered==TRUE, na.rm = TRUE),
    # In how many RP <= threshold
    count_BBsmaller2 = sum(exceed_95_emp_centered==FALSE, na.rm = TRUE),
    # "d1_d9_count" = total biomes (24?) minus those greater than threshold
    biomes_rest = 24 - sum(exceed_95_emp_centered==TRUE, na.rm = TRUE),
    # sum of RP_centered non-top winners
    sum_BBsmaller2 = sum(mean_rating_centered[exceed_95_emp_centered==FALSE], na.rm = TRUE),
    sum_BBall = sum(mean_rating_centered, na.rm = TRUE),
    .groups = "drop")

# Estimating the interbiome-performance

```

```

RP_summary_otus$AverageRP<- (RP_summary_otus$sum_BBsmaller2)/(RP_summary_otus$count_BBsmaller2+1)

#Biome occurrence
RP_summary_otus$biome_occ<-RP_summary_otus$count_BBgreater2+RP_summary_otus$count_BBsmaller2

##Set occurrence categories
RP_summary_otus <- RP_summary_otus %>%
  mutate(
    biome_occ_class = case_when(
      biome_occ == 24           ~ "Ubiquitous Top5%", # exactly 24
      biome_occ < 24           ~ "Restricted",
      TRUE                     ~ NA_character_),biome_occ_class = factor(
        biome_occ_class,
        ordered = TRUE))

#Reorder biome levels co-occurrence
RP_summary_otus$biome_occ_class<-factor(RP_summary_otus$biome_occ_class,
                                           c("Ubiquitous Top5%","Restricted"))

#For the OTU that was top winner in 24, we manuall set it the max observed AverageRP
RP_summary_otus$AverageRP[RP_summary_otus$count_BBgreater2==24]<-max(RP_summary_otus$AverageRP)

# Estimate Spearman correlation between top5% occurrence and interbiome performance

res.correlation<-cor.test(~count_BBgreater2+AverageRP,data=RP_summary_otus[RP_summary_otus$biome_occ_cl]

res.correlation

## 
## Spearman's rank correlation rho
##
## data: count_BBgreater2 and AverageRP
## S = 2753960, p-value < 2.2e-16
## alternative hypothesis: true rho is not equal to 0
## sample estimates:
##          rho
## 0.6112806

### Find the top 1% performers ###

# Step 1: Filter only player_ids
df_Elo_corr<-df_MAP_Elo.corr #Just to keep the original
players_top <- df_Elo_corr%>%
  filter(exceed_99_emp_centered== TRUE) %>% # 1% top winners
  pull(player_id) %>%
  unique()

# Step 2: Filter df to include only those player_ids
filtered_df <- df_Elo_corr %>%
  filter(player_id %in% players_top)

# Step 4: Count and calculate percentages per player_id
df_top<- filtered_df %>%

```

```

dplyr::select(player_id, biome, mean_rating_centered, exceed_99_emp_centered)

# Save results
summary.tmp99<-df_top

#Step 5: Estimations
RP_summary_otus_99 <- summary.tmp99 %>%
  group_by(player_id) %>%
  summarise(
    # In how many are greater than threshold
    count_BBgreater2 = sum(exceed_99_emp_centered==TRUE, na.rm = TRUE),
    # In how many RP <= threshold
    count_BBsmaller2 = sum(exceed_99_emp_centered==FALSE, na.rm = TRUE),
    # "d1_d9_count" = total biomes (24?) minus those greater than threshold
    biomes_rest = 24 - sum(exceed_99_emp_centered==TRUE, na.rm = TRUE),
    # sum of RP_centered non-top winners
    sum_BBsmaller2 = sum(mean_rating_centered[exceed_99_emp_centered==FALSE], na.rm = TRUE),
    .groups = "drop")
#change names
names(RP_summary_otus_99)<-c("player_id","count_BBgreater2.99","count_BBsmaller2.99","biomes_rest.99", "sum_BBsmaller2.99")

## Combine both top5% and top1% estimations
RP_summary_otus <- merge(RP_summary_otus,RP_summary_otus_99,by="player_id",all.x=T)
tibble(RP_summary_otus)

## # A tibble: 20,942 x 13
##   player_id      count_BBgreater2 count_BBsmaller2 biomes_rest sum_BBsmaller2
##   <chr>          <int>           <int>        <dbl>       <dbl>
## 1 90_1;96_1;97_1;~         1            13          23        0.558
## 2 90_1;96_1;97_35~        3            19          21        3.22
## 3 90_1;96_1115;97~        3            19          21        1.25
## 4 90_1;96_11286;9~        1            12          23        0.310
## 5 90_1;96_11646;9~        2            16          22        2.01
## 6 90_1;96_11925;9~        1            14          23        0.971
## 7 90_1;96_1238;97~        1            19          23        3.33
## 8 90_1;96_1238;97~        1            17          23        1.25
## 9 90_1;96_13029;9~        2            10          22        0.598
## 10 90_1;96_13030;9~       1            20          23        2.32
## # i 20,932 more rows
## # i 8 more variables: sum_BBall <dbl>, AverageRP <dbl>, biome_occ <int>,
## #   biome_occ_class <ord>, count_BBgreater2.99 <int>,
## #   count_BBsmaller2.99 <int>, biomes_rest.99 <dbl>, sum_BBsmaller2.99 <dbl>
```

2.2.1 Biome occurrence of top 5% performers (Figure 4B)

```

# Top panel (Occupancy distribution)
df.agg.occ.biomes.V2<- aggregate(player_id~biome_occ ,data=RP_summary_otus, length)

df.agg.occ.biomes.V2<-df.agg.occ.biomes.V2%>%
  mutate(
    biome_occ_class = case_when(
```

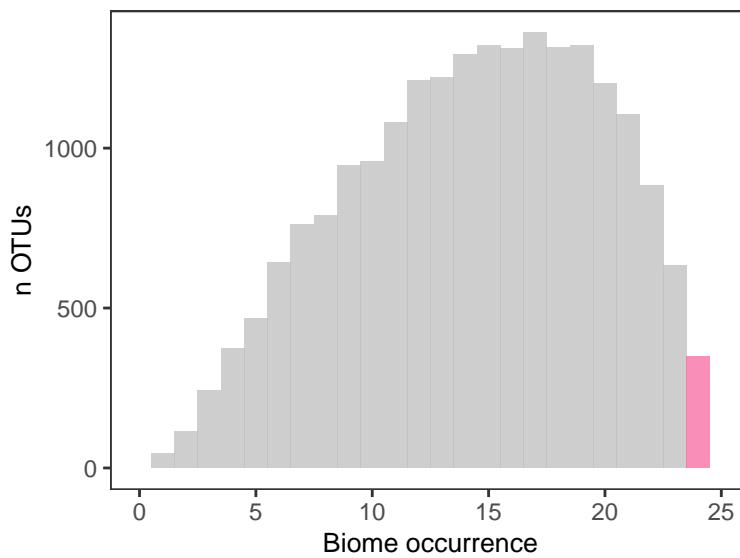
```

biome.occ == 24           ~ "Ubiquitous Top5%", # exactly 24
biome.occ < 24            ~ "Restricted",
TRUE                      ~ NA_character_),biome_occ_class = factor(
biome_occ_class,
ordered = TRUE))

df.agg.occ.biomes.V2$biome_occ_class<-factor(df.agg.occ.biomes.V2$biome_occ_class,
                                               c("Ubiquitous Top5%","Restricted"))

# Plot
bar.plot.top.panel<-ggplot(df.agg.occ.biomes.V2, aes(x=biome.occ,y=player_id,fill=biome_occ_class)) +
  geom_bar(stat="identity",width = 1,alpha=0.75) +
  scale_fill_manual(values=c("#f768a1","grey"),name=NULL) +
  theme_bw() +theme(legend.position = "none",axis.title = element_text(size=10),
                    panel.grid.major = element_blank(),panel.grid.minor = element_blank())+labs(y="n OTU"
bar.plot.top.panel

```



2.2.2 Distribution top 5% performers based on BB-scores (Figure 4B)

```

# Bottom panel (Distribution top winners)
df.agg.occ.biomes.V3<- aggregate(player_id~count_BBgreater2+biome_occ_class ,data=RP_summary_otus, length.out=1)

df.agg.occ.biomes.V3$biome_occ_class<-factor(df.agg.occ.biomes.V3$biome_occ_class,
                                               c("Restricted","Ubiquitous Top5%"))

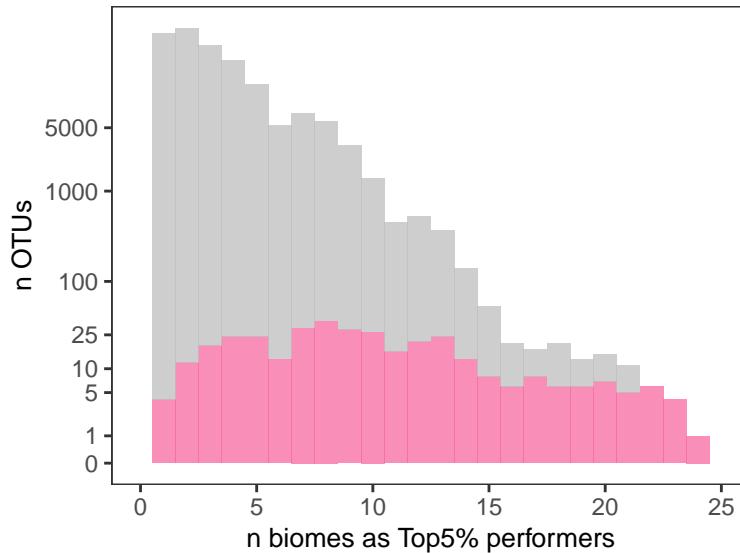
bar.plot.bottom.panel <- ggplot(df.agg.occ.biomes.V3, aes(x = count_BBgreater2, y = player_id+1,fill=biome_occ_class)) +
  geom_col(width = 1, alpha = 0.75) +
  scale_y_log10(
    breaks = c(0, 1, 5,10, 25, 100,1000, 5000) + 1, # match shift
    labels = function(b) b - 1) +
  xlim(0, 25) +
  theme_bw() +

```

```

theme(legend.position = "none",axis.title = element_text(size=10),
      panel.grid.major = element_blank(),
      panel.grid.minor = element_blank()) +
  labs(y = expression("n OTUs"), x = "n biomes as Top5% performers")+
  scale_fill_manual(values=c("grey","#f768a1"),name=NULL)
bar.plot.bottom.panel

```



2.3 Merge dataset with taxonomy

```

#set taxonomy
RP_summary_otus_tax<-merge(RP_summary_otus, taxonomy.table,by.x ="player_id",by.y="OTU",all.x=T)

RP_summary_otus_tax <- RP_summary_otus_tax %>%
  mutate(
    # turn empty strings into NA for consistency
    Family = na_if(Family, ""),
    Order = na_if(Order, ""),
    Class = na_if(Class, ""),
    # build a fallback label for each row:
    Genus = case_when(
      !is.na(Genus) & Genus != "" ~ Genus,
      !is.na(Family) ~ paste0("unc_", Family),
      !is.na(Order) ~ paste0("unc_", Order),
      !is.na(Class) ~ paste0("unc_", Class),
      TRUE ~ NA_character_
    )
  )

```

2.3.1 Principal panel top5% winners versus average BB-score (Figure 4A)

Inter-biome performance (average normalized BB-score across all biomes) of OTUs that were in the top 5% of performers in 1 or more biomes. The subset of ubiquitous microbes detected in all 24 biomes are highlighted in pink. Six ubiquitous microbes were in the top 1% of performers in more than half of the biomes and their average BB-score was >99.9% of all OTUs (in green).

```
# add flag columns for conditions to highlight (Ubiquitous and high performers)
RP_summary_otus_tax2 <- RP_summary_otus_tax %>%
  mutate(
    ring = RP_summary_otus$count_BBgreater2.99 >= 12 & count_BBgreater2.99>=12 & AverageRP> quantile(RP,
    stroke_size = ifelse(ring, 1.1, 0.2)    # adjust as you like
  )

pos <- position_dodge2(width = 0.5, preserve = "single")

RP_summary_otus_tax2$flag.colour<-as.character(RP_summary_otus_tax2$biome_occ_class)
RP_summary_otus_tax2$flag.colour[RP_summary_otus_tax2$ring==TRUE]<-"Ubiquitous Top1%"
RP_summary_otus_tax2$flag.colour<-as.factor(RP_summary_otus_tax2$flag.colour)

##Check correlation between the 2 parameters (Spearman correlation)
res.correlation<-cor.test(~count_BBgreater2+AverageRP,RP_summary_otus_tax2[RP_summary_otus_tax2$biome_o
res.correlation

##
## Spearman's rank correlation rho
##
## data: count_BBgreater2 and AverageRP
## S = 2753960, p-value < 2.2e-16
## alternative hypothesis: true rho is not equal to 0
## sample estimates:
##      rho
## 0.6112806

principal.mappinng.OTUs.BB <- ggplot(
  RP_summary_otus_tax2,
  aes(x = factor(count_BBgreater2), y = AverageRP, fill = flag.colour)
) +
  # Restricted: plot but don't show in legend
  geom_point(
    data = dplyr::filter(RP_summary_otus_tax2, flag.colour == "Restricted"),
    shape = 21, size = 2.2, alpha = 0.5, color = "black",
    position = pos, show.legend = FALSE
  ) +
  # Ubiquitous
  geom_point(
    data = dplyr::filter(RP_summary_otus_tax2, flag.colour == "Ubiquitous Top5%"),
    shape = 21, size = 2.4, alpha = 0.75, color = "black", position = pos
  ) +
  # Ubiquitous 1%
  geom_point(
```

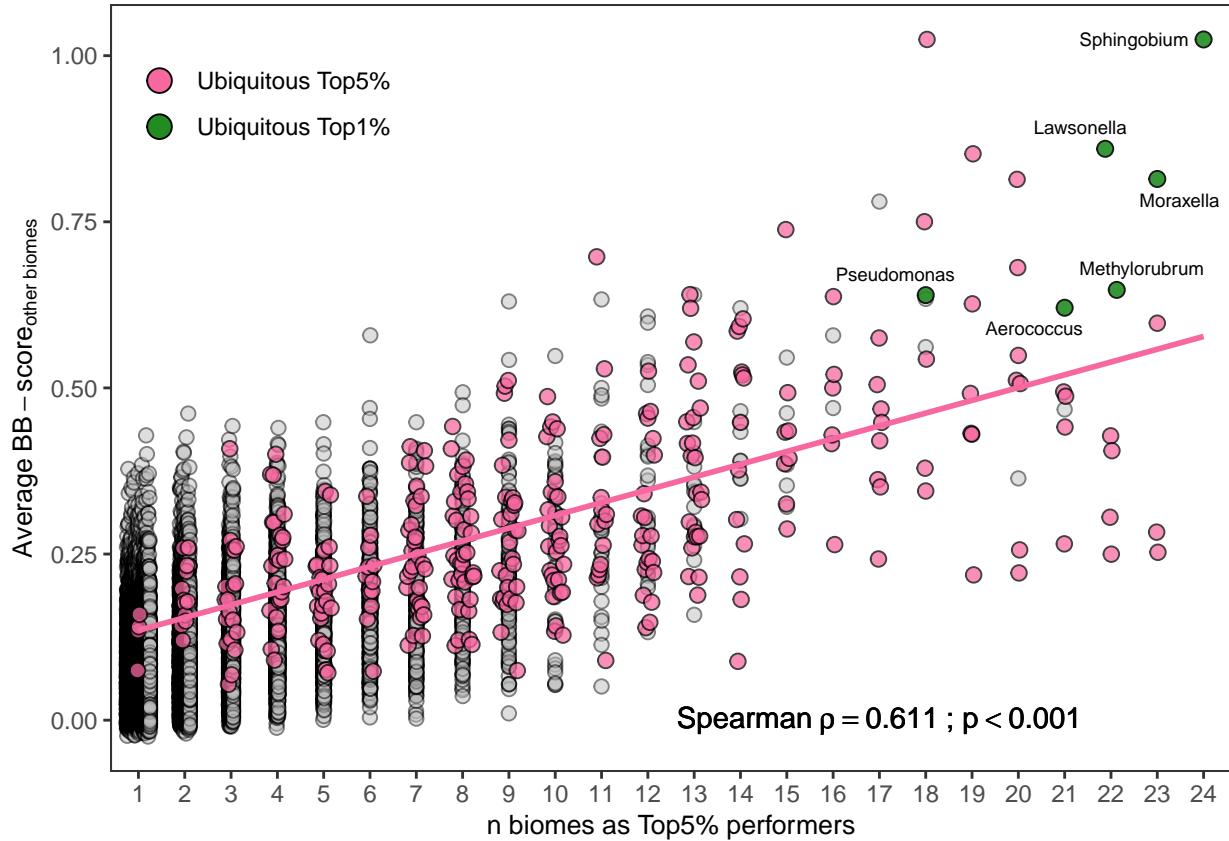
```

    data = dplyr::filter(RP_summary_otus_tax2, flag.colour == "Ubiquitous Top1%"),
    shape = 21, size = 2.5, alpha = 0.9, color = "black", position = pos
) +
scale_fill_manual(
  name = NULL,
  values = c("Restricted" = "grey",
             "Ubiquitous Top5%" = "#f768a1",
             "Ubiquitous Top1%" = "#228B22"),
  breaks = c("Ubiquitous Top5%", "Ubiquitous Top1%") # <- removes "Restricted" from legend
) +
theme_bw() +
scale_x_discrete(breaks = as.character(1:24)) +
labs(x = "n biomes as Top5% performers", y = expression(Average~BB-score["other biomes"])) +
ggrepel::geom_text_repel(
  data = dplyr::filter(RP_summary_otus_tax2, ring),
  aes(label = Genus), size = 2.5, max.overlaps = Inf,
  box.padding = 0.3, point.padding = 0.2, segment.size = 0.2
) +
theme(panel.grid.major = element_blank(), panel.grid.minor = element_blank(), axis.title = element_text(),
      legend.position = c(0.01, 0.95), legend.justification = c(0, 1)) +
geom_smooth(
  data = subset(RP_summary_otus_tax2, biome_occ_class == "Ubiquitous Top5%"),
  aes(count_BBgreater2, AverageRP, colour=biome_occ_class, fill=biome_occ_class),
  se = FALSE, method = "lm", colour = "#f768a1", show.legend = FALSE
) +
guides(fill = guide_legend(override.aes = list(size = 4, alpha = 1))) +
annotate("text", x=17, y=0.0,
        label=bquote(Spearman~rho==.(round(res.correlation$estimate,3))~"; p"~<~0.001))

## Warning: A numeric 'legend.position' argument in 'theme()' was deprecated in ggplot2
## 3.5.0.
## i Please use the 'legend.position.inside' argument of 'theme()' instead.
## This warning is displayed once every 8 hours.
## Call 'lifecycle::last_lifecycle_warnings()' to see where this warning was
## generated.

principal.mappinng.OTUs.BB

```



2.4 Get names of the widespread and thos with high interbiome performance

```
names.taxa.top <- filter(RP_summary_otus_tax2, count_BBgreater2 >= 12 & AverageRP > quantile(RP_summary_otus_tax2, 0.75))
tibble(names.taxa.top[, 1:5])
```

```
## # A tibble: 6 x 5
##   player_id      count_BBgreater2 count_BBsmaller2 biomes_rest sum_BBsmaller2
##   <chr>           <int>            <int>        <dbl>          <dbl>
## 1 90_10;96_1541;97~       22              2          2             2.58
## 2 90_13;96_938;97~       21              3          3             2.48
## 3 90_15368;96_177;~      22              2          2             1.94
## 4 90_20;96_774;97~       24              0          0             0
## 5 90_23;96_61;97_8~      18              6          6             4.48
## 6 90_80;96_1834;97~      23              1          1             1.63
```

2.4.1 Distribution of the observed top winners across biomes (Figure 4D)

Heatmap of biome-level occupancy (% of samples per biome) of the six top-performing microbes. The numbers in parentheses indicate the number of biomes in which each lineage was ranked within the top 1% of performers. The color bar indicates relative occupancy.

```

winners.info<-data.frame(read.csv("../data/selected_OTUs_across_biomes.csv"))
#Intermediate step
#names.taxa.top<-filter(RP_summary_otus_tax2, count_BBgreater2>=12& AverageRP>quantile(RP_summary_otus_)

## Add names to the winners
names.taxa.top$short.name<-paste0(names.taxa.top$Genus, ".", str_split_fixed(names.taxa.top$RepSequenceID, "#names$short.name<-str_replace_all(names$short.name, "[ : ]", " . ")
```

```

names.taxa.top<-merge(names.taxa.top[,c(1,25)],winners.info,by.x="player_id",by.y="otu",all.x=T)
names.taxa.top<-merge(names.taxa.top,df.elo.diversity[,c(2,4)],by.x="biome",by.y="MAP_biomes")
```

```

# Sort biomes
levels_sorted<-c("airborne","animal-urogenital","animal-proximalgut","animal-distalgut","animal-oral",
                 "plant-rhizosphere","plant-phyllosphere","plant-endosphere","plant-spermophere",
                 "freshwater-sediments","freshwater-water","freshwater-biofilm",
                 "freshwater-peatlands(peat/bog)","freshwater-peatlands(water)",
                 "saline-sediments","saline-water","saline-biofilm",
                 "soil-agricultural","soil-desert","soil-tundra","soil-forest","soil-grassland")
names.taxa.top$MAP_biomesFullName<-as.factor(names.taxa.top$MAP_biomesFullName)
names.taxa.top$MAP_biomesFullName<-factor(names.taxa.top$MAP_biomesFullName,levels_sorted)
```

```

occ_mat <- names.taxa.top %>%
  dplyr::select(short.name = short.name, MAP_biomesFullName, occurrence_rate) %>%
  group_by(short.name, MAP_biomesFullName) %>%
  summarise(value = mean(occurrence_rate, na.rm = TRUE), .groups = "drop") %>%
  pivot_wider(names_from = MAP_biomesFullName, values_from = value, values_fill = 0) %>%
  column_to_rownames("short.name") %>%
  as.matrix()
```

```

row.names(occ_mat)<-c("Aerococcus viridans (12)", "Lawsonella clevelandensis (13)",
                      "Methylobacter [Methylobacterium] populi (15)",
                      "Faucicola [Moraxella] osloensis (21)", "Pseudomonas fluorescens complex (12)",
                      "Sphingobium yanoikuyaе (20)" )
```

```

occ_mat.sorted<-occ_mat[c(4,6,3,2,1,5),]
```

```

# Define breaks from 1 to 100
my_breaks <- seq(0, 57, length.out = 40)
```

```

mat_perc <- occ_mat.sorted * 100
```

```

# --- categories & colors ---
cat_levels <- c("airborne", "animal", "plant", "freshwater", "saline", "soil")
group_cols <- c(
  airborne = "black",
  animal = "#e67e22", # orange
  plant = "#2ca02c", # green
  freshwater = "#74a9cf", # light blue
  saline = "#045a8d", # dark blue
  soil = "#8c510a" # brown
)
# group assignment
```

```

biome_full <- colnames(mat_perc)
biome_group <- sub("-.*$", "", biome_full)
biome_group <- dplyr::recode(biome_group, plants = "plant") # unify naming
cat_fac     <- factor(biome_group, levels = cat_levels)

# shorten labels (after "-")
biome_short <- ifelse(grepl("-", biome_full),
                      sub("^[^-]+-", "", biome_full),
                      biome_full)

# block labels: capitalize, omit airborne
block_labels <- ifelse(cat_levels == "airborne", "", str_to_title(cat_levels))

# bottom annotation with group-colored boxes and white labels
ha_bottom <- HeatmapAnnotation(
  Category = anno_block(
    gp        = gpar(fill = group_cols[cat_levels], col = NA),
    labels   = block_labels,
    labels_gp = gpar(col = "white", fontsize = 10),
    which    = "column"
  )
)

col_fun <- circlize::colorRamp2(
  c(0, 0.001, 10, 20, 40, 60),
  c("grey85", "white", "#FFF7BC", "gold", "orange", "darkred"))

```

```

ht <- Heatmap(
  mat_perc,
  name = "%Occupancy",
  col  = col_fun,
  cluster_rows  = FALSE,
  cluster_columns = FALSE,
  column_split   = cat_fac,
  column_labels   = biome_short,
  column_names_side = "bottom",
  column_names_rot = 45,
  column_names_gp  = gpar(col = group_cols[as.character(cat_fac)], fontsize = 9),
  row_names_side   = "left",
  row_names_gp     = gpar(fontsize = 8, fontface = "italic"),
  border  = TRUE,
  rect_gp = gpar(col = "grey40", lwd = 0.75),
  bottom_annotation = ha_bottom,
  column_title = NULL,
  show_heatmap_legend = FALSE,           # <-- important
  heatmap_legend_param = list(direction="horizontal", fontsize=9)
)

# 2) Your custom legend
lgd <- Legend(title = "%Occupancy", col_fun = col_fun, direction = "horizontal")

# 3) Capture EVERYTHING in a single grob; don't draw beforehand
combined_grob <- grid.grabExpr({

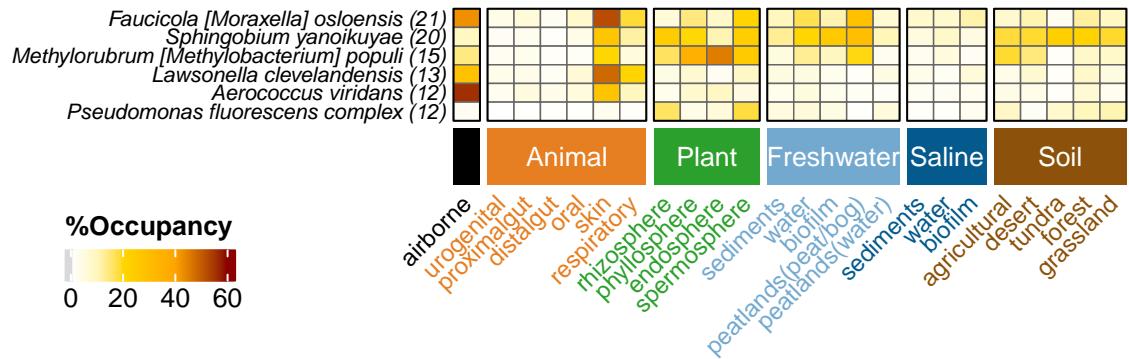
```

```

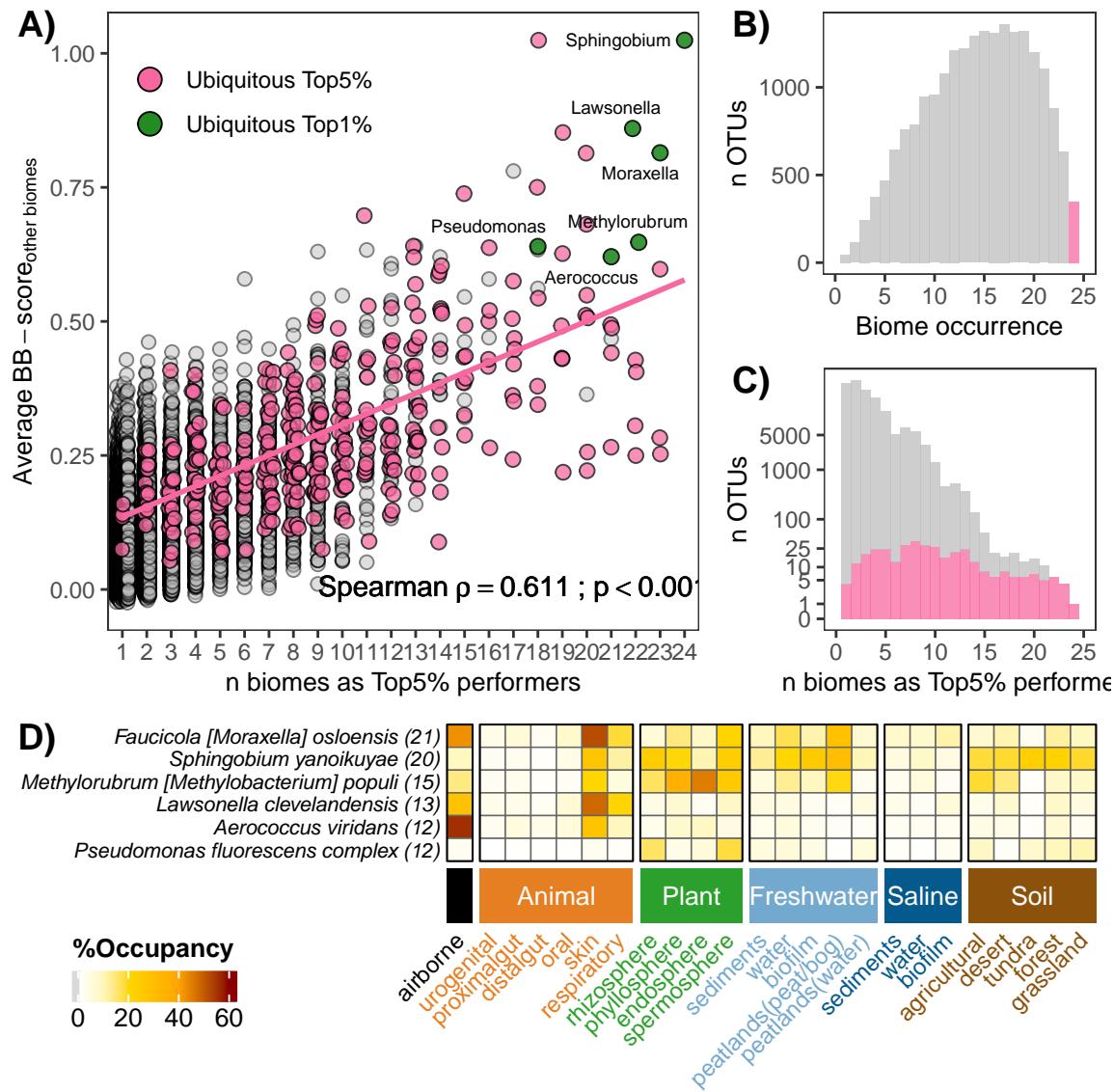
draw(ht, newpage = FALSE)                      # draw once, inside grabExpr
pushViewport(viewport(x = unit(-55, "mm"),    # adjust as needed
                     y = unit(-10, "mm"),
                     just = c("left", "bottom")))
draw(lgd)
upViewport()
})

ggdraw(combined_grob)

```



2.4.2 Figure 4



2.4.3 Screen for the pathogens and SKAPE bacteria (example, Table S7)

```
#Identification of the WHO bacterial priority pathogens list 2024
#1) Acinetobacter baumannii
# 90_8;96_11;97_11;98_13;99_14

#2) Pseudomonas aeruginosa
#"90_23;96_36;97_37;98_40;99_42"

#3) Neisseria gonorrhoeae subclade
# 90_15355;96_42;97_44;98_48;99_52

#4) Streptococcus pneumoniae
```

```

#90_4;96_4;97_4;98_4;99_4

#5) Haemophilus influenzae
# 90_18;96_30;97_31;98_34;99_36

#6) Shigella spp (KF080510)
# 90_11728;96_51054;97_65764;98_87812;99_131241

#7) Enterococcus faecium
# 90_13;96_19;97_19;98_22;99_23

#8) Staphylococcus aureus
# 90_3;96_3;97_3;98_3;99_3

#9) Klebsiella pneumoniae
# 90_6;96_7;97_7;98_9;99_10

#Calculate percentiles per biome

biomes <- df_MAP_Elo.corr %>% distinct(biome)
stats <- df_MAP_Elo.corr %>%
  group_by(biome) %>%
  mutate(biome_percentile = percent_rank(mean_rating) * 100) %>%
  ungroup()

#Query
id <- "90_6;96_7;97_7;98_9;99_10" #Klebsiella pneumoniae

#Calculate players stats
player <- stats %>%
  filter(player_id == id) %>%
  group_by(biome) %>%
  summarise(player_mean_rating = round(mean(mean_rating), 2),
            biome_percentile = round(mean(biome_percentile), 2),
            .groups = "drop")
#summary
out <- biomes %>% left_join(player, by = "biome") %>% arrange(desc(biome_percentile))
t(out)

## [,1]      [,2]      [,3]      [,4]      [,5]
## biome      "animal.3" "animal.2" "plant.2" "plant.3" "animal.6"
## player_mean_rating "817.27" "758.08" "707.19" "694.09" "630.64"
## biome_percentile   "99.81"   "99.38"   "99.19"   "97.92"   "97.62"
## [,6]      [,7]      [,8]      [,9]
## biome      "plant.4" "freshwater.3" "animal.5" "animal.1"
## player_mean_rating "719.24" "795.89"   "775.31"  "384.49"
## biome_percentile   "97.45"   "97.36"    "96.97"   "96.84"
## [,10]     [,11]     [,12]     [,13]     [,14]
## biome      "freshwater.2" "plant.1"  "freshwater.1" "animal.4" "soil.1"
## player_mean_rating "812.43"   "903.40"  "870.04"   "793.29"  "902.94"
## biome_percentile   "96.70"   "96.63"   "95.59"   "94.47"   "93.79"
## [,15]     [,16]     [,17]     [,18]     [,19]     [,20]

```

```

## biome          "soil.5" "soil.4" "saline.1" "saline.2" "saline.3" "soil.2"
## player_mean_rating "924.21" "832.25" "868.75"   "846.52"   "843.90"   "862.35"
## biome_percentile  "90.02"   "88.93"   "88.26"    "82.98"    "79.52"    "61.99"
##                [,21]      [,22]      [,23]      [,24]
## biome          "peatland.1" "airborne.1" "peatland.2" "soil.3"
## player_mean_rating "849.70"   "865.49"   "870.57"    NA
## biome_percentile  "51.77"    "51.59"    " 9.32"    NA

```

3 Network analysis

The biome connectivity network was constructed from the presence/absence of the top 5% BB-scores OTUs within each biome using the Jaccard dissimilarity. To assess whether similarity was higher than by chance, we used the maximum likelihood estimation from the CooccurrenceAffinity R-package.

3.1 Prepare matrix presence-absences

3.2 Construct Netowrk based on Jaccard dissimilarity.

The biome connectivity network used the presence/absence of the top 5% BB-scores OTUs within each biome using the Jaccard dissimilarity. To determine whether similarity was higher than by chance, we used the maximum likelihood estimation (alpha) from the CooccurrenceAffinity R-package.

```

# 2) Compute pairwise Jaccard distances
res <- affinity(data = presence_absence, row.or.col = "row",datatype = "binary",squarematrix = c("jaccard"))

res.affinity<-res$all
res.affinity$p_value<-as.numeric(res.affinity$p_value)
res.affinity$padjust<-p.adjust(res.affinity$p_value,method="bonferroni")

#Save data for network visualization in Gephi
write.csv(file="..../data/network.table.top5perc.csv",res.affinity,row.names = FALSE)

```

3.3 Network properties

Network analysis and small-worldness index (SWI) were computed, and its significance was assessed by a Monte Carlo randomization test (Manly,2018. Randomization, bootstrap and Monte Carlo methods in biology) against 1000 degree-preserving null networks.

```

# Filter out a>0 and p-adjust <0.05 as in Gephi network
edges <- res.affinity %>%
  filter(alpha_mle > 0, padjust < 0.05) %>%
  dplyr::select(entity_1, entity_2, jaccard)

#Prepare adjance matrix from dataframe
nodes <- sort(unique(c(edges$entity_1, edges$entity_2)))
A <- matrix(0, length(nodes), length(nodes), dimnames = list(nodes, nodes))

for(i in 1:nrow(edges)){
  a <- edges$entity_1[i]; b <- edges$entity_2[i]
  A[a, b] <- A[b, a] <- 1
}

```

```

}

# Network parameters
set.seed(1)
smallworldness(A, B = 1000)

##      smallworldness      trans_target average_length_target
##      1.1847561          0.7179648          1.5688406
##      trans_rnd_M         trans_rnd_lo       trans_rnd_up
##      0.5897619          0.5615578          0.6218593
##  average_length_rnd_M average_length_rnd_lo average_length_rnd_up
##      1.5267971          1.5108696          1.5471014

```

3.4 Smallworldness index (SWI) by Monte Carlo randomization test

```

#1) Get observed index
set.seed(1)
sw_obs <- smallworldness(A, B = 0)
sw_obs[1]

## smallworldness
##      1.193143

#2) Extract network from adjancent matrix
g_obs <- graph_from_adjacency_matrix(A, mode = "undirected", diag = FALSE)

#3) Estimate null distribution
set.seed(1)
B <- 1000
sw_null <- numeric(B)

for (b in seq_len(B)) {
  g_rand <- igraph::sample_degseq(igraph::degree(g_obs), method = "vl")    # preserves degree sequence
  tmp_sw<- smallworldness(g_rand, B = 0)
  sw_null[b] <-tmp_sw[["smallworldness"]]\# Save the result from each iteration
}

#4) Estimate P-value by comparing the observed SWI vs the SWI-NULL distribution data
B <- length(sw_null)
p_one_sided <- (sum(sw_null >= sw_obs[1]) + 1) / (B + 1)

print (paste("Pvalue Monte Carlo test =",round(p_one_sided,3)))

## [1] "Pvalue Monte Carlo test = 0.001"

```