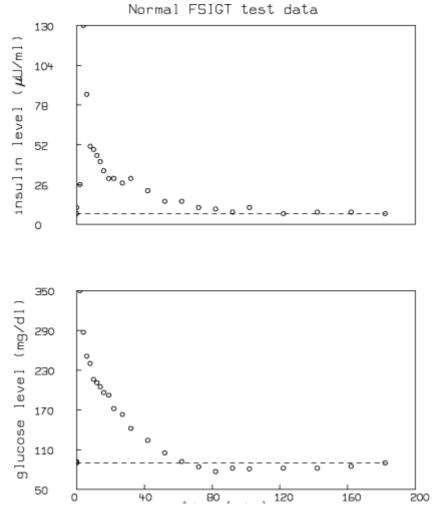
Minimal Models for Glucose and Insulin Kinetics

Minimal models of glucose and insulin plasma levels in dogs and humans during frequently-sampled intravenous glucose tolerance (FSIGT) tests have been developed and used by Dr. Richard N. Bergman and co-workers since the 1970's. (See references 1-5.) In a typical FSIGT test, blood samples are taken from a fasting subject at regular intervals of time, following a single intravenous injection of glucose. The blood samples are then analyzed for glucose and insulin content. The figure below shows a typical response from a normal subject.



Qualitatively, the glucose level in plasma starts at a peak due to the injection, drops to a minimum which is below the basal (pre-injection) glucose level, and then gradually returns to the basal level. The insulin level in plasma rapidly rises to a peak immediately after the injection, drops to a lower level which is still above the basal insulin level, rises again to a lesser peak, and then gradually drops to the basal level. Depending on the state of the subject, there can be wide variations from this response; for example, the glucose level may not drop below basal level, the first peak in insulin level may have different amplitude, there may be no secondary peak in insulin level, or there may be more than two peaks in insulin level.

The glucose and insulin minimal models provide a quantitative and parsimonious description of glucose and insulin concentrations in the blood samples following the glucose injection. The glucose minimal model involves two physiologic compartments: a plasma compartment and an interstitial tissue compartment; the insulin minimal model involves only a single plasma compartment. The glucose and insulin minimal models allow us to characterize the FSIGT test data in terms of four metabolic indices:

• S_I = insulin sensitivity: a measure of the dependence of fractional glucose disappearance on plasma insulin,

- S_G = glucose effectiveness: a measure of the fractional ability of glucose to lower its own concentration in plasma independent of increased insulin,
- f_1 = first phase pancreatic responsivity: a measure of the size of the first peak in plasma insulin due to the glucose injection, and
- f₂ = second phase pancreatic responsivity: a measure of the size of the second peak of plasma insulin which follows the first peak and the refractory period.

This paper will demonstrate the use of the mathematical modeling computer program MLAB to simulate insulin and glucose plasma levels during an FSIGT test and determine values of the metabolic indices from a data set via curve-fitting.

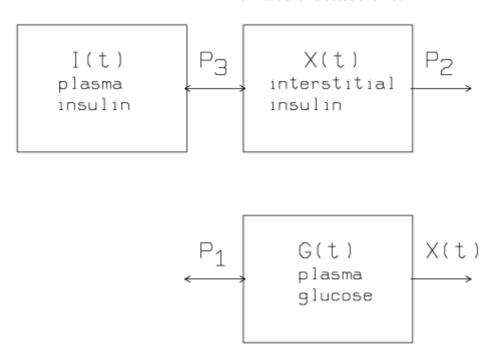
Reference 3 provides the following FSIGT test data (also shown in the graphs above) from a normal individual:

time (minutes) glucose level	(mg/dl) insulin level ((mU/mI)
------------------------------	-------------------------	---------

0	92	11
2	350	26
4	287	130
6	251	85
8	240	51
10	216	49
12	211	45
14	205	41
16	196	35
19	192	30
22	172	30
27	163	27
32	142	30
42	124	22
52	105	15
62	92	15
72	84	11
82	77	10
92	82	8
102	81	11
122	82	7
142	82	8
162	85	8
182	90	7

Using a spreadsheet program, word processor, or the MLAB file edit command directly, these numbers can be stored in an ASCII text file named "TEST.DAT".

The following diagram summarizes the minimal model for glucose kinetics:



Insulin leaves or enters the interstitial tissue compartment at a rate proportional to the difference between the plasma insulin level, I(t), and the basal level, I_b ; if the plasma insulin level falls below the basal level, insulin leaves the interstitial tissue compartment, and if the plasma insulin level rises above the basal level, insulin enters the interstitial tissue compartment. Insulin also disappears from the interstitial tissue compartment via a second pathway at a rate proportional to the amount of insulin in the interstitial tissue compartment.

Glucose leaves or enters the plasma compartment at a rate proportional to the difference between the plasma glucose level, G(t), and the basal level, G_b ; if the plasma glucose level falls below the basal level, glucose enters the plasma compartment, and if the glucose level rises above the basal level, glucose leaves the plasma compartment. Glucose also disappears from the plasma compartment via a second pathway at a rate proportional to the amount of insulin in the interstitial tissue.

The differential equations corresponding to the glucose minimal model are:

$$\frac{dX(t)}{dt} = -p_2 \cdot X(t) + p_3 \cdot (I(t) - I_b), \text{ and}$$

$$\frac{dG(t)}{dt} = -X(t) \cdot G(t) + p_1 \cdot (G_b - G(t))$$

with $G(0) = G_0$ and X(0) = 0. In these equations, \boldsymbol{t} is time, G(t) is the plasma glucose concentration at time \boldsymbol{t} , I(t) is the plasma insulin concentration at time \boldsymbol{t} , and X(t) is the interstitial insulin at time \boldsymbol{t} . G_b is the basal plasma glucose concentration and I_b is the basal plasma insulin concentration. Basal plasma concentrations of glucose and insulin are typically measured either before, or 180 minutes after, administration of glucose. There are four unknown parameters in this model: p_1 , p_2 , p_3 , and G_0 .

Note that in this model, glucose is utilized at the constant rate p_1 , when we neglect *feedback* effects due to interstitial insulin as represented by the term $-X(t) \cdot G(t)$. An additional amount of plasma insulin will cause the amount of interstitial insulin to change, which in turn, will cause the rate of glucose utilization to change. The *insulin sensitivity* is defined as $S_I = p_3/p_2$ and the *glucose effectiveness* is defined as $S_G = p_1$.

The following MLAB commands in the do-file "g.do" estimate values for the parameters p_1 , p_2 , p_3 , and G_0 given the time course of plasma glucose and insulin. The values of the parameters found minimize, in the least squares sense, the weighted difference between the measured time course of plasma glucose and the parameter-dependent solution to the glucose minimal model differential equations. The plasma insulin concentration function is obtained by linear interpolation of the time-insulin values listed in TEST.DAT. This is done by employing the MLAB function "LOOKUP".

```
"file: g.do = glucose minimal model"
"get data consisting of time values in column 1, plasma glucose"
"levels in column 2, and plasma insulin levels in column 3. Set" "n to the number of time values. Set gdat to the (time,glucose level)"
"ordered pairs. Set idat to the (time,insulin level) ordered pairs.'
data = read(test, 50, 3);
m = nrows(data);
gdat = data col (1,2);
idat = data col (1,3);
"define the glucose minimal model:"
" t is time'
  g(t) is plasma glucose level"
  i(t) is plasma insulin level, empirically-defined by interpolation in idat"
" x(t) is interstitial insulin"
  gb is basal (180 min) plasma glucose level"
" ib is basal (180 min) plasma insulin level"
fct g't(t) = -(p1+x(t))*g(t)+p1*gb
fct x't(t) = -p2*x(t)+p3*(i(t)-ib)
fct i(t) = lookup(idat,t)
init g(0) = g0
init x(0) = 0.0
gb = gdat(m,2) ib =
idat(m,2)
"define weights for glucose level data, censoring data up to time t = 8"
fct wf(i) = if gdat(i,1) < 8 then 0 else if gdat(i,1) = 8 then 10 else 1
wts = wf on 1:m
"define constraints for p1, p2, p3, and g0"
constraints q = \{p1>0, p2>0, p3>0, g0>0\}
"give initial estimates for parameters p1,p2,p3, and g0"
p1 = .03082; p2 = .02093; p3 = .00001062; g0 = 287;
"fit the model to the weighted data with defined constraints"
fit (p1,p2,p3,g0), g to gdat with weight wts constraints q
type "glucose effectiveness",p1
type "insulin sensitivity",p3/p2
"draw 3 graphs:1-plasma insulin level function and data vs time"
                2-interstitial insulin vs time"
                 3-fitted glucose level function and data vs time
"horizontal dashed lines show basal levels"
top title "GLUCOSE MINIMAL MODEL"
draw idat pt circle ptsize .01
draw shape(2,2,list(idat(1,1),ib,idat(m,1),ib)) lt dashed pt circle ptsize .01
left title " insulin level ('15Tm'RU/ml)"
delete w.xaxis
frame .25 to .75, .667 to 1
w1 = w
draw points(x,gdat(1,1):gdat(m,1)!200)
left title interstitial insulin"
delete w.xaxis
frame .25 to .75, .334 to .666
w2 = w
draw points(g,gdat(1,1):gdat(m,1)!200)
```

draw gdat lt none pt circle ptsize .01
draw shape(2,2,list(gdat(1,1),gb,gdat(m,1),gb)) lt dashed pt circle ptsize .01
left title "glucose level (mg/dl)"
bottom title "time v(min)"
frame .25 to .75, 0 to .333
view

save idat,gdat,ib,gb,g0,m in tmp.sav

The MLAB log-file and picture generated by executing the do-file "g.do" follow:

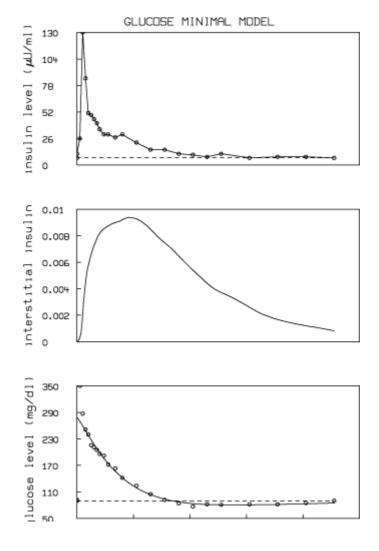
* do "q.do" final parameter values value dependency parameter error Ρ1 0.02649256302 0.01367779755 0.9976038298 P2 0.02543609572 0.029223424 0.9932825935 **P3** 1.281692067e-05 1.516217536e-05 0.9978008452 279.1123014 15.38803832 0.9901777503 G05 iterations **CONVERGED** best weighted sum of squares = 4.237885e+02weighted root mean square error = 4.603197e+00 weighted deviation fraction = 1.469631e-02

glucose effectiveness
P1 = .026492563

R squared = 6.905055e-01 no active constraints

insulin sensitivity
= 5.03887106E-4

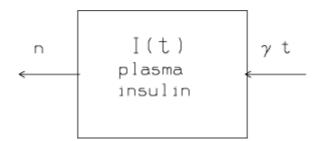
creating save file: TMP.SAV



The insulin sensitivity, S_I , for this data set is estimated as $5.039 \times 10^{-4} \, \text{min}^{-1} \cdot (\text{mU/ml})^{-1}$ which is within the normal range reported in reference 2: 2.1 to $18.2 \times 10^{-4} \, \text{min}^{-1} \cdot (\text{mU/ml})^{-1}$. The glucose utilization, S_G , for this data set is estimated as $0.0265 \, \text{min}^{-1}$, which is also within the normal range reported in reference 2: $0.0026 \, \text{to} \, 0.039 \, \text{min}^{-1}$.

Next we examine the minimal model for insulin kinetics.

The following diagram summarizes the minimal model for insulin kinetics:



The course of plasma glucose in time, G(t), is given by linear interpolation of the time-glucose values listed in TEST.DAT. Insulin enters the plasma insulin compartment at a rate proportional to the product of time and the concentration of glucose above a threshold amount. Here, time is the interval, in minutes, from the glucose injection. If the plasma glucose level drops below the threshold amount, the rate of insulin entering the plasma compartment is zero. Insulin is cleared from the plasma compartment at a rate proportional to the amount of insulin in the plasma compartment.

The minimal model for insulin kinetics is given by the equation:

$$\frac{dI(t)}{dt} = \hat{i} - n \cdot I(t) + g \cdot (G(t) - h) \cdot t, \text{ if } G(t) > h$$

$$\frac{dI(t)}{dt} = \hat{i} - n \cdot I(t), \text{ otherwise.}$$

with $I(0) = I_0$. n is the insulin clearance fraction, h is roughly the basal glucose plasma level, and g is a measure of the secondary pancreatic response to glucose. The *first phase pancreatic responsivity* is defined as $f_1 = (I_{max} - I_b)/[n \cdot (G_0 - G_b)]$ where I_{max} is the maximum insulin response. The *second phase pancreatic responsivity* is defined as $f_2 = g \times 10^4$.

The following MLAB commands in the do-file GI.DO estimate values for the parameters n, g, h, and I_0 given the time course of plasma glucose. The values of the parameters found minimize (in the weighted least squares sense) the difference between the measured time course of plasma insulin and the parameter-dependent solution to the insulin minimal model differential equations.

```
"file: gi.do = insulin minimal model"
reset

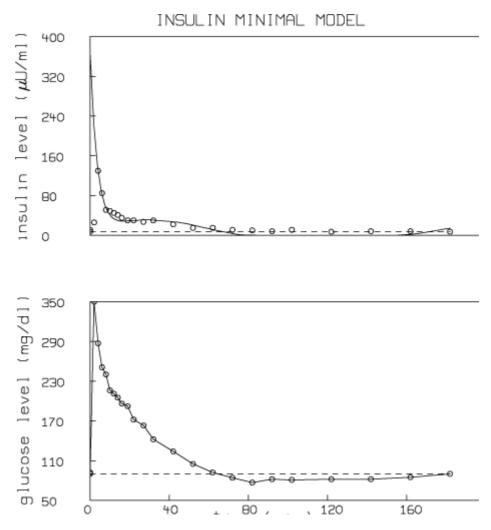
"read insulin, glucose levels from temporary save file"
use tmp.sav

"define the insulin minimal model"
fct i't(t) = -n*i+gama*(if g(t) < h then 0 else (g(t)-h))*t
fct g(t) = lookup(gdat,t)
init i(0) = i0

"determine weights for insulin data"
fct wf(i) = if idat(i,1) < 3 then 0 else if idat(i,1) <= 8 then 10 else 1
wts = wf on 1:m

"define a constraint set for all the parameters"
constraints g1 = {n > 0, gama > 0, h > 0, i0 > 0}
```

```
"provide initial guesses for the paramters"
n = .3; gama = .003349; h = 89.50; i0 = 410.4
"fit the insulin minimal model to the insulin data"
DISASTERSW = -1
fit (n,gama,h,i0), i to idat with weight wts constraints q1
type "phase 1 pancreas responsivity",(maxv(idat)-ib)/(n*(g0-gb)) type
'phase 2 pancreas responsivity",10000*gama
"draw 2 graphs:1-fitted plasma insulin level and data vs time"
      2-glucose plasma level data vs time"
top title "INSULIN MINIMAL MODEL"
draw idat lt none pt circle ptsize .01
draw points(i,idat(1,1):idat(m,1)!100)
draw shape(2,2,list(idat(1,1),ib,idat(m,1),ib)) lt dashed pt circle ptsize .01
left title " insulin level ('15Tm'RU/ml)"
delete w.xaxis
frame .15 to .85, .5 to 1
w1 = w
draw gdat pt circle ptsize .01
draw shape(2,2,list(gdat(1,1),gb,gdat(m,1),gb)) lt dashed pt circle ptsize .01
left title "glucose level (mg/dl)'
bottom title "time v(min)
frame .15 to .85, 0 to .5
view
The log-file and picture generated by executing the do-file GI.DO follow:
* do "gi.do"
Using: M,GO,GB,IB,GDAT,IDAT
final parameter values
    value
                     error
                                  dependency
                                                 parameter
                0.01603033225
0.2673230345
                                 0.9611152205
                                                     N
0.004074459794
                0.0006378976592 0.7211259427
                                                   GAMA
83.74403736
                2.329398193
                                 0.2073681454
                                                     Н
363.666326
                26.5919455
                                 0.9471979611
                                                    Ι0
4 iterations
CONVERGED
best weighted sum of squares = 1.292680e+03
weighted root mean square error = 8.039529e+00
weighted deviation fraction = 2.772192e-02
no active constraints
        phase 1 pancreas responsivity
       = 3.46163989
        phase 2 pancreas responsivity
       = 40.7445979
```



The phase 1 pancreas responsivity, f_1 , is estimated as 3.462 min·(mU/ml)(mg/dl)⁻¹ for this data set. This is within the normal range for f_1 reported in reference 3 as 2.0 to 4.0. The phase 2 pancreas responsivity, f_2 , is estimated as 40.745 min⁻²·(mU/ml)(mg/dl)⁻¹. This is slightly higher than the normal range for f_2 reported in reference 3 as 20 to 35.

Note that when performing the least squares minimization between the solution of the insulin minimal model equation and the measured plasma insulin levels, relative weights of 0, 10, and 1, were assigned to the plasma insulin levels so that less reliable values would not adversely effect the estimation of parameters and more reliable values would be more heavily weighted. Using appropriate weights is generally important in fitting forms of the minimal model. While there are many ways of weighing data, this particular method was suggested by Walton (reference 6).

The glucose minimal model provides differential equations for the plasma glucose and interstitial insulin levels. The insulin minimal model provides a differential equation for the plasma insulin level. It is possible to combine all three differential equations into one model. This is demonstrated in the following do-file, GGI.DO:

```
"file: ggi.do - combined glucose and insulin minimal models" read insulin, glucose levels from temporary save file" use tmp.sav

"define the combined glucose-insulin minimal model" fct i't(t) = -n*i+gama*(if g < h then 0 else (g-h))*t fct g't(t) = -(p1+x)*g+p1*gb fct x't(t) = -p2*x+p3*(i-ib) init i(0) = i0 init g(0) = g0 init x(0) = 0
```

"determine weights for glucose level data, censoring data up to time t = 8"

364.8353065

291.2242018

5 iterations CONVERGED

 $R \ squared = 3.943754e-01$ no active constraints

```
06/04/2024, 04:27
                                             Minimal Models for Glucose and Insulin
 fct wg(i) = if gdat(i,1) < 8 then 0 else if gdat(i,1) = 8 then 10 else 1
 wqs = wq on 1:m
 "determine weights for insulin level data"
 fct wi(i) = if idat(i,1) < 3 then 0 else if idat(i,1) <= 8 then 10 else 1
 wis = wi on 1:m
 "define a constraint set for all of the parameters"
 constraints q1 = \{n>0, gama>0, h>0, p1>0, p2>0, p3>0, i0>0, g0>0\}
 "provide initial guesses for the parameters"
 n = .3; gama = .003349; h = 89.50; p1 = .03082; p2 = .02093
 p3 = .00001062; i0 = 403.4; g0 = 287
 "fit both of the models to the data"
 disastersw = -1
 fit (n,gama,h,p1,p2,p3,i0,g0), i to idat with weight wis, \
    g to gdat with weight wgs, constraints q1
 type "glucose effectiveness",p1
 type "insulin sensitivity",p3/p2
 type "phase 1 pancreas responsivity", (\max(idat)-ib)/(n*(g0-gb)) type "phase 2 pancreas responsivity", 10000*gama
 Here is the resulting MLAB log-file and picture:
 final parameter values
     value
                        error
                                     dependency
                                                      parameter
 0.2658844452
                  0.01153178897
                                    0.9621356273
                                                          N
 0.003911687955 0.0004693022146 0.7457248671
                                                        GAMA
 79.03532257
                  2.421480638
                                    0.4908060854
                                                          Н
                                                         Ρ1
 0.03168360775
                  0.004806175221
                                    0.9712465296
                                                         P2
 0.0123362991
                  0.006558350386
                                    0.888606309
```

0.9496151759

0.8836723395

Р3

Ι0

G0

4.891692162e-06 1.923309135e-06 0.9627481277

18.88965714

5.82052435

best weighted sum of squares = 1.278572e+03 weighted root mean square error = 5.653697e+00 weighted deviation fraction = 1.545963e-02

phase 1 pancreas responsivity

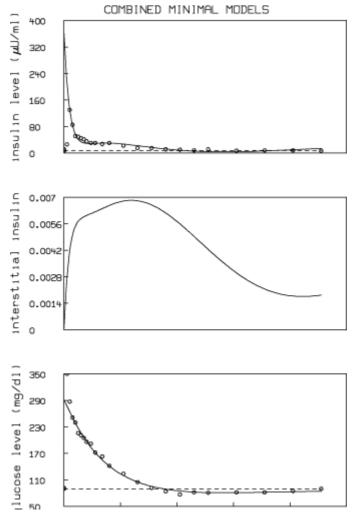
phase 2 pancreas responsivity

glucose effectiveness P1 = 3.16836078E-2

insulin sensitivity = 3.96528337E-4

= 3.27088221

= 39.1168796



These results show that, as is common, there is not a unique set of parameters that characterize a FSIGT test data set. The combined minimal model is seen to generate slightly lower values for glucose effectiveness and insulin sensitivity than the glucose minimal model, and slightly higher values for phase 1 and 2 pancreas responsivity than the insulin minimal model.

There are various devices that could be explored in order to improve the family of models studied here. First, these models employed the MLAB operator "LOOKUP" to linearly interpolate glucose and insulin plasma time course data. Alternatively, the MLAB operator "SMOOTHSPLINE" could be used to provide glucose and insulin plasma time course curves that are not only continuous, but also have continuous first and second derivatives.

Second, several authors have augmented the insulin minimum model to account for plasma levels of C-peptide (see references 7-9). It is a straightforward exercise to implement the C-Peptide minimal model using MLAB.

This paper has shown how MLAB can be used to calculate diagnostically important metabolic indices which arise in the glucose and insulin minimal models from frequently-sampled intravenous glucose tolerance test data. The MLAB program is generally an ideal tool for the study of compartmental models. You can contact Civilized Software for further examples in neurophysiology and pharmacology.

References:

- 1. Saad, M.F., Anderson, R.L., Laws, A., Watanabe, R.M., Kades, W.W., Chen, Y.-D.I., Sands, R.E., Pei, D., Savage, P.J., Bergman, R.N., *A Comparison Between the Minimal Model and the Glucose Clamp in the Assessment of Insulin Sensitivity Across the Spectrum of Glucose Tolerance* **Diabetes** 43 (1994) pp. 1114-21.
- 2. Steil, G.M., Volund, A., Kahn, S.E., Bergman, R.N.: *Reduced Sample Number for Calculation of Insulin Sensitivity and Glucose Effectiveness From the Minimal Model*, **Diabetes** 42 (1993) pp. 250-256.

- 3. Pacini, G., Bergman, R.N.: MINMOD: A Computer Program to Calculate Insulin Sensitivity and Pancreatic Responsivity from the Frequently Sampled Intravenous Glucose Tolerance Test, Computer Methods and Programs in Biomedicine 23 (1986) pp. 113-22.
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- 5. Bergman, R.N., Ider, Y.Z., Bowden, C.R., Cobelli, C.: *Quantitative Estimation of Insulin Sensitivity*, **American Journal of Physiology** 236 (1979) pp. E667-77.
- 6. Walton, C.W., Wynn Department of Metabolic Medicine, Imperial College of Science, Technology, and Medicine, *private communication*, April 1998.
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- 8. Cobelli, C., Pacini, G.: *Insulin Secretion and Hepatic Extraction in Humans by Minimal Modeling of C-Peptide and Insulin Kinetics*, **Diabetes** 37 (1988) 223-231.
- 9. Volund, A., Polonsky, K.S., Bergman, R.N.: *Calculated Pattern of Intraportal Insulin Appearance Without Independent Assessment of C-Peptide Kinetics*, **Diabetes** 36 (1987) 1195-1202.