

Supplemental Information
Cell Metabolism, *Volume 14*

**Calorie Restriction-like Effects of 30 Days of Resveratrol
Supplementation on Energy Metabolism and Metabolic
Profile in Obese Humans**

Silvie Timmers, Ellen Konings, Lena Bilet, Riekelt H. Houtkooper, Tineke van de Weijer, Gijs H. Goossens, Joris Hoeks, Sophie van der Krieken, Dongryeol Ryu, Sander Kersten, Esther Moonen-Kornips, Matthijs K.C. Hesselink, Iris Kunz, Vera B. Schrauwen-Hinderling, Ellen Blaak, Johan Auwerx, and Patrick Schrauwen

Inventory

Figure S1 corresponds to figure 3

Table S1 corresponds to table 2

Table S2 corresponds to table 3

Table S3 corresponds to figure 2

Table S4 does not correspond to a figure/table but is a summary of our findings and adds to the comprehension of the conclusion of the manuscript.

FIGURE S1

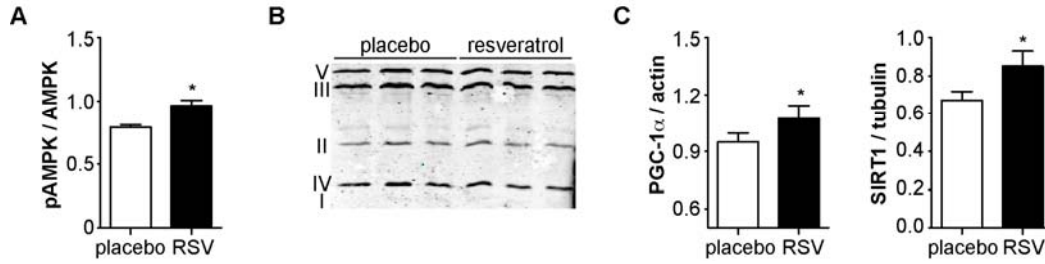


Figure S1. Western Blot Analysis

(A) Quantitative analysis of the effect of resveratrol (RSV) on the phosphorylation of AMPK on Thr¹⁷² of the α -subunit in vastus lateralis muscle. Data are presented as the ratio of pAMPK/AMPK (n=9).

(B) Representative western blot for the individual complexes of the electron transport chain (n=11, a subset of 3 subjects is shown).

(C) Quantitative analysis of the western blots of PGC-1 α and SIRT1. Data are normalized for the loading control, actin for PGC-1 α and tubulin for SIRT1. Values are given as means \pm SEM (n=11 for PGC-1 α , n=8 for SIRT1), * p<0.05.

Table S1. Blood Safety Parameters

| | Placebo | Resveratrol | p-value |
|---|--------------------|--------------------|----------------|
| Erythrocytes ($10^{12}/\text{l}$) | 4.85 \pm 0.09 | 4.81 \pm 0.09 | 0.52 |
| Hemoglobin (mmol/l) | 9.07 \pm 0.17 | 8.95 \pm 0.15 | 0.17 |
| Hematocrit (%) | 44.00 \pm 1.00 | 43.00 \pm 1.00 | 0.35 |
| MCV (fl) | 90.18 \pm 1.73 | 89.73 \pm 1.72 | 0.10 |
| MCHC (mmol/l) | 20.77 \pm 0.10 | 20.81 \pm 0.11 | 0.79 |
| Trombocytes ($10^9/\text{l}$) | 237.55 \pm 11.88 | 228.90 \pm 10.60 | 0.10 |
| RDW (%) | 13.25 \pm 0.15 | 13.48 \pm 0.26 | 0.40 |
| Leucocytes ($10^9/\text{l}$) | 7.03 \pm 0.44 | 6.48 \pm 0.39 | 0.03 |
| PT (sec) | 10.62 \pm 0.06 | 10.58 \pm 0.09 | 0.65 |
| aPTT (sec) | 29.00 \pm 0.83 | 30.63 \pm 1.88 | 0.48 |
| INR | 1.00 \pm 0.01 | 1.00 \pm 0.01 | 0.90 |
| Sodium (mmol/l) | 140.18 \pm 0.35 | 139.64 \pm 0.58 | 0.40 |
| Potassium (mmol/l) | 4.26 \pm 0.07 | 4.18 \pm 0.07 | 0.57 |
| Phosphate (mmol/l) | 0.88 \pm 0.03 | 0.92 \pm 0.04 | 0.08 |
| Chloride (mmol/l) | 106.36 \pm 0.94 | 106.18 \pm 0.60 | 0.83 |
| Urea (mmol/l) | 4.50 \pm 0.24 | 4.37 \pm 0.21 | 0.15 |
| Creatinine ($\mu\text{mol/l}$) | 76.10 \pm 3.83 | 75.82 \pm 3.28 | 0.83 |
| Alkaline phosphatase (U/l) | 79.00 \pm 4.11 | 76.55 \pm 3.81 | 0.30 |
| Gamma GT (U/l) | 29.60 \pm 3.71 | 28.00 \pm 2.61 | 0.44 |
| ASAT (U/l) | 19.00 \pm 1.46 | 18.91 \pm 1.11 | 0.93 |
| ALAT (U/l) | 31.91 \pm 2.21 | 28.09 \pm 1.54 | 0.02 |
| Bilirubin ($\mu\text{mol/l}$) | 14.25 \pm 1.00 | 13.86 \pm 1.07 | 0.42 |
| Total protein (g/l) | 66.55 \pm 1.24 | 65.39 \pm 0.80 | 0.14 |
| Albumin (g/l) | 38.02 \pm 0.79 | 37.33 \pm 0.55 | 0.18 |

On day 30, blood of the subjects was analyzed for several clinical chemistry, hematology and coagulation markers in order to confirm that resveratrol supplementation did not cause adverse effects. Values are given as means \pm SEM (n=11).

Table S2. Time Course RQ and EE

| | | Placebo | Resveratrol | | Placebo | Resveratrol | |
|--------------|-------------------------|-------------|-------------|---------|--------------|--------------|---------|
| Time (h:min) | Activity | RQ | | p-value | EE (MJ/d) | | p-value |
| - | Sleeping metabolic rate | 0.87 ± 0.01 | 0.89 ± 0.01 | 0.09 | 8.09 ± 0.24 | 7.75 ± 0.23 | 0.007 |
| 9.00-10.00 | breakfast | 0.87 ± 0.01 | 0.89 ± 0.01 | 0.17 | 11.86 ± 0.57 | 11.11 ± 0.46 | 0.05 |
| 10.00-13.00 | stepping + snack | 0.91 ± 0.01 | 0.94 ± 0.01 | 0.02 | 14.40 ± 0.32 | 13.86 ± 0.32 | 0.07 |
| 13.00-15.30 | lunch | 0.89 ± 0.01 | 0.89 ± 0.01 | 0.44 | 11.75 ± 0.44 | 11.65 ± 0.47 | 0.44 |
| 15.30-18.00 | stepping + snack | 0.90 ± 0.01 | 0.93 ± 0.01 | 0.01 | 14.85 ± 0.31 | 14.55 ± 0.25 | 0.24 |
| 18.00-20.00 | diner | 0.89 ± 0.01 | 0.90 ± 0.01 | 0.35 | 12.62 ± 0.39 | 12.26 ± 0.43 | 0.16 |
| 20.00-24.00 | stepping + snack | 0.91 ± 0.01 | 0.92 ± 0.01 | 0.33 | 13.16 ± 0.21 | 13.49 ± 0.23 | 0.27 |
| - | Sleeping metabolic rate | 0.88 ± 0.01 | 0.90 ± 0.01 | 0.21 | 8.06 ± 0.22 | 7.90 ± 0.18 | 0.06 |

Detailed time course of RQ and energy expenditure (EE) over the last 24h of resveratrol or placebo supplementation. Values are given as means ± SEM (n=10).

Table S3. Gene Set Enrichment Analysis

Gene Sets that Are Upregulated by Resveratrol

| NAME | SIZE | NES | FDR q-val |
|---|------|-----------|-------------|
| KEGG_DRUG METABOLISM - CYTOCHROME P450 | 65 | 1.9769608 | 0.055391833 |
| REACT_OLFACTORY SIGNALING PATHWAY | 326 | 1.883034 | 0.09561254 |
| REACT_RESPIRATORY ELECTRON TRANSPORT, ATP SYNTHESIS BY CHEMIOSMOTIC COUPLING, AND HEAT PRODUCTION BY UNCOUPLING PROTEINS. | 80 | 1.8434185 | 0.10389777 |
| WIP_HS_ELECTRON_TRANSPORT_CHAIN | 88 | 1.8122538 | 0.11256053 |
| KEGG_OLFACTORY TRANSDUCTION | 362 | 1.8074874 | 0.09481991 |
| KEGG_NEUROACTIVE LIGAND-RECEPTOR INTERACTION | 272 | 1.77137 | 0.11674452 |
| KEGG_METABOLISM OF XENOBIOTICS BY CYTOCHROME P450 | 74 | 1.7553229 | 0.12136926 |
| REACT_RESPIRATORY ELECTRON TRANSPORT | 64 | 1.7295214 | 0.14394456 |
| WIP_HS_METAPATHWAY_BIOTRANSFORMATION | 168 | 1.7112943 | 0.15443122 |
| KEGG_STEROID HORMONE BIOSYNTHESIS | 49 | 1.6833845 | 0.18774375 |

Gene Sets that Are Downregulated by Resveratrol

| NAME | SIZE | NES | FDR q-val |
|--|------|------------|-----------|
| WIP_HS_T_CELL_RECEPTOR_SIGNALING_PATHWAY | 133 | -3.1916971 | 0 |
| REACT_TCR SIGNALING | 64 | -3.005096 | 0 |
| REACT_COSTIMULATION BY THE CD28 FAMILY | 66 | -2.9632292 | 0 |
| REACT_GENERATION OF SECOND MESSENGER MOLECULES | 33 | -2.9366267 | 0 |
| REACT_INTERFERON GAMMA SIGNALING | 69 | -2.915365 | 0 |
| NCI_TCR_PATHWAY | 63 | -2.9115481 | 0 |
| NCI_CXCR4_PATHWAY | 100 | -2.8939204 | 0 |
| NCI_CD8TCRPATHWAY | 52 | -2.8382792 | 0 |
| REACT_CYTOKINE SIGNALING IN IMMUNE SYSTEM | 201 | -2.8212645 | 0 |

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|--|-----|------------|---|
| NCI_PDGFRBPATHWAY | 124 | -2.8036184 | 0 |
| REACT_PD-1 SIGNALING | 25 | -2.7586458 | 0 |
| KEGG_LEISHMANIASIS | 66 | -2.6811383 | 0 |
| NCI_TXA2PATHWAY | 54 | -2.6645982 | 0 |
| REACT_PHOSPHORYLATION OF CD3 AND TCR ZETA CHAINS | 22 | -2.6398609 | 0 |
| REACT_DOWNSTREAM TCR SIGNALING | 47 | -2.639546 | 0 |
| KEGG_ANTIGEN PROCESSING AND PRESENTATION | 67 | -2.6350987 | 0 |
| REACT_SIGNALING BY INTERLEUKINS | 102 | -2.6347778 | 0 |
| REACT_INTERLEUKIN-3, 5 AND GM-CSF SIGNALING | 48 | -2.6297154 | 0 |
| REACT_TRANSLOCATION OF ZAP-70 TO IMMUNOLOGICAL SYNAPSE | 20 | -2.615988 | 0 |
| REACT_INTERFERON SIGNALING | 106 | -2.6144912 | 0 |
| KEGG_VIRAL MYOCARDITIS | 68 | -2.610482 | 0 |
| KEGG_OSTEOCLAST DIFFERENTIATION | 124 | -2.5870695 | 0 |
| REACT_IMMUNOREGULATORY INTERACTIONS BETWEEN A LYMPHOID AND A NON-LYMPHOID CELL | 67 | -2.5784707 | 0 |
| NCI_FCR1PATHWAY | 58 | -2.5762885 | 0 |
| WIP_HS_IL-3_SIGNALING_PATHWAY | 44 | -2.5416484 | 0 |
| BIOC_CTLA4PATHWAY | 17 | -2.5320008 | 0 |
| REACT_INTERLEUKIN-2 SIGNALING | 37 | -2.5155833 | 0 |
| REACT_GPVI-MEDIATED ACTIVATION CASCADE | 32 | -2.5145288 | 0 |
| REACT_CELL SURFACE INTERACTIONS AT THE VASCULAR WALL | 90 | -2.504863 | 0 |
| WIP_HS_IL-2_SIGNALING_PATHWAY | 36 | -2.501475 | 0 |
| BIOC_HIVNEFPATHWAY | 52 | -2.4811022 | 0 |
| NCI_IL8CXCR2_PATHWAY | 33 | -2.4680712 | 0 |
| REACT_FORMATION OF PLATELET PLUG | 256 | -2.4612563 | 0 |
| WIP_HS_B_CELL_RECEPTOR_SIGNALING_PATHWAY | 93 | -2.457269 | 0 |
| KEGG_INFLUENZA A | 165 | -2.4498298 | 0 |
| KEGG_FC GAMMA R-MEDIATED PHAGOCYTOSIS | 90 | -2.441085 | 0 |
| KEGG_GRAFT-VERSUS-HOST DISEASE | 38 | -2.436977 | 0 |

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| NCI_IL12_2PATHWAY | 62 | -2.4258018 | 0 |
| WIP_HS_TYPE_II_INTERFERON_SIGNALING_(IFNG) | 36 | -2.4174535 | 0 |
| REACT_SIGNAL REGULATORY PROTEIN (SIRP) FAMILY INTERACTIONS | 16 | -2.4125907 | 0 |
| WIP_HS_IL-4_SIGNALING_PATHWAY | 46 | -2.3988845 | 0 |
| REACT_ADAPTIVE IMMUNITY SIGNALING | 406 | -2.3978803 | 0 |
| NCI_TCPTP_PATHWAY | 40 | -2.3929656 | 0 |
| KEGG_PHAGOSOME | 142 | -2.3906705 | 0 |
| KEGG_ALLOGRAFT REJECTION | 35 | -2.390225 | 0 |
| REACT_REGULATION OF SIGNALING BY CBL | 22 | -2.3880403 | 0 |
| NCI_IL8CXCR1_PATHWAY | 27 | -2.3865337 | 0 |
| REACT_INTERACTIONS OF THE IMMUNOGLOBULIN SUPERFAMILY (IGSF) MEMBER PROTEINS | 49 | -2.3851216 | 0 |
| NCI_PI3KCI PATHWAY | 47 | -2.3776176 | 0 |
| REACT_ANTIGEN PRESENTATION_ FOLDING, ASSEMBLY AND PEPTIDE LOADING OF CLASS I MHC | 25 | -2.3743033 | 0 |
| WIP_HS_IL-7_SIGNALING_PATHWAY | 26 | -2.370715 | 0 |
| WIP_HS_REGULATION_OF_TOLL-LIKE_RECEPTOR_SIGNALING_PATHWAY | 141 | -2.367435 | 0 |
| KEGG_CHEMOKINE SIGNALING PATHWAY | 182 | -2.3660667 | 0 |
| REACT_PLATELET ACTIVATION | 238 | -2.3499827 | 3.52E-05 |
| KEGG_SHIGELLOSIS | 61 | -2.3481314 | 3.46E-05 |
| KEGG_NATURAL KILLER CELL MEDIATED CYTOTOXICITY | 127 | -2.3426235 | 3.40E-05 |
| KEGG_TUBERCULOSIS | 170 | -2.3404236 | 3.34E-05 |
| BIOC_TCRPATHWAY | 42 | -2.3288894 | 3.28E-05 |
| KEGG_SALMONELLA INFECTION | 81 | -2.3285587 | 3.22E-05 |
| NCI_IFNGPATHWAY | 42 | -2.3255553 | 6.49E-05 |
| NCI_GMCSF_PATHWAY | 35 | -2.323727 | 9.61E-05 |
| WIP_HS_LEUKOCYTE_TARBASE | 124 | -2.3193054 | 9.46E-05 |
| KEGG_CELL ADHESION MOLECULES (CAMs) | 129 | -2.3113422 | 1.25E-04 |
| NCI_TNFPATHWAY | 46 | -2.3104668 | 1.23E-04 |
| WIP_HS_IL-6_SIGNALING_PATHWAY | 44 | -2.3059502 | 1.21E-04 |

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| KEGG_APOPTOSIS | 82 | -2.304132 | 1.50E-04 |
| REACT_RESPONSE TO ELEVATED PLATELET CYTOSOLIC CA2+ | 81 | -2.3031735 | 1.47E-04 |
| REACT_PLATELET ACTIVATION TRIGGERS | 81 | -2.3018825 | 1.45E-04 |
| REACT_HEMOSTASIS | 471 | -2.3016825 | 1.43E-04 |
| NCI_IL12_STAT4PATHWAY | 32 | -2.299623 | 1.41E-04 |
| WIP_HS_IL-5_SIGNALING_PATHWAY | 34 | -2.2968433 | 1.39E-04 |
| KEGG_T CELL RECEPTOR SIGNALING PATHWAY | 106 | -2.289124 | 1.37E-04 |
| WIP_HS_TOLL-LIKE_RECEPTOR_SIGNALING_PATHWAY | 100 | -2.2786198 | 1.65E-04 |
| WIP_HS_TCR_SIGNALING | 22 | -2.2754219 | 1.63E-04 |
| NCI_INTEGRIN_A4B1_PATHWAY | 32 | -2.2745643 | 1.61E-04 |
| KEGG_LEUKOCYTE TRANSENDOTHELIAL MIGRATION | 112 | -2.2721574 | 1.59E-04 |
| NCI_IL2_STAT5PATHWAY | 30 | -2.2624764 | 1.57E-04 |
| BIOC_RHOPATHWAY | 30 | -2.2533038 | 1.55E-04 |
| BIOC_NKCELLSPATHWAY | 20 | -2.2500315 | 1.77E-04 |
| NCI_NECTIN_PATHWAY | 28 | -2.2460837 | 1.75E-04 |
| REACT_INTERLEUKIN RECEPTOR SHC SIGNALING | 27 | -2.2445626 | 1.73E-04 |
| NCI_AVB3_OPN_PATHWAY | 31 | -2.2435489 | 1.96E-04 |
| REACT_INTEGRIN CELL SURFACE INTERACTIONS | 83 | -2.2385252 | 1.94E-04 |
| NCI_RAC1_PATHWAY | 52 | -2.2383287 | 1.92E-04 |
| KEGG_PROTEIN PROCESSING IN ENDOPLASMIC RETICULUM | 165 | -2.2322483 | 1.89E-04 |
| KEGG_TOXOPLASMOSIS | 127 | -2.224728 | 2.34E-04 |
| NCI_THROMBIN_PAR1_PATHWAY | 42 | -2.22046 | 2.31E-04 |
| NCI_VEGFR1_2_PATHWAY | 68 | -2.2173662 | 2.29E-04 |
| KEGG_HERPES SIMPLEX INFECTION | 176 | -2.2170615 | 2.49E-04 |
| BIOC_IL7PATHWAY | 16 | -2.2165563 | 2.46E-04 |
| NCI_FASPATHWAY | 38 | -2.2125952 | 2.66E-04 |
| BIOC_AMIPATHWAY | 21 | -2.2095914 | 2.63E-04 |
| KEGG_NOD-LIKE RECEPTOR SIGNALING PATHWAY | 58 | -2.206224 | 2.82E-04 |
| BIOC_ECMPATHWAY | 22 | -2.2058637 | 3.00E-04 |

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| BIOC_NFKBPATHWAY | 22 | -2.1963716 | 4.02E-04 |
| REACT_CTLA4 INHIBITORY SIGNALING | 21 | -2.1920862 | 4.84E-04 |
| KEGG_STAPHYLOCOCCUS AUREUS INFECTION | 47 | -2.1917832 | 4.79E-04 |
| REACT_ACTIVATION OF CHAPERONES BY IRE1ALPHA | 45 | -2.1906514 | 4.74E-04 |
| NCI_PTP1BPATHWAY | 51 | -2.186575 | 4.89E-04 |
| KEGG_INTESTINAL IMMUNE NETWORK FOR IGA PRODUCTION | 46 | -2.1859972 | 4.85E-04 |
| KEGG_TOLL-LIKE RECEPTOR SIGNALING PATHWAY | 100 | -2.1823518 | 5.00E-04 |
| KEGG_TYPE I DIABETES MELLITUS | 41 | -2.1822712 | 4.95E-04 |
| REACT_PLATELET DEGRANULATION | 76 | -2.1792736 | 4.90E-04 |
| BIOC_CSKPATHWAY | 21 | -2.1780436 | 4.86E-04 |
| WIP_HS_ADIPOCYTE_TARBASE | 17 | -2.1777108 | 4.81E-04 |
| KEGG_B CELL RECEPTOR SIGNALING PATHWAY | 73 | -2.1770425 | 4.76E-04 |
| BIOC_TNFR2PATHWAY | 18 | -2.1720812 | 5.47E-04 |
| KEGG_PATHOGENIC ESCHERICHIA COLI INFECTION | 53 | -2.1687157 | 5.42E-04 |
| NCI_IL2_1PATHWAY | 55 | -2.1679046 | 5.37E-04 |
| KEGG_MEASLES | 133 | -2.163485 | 5.69E-04 |
| KEGG_HEMATOPOIETIC CELL LINEAGE | 80 | -2.1606114 | 5.64E-04 |
| WIP_HS_INTEGRIN_CELL_SURFACE_INTERACTIONS | 15 | -2.1601222 | 5.59E-04 |
| NCI_AMB2_NEUTROPHILS_PATHWAY | 41 | -2.1547117 | 5.89E-04 |
| BIOC_GLEEVECPATHWAY | 22 | -2.1534536 | 5.84E-04 |
| NCI_IL27PATHWAY | 26 | -2.1517205 | 5.79E-04 |
| REACT_INFLAMMASOMES | 17 | -2.1466086 | 6.25E-04 |
| KEGG_MALARIA | 49 | -2.1440923 | 6.37E-04 |
| KEGG_HTLV-I INFECTION | 258 | -2.143463 | 6.32E-04 |
| NCI_INTEGRIN5_PATHWAY | 17 | -2.141736 | 6.27E-04 |
| NCI_IL6_7PATHWAY | 45 | -2.1412413 | 6.21E-04 |
| REACT_CD28 CO-STIMULATION | 29 | -2.1404538 | 6.33E-04 |
| KEGG_REGULATION OF ACTIN CYTOSKELETON | 208 | -2.1323092 | 6.93E-04 |
| BIOC_LAIRPATHWAY | 16 | -2.1301816 | 7.36E-04 |

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| NCI_HIVNEFPATHWAY | 35 | -2.1283083 | 7.47E-04 |
| NCI_TOLL_ENDOGENOUS_PATHWAY | 26 | -2.1151865 | 8.21E-04 |
| KEGG_PRIMARY IMMUNODEFICIENCY | 35 | -2.106903 | 9.41E-04 |
| REACT_THE ROLE OF NEF IN HIV-1 REPLICATION AND DISEASE PATHOGENESIS | 28 | -2.100416 | 9.97E-04 |
| REACT_ASSOCIATION OF TRIC_CCT WITH TARGET PROTEINS DURING BIOSYNTHESIS | 29 | -2.098923 | 9.89E-04 |
| WIP_HS_KIT_RECEPTOR_SIGNALING_PATHWAY | 56 | -2.0973399 | 0.001026911 |
| KEGG_ACUTE MYELOID LEUKEMIA | 57 | -2.0968208 | 0.001034313 |
| WIP_HS_MAPK_SIGNALING_PATHWAY | 160 | -2.093754 | 0.001071812 |
| BIOC_FASPATHWAY | 27 | -2.087879 | 0.001092478 |
| BIOC_NO2IL12PATHWAY | 15 | -2.0841186 | 0.001129477 |
| BIOC_CASPASEPATHWAY | 21 | -2.082723 | 0.001121049 |
| BIOC_KERATINOCYTEPATHWAY | 43 | -2.07871 | 0.00117042 |
| WIP_HS_INTERLEUKIN-3_5_AND_GM-CSF_SIGNALING | 16 | -2.0713346 | 0.001339471 |
| NCI_INTEGRIN2_PATHWAY | 28 | -2.0658157 | 0.001404977 |
| WIP_HS_TNF-ALPHA-NF-KB_SIGNALING_PATHWAY | 185 | -2.0609932 | 0.001466814 |
| KEGG_RHEUMATOID ARTHRITIS | 87 | -2.0600512 | 0.001470178 |
| WIP_HS_INTERFERON_TYPE_I | 28 | -2.0600033 | 0.001459677 |
| NCI_BCR_5PATHWAY | 66 | -2.0594187 | 0.001477417 |
| NCI_IL4_2PATHWAY | 60 | -2.059172 | 0.001467013 |
| REACT_SEMAPHORIN INTERACTIONS | 65 | -2.0533848 | 0.001567565 |
| BIOC_IL2RBPATWAY | 34 | -2.0504084 | 0.001611794 |
| REACT_SIGNALING BY RHO GTPASES | 121 | -2.0455878 | 0.00165782 |
| REACT_INTEGRIN ALPHAIIIB BETA3 SIGNALING | 27 | -2.0452664 | 0.001646465 |
| KEGG_BIOSYNTHESIS OF UNSATURATED FATTY ACIDS | 21 | -2.037153 | 0.001758598 |
| REACT_CELL-EXTRACELLULAR MATRIX INTERACTIONS | 18 | -2.0304053 | 0.001896827 |
| KEGG_OTHER GLYCAN DEGRADATION | 17 | -2.0297048 | 0.001897774 |
| NCI_ANGIOPOIETINRECEPTOR_PATHWAY | 48 | -2.0268133 | 0.001965854 |
| WIP_HS_RANKL-RANK_SIGNALING_PATHWAY | 55 | -2.0247173 | 0.001966095 |
| NCI_IL1PATHWAY | 32 | -2.0234203 | 0.002030815 |

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| NCI_MYC_REPRESSPATHWAY | 62 | -2.0197527 | 0.002136653 |
| REACT_RHO GTPASE CYCLE | 121 | -2.0185149 | 0.002162162 |
| WIP_HS_CELL_SURFACE_INTERACTIONS_AT_THE_VASCULAR_WALL | 36 | -2.0149705 | 0.002214421 |
| BIOC_EGFPATHWAY | 26 | -2.012483 | 0.002250164 |
| BIOC_PTENPATHWAY | 16 | -2.0056415 | 0.002337149 |
| NCI_ERBB1_RECEPTOR_PROXIMAL_PATHWAY | 33 | -2.0043683 | 0.002335085 |
| REACT_PROTEIN FOLDING | 54 | -2.00279 | 0.002369915 |
| BIOC_GSK3PATHWAY | 26 | -2.0026329 | 0.00238005 |
| NCI_IL2_PI3KPATHWAY | 37 | -1.9945135 | 0.002588744 |
| BIOC_P53HYPOXIAPATHWAY | 20 | -1.9921327 | 0.002634367 |
| KEGG_CHAGAS DISEASE (AMERICAN TRYPA NOSOMIASIS) | 101 | -1.9900556 | 0.002702796 |
| KEGG_BACTERIAL INVASION OF EPITHELIAL CELLS | 70 | -1.9867301 | 0.002746774 |
| NCI_CMYB_PATHWAY | 82 | -1.9857032 | 0.002730127 |
| REACT_P75NTR SIGNALS VIA NF-KB | 16 | -1.9785255 | 0.002988969 |
| NCI_ECADHERIN_STABILIZATION_PATHWAY | 40 | -1.9779519 | 0.002983994 |
| WIP_HS_EBV_LMP1_SIGNALING | 22 | -1.9777207 | 0.002978202 |
| NCI_EPHA2_FWDPATHWAY | 16 | -1.977513 | 0.002960579 |
| KEGG_SYSTEMIC LUPUS ERYTHEMATOSUS | 116 | -1.9758341 | 0.002966265 |
| REACT_UNFOLDED PROTEIN RESPONSE | 60 | -1.9718972 | 0.003079111 |
| REACT_NUCLEOTIDE-BINDING DOMAIN, LEUCINE RICH REPEAT CONTAINING RECEPTOR (NLR) SIGNALING PATHWAYS | 51 | -1.9718379 | 0.003061209 |
| REACT_CHAPERONIN-MEDIATED PROTEIN FOLDING | 49 | -1.9717852 | 0.003054488 |
| REACT_VIRAL DSRNA_TLR3_TRIF COMPLEX ACTIVATES RIP1 | 25 | -1.9706969 | 0.003047578 |
| NCI_GLYPICAN_1PATHWAY | 27 | -1.9691544 | 0.003063144 |
| NCI_ENDOTHELINPATHWAY | 62 | -1.9676932 | 0.003090687 |
| NCI_CASPASE_PATHWAY | 49 | -1.9672807 | 0.003084248 |
| NCI_SYNDECAN_4_PATHWAY | 31 | -1.9656968 | 0.00310095 |
| NCI_FAK_PATHWAY | 57 | -1.9642185 | 0.003172604 |
| WIP_HS_FOCAL_ADHESION | 184 | -1.9635513 | 0.003199586 |

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| KEGG_ADHERENS JUNCTION | 73 | -1.9560665 | 0.00335038 |
| NCI_ECADHERIN_NASCENTAJ_PATHWAY | 38 | -1.9552695 | 0.003376116 |
| NCI_LYSOPHOSPHOLIPID_PATHWAY | 65 | -1.9501927 | 0.003631476 |
| KEGG_LYSOSOME | 122 | -1.9470923 | 0.003708638 |
| WIP_HS_APOPTOSIS_MODULATION_BY_HSP70 | 18 | -1.9468039 | 0.003699987 |
| BIOC_PDGFPATHWAY | 26 | -1.9449476 | 0.003809317 |
| NCI_IL3_PATHWAY | 24 | -1.9431885 | 0.00386226 |
| REACT_INTERFERON ALPHA_BETA SIGNALING | 64 | -1.9402235 | 0.003949524 |
| KEGG_CHRONIC MYELOID LEUKEMIA | 73 | -1.9374653 | 0.004023213 |
| NCI_LYMPHANGIOGENESIS_PATHWAY | 24 | -1.9281691 | 0.004251898 |
| REACT_G ALPHA (12_13) SIGNALLING EVENTS | 77 | -1.9246988 | 0.004405566 |
| REACT_FORMATION OF TUBULIN FOLDING INTERMEDIATES BY CCT_TRIC | 21 | -1.9246123 | 0.00438262 |
| KEGG_PERTUSSIS | 71 | -1.9242588 | 0.004390685 |
| BIOC_IL6PATHWAY | 21 | -1.9214544 | 0.004440921 |
| WIP_HS_INTERFERON_ALPHA-BETA_SIGNALING | 24 | -1.9210826 | 0.004418148 |
| BIOC_IL2PATHWAY | 22 | -1.917379 | 0.004557698 |
| REACT_NEF-MEDIATES DOWN MODULATION OF CELL SURFACE RECEPTORS BY RECRUITING THEM TO CLATHRIN ADAPTERS | 21 | -1.9168459 | 0.004575906 |
| KEGG_FC EPSILON RI SIGNALING PATHWAY | 77 | -1.9162374 | 0.004592581 |
| BIOC_FCER1PATHWAY | 37 | -1.9153317 | 0.004579675 |
| REACT_NEPHRIN INTERACTIONS | 22 | -1.9141845 | 0.004586953 |
| WIP_HS_INFLAMMATORY_RESPONSE_PATHWAY | 32 | -1.9090894 | 0.00481389 |
| NCI_CD40_PATHWAY | 30 | -1.9033732 | 0.005008108 |
| NCI_EPHRINBREVPATHWAY | 30 | -1.9024303 | 0.005031189 |
| NCI_P38ALPHABETAPATHWAY | 30 | -1.9004377 | 0.005113904 |
| WIP_HS_EGF_RECEPTOR_SIGNALING_PATHWAY | 143 | -1.8966523 | 0.005195555 |
| REACT_DOWN-STREAM SIGNAL TRANSDUCTION | 36 | -1.8949499 | 0.005247768 |
| REACT_TRAF6 MEDIATED INDUCTION OF NFKB AND MAP KINASES UPON TLR7_8 OR 9 ACTIVATION | 73 | -1.8872705 | 0.005669379 |
| REACT_PLATELET SENSITIZATION BY LDL | 15 | -1.8831625 | 0.00586245 |

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| BIOC_TIDPATHWAY | 19 | -1.8829594 | 0.00585268 |
| REACT_P75 NTR RECEPTOR-MEDIATED SIGNALLING | 86 | -1.8829311 | 0.00582481 |
| WIP_HS_MUSCLE_CELL_TARBASE | 328 | -1.8811718 | 0.005936944 |
| NCI_EPOPATHWAY | 33 | -1.8811679 | 0.00590894 |
| NCI_TRAIL_PATHWAY | 28 | -1.8807642 | 0.005890639 |
| NCI_CERAMIDE_PATHWAY | 45 | -1.8801349 | 0.005880735 |
| REACT_CD28 DEPENDENT PI3K_AKT SIGNALING | 19 | -1.8781047 | 0.005975447 |
| WIP_HS_APOPTOSIS | 83 | -1.8780979 | 0.005947783 |
| NCI_CD8TCRDOWNSTREAMPATHWAY | 67 | -1.8759261 | 0.006069485 |
| WIP_HS_G13_SIGNALING_PATHWAY | 37 | -1.8706902 | 0.006389288 |
| REACT_PREFOLDIN MEDIATED TRANSFER OF SUBSTRATE TO CCT_TRIC | 27 | -1.8702604 | 0.006377927 |
| WIP_HS_INTEGRIN-MEDIATED_CELL_ADHESION | 99 | -1.8698312 | 0.006375916 |
| NCI_FGF_PATHWAY | 54 | -1.8683932 | 0.006410027 |
| WIP_HS_NOD_PATHWAY | 39 | -1.866042 | 0.006499623 |
| NCI_IL23PATHWAY | 37 | -1.8649479 | 0.006569438 |
| WIP_HS_SQUAMOUS_CELL_TARBASE | 117 | -1.8642224 | 0.006576047 |
| REACT_TAK1 ACTIVATES NFkB BY PHOSPHORYLATION AND ACTIVATION OF IKKS COMPLEX | 22 | -1.8636843 | 0.006573375 |
| KEGG_FOCAL ADHESION | 198 | -1.8626368 | 0.006642122 |
| NCI_AP1_PATHWAY | 69 | -1.8617864 | 0.006683086 |
| WIP_HS_IL-1_PATHWAY | 53 | -1.8615816 | 0.006653774 |
| REACT_TOLL LIKE RECEPTOR 3 (TLR3) CASCADE | 67 | -1.860271 | 0.006728928 |
| KEGG_RENAL CELL CARCINOMA | 70 | -1.8601748 | 0.006699672 |
| NCI_INTEGRIN_CS_PATHWAY | 26 | -1.8574877 | 0.006853806 |
| REACT_SEMA4D IN SEMAPHORIN SIGNALING | 28 | -1.8569036 | 0.006858972 |
| BIOC_GCRPATHWAY | 17 | -1.8511567 | 0.007214225 |
| NCI_HDAC_CLASSI_PATHWAY | 65 | -1.8505273 | 0.007251964 |
| WIP_HS_SIGNALING_OF_HEPATOCYTE_GROWTH_FACTOR_RECEPTOR | 33 | -1.8479131 | 0.007381756 |
| REACT_MYD88 DEPENDENT CASCADE INITIATED ON ENDOSOME | 74 | -1.8473257 | 0.007400869 |

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| NCI_ERBB2ERBB3PATHWAY | 43 | -1.8446246 | 0.007595204 |
| REACT_CLATHRIN DERIVED VESICLE BUDDING | 60 | -1.8427671 | 0.007696442 |
| REACT_TOLL LIKE RECEPTOR 7_8 (TLR7_8) CASCADE | 74 | -1.8420358 | 0.007755715 |
| KEGG_AUTOIMMUNE THYROID DISEASE | 49 | -1.8403913 | 0.007855522 |
| WIP_HS_FAS_PATHWAY_AND_STRESS_INDUCED_OF_HSP_REGULATION | 38 | -1.8394657 | 0.007904647 |
| NCI_CXCR3PATHWAY | 43 | -1.8386271 | 0.007897158 |
| BIOC_CERAMIDEPATHWAY | 21 | -1.8383093 | 0.007881178 |
| BIOC_CHEMICALPATHWAY | 20 | -1.8375019 | 0.007938497 |
| REACT_SIGNALING BY EGFR | 51 | -1.8334502 | 0.008200912 |
| BIOC_TOLLPATHWAY | 32 | -1.831022 | 0.00837821 |
| BIOC_METPATHWAY | 35 | -1.8293363 | 0.008481098 |
| NCI_NFKAPPABATYPICALPATHWAY | 17 | -1.8291018 | 0.008487571 |
| BIOC_CCR5PATHWAY | 17 | -1.8283324 | 0.008485848 |
| NCI_ILK_PATHWAY | 44 | -1.8273059 | 0.008522934 |
| KEGG_NON-SMALL CELL LUNG CANCER | 54 | -1.8256892 | 0.008672221 |
| REACT_TRANS-GOLGI NETWORK VESICLE BUDDING | 60 | -1.825211 | 0.008662016 |
| BIOC_CXCR4PATHWAY | 23 | -1.8247247 | 0.008659306 |
| NCI_P75NTRPATHWAY | 66 | -1.823523 | 0.008752542 |
| KEGG_NEUROTROPHIN SIGNALING PATHWAY | 126 | -1.8230648 | 0.008764952 |
| WIP_HS_INSULIN_SIGNALING | 160 | -1.8215282 | 0.008853972 |
| BIOC_NKTPATHWAY | 28 | -1.8185842 | 0.009043474 |
| REACT_COOPERATION OF PREFOLDIN AND TRIC_CCT IN ACTIN AND TUBULIN FOLDING | 28 | -1.8184261 | 0.009008421 |
| REACT_SIGNALLING BY NGF | 222 | -1.8174424 | 0.009035066 |
| BIOC_ACTINYPATHWAY | 18 | -1.8167003 | 0.009054083 |
| REACT_TOLL LIKE RECEPTOR 9 (TLR9) CASCADE | 76 | -1.8165551 | 0.009019394 |
| WIP_HS_EICOSANOID_SYNTHESIS | 19 | -1.8153262 | 0.009083658 |
| KEGG_PANCREATIC CANCER | 70 | -1.814876 | 0.009079379 |
| NCI_HDAC_CLASSIII_PATHWAY | 36 | -1.8122996 | 0.009247158 |
| BIOC_CALCINEURINPATHWAY | 18 | -1.8103873 | 0.009341 |

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| KEGG_AFRICAN TRYPANOSOMIASIS | 34 | -1.8101227 | 0.009374057 |
| WIP_HS_EPO_RECEPTOR_SIGNALING | 26 | -1.808065 | 0.009520139 |
| REACT_TRIGLYCERIDE BIOSYNTHESIS | 33 | -1.7988191 | 0.010253353 |
| KEGG_COLORECTAL CANCER | 62 | -1.7947834 | 0.010588672 |
| NCI_NFAT_TFPATHWAY | 48 | -1.7945398 | 0.010556559 |
| BIOC_MAPKPATHWAY | 84 | -1.792605 | 0.010657342 |
| REACT_MEMBRANE TRAFFICKING | 145 | -1.7906474 | 0.010846768 |
| NCI_RHOA_REG_PATHWAY | 43 | -1.7895484 | 0.010865686 |
| REACT_PLATELET AGGREGATION (PLUG FORMATION) | 34 | -1.786117 | 0.011204754 |
| BIOC_TPOPATHWAY | 22 | -1.7855176 | 0.011221645 |
| BIOC_DEATHPATHWAY | 32 | -1.7779057 | 0.011974087 |
| KEGG_CARBOHYDRATE DIGESTION AND ABSORPTION | 38 | -1.7734265 | 0.012349037 |
| NCI_ERBB1_DOWNSTREAM_PATHWAY | 106 | -1.7707475 | 0.012598489 |
| WIP_HS_LYMPHOCYTE_TARBASE | 407 | -1.770456 | 0.012567803 |
| WIP_HS_TGF_BETA_SIGNALING_PATHWAY_NETPATH | 117 | -1.7638924 | 0.013362426 |
| NCI_PDGFRA_PATHWAY | 21 | -1.7587968 | 0.013894224 |
| NCI_HIF1A_PATHWAY | 18 | -1.7581452 | 0.013917537 |
| NCI_ER_NONGENOMIC_PATHWAY | 39 | -1.7579918 | 0.01388214 |
| REACT_SMOOTH MUSCLE CONTRACTION | 24 | -1.7574564 | 0.013904452 |
| KEGG_SPHINGOLIPID METABOLISM | 40 | -1.7570717 | 0.013904913 |
| NCI_AVB3_INTEGRIN_PATHWAY | 73 | -1.7557452 | 0.013980775 |
| NCI_ALPHASYNUCLEIN_PATHWAY | 32 | -1.7547078 | 0.014057365 |
| REACT_TRAF6 MEDIATED INDUCTION OF PROINFLAMMATORY CYTOKINES | 62 | -1.7539696 | 0.014121322 |
| NCI_THROMBIN_PAR4_PATHWAY | 15 | -1.7533691 | 0.014135659 |
| BIOC_TNFR1_PATHWAY | 28 | -1.7531135 | 0.014128104 |
| BIOC_TH1TH2_PATHWAY | 17 | -1.7511908 | 0.014305572 |
| REACT_SEMA4D INDUCED CELL MIGRATION AND GROWTH-CONE COLLAPSE | 24 | -1.7510194 | 0.014284329 |
| WIP_HS_HYPERTROPHY_MODEL | 20 | -1.7497274 | 0.014500368 |
| NCI_KIT_PATHWAY | 52 | -1.7475505 | 0.014736447 |

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| NCI_RAC1_REG_PATHWAY | 38 | -1.7463287 | 0.014809483 |
| NCI_VEGFR1_PATHWAY | 28 | -1.745497 | 0.014873933 |
| KEGG_PHOSPHATIDYLINOSITOL SIGNALING SYSTEM | 78 | -1.7431507 | 0.015185053 |
| BIOC_MTORPATHWAY | 21 | -1.7401983 | 0.015515841 |
| WIP_HS_RIG-I-MDA5_MEDIATED_INDUCTION_OF_IFN-ALPHA-BETA_PATHWAYS | 21 | -1.7354453 | 0.016160844 |
| WIP_HS_EPITHELIUM_TARBASE | 269 | -1.7301913 | 0.016720043 |
| BIOC_PAR1PATHWAY | 19 | -1.7265502 | 0.017169883 |
| REACT_GOLGI ASSOCIATED VESICLE BIOGENESIS | 53 | -1.7227432 | 0.017502205 |
| BIOC_IL12PATHWAY | 20 | -1.7191168 | 0.01811797 |
| KEGG_PATHWAYS IN CANCER | 324 | -1.716018 | 0.018533086 |
| BIOC_BIOPEPTIDESPATHWAY | 37 | -1.7141166 | 0.018775648 |
| BIOC_TALL1PATHWAY | 15 | -1.7121788 | 0.019059075 |
| REACT_SPHINGOLIPID METABOLISM | 25 | -1.7065523 | 0.019786797 |

Table S4. Comparison of Calorie Restriction with Resveratrol

| | Calorie restriction | | Resveratrol | |
|----------------------------|--------------------------------|--------|--------------------|--------|
| | Rodents | Humans | Rodents | Humans |
| Body weight | ↓ | ↓ | ≠↓ | ≠ |
| Insulin | ↓ | ↓ | ↓ | ↓ |
| Energy expenditure | | | | |
| Total | ≠↓ | ↓ | ↑ | ≠ |
| Postprandial | ↓ | ↓ | ? | ↓ |
| Sedentary/Sleep | ↓ | ↓ | ? | ↓ |
| Fat mass and fat-free mass | ↓ | ↓ | ↓ | ? |
| Liver fat | ↓ | ↓ | ↓ | ↓ |
| Intramyocellular lipids | ↓ | ↓ | ↓ | ↑ |
| Insulin sensitivity | ↑ | ↑ | ↑ | ↑ |
| Inflammation markers | ↓ | ↓ | ↓ | ↓ |
| Mitochondrial efficiency | ↑ | ↑ | ↑ | ↑ |

Comparison of chronic calorie restriction in rodents and overweight humans to the effects of 30 days of resveratrol supplementation in obese humans on markers of ageing. To make the comparison complete, we also included the effects of resveratrol in rodents. The measured effects of calorie restriction and resveratrol on the above markers of aging were obtained from references that are included in the supplemental references.

↓ decrease, ↑ increase, ≠ similar, ? currently unknown

Supplemental Experimental Procedures

Analysis of resveratrol and dihydroresveratrol

To check compliance, the analytics of resveratrol (original and metabolites) were processed in a blinded fashion by a liquid chromatography mass spectrometry (LC-MS) system in both the resveratrol- and placebo-supplemented period by DSM Nutritional Products, Ltd. Kaiseraugst, Switzerland. Plasma collected on day 0, 7, 14, 21, 30 and during the stay in the respiration chamber (day 29) was processed for the determination of “free” trans-resveratrol and dihydroresveratrol (aglycone) and “total” trans-resveratrol and dihydroresveratrol (aglycone + glucuronide conjugates). After addition of an internal standard and liquid-liquid extraction (“free” analyte) or pre-digesting by β -glucuronidase followed by liquid-liquid extraction (“total” analyte), the samples were injected in a C18 column. Detection was performed using MS in the SIM mode.

Plasma biochemistry

In the morning of day 30 -after a standardized overnight fast for 12 hours- and during the postprandial microdialysis test, blood samples were withdrawn for the determination of plasma glucose, insulin, non-esterified fatty acids (NEFA) and triglycerides (TG) levels as described (Phielix et al., 2008). Glycerol and lactate levels during the microdialysis test were analyzed enzymatically on a Cobas Mira automated spectrophotometer. The concentration of leptin and adiponectin in plasma were analyzed by radioimmunoassay using commercial kits (Millipore Corporation, Billerica, MA, USA) in accordance with the manufacturer’s instructions. High-sensitivity C-reactive protein was measured

with a commercially available kit (Horiba ABX, Montpellier, France). Plasma inflammatory markers (IL-1b, IL-6, IL-8 and TNF- α) were measured with a commercially available Multi Spot ELISA kit (Meso Scale Discovery, Gaithersburg, MD, USA).

Molecular and protein expression

Mitochondrial DNA (mtDNA) copy number, the ratio of NADH dehydrogenase subunit one (ND1) to lipoprotein lipase (LPL) (mtDNA/nuclear DNA) was determined as described (Phielix et al., 2008).

Oxidative phosphorylation (OXPHOS), SIRT1, and PGC-1 α protein levels were measured in whole muscle by western blotting as an additional reflection of mitochondrial density, as described (Timmers et al., 2011). OXPHOS proteins were detected using a monoclonal antibody cocktail of five monoclonal antibodies directed against the different OXPHOS complexes (MS601, MitoSciences, Eugene, OR). SIRT1 was detected using a polyclonal antibody (#2493 Cell Signaling, Technology, Inc., Beverly MA, USA). PGC1 α was measured using a polyclonal antibody (Santa Cruz, Heidelberg, Germany). Total and the phosphorylated subunit α of AMPK (#2531 and #2532 Cell Signalling Technology, Inc., Beverly MA, USA) were detected as described (Feige et al., 2008).

Postprandial substrate utilization and tissue lipolysis

In 10 subjects, the lipolytic effects of resveratrol in adipose tissue and skeletal muscle were successfully determined by microdialysis, essentially according to (Goossens et al., 2004).

To study the response of tissue lipolysis on a mixed meal, subjects were instructed to consume a liquid meal within 5 minutes. The total energy content of the shake was 2.6 MJ and consisted of 32.6 En% carbohydrates, 61.2 En% fat and 6.3 En% protein. Blood samples were taken before ingestion of the liquid test meal ($t = -30$ and 0 min) and for 6h after meal ingestion at $t = 30, 60, 90, 120, 180, 240, 300$ and 360 min to determine glycerol, glucose, pyruvate and lactate in the plasma. Microdialysate was collected from the probes in 30 min fractions during the baseline period and during the early postprandial period (0 - 120 min) and at 1h fractions during the last 4h postprandially (120 - 360 min) to determine glycerol, glucose, pyruvate and lactate levels. Microdialysates were measured by Pronexus Analytical AB (Stockholm, Sweden) on a CMA 600 Microdialysis Analyzer (CMA Microdialysis AB, Solna, Sweden). Energy expenditure and substrate utilization were measured, before and for 6h after ingestion of the liquid test meal, using a ventilated hood system that analyses gasses every 15 seconds (Omnical, Maastricht University, The Netherlands) (Schoffelen et al., 1997). Metabolic rate was calculated from VO_2 (L/min) and VCO_2 (L/min) according to the equations of Frayn (Frayn, 1983). Nitrogen excretion was calculated based on the assumption that protein oxidation represents ~15% of total energy expenditure. Energy expenditure was calculated using the formula of Weir (Weir, 1949).

Microarray

Total RNA was prepared from human muscle using Trizol reagent (Invitrogen, Breda, The Netherlands), treated with DNase and purified on columns using

the RNeasy Mini Kit (Qiagen, Venlo, The Netherlands). RNA quantity and quality was assessed spectrophotometrically (ND-1000, NanoDrop Technologies, Wilmington, USA) and with 6000 Nano chips (Bioanalyzer 2100; Agilent, Amstelveen, The Netherlands), respectively.

The Ambion WT Expression kit (Life Technologies, P/N 4411974) and the Affymetrix GeneChip WT Terminal Labeling kit (Affymetrix, Santa Clara, CA; P/N 900671) were used for the preparation of labeled cDNA from 100ng of total RNA. Labeled samples were hybridized on Affymetrix GeneChip human Gene 1.1 ST arrays, provided in plate format. Hybridization, washing and scanning of the array plates was performed on an Affymetrix GeneTitan Instrument, according to the manufacturer's recommendations. Detailed protocols can be found in the Affymetrix WT Terminal Labeling and Hybridization User Manual (P/N 702808 revision 4), and are also available upon request.

Quality control of the datasets obtained from the scanned Affymetrix arrays was performed using Bioconductor (Gentleman et al., 2004) packages integrated in an on-line pipeline (Lin et al., 2011). Various advanced quality metrics, diagnostic plots, pseudo-images and classification methods were applied for selection of ascertain only excellent quality arrays were used in the subsequent analyses (Heber and Sick, 2006). An extensive description of the applied criteria is available upon request.

The more than 803000 probes on the Human Gene 1.1 ST array were redefined according to Dai et al. (Dai et al., 2005) utilizing current genome information, yielding 19738 unique genes. Probes were reorganized based on the Entrez Gene database, build 37, version 2 (remapped CDF v14).

Normalized expression estimates were obtained from the raw intensity values using the robust multiarray analysis (RMA) preprocessing algorithm (Irizarry et al., 2003). All genes represented on the array were considered for the unbiased Gene Set Enrichment Analysis (GSEA) (Subramanian et al., 2005). GSEA was run using 1000 permutations per gene set.

Unsupervised hierarchical clustering was performed using complete linkage and Pearson rank correlation distance on the normalized transcripts using software implemented in Genepattern

(<http://www.broadinstitute.org/cancer/software/genepattern/>) (de Hoon et al., 2004; Reich et al., 2006). The z-score was calculated by subtracting the mean expression value for each transcript from each of the values and then dividing the resulting values by the standard deviation. Color in the heat-maps reflects the relative transcript abundance level with red being higher and blue lower than the mean transcript abundance value. Transcript ordering is determined as in hierarchical clustering using the distance function 1-correlation. Mouse microarray data (Lagouge et al., 2006) were reanalysed using KEGG pathway gene sets in the GSEA software (Broad institute) (Subramanian et al., 2005).

Supplemental References

Ahn, J., Cho, I., Kim, S., Kwon, D., and Ha, T. (2008). Dietary resveratrol alters lipid metabolism-related gene expression of mice on an atherogenic diet. *J Hepatol* 49, 1019-1028.

Aubin, M.C., Lajoie, C., Clement, R., Gosselin, H., Calderone, A., and Perrault, L.P. (2008). Female rats fed a high-fat diet were associated with vascular dysfunction and cardiac fibrosis in the absence of overt obesity and hyperlipidemia: therapeutic potential of resveratrol. *J Pharmacol Exp Ther* 325, 961-968.

Ballor, D.L. (1991). Effect of dietary restriction and/or exercise on 23-h metabolic rate and body composition in female rats. *J Appl Physiol* 71, 801-806.

Barger, J.L., Kayo, T., Vann, J.M., Arias, E.B., Wang, J., Hacker, T.A., Wang, Y., Raederstorff, D., Morrow, J.D., Leeuwenburgh, C., Allison, D.B., Saupe, K.W., Cartee, G.D., Weindruch, R., and Prolla, T.A. (2008). A low dose of dietary resveratrol partially mimics caloric restriction and retards aging parameters in mice. *PLoS One* 3, e2264.

Barger, J.L., Walford, R.L., and Weindruch, R. (2003). The retardation of aging by caloric restriction: its significance in the transgenic era. *Exp Gerontol* 38, 1343-1351.

Barzilai, N., Banerjee, S., Hawkins, M., Chen, W., and Rossetti, L. (1998). Caloric restriction reverses hepatic insulin resistance in aging rats by decreasing visceral fat. *J Clin Invest* 101, 1353-1361.

Baur, J.A. (2010). Resveratrol, sirtuins, and the promise of a DR mimetic. *Mech Ageing Dev* 131, 261-269.

Baur, J.A., Pearson, K.J., Price, N.L., Jamieson, H.A., Lerin, C., Kalra, A., Prabhu, V.V., Allard, J.S., Lopez-Lluch, G., Lewis, K., Pistell, P.J., Poosala, S., Becker, K.G., Boss, O., Gwinn, D., Wang, M., Ramaswamy, S., Fishbein, K.W., Spencer, R.G., Lakatta, E.G., Le Couteur, D., Shaw, R.J., Navas, P., Puigserver, P., Ingram, D.K., de Cabo, R., and Sinclair, D.A. (2006). Resveratrol improves health and survival of mice on a high-calorie diet. *Nature* 444, 337-342.

Bertrand, H.A., Lynd, F.T., Masoro, E.J., and Yu, B.P. (1980). Changes in adipose mass and cellularity through the adult life of rats fed ad libitum or a life-prolonging restricted diet. *J Gerontol* 35, 827-835.

Bhat, K.P.L., Kosmeder, J.W., 2nd, and Pezzuto, J.M. (2001). Biological effects of resveratrol. *Antioxid Redox Signal* 3, 1041-1064.

Bruss, M.D., Khambatta, C.F., Ruby, M.A., Aggarwal, I., and Hellerstein, M.K. (2010). Calorie restriction increases fatty acid synthesis and whole body fat oxidation rates. *Am J Physiol Endocrinol Metab* 298, E108-116.

Bujanda, L., Hijona, E., Larzabal, M., Beraza, M., Aldazabal, P., Garcia-Urkia, N., Sarasqueta, C., Cosme, A., Irastorza, B., Gonzalez, A., and Arenas, J.I., Jr. (2008). Resveratrol inhibits nonalcoholic fatty liver disease in rats. *BMC Gastroenterol* 8, 40.

Chen, L.L., Zhang, H.H., Zheng, J., Hu, X., Kong, W., Hu, D., Wang, S.X., and Zhang, P. (2011). Resveratrol attenuates high-fat diet-induced insulin resistance by influencing skeletal muscle lipid transport and subsarcolemmal mitochondrial beta-oxidation. *Metabolism*.

Civitarese, A.E., Carling, S., Heilbronn, L.K., Hulver, M.H., Ukropcova, B., Deutsch, W.A., Smith, S.R., and Ravussin, E. (2007). Calorie restriction increases muscle mitochondrial biogenesis in healthy humans. *PLoS Med* 4, e76.

Corton, J.C., Apte, U., Anderson, S.P., Limaye, P., Yoon, L., Latendresse, J., Dunn, C., Everitt, J.I., Voss, K.A., Swanson, C., Kimbrough, C., Wong, J.S., Gill, S.S., Chandraratna, R.A., Kwak, M.K., Kensler, T.W., Stulnig, T.M., Steffensen, K.R., Gustafsson, J.A., and Mehendale, H.M. (2004). Mimetics of caloric restriction include agonists of lipid-activated nuclear receptors. *J Biol Chem* 279, 46204-46212.

Dai, M., Wang, P., Boyd, A.D., Kostov, G., Athey, B., Jones, E.G., Bunney, W.E., Myers, R.M., Speed, T.P., Akil, H., Watson, S.J., and Meng, F. (2005). Evolving gene/transcript definitions significantly alter the interpretation of GeneChip data. *Nucleic Acids Res* 33, e175.

Das, S.K., Gilhooly, C.H., Golden, J.K., Pittas, A.G., Fuss, P.J., Cheatham, R.A., Tyler, S., Tsay, M., McCrory, M.A., Lichtenstein, A.H., Dallal, G.E., Dutta, C., Bhapkar, M.V., Delany, J.P., Saltzman, E., and Roberts, S.B. (2007). Long-term effects of 2 energy-restricted diets differing in glycemic load on dietary adherence, body composition, and metabolism in CALERIE: a 1-y randomized controlled trial. *Am J Clin Nutr* 85, 1023-1030.

de Hoon, M.J., Imoto, S., Nolan, J., and Miyano, S. (2004). Open source clustering software. *Bioinformatics* 20, 1453-1454.

Deng, J.Y., Hsieh, P.S., Huang, J.P., Lu, L.S., and Hung, L.M. (2008). Activation of estrogen receptor is crucial for resveratrol-stimulating muscular glucose uptake via both insulin-dependent and -independent pathways. *Diabetes* 57, 1814-1823.

Dengel, D.R., Pratley, R.E., Hagberg, J.M., Rogus, E.M., and Goldberg, A.P. (1996). Distinct effects of aerobic exercise training and weight loss on glucose homeostasis in obese sedentary men. *J Appl Physiol* 81, 318-325.

Dhahbi, J.M., Mote, P.L., Fahy, G.M., and Spindler, S.R. (2005). Identification of potential caloric restriction mimetics by microarray profiling. *Physiol Genomics* 23, 343-350.

Dhahbi, J.M., Tsuchiya, T., Kim, H.J., Mote, P.L., and Spindler, S.R. (2006). Gene expression and physiologic responses of the heart to the initiation and withdrawal of caloric restriction. *J Gerontol A Biol Sci Med Sci* 61, 218-231.

Drapeau, S., Doucet, E., Rabasa-Lhoret, R., Brochu, M., Prud'homme, D., and Imbeault, P. (2011). Improvement in insulin sensitivity by weight loss does not affect hyperinsulinemia-mediated reduction in total and high molecular weight adiponectin: a MONET study. *Appl Physiol Nutr Metab* 36, 191-200.

Duffy, P.H., Feuers, R., Nakamura, K.D., Leahey, J., and Hart, R.W. (1990). Effect of chronic caloric restriction on the synchronization of various physiological measures in old female Fischer 344 rats. *Chronobiol Int* 7, 113-124.

Edwards, M.G., Anderson, R.M., Yuan, M., Kendzierski, C.M., Weindruch, R., and Prolla, T.A. (2007). Gene expression profiling of aging reveals activation of a p53-mediated transcriptional program. *BMC Genomics* 8, 80.

Escriva, F., Gavete, M.L., Fermin, Y., Perez, C., Gallardo, N., Alvarez, C., Andres, A., Ros, M., and Carrascosa, J.M. (2007). Effect of age and moderate food restriction on insulin sensitivity in Wistar rats: role of adiposity. *J Endocrinol* 194, 131-141.

Fan, J.G., Zhong, L., Xu, Z.J., Tia, L.Y., Ding, X.D., Li, M.S., and Wang, G.L. (2003). Effects of low-calorie diet on steatohepatitis in rats with obesity and hyperlipidemia. *World J Gastroenterol* 9, 2045-2049.

Feige, J.N., Lagouge, M., Canto, C., Strehle, A., Houten, S.M., Milne, J.C., Lambert, P.D., Matak, C., Elliott, P.J., and Auwerx, J. (2008). Specific SIRT1 activation mimics low energy levels and protects against diet-induced metabolic disorders by enhancing fat oxidation. *Cell Metab* 8, 347-358.

Fontana, L., Meyer, T.E., Klein, S., and Holloszy, J.O. (2004). Long-term calorie restriction is highly effective in reducing the risk for atherosclerosis in humans. *Proc Natl Acad Sci U S A* 101, 6659-6663.

Frayn, K.N. (1983). Calculation of substrate oxidation rates in vivo from gaseous exchange. *J Appl Physiol* 55, 628-634.

Fu, C., Hickey, M., Morrison, M., McCarter, R., and Han, E.S. (2006). Tissue specific and non-specific changes in gene expression by aging and by early stage CR. *Mech Ageing Dev* 127, 905-916.

Gentleman, R.C., Carey, V.J., Bates, D.M., Bolstad, B., Dettling, M., Dudoit, S., Ellis, B., Gautier, L., Ge, Y., Gentry, J., Hornik, K., Hothorn, T., Huber, W., Iacus, S., Irizarry, R., Leisch, F., Li, C., Maechler, M., Rossini, A.J., Sawitzki, G., Smith, C., Smyth, G., Tierney, L., Yang, J.Y., and Zhang, J. (2004).

Bioconductor: open software development for computational biology and bioinformatics. *Genome Biol* 5, R80.

Gonzalez-Rodriguez, A., Mas Gutierrez, J.A., Sanz-Gonzalez, S., Ros, M., Burks, D.J., and Valverde, A.M. (2010). Inhibition of PTP1B restores IRS1-mediated hepatic insulin signaling in IRS2-deficient mice. *Diabetes* 59, 588-599.

Goodpaster, B.H., Kelley, D.E., Wing, R.R., Meier, A., and Thaete, F.L. (1999). Effects of weight loss on regional fat distribution and insulin sensitivity in obesity. *Diabetes* 48, 839-847.

Goodpaster, B.H., Theriault, R., Watkins, S.C., and Kelley, D.E. (2000). Intramuscular lipid content is increased in obesity and decreased by weight loss. *Metabolism* 49, 467-472.

Goodrick, C.L., Ingram, D.K., Reynolds, M.A., Freeman, J.R., and Cider, N. (1990). Effects of intermittent feeding upon body weight and lifespan in inbred mice: interaction of genotype and age. *Mech Ageing Dev* 55, 69-87.

Goossens, G.H., Blaak, E.E., Saris, W.H., and van Baak, M.A. (2004). Angiotensin II-induced effects on adipose and skeletal muscle tissue blood flow and lipolysis in normal-weight and obese subjects. *J Clin Endocrinol Metab* 89, 2690-2696.

Haufe, S., Engeli, S., Kast, P., Bohnke, J., Utz, W., Haas, V., Hermsdorf, M., Mahler, A., Wiesner, S., Birkenfeld, A.L., Sell, H., Otto, C., Mehling, H., Luft, F.C., Eckel, J., Schulz-Menger, J., Boschmann, M., and Jordan, J. (2011). Randomized comparison of reduced fat and reduced carbohydrate hypocaloric diets on intrahepatic fat in overweight and obese human subjects. *Hepatology* 53, 1504-1514.

Havel, P.J. (2002). Control of energy homeostasis and insulin action by adipocyte hormones: leptin, acylation stimulating protein, and adiponectin. *Curr Opin Lipidol* 13, 51-59.

Heber, S., and Sick, B. (2006). Quality assessment of Affymetrix GeneChip data. *OMICS* 10, 358-368.

Heilbronn, L.K., de Jonge, L., Frisard, M.I., DeLany, J.P., Larson-Meyer, D.E., Rood, J., Nguyen, T., Martin, C.K., Volaufova, J., Most, M.M., Greenway, F.L., Smith, S.R., Deutsch, W.A., Williamson, D.A., and Ravussin, E. (2006). Effect of 6-month calorie restriction on biomarkers of longevity, metabolic adaptation, and oxidative stress in overweight individuals: a randomized controlled trial. *JAMA* 295, 1539-1548.

Higami, Y., Pugh, T.D., Page, G.P., Allison, D.B., Prolla, T.A., and Weindruch, R. (2004). Adipose tissue energy metabolism: altered gene expression profile of mice subjected to long-term caloric restriction. *FASEB J* 18, 415-417.

Huang, J.P., Huang, S.S., Deng, J.Y., Chang, C.C., Day, Y.J., and Hung, L.M. (2010). Insulin and resveratrol act synergistically, preventing cardiac

dysfunction in diabetes, but the advantage of resveratrol in diabetics with acute heart attack is antagonized by insulin. *Free Radic Biol Med* 49, 1710-1721.

Irizarry, R.A., Hobbs, B., Collin, F., Beazer-Barclay, Y.D., Antonellis, K.J., Scherf, U., and Speed, T.P. (2003). Exploration, normalization, and summaries of high density oligonucleotide array probe level data. *Biostatistics* 4, 249-264.

Kim, J.Y., Kim, D.H., Choi, J., Park, J.K., Jeong, K.S., Leeuwenburgh, C., Yu, B.P., and Chung, H.Y. (2009). Changes in lipid distribution during aging and its modulation by calorie restriction. *Age (Dordr)* 31, 127-142.

Kim, Y.H., Kim, Y.S., Kang, S.S., Cho, G.J., and Choi, W.S. (2010). Resveratrol inhibits neuronal apoptosis and elevated Ca²⁺/calmodulin-dependent protein kinase II activity in diabetic mouse retina. *Diabetes* 59, 1825-1835.

Kumar, A., and Sharma, S.S. (2010). NF-kappaB inhibitory action of resveratrol: a probable mechanism of neuroprotection in experimental diabetic neuropathy. *Biochem Biophys Res Commun* 394, 360-365.

Labayen, I., Ortega, F.B., Ruiz, J.R., Lasa, A., Simon, E., and Margareto, J. (2011). Role of baseline leptin and ghrelin levels on body weight and fat mass changes after an energy-restricted diet intervention in obese women: effects on energy metabolism. *J Clin Endocrinol Metab* 96, E996-1000.

Lagouge, M., Argmann, C., Gerhart-Hines, Z., Meziane, H., Lerin, C., Daussin, F., Messadeq, N., Milne, J., Lambert, P., Elliott, P., Geny, B., Laakso, M., Puigserver, P., and Auwerx, J. (2006). Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1alpha. *Cell* 127, 1109-1122.

Larson-Meyer, D.E., Heilbronn, L.K., Redman, L.M., Newcomer, B.R., Frisard, M.I., Anton, S., Smith, S.R., Alfonso, A., and Ravussin, E. (2006). Effect of calorie restriction with or without exercise on insulin sensitivity, beta-cell function, fat cell size, and ectopic lipid in overweight subjects. *Diabetes Care* 29, 1337-1344.

Lee, C.K., Allison, D.B., Brand, J., Weindruch, R., and Prolla, T.A. (2002). Transcriptional profiles associated with aging and middle age-onset caloric restriction in mouse hearts. *Proc Natl Acad Sci U S A* 99, 14988-14993.

Lee, C.K., Klopp, R.G., Weindruch, R., and Prolla, T.A. (1999). Gene expression profile of aging and its retardation by caloric restriction. *Science* 285, 1390-1393.

Lefevre, M., Redman, L.M., Heilbronn, L.K., Smith, J.V., Martin, C.K., Rood, J.C., Greenway, F.L., Williamson, D.A., Smith, S.R., and Ravussin, E. (2009). Caloric restriction alone and with exercise improves CVD risk in healthy non-obese individuals. *Atherosclerosis* 203, 206-213.

Lim, E.L., Hollingsworth, K.G., Aribisala, B.S., Chen, M.J., Mathers, J.C., and Taylor, R. (2011). Reversal of type 2 diabetes: normalisation of beta cell function in association with decreased pancreas and liver triacylglycerol. *Diabetologia* 54, 2506-2514.

Lin, K., Kools, H., de Groot, P.J., Gavai, A.K., Basnet, R.K., Cheng, F., Wu, J., Wang, X., Lommen, A., Hooiveld, G.J., Bonnema, G., Visser, R.G., Muller, M.R., and Leunissen, J.A. (2011). MADMAX – Management and analysis database for multiple -omics experiments. *J Integr Bioinform* 8, 160.

Macarulla, M.T., Alberdi, G., Gomez, S., Tueros, I., Bald, C., Rodriguez, V.M., Martinez, J.A., and Portillo, M.P. (2009). Effects of different doses of resveratrol on body fat and serum parameters in rats fed a hypercaloric diet. *J Physiol Biochem* 65, 369-376.

Martin, C.K., Anton, S.D., Han, H., York-Crowe, E., Redman, L.M., Ravussin, E., and Williamson, D.A. (2007a). Examination of cognitive function during six months of calorie restriction: results of a randomized controlled trial. *Rejuvenation Res* 10, 179-190.

Martin, C.K., Heilbronn, L.K., de Jonge, L., DeLany, J.P., Volaufova, J., Anton, S.D., Redman, L.M., Smith, S.R., and Ravussin, E. (2007b). Effect of calorie restriction on resting metabolic rate and spontaneous physical activity. *Obesity (Silver Spring)* 15, 2964-2973.

Meites, J. (1989). Evidence that underfeeding acts via the neuroendocrine system to influence aging processes. *Prog Clin Biol Res* 287, 169-180.

Milne, J.C., Lambert, P.D., Schenk, S., Carney, D.P., Smith, J.J., Gagne, D.J., Jin, L., Boss, O., Perni, R.B., Vu, C.B., Bemis, J.E., Xie, R., Disch, J.S., Ng, P.Y., Nunes, J.J., Lynch, A.V., Yang, H., Galonek, H., Israelian, K., Choy, W., Iffland, A., Lavu, S., Medvedik, O., Sinclair, D.A., Olefsky, J.M., Jirousek, M.R., Elliott, P.J., and Westphal, C.H. (2007). Small molecule activators of SIRT1 as therapeutics for the treatment of type 2 diabetes. *Nature* 450, 712-716.

Nadeau, K.J., Ehlers, L.B., Aguirre, L.E., Moore, R.L., Jew, K.N., Ortmeyer, H.K., Hansen, B.C., Reusch, J.E., and Draznin, B. (2006). Exercise training and calorie restriction increase SREBP-1 expression and intramuscular triglyceride in skeletal muscle. *Am J Physiol Endocrinol Metab* 291, E90-98.

Niskanen, L., Uusitupa, M., Sarlund, H., Siitonen, O., Paljarvi, L., and Laakso, M. (1996). The effects of weight loss on insulin sensitivity, skeletal muscle composition and capillary density in obese non-diabetic subjects. *Int J Obes Relat Metab Disord* 20, 154-160.

Palsamy, P., and Subramanian, S. (2008). Resveratrol, a natural phytoalexin, normalizes hyperglycemia in streptozotocin-nicotinamide induced experimental diabetic rats. *Biomed Pharmacother* 62, 598-605.

- Palsamy, P., and Subramanian, S. (2010). Ameliorative potential of resveratrol on proinflammatory cytokines, hyperglycemia mediated oxidative stress, and pancreatic beta-cell dysfunction in streptozotocin-nicotinamide-induced diabetic rats. *J Cell Physiol* 224, 423-432.
- Pearson, K.J., Baur, J.A., Lewis, K.N., Peshkin, L., Price, N.L., Labinskyy, N., Swindell, W.R., Kamara, D., Minor, R.K., Perez, E., Jamieson, H.A., Zhang, Y., Dunn, S.R., Sharma, K., Pleshko, N., Woollett, L.A., Csiszar, A., Ikeno, Y., Le Couteur, D., Elliott, P.J., Becker, K.G., Navas, P., Ingram, D.K., Wolf, N.S., Ungvari, Z., Sinclair, D.A., and de Cabo, R. (2008). Resveratrol delays age-related deterioration and mimics transcriptional aspects of dietary restriction without extending life span. *Cell Metab* 8, 157-168.
- Petersen, K.F., Dufour, S., Befroy, D., Lehrke, M., Hendler, R.E., and Shulman, G.I. (2005). Reversal of nonalcoholic hepatic steatosis, hepatic insulin resistance, and hyperglycemia by moderate weight reduction in patients with type 2 diabetes. *Diabetes* 54, 603-608.
- Phielix, E., Schrauwen-Hinderling, V.B., Mensink, M., Lenaers, E., Meex, R., Hoeks, J., Kooi, M.E., Moonen-Kornips, E., Sels, J.P., Hesselink, M.K., and Schrauwen, P. (2008). Lower intrinsic ADP-stimulated mitochondrial respiration underlies in vivo mitochondrial dysfunction in muscle of male type 2 diabetic patients. *Diabetes* 57, 2943-2949.
- Racette, S.B., Weiss, E.P., Villareal, D.T., Arif, H., Steger-May, K., Schechtman, K.B., Fontana, L., Klein, S., and Holloszy, J.O. (2006). One year of caloric restriction in humans: feasibility and effects on body composition and abdominal adipose tissue. *J Gerontol A Biol Sci Med Sci* 61, 943-950.
- Rahman, M., Halade, G.V., Bhattacharya, A., and Fernandes, G. (2009). The fat-1 transgene in mice increases antioxidant potential, reduces pro-inflammatory cytokine levels, and enhances PPAR-gamma and SIRT-1 expression on a calorie restricted diet. *Oxid Med Cell Longev* 2, 307-316.
- Ramadori, G., Gautron, L., Fujikawa, T., Vianna, C.R., Elmquist, J.K., and Coppari, R. (2009). Central administration of resveratrol improves diet-induced diabetes. *Endocrinology* 150, 5326-5333.
- Redman, L.M., Heilbronn, L.K., Martin, C.K., Alfonso, A., Smith, S.R., and Ravussin, E. (2007). Effect of calorie restriction with or without exercise on body composition and fat distribution. *J Clin Endocrinol Metab* 92, 865-872.
- Reich, M., Liefeld, T., Gould, J., Lerner, J., Tamayo, P., and Mesirov, J.P. (2006). GenePattern 2.0. *Nat Genet* 38, 500-501.
- Rivera, L., Moron, R., Zarzuelo, A., and Galisteo, M. (2009). Long-term resveratrol administration reduces metabolic disturbances and lowers blood pressure in obese Zucker rats. *Biochem Pharmacol* 77, 1053-1063.
- Rocha, K.K., Souza, G.A., Ebaid, G.X., Seiva, F.R., Cataneo, A.C., and Novelli, E.L. (2009). Resveratrol toxicity: effects on risk factors for

atherosclerosis and hepatic oxidative stress in standard and high-fat diets. *Food Chem Toxicol* 47, 1362-1367.

Russell, J.C., Epling, W.F., Pierce, D., Amy, R.M., and Boer, D.P. (1987). Induction of voluntary prolonged running by rats. *J Appl Physiol* 63, 2549-2553.

Schoffelen, P.F., Westerterp, K.R., Saris, W.H., and Ten Hoor, F. (1997). A dual-respiration chamber system with automated calibration. *J Appl Physiol* 83, 2064-2072.

Selman, C., Kerrison, N.D., Cooray, A., Piper, M.D., Lingard, S.J., Barton, R.H., Schuster, E.F., Blanc, E., Gems, D., Nicholson, J.K., Thornton, J.M., Partridge, L., and Withers, D.J. (2006). Coordinated multitissue transcriptional and plasma metabolomic profiles following acute caloric restriction in mice. *Physiol Genomics* 27, 187-200.

Shang, J., Chen, L.L., Xiao, F.X., Sun, H., Ding, H.C., and Xiao, H. (2008). Resveratrol improves non-alcoholic fatty liver disease by activating AMP-activated protein kinase. *Acta Pharmacol Sin* 29, 698-706.

Sharma, S., Chopra, K., and Kulkarni, S.K. (2007). Effect of insulin and its combination with resveratrol or curcumin in attenuation of diabetic neuropathic pain: participation of nitric oxide and TNF-alpha. *Phytother Res* 21, 278-283.

Sohal, R.S., and Weindruch, R. (1996). Oxidative stress, caloric restriction, and aging. *Science* 273, 59-63.

Su, H.C., Hung, L.M., and Chen, J.K. (2006). Resveratrol, a red wine antioxidant, possesses an insulin-like effect in streptozotocin-induced diabetic rats. *Am J Physiol Endocrinol Metab* 290, E1339-1346.

Subramanian, A., Tamayo, P., Mootha, V.K., Mukherjee, S., Ebert, B.L., Gillette, M.A., Paulovich, A., Pomeroy, S.L., Golub, T.R., Lander, E.S., and Mesirov, J.P. (2005). Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A* 102, 15545-15550.

Sun, C., Zhang, F., Ge, X., Yan, T., Chen, X., Shi, X., and Zhai, Q. (2007). SIRT1 improves insulin sensitivity under insulin-resistant conditions by repressing PTP1B. *Cell Metab* 6, 307-319.

Thirunavukkarasu, M., Penumathsa, S.V., Koneru, S., Juhasz, B., Zhan, L., Otani, H., Bagchi, D., Das, D.K., and Maulik, N. (2007). Resveratrol alleviates cardiac dysfunction in streptozotocin-induced diabetes: Role of nitric oxide, thioredoxin, and heme oxygenase. *Free Radic Biol Med* 43, 720-729.

Timmers, S., de Vogel-van den Bosch, J., Hesselink, M.K., van Beurden, D., Schaart, G., Ferraz, M.J., Losen, M., Martinez-Martinez, P., De Baets, M.H., Aerts, J.M., and Schrauwen, P. (2011). Paradoxical increase in TAG and DAG content parallel the insulin sensitizing effect of unilateral DGAT1 overexpression in rat skeletal muscle. *PLoS One* 6, e14503.

Tsuchiya, T., Dhahbi, J.M., Cui, X., Mote, P.L., Bartke, A., and Spindler, S.R. (2004). Additive regulation of hepatic gene expression by dwarfism and caloric restriction. *Physiol Genomics* 17, 307-315.

Tunali-Akbay, T., Sehirli, O., Ercan, F., and Sener, G. (2010). Resveratrol protects against methotrexate-induced hepatic injury in rats. *J Pharm Pharm Sci* 13, 303-310.

Um, J.H., Park, S.J., Kang, H., Yang, S., Foretz, M., McBurney, M.W., Kim, M.K., Viollet, B., and Chung, J.H. (2010). AMP-activated protein kinase-deficient mice are resistant to the metabolic effects of resveratrol. *Diabetes* 59, 554-563.

Vang, O., Ahmad, N., Baile, C.A., Baur, J.A., Brown, K., Csiszar, A., Das, D.K., Delmas, D., Gottfried, C., Lin, H.Y., Ma, Q.Y., Mukhopadhyay, P., Nalini, N., Pezzuto, J.M., Richard, T., Shukla, Y., Surh, Y.J., Szekeres, T., Szkudelski, T., Walle, T., and Wu, J.M. (2011). What is new for an old molecule? Systematic review and recommendations on the use of resveratrol. *PLoS One* 6, e19881.

Walford, R.L., Harris, S.B., and Gunion, M.W. (1992). The calorically restricted low-fat nutrient-dense diet in Biosphere 2 significantly lowers blood glucose, total leukocyte count, cholesterol, and blood pressure in humans. *Proc Natl Acad Sci U S A* 89, 11533-11537.

Walford, R.L., Mock, D., MacCallum, T., and Laseter, J.L. (1999). Physiologic changes in humans subjected to severe, selective calorie restriction for two years in biosphere 2: health, aging, and toxicological perspectives. *Toxicol Sci* 52, 61-65.

Wang, Z., Al-Regaiey, K.A., Masternak, M.M., and Bartke, A. (2006). Adipocytokines and lipid levels in Ames dwarf and calorie-restricted mice. *J Gerontol A Biol Sci Med Sci* 61, 323-331.

Weir, J.B. (1949). New methods for calculating metabolic rate with special reference to protein metabolism. *J Physiol* 109, 1-9.

Weiss, E.P., Racette, S.B., Villareal, D.T., Fontana, L., Steger-May, K., Schechtman, K.B., Klein, S., and Holloszy, J.O. (2006). Improvements in glucose tolerance and insulin action induced by increasing energy expenditure or decreasing energy intake: a randomized controlled trial. *Am J Clin Nutr* 84, 1033-1042.

Weyer, C., Walford, R.L., Harper, I.T., Milner, M., MacCallum, T., Tataranni, P.A., and Ravussin, E. (2000). Energy metabolism after 2 y of energy restriction: the biosphere 2 experiment. *Am J Clin Nutr* 72, 946-953.

Williamson, D.A., Martin, C.K., York-Crowe, E., Anton, S.D., Redman, L.M., Han, H., and Ravussin, E. (2007). Measurement of dietary restraint: validity tests of four questionnaires. *Appetite* 48, 183-192.

Zhang, H., Morgan, B., Potter, B.J., Ma, L., Dellsperger, K.C., Ungvari, Z., and Zhang, C. (2010). Resveratrol improves left ventricular diastolic relaxation in type 2 diabetes by inhibiting oxidative/nitrative stress: in vivo demonstration with magnetic resonance imaging. *Am J Physiol Heart Circ Physiol* 299, H985-994.

Zheng, Y., Zhang, W., Pendleton, E., Leng, S., Wu, J., Chen, R., and Sun, X.J. (2009). Improved insulin sensitivity by calorie restriction is associated with reduction of ERK and p70S6K activities in the liver of obese Zucker rats. *J Endocrinol* 203, 337-347.