Chapter 7
Practical

#### 7.1 Practical 1: Linear Mixed Models with R



- We will illustrate some basic linear mixed models analysis
- We will use the PBC dataset; this is available as the object <a href="pbc2">pbc2</a> in the R workspace you have received
- We will need the following variables
  - \* id: patient id number
  - \* serBilir: serum bilirubin (the response variable of interest)
  - \* year: follow-up times in years
  - \* drug: the randomized treatment
  - \* sex: the gender of the patients
  - \* age: the age of the patients



- The response variable we will use will be the natural logarithm of serBilir
- We start with some descriptive plots; load the lattice package using:
   library("lattice") (or your favorite graphics package, e.g., ggplot2)
- T1: Plot the average longitudinal evolutions of the two treatment groups using loess. Should we or should we not trust this plot?
- T2: Do the same plot for sex



- T3: Create the plot of the subject-specific longitudinal trajectories
  - it will be useful to save the plots in a pdf, using pdf() before executing the plot and dev.off() afterwards
- T4: As an initial analysis we will test for a treatment effect using the AUC
  - ▷ calculate the AUC for each subject (see p. 31)
  - be do a t-test for the difference in the AUC between the two treatment groups



- We will proceed by fitting appropriate linear mixed models to the data
- One approach to graphically investigate the variance function over time is to smooth the squared OLS residuals
  - in order the OLS residuals to correctly reflect the properties of the marginal covariance matrix of the response variable, it is important to remove all systematic trends
  - > hence we want to fit an elaborate mean structure linear model
  - > we will allow for nonlinear time evolutions using natural cubic splines
  - ▷ correct for sex, drug and age + interactions of the time effect with sex and drug



- A bit of motivation and background for splines: When modeling continuous covariates it is customary to assume that such covariates affect linearly the response
- However, this assumption is very restrictive, and in many real applications it may not hold
  - ▷ increasing age from 20y to 25y does not increase the risk in the same amount as increasing age from 60y to 65y
  - > similar conjectures also can be made for the time effect in a longitudinal setting
- Wrongly assuming linearity may affect the resulting inference for such covariates as well as the predictive ability of the model



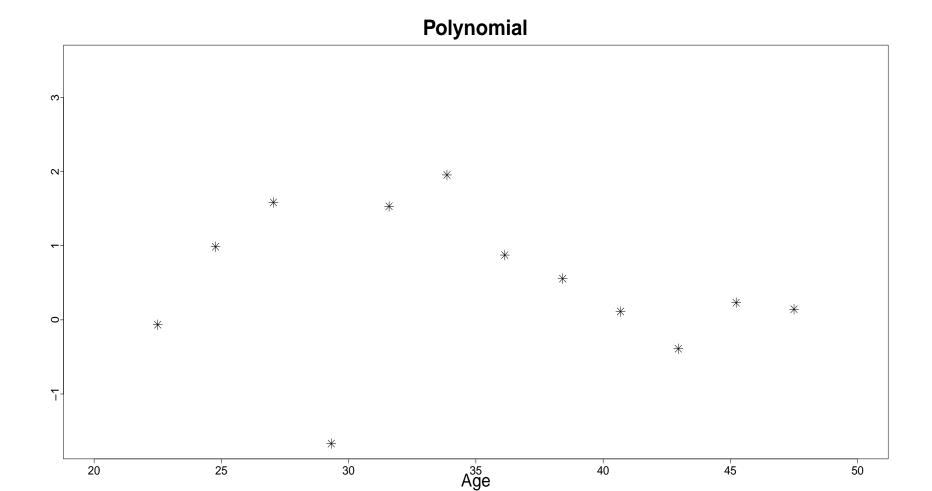
• Therefore, it is highly advisable not to restrict a priori the effects of continuous predictors to be linear and let the data tell you the true story

• The easiest way to relax linearity is to assume polynomial effects

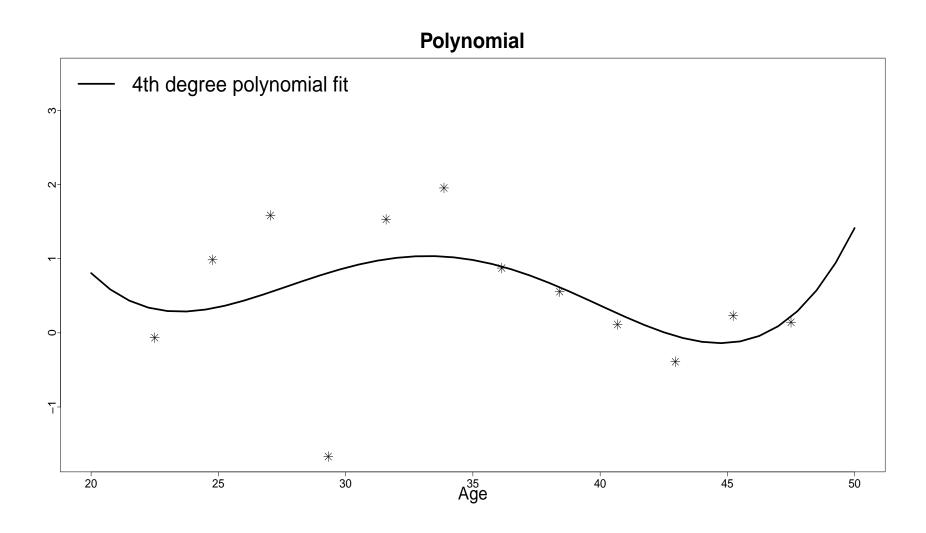
$$\beta_0 + \beta_1 X_i + \beta_2 X_i^2 + \beta_3 X_i^3 + \dots$$

- However, polynomials have some disadvantages, namely
  - $\triangleright$  they are not local  $\Rightarrow$  changing one data point will affect the overall fit
  - > numerically ill-conditioned (however, not too worrisome with modern software)

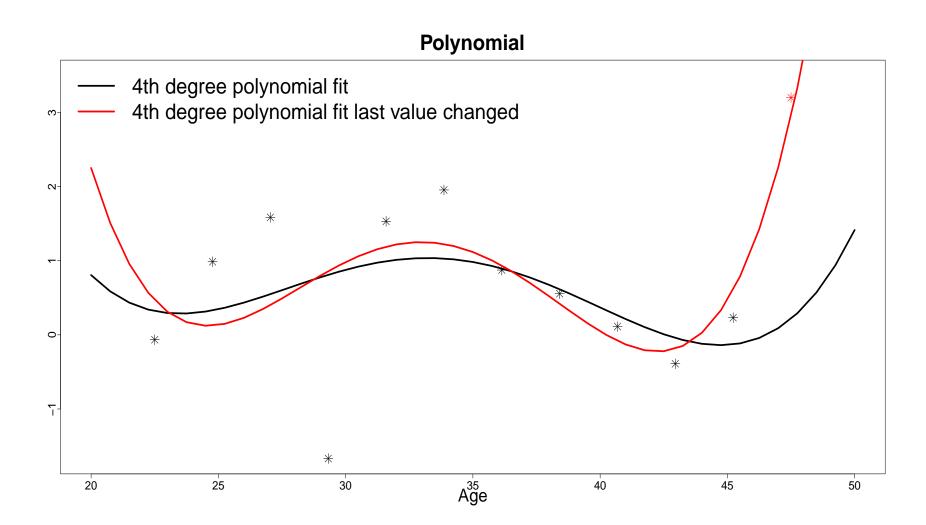










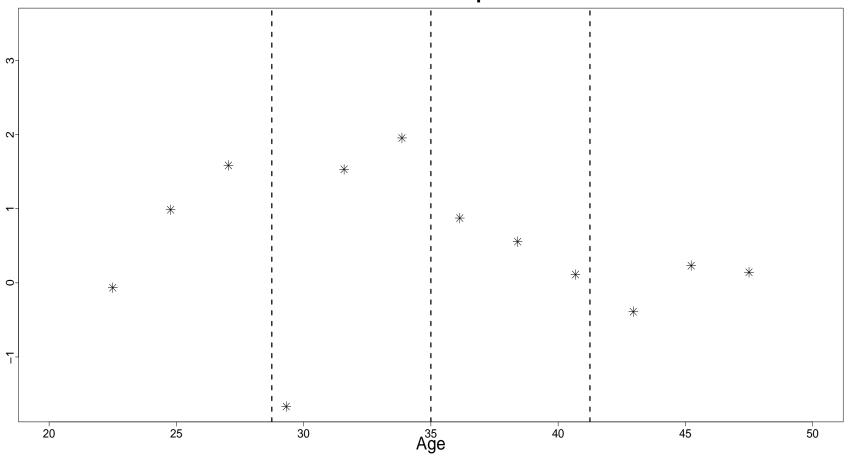




- An alternative approach to relax the linearity assumption of continuous predictors is to use regression splines
- Idea behind regression splines: use polynomials but locally
  - > split the range of values of the continuous predictor into subintervals using a series of knots
  - b within each subinterval assume that the effect of the predictor is nonlinear and can be approximated by a cubic polynomial
  - put extra smoothness assumptions, i.e., the cubic polynomial fits between neighboring subintervals must be connected



#### **Natural Cubic Splines**



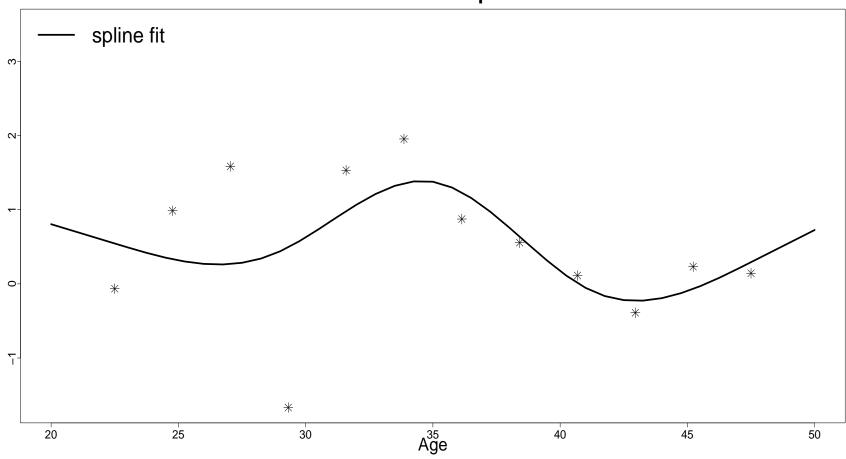


- There are several types of regression splines available
  - ▷ advisable to use natural cubic splines, which assume linearity outside the boundary knots – better statistical properties
- Other approaches (we are not going to discuss them here)
  - ▶ penalized splines
  - ▷ local regression

  - ▷ . . .

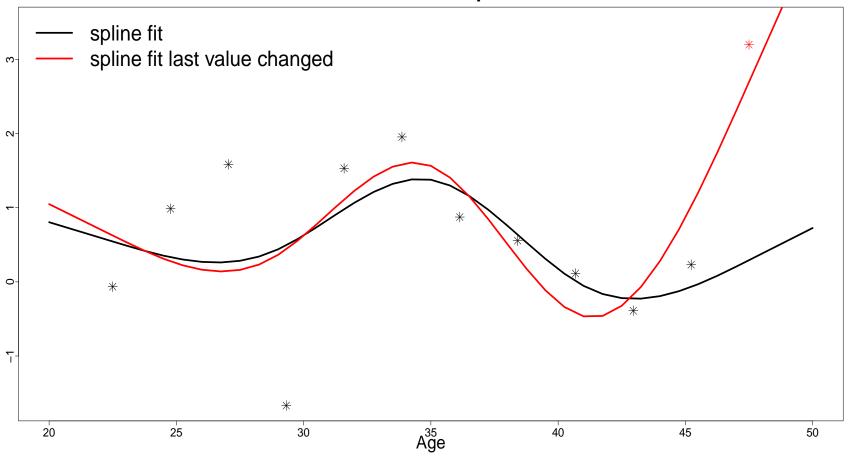


#### **Natural Cubic Splines**





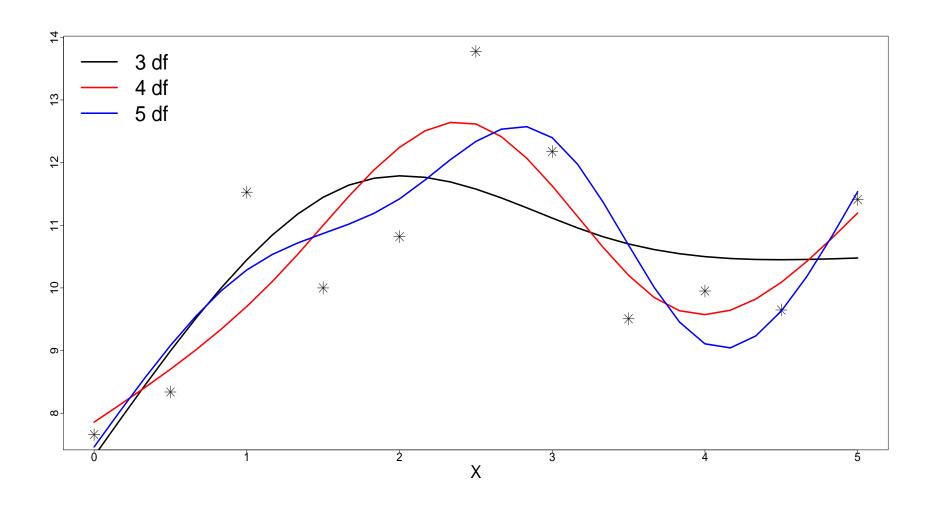
#### **Natural Cubic Splines**





- As also in the case of the polynomials, we can tune the degree of nonlinearity by specifying the degrees of freedom for the spline
  - ▷ increasing the degrees of freedom results in more flexible modeling
  - bias-variance tradeoff







- T5: Calculate the squared OLS residuals for the above defined linear regression model, and do the loess plot
  - ▷ load package splines using library("splines") in order to make the spline functions available
  - be the function that can be used to fit natural cubic splines is ns() and it can be directly included in a model formula
  - b fit the above defined model using function lm()
  - > extract the residuals using function resid()
  - ▷ make the plot of the squared residuals using xyplot() (or your favorite plotting function)



- We will start our model-building exercise. . .
- General recipe: First model the covariance structure and then the mean structure
  - > start with an elaborate mean model (i.e., in order to be more or less certain that we have removed all systematic trends)
  - build up the random-effects structure, starting from random intercepts, random intercepts and random slopes, etc. until you find a satisfying model
  - > then return to the mean structure and simplify it if required



- T6: Fit a linear mixed model with mean structure the same as the one you used in the simple linear model to calculate the OLS residuals in T5, and random intercepts you will need to load package **nlme** first using library("nlme")
- T7: Continue on elaborating the random-effects structure and perform likelihood ratio tests (using function anova()) to see if the additional random effects are required
  - > random intercepts & random slopes
  - > random intercepts & splines for the time effect



- Technical/Theorical Issue: Consider the hypothesis test between the random intercepts and the random intercepts & random slopes models
  - > random intercepts model

$$y_{ij} = X\beta + b_{i0} + \varepsilon_{ij}, \qquad b_{i0} \sim \mathcal{N}(0, \sigma_{b1}^2)$$

$$y_{ij} = X\beta + b_{i0} + b_{i1}t + \varepsilon_{ij}, \qquad b_{i0} \sim \mathcal{N}(0, D)$$

with

$$D = \begin{bmatrix} \sigma_{b1}^2 & \sigma_{b12} \\ \sigma_{b12} & \sigma_{b2}^2 \end{bmatrix}$$



• Hence, the hypotheses to be tested are

$$H_0: \quad \sigma_{b2}^2 = \sigma_{b12} = 0$$

$$H_a: \quad \sigma_{b2}^2 \neq 0 \text{ or } \sigma_{b12} \neq 0$$

- ullet What is the problem? The null hypothesis for  $\sigma^2_{b2}$  is on the boundary of its corresponding parameter space
  - $\triangleright$  statistical tests derived from standard ML theory assume the  $H_0$  is an interior point of the parameter space
  - $\triangleright$  the classical asymptotic  $\chi^2$  distribution for the likelihood ratio test statistic does not apply



- ullet For simple settings (as the one above), it has been proposed to use a mixture of  $\chi^2$  distributions
  - ▷ nonetheless, it has been suggested that this does not always work satisfactorily (e.g., see package **RLRsim** and the references therein)
- ullet Here we will just use the  $\chi^2$  distribution and be a bit conservative
- T8: Continue by relaxing the fixed-effects structure
  - > start be checking if all interaction terms can be dropped using a likelihood ratio test



- Technical/Theorical Issue: By default <a href="mailto:linear">line</a> fits linear mixed models using REML
  - $ightharpoonup \mathsf{REML}$  estimation proceeds by transforming the response variable using the design matrix X
  - $\triangleright$  hence, by comparing linear mixed models with different fixed-effect structures, we are actually comparing models with different response variables  $\Rightarrow$  LRT is not valid in models with different response variables
- T9: Re-fit the mixed model you ended up with in T8 using maximum likelihood instead of REML, and redo the LRT (check argument method of lme())
  - > continue by checking if any main effects may be dropped



• T10: For the final model use function summary() to obtain a detailed output and interpret the results

#### 7.2 Practical 2: Cox Models



• We will perform some basic survival analysis calculations and fit a series of Cox models for the AIDS dataset

• Start R and load package **survival**, using **library**("survival")

• Load the R workspace with the AIDS dataset



- We will need the following variables
  - \* Time: observed event times in years
  - \* death: the death indicator
  - \* drug: the randomized treatment
  - \* gender: the sex of the patients
  - \* AZT: intolerance or failure
  - \* CD4: the square root CD4 cell count at baseline
- T1: Calculate and plot the Kaplan-Meier estimator for the time to death
  - > to compute the Kaplan-Meier estimator you will need function survfit()
  - b to plot it, just use the plot() function on the resulting object



- T2: Calculate and plot the Kaplan-Meier estimator for the time to death, separately for the two treatment groups
  - ▷ what do you observe?
- T3: Calculate and plot the Kaplan-Meier estimator for the time to death, separately for males and females
- T4: Calculate the log-rank tests for the two treatment groups and for males versus females
  - > you will need function survdiff(), which has a very similar syntax as survfit()



• T5: We are interesting in studying the relationship between the hazard for death, and drug, gender, AZT, and CD4. Fit a Cox model that relaxes the linearity assumption for the effect of CD4 using natural cubic splines (you need function ns()). In addition, assume that there is an effect drug, gender and AZT on the hazard for death, but the effect of these predictors is different for different levels of CD4 cell count

□ use the summary() method and try to interpret the results

- T6: Use a likelihood ratio test to test whether the model can be reduced by dropping all interaction terms
  - ▷ use the anova() function



- T7: Use the summary() method to obtain a detailed summary of the second fitted model. What is the interpretation of the estimated coefficient for drug? In addition, in the output you have values for exp(coef) and exp(-coef). What do these values represent?
- The main motivation to introduce the semiparametric Cox model was to avoid the impact of a possibly wrong assumption for the distribution of the event times
- However, all statistical models make assumptions in the Cox model we make no assumption for the distribution of  $T_i^*$  but we do make other assumptions:



- If PH is seriously violated, then the results we obtain from the Cox model may not be trustworthy!
- In practice, PH means that the effect of a covariate in the risk for an event is constant over time

- Some times the PH assumption may not be reasonable, e.g.,
  - $\triangleright$  the new treatment requires a time period to start working  $\Rightarrow$  at the beginning of follow-up the risk for the treatment group is the same as in the control group, however we expect that later the risk for the treatment group will decrease

▷ . . .



• To check the PH assumption we will (hypothetically) consider an extension of the Cox model, namely the Cox model with a *time-dependent coefficient* 

$$h_i(t) = h_0(t) \exp\{X_i \beta(t)\}\$$

where, the effect of X on the hazard varies with time

ullet Grambsch and Therneau (Biometrika, 1994) have shown that, if  $\widehat{\beta}$  is the estimated coefficient from the ordinary (time-independent) Cox model, then

$$\beta(t) \approx \widehat{\beta} + E\{s^*(t)\}$$

where  $s^*(t)$  is the scaled Schoenfeld residual



- The formula and rationale behind the scaled Schoenfeld residuals is rather technical 

  ▷ we will not give them here (see Therneau & Grambsch (2000) for more info)
- Plotting scaled Schoenfeld residuals against time or suitable transformation of time, reveals violations of the PH assumption
- An additional advantage of the scaled Schoenfeld residuals is that they can be used to statistically test PH (though this is not advisable)



- T8: In R, plots of the Schoenfeld residuals are calculated by function cox.zph()
  - □ use this function on the final Cox model you fitted above
  - > use the plot() function to produce the plots (before running plot(), run
    par(mfrow = c(3, 3)))
  - b we will interpret together the results...

• T9: Check if conclusions change by using other transformations of the time variable (i.e., argument transform of cox.zph())

Chapter 8
Closing

#### 8.1 Concluding Remarks



#### When we need mixed effects models?

b to analyze correlated data structures, e.g.,

- \* longitudinal data
- \* hierarchical data

\*

#### • How mixed models work?

- > a model for the responses given random effects
- ▷ a model for the random effects

#### 8.1 Concluding Remarks (cont'd)



#### What we did not cover

▷ . . .

The End!