

# Single cell transcriptomics

Analysis tools & QC

cells ->

genes ->

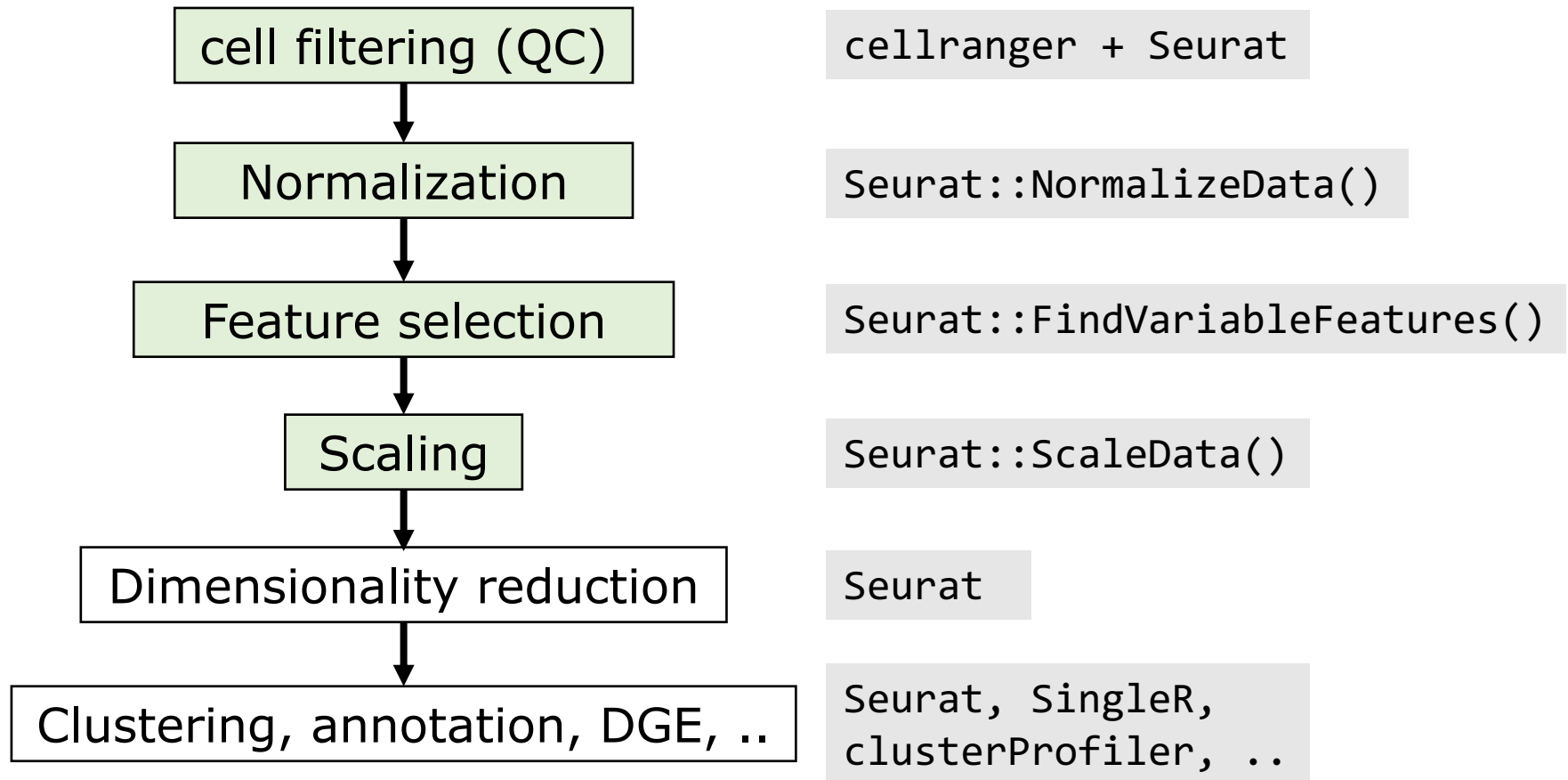
|         | ATAC-1 | CCG-1 | GCGA-1 |
|---------|--------|-------|--------|
| RPL22   | 5      | 13    | 3      |
| PARK7   | 0      | 9     | 3      |
| ENO1    | 1      | 35    | 0      |
| PLA2G2A | 0      | 0     | 0      |
| CAMK2N1 | 0      | 6     | 0      |
| CDC42   | 0      | 8     | 1      |
| C1QA    | 0      | 0     | 25     |
| C1QC    | 0      | 0     | 25     |
| C1QB    | 0      | 0     | 29     |
| ID3     | 0      | 35    | 0      |
| RPL11   | 18     | 29    | 16     |
| CLIC4   | 0      | 4     | 0      |

# Frequently used analysis tools

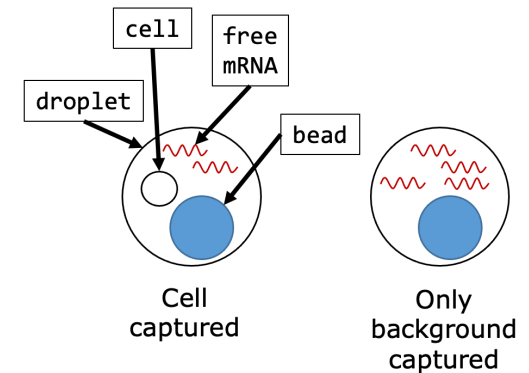
- Major tools perform (at least) the following:
  - QC
  - normalization & scaling
  - dimensionality reduction
- scanpy (python)
- scater + scan (R, Bioconductor)
- monocle3 (R, beta on github)
- Seurat (R, CRAN)



# Analysis overview



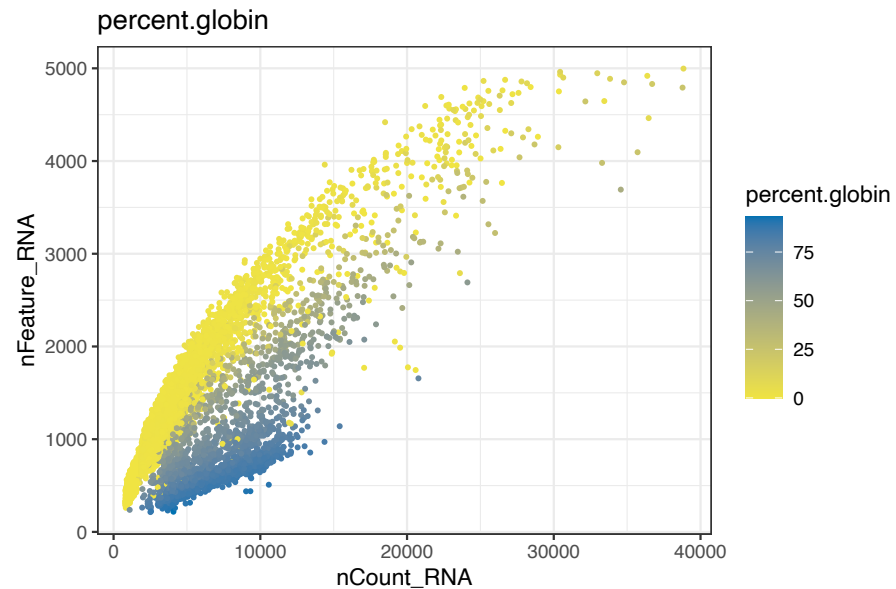
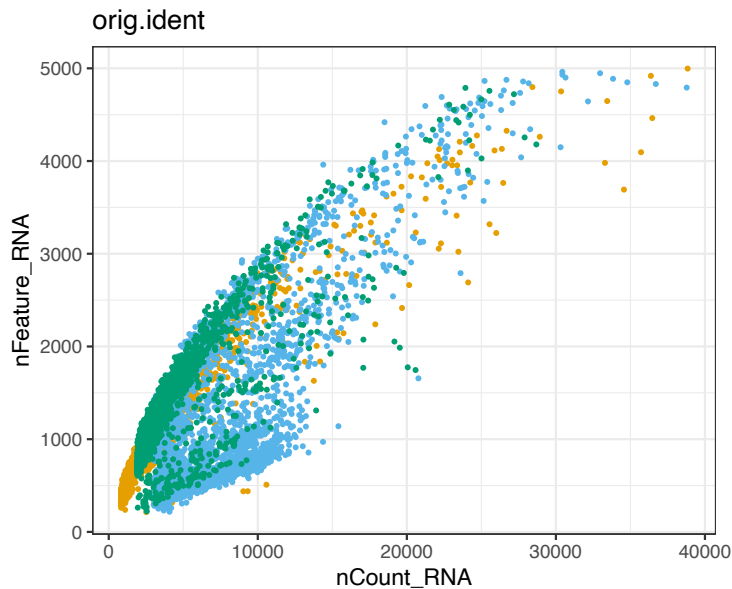
# Cell filtering

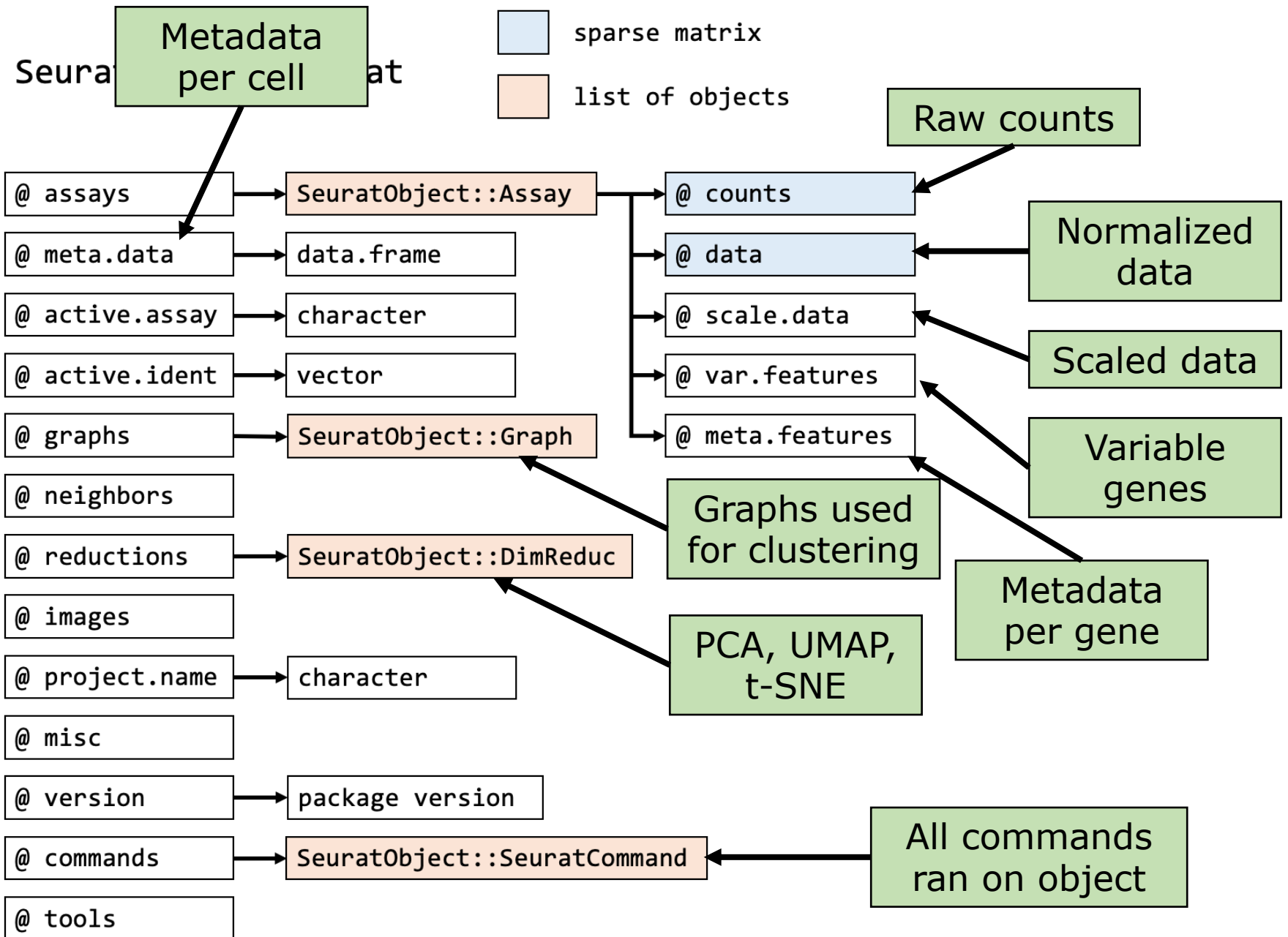


- Cellranger:
  - cell calling (filter against low #UMI)
- Manually (e.g. with Seurat):
  - #UMI: high -> possible doublet
  - #detected genes
  - % mitochondrial UMI: dying cells
  - % ribosomal UMI
  - % globin UMI
  - Relationships between variables

# Cell filtering

Often it makes sense to look at relationships





# Normalization & scaling

- **Normalization** (per cell): remove technical effects (i.e. library size)
- **Scaling** (per gene): standardize range, mean and variance

Both are mainly for the purpose of dimensionality reduction



# Regress out variables

- Typically done after visualization with PCA/UMAP/t-SNE
- Embedding might be according to e.g. cell cycle phase -> but you want cell type

