### Impacts of climate change on high priority fruit fly species in Australia

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## Abstract

Tephritid fruit flies are among the most destructive horticultural pests and pose risks to Australia's multi-billion-dollar horticulture industry. Currently, there are 11 pest fruit fly species of economic concern present in various regions of Australia. Of these, nine are native to this continent (*Bactrocera aquilonis, B. bryoniae, B. halfordiae, B. jarvisi, B. kraussi, B. musae, B. neohumeralis, B. tryoni* and *Zeugodacus cucumis*), while *B. frauenfeldi* and *Ceratitis capitata* are introduced. To varying degrees these species are costly to Australia's horticulture through in-farm management, monitoring to demonstrate pest freedom, quarantine and trade restrictions, and crop losses. Here, we used a common species distribution modelling approach, Maxent, to assess habitat suitability for these 11 species under current and future climate scenarios. These projections indicate that the Wet Tropics is likely to be vulnerable to all 11 species. The east coast of Australia will likely remain vulnerable to multiple species until at least 2070. Both the Cape York Peninsula and Northern Territory are also likely to be vulnerable, however, extrapolation to novel climates in these areas decrease confidence in model projections. The climate suitability of current major horticulture regions in north-

26 western Australia, the Northern Territory, southern-central regions of New South Wales and

southern Victoria to these pests is projected to increase as climate changes. Our study highlights

areas at risk of pest range expansion in the future, to guide Australia's horticulture industry in

developing effective monitoring and management strategies.

**Keywords:** Tephritidae, fruit flies, species distribution modelling, habitat suitability, climate

change

capitata) [3] being highly polyphagous.

# Introduction

Tephritid fruit flies are one of the most destructive and economically significant pest insect families, attacking a wide range of fruit and vegetables. While the family contains more than 4000 species, around 350 are recognized as economically important horticultural pests [1] that have significant impacts on global horticultural production and market access. In Australia, the average annual value of crops susceptible to fruit flies is ~\$4.8 billion [1], and the National Fruit Fly Strategy has identified 46 species as 'high priority pests' [2] of concern. The majority of these species are exotic to Australia, primarily found in South-East Asia and the South Pacific [1, 2], and are yet to establish populations in Australia. Of the 11 species that are currently present in Australia [1-3] (Table 1), seven are reported to cause significant economic losses (Bactrocera aquilonis, B. jarvisi, B. neohumeralis, B. musae, B. tryoni, Ceratitis capitata, and Zeugodacus cucumis) [1, 4]. Combined, these species infest a wide variety of hosts, with some (e.g. B. frauenfeldi, B. jarvisi, B. neohumeralis, B. tryoni and Ceratitis

The distributions of Australia's pest fruit fly species are influenced by their climatic tolerances

and the distributions of their hosts. Bactrocera originated in tropical regions, and have their

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highest richness in rainforests [5]. However, over the last 100 years, as horticulture has proliferated across Australia, some species have expanded their geographic range and host breadth [6]. Of the 11 high priority fruit fly species presently on the continent, three are currently restricted to north-east Queensland (B. frauenfeldi, B. kraussi and B. musae) [7]. In contrast, the geographic range of *B. neohumeralis* (Lesser Queensland fruit fly) extends along eastern Australia, from Queensland to central New South Wales (NSW) [3, 4, 7]. Previous climatic analysis indicates that this species also has the potential to establish elsewhere in northern Australia [4]. The remaining species have substantially wider climate tolerances, and are found across broad regions of the continent. For instance, B. tryoni (Qfly) ranges across much of eastern Australia, eastern Oueensland and northern regions of the Northern Territory [8]. Bactrocera jarvisi (Jarvis' fruit fly) extends from northwest Western Australia, across the Northern Territory to northern Queensland and the Torres Strait Islands [4, 9], and, in favourable years, may spread down the east coast of Australia into northern coastal NSW [4, 9]. Hence, B. jarvisi and B. tryoni have overlapping geographic ranges and infest many of the same hosts [4]. Ceratitis capitata (Medfly) originated from the Afrotropical region [10], and was introduced into the Perth area (Western Australia) in the late 1800s [4]. Before quarantine controls were developed, this species spread to NSW, Victoria, and other parts of Australia [11]. However, for reasons that remain unclear, Ofly is believed to have displaced Medfly throughout most of its former Australian range [12], and now Medfly is confined to Western Australia, with occasional detections in South Australia [13]. Under current climate conditions, most of these 11 fruit fly species pose threats to Australia's horticulture industries, as well as to backyard growers. As such, controlling fruit flies is imperative for the viability of Australian horticulture, necessitating in-farm management and

pest treatment, monitoring to demonstrate pest freedom, and quarantine and trade restrictions

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[1, 2]. These controls, along with loss of market access, are estimated to cost Australian growers \$100 million per annum [4], in addition to losses of up to \$159 million per annum due to infestation of fruit and vegetable crops [14]. For those areas where fruit flies are found, the annual cost, as reported in 2012, of bait and cover spray, as well as post-harvest treatments, amount to \$269 ha<sup>-1</sup> and \$62.36 tonne<sup>-1</sup>, respectively [15], while maintaining fruit fly free areas is estimated to exceed \$28 million per annum based on data from 2009-2011 [16]. However, restrictions were recently placed on the use of insecticides to control fruit flies due to concerns about toxicity [17], with dimethoate and fenthion suspended or highly restricted for many horticultural crops [17-20]. Other approaches, including Sterile Insect Techniques, are now being explored. Regardless, it has been estimated that the annual likelihood of an incursion by an exotic fruit fly species is 21% [15], and the annual cost of eradicating these incursions is ~\$13 million [16], with rapid responses to outbreaks being crucial for eradication [21]. Even brief incursions can result in significant economic damage due to market access restrictions that may be imposed. However, climate change is likely to alter the distribution of suitable habitat for fruit fly species and areas vulnerable to outbreaks, and this could have serious repercussions for Australian horticulture [22]. Previous studies [22-24] have used the semi-mechanistic species distribution model (SDM), CLIMEX, to estimate the potential geographic distributions of several high priority fruit fly species, based on their performance along climatic gradients. While highly useful in furthering our understanding of climate impacts on fruit flies, these studies have either focused on other countries [24-26] or have explored global patterns of the distribution of suitable climate [22, 23]. Here we assess how climate change may result in shifts to the distribution of suitable

habitat for the 11 high priority fruit fly species present in Australia, using the correlative SDM, Maxent [27]. This SDM has been used extensively to assess the distribution of suitable habitat for a broad range of pests and invasive species [26, 28-31]. We also highlight areas at risk of pest range expansion, to guide Australia's horticulture industries in development of effective monitoring and management strategies.

Table 1. Eleven economically-significant tephritid pest species present in Australia.

Species	Common name	Geographical Range	Major Commercial Hosts [32]	References
Bactrocera aquilonis (May)	Northern Territory fruit fly	Top End of the Northern Territory (NT), northern areas of Western Australia	Bell pepper, tomato, lemon, mandarin, grapefruit, apple, mango, peach	1
Bactrocera bryoniae (Tryon)	N/A	Torres Strait Islands, mainland Queensland, northern Western Australia, NT, NSW as far south as Sydney	Chilli, tomato	3
Bactrocera halfordiae (Tryon)	Halfordia fruit fly	North Queensland south to the Sydney region in NSW	Citrus	3
Bactrocera jarvisi (Tryon)	Jarvis' fruit fly	North-western Western Australia, NT, north-west Queensland, eastern Australia from Cape York to Sydney, NSW	Mango, peach, banana, pear, apple, pawpaw, persimmon	3
Bactrocera kraussi (Hardy)	Krauss' fruit fly	Torres Strait Islands, northeast Queensland as far south as Townsville	Grapefruit, mandarin, orange, mango, peach and banana	3,7
Bactrocera musae (Tryon)	Banana fruit fly	Torres Strait Islands, northeast Queensland as far south as Townsville	Banana	3,7
Bactrocera neohumeralis (Tryon)	Lesser Queensland fruit fly	Torres Strait Islands, eastern Queensland, northern NSW	Mango, papaw, persimmon, avocado, banana, passionfruit, apple, apricot, plum, peach,	3

			citrus, capsicum, chilli, tomato	
Bactrocera tryoni (Froggatt)	Queensland fruit fly (Qfly)	Central and Top End of NT, eastern Australia, Victoria	Mango, papaw, avocado, grapefruit, passionfruit, strawberry, peach, pear, apple, banana, persimmon, chilli, capsicum, tomato, eggplant	3,31
Zeugodacus cucumis (French) (formerly Bactrocera cucumis)	Cucumber fruit fly	Eastern Queensland, north-eastern NSW, NT	Cucumber, pumpkin, zucchini, squash, passionfruit, tomato, pawpaw	3
Bactrocera frauenfeldi (Schiner)	Mango fruit fly	Native to Papua New Guinea and surrounding islands, spread to Torres Strait Islands and northern Queensland as far south as Townsville	Mango, banana, passionfruit, citrus, chilli	7
Ceratitis capitata (Wiedemann)	Mediterranean fruit fly (Medfly)	Native to Africa, spread to the Mediterranean regions, Western Australia, occasional detections in South Australia and NT are eradicated.	Mango, papaw, apple, peach, pear, citrus	3, 28, 31

Table 1. Eleven tephritid pest species present in Australia, including nine natives (*B. aquilonis*, *B. bryoniae*, *B. halfordiae*, *B. jarvisi*, *B. kraussi*, *B. musae*, *B. neohumeralis*, *B. tryoni* and *Z. cucumis*) and two introduced species (*B. frauenfeldi* and *C. capitata*), and their major commercial hosts.

# Methodology

#### Species occurrence data

We collected occurrence data for the 11 species from five main sources: the Australian Plant Pest Database (APPD; <a href="http://www.planthealthaustralia.com.au/resources/australian-plant-pest-database">http://www.planthealthaustralia.com.au/resources/australian-plant-pest-database</a>, accessed 15th March 2017), the Atlas of Living Australia (ALA; <a href="http://www.ala.org.au">http://www.ala.org.au</a>, 22nd December, 2016), the Global Biodiversity Information Facility

(GBIF, <a href="https://www.gbif.org">https://www.gbif.org</a>, 28th June, 2017), trap data, and existing literature. APPD is a national digital database of plant pest and pathogen specimens held within herbaria and insect collections across Australia. It is a powerful tool for market access and emergency responses to pest incursion, and supports associated research activities. ALA is Australia's largest digital database of species occurrence records, containing information from a wide array of data providers including Australia's major museums and government departments. GBIF provides similar data at a global scale. Before downloading data from APPD, ALA and GBIF, we applied filters to restrict records to those that were resolved to species-level, were dated no earlier than 1 January 1950, contained valid geographic coordinates, and were not flagged as 'environmental outliers'.

We also collected trap data from various state government departments (Biosecurity and Food Safety, Department of Primary Industries, NSW; Biosecurity Queensland and the Queensland Department of Agriculture and Fisheries; Department of Economic Development, Jobs, Transport and Resources, Victoria; and Department of Primary Industries and Regions South Australia (PIRSA)). Trap data from these sources were collected at different periods from 1996 to 2017. Finally, we also obtained occurrence data from the literature [1-4, 6, 7, 11, 33-37].

#### Major commercial fruit and vegetable hosts

For each of the 11 fruit fly species, we compiled information on the major commercial hosts on which infestation has been recorded. For this purpose, we defined major fruit and vegetable host species according to the Australian Horticulture Statistics Handbook (HSHB; www.horticulture.com.au) for the year 2016/2017 [32]. This document consolidates horticulture statistics of interest to industry members and other stakeholders. The data contained in HSHB were derived from the Australian Bureau of Statistics, projects funded by

Hort Innovation, international trade sources and horticulture industry representative bodies where available.

#### Climate data

For current and future climate conditions we used the bioclimatic variables available within the WorldClim database, at a spatial resolution of 30 arc-seconds [38] (approximately 1 km; http://www.worldclim.org). These data, based on meteorological records for the period 1960–1990, comprise 19 climatic variables, 11 of which are temperature-based while eight relate to precipitation. Combined, the data represent annual trends, seasonality, and limiting or extreme environmental conditions. Assuming that host plants are available, temperature and moisture are the key factors influencing fruit fly reproduction and survival [18, 39]. Thus, these variables were chosen as predictor candidates based on the fruit flies' biology and ecological requirements, and similar habitat suitability studies undertaken on other insects [40]. For each species, we identified a set of ecologically-relevant variables, with minimal collinearity, that resulted in high predictive power for the model [41] (described below).

When projecting the future suitability of habitat, we considered a range of climate scenarios to acknowledge this important aspect of uncertainty. We choose six global climate models (GCMs) recommended by CSIRO as being useful for Australian climate impact assessments [42], and for which data were available at our chosen spatial resolution for the Representative Concentration Pathway 8.5 (RCP8.5) [43]. These GCMs included: CanESM2 (The Second Generation of Canadian Earth System Model); ACCESS1.0 (The Australian Community Climate and Earth System Simulator); MIROC5 (Model for Interdisciplinary Research on Climate); HadGEM2-CC (Hadley Centre Global Environmental Model Version 2 Carbon Cycle); NorESM1-M (The Norwegian Earth System Model-Part-1); and GFDL-ESM2M

(Global Coupled Climate Carbon Earth System Model Part-1). The CanESM2 model projects a hot future with drying across central regions and higher precipitation in the north-east. The ACCESS1.0 model projects a hot and dry future across most areas of Australia, while MIROC5 projects moderate warming, with drying in the north-east and south-west but higher precipitation in central Australia. NorESM1-M projects moderate warming. HadGEM2-CC and GFDL-ESM2M project a hot future with warming typically in central regions. We downloaded the 19 bioclimatic variables from these six models for 20-year periods centred on 2030, 2050 and 2070 (http://www.climatechangeinaustralia.gov.au). These data were then reprojected to a spatial resolution of 1 × 1 km (Australian Albers Equal Area, EPSG: 3577) via bilinear interpolation, using the gdalwarp function provided by the R package gdalUtils [44] in R version 3.3.3 [45].

#### **Species Distribution Models**

We used the machine learning approach, Maxent (v3.3.3k [27]), to assess habitat suitability for species under current and future climate scenarios. Maxent accommodates presence-only data and has performed well in multimodel assessments [46]. It produces a continuous probability surface, which can be interpreted as an index of habitat suitability given the predictor variables included in model calibration. Detailed descriptions of Maxent are given elsewhere [47, 48]. We optimized models by assessing the effects of different combinations of feature types, of competing predictor sets deemed ecologically sensible *a priori*, and of the extent of regularization on model performance. We found that Maxent performed best when product (first-order interactions), linear and quadratic features were used, with a regularization multiplier of 1 (the default), and used this configuration to calibrate our final models.

Maxent requires background data, to which it compares the environmental characteristics of

presence locations. There is flexibility for users to specify which points to use as background, as well as the number of records and the spatial extent from which they are chosen [47]. Following Ihlow *et al* [49], we manually generated background points by randomly selecting 100,000 cells from terrestrial areas within 200 km of occurrence records of the target species. Our choice of background achieves a balance between fine-scale discrimination of suitable and unsuitable sites along environmental gradients, and generalization of model predictions.

To assess model performance, we used five-fold cross-validation to reduce model errors that may occur from the random splitting of data into test and training subsets. The performance of each model was evaluated using the area under the receiver operating characteristic curve (AUC), which describes the consistency with which a model ranks randomly chosen presence sites as more suitable than randomly chosen background sites. AUC ranges from 0 to 1, with a value of 0.50 indicating discrimination ability no better than random, while values greater than 0.75 indicates that the model has a discriminative ability that is better than "fair" [50]. Cross-validated AUC scores were presumed to reflect the performance of a single final model for each species, which used all available data.

Following previous studies of pest species [26], continuous habitat suitability scores projected by Maxent models were converted to binary layers (0 = unsuitable, 1 = suitable) using the 10th percentile training presence threshold (i.e. the value that corresponds to 10% training omission). We note that the selection of a threshold value is subjective and may vary depending upon the goals of the study [51], thus we also provide continuous output for current climate as supplemental data (S1-11 Figs). For each species, the six binary suitability grids (i.e., one for each GCM, with cells assigned 0 when unsuitable and 1 when suitable) for each time period were summed to produce a consensus map, identifying agreement about the suitability of grid

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cells across the six climate scenarios. Each species' consensus map was then converting to a binary map indicating whether cells were projected to be suitable under the majority of GCMs (i.e., suitable in  $\leq 4$  GCMs = 0, suitable in 4 or more = 1). The resulting binary maps were summed across species to identify hotspots - grid cells suitable for multiple pest species. Finally, we compared the distribution of hotspots to that of major horticultural crops. When projecting models, extrapolation to conditions beyond the range of the training data may be unreliable. Following Elith et al. [52] we developed MESS (multivariate environmental similarity surface) maps to identify regions of extrapolation [52]. By revealing areas with novel environmental conditions, MESS maps can be used as a projection mask, highlighting regions for which projections are unreliable, or as a quantitative measure of prediction uncertainty [52]. To assess how novel environments may alter projections of habitat suitability, for each species we visually compared maps of habitat suitability that included projections made in areas with novel environmental conditions to maps that classified these areas as unsuitable. All modelling and post-modelling analyses and calculation of statistics were performed in R version 3.1.2 [53]. We used the sp [54] and raster [55] packages for preparation and manipulation of spatial data, the dismo package to fit Maxent models, and custom R code for rapid projection of fitted models. Results **Model Performance** Model performance for all species was better than random, with average cross-validated AUC ranging from 0.815 (SD = 0.05; B. frauenfeldi) to 0.907 (SD = 0.02; B. neohumeralis)

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(S1 Table).

**Bactrocera aquilonis:** Our model suggested that suitable habitat for B. aquilonis currently exists in the northern regions of the Northern Territory and Western Australia, as well as in northern Queensland, where this fly has not been reported (S1A-1B Figs). The variables with the highest permutation importance were precipitation of the wettest guarter (68.9%) and annual mean temperature (28.9%) (S1 Table). As the century progresses, the geographic extent of suitable habitat for this species is projected to increase and shift southwards under all six scenarios (S1C-1E Figs). Further, most areas currently suitable are projected to remain so until at least 2070. By 2030, 21.5% of Australia (i.e. ~1,600,100 km<sup>2</sup>) is projected to be climatically suitable under at least one climate scenario (S3 Table), and this is projected to increase to 39.9% ( $\sim 3,000,100 \text{ km}^2$ ) by 2070 (S3 Table). However, a substantially smaller area is projected suitable under all six scenarios (2030 = 9.2%,  $\sim$ 700,100 km<sup>2</sup>; 2070 = 20.7%,  $\sim$ 1,590,800 km<sup>2</sup>) (S3 Table). This includes northern Western Australia, much of the Northern Territory, and north-western Queensland (S1C-1E Figs). Key horticultural crops for B. aquilonis are Mangifera indica (mango), Citrus × paradisi (grapefruit), Malus domestica (apple), Prunus persica (peach) and Citrus sp. (citrus) (S4 **Table).** The major regions where these crops are currently grown include the Northern Territory and north-east Western Australia. These regions will likely remain suitable for B. aquilonis until at least 2070. Similarly, fruit growing regions in the Wet Tropics (north-east Queensland) are likely to increase in suitability in the future. Other major host-plant growing regions in the south and east of the continent will likely remain unsuitable (S1 Fig).

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**Bactrocera bryoniae:** Current suitable habitat for B. bryoniae is projected to occur along the northern and eastern coastlines (S2A-2B Figs). Temperature annual range and precipitation of the driest month contributed the most to the model for this species (42.2% and 27.4%, respectively) (S1 Table). By 2070, suitable habitat is projected to increase under all scenarios except GFDL-ESM2M (which projects a hot, very dry future) (S2C-2E Figs; S2 Table), expanding to the southern coastlines of Victoria and Western Australia. By 2030, 13.5% of Australia (i.e. ~1,038,000 km<sup>2</sup>) is projected to be suitable for B. bryoniae under at least one of the climate scenarios, increasing to 20.6% (~1,500,700 km<sup>2</sup>) by 2070 (S3 Table). Under 1-3 scenarios, suitable habitat is projected to shift inland in Queensland and NSW. However, the amount of habitat projected to remain suitable under all six scenarios remains relatively stable from 2030-2070 (i.e. spanning 6.5–7.1% of the continent) (S3 Table). The major horticultural host for B. bryoniae is Capsicum annuum (chilli) (S4 Table). Our model indicates that key growing regions for this crop in Queensland currently contain suitable habitat for B. bryoniae, and this will continue to be the case until at least 2070 (S2 Fig). **Bactrocera frauenfeldi:** Currently, suitable habitat for this species is projected to be mostly confined to Cape York Peninsula and the Wet Tropics, although there are also small areas in northern Western Australia and the Northern Territory that are classified as suitable, but from which the species has not been recorded (S3A-3B Figs). The most important variable in the model for B. frauenfeldi was precipitation of the wettest quarter (75.4%) (S1 Table). As the century progresses, suitable habitat is projected to increase under all scenarios except

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CanESM2 (S2 Table). This scenario projects a hot, very dry future, leading to loss of suitable habitat in northern Oueensland by 2050. In contrast, under the ACCESS1.0 scenario (a hot, dry scenario) total range size may increase by 43.6% by 2030. However, by 2070, the Cape York Peninsula region is projected to become unsuitable (S2 Table). Overall, 3.3% of Australia (i.e. ~250,400 km<sup>2</sup>) is projected to be suitable for *B. frauenfeldi* under at least one scenario by 2030, increasingly slightly to 4% ( $\sim 300,000 \text{ km}^2$ ) by 2070 (S3 Table). Only 1.3% ( $\sim 100,000 \text{ km}^2$ ) of the continent is projected to be suitable under all six scenarios by 2070 (S3 Table). The major crops for B. frauenfeldi are Mangifera indica (mango) and Carcica papaya (pawpaw) (S4 Table). Major production regions in north-western Northern Territory may remain suitable for this species until at least 2070, although there is substantial uncertainty across the climate scenarios. In contrast, it is very likely that the Wet Tropics will remain suitable until at least 2070, irrespective of the climate scenario (S3 Fig). **Bactrocera halfordiae:** Suitable habitat for *B. halfordiae* is currently found in the Wet Tropics and subtropics from north Queensland to eastern New South Wales (S4A-4B Figs). Precipitation of the driest month (66.8%) and annual mean temperature (32.3%) contributed most to this model (S1 Table). The geographic extent of suitable habitat is projected to vary considerably across the six climate scenarios, however as the century progresses, gains in new habitat may exceed losses under some scenarios (e.g. see ACCESS and MIROC5 in S2 Table). In contrast, under the CanESM2 scenario (which projects a hot future, drying across central regions and higher precipitation in the north-east), total range size may decline by 65.1% by 2030 (S2 Table), mostly due to contractions in the south and east, although limited gains in habitat may occur in

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northern Australia. Similarly, under the HadGEM2 scenario (which projects substantial drying across some regions of the continent), 51.4% of current suitable habitat is projected to be lost by 2030, although by 2070, range expansions may exceed losses (S2 Table). By 2030, 6.9% of Australia (i.e. ~500,800 km<sup>2</sup>) is projected to be suitable for *B. halfordiae* under at least one climate scenario, increasing to 9.9% (~750,900 km²) by 2070 (S3 Table). Comparing the current and future distribution of suitable habitat for this fly with the major growing regions for its host plants indicates that crops in the Wet Tropics may continue to be at risk, until at least 2070. However, only 1–2 scenarios project horticultural regions in southern Oueensland to retain suitable climate (S4C-4E Figs). Although horticultural regions along the NSW-Victorian border are currently unsuitable for B. halfordiae, some models project these areas to become suitable between 2050–2070 (S4 Fig). Bactrocera jarvisi: Current suitable habitat for this species is projected to be mostly confined to northern Western Australia, the Top End of the Northern Territory, and eastern Australia from Cape York to NSW (S5A-5B Figs). Annual mean temperature (38.0%) and precipitation of driest month (37.2%) had the highest contributions to the model for this species (S1 Table). There is substantial consensus across the six scenarios that regions currently suitable for B. jarvisi will remain so until at least 2070 (S5C-5E Figs; S2 Table). In addition, across some models, gains are projected to occur in Western Australia, the Northern Territory and central Queensland. For instance, under the CanESM2 scenario, total range size may increase by 45.5% to 58.7% between 2030 and 2070. Similarly, under the hot, very dry scenario projected by ACCESS1.0, 62.5% of future suitable habitat may occur in new areas by 2070 (S2 Table). By 2030, 25.8% of Australia (i.e.~1,980,000 km²) is projected to be suitable for *B. jarvisi* under at least one of the climate scenarios, and due to subsequent gains in suitable habitat, this may

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increase to 44.3% ( $\sim 3,390,400 \text{ km}^2$ ) by 2070 (S3 Table). Comparing the distribution of suitable habitat for this fly with that of its major host crops indicates that crops currently grown in the Top End of the Northern Territory, and in eastern Australia from Cape York to New South Wales, may continue to be at risk until at least 2070. Other major host-plant growing regions in the south and west of the continent will also remain suitable for this species until 2070 (S5 Fig). **Bactrocera kraussi:** Suitable habitat for B. kraussi is projected to occur across the northern tip of Australia and northeast Oueensland, as far south as Townsville (S6A-6B Figs). Precipitation of the wettest quarter (75.19%) had the highest contribution to the model of B. kraussi (S1 Table). The geographic extent of suitable habitat is projected to increase under all six scenarios (S6C-6E Figs; S2 Table). For example, under the NorESM1-M scenario (a moderate warming scenario with little precipitation change), an area equivalent to 29.6% of current suitable habitat is projected to be gained by 2070 (S2 Table). By 2030, 3.1% of Australia (i.e.~240,600 km<sup>2</sup>) is projected to be suitable for B. kraussi under one or more of the climate scenarios, increasing to 3.8% (~280,300 km<sup>2</sup>) by 2070 (S3 Table). This includes north-east Oueensland (S6C-6E Figs). Horticultural production regions in northeast Queensland as far south as Townsville may remain suitable for this species by 2070, although production regions in the south are likely to remain unsuitable (S6 Fig). **Bactrocera musae:** Current suitable habitat for B. musae is predicted from the Torres Strait Islands through to the Wet Tropics (S7A-7B Figs). The most important variable in the model

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for B. musae was precipitation of the wettest quarter (78.7%) (S1 Table). Projections for this species under climate change are similar to those for *B. kraussi*. Under the CanESM2 scenario, 22.2% of current suitable habitat is projected to be lost by 2030, although by 2070, range expansions are projected to exceed losses (S2 Table). However, in NorESM1-M (a moderate warming scenario with little precipitation change), total range size may increase by ~43.5% by 2070 (S2 Table). By 2030, 3.4% of Australia (i.e.~260,100 km<sup>2</sup>) is projected to be suitable for B. musae under at least one of the climate scenarios (S3 Table), increasing to 4.2% (~300,600 km<sup>2</sup>) by 2070 (S3 Table). B. musae mainly attacks Musa × paradisiaca (banana), the production areas for which are located primarily in tropical and subtropical regions of the continent (S4 Table). The major commercial growing region in the Wet Tropics is projected to remain climatically suitable for this species until at least 2070 (S8 Fig). **Bactrocera neohumeralis:** Current suitable habitat for this species is projected to be mostly confined to the Torres Strait Islands, eastern Queensland, and north eastern NSW south to Wollongong (S8A-8B Figs). Precipitation of the wettest month (47.4%) contributed most to the model for *B. neohumeralis* (S1 Table). As the century progresses, considerable differences in suitable habitat are projected across the six scenarios. For example, under the CanESM2 scenario, 24.2% of current suitable habitat is projected to be lost by 2030, although by 2070, range expansions are projected to exceed losses (S2 Table). Similarly, under the hot, very dry scenario simulated by GFDL-ESM2M, total range size may decline by 32.4% by 2030, mostly due to contractions in the south and east,

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although limited gains in habitat may occur in northern Australia (S8C-8E Figs; S2 Table). By 2030, ~6.9% of Australia (i.e.~500,500 km<sup>2</sup>) is projected to be suitable for *B. neohumeralis* under at least one of the climate scenarios, increasing to 14% (~1,000,100 km<sup>2</sup>) by 2070 (S3 Table). Production regions in eastern Queensland and north-eastern NSW will likely remain suitable for this species until at least 2070, although there is substantial uncertainty across the climate scenarios. In contrast, regions along the NSW-Victorian border and further south are projected to remain unsuitable for B. neohumeralis (S8 Fig). Bactrocera tryoni: Highly suitable habitat for B. tryoni is projected to occur along southwestern Western Australia, south-eastern South Australia, Victoria, and eastern Australia from Cape York to NSW (S9A-9B Figs). Coastal zones in northern Western Australia, the Northern Territory and the eastern half of Tasmania have moderate suitability (S9A-9B Figs). Annual mean temperature (33.06%) and mean temperature of the coldest month (32.42%) had the highest contributions to the model for this species (S1 Table). The geographic extent of suitable habitat varies across the six climate scenarios. However, as the century progresses, gains in new habitat may exceed losses under some scenarios (e.g. see ACCESS1.0, MIROC5 and NorESM1-M; S2 Table). In contrast, under the GFDL-ESM2M scenario, total range size may decline by 47.9% by 2030 (S2 Table), mostly due to contractions in the south and east. Under the ACCESS1.0 scenario, 26.1% of current suitable habitat is projected to be lost by 2030, although by 2070, range expansions are projected for some regions (S2 Table). By 2030, 26.1% of Australia (i.e. $\sim$ 1,990,800 km<sup>2</sup>) is projected to be suitable for B. tryoni under at least one scenario, increasing to 39.6% (~3,000,500 km<sup>2</sup>) by 2070 (S3 Table).

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This includes south-west Western Australia, most of Victoria and eastern Tasmania, eastern Queensland and the northern reaches of the continent (S9C-9E Figs). Comparing the distribution of suitable habitat for this fly with that of its major host crops indicates that key regions for these crops in the Top End of Northern Territory, eastern Australia from Cape York to NSW, Victoria, and some parts of Tasmania, may remain suitable for B. tryoni until at least 2070. Major host-plant growing regions in South Australia may also remain suitable for this species until 2070 (S9 Fig). Ceratitis capitata: Our model suggests that suitable habitat for C. capitata exists throughout Western Australia, the Northern Territory, the east coast of Queensland to NSW and South Australia (S10A-10B Figs). Annual mean temperature (47.2%) and mean temperature of the coldest month (46.2%) contributed most to the model for this species (S1 Table). Under the future climate scenarios, the geographic extent of suitable habitat is projected to increase and shift south **(S10C-10E Figs).** By 2030, 28.3% of Australia (i.e. ~2,100,400 km<sup>2</sup>) is projected to be suitable for *C. capitata* under one or more scenarios (S3 Table), increasing to 47.1% (~3,600,800 km<sup>2</sup>) by 2070 (S3 Table). However, under all six scenarios, 13.9% (2030; ~1,000,000 km<sup>2</sup>) to 17.3% (2070; ~1,300,000 km<sup>2</sup>) of Australia is likely to be suitable for *C. capitata* (S3 Table). Climate suitability within production regions in the south and west of the continent may gradually increase for this species from 2030 to 2070 (S10 Fig). **Zeugodacus cucumis:** Suitable habitat for *Z. cucumis* is projected to occur along the northern region of Western Australia and the Northern Territory, north-east Queensland, and south along the east coast to NSW (S11A-11B Figs). Precipitation of the driest quarter (54.3%) and mean temperature of the coldest quarter (36.2%) had the highest permutation importance in the model for this species (S1 Table).

Under future climate scenarios, the geographic extent of suitable habitat is projected to increase, expanding southward and inland, with most areas that are currently suitable projected to remain so until at least 2070 (S11C-11E Figs). However, there is considerable variation among projections for inland regions, indicating higher uncertainty about the future suitability of these regions. For instance, under a scenario where most regions, except central Australia, are projected to become drier (ACCESS1.0), total range size may increase by 68.2% by 2070 (S2 Table), with areas of expansion projected to occur in northern Western Australia, central Australia, eastern Queensland and NSW (S11C-11E Figs). Under a warmer/wetter scenario (MIROC5), a substantial westward range expansion is projected in eastern Australia (S2 Table). Hence, although 35.4% (~2,700,900 km²) of Australia is projected to be suitable for this species under at least one scenario for 2070, a much smaller extent is suitable under all six scenarios (10.3%, ~789,900 km²) (S3 Table).

Major commercial growing regions for host crops in Queensland and the Northern Territory are projected to remain climatically suitable for this species until at least 2070 (S11 Fig). Other major host-plant growing regions in the south and west of the continent will likely remain unsuitable under the time periods considered in this study (S11F Fig).

#### **Future hotspots of pest fruit flies**

For each time period, we stacked habitat suitability maps for all species, to identify regions most likely to contain suitable climate conditions for multiple pest species (i.e. hotspots). As the century progresses, the geographic extent of suitable habitat for most of the 11 species is

projected to expand and shift south regardless of whether novel environments are included or excluded (**Fig 1, S12 Fig, Table 2 and S5 Table**). When regions containing novel climate are included, 31.6% of Australia (i.e. ~2,400,800 km²) is projected to be currently suitable for at least one of the 11 species. By 2070, this may increase to 52.9% (~4,000,800 km²) (**Table 2**). only a small region within Queensland's Wet Tropics (5,621km²) is projected suitable for all 11 species.

Fig 1. Hotspot maps of habitat suitability for the 11 fruit fly species under climate change, when novel environments are included. Hotspot maps of current and future habitat suitability for 11 fruit flies. Suitability was modelled with Maxent, and thresholded using the 10th percentile of suitability at training presence localities. These maps include projections under novel environments. Colours indicate the number of species for which habitat is projected to be suitable under the majority ( $\geq 4$ ) future climate scenarios. Figure was created in R version 3.3.3 [45] (https://www.R-project.org/).

Table 2. Area (km²) and percentage (%) of Australia projected to be suitable for the 11 fruit fly species considered in this study, under current and future climates.

	Suitable area km² (% of Australia)				
Count	Current	2030	2050	2070	
0	5,250,277 (68.4%)	5,175,668 (67.5%)	4,567,601 (59.5%)	3,607,211 (47.0%)	
1	847,090 (11.0%)	738,118 (9.6%)	887,083 (11.6%)	1,122,410 (14.6%)	
2	850,401 (11.1%)	806,220 (10.5%)	1,133,009 (14.8%)	1,343,740 (17.5%)	
3	176,163 (2.3%)	257,558 (3.4%)	228,976 (2.9%)	501,960 (6.5%)	
4	93,709 (1.2%)	148,806 (1.9%)	248,057 (3.2%)	365,032 (4.8%)	
5	121,923 (1.6%)	196,041 (2.6%)	225,006 (2.9%)	299,590 (3.9%)	
6	89,766 (1.2%)	144,056 (1.9%)	141,026 (1.8%)	176,864 (2.3%)	
7	101,763 (1.3%)	46,306 (0.6%)	78,803 (1.0%)	89,042 (1.2%)	
8	32,385 (0.4%)	30,536 (0.4%)	37,543 (0.5%)	43,472 (0.6%)	
9	57,695 (0.8%)	102,927 (1.3%)	109,316 (1.4%)	10,8250 (1.4%)	
10	48,537 (0.6%)	21,554 (0.3%)	10,807 (0.1%)	9,888 (0.1%)	

11 3,371 (0.0%) 5,290 (0.1%) 5,853 (0.1%) 486 Table 2. Each row of the table indicates the area (and percentage) projected to be suitable now, in 2030, 2050, and 2070, for n species, where n is given in the "Count" column. Thus, the first row (with Count = 0) gives the 487 488 area projected to be unsuitable for all 11 species, the row with Count = 1 gives the area projected to be suitable 489 for any one of the 11 species, and the row with Count = 11 gives the area projected to be suitable for all 11 490 species. 491 When novel environments are excluded from maps, 31.5% of Australia is projected to be 492 currently suitable for at least one of the species, decreasing to 28% (~2,100,500 km<sup>2</sup>) by 2070 493 (S5 Table). Hence, exclusion of novel environments substantially impacts the size of suitable 494 habitat (i.e., a substantial area of suitable habitat is projected to occur in areas with novel 495 climatic conditions). However, parts of the Wet Tropics bioregion is projected to remain 496 suitable for all 11 species (70 km<sup>2</sup>) and the major commercial host plants within this bioregion 497 may continue to be at risk of invasion by most or all of these high priority species. 498 499 Major commercial host plant regions along the coastal strip of south-east Queensland and 500 north-east NSW are likely to have areas that are suitable under all future scenarios for B. 501 bryoniae, B. jarvisi, C. capitata and Z. cucumis (S2, S5, S10 and S11 Figs). Under some 502 503 scenarios, these regions may also be suitable for B. halfordiae, B. neohumeralis and B. tryoni (S4, S8 and S9 Figs). Some major commercial host plant regions in southern NSW and 504 Victoria are also projected to be suitable for B. jarvisi, B. tryoni and C. capitata under all 505 scenarios (S5, S9 and S10 Figs) and for B. halfordiae, B. neohumeralis and Z. cucumis under 506 a limited number of scenarios (S4, S8 and S11 Figs). 507 508 In south-west Western Australia, major horticulture regions are likely to remain suitable for B. 509 jarvisi, B. tryoni and C. capitata, although the latter species is currently not found in this region 510 511 (S5, S9 and S10 Figs). Commercial horticulture regions in the top-end of the Northern

Territory are also likely to be suitable for *B. jarvisi*, *B. kraussi*, *B. musae*, *B. tryoni* and *Z. cucumis* under all scenarios, and *B. frauenfeldi* under some climate scenarios. Horticultural regions in Tasmania are projected as suitable for *B. jarvisi*, *B. tryoni* and *C. capitata* (S5, S9 and S10 Figs).

### Discussion

Our study suggests that the Wet Tropics bioregion has climatically suitable habitat for the largest number of high priority tephritid pest species both now and as a result of climate changes projected to occur through to 2070. Cape York Peninsula and the Northern Territory are also likely to be vulnerable, although novel climates are projected to occur in these regions, and the extrapolation of SDMs to these conditions may be unreliable. The east coast of Australia is also likely to remain suitable for multiple species until at least 2070. As such, major horticulture regions in north-western Australia, the Northern Territory, southern-central regions of NSW, southern Victoria and north Tasmania may become increasingly suitable to high priority fruit flies. Two species, *B. tryoni* (Qfly) and *C. capitata* (Medfly), are projected to have suitable conditions in all states and territories of Australia, under all considered climate change scenarios, until at least 2070.

Models for both Qfly and Medfly were driven primarily by temperature parameters, rather than precipitation. Previous studies have identified climatic constraints on the distribution of Qfly. For instance, it has been reported that Qfly pupae do not survive in the winter months in Melbourne and near Sydney [56], and adults fail to emerge later than mid-April [57]. Further, many subtropical sites in Queensland are marginal in winter for Qfly breeding and general

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activity [8, 57]. As such, slight temperature increases associated with climate change are projected to substantially elevate the threat that this species poses to horticultural industries [58]. For instance, using data from the late 1990s, it was estimated that annual control costs for apple growers around Adelaide may increase by between \$346,000 and \$1.3 million with a 0.5–2°C increase in temperature [58].

With the exception of Western Australia, all Australian states and territories are currently free from Medfly, with market access protocols inhibiting movement into other states [21], and incursions met with immediate eradication programs [13]. Our model of current habitat indicates suitable conditions for Medfly around most of Australia's coastal regions. In addition to identifying suitability in the subtropical coastal fringe of Queensland, our model suggested that much of the low-altitude regions in the south-east, including parts of Tasmania, are also suitable. This is consistent with previous work using CLIMEX to estimate the potential distribution of Medfly [25]. Competition with Ofly may be responsible for exclusion of Medfly from much of Queensland [25], and similar biotic interactions may suppress the species elsewhere [13]. However, Medfly may be more tolerant to low temperatures and dry summers than Ofly [4], rendering Medfly the stronger competitor in areas with these conditions. Medfly was recorded in Tasmania in the 1920s but reportedly failed to survive an unseasonably hot and dry summer [4]. Due to their age, these records were not used to calibrate our model, yet our projections indicate that Tasmania continues to have conditions suitable for this species. Indeed, while strong market access protocols have excluded Ofly from Tasmania for around 90 years, our projections of Tasmania's suitability were validated upon discovery of an outbreak in 2018.

Allwood and Angeles [59] reported that B. jarvisi is recognized as a pest in north-western

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Australia, infesting mango, guava and pomegranates (as reported in [60]). Dominiak and Worsley [9] concluded that the current south-eastern range limit lies north of the Oueensland-NSW border (~25.5° south), while the south-western limit lies at approximately 18° south. However, previous analysis suggested that this species' current climatic range could extend into the cooler temperate areas of southern NSW, and eastern and northern Victoria [4]. Our models partly agree, indicating that suitable conditions currently occur along the east coast of Victoria. This species can also withstand very warm conditions, with eggs known to be more heat tolerant than those of the sympatric Qfly, surviving temperatures of 48.2°C [60]. Given that these species infest many of the same hosts, competition is likely, hence eradication of Ofly may result in the competitive release of B. jarvisi, increasing the threat it poses to horticulture [4, 60]. Further, as the cultivation of *B. jarvisi* host plants expands geographically, this species may increase in abundance and extend its range, potentially becoming a major pest in north-western Australia [6, 36]. While widespread throughout Oueensland, Z. cucumis currently has a restricted distribution in the Northern Territory, although there is a disputed single record from northern Western Australia [61]. Both Fitt [62] and the Horticultural Policy Council [4] reported that if the cucurbit industry expands in the Northern Territory, the pest status of *Z. cucumis* may increase. However, while the species has been trapped frequently in the Northern Territory, it has not been found on cucurbits growing in this region [6]. In NSW, Z. cucumis appears to be currently limited to regions close to the Queensland border, with rare detection as far south as Sydney [61] It has not been detected in the (former) Fruit Fly Exclusion Zone in southern NSW [37]. Our model also estimates the southern limit of suitable climate for this species to be around Sydney. However, with climate change this may extend further southward, with parts of

Victoria projected to become increasingly suitable over time, depending on the climate change scenario.

Bactrocera neohumeralis presently occurs from the western Cape York Peninsula, Queensland, south to Sydney, NSW [3, 7, 37]. Our models agree that suitable conditions occur from Cape York Peninsula to north of Sydney, although as climate changes, the range of this species may extend southward and, under some scenarios, into parts of Victoria. Previous climatic analysis also suggested that this species is well adapted to conditions on the east coast of Queensland, with large populations occurring in areas north of Townsville [4]. Similar ecological characteristics are shared by *B. neohumeralis* and Qfly [63], yet while Qfly is prevalent in subtropical and temperate areas of Queensland and NSW, *B. neohumeralis* is more prevalent in northern wet tropical areas [4, 5, 64]. The reason for this difference between the geographical ranges of these species is unclear, as both are polyphagous and use similar host fruits for their larval development [63, 64].

Our model for *B. aquilonis* indicates that suitable conditions for this species are currently found in northern Queensland, although it is presently only known from north-western Australia [5]. The hosts of this species now include 40 commercial crops [6]. Expansion of the range of this species, or the growth of host plant industries in north-western Australia may necessitate the development of new monitoring, control and disinfestation procedures [60]. In addition, it has been argued that if *B. aquilonis* hybridises with Qfly, and the resulting strain may have greater potential for spread than *B. aquilonis* [4]. This, in turn, would require that disinfestation procedures should be developed for the hybrids [60].

The Australian distribution of *B. bryoniae* ranges from the Torres Strait Islands, across northern

Australia, and along the east coast to north of Sydney, NSW. Our results indicate that suitable habitat exists in Victoria, i.e. south of the species' known range. However, previous studies have demonstrated that populations in northern NSW experience a marked decline in abundance through November–January [37]. This may be explained by a decline in the fruiting and flowering of native host trees, or seasonal climatic constraints that are not reflected in our model [37], which may also explain their absence in Victoria.

Northern Queensland has the highest diversity of fruit flies in Australia, and some species with significant economic impacts are found only in this region [7]. The distribution of *B. kraussi*, *B. musae* and *B. frauenfeldi* is limited to north Queensland [3, 65], with recent trap data suggesting that these species do not occur south of Townsville [7]. Royer et al. [35] predicted that *B. frauenfeldi* also has suitable habitat in the Northern Territory and northern Western Australia, which is also suggested by our model. The availability of hosts does not appear to limit the range of *B. frauenfeldi*, which has expanded in northern Queensland due to continued planting of hosts, such as mango and guava [35]. Further increases within these horticulture industries in northern Queensland may increase the pest status of this fly [65].

#### **Model Errors and Uncertainties**

SDMs are useful for developing a broad understanding of how the distribution of suitable habitat may be influenced by climate change. However, the output of SDMs is known to be influenced by characteristics of the occurrence sample, including its size [66], sampling bias [67], and spatial autocorrelation [68], as well as the extent of the study area, selection of predictor variables [69], and selection of background points [70]. We addressed these issues by: (1) exploring alternate settings in Maxent to optimise models and reduce overfitting that may generate unreliable estimates [47]; (3) reducing the number of predictor variables by

assessing collinearity; and (4) critically examining response curves.

In addition, we acknowledge that the selection of a threshold for converting Maxent's continuous output into binary data (typically defined as distinguishing between "suitable" and "unsuitable" conditions) can be subjective and problematic. A region classified as unsuitable may not be free of the pest; rather, these areas are considered less likely to support a population compared with regions above the threshold. In reality, the choice of threshold is based upon a comparison of the importance of false positives (FP) and false negatives (FN) [71]. For invasive species, the latter may be more serious because they can result in an underestimate of the geographic extent of suitable habitat, and hence, invasion risk [72]. This, in turn, can lead to poor decision-making and failure to establish appropriate surveillance or containment measures. Hence, in this context a precautionary approach to defining a threshold, as undertaken in the present study, is warranted. However, since overprediction of suitable habitat can also prove problematic (potentially leading to ineffective allocation of monitoring resources), we provide continuous (unthresholded) model output, permitting stakeholders to modify this threshold according to their objectives.

Sampling bias is another challenge faced when fitting correlative SDMs, particularly when incorporating data from sources of incidental observations such as museums and natural history collections [73]. As such, it is difficult to determine whether a species is observed in a particular environment because of habitat preferences or because that region has received the largest search effort [70, 73]. For presence-background approaches to habitat modelling, a target-group background sampling strategy goes some way to handling biased occurrence samples [74]. However, while imposing environmental bias on the background counteracts similar bias in the occurrence sample, this strategy may increase the extent of novel environments to which

the model must be extrapolated.

While SDMs consider exposure to climate change, species responses may also include microevolution [75] or plasticity [76]. As accessibility to genomic data increases, and experiments on plasticity are conducted, SDM output can be refined [77]. In addition, as mean conditions change, so too will the distribution and magnitude of extremes. Presently, there has been little work undertaken to assess how different fruit fly species tolerate extreme weather events such as heatwaves and moisture stress.

We also note that our analysis does not take into consideration the potential necessity for horticultural industries to shift geographically to adapt to climate change. Analysing potential shifts in climatic suitability for horticultural crops is complicated by our capacity to modify the environment (e.g. through irrigation), and thus was beyond the scope of this study.

## Conclusions

Surveillance activities, pre- and post-harvest treatment, and control activities for fruit flies present a substantial cost to Australia's horticultural industries [2, 4, 14]. Climate change is likely to alter the distribution of suitable habitat for these species, and this could have ramifications for Australia's horticultural industries. Our analysis highlights that the major horticultural production regions are likely to remain suitable for multiple economically important fruit fly species as climate changes. Furthermore, given that knowledge of current species distributions remains the basis for market access decisions, the potential for range shifts to occur is of critical interest to horticultural industries.

Our model projections identify geographic regions likely to be suitable for 11 high priority, economically important pest fruit fly species, now and in the future. Outputs from this study provide guidance to pest managers, such that they can assess pest risks and design appropriate ongoing surveillance strategies. Our results emphasize the importance of vigilance and preparedness across Australia, to prevent further range expansion of these 11 species, and underscore the need for ongoing research and development into monitoring, control, and eradication tools.

# **Supporting Information**

S1-11 Figs. Climatic habitat suitability for 11 tephritid fruit flies under various future climate scenarios. (1) Bactrocera aquilonis (2) Bactrocera bryoniae (3) Bactrocera frauenfeldi (4) Bactrocera halfordiae (5) Bactrocera jarvisi (6) Bactrocera kraussi (7) Bactrocera musae (8) Bactrocera neohumeralis (9) Bactrocera tryoni (10) Ceratitis capitata. (11) Zeugodacus cucumis. (A) current habitat suitability modelled using Maxent – values close to zero represent areas with low climatic suitability while values closer to one indicate higher climatic suitability; (B) areas considered "suitable" (i.e., with habitat suitability values above the 10th percentile at training presence sites, shown in red); (C, D, E) agreement about the suitability of habitat for the species across six climate scenarios for 2030, 2050 and 2070, respectively; (F) the location of Australian occurrence records of the species, based on specimens from natural history collections, literature and State Government trapping programs,

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and major commercial horticultural hosts, according to the Australian Horticulture Statistics Handbook (HSHB; www.horticulture.com.au). S12 Fig. Hotspot maps of habitat suitability for the 11 fruit fly species under climate change, when novel environments are excluded. Hotspot maps of current and future habitat suitability for 11 fruit flies with regions containing novel environments considered unsuitable. Suitability was modelled with Maxent, and thresholded using the 10th percentile at training presence localities. Colours indicate the number of species for which habitat is predicted to be suitable under the majority ( $\geq 4$ ) future climate scenarios. Figure was created in R version 3.3.3[1] (https://www.R-project.org/). S1 Table. Model performance and bioclimatic variables used to investigate the suitability of habitat for tephritid fruit fly species. AUC value indicates the area under the receiver operating characteristic curve (average of 5 cross-validated replicates), which was used to evaluate model performance; SD (standard deviation); and HPI (highest permutation importance, %) of bioclimatic variables contributing to the model where BIO01: annual mean temperature, BIO02: mean diurnal range; BIO03: isothermality; BIO06: minimum temperature of the coldest month; BIO07: temperature annual range; BIO11: mean temperature of the coldest quarter: BIO13: precipitation of wettest month: BIO14: precipitation of the driest month; BIO16: precipitation of the wettest quarter; BIO17: precipitation of the driest quarter and BIO19: precipitation of the coldest quarter. S2 Table. Projected changes in the area of suitable habitat for all 11 fruit fly species, under six future climate scenarios, relative to the current period. (1) Bactrocera aquilonis

(2) Bactrocera bryoniae (3) Bactrocera frauenfeldi (4) Bactrocera halfordiae (5) Bactrocera

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jarvisi (6) Bactrocera kraussi (7) Bactrocera musae (8) Bactrocera neohumeralis (9) Bactrocera tryoni (10) Ceratitis capitata (11) Zeugodacus cucumis. For each species, the first column indicates the GCM (Global Climate Model) for three time periods 2030, 2050 and 2070. Other columns: % Lost refers to the percentage of currently suitable habitat projected to become unsuitable in the future; % Gained refers to the percentage of future suitable habitat that is in areas currently unsuitable; Range Changed refers to the change (%) between the size of current and future suitable habitat (positive numbers indicate an increase in range size, negative numbers indicate a decrease). S3 Table. Area (km²) and percentage of Australia projected to be suitable for 11 fruit flies under six future climate scenarios. In the column 'Climate scenarios', 0 refers to the area projected to be unsuitable across all six scenarios; 1 refers to the area projected to be suitable under any one of the six scenarios...6 refers to the area projected to be suitable under all six scenarios. S4 Table. Major commercial fruits and vegetables host species to the Australian Horticulture Statistics Handbook (HSHB; www.horticulture.com.au). Pest status is based on Hancock et al<sup>3</sup>, where "major" indicates that there have been many records of the fly infesting that host. S5 Table. Area (km<sup>2</sup>) and percentage (%) of Australia projected to be suitable for the 11 fruit fly species considered in this study, when novel environments have been excluded. Each row of the table indicates the area (and percentage) projected to be suitable now, in 2030, 2050, and 2070, for *n* species, where *n* is given in the "Count" column. Thus, the first row (with Count = 0) gives the area projected to be unsuitable for all 11 species, the row with Count = 1

gives the area projected to be suitable for any one of the 11 species, and the row with Count =

11 gives the area projected to be suitable for all 11 species. Values shown here apply when

cells with novel environmental conditions are considered unsuitable.

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### Author contributors

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## References

- 778 1. Plant Health Australia. The Australian Handbook for the Identification of Fruit Flies.
- Version 3.1.: Plant Health Australia. Canberra, ACT.; 2018.
- 781 2. Plant Health Australia. Draft National Fruit Fly Strategy. Plant Health Australia.

782 DEAKIN, ACT, 2600; 2008.

Hancock DL, Hamacek EL, Lloyd AC, Elson-Harris MM. The distribution and host plants of fruit flies (Diptera: Tephritidae) in Australia: Department of Primary Industries, Queensland; 2000. 1-75 p.

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793

797

801

805

810

814

817

820

825

827

- Horticultural Policy Council. The impact of fruit flies on Australian horticulture: Report to the Minister for Primary Industries and Energy, Canberra; 1991.
- Drew RAI. The tropical fruit flies (Diptera: Tephritidae: Dacinae) of the Australasian
   and Oceanian regions. 1989. Report No.: 0079-8835.
- 5794 6. Smith ESC, Chin D, Allwood AJ, Collins SG. A revised host list of fruit flies
   795 (Diptera: Tephritidae) from the Northern Territory of Australia. Queensland Journal
   796 of Agricultural and Animal Sciences. 1988;45(1):19-28.
- 7. Royer JE, Hancock DL. New distribution and lure records of Dacinae (Diptera: Tephritidae) from Queensland, Australia, and description of a new species of *Dacus* Fabricius. Australian Journal of Entomology. 2012;51(4):239-47.
- 802 8. Sultana S, Baumgartner JB, Dominiak BC, Royer JE, Beaumont LJ. Potential impacts of climate change on habitat suitability for the Queensland fruit fly. Scientific Reports. 2017;7(1):13025.
- 9. Dominiak BC, Worsley P. Review of the southern boundary of Jarvis fruit fly 'Bactrocera jarvisi' (Tyron)(Diptera: Tephritidae: Dacinae) and its likley southern distribution in Australia. General and Applied Entomology: The Journal of the Entomological Society of New South Wales. 2017;45:1-7.
- De Meyer M, Copeland R, Wharton R, McPheron B. On the geographic origin of the Medfly *Ceratitis capitata* (Weidemann)(Diptera: Tephritidae). Proceedings of the 6th International Fruit Fly Symposium, Stellenbosch, South Africa. 2002:45-53.
- White IM, Elson-Harris MM. Fruit Flies of Economic Significance: Their Identification and Bionomics: CAB International; 1992.
- Permkam S, Hancock D. Australian Ceratitinae (Diptera: Tephritidae). Invertebrate Systematics. 1994;8(6):1325-41.
- Dominiak BC, Mapson R. Revised distribution of *Bactrocera tryoni* in eastern
  Australia and effect on possible incursions of Mediterranean fruit fly: development of
  Australia's eastern trading block. Journal of Economic Entomology. 2017;110(6):245965.
- Plant Health Australia. Prevent fruit fly. Fruit Fly Research. 2016.
- Abdalla A, Millist N, Buetre B, Bowen B. Benefit-cost analysis of the national fruit fly strategy action plan. Australian Bureau of Agricultural and Resource Economics and Sciences, Canberra; 2012.
- 832 16. PHA. Economic assessment of the implementation of the proposed National Fruit Fly

Strategy: Part 1 (Plant Health Australia, August, Canberra., 2009).

35

834

837

841

844

847

852

856

861

864

869

873

- Australian Pesticides and Veterinary Medicines Authority. Dimethoate Residues and Dietary Risk Assessment Report. August 2011. 2011.
- Clarke AR, Powell KS, Weldon CW, Taylor PW. The ecology of *Bactrocera tryoni*(Diptera: Tephritidae): what do we know to assist pest management? Annals of
  Applied Biology. 2011;158(1):26-54.
- Australian Pesticides and Veterinary Medicines Authority. Fenthion Residues and Dietary Risk Assessment Report. September,2012.
- Dominiak BC, Ekman JH. The rise and demise of control options for fruit fly in Australia. Crop Protection. 2013;51:57-67.
- Jessup A, Dalton S, Slogget R. Determination of host status of table grapes to
  Queensland fruit fly,'*Bactrocera tryoni*'(Froggatt)(Diptera: Tephritidae), for export to
  New Zealand. General and Applied Entomology: The Journal of the Entomological
  Society of New South Wales. 1998;28:73.
- Stephens AEA, Stringer LD, Suckling DM. Advance, retreat, resettle? Climate change could produce a zero-sum game for invasive species. Austral Entomology. 2016;55:177-84.
- Hill MP, Bertelsmeier C, Clusella-Trullas S, Garnas J, Robertson MP, Terblanche JS.
  Predicted decrease in global climate suitability masks regional complexity of invasive fruit fly species response to climate change. Biological Invasions. 2016;18(4):1105-19.
- Kriticos D. Risks of establishment of fruit flies in New Zealand under climate change.
  New Zealand Forest Research Institute, Rotorua; 2007.
- Vera MT, Rodriguez R, Segura DF, Cladera JL, Sutherst RW. Potential geographical distribution of the Mediterranean fruit fly, *Ceratitis capitata* (Diptera: Tephritidae), with emphasis on Argentina and Australia. Environmental Entomology. 2002;31(6):1009-22.
- Aguilar G, Blanchon D, Foote H, Pollonais C, Mosee A. Queensland fruit fly invasion of New Zealand: predicting area suitability under future climate change scenarios.
   Perspectives in Biosecurity Research Series 2015;2:1-13.
- Phillips SJ, Anderson RP, Schapire RE. Maximum entropy modeling of species geographic distributions. Ecological Modelling. 2006;190(3):231-59.
- Kumar S, Neven LG, Yee WL. Evaluating correlative and mechanistic niche models for assessing the risk of pest establishment. Ecosphere. 2014;5(7):1-23.
- Kumar S, Neven LG, Yee WL. Assessing the potential for establishment of Western Cherry Fruit Fly using ecological niche modeling. Journal of Economic Entomology. 2014;107(3):1032-44.

Kumar S, Neven LG, Zhu H, Zhang R. Assessing the global risk of establishment of *Cydia pomonella* (Lepidoptera: Tortricidae) using CLIMEX and MaxEnt niche models. Journal of Economic Entomology. 2015;108(4):1708-19.

36

883

887

891

894

899

903

907

911

914

918

921

926

- 888 31. Kumar S, Yee WL, Neven LG. Mapping global potential risk of establishment of *Rhagoletis pomonella* (Diptera: Tephritidae) using MaxEnt and CLIMEX niche models. Journal of Economic Entomology. 2016;109(5):2043-53.
- The Australian horticulture statistics handbook. Horticulture Innovation Australian Limited 2016/17.
- B95 33. Dominiak BC, Daniels D. Review of the past and present distribution of
   B96 Mediterranean fruit fly (*Ceratitis capitata* Wiedemann) and Queensland fruit fly
   B97 (*Bactrocera tryoni* Froggatt) in Australia. Australian Journal of Entomology.
   B98 2012;51(2):104-15.
- 900 34. Dominiak BC. Review of grapes Vitis sp. as an occasional host for Queensland fruit fly Bactrocera. tryoni (Froggatt)(Diptera: Tephritidae). Crop Protection. 2011;30(8):958-61.
- Royer JE, Wright CL, Hancock DL. *Bactrocera frauenfeldi* (Diptera: Tephritidae), an invasive fruit fly in Australia that may have reached the extent of its spread due to environmental variables. Austral Entomology. 2016;55(1):100-11.
- 908 36. May AWS. An investigation of fruit flies (Trypertidae: Diptera) in Queensland 1.
  909 Introduction, species, pest status and distribution. Queensland Journal of Agricultural
  910 Science. 1963;20:1-82.
- 912 37. Gillespie P. Observations on fruit flies (Diptera :Tephritidae) in New South Wales. 913 Gen Appl Ent Vol. 2003;32:41-8.
- Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A. Very high resolution
   interpolated climate surfaces for global land areas. International Journal of
   Climatology. 2005;25(15):1965-78.
- 919 39. Bateman MA. The ecology of fruit flies. Annual Review of Entomology. 1972;17(1):493-518.
- 922 40. De Meyer M, Robertson MP, Mansell MW, Ekesi S, Tsuruta K, Mwaiko W, et al. Ecological niche and potential geographic distribution of the invasive fruit fly *Bactrocera invadens* (Diptera, Tephritidae). Bulletin of Entomological Research. 2010;100(01):35-48.
- 927 41. Beaumont LJ, Graham E, Duursma DE, Wilson PD, Cabrelli A, Baumgartner JB, et 928 al. Which species distribution models are more (or less) likely to project broad-scale, 929 climate-induced shifts in species ranges? Ecological Modelling. 2016;342:135-46.
- 42. CSIRO & BoM. Climate change in Australia information for Australia's natural
   resource management regions. Technical Report, CSIRO and Bureau of Meteorology,

933 Australia.; 2015.

37

934

938

941

944

948

952

955

959

962

966

969

972

974

977

- 935 43. Moss RH, Edmonds JA, Hibbard KA, Manning MR, Rose SK, Van Vuuren DP, et al. The next generation of scenarios for climate change research and assessment. Nature. 2010;463(7282):747-56.
- 939 44. Greenberg J, Mattiuzzi M. gdalUtils: Wrappers for the Geospatial Data Abstraction 940 Library (GDAL) Utilities. R Package Version 2017. 2015.
- 942 45. R Core Team. R: A language and environment for statistical computing: R Foundation for Statistical Computing. Vienna, Austria; 2017.
- Elith J, Graham CH, Anderson RP, Dudík M, Ferrier S, Guisan A, et al. Novel
   methods improve prediction of species' distributions from occurrence data.
   Ecography. 2006;29:129-51.
- 949 47. Merow C, Smith MJ, Silander JA. A practical guide to MaxEnt for modeling species' distributions: what it does, and why inputs and settings matter. Ecography. 2013;36(10):1058-69.
- 953 48. Elith J, Phillips SJ, Hastie T, Dudík M, Chee YE, Yates CJ. A statistical explanation of MaxEnt for ecologists. Diversity and Distributions. 2011;17(1):43-57.
- Hlow F, Dambach J, Engler JO, Flecks M, Hartmann T, Nekum S, et al. On the brink of extinction? How climate change may affect global chelonian species richness and distribution. Global Change Biology. 2012;18(5):1520-30.
- 960 50. Swets JA. Measuring the accuracy of diagnostic systems. Science. 1988;240(4857):1285.
- 963 51. Wilson KA, Westphal MI, Possingham HP, Elith J. Sensitivity of conservation 964 planning to different approaches to using predicted species distribution data. 965 Biological Conservation. 2005;122(1):99-112.
- 967 52. Elith J, Kearney M, Phillips S. The art of modelling range-shifting species. Methods in Ecology and Evolution. 2010;1(4):330-42.
- 970 53. R Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria2014.
- 973 54. Pebesma EJRSB. Classes and methods for spatial data in R. R News 5 (2). 2005.
- 975 55. Hijmans RJ. Raster: Geographic Data Analysis and Modeling. R Package Version 2.4-15. http://CRAN.R-project.org/package=raster. 2015.
- 978 56. O'Loughlin GT, East RA, Meats A. Survival, development rates and generation times of the Queensland fruit fly, *Dacus tryoni*, in a marginally favourable climate: experiments in Victoria. Australian Journal of Zoology. 1984;32(3):353-61.
- 982 57. Muthuthantri S, Maelzer D, Zalucki MP, Clarke AR. The seasonal phenology of

983 *Bactrocera tryoni* (Froggatt)(Diptera: Tephritidae) in Queensland. Austral Entomology. 2010;49(3):221-33.

38

985

989

992

995

999

1003

1006

1010

1013

1017

1021

1024

- Sutherst RW, Collyer BS, Yonow T. The vulnerability of Australian horticulture to
   the Queensland fruit fly, *Bactrocera (Dacus) tryoni*, under climate change. Australian
   Journal of Agricultural Research. 2000;51(4):467-80.
- Allwood A, Angeles T. Host records of fruit flies (family Tephritidae) in the Northern
   Territory. Queensland Journal of Agricultural and Animal Sciences. 1979.
- 993 60. Cameron EC. Fruit Fly Pests of Northwestern Australia [Thesis]: University of Sydney.; 2006.
- 996 61. Dominiak BC, Worsley P. Review of cucumber fruit fly, *Bactrocera cucumis* 997 (French)(Diptera: Tephritidae: Dacinae) in Australia: Part 1, host range, surveillance 998 and distribution. Crop Protection. 2018;106:79-85.
- Fitt GP. New records of *Dacus (Austrodacus) cucumis* French from the Northern Territory, Australia (Diptera: Tephritidae). Australian Journal of Entomology. 1980;19(3):240.
- Gibbs G. The comparative ecology of two closely related, sympatric species of Dacus (Diptera) in Queensland. Australian Journal of Zoology. 1967;15(6):1123-39.
- Wang Y, Yu H, Raphael K, Gilchrist A. Genetic delineation of sibling species of the pest fruit fly *Bactocera* (Diptera: Tephritidae) using microsatellites. Bulletin of Entomological Research. 2003;93(4):351-60.
- Drew R, Hooper G, Bateman M. Economic fruit flies of the South Pacific Region.
   Economic fruit flies of the South Pacific Region. 1978.
- 1014 66. Wisz MS, Hijmans R, Li J, Peterson AT, Graham C, Guisan A, et al. Effects of
   1015 sample size on the performance of species distribution models. Diversity and
   1016 distributions. 2008;14(5):763-73.
- 1018 67. Syfert MM, Smith MJ, Coomes DA. The effects of sampling bias and model complexity on the predictive performance of MaxEnt species distribution models. PLoS ONE. 2013;8(2):e55158.
- Veloz SD. Spatially autocorrelated sampling falsely inflates measures of accuracy for presence-only niche models. Journal of Biogeography. 2009;36(12):2290-9.
- Guillén D, Sánchez R. Expansion of the national fruit fly control programme in
   Argentina. Area-wide Control of Insect Pests. 2007:653-60.
- Phillips SJ. Transferability, sample selection bias and background data in presenceonly modelling: a response to Peterson et al.(2007). Ecography. 2008;31(2):272-8.
- Franklin J. Mapping species distributions: spatial inference and prediction: Cambridge University Press; 2010.

Pheloung P, Williams P, Halloy S. A weed risk assessment model for use as a biosecurity tool evaluating plant introductions. Journal of Environmental Management. 1999;57(4):239-51.

- Newbold T, Reader T, El-Gabbas A, Berg W, Shohdi WM, Zalat S, et al. Testing the accuracy of species distribution models using species records from a new field survey. Oikos. 2010;119(8):1326-34.
- 1042 74. Elith J. Predicting distributions of invasive species. 2015:1-28.

39

1033

1037

1041

1043

1047

1051

- Salamin N, Wüest RO, Lavergne S, Thuiller W, Pearman PB. Assessing rapid
   evolution in a changing environment. Trends in Ecology & Evolution.
   2010;25(12):692-8.
- 1048 76. Charmantier A, McCleery RH, Cole LR, Perrins C, Kruuk LE, Sheldon BC. Adaptive phenotypic plasticity in response to climate change in a wild bird population. Science. 2008;320(5877):800-3.
- Bush A, Mokany K, Catullo R, Hoffmann A, Kellermann V, Sgrò C, et al.
   Incorporating evolutionary adaptation in species distribution modelling reduces projected vulnerability to climate change. Ecology Letters. 2016;19(12):1468-78.

