

# Introduction to R

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## Overview

### What is R?

R is a powerful statistical environment and programming language for the analysis and visualization of data. The associated Bioconductor and CRAN package repositories provide many additional R packages for statistical data analysis for a wide array of research areas. The R software is free and runs on all common operating systems.

### Why Using R?

- Complete statistical environment and programming language
- Efficient functions and data structures for data analysis
- Powerful graphics
- Access to fast growing number of analysis packages
- Most widely used language in bioinformatics
- Is standard for data mining and biostatistical analysis
- Technical advantages: free, open-source, available for all OSs

### Books and Documentation

- simpleR - Using R for Introductory Statistics (John Verzani, 2004) - [URL](#)
- Bioinformatics and Computational Biology Solutions Using R and Bioconductor (Gentleman et al., 2005) - [URL](#)
- More on this see “Finding Help” section in UCR Manual - [URL](#)

### R Working Environments

#### R Projects and Interfaces

Some R working environments with support for syntax highlighting and utilities to send code to the R console:

- RStudio: excellent choice for beginners ([Cheat Sheet](#))
- Basic R code editors provided by Rguis
- gedit, Rgedit, RKWard, Eclipse, Tinn-R, Notepad++, NppToR
- Vim-R-Tmux: R working environment based on vim and tmux
- Emacs (ESS add-on package)

#### Example: RStudio

New integrated development environment (IDE) for R. Highly functional for both beginners and advanced.

RStudio IDE

Some useful shortcuts: **Ctrl+Enter** (send code), **Ctrl+Shift+C** (comment/uncomment), **Ctrl+1/2** (switch window focus)

### Example: Nvim-R-Tmux

Terminal-based Working Environment for R: Nvim-R-Tmux.

Nvim-R-Tmux IDE for R

## R Package Repositories

- CRAN (>11,000 packages) general data analysis - [URL](#)
- Bioconductor (>1,100 packages) bioscience data analysis - [URL](#)
- Omegahat (>90 packages) programming interfaces - [URL](#)

## Installation of R Packages

1. Install R for your operating system from CRAN.
2. Install RStudio from RStudio.
3. Install CRAN Packages from R console like this:

```
install.packages(c("pkg1", "pkg2"))  
install.packages("pkg.zip", repos=NULL)
```

4. Install Bioconductor packages as follows:

```
source("http://www.bioconductor.org/biocLite.R")  
library(BiocInstaller)  
BiocVersion()  
biocLite()  
biocLite(c("pkg1", "pkg2"))
```

5. For more details consult the Bioc Install page and BiocInstaller package.

## Getting Around

### Startup and Closing Behavior

- **Starting R:** The R GUI versions, including RStudio, under Windows and Mac OS X can be opened by double-clicking their icons. Alternatively, one can start it by typing R in a terminal (default under Linux).
- **Startup/Closing Behavior:** The R environment is controlled by hidden files in the startup directory: `.RData`, `.Rhistory` and `.Rprofile` (optional).
- **Closing R:**

```
q()
```

Save workspace image? [y/n/c]:

- **Note:** When responding with `y`, then the entire R workspace will be written to the `.RData` file which can become very large. Often it is sufficient to just save an analysis protocol in an R source file. This way one can quickly regenerate all data sets and objects.

## Navigating directories

Create an object with the assignment operator `<-` or `=`

```
object <- ...
```

List objects in current R session

```
ls()
```

Return content of current working directory

```
dir()
```

Return path of current working directory

```
getwd()
```

Change current working directory

```
setwd("/home/user")
```

## Basic Syntax

General R command syntax

```
object <- function_name(arguments)
object <- object[arguments]
```

Finding help

```
?function_name
```

Load a library/package

```
library("my_library")
```

List functions defined by a library

```
library(help="my_library")
```

Load library manual (PDF or HTML file)

```
vignette("my_library")
```

Execute an R script from within R

```
source("my_script.R")
```

Execute an R script from command-line (the first of the three options is preferred)

```
$ Rscript my_script.R
$ R CMD BATCH my_script.R
$ R --slave < my_script.R
```

# Data Types

## Numeric data

Example: 1, 2, 3, ...

```
x <- c(1, 2, 3)
x
```

```
## [1] 1 2 3
```

```
is.numeric(x)
```

```
## [1] TRUE
```

```
as.character(x)
```

```
## [1] "1" "2" "3"
```

## Character data

Example: "a", "b", "c", ...

```
x <- c("1", "2", "3")
x
```

```
## [1] "1" "2" "3"
```

```
is.character(x)
```

```
## [1] TRUE
```

```
as.numeric(x)
```

```
## [1] 1 2 3
```

## Complex data

Example: mix of both

```
c(1, "b", 3)
```

```
## [1] "1" "b" "3"
```

## Logical data

Example: TRUE of FALSE

```
x <- 1:10 < 5
x
```

```
## [1] TRUE TRUE TRUE TRUE FALSE FALSE FALSE FALSE FALSE
```

```
!x
```

```
## [1] FALSE FALSE FALSE FALSE TRUE TRUE TRUE TRUE TRUE
```

```
which(x) # Returns index for the 'TRUE' values in logical vector
```

```
## [1] 1 2 3 4
```

# Data Objects

## Object types

### Vectors (1D)

Definition: numeric or character

```
myVec <- 1:10; names(myVec) <- letters[1:10]
myVec[1:5]
```

```
## a b c d e
## 1 2 3 4 5
```

```
myVec[c(2,4,6,8)]
```

```
## b d f h
## 2 4 6 8
```

```
myVec[c("b", "d", "f")]
```

```
## b d f
## 2 4 6
```

### Factors (1D)

Definition: vectors with grouping information

```
factor(c("dog", "cat", "mouse", "dog", "dog", "cat"))
```

```
## [1] dog   cat   mouse dog   dog   cat
## Levels: cat dog mouse
```

### Matrices (2D)

Definition: two dimensional structures with data of same type

```
myMA <- matrix(1:30, 3, 10, byrow = TRUE)
class(myMA)
```

```
## [1] "matrix"
```

```
myMA[1:2,]
```

```
##      [,1] [,2] [,3] [,4] [,5] [,6] [,7] [,8] [,9] [,10]
## [1,]    1    2    3    4    5    6    7    8    9    10
## [2,]   11   12   13   14   15   16   17   18   19   20
```

```
myMA[1, , drop=FALSE]
```

```
##      [,1] [,2] [,3] [,4] [,5] [,6] [,7] [,8] [,9] [,10]
## [1,]    1    2    3    4    5    6    7    8    9    10
```

### Data Frames (2D)

Definition: two dimensional objects with data of variable types

```
myDF <- data.frame(Col1=1:10, Col2=10:1)
myDF[1:2, ]
```

```
##   Col1 Col2
## 1    1   10
## 2    2    9
```

## Arrays

Definition: data structure with one, two or more dimensions

## Lists

Definition: containers for any object type

```
myL <- list(name="Fred", wife="Mary", no.children=3, child.ages=c(4,7,9))
myL
```

```
## $name
## [1] "Fred"
##
## $wife
## [1] "Mary"
##
## $no.children
## [1] 3
##
## $child.ages
## [1] 4 7 9
```

```
myL[[4]][1:2]
```

```
## [1] 4 7
```

## Functions

Definition: piece of code

```
myfct <- function(arg1, arg2, ...) {
  function_body
}
```

## Subsetting of data objects

(1.) Subsetting by positive or negative index/position numbers

```
myVec <- 1:26; names(myVec) <- LETTERS
myVec[1:4]
```

```
## A B C D
## 1 2 3 4
```

## (2.) Subsetting by same length logical vectors

```
myLog <- myVec > 10
myVec[myLog]
```

```
## K L M N O P Q R S T U V W X Y Z
## 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26
```

## (3.) Subsetting by field names

```
myVec[c("B", "K", "M")]
```

```
## B K M
## 2 11 13
```

## (4.) Subset with \$ sign: references a single column or list component by its name

```
iris$Species[1:8]
```

```
## [1] setosa setosa setosa setosa setosa setosa setosa setosa
## Levels: setosa versicolor virginica
```

# Important Utilities

## Combining Objects

The `c` function combines vectors and lists

```
c(1, 2, 3)
```

```
## [1] 1 2 3
```

```
x <- 1:3; y <- 101:103
c(x, y)
```

```
## [1] 1 2 3 101 102 103
```

```
iris$Species[1:8]
```

```
## [1] setosa setosa setosa setosa setosa setosa setosa setosa
## Levels: setosa versicolor virginica
```

The `cbind` and `rbind` functions can be used to append columns and rows, respectively.

```
ma <- cbind(x, y)
ma
```

```
##      x      y
## [1,] 1 101
## [2,] 2 102
## [3,] 3 103
```

```
rbind(ma, ma)
```

```
##      x      y
## [1,] 1 101
## [2,] 2 102
## [3,] 3 103
## [4,] 1 101
## [5,] 2 102
```

```
## [6,] 3 103
```

## Accessing Dimensions of Objects

Length and dimension information of objects

```
length(iris$Species)
```

```
## [1] 150
```

```
dim(iris)
```

```
## [1] 150 5
```

## Accessing Name Slots of Objects

Accessing row and column names of 2D objects

```
rownames(iris)[1:8]
```

```
## [1] "1" "2" "3" "4" "5" "6" "7" "8"
```

```
colnames(iris)
```

```
## [1] "Sepal.Length" "Sepal.Width" "Petal.Length" "Petal.Width" "Species"
```

Return name field of vectors and lists

```
names(myVec)
```

```
## [1] "A" "B" "C" "D" "E" "F" "G" "H" "I" "J" "K" "L" "M" "N" "O" "P" "Q" "R" "S" "T" "U" "V" "W" "X"  
## [25] "Y" "Z"
```

```
names(myL)
```

```
## [1] "name" "wife" "no.children" "child.ages"
```

## Sorting Objects

The function `sort` returns a vector in ascending or descending order

```
sort(10:1)
```

```
## [1] 1 2 3 4 5 6 7 8 9 10
```

The function `order` returns a sorting index for sorting an object

```
sortindex <- order(iris[,1], decreasing = FALSE)
```

```
sortindex[1:12]
```

```
## [1] 14 9 39 43 42 4 7 23 48 3 30 12
```

```
iris[sortindex,][1:2,]
```

```
## Sepal.Length Sepal.Width Petal.Length Petal.Width Species  
## 14 4.3 3.0 1.1 0.1 setosa  
## 9 4.4 2.9 1.4 0.2 setosa
```

```
sortindex <- order(-iris[,1]) # Same as decreasing=TRUE
```



Sorting multiple columns

```
iris[order(iris$Sepal.Length, iris$Sepal.Width),][1:2,]
```

```
##      Sepal.Length Sepal.Width Petal.Length Petal.Width Species
## 14             4.3         3.0          1.1         0.1  setosa
## 9              4.4         2.9          1.4         0.2  setosa
```

## Operators and Calculations

### Comparison Operators

Comparison operators: ==, !=, <, >, <=, >=

```
1==1
```

```
## [1] TRUE
```

Logical operators: AND: &, OR: |, NOT: !

```
x <- 1:10; y <- 10:1
```

```
x > y & x > 5
```

```
## [1] FALSE FALSE FALSE FALSE FALSE  TRUE  TRUE  TRUE  TRUE  TRUE
```

### Basic Calculations

To look up math functions, see [Function Index](#) here

```
x + y
```

```
## [1] 11 11 11 11 11 11 11 11 11 11 11
```

```
sum(x)
```

```
## [1] 55
```

```
mean(x)
```

```
## [1] 5.5
```

```
apply(iris[1:6,1:3], 1, mean)
```

```
##      1      2      3      4      5      6
## 3.333333 3.100000 3.066667 3.066667 3.333333 3.666667
```

## Reading and Writing External Data

### Import of tabular data

Import of a tab-delimited tabular file

```
myDF <- read.delim("myData.xls", sep="\t")
```

Import of Excel file. Note: working with tab- or comma-delimited files is more flexible and preferred.

```
library(gdata)
myDF <- read.xls("myData.xls")
```

Import of Google Sheets. The following example imports a sample Google Sheet from here. Detailed instructions for interacting from R with Google Sheets with the required `googlesheets` package are here.

```
library("googlesheets"); library("dplyr"); library(knitr)
gs_auth() # Creates authorizaton token (.httr-oauth) in current directory if not present
sheetid <- "1U-32UcwZP1k3saKeaHimbvEA0fZRdNHNkWK2GI1rpPM"
gap <- gs_key(sheetid)
mysheet <- gs_read(gap, skip=4)
myDF <- as.data.frame(mysheet)
myDF
```

## Export of tabular data

```
write.table(myDF, file="myfile.xls", sep="\t", quote=FALSE, col.names=NA)
```

## Line-wise import

```
myDF <- readLines("myData.txt")
```

## Line-wise export

```
writeLines(month.name, "myData.txt")
```

## Copy and paste into R

On Windows/Linux systems

```
read.delim("clipboard")
```

On Mac OS X systems

```
read.delim(pipe("pbpaste"))
```

## Copy and paste from R

On Windows/Linux systems

```
write.table(iris, "clipboard", sep="\t", col.names=NA, quote=F)
```

On Mac OS X systems

```
zz <- pipe('pbcopy', 'w')
write.table(iris, zz, sep="\t", col.names=NA, quote=F)
close(zz)
```

## Homework 3A

Homework 3A: Object Subsetting Routines and Import/Export

## Useful R Functions

### Unique entries

Make vector entries unique with `unique`

```
length(iris$Sepal.Length)
```

```
## [1] 150
```

```
length(unique(iris$Sepal.Length))
```

```
## [1] 35
```

### Count occurrences

Count occurrences of entries with `table`

```
table(iris$Species)
```

```
##
```

```
##      setosa versicolor  virginica
```

```
##          50          50          50
```

### Aggregate data

Compute aggregate statistics with `aggregate`

```
aggregate(iris[,1:4], by=list(iris$Species), FUN=mean, na.rm=TRUE)
```

```
##      Group.1 Sepal.Length Sepal.Width Petal.Length Petal.Width
```

```
## 1      setosa      5.006      3.428      1.462      0.246
```

```
## 2 versicolor      5.936      2.770      4.260      1.326
```

```
## 3  virginica      6.588      2.974      5.552      2.026
```

### Intersect data

Compute intersect between two vectors with `%in%`

```
month.name %in% c("May", "July")
```

```
## [1] FALSE FALSE FALSE FALSE  TRUE FALSE  TRUE FALSE FALSE FALSE FALSE
```

### Merge data frames

Join two data frames by common field entries with `merge` (here row names by `x=0`). To obtain only the common rows, change `all=TRUE` to `all=FALSE`. To merge on specific columns, refer to them by their position numbers or their column names.

```

frame1 <- iris[sample(1:length(iris[,1]), 30), ]
frame1[1:2,]

##      Sepal.Length Sepal.Width Petal.Length Petal.Width  Species
## 110           7.2         3.6         6.1         2.5  virginica
## 60            5.2         2.7         3.9         1.4  versicolor
dim(frame1)

## [1] 30  5

my_result <- merge(frame1, iris, by.x = 0, by.y = 0, all = TRUE)
dim(my_result)

## [1] 150 11

```

## dplyr Environment

Modern object classes and methods for handling `data.frame` like structures are provided by the `dplyr` and `data.table` packages. The following gives a short introduction to the usage and functionalities of the `dplyr` package. More detailed tutorials on this topic can be found [here](#):

- `dplyr`: A Grammar of Data Manipulation
- Introduction to `dplyr`
- Tutorial on `dplyr`
- Cheatsheet for Joins from Jenny Bryan
- Tibbles
- Intro to `data.table` package
- Big data with `dplyr` and `data.table`
- Fast lookups with `dplyr` and `data.table`

## Installation

The `dplyr` environment has evolved into an ecosystem of packages. To simplify package management, one can install and load the entire collection via the `tidyverse` package. For more details on `tidyverse` see [here](#).

```
install.packages("tidyverse")
```

## Construct a data frame (tibble)

```

library(tidyverse)
as_data_frame(iris) # coerce data.frame to data frame tbl

## # A tibble: 150 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##   <dbl>         <dbl>         <dbl>         <dbl> <fctr>
## 1         5.1         3.5         1.4         0.2  setosa
## 2         4.9         3.0         1.4         0.2  setosa
## 3         4.7         3.2         1.3         0.2  setosa
## 4         4.6         3.1         1.5         0.2  setosa
## 5         5.0         3.6         1.4         0.2  setosa
## 6         5.4         3.9         1.7         0.4  setosa

```

```
## 7      4.6      3.4      1.4      0.3 setosa
## 8      5.0      3.4      1.5      0.2 setosa
## 9      4.4      2.9      1.4      0.2 setosa
## 10     4.9      3.1      1.5      0.1 setosa
## # ... with 140 more rows
```

Alternative functions producing the same result include `as_tibble` and `tbl_df`:

```
as_tibble(iris) # newer function provided by tibble package
tbl_df(iris) # this alternative exists for historical reasons
```

## Reading and writing tabular files

While the base R read/write utilities can be used for **data frames**, best time performance with the least amount of typing is achieved with the export/import functions from the **readr** package. For very large files the **fread** function from the **data.table** package achieves the best time performance.

### Import with readr

Import functions provided by **readr** include:

- `read_csv()`: comma separated (CSV) files
- `read_tsv()`: tab separated files
- `read_delim()`: general delimited files
- `read_fwf()`: fixed width files
- `read_table()`: tabular files where columns are separated by white-space.
- `read_log()`: web log files

Create a sample tab delimited file for import

```
write_tsv(iris, "iris.txt") # Creates sample file
```

Import with `read_tsv`

```
iris_df <- read_tsv("iris.txt") # Import with read_tsv from readr package
iris_df
```

```
## # A tibble: 150 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##   <dbl>         <dbl>         <dbl>         <dbl>    <chr>
## 1         5.1         3.5         1.4         0.2    setosa
## 2         4.9         3.0         1.4         0.2    setosa
## 3         4.7         3.2         1.3         0.2    setosa
## 4         4.6         3.1         1.5         0.2    setosa
## 5         5.0         3.6         1.4         0.2    setosa
## 6         5.4         3.9         1.7         0.4    setosa
## 7         4.6         3.4         1.4         0.3    setosa
## 8         5.0         3.4         1.5         0.2    setosa
## 9         4.4         2.9         1.4         0.2    setosa
## 10        4.9         3.1         1.5         0.1    setosa
## # ... with 140 more rows
```

To import Google Sheets directly into R, see [here](#).

## Fast table import with fread

The `fread` function from the `data.table` package provides the best time performance for reading large tabular files into R.

```
library(data.table)
iris_df <- as_data_frame(fread("iris.txt")) # Import with fread and conversion to tibble
iris_df
```

```
## # A tibble: 150 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##   <dbl>         <dbl>         <dbl>         <dbl>   <chr>
## 1         5.1         3.5         1.4         0.2   setosa
## 2         4.9         3.0         1.4         0.2   setosa
## 3         4.7         3.2         1.3         0.2   setosa
## 4         4.6         3.1         1.5         0.2   setosa
## 5         5.0         3.6         1.4         0.2   setosa
## 6         5.4         3.9         1.7         0.4   setosa
## 7         4.6         3.4         1.4         0.3   setosa
## 8         5.0         3.4         1.5         0.2   setosa
## 9         4.4         2.9         1.4         0.2   setosa
## 10        4.9         3.1         1.5         0.1   setosa
## # ... with 140 more rows
```

Note: to ignore lines starting with comment signs, one can pass on to `fread` a shell command for preprocessing the file. The following example illustrates this option.

```
fread("grep -v '^#' iris.txt")
```

## Export with readr

Export function provided by `readr` include

- `write_delim()`: general delimited files
- `write_csv()`: comma separated (CSV) files
- `write_excel_csv()`: excel style CSV files
- `write_tsv()`: tab separated files

For instance, the `write_tsv` function writes a `data frame` to a tab delimited file with much nicer default settings than the base R `write.table` function.

```
write_tsv(iris_df, "iris.txt")
```

## Column and row binds

The equivalents to base R's `rbind` and `cbind` are `bind_rows` and `bind_cols`, respectively.

```
bind_cols(iris_df, iris_df)
```

```
## # A tibble: 150 × 10
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species Sepal.Length Sepal.Width Petal.Length
##   <dbl>         <dbl>         <dbl>         <dbl>   <chr>         <dbl>         <dbl>         <dbl>
## 1         5.1         3.5         1.4         0.2   setosa         5.1         3.5         1.4
## 2         4.9         3.0         1.4         0.2   setosa         4.9         3.0         1.4
## 3         4.7         3.2         1.3         0.2   setosa         4.7         3.2         1.3
## 4         4.6         3.1         1.5         0.2   setosa         4.6         3.1         1.5
```

```
## 5      5.0      3.6      1.4      0.2 setosa      5.0      3.6      1.4
## 6      5.4      3.9      1.7      0.4 setosa      5.4      3.9      1.7
## 7      4.6      3.4      1.4      0.3 setosa      4.6      3.4      1.4
## 8      5.0      3.4      1.5      0.2 setosa      5.0      3.4      1.5
## 9      4.4      2.9      1.4      0.2 setosa      4.4      2.9      1.4
## 10     4.9      3.1      1.5      0.1 setosa      4.9      3.1      1.5
## # ... with 140 more rows, and 2 more variables: Petal.Width <dbl>, Species <chr>

bind_rows(iris_df, iris_df)

## # A tibble: 300 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##   <dbl>        <dbl>        <dbl>        <dbl>    <chr>
## 1         5.1         3.5         1.4         0.2  setosa
## 2         4.9         3.0         1.4         0.2  setosa
## 3         4.7         3.2         1.3         0.2  setosa
## 4         4.6         3.1         1.5         0.2  setosa
## 5         5.0         3.6         1.4         0.2  setosa
## 6         5.4         3.9         1.7         0.4  setosa
## 7         4.6         3.4         1.4         0.3  setosa
## 8         5.0         3.4         1.5         0.2  setosa
## 9         4.4         2.9         1.4         0.2  setosa
## 10        4.9         3.1         1.5         0.1  setosa
## # ... with 290 more rows
```

## Extract column as vector

The subsetting operators `[]` and `$` can be used to extract from a data frame single columns as vector.

```
iris_df[[5]][1:12]

## [1] "setosa" "setosa" "setosa" "setosa" "setosa" "setosa" "setosa" "setosa" "setosa" "setosa"
## [11] "setosa" "setosa"

iris_df$Species[1:12]

## [1] "setosa" "setosa" "setosa" "setosa" "setosa" "setosa" "setosa" "setosa" "setosa" "setosa"
## [11] "setosa" "setosa"
```

## Important dplyr functions

1. `filter()` and `slice()`
2. `arrange()`
3. `select()` and `rename()`
4. `distinct()`
5. `mutate()` and `transmute()`
6. `summarise()`
7. `sample_n()` and `sample_frac()`

## Slice and filter functions

### Filter function

```
filter(iris_df, Sepal.Length > 7.5, Species=="virginica")
```

```
## # A tibble: 6 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width  Species
##   <dbl>        <dbl>        <dbl>        <dbl>    <chr>
## 1         7.6         3.0         6.6         2.1  virginica
## 2         7.7         3.8         6.7         2.2  virginica
## 3         7.7         2.6         6.9         2.3  virginica
## 4         7.7         2.8         6.7         2.0  virginica
## 5         7.9         3.8         6.4         2.0  virginica
## 6         7.7         3.0         6.1         2.3  virginica
```

### Base R code equivalent

```
iris_df[iris_df[, "Sepal.Length"] > 7.5 & iris_df[, "Species"]=="virginica", ]
```

```
## # A tibble: 6 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width  Species
##   <dbl>        <dbl>        <dbl>        <dbl>    <chr>
## 1         7.6         3.0         6.6         2.1  virginica
## 2         7.7         3.8         6.7         2.2  virginica
## 3         7.7         2.6         6.9         2.3  virginica
## 4         7.7         2.8         6.7         2.0  virginica
## 5         7.9         3.8         6.4         2.0  virginica
## 6         7.7         3.0         6.1         2.3  virginica
```

### Including boolean operators

```
filter(iris_df, Sepal.Length > 7.5 | Sepal.Length < 5.5, Species=="virginica")
```

```
## # A tibble: 7 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width  Species
##   <dbl>        <dbl>        <dbl>        <dbl>    <chr>
## 1         7.6         3.0         6.6         2.1  virginica
## 2         4.9         2.5         4.5         1.7  virginica
## 3         7.7         3.8         6.7         2.2  virginica
## 4         7.7         2.6         6.9         2.3  virginica
## 5         7.7         2.8         6.7         2.0  virginica
## 6         7.9         3.8         6.4         2.0  virginica
## 7         7.7         3.0         6.1         2.3  virginica
```

### Subset rows by position

dplyr approach

```
slice(iris_df, 1:2)
```



```
## # A tibble: 2 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##         <dbl>      <dbl>      <dbl>      <dbl>    <chr>
## 1         5.1        3.5        1.4        0.2   setosa
## 2         4.9        3.0        1.4        0.2   setosa
```

Base R code equivalent

```
iris_df[1:2,]
```

```
## # A tibble: 2 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##         <dbl>      <dbl>      <dbl>      <dbl>    <chr>
## 1         5.1        3.5        1.4        0.2   setosa
## 2         4.9        3.0        1.4        0.2   setosa
```

### Subset rows by names

Since data frames do not contain row names, row wise subsetting via the `[,]` operator cannot be used. However, the corresponding behavior can be achieved by passing to `select` a row position index obtained by basic R intersect utilities such as `match`.

Create a suitable test data frame

```
df1 <- bind_cols(data_frame(ids1=paste0("g", 1:10)), as_data_frame(matrix(1:40, 10, 4, dimnames=list(1:10, 1:4))))
df1
```

```
## # A tibble: 10 × 5
##   ids1    CA1    CA2    CA3    CA4
##   <chr> <int> <int> <int> <int>
## 1    g1      1    11    21    31
## 2    g2      2    12    22    32
## 3    g3      3    13    23    33
## 4    g4      4    14    24    34
## 5    g5      5    15    25    35
## 6    g6      6    16    26    36
## 7    g7      7    17    27    37
## 8    g8      8    18    28    38
## 9    g9      9    19    29    39
## 10   g10     10    20    30    40
```

dplyr approach

```
slice(df1, match(c("g10", "g4", "g4"), df1$ids1))
```

```
## # A tibble: 3 × 5
##   ids1    CA1    CA2    CA3    CA4
##   <chr> <int> <int> <int> <int>
## 1   g10     10     20     30     40
## 2    g4      4     14     24     34
## 3    g4      4     14     24     34
```

Base R equivalent

```
df1_old <- as.data.frame(df1)
rownames(df1_old) <- df1_old[,1]
df1_old[c("g10", "g4", "g4"),]
```

```
##      ids1 CA1 CA2 CA3 CA4
## g10   g10 10  20  30  40
## g4    g4   4  14  24  34
## g4.1  g4   4  14  24  34
```

## Sorting with arrange

Row-wise ordering based on specific columns

dplyr approach

```
arrange(iris_df, Species, Sepal.Length, Sepal.Width)
```

```
## # A tibble: 150 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##         <dbl>         <dbl>         <dbl>         <dbl>    <chr>
## 1           4.3           3.0           1.1           0.1  setosa
## 2           4.4           2.9           1.4           0.2  setosa
## 3           4.4           3.0           1.3           0.2  setosa
## 4           4.4           3.2           1.3           0.2  setosa
## 5           4.5           2.3           1.3           0.3  setosa
## 6           4.6           3.1           1.5           0.2  setosa
## 7           4.6           3.2           1.4           0.2  setosa
## 8           4.6           3.4           1.4           0.3  setosa
## 9           4.6           3.6           1.0           0.2  setosa
## 10          4.7           3.2           1.3           0.2  setosa
## # ... with 140 more rows
```

For ordering descendingly use desc() function

```
arrange(iris_df, desc(Species), Sepal.Length, Sepal.Width)
```

```
## # A tibble: 150 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##         <dbl>         <dbl>         <dbl>         <dbl>    <chr>
## 1           4.9           2.5           4.5           1.7 virginica
## 2           5.6           2.8           4.9           2.0 virginica
## 3           5.7           2.5           5.0           2.0 virginica
## 4           5.8           2.7           5.1           1.9 virginica
## 5           5.8           2.7           5.1           1.9 virginica
## 6           5.8           2.8           5.1           2.4 virginica
## 7           5.9           3.0           5.1           1.8 virginica
## 8           6.0           2.2           5.0           1.5 virginica
## 9           6.0           3.0           4.8           1.8 virginica
## 10          6.1           2.6           5.6           1.4 virginica
## # ... with 140 more rows
```

Base R code equivalent

```
iris_df[order(iris_df$Species, iris_df$Sepal.Length, iris_df$Sepal.Width), ]
```

```
## # A tibble: 150 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##         <dbl>         <dbl>         <dbl>         <dbl>    <chr>
## 1           4.3           3.0           1.1           0.1  setosa
## 2           4.4           2.9           1.4           0.2  setosa
## 3           4.4           3.0           1.3           0.2  setosa
```

```
## 4      4.4      3.2      1.3      0.2 setosa
## 5      4.5      2.3      1.3      0.3 setosa
## 6      4.6      3.1      1.5      0.2 setosa
## 7      4.6      3.2      1.4      0.2 setosa
## 8      4.6      3.4      1.4      0.3 setosa
## 9      4.6      3.6      1.0      0.2 setosa
## 10     4.7      3.2      1.3      0.2 setosa
## # ... with 140 more rows
```

```
iris_df[order(iris_df$Species, decreasing=TRUE), ]
```

```
## # A tibble: 150 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##   <dbl>         <dbl>         <dbl>         <dbl>    <chr>
## 1         6.3         3.3         6.0         2.5 virginica
## 2         5.8         2.7         5.1         1.9 virginica
## 3         7.1         3.0         5.9         2.1 virginica
## 4         6.3         2.9         5.6         1.8 virginica
## 5         6.5         3.0         5.8         2.2 virginica
## 6         7.6         3.0         6.6         2.1 virginica
## 7         4.9         2.5         4.5         1.7 virginica
## 8         7.3         2.9         6.3         1.8 virginica
## 9         6.7         2.5         5.8         1.8 virginica
## 10        7.2         3.6         6.1         2.5 virginica
## # ... with 140 more rows
```

## Select columns with select

Select specific columns

```
select(iris_df, Species, Petal.Length, Sepal.Length)
```

```
## # A tibble: 150 × 3
##   Species Petal.Length Sepal.Length
##   <chr>         <dbl>         <dbl>
## 1 setosa         1.4         5.1
## 2 setosa         1.4         4.9
## 3 setosa         1.3         4.7
## 4 setosa         1.5         4.6
## 5 setosa         1.4         5.0
## 6 setosa         1.7         5.4
## 7 setosa         1.4         4.6
## 8 setosa         1.5         5.0
## 9 setosa         1.4         4.4
## 10 setosa        1.5         4.9
## # ... with 140 more rows
```

Select range of columns by name

```
select(iris_df, Sepal.Length : Petal.Width)
```

```
## # A tibble: 150 × 4
##   Sepal.Length Sepal.Width Petal.Length Petal.Width
##   <dbl>         <dbl>         <dbl>         <dbl>
## 1         5.1         3.5         1.4         0.2
## 2         4.9         3.0         1.4         0.2
```

```
## 3      4.7      3.2      1.3      0.2
## 4      4.6      3.1      1.5      0.2
## 5      5.0      3.6      1.4      0.2
## 6      5.4      3.9      1.7      0.4
## 7      4.6      3.4      1.4      0.3
## 8      5.0      3.4      1.5      0.2
## 9      4.4      2.9      1.4      0.2
## 10     4.9      3.1      1.5      0.1
## # ... with 140 more rows
```

Drop specific columns (here range)

```
select(iris_df, -(Sepal.Length : Petal.Width))
```

```
## # A tibble: 150 × 1
##   Species
##   <chr>
## 1  setosa
## 2  setosa
## 3  setosa
## 4  setosa
## 5  setosa
## 6  setosa
## 7  setosa
## 8  setosa
## 9  setosa
## 10 setosa
## # ... with 140 more rows
```

## Renaming columns with rename

dplyr approach

```
rename(iris_df, new_col_name = Species)
```

```
## # A tibble: 150 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width new_col_name
##   <dbl>         <dbl>         <dbl>         <dbl>         <chr>
## 1      5.1      3.5      1.4      0.2      setosa
## 2      4.9      3.0      1.4      0.2      setosa
## 3      4.7      3.2      1.3      0.2      setosa
## 4      4.6      3.1      1.5      0.2      setosa
## 5      5.0      3.6      1.4      0.2      setosa
## 6      5.4      3.9      1.7      0.4      setosa
## 7      4.6      3.4      1.4      0.3      setosa
## 8      5.0      3.4      1.5      0.2      setosa
## 9      4.4      2.9      1.4      0.2      setosa
## 10     4.9      3.1      1.5      0.1      setosa
## # ... with 140 more rows
```

Base R code approach

```
colnames(iris_df)[colnames(iris_df)=="Species"] <- "new_col_names"
```

## Obtain unique rows with distinct

dplyr approach

```
distinct(iris_df, Species, .keep_all=TRUE)
```

```
## # A tibble: 3 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##   <dbl>         <dbl>         <dbl>         <dbl>     <chr>
## 1         5.1         3.5         1.4         0.2     setosa
## 2         7.0         3.2         4.7         1.4 versicolor
## 3         6.3         3.3         6.0         2.5  virginica
```

Base R code approach

```
iris_df[!duplicated(iris_df$Species),]
```

```
## # A tibble: 3 × 5
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species
##   <dbl>         <dbl>         <dbl>         <dbl>     <chr>
## 1         5.1         3.5         1.4         0.2     setosa
## 2         7.0         3.2         4.7         1.4 versicolor
## 3         6.3         3.3         6.0         2.5  virginica
```

## Add columns

mutate

The mutate function allows to append columns to existing ones.

```
mutate(iris_df, Ratio = Sepal.Length / Sepal.Width, Sum = Sepal.Length + Sepal.Width)
```

```
## # A tibble: 150 × 7
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species Ratio Sum
##   <dbl>         <dbl>         <dbl>         <dbl>     <chr> <dbl> <dbl>
## 1         5.1         3.5         1.4         0.2     setosa 1.457143 8.6
## 2         4.9         3.0         1.4         0.2     setosa 1.633333 7.9
## 3         4.7         3.2         1.3         0.2     setosa 1.468750 7.9
## 4         4.6         3.1         1.5         0.2     setosa 1.483871 7.7
## 5         5.0         3.6         1.4         0.2     setosa 1.388889 8.6
## 6         5.4         3.9         1.7         0.4     setosa 1.384615 9.3
## 7         4.6         3.4         1.4         0.3     setosa 1.352941 8.0
## 8         5.0         3.4         1.5         0.2     setosa 1.470588 8.4
## 9         4.4         2.9         1.4         0.2     setosa 1.517241 7.3
## 10        4.9         3.1         1.5         0.1     setosa 1.580645 8.0
## # ... with 140 more rows
```

transmute

The transmute function does the same as mutate but drops existing columns

```
transmute(iris_df, Ratio = Sepal.Length / Sepal.Width, Sum = Sepal.Length + Sepal.Width)
```

```
## # A tibble: 150 × 2
##   Ratio Sum
##   <dbl> <dbl>
```

```
## 1  1.457143  8.6
## 2  1.633333  7.9
## 3  1.468750  7.9
## 4  1.483871  7.7
## 5  1.388889  8.6
## 6  1.384615  9.3
## 7  1.352941  8.0
## 8  1.470588  8.4
## 9  1.517241  7.3
## 10 1.580645  8.0
## # ... with 140 more rows
```

`bind_cols`

The `bind_cols` function is the equivalent of `cbind` in base R. To add rows, use the corresponding `bind_rows` function.

```
bind_cols(iris_df, iris_df)
```

```
## # A tibble: 150 × 10
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species Sepal.Length Sepal.Width Petal.Length
##   <dbl>         <dbl>         <dbl>         <dbl>   <chr>         <dbl>         <dbl>         <dbl>
## 1         5.1         3.5         1.4         0.2   setosa         5.1         3.5         1.4
## 2         4.9         3.0         1.4         0.2   setosa         4.9         3.0         1.4
## 3         4.7         3.2         1.3         0.2   setosa         4.7         3.2         1.3
## 4         4.6         3.1         1.5         0.2   setosa         4.6         3.1         1.5
## 5         5.0         3.6         1.4         0.2   setosa         5.0         3.6         1.4
## 6         5.4         3.9         1.7         0.4   setosa         5.4         3.9         1.7
## 7         4.6         3.4         1.4         0.3   setosa         4.6         3.4         1.4
## 8         5.0         3.4         1.5         0.2   setosa         5.0         3.4         1.5
## 9         4.4         2.9         1.4         0.2   setosa         4.4         2.9         1.4
## 10        4.9         3.1         1.5         0.1   setosa         4.9         3.1         1.5
## # ... with 140 more rows, and 2 more variables: Petal.Width <dbl>, Species <chr>
```

## Summarize data

Summary calculation on single column

```
summarize(iris_df, mean(Petal.Length))
```

```
## # A tibble: 1 × 1
##   `mean(Petal.Length)`
##   <dbl>
## 1         3.758
```

Summary calculation on many columns

```
summarize_all(iris_df[,1:4], mean)
```

```
## # A tibble: 1 × 4
##   Sepal.Length Sepal.Width Petal.Length Petal.Width
##   <dbl>         <dbl>         <dbl>         <dbl>
## 1     5.843333     3.057333         3.758         1.199333
```

Summarize by grouping column

```
summarize(group_by(iris_df, Species), mean(Petal.Length))
```

```
## # A tibble: 3 × 2
##   Species `mean(Petal.Length)`
##   <chr>      <dbl>
## 1 setosa      1.462
## 2 versicolor  4.260
## 3 virginica   5.552
```

Aggregate summaries

```
summarize_all(group_by(iris_df, Species), mean)
```

```
## # A tibble: 3 × 5
##   Species Sepal.Length Sepal.Width Petal.Length Petal.Width
##   <chr>      <dbl>      <dbl>      <dbl>      <dbl>
## 1 setosa      5.006      3.428      1.462      0.246
## 2 versicolor  5.936      2.770      4.260      1.326
## 3 virginica   6.588      2.974      5.552      2.026
```

Note: `group_by` does the looping for the user similar to `aggregate` or `tapply`.

## Merging data frames

The `dplyr` package provides several join functions for merging **data frames** by a common key column similar to the `merge` function in base R. These `*_join` functions include:

- `inner_join()`: returns join only for rows matching among both **data tables**
- `full_join()`: returns join for all (matching and non-matching) rows of two **data tables**
- `left_join()`: returns join for all rows in first **data table**
- `right_join()`: returns join for all rows in second **data table**
- `anti_join()`: returns for first **data table** only those rows that have no match in the second one

Sample data frames to illustrate `*.join` functions.

```
df1 <- bind_cols(data_frame(ids1=paste0("g", 1:10)), as_data_frame(matrix(1:40, 10, 4, dimnames=list(1:10, 1:4))))
df1
```

```
## # A tibble: 10 × 5
##   ids1  CA1  CA2  CA3  CA4
##   <chr> <int> <int> <int> <int>
## 1 g1      1    11    21    31
## 2 g2      2    12    22    32
## 3 g3      3    13    23    33
## 4 g4      4    14    24    34
## 5 g5      5    15    25    35
## 6 g6      6    16    26    36
## 7 g7      7    17    27    37
## 8 g8      8    18    28    38
## 9 g9      9    19    29    39
## 10 g10    10    20    30    40
```

```
df2 <- bind_cols(data_frame(ids2=paste0("g", c(2,5,11,12))), as_data_frame(matrix(1:16, 4, 4, dimnames=list(1:4, 1:4))))
df2
```

```
## # A tibble: 4 × 5
##   ids2  CB1  CB2  CB3  CB4
```

```
##   <chr> <int> <int> <int> <int>
## 1    g2      1      5      9     13
## 2    g5      2      6     10     14
## 3   g11      3      7     11     15
## 4   g12      4      8     12     16
```

## Inner join

```
inner_join(df1, df2, by=c("ids1"="ids2"))
```

```
## # A tibble: 2 × 9
##   ids1  CA1  CA2  CA3  CA4  CB1  CB2  CB3  CB4
##   <chr> <int> <int> <int> <int> <int> <int> <int> <int>
## 1    g2      2    12    22    32      1      5      9    13
## 2    g5      5    15    25    35      2      6     10    14
```

## Left join

```
left_join(df1, df2, by=c("ids1"="ids2"))
```

```
## # A tibble: 10 × 9
##   ids1  CA1  CA2  CA3  CA4  CB1  CB2  CB3  CB4
##   <chr> <int> <int> <int> <int> <int> <int> <int> <int>
## 1    g1      1    11    21    31    NA    NA    NA    NA
## 2    g2      2    12    22    32      1      5      9    13
## 3    g3      3    13    23    33    NA    NA    NA    NA
## 4    g4      4    14    24    34    NA    NA    NA    NA
## 5    g5      5    15    25    35      2      6     10    14
## 6    g6      6    16    26    36    NA    NA    NA    NA
## 7    g7      7    17    27    37    NA    NA    NA    NA
## 8    g8      8    18    28    38    NA    NA    NA    NA
## 9    g9      9    19    29    39    NA    NA    NA    NA
## 10   g10     10    20    30    40    NA    NA    NA    NA
```

## Right join

```
right_join(df1, df2, by=c("ids1"="ids2"))
```

```
## # A tibble: 4 × 9
##   ids1  CA1  CA2  CA3  CA4  CB1  CB2  CB3  CB4
##   <chr> <int> <int> <int> <int> <int> <int> <int> <int>
## 1    g2      2    12    22    32      1      5      9    13
## 2    g5      5    15    25    35      2      6     10    14
## 3   g11     NA     NA     NA     NA      3      7     11    15
## 4   g12     NA     NA     NA     NA      4      8     12    16
```

## Full join

```
full_join(df1, df2, by=c("ids1"="ids2"))
```



```
## # A tibble: 12 × 9
##   ids1    CA1    CA2    CA3    CA4    CB1    CB2    CB3    CB4
##   <chr> <int> <int> <int> <int> <int> <int> <int> <int>
## 1    g1      1    11    21    31    NA    NA    NA    NA
## 2    g2      2    12    22    32      1     5     9    13
## 3    g3      3    13    23    33    NA    NA    NA    NA
## 4    g4      4    14    24    34    NA    NA    NA    NA
## 5    g5      5    15    25    35      2     6    10    14
## 6    g6      6    16    26    36    NA    NA    NA    NA
## 7    g7      7    17    27    37    NA    NA    NA    NA
## 8    g8      8    18    28    38    NA    NA    NA    NA
## 9    g9      9    19    29    39    NA    NA    NA    NA
## 10   g10     10    20    30    40    NA    NA    NA    NA
## 11   g11     NA    NA    NA    NA      3     7    11    15
## 12   g12     NA    NA    NA    NA      4     8    12    16
```

## Anti join

```
anti_join(df1, df2, by=c("ids1"="ids2"))
```

```
## # A tibble: 8 × 5
##   ids1    CA1    CA2    CA3    CA4
##   <chr> <int> <int> <int> <int>
## 1   g10     10     20     30     40
## 2    g9      9     19     29     39
## 3    g8      8     18     28     38
## 4    g7      7     17     27     37
## 5    g6      6     16     26     36
## 6    g4      4     14     24     34
## 7    g3      3     13     23     33
## 8    g1      1     11     21     31
```

For additional join options users want to consult the `*_join` help pages.

## Chaining

To simplify chaining of several operations, `dplyr` provides the `%>%` operator. where `x %>% f(y)` turns into `f(x, y)`. This way one can pipe together multiple operations by writing them from left-to-right or top-to-bottom. This makes for easy to type and readable code.

### Example 1

Series of data manipulations and export

```
iris_df %>% # Declare data frame to use
  select(Sepal.Length:Species) %>% # Select columns
  filter(Species=="setosa") %>% # Filter rows by some value
  arrange(Sepal.Length) %>% # Sort by some column
  mutate(Subtract=Petal.Length - Petal.Width) # Calculate and append
```

```
## # A tibble: 50 × 6
##   Sepal.Length Sepal.Width Petal.Length Petal.Width Species Subtract
##   <dbl>         <dbl>         <dbl>         <dbl>    <chr>    <dbl>
```

```
## 1      4.3      3.0      1.1      0.1 setosa      1.0
## 2      4.4      2.9      1.4      0.2 setosa      1.2
## 3      4.4      3.0      1.3      0.2 setosa      1.1
## 4      4.4      3.2      1.3      0.2 setosa      1.1
## 5      4.5      2.3      1.3      0.3 setosa      1.0
## 6      4.6      3.1      1.5      0.2 setosa      1.3
## 7      4.6      3.4      1.4      0.3 setosa      1.1
## 8      4.6      3.6      1.0      0.2 setosa      0.8
## 9      4.6      3.2      1.4      0.2 setosa      1.2
## 10     4.7      3.2      1.3      0.2 setosa      1.1
## # ... with 40 more rows

# write_tsv("iris.txt") # Export to file, omitted here to show result
```

## Example 2

Series of summary calculations for grouped data (group\_by)

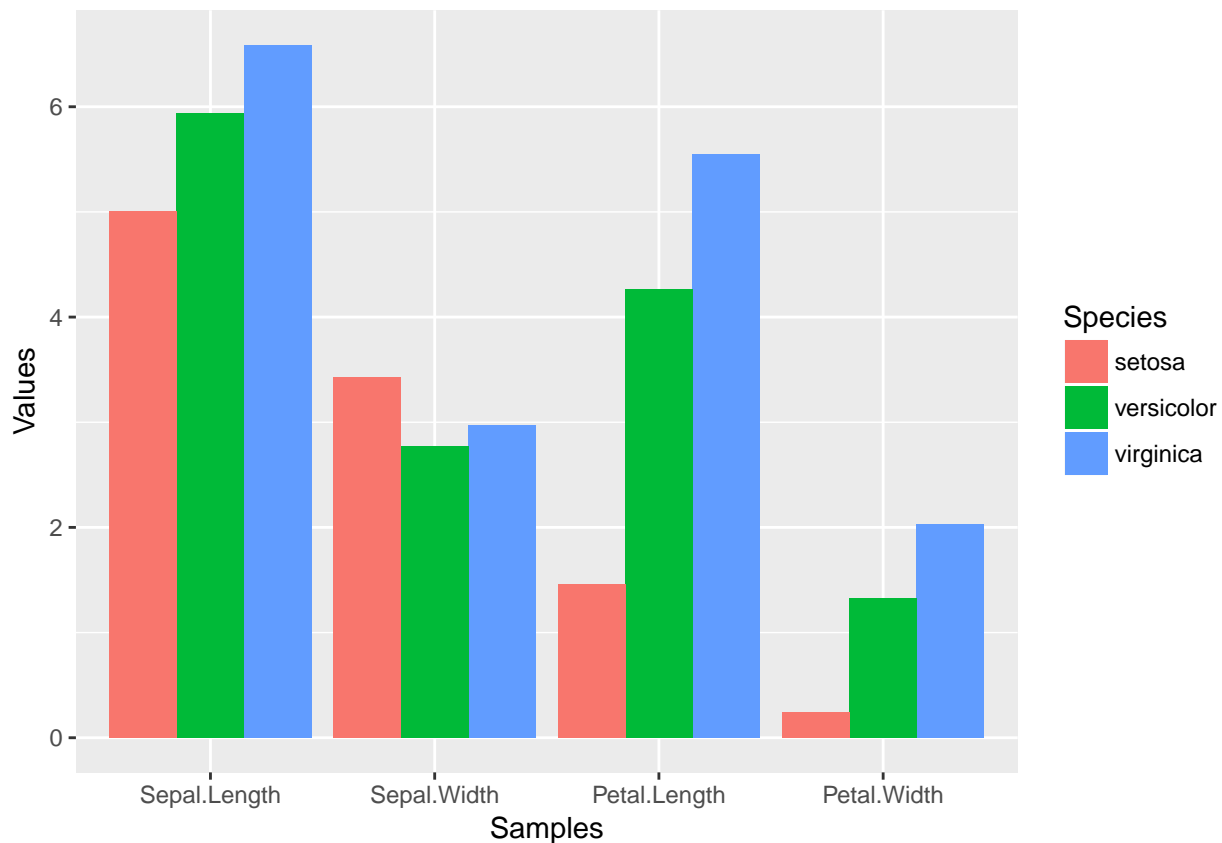
```
iris_df %>% # Declare data frame to use
  group_by(Species) %>% # Group by species
  summarize(Mean_Sepal.Length=mean(Sepal.Length),
            Max_Sepal.Length=max(Sepal.Length),
            Min_Sepal.Length=min(Sepal.Length),
            SD_Sepal.Length=sd(Sepal.Length),
            Total=n())
```

```
## # A tibble: 3 × 6
##   Species Mean_Sepal.Length Max_Sepal.Length Min_Sepal.Length SD_Sepal.Length Total
##   <chr>      <dbl>          <dbl>          <dbl>          <dbl> <int>
## 1 setosa      5.006            5.8            4.3            0.3524897 50
## 2 versicolor 5.936            7.0            4.9            0.5161711 50
## 3 virginica   6.588            7.9            4.9            0.6358796 50
```

## Example 3

Combining dplyr chaining with ggplot

```
iris_df %>%
  group_by(Species) %>%
  summarize_all(mean) %>%
  reshape2::melt(id.vars=c("Species"), variable.name = "Samples", value.name="Values") %>%
  ggplot(aes(Samples, Values, fill = Species)) +
  geom_bar(position="dodge", stat="identity")
```



## SQLite Databases

SQLite is a lightweight relational database solution. The RSQLite package provides an easy to use interface to create, manage and query SQLite databases directly from R. Basic instructions for using SQLite from the command-line are available [here](#). A short introduction to RSQLite is available [here](#).

### Loading data into SQLite databases

The following loads two `data.frames` derived from the `iris` data set (here `mydf1` and `mydf2`) into an SQLite database (here `test.db`).

```
library(RSQLite)
mydb <- dbConnect(SQLite(), "test.db") # Creates database file test.db
mydf1 <- data.frame(ids=paste0("id", seq_along(iris[,1])), iris)
mydf2 <- mydf1[sample(seq_along(mydf1[,1]), 10),]
dbWriteTable(mydb, "mydf1", mydf1)
```

```
## [1] TRUE
```

```
dbWriteTable(mydb, "mydf2", mydf2)
```

```
## [1] TRUE
```

## List names of tables in database

```
dbListTables(mydb)
```

```
## [1] "mydf1" "mydf2"
```

## Import table into data.frame

```
dbGetQuery(mydb, 'SELECT * FROM mydf2')
```

##	ids	Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
## 1	id8	5.0	3.4	1.5	0.2	setosa
## 2	id28	5.2	3.5	1.5	0.2	setosa
## 3	id111	6.5	3.2	5.1	2.0	virginica
## 4	id65	5.6	2.9	3.6	1.3	versicolor
## 5	id92	6.1	3.0	4.6	1.4	versicolor
## 6	id108	7.3	2.9	6.3	1.8	virginica
## 7	id140	6.9	3.1	5.4	2.1	virginica
## 8	id37	5.5	3.5	1.3	0.2	setosa
## 9	id101	6.3	3.3	6.0	2.5	virginica
## 10	id118	7.7	3.8	6.7	2.2	virginica

## Query database

```
dbGetQuery(mydb, 'SELECT * FROM mydf1 WHERE "Sepal.Length" < 4.6')
```

##	ids	Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
## 1	id9	4.4	2.9	1.4	0.2	setosa
## 2	id14	4.3	3.0	1.1	0.1	setosa
## 3	id39	4.4	3.0	1.3	0.2	setosa
## 4	id42	4.5	2.3	1.3	0.3	setosa
## 5	id43	4.4	3.2	1.3	0.2	setosa

## Join tables

The two tables can be joined on the shared `ids` column as follows.

```
dbGetQuery(mydb, 'SELECT * FROM mydf1, mydf2 WHERE mydf1.ids = mydf2.ids')
```

##	ids	Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species	ids	Sepal.Length
## 1	id8	5.0	3.4	1.5	0.2	setosa	id8	5.0
## 2	id28	5.2	3.5	1.5	0.2	setosa	id28	5.2
## 3	id37	5.5	3.5	1.3	0.2	setosa	id37	5.5
## 4	id65	5.6	2.9	3.6	1.3	versicolor	id65	5.6
## 5	id92	6.1	3.0	4.6	1.4	versicolor	id92	6.1
## 6	id101	6.3	3.3	6.0	2.5	virginica	id101	6.3
## 7	id108	7.3	2.9	6.3	1.8	virginica	id108	7.3
## 8	id111	6.5	3.2	5.1	2.0	virginica	id111	6.5
## 9	id118	7.7	3.8	6.7	2.2	virginica	id118	7.7
## 10	id140	6.9	3.1	5.4	2.1	virginica	id140	6.9

##	Sepal.Width	Petal.Length	Petal.Width	Species
----	-------------	--------------	-------------	---------

## 1	3.4	1.5	0.2	setosa
## 2	3.5	1.5	0.2	setosa
## 3	3.5	1.3	0.2	setosa
## 4	2.9	3.6	1.3	versicolor
## 5	3.0	4.6	1.4	versicolor
## 6	3.3	6.0	2.5	virginica
## 7	2.9	6.3	1.8	virginica
## 8	3.2	5.1	2.0	virginica
## 9	3.8	6.7	2.2	virginica
## 10	3.1	5.4	2.1	virginica

## Graphics in R

### Advantages

- Powerful environment for visualizing scientific data
- Integrated graphics and statistics infrastructure
- Publication quality graphics
- Fully programmable
- Highly reproducible
- Full LaTeX and Markdown support via `knitr` and R `markdown`
- Vast number of R packages with graphics utilities

### Documentation for R Graphics

#### General

- Graphics Task Page - URL
- R Graph Gallery - URL
- R Graphical Manual - URL
- Paul Murrell's book R (Grid) Graphics - URL

#### Interactive graphics

- `rggobi` (GGobi) - URL
- `iplots` - URL
- Open GL (`rgl`) - URL

### Graphics Environments

#### Viewing and saving graphics in R

- On-screen graphics
- `postscript`, `pdf`, `svg`
- `jpeg`, `png`, `wmf`, `tiff`, ...

#### Four major graphic environments

(a) Low-level infrastructure

- R Base Graphics (low- and high-level)
- `grid`: Manual

(b) High-level infrastructure \begin{itemize}

- `lattice`: Manual, Intro, Book
- `ggplot2`: Manual, Intro, Book

## Base Graphics: Overview

### Important high-level plotting functions

- `plot`: generic x-y plotting
- `barplot`: bar plots
- `boxplot`: box-and-whisker plot
- `hist`: histograms
- `pie`: pie charts
- `dotchart`: cleveland dot plots
- `image`, `heatmap`, `contour`, `persp`: functions to generate image-like plots
- `qqnorm`, `qqline`, `qqplot`: distribution comparison plots
- `pairs`, `coplot`: display of multivariate data

### Help on graphics functions

- `?myfct`
- `?plot`
- `?par`

### Preferred Object Types

- Matrices and data frames
- Vectors
- Named vectors

## Scatter Plots

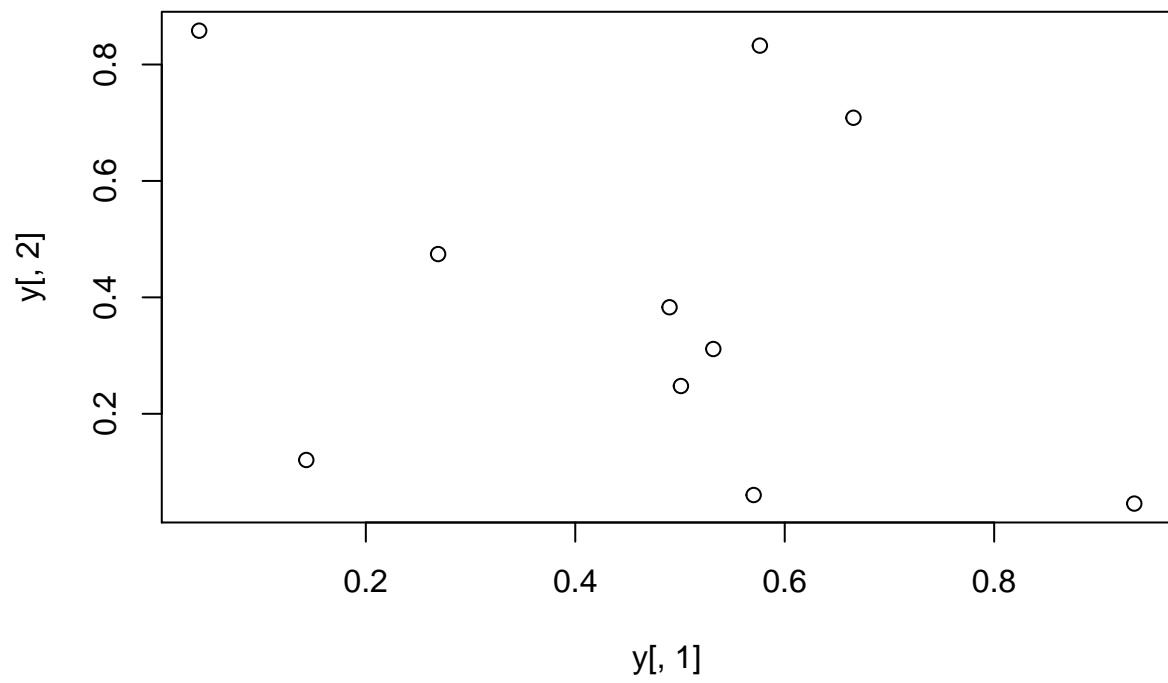
### Basic Scatter Plot

Sample data set for subsequent plots

```
set.seed(1410)
y <- matrix(runif(30), ncol=3, dimnames=list(letters[1:10], LETTERS[1:3]))
```

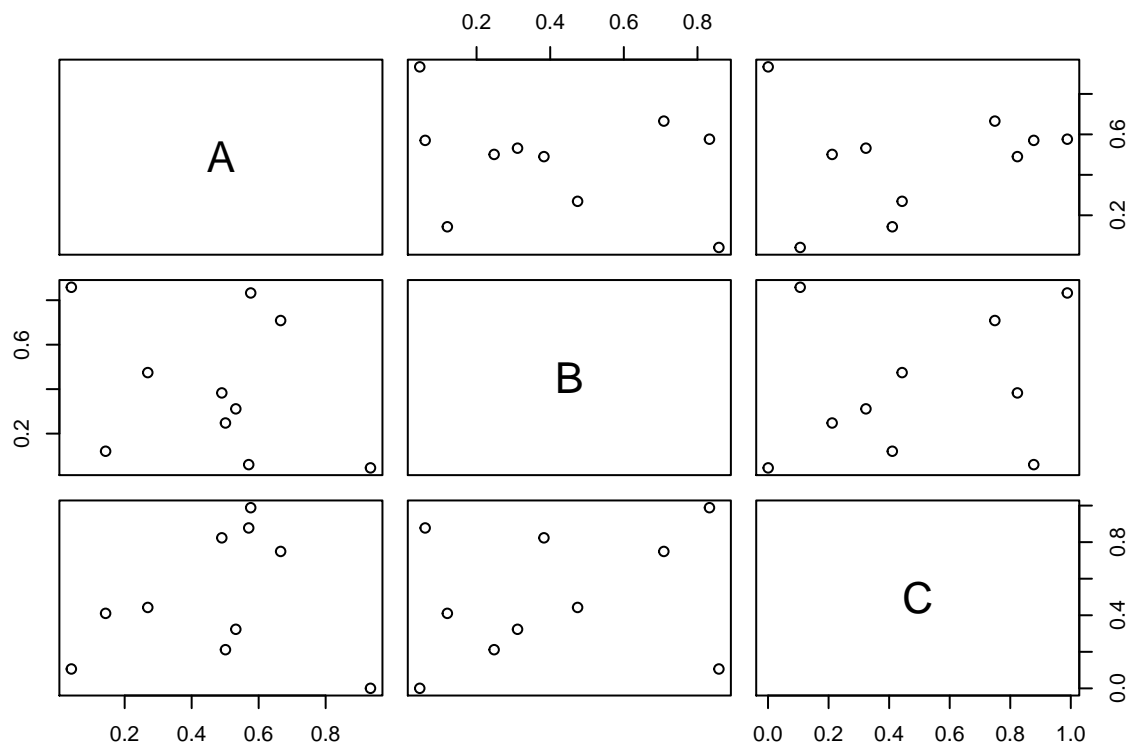
Plot data

```
plot(y[,1], y[,2])
```



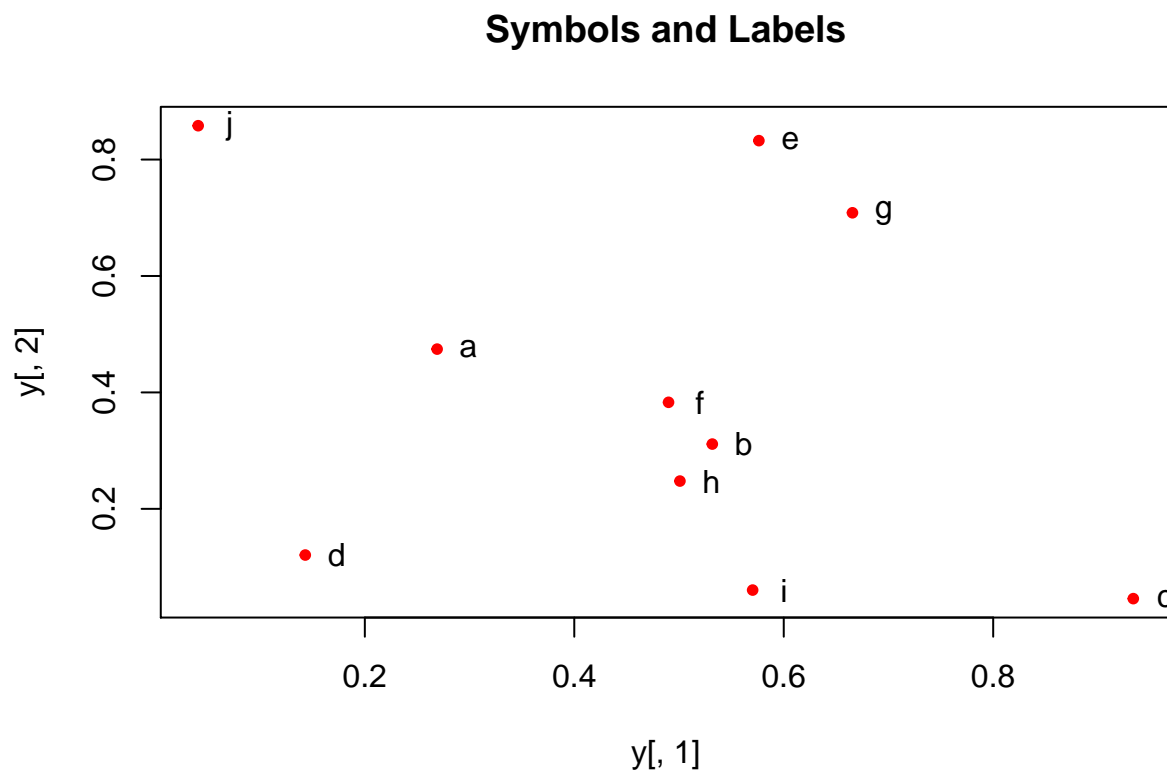
All pairs

```
pairs(y)
```



With labels

```
plot(y[,1], y[,2], pch=20, col="red", main="Symbols and Labels")
text(y[,1]+0.03, y[,2], rownames(y))
```



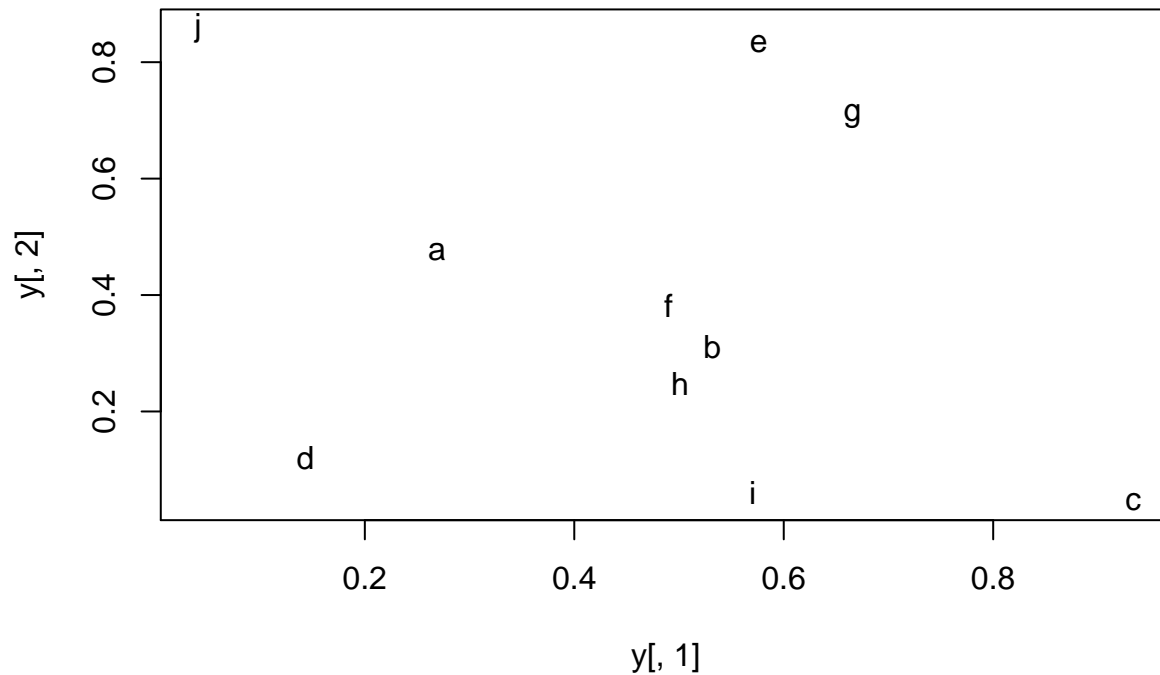
More examples

Print instead of symbols the row names

```
plot(y[,1], y[,2], type="n", main="Plot of Labels")
text(y[,1], y[,2], rownames(y))
```



## Plot of Labels



### Usage of important plotting parameters

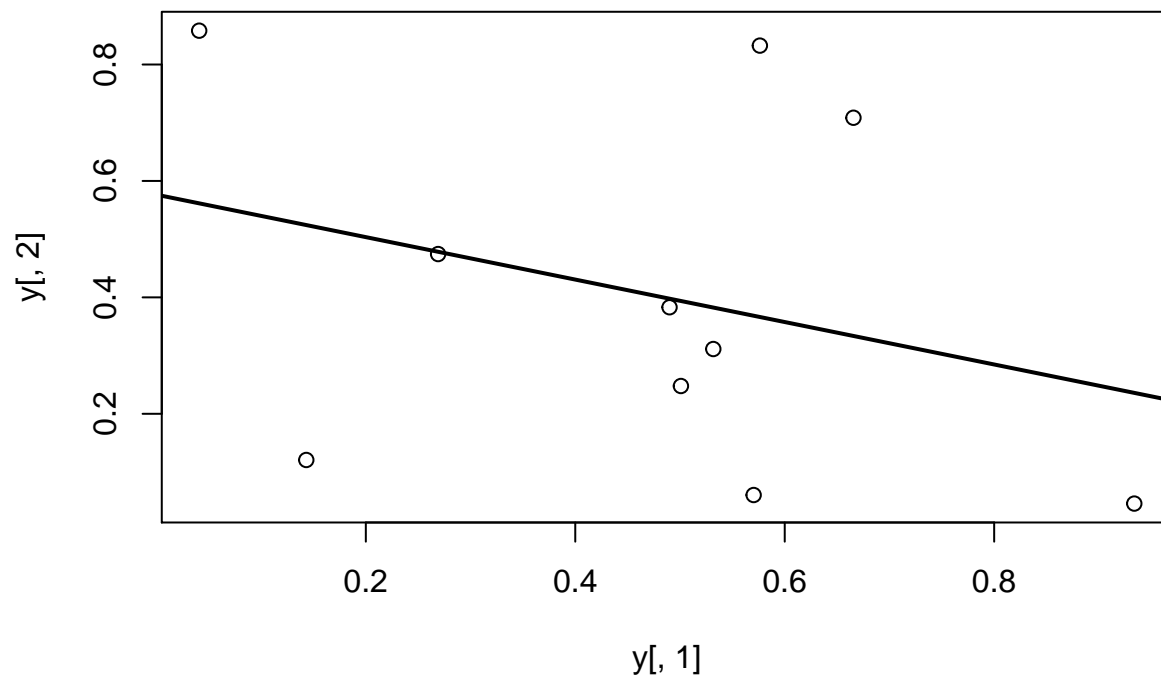
```
grid(5, 5, lwd = 2)
op <- par(mar=c(8,8,8,8), bg="lightblue")
plot(y[,1], y[,2], type="p", col="red", cex.lab=1.2, cex.axis=1.2,
      cex.main=1.2, cex.sub=1, lwd=4, pch=20, xlab="x label",
      ylab="y label", main="My Main", sub="My Sub")
par(op)
```

### \_\_Important arguments\_\_

- `mar`: specifies the margin sizes around the plotting area in order: `c(bottom, left, top, right)`
- `col`: color of symbols
- `pch`: type of symbols, samples: `example(points)`
- `lwd`: size of symbols
- `cex.*`: control font sizes
- For details see `?par`

### Add regression line

```
plot(y[,1], y[,2])
myline <- lm(y[,2]~y[,1]); abline(myline, lwd=2)
```



```
summary(myline)
```

```
##
## Call:
## lm(formula = y[, 2] ~ y[, 1])
##
## Residuals:
```

	Min	1Q	Median	3Q	Max
	-0.40357	-0.17912	-0.04299	0.22147	0.46623

```
##
## Coefficients:
```

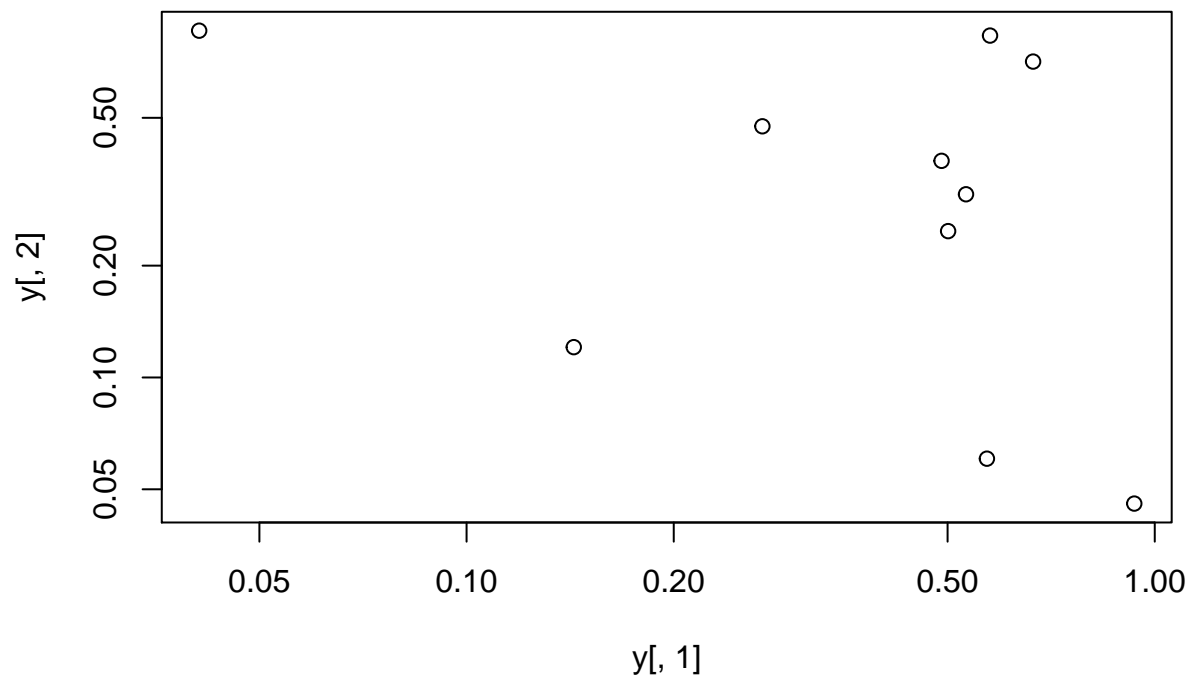
	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	0.5764	0.2110	2.732	0.0258 *
y[, 1]	-0.3647	0.3959	-0.921	0.3839

```
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.3095 on 8 degrees of freedom
## Multiple R-squared:  0.09589,    Adjusted R-squared:  -0.01712
## F-statistic: 0.8485 on 1 and 8 DF,  p-value: 0.3839
```

## Log scale

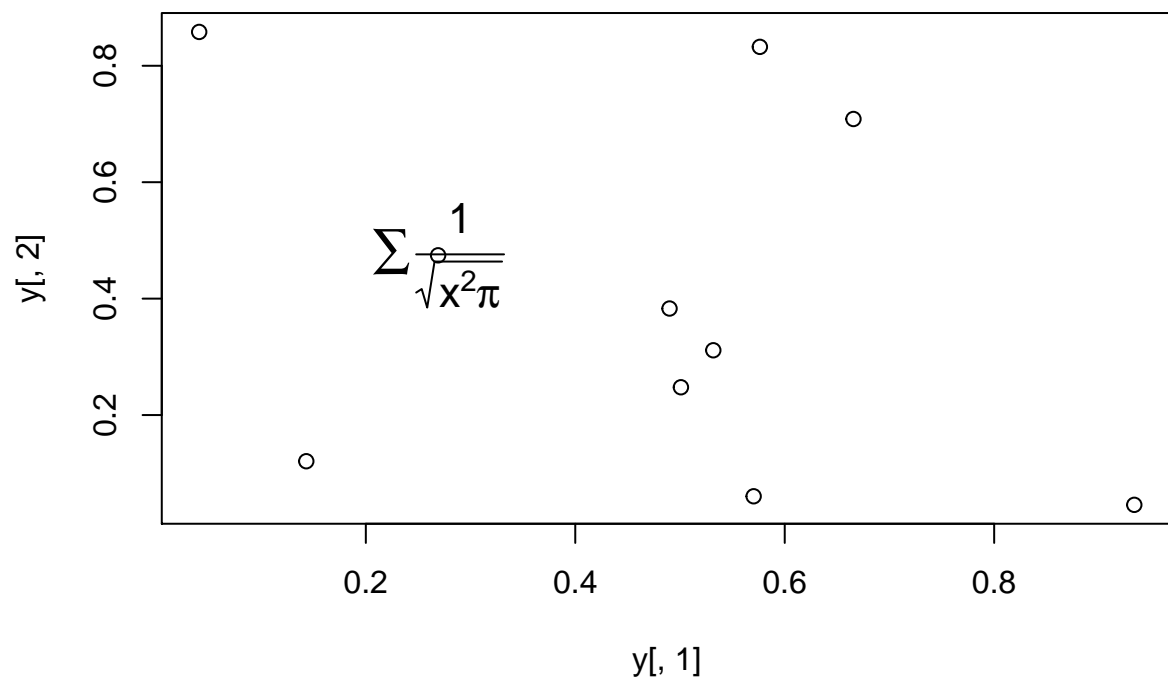
Same plot as above, but on log scale

```
plot(y[,1], y[,2], log="xy")
```



Add a mathematical expression

```
plot(y[,1], y[,2]); text(y[1,1], y[1,2], expression(sum(frac(1,sqrt(x^2*pi)))), cex=1.3)
```



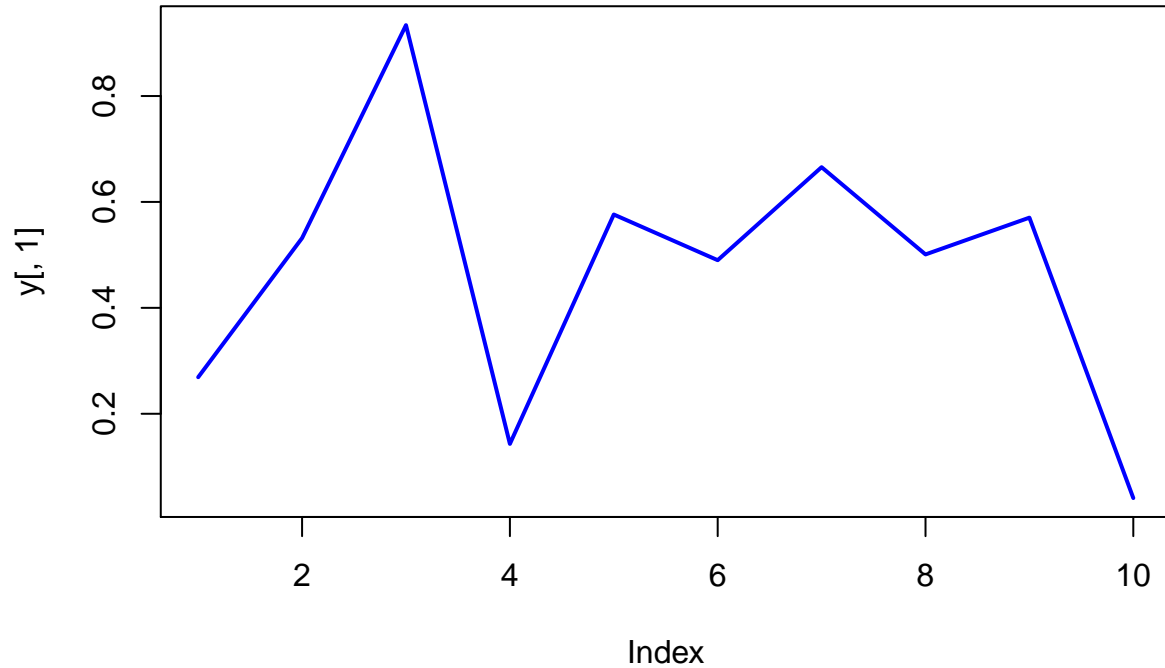
## Homework 3B

Homework 3B: Scatter Plots

## Line Plots

Single data set

```
plot(y[,1], type="l", lwd=2, col="blue")
```



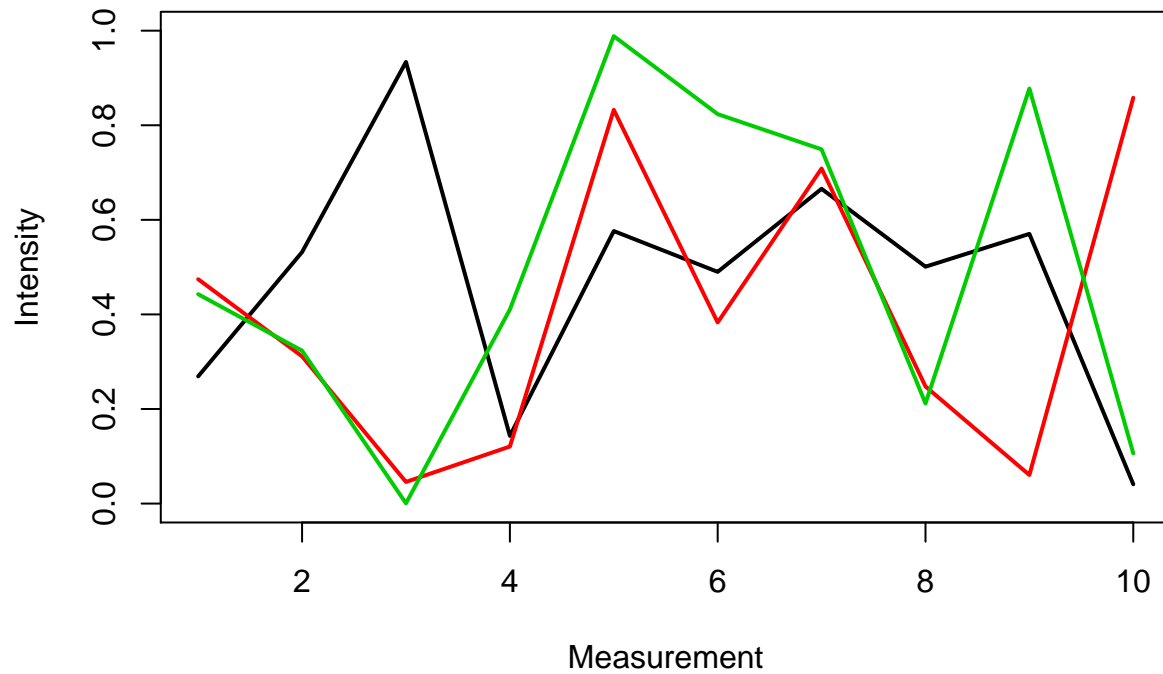
## Many Data Sets

Plots line graph for all columns in data frame `y`. The `split.screen` function is used in this example in a for loop to overlay several line graphs in the same plot.

```
split.screen(c(1,1))
```

```
## [1] 1
```

```
plot(y[,1], ylim=c(0,1), xlab="Measurement", ylab="Intensity", type="l", lwd=2, col=1)
for(i in 2:length(y[,1])) {
  screen(1, new=FALSE)
  plot(y[,i], ylim=c(0,1), type="l", lwd=2, col=i, xaxt="n", yaxt="n", ylab="", xlab="", main="", bty="n")
}
```

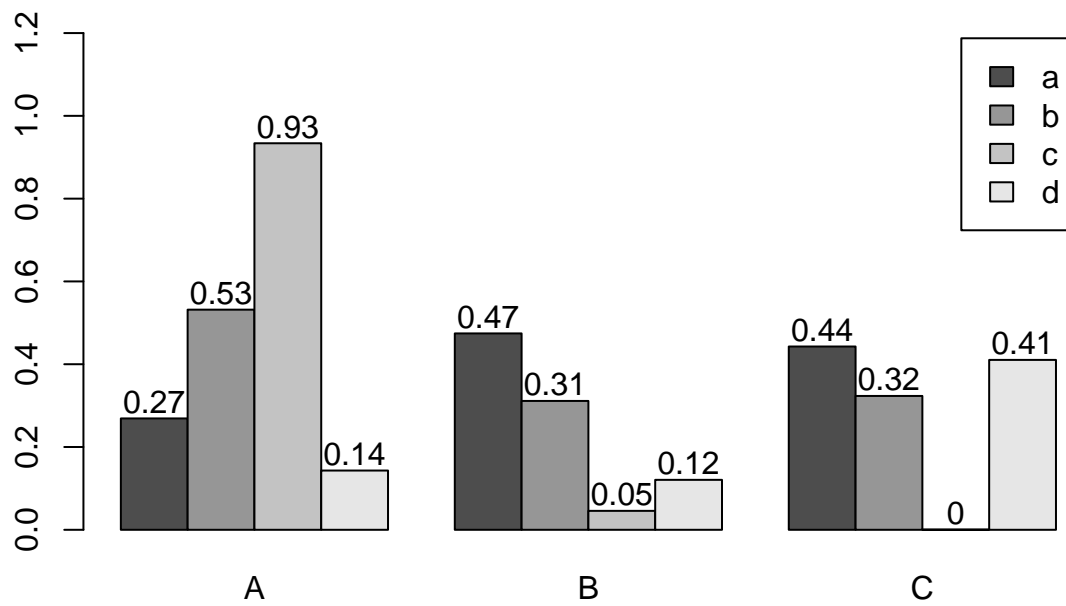


```
close.screen(all=TRUE)
```

## Bar Plots

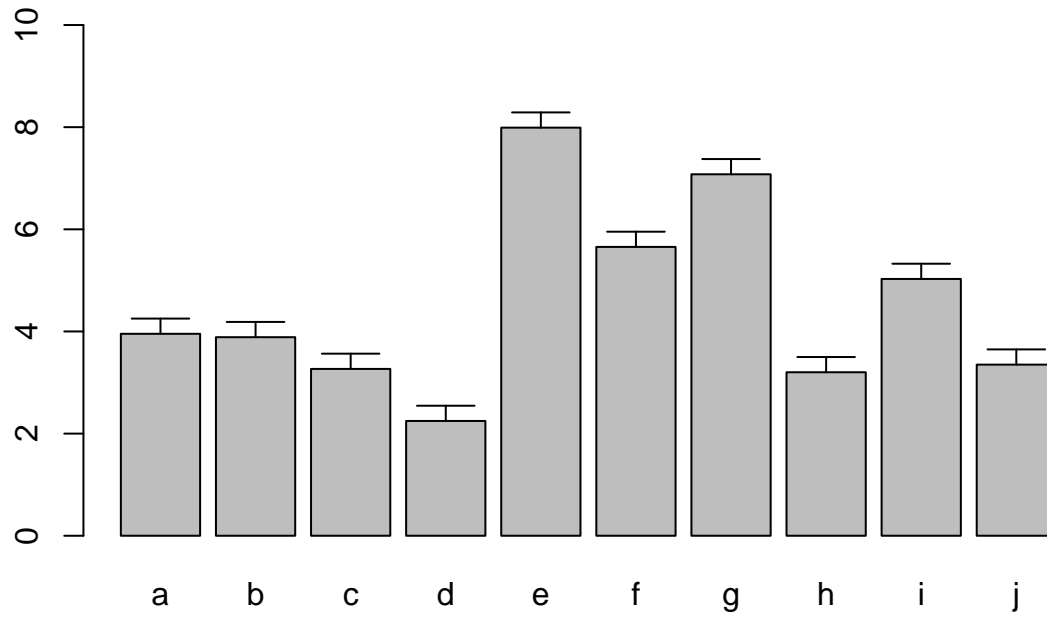
### Basics

```
barplot(y[1:4,], ylim=c(0, max(y[1:4,])+0.3), beside=TRUE, legend=letters[1:4])
text(labels=round(as.vector(as.matrix(y[1:4,])),2), x=seq(1.5, 13, by=1) + sort(rep(c(0,1,2), 4)), y=as
```



## Error Bars

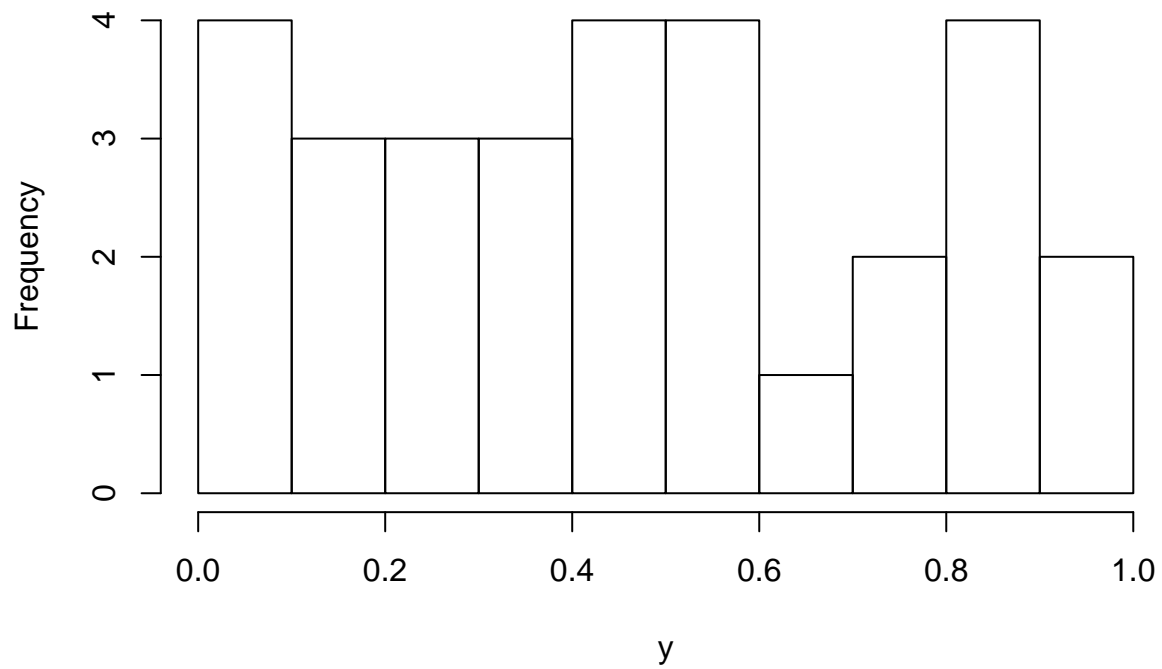
```
bar <- barplot(m <- rowMeans(y) * 10, ylim=c(0, 10))  
stdev <- sd(t(y))  
arrows(bar, m, bar, m + stdev, length=0.15, angle = 90)
```



## Histograms

```
hist(y, freq=TRUE, breaks=10)
```

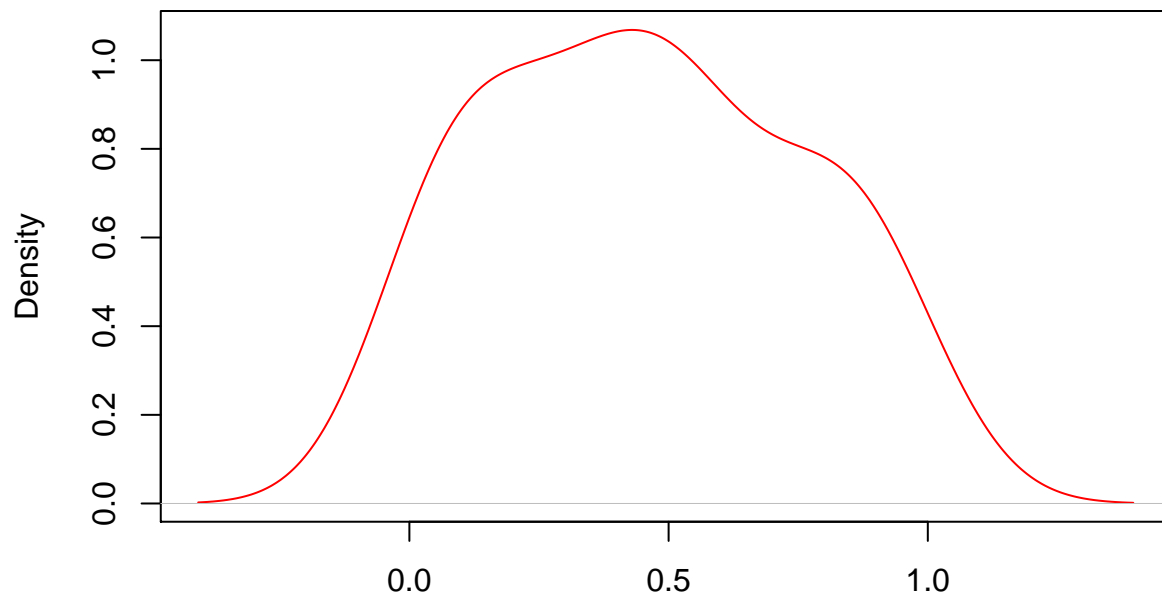
**Histogram of y**



**Density Plots**

```
plot(density(y), col="red")
```

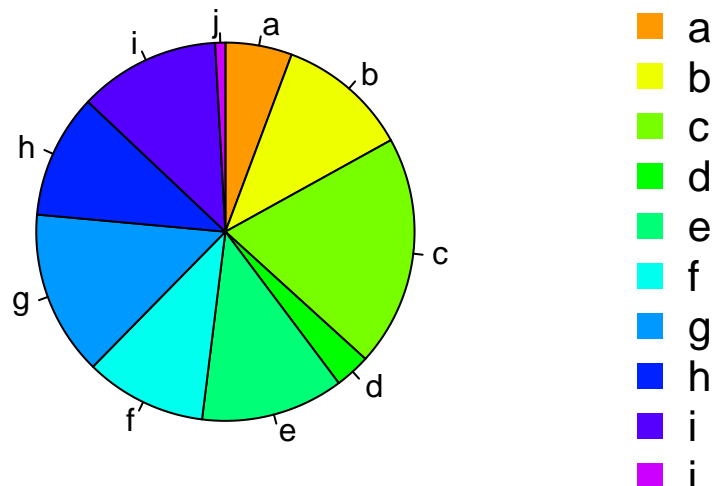
**density.default(x = y)**



N = 30 Bandwidth = 0.136

## Pie Charts

```
pie(y[,1], col=rainbow(length(y[,1]), start=0.1, end=0.8), clockwise=TRUE)
legend("topright", legend=row.names(y), cex=1.3, bty="n", pch=15, pt.cex=1.8,
col=rainbow(length(y[,1]), start=0.1, end=0.8), ncol=1)
```



## Color Selection Utilities

Default color palette and how to change it

```
palette()
```

```
## [1] "black" "red" "green3" "blue" "cyan" "magenta" "yellow" "gray"
```

```
palette(rainbow(5, start=0.1, end=0.2))
```

```
palette()
```

```
## [1] "#FF9900" "#FFBF00" "#FFE600" "#F2FF00" "#CCFF00"
```

```
palette("default")
```

The `gray` function allows to select any type of gray shades by providing values from 0 to 1

```
gray(seq(0.1, 1, by= 0.2))
```

```
## [1] "#1A1A1A" "#4D4D4D" "#808080" "#B3B3B3" "#E6E6E6"
```

Color gradients with `colorpanel` function from `gplots` library

```
library(gplots)
```

```
colorpanel(5, "darkblue", "yellow", "white")
```

```
## [1] "#00008B" "#808046" "#FFFF00" "#FFFF80" "#FFFFFF"
```

Much more on colors in R see Earl Glynn's color chart [here](#)

## Saving Graphics to File

After the `pdf()` command all graphs are redirected to file `test.pdf`. Works for all common formats similarly: jpeg, png, ps, tiff, ...



```
pdf("test.pdf")
plot(1:10, 1:10)
dev.off()
```

Generates Scalable Vector Graphics (SVG) files that can be edited in vector graphics programs, such as Inkscape.

```
library("RSvgDevice")
devSVG("test.svg")
plot(1:10, 1:10)
dev.off()
```

## Homework 3C

Homework 3C: Bar Plots

## Analysis Routine

### Overview

The following exercise introduces a variety of useful data analysis utilities in R.

### Analysis Routine: Data Import

- **Step 1:** To get started with this exercise, direct your R session to a dedicated workshop directory and download into this directory the following sample tables. Then import the files into Excel and save them as tab delimited text files.
  - MolecularWeight\_tair7.xls
  - TargetP\_analysis\_tair7.xls

#### Import the tables into R

Import molecular weight table

```
my_mw <- read.delim(file="MolecularWeight_tair7.xls", header=T, sep="\t")
my_mw[1:2,]
```

Import subcellular targeting table

```
my_target <- read.delim(file="TargetP_analysis_tair7.xls", header=T, sep="\t")
my_target[1:2,]
```

Online import of molecular weight table

```
my_mw <- read.delim(file="http://faculty.ucr.edu/~tgirke/Documents/R_BioCond/Samples/MolecularWeight_ta",
my_mw[1:2,]
```

```
## Sequence.id Molecular.Weight.Da. Residues
## 1 AT1G08520.1 83285 760
## 2 AT1G08530.1 27015 257
```

Online import of subcellular targeting table

```
my_target <- read.delim(file="http://faculty.ucr.edu/~tgirke/Documents/R_BioCond/Samples/TargetP_analysis")
my_target[1:2,]
```

```
##      GeneName Loc   cTP   mTP   SP other
## 1 AT1G08520.1   C 0.822 0.137 0.029 0.039
## 2 AT1G08530.1   C 0.817 0.058 0.010 0.100
```

## Merging Data Frames

- **Step 2:** Assign uniform gene ID column titles

```
colnames(my_target)[1] <- "ID"
colnames(my_mw)[1] <- "ID"
```

- **Step 3:** Merge the two tables based on common ID field

```
my_mw_target <- merge(my_mw, my_target, by.x="ID", by.y="ID", all.x=T)
```

- **Step 4:** Shorten one table before the merge and then remove the non-matching rows (NAs) in the merged file

```
my_mw_target2a <- merge(my_mw, my_target[1:40,], by.x="ID", by.y="ID", all.x=T) # To remove non-matching rows
my_mw_target2 <- na.omit(my_mw_target2a) # Removes rows containing "NAs" (non-matching rows).
```

- **Homework 3D:** How can the merge function in the previous step be executed so that only the common rows among the two data frames are returned? Prove that both methods - the two step version with `na.omit` and your method - return identical results.
- **Homework 3E:** Replace all NAs in the data frame `my_mw_target2a` with zeros.

## Filtering Data

- **Step 5:** Retrieve all records with a value of greater than 100,000 in 'MW' column and 'C' value in 'Loc' column (targeted to chloroplast).

```
query <- my_mw_target[my_mw_target[, 2] > 100000 & my_mw_target[, 4] == "C", ]
query[1:4, ]
```

```
##      ID Molecular.Weight.Da. Residues   Loc   cTP   mTP   SP other
## NA      <NA>                NA      NA <NA>    NA    NA    NA    NA
## NA.1      <NA>                NA      NA <NA>    NA    NA    NA    NA
## NA.2      <NA>                NA      NA <NA>    NA    NA    NA    NA
## 219 AT1G02730.1            132588    1181   C 0.972 0.038 0.008 0.045
```

```
dim(query)
```

```
## [1] 1092    8
```

- **Homework 3F:** How many protein entries in the `my_mw_target` data frame have a MW of greater than 4,000 and less than 5,000. Subset the data frame accordingly and sort it by MW to check that your result is correct.

## String Substitutions

- **Step 6:** Use a regular expression in a substitute function to generate a separate ID column that lacks the gene model extensions. `<>=`

```
my_mw_target3 <- data.frame(loci=gsub("\\\\.*", "", as.character(my_mw_target[,1]), perl = TRUE), my_mw_target3[1:3,1:8])
```

```
##      loci      ID Molecular.Weight.Da. Residues  Loc cTP mTP SP
## 1 AT1G01010 AT1G01010.1      49426      429 <NA>  NA  NA NA
## 2 AT1G01020 AT1G01020.1      28092      245 <NA>  NA  NA NA
## 3 AT1G01020 AT1G01020.2      21711      191 <NA>  NA  NA NA
```

- **Homework 3G:** Retrieve those rows in `my_mw_target3` where the second column contains the following identifiers: `c("AT5G52930.1", "AT4G18950.1", "AT1G15385.1", "AT4G36500.1", "AT1G67530.1")`. Use the `%in%` function for this query. As an alternative approach, assign the second column to the row index of the data frame and then perform the same query again using the row index. Explain the difference of the two methods.

## Calculations on Data Frames

- **Step 7:** Count the number of duplicates in the `loci` column with the `table` function and append the result to the data frame with the `cbind` function.

```
mycounts <- table(my_mw_target3[,1])[my_mw_target3[,1]]
my_mw_target4 <- cbind(my_mw_target3, Freq=mycounts[as.character(my_mw_target3[,1])])
```

- **Step 8:** Perform a vectorized division of columns 3 and 4 (average AA weight per protein)

```
data.frame(my_mw_target4, avg_AA_WT=(my_mw_target4[,3] / my_mw_target4[,4]))[1:2,5:11]
```

```
##      Loc cTP mTP SP other Freq.Var1 Freq.Freq
## 1 <NA>  NA  NA NA      NA AT1G01010      1
## 2 <NA>  NA  NA NA      NA AT1G01020      2
```

- **Step 9:** Calculate for each row the mean and standard deviation across several columns

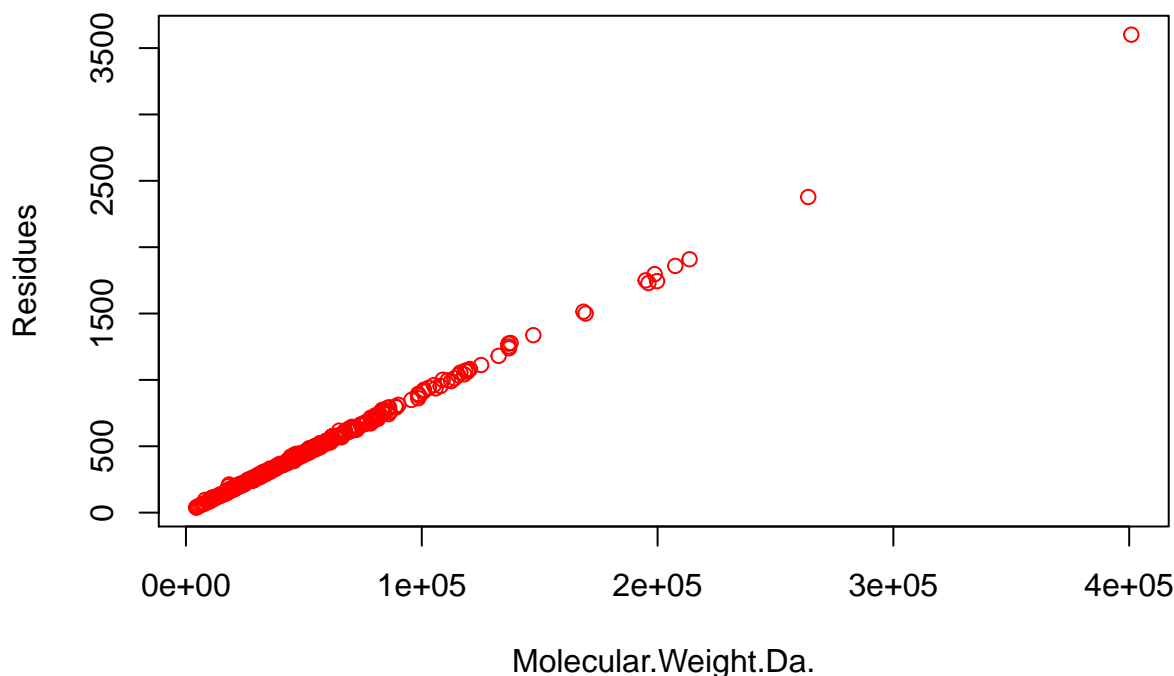
```
mymean <- apply(my_mw_target4[,6:9], 1, mean)
mystdev <- apply(my_mw_target4[,6:9], 1, sd, na.rm=TRUE)
data.frame(my_mw_target4, mean=mymean, stdev=mystdev)[1:2,5:12]
```

```
##      Loc cTP mTP SP other Freq.Var1 Freq.Freq mean
## 1 <NA>  NA  NA NA      NA AT1G01010      1  NA
## 2 <NA>  NA  NA NA      NA AT1G01020      2  NA
```

## Plotting Example

- **Step 10:** Generate scatter plot columns: 'MW' and 'Residues'

```
plot(my_mw_target4[1:500,3:4], col="red")
```



## Export Results and Run Entire Exercise as Script

- **Step 11:** Write the data frame `my_mw_target4` into a tab-delimited text file and inspect it in Excel.

```
write.table(my_mw_target4, file="my_file.xls", quote=F, sep="\t", col.names = NA)
```

- **Homework 3H:** Write all commands from this exercise into an R script named `exerciseRbasics.R`, or download it from [here](#). Then execute the script with the `source` function like this: `source("exerciseRbasics.R")`. This will run all commands of this exercise and generate the corresponding output files in the current working directory.

```
source("exerciseRbasics.R")
```

## R Markdown

### Overview

R Markdown combines markdown (an easy to write plain text format) with embedded R code chunks. When compiling R Markdown documents, the code components can be evaluated so that both the code and its output can be included in the final document. This makes analysis reports highly reproducible by allowing to automatically regenerate them when the underlying R code or data changes. R Markdown documents (`.Rmd` files) can be rendered to various formats including HTML and PDF. The R code in an `.Rmd` document is processed by `knitr`, while the resulting `.md` file is rendered by `pandoc` to the final output formats (*e.g.* HTML or PDF). Historically, R Markdown is an extension of the older `Sweave/Latex` environment. Rendering of mathematical expressions and reference management is also supported by R Markdown using embedded LaTeX syntax and Bibtex, respectively.

## Quick Start

### Install R Markdown

```
install.packages("rmarkdown")
```

### Initialize a new R Markdown (Rmd) script

To minimize typing, it can be helpful to start with an R Markdown template and then modify it as needed. Note the file name of an R Markdown script needs to have the extension `.Rmd`. Template files for the following examples are available here:

- R Markdown sample script: `sample.Rmd`
- Bibtex file for handling citations and reference section: `bibtex.bib`

Users want to download these files, open the `sample.Rmd` file with their preferred R IDE (*e.g.* RStudio, vim or emacs), initialize an R session and then direct their R session to the location of these two files.

### Metadata section

The metadata section (YAML header) in an R Markdown script defines how it will be processed and rendered. The metadata section also includes both title, author, and date information as well as options for customizing the output format. For instance, PDF and HTML output can be defined with `pdf_document` and `html_document`, respectively. The `BiocStyle::` prefix will use the formatting style of the `BiocStyle` package from Bioconductor.

```
---
title: "My First R Markdown Document"
author: "Author: First Last"
date: "Last update: 13 April, 2018"
output:
  BiocStyle::html_document:
    toc: true
    toc_depth: 3
    fig_caption: yes

fontsize: 14pt
bibliography: bibtex.bib
---
```

### Render Rmd script

An R Markdown script can be evaluated and rendered with the following `render` command or by pressing the `knit` button in RStudio. The `output_format` argument defines the format of the output (*e.g.* `html_document`). The setting `output_format="all"` will generate all supported output formats. Alternatively, one can specify several output formats in the metadata section as shown in the above example.

```
rmarkdown::render("sample.Rmd", clean=TRUE, output_format="html_document")
```

The following shows two options how to run the rendering from the command-line.

```
$ echo "rmarkdown::render('sample.Rmd', clean=TRUE)" | R --slave
$ Rscript -e "rmarkdown::render('sample.Rmd', clean=TRUE)"
```

Alternatively, one can use a Makefile to evaluate and render an R Markdown script. A sample Makefile for rendering the above `sample.Rmd` can be downloaded [here](#). To apply it to a custom `Rmd` file, one needs open the Makefile in a text editor and change the value assigned to `MAIN` (line 13) to the base name of the corresponding `.Rmd` file (*e.g.* assign `systemPipeRNAseq` if the file name is `systemPipeRNAseq.Rmd`). To execute the Makefile, run the following command from the command-line.

```
$ make -B
```

## R code chunks

R Code Chunks can be embedded in an R Markdown script by using three backticks at the beginning of a new line along with arguments enclosed in curly braces controlling the behavior of the code. The following lines contain the plain R code. A code chunk is terminated by a new line starting with three backticks. The following shows an example of such a code chunk. Note the backslashes are not part of it. They have been added to print the code chunk syntax in this document.

```
```\{r code_chunk_name, eval=FALSE\}  
x <- 1:10  
```
```

The following lists the most important arguments to control the behavior of R code chunks:

- `r`: specifies language for code chunk, here R
- `code_chunk_name`: name of code chunk; this name needs to be unique
- `eval`: if assigned `TRUE` the code will be evaluated
- `warning`: if assigned `FALSE` warnings will not be shown
- `message`: if assigned `FALSE` messages will not be shown
- `cache`: if assigned `TRUE` results will be cached to reuse in future rendering instances
- `fig.height`: allows to specify height of figures in inches
- `fig.width`: allows to specify width of figures in inches

For more details on code chunk options see [here](#).

## Learning Markdown

The basic syntax of Markdown and derivatives like `kramdown` is extremely easy to learn. Rather than providing another introduction on this topic, here are some useful sites for learning Markdown:

- [Markdown Intro on GitHub](#)
- [Markdown Cheet Sheet](#)
- [Markdown Basics from RStudio](#)
- [R Markdown Cheat Sheet](#)
- [kramdown Syntax](#)

## Tables

There are several ways to render tables. First, they can be printed within the R code chunks. Second, much nicer formatted tables can be generated with the functions `kable`, `pander` or `xtable`. The following example uses `kable` from the `knitr` package.

```
library(knitr)  
kable(iris[1:12,])
```

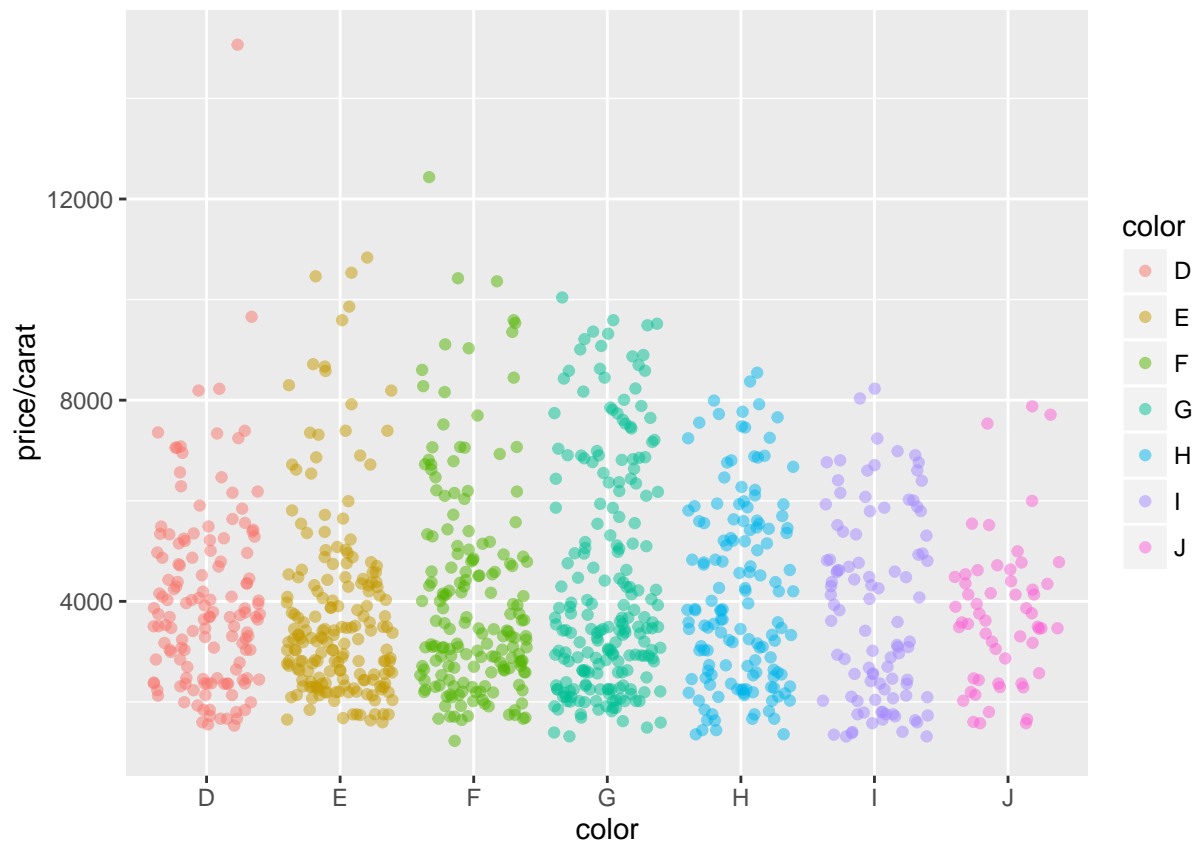
| Sepal.Length | Sepal.Width | Petal.Length | Petal.Width | Species |
|--------------|-------------|--------------|-------------|---------|
| 5.1          | 3.5         | 1.4          | 0.2         | setosa  |

| Sepal.Length | Sepal.Width | Petal.Length | Petal.Width | Species |
|--------------|-------------|--------------|-------------|---------|
| 4.9          | 3.0         | 1.4          | 0.2         | setosa  |
| 4.7          | 3.2         | 1.3          | 0.2         | setosa  |
| 4.6          | 3.1         | 1.5          | 0.2         | setosa  |
| 5.0          | 3.6         | 1.4          | 0.2         | setosa  |
| 5.4          | 3.9         | 1.7          | 0.4         | setosa  |
| 4.6          | 3.4         | 1.4          | 0.3         | setosa  |
| 5.0          | 3.4         | 1.5          | 0.2         | setosa  |
| 4.4          | 2.9         | 1.4          | 0.2         | setosa  |
| 4.9          | 3.1         | 1.5          | 0.1         | setosa  |
| 5.4          | 3.7         | 1.5          | 0.2         | setosa  |
| 4.8          | 3.4         | 1.6          | 0.2         | setosa  |

## Figures

Plots generated by the R code chunks in an R Markdown document can be automatically inserted in the output file. The size of the figure can be controlled with the `fig.height` and `fig.width` arguments.

```
library(ggplot2)
dsmall <- diamonds[sample(nrow(diamonds), 1000), ]
ggplot(dsmall, aes(color, price/carat)) + geom_jitter(alpha = I(1 / 2), aes(color=color))
```



Sometimes it can be useful to explicitly write an image to a file and then insert that image into the final document by referencing its file name in the R Markdown source. For instance, this can be useful for time consuming analyses. The following code will generate a file named `myplot.png`. To insert the file in the final document, one can use standard Markdown or HTML syntax, *e.g.*: ``.

```
png("myplot.png")
ggplot(dsmall, aes(color, price/carat)) + geom_jitter(alpha = I(1 / 2), aes(color=color))
dev.off()

## pdf
## 2
```

## Inline R code

To evaluate R code inline, one can enclose an R expression with a single back-tick followed by `r` and then the actual expression. For instance, the back-ticked version of `'r 1 + 1'` evaluates to 2 and `'r pi'` evaluates to 3.1415927.

## Mathematical equations

To render mathematical equations, one can use standard Latex syntax. When expressions are enclosed with single `$` signs then they will be shown inline, while enclosing them with double `$$` signs will show them in display mode. For instance, the following Latex syntax `d(X,Y) = \sqrt{\sum_{i=1}^n (x_i - y_i)^2}` renders in display mode as follows:

$$d(X, Y) = \sqrt{\sum_{i=1}^n (x_i - y_i)^2}$$

## Citations and bibliographies

Citations and bibliographies can be autogenerated in R Markdown in a similar way as in Latex/Bibtex. Reference collections should be stored in a separate file in Bibtex or other supported formats. To cite a publication in an R Markdown script, one uses the syntax `[@<id1>]` where `<id1>` needs to be replaced with a reference identifier present in the Bibtex database listed in the metadata section of the R Markdown script (*e.g.* `bibtex.bib`). For instance, to cite Lawrence et al. (2013), one uses its reference identifier (*e.g.* `Lawrence2013-kt`) as `<id1>` (Lawrence et al. 2013). This will place the citation inline in the text and add the corresponding reference to a reference list at the end of the output document. For the latter a special section called **References** needs to be specified at the end of the R Markdown script. To fine control the formatting of citations and reference lists, users want to consult this the corresponding R Markdown page. Also, for general reference management and outputting references in Bibtex format Paperpile can be very helpful.

## Viewing R Markdown report on HPCC cluster

R Markdown reports located on UCR's HPCC Cluster can be viewed locally in a web browser (without moving the source HTML) by creating a symbolic link from a user's `.html` directory. This way any updates to the report can be viewed immediately without creating another copy of the HTML file. For instance, if user `ttest` has generated an R Markdown report under `~/bigdata/today/rmarkdown/sample.html`, then the proper symbolic link to this file can be created as follows:

```
cd ~/.html
ln -s ~/bigdata/today/rmarkdown/sample.html sample.html
```

After this one can view the report in a web browser using this URL <http://biocluster.ucr.edu/~ttest/rmarkdown/sample.html>. If necessary access to the URL can be restricted with a password following the instructions here.



# Shiny Web Apps

## What is Shiny?

Shiny is an R-based environment for building interactive web applications for data analysis and exploration. Since most JavaScript code is autogenerated by the environment, basic R knowledge is sufficient for developing Shiny apps. They can be deployed on local computers or web servers including custom and cloud-based servers (e.g. AWS, GCP, shinyapp.io service). The basic structure of a Shiny app is an `app.R` script containing the following components:

1. User interface

```
ui <- fluidPage()
```

2. Server function

```
server <- function(input, output) {}
```

3. Statement to run shiny app

```
shinyApp(ui = ui, server = server)
```

Alternatively, the `ui` and `server` functions can be organized in two script files, a `ui.R` and a `server.R` script, respectively.

## Develop and test Shiny app locally

Open R and set session to parent directory (here `myappdir`) containing shiny script `app.R`, and then run it with the `runApp()` function. A sample `app.R` script for testing can be downloaded from [here](#).

```
library(shiny)
runApp("myappdir") # To show code in app, add argument: display.mode="showcase"
```

This will open the app in a web browser.

## Deploy on web server

This can be done on local or cloud systems. An easy solution is to get an account on shinyapps.io and then deploy Shiny apps there. For details, see [here](#).

```
setwd("myappdir")
library(rsconnect)
deployApp()
```

## Example Shiny app

The following Shiny app is hosted on [shinyapps.io](#) and embedded into the markdown (or html) source of this page using the following iframe syntax:

```
<iframe src="https://tgirke.shinyapps.io/diamonds/" style="border: none; width: 880px; height: 900px"></iframe>
```

## Learning Shiny

The Shiny section on the Rstudio site contains excellent tutorials. In addition, users may want to explore the example apps included in the `shiny` package. This can be done by loading the individual examples (see [here](#)) or saving the code to a user writable directory like this:

```
mydir <- system.file("examples", package="shiny")
dir.create('my_shiny_test_dir')
file.copy(mydir, "my_shiny_test_dir", recursive=TRUE)
setwd("my_shiny_test_dir/examples")
runApp("01_hello") # Runs first example app in directory
dir() # Lists available Shiny examples (directories).
```

## Session Info

```
sessionInfo()
```

```
## R version 3.3.3 (2017-03-06)
## Platform: x86_64-pc-linux-gnu (64-bit)
## Running under: Ubuntu 14.04.5 LTS
##
## locale:
##  [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C              LC_TIME=en_US.UTF-8
##  [4] LC_COLLATE=en_US.UTF-8    LC_MONETARY=en_US.UTF-8   LC_MESSAGES=en_US.UTF-8
##  [7] LC_PAPER=en_US.UTF-8      LC_NAME=C                 LC_ADDRESS=C
## [10] LC_TELEPHONE=C           LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C
##
## attached base packages:
## [1] methods      stats      graphics  utils      datasets  grDevices  base
##
## other attached packages:
## [1] knitr_1.14      plots_3.0.1    RSQLite_1.0.0  DBI_0.5-1    ggplot2_2.1.0  limma_3.30.0
## [7] BiocStyle_2.2.0
##
## loaded via a namespace (and not attached):
##  [1] Rcpp_0.12.7      magrittr_1.5      munsell_0.4.3     colorspace_1.2-7  highr_0.6
##  [6] stringr_1.1.0    plyr_1.8.4        caTools_1.17.1    tools_3.3.3       grid_3.3.3
## [11] gtable_0.2.0     KernSmooth_2.23-15 htmltools_0.3.5    gtools_3.5.0      yaml_2.1.13
## [16] assertthat_0.1   digest_0.6.10     tibble_1.2        formatR_1.4       codetools_0.2-15
## [21] bitops_1.0-6     evaluate_0.10     rmarkdown_1.1     labeling_0.3       gdata_2.17.0
## [26] stringi_1.1.2    scales_0.4.0
```

## References

Lawrence, Michael, Wolfgang Huber, Hervé Pagès, Patrick Aboyoun, Marc Carlson, Robert Gentleman, Martin T Morgan, and Vincent J Carey. 2013. “Software for Computing and Annotating Genomic Ranges.” *PLoS Comput. Biol.* 9 (8): e1003118. doi:10.1371/journal.pcbi.1003118.