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# The effects of resveratrol on metabolic status in patients with type 2 diabetes mellitus and coronary heart disease

Asma Hoseini<sup>1</sup>, Gholamreza Namazi<sup>1†</sup>, Alireza Farrokhan<sup>2</sup>, Željko Reiner<sup>3</sup>, Esmat Aghadavod<sup>1</sup>, Fereshteh Bahmani<sup>1</sup>, Zatollah Asemi<sup>1\*</sup>

<sup>1</sup> Research Center for Biochemistry and Nutrition in Metabolic Diseases, Kashan University of Medical Sciences, Kashan, Iran

<sup>2</sup> Department of Cardiology, School of Medicine, Kashan University of Medical Sciences, Kashan, Iran

<sup>3</sup> Department of Internal Medicine, University Hospital Centre Zagreb, School of Medicine, University of Zagreb, Zagreb, Croatia

\* Correspondence: Z. Asemi, PhD. Tel: +98 315 546 3378, Email: asemi\_r@yahoo.com

<sup>†</sup> Co-correspondence: E-mail addresses: namazi-gh@kaums.ac.ir

16 Number of words (Text): 2400

## 17 Number of words (Abstract): 245

18 Number of Tables: 3

## 19 Supplemental file: 1

20 Number of Figure: 3

**22 Abstract**

23 This study was performed to investigate the effects of resveratrol on metabolic status in patients  
24 with type 2 diabetes mellitus (T2DM) and coronary heart disease (CHD). This randomized,  
25 double-blind, placebo-controlled trial was performed in 56 patients with T2DM and CHD.  
26 Patients were randomly divided into two groups to receive either 500 mg resveratrol/day (n=28)  
27 or placebo (n=28) for 4 weeks. Resveratrol reduced fasting glucose ( $\beta$  -10.04 mg/dL; 95% CI, -  
28 18.23, -1.86; P=0.01), insulin ( $\beta$  -1.09  $\mu$ IU/mL; 95% CI, -1.93, -0.24; P=0.01) and insulin  
29 resistance ( $\beta$  -0.48; 95% CI, -0.76, -0.21; P=0.001), and significantly increased insulin sensitivity  
30 ( $\beta$  0.006; 95% CI, 0.001, 0.01; P=0.02) when compared with the placebo. Resveratrol also  
31 significantly increased HDL-cholesterol levels ( $\beta$  3.38 mg/dL; 95% CI, 1.72, 5.05; P<0.001) and  
32 significantly decreased total-/HDL-cholesterol ratio ( $\beta$  -0.36; 95% CI, -0.59, -0.13; P=0.002)  
33 when compared with the placebo. Additionally, resveratrol caused a significant increase in total  
34 antioxidant capacity (TAC) ( $\beta$  58.88 mmol/L; 95% CI, 17.33, 100.44; P=0.006) and a significant  
35 reduction in malondialdehyde (MDA) levels ( $\beta$  -0.21  $\mu$ mol/L; 95% CI, -0.41, -0.005; P=0.04)  
36 when compared with the placebo. Resveratrol upregulated PPAR- $\gamma$  (P=0.01) and sirtuin 1  
37 (SIRT1) (P=0.01) in peripheral blood mononuclear cell (PBMC) of T2DM patients with CHD.  
38 Resveratrol supplementation did not have any effect on inflammatory markers. 4-week  
39 supplementation with resveratrol in patients with T2DM and CHD had beneficial effects on  
40 glycemic control, HDL-cholesterol, total-/HDL-cholesterol ratio, TAC and MDA levels.  
41 Resveratrol also upregulated PPAR- $\gamma$  and SIRT1 in PBMC of T2DM patients with CHD.

42 **Keywords:** Resveratrol, coronary heart disease, type 2 diabetes mellitus, HDL-cholesterol,  
43 antioxidant capacity

**45      Introduction**

46      Type 2 diabetes mellitus (T2DM) is one of the most wide spread chronic diseases in almost all  
47      countries and is one of the biggest global public health problems [1, 2]. T2DM increases the risk  
48      of micro- and macrovascular disease [3, 4]. Coronary heart disease (CHD) is one of the major  
49      complications related to T2DM [5, 6]. T2DM is associated with several metabolic complications,  
50      including not only hyperglycemia, but also insulin resistance, dyslipidemia, increased  
51      inflammatory cytokines and oxidative damage [7]. Both T2DM and CHD are multifactorial  
52      caused diseases which, beside the environmental factors, are caused by interference of many  
53      genes having impact on pathogenesis through different mechanisms and pathways [8, 9].

54

55      Resveratrol (3,5,4'-trihydroxy-trans-stilbene) is a naturally occurring polyphenolic compound  
56      originating from food sources, particularly from different plants such as berries, grapes, rhubarb  
57      and peanuts. After it gained popularity in 1992 different possible targets for its pharmacological  
58      effects have been identified [10, 11]. Some observational studies have shown that resveratrol has  
59      several effects in the prevention and treatment of a wide variety of metabolic diseases. These  
60      benefits are wide ranging and according to some indications include anticancer, antioxidant, anti-  
61      inflammatory, anti-diabetic, cardioprotective and even maybe anti-aging effects [12]. Previous  
62      evidence has shown that resveratrol supplementation can decrease the risk of CHD and improve  
63      serum lipids [13, 14]. Moreover, resveratrol is thought to activate sirtuin 1 (SIRT1) gene [15],  
64      inhibiting RelA acetylation and to promote inhibitor protein- $\kappa$ B $\alpha$  degradation, which decreases  
65      nuclear factor kappa B (NF- $\kappa$ B)-induced expression of tumor necrosis-alpha (TNF- $\alpha$ ), interleukin  
66      (IL)-1 $\beta$ , IL(-6), and metalloproteases [16]. A study published several years ago reported that  
67      resveratrol is able to decrease oxidized LDL-cholesterol and apolipoprotein B. In addition, this  
68      study has demonstrated the ability to protect endothelial cells from lipid damage [17]. Resveratrol

69 also increases endothelial nitric oxide (NO) production by upregulation of endothelial nitric oxide  
70 synthase (eNOS) expression, enhancement of eNOS enzymatic activity, and prevention of eNOS  
71 uncoupling [18]. It reduces endothelial oxidative stress, and endothelin-1 synthesis but it reduces  
72 oxidative stress in smooth muscle cells as well, it inhibits smooth muscle cell proliferation, it  
73 prevents arterial stiffness and vascular remodeling which can all contribute to its antiatherogenic  
74 effects and prevention of CHD [19]. However, some studies did not find any significant change  
75 on inflammatory markers, glycemic control and serum lipids in patients with T2DM after  
76 resveratrol supplementation [20].

77

78 Resveratrol possibly improves heart function by influencing inflammatory markers and  
79 oxidation process, platelet aggregation and endothelial function. This polyphenolic flavonoid has  
80 also effects on metabolic modulation [21]. Since resveratrol increases SIRT1 expression it  
81 improves glucose homeostasis and decreases insulin resistance [22, 23] but it also improves  
82 cardiac function through SIRT1 activation [24]. SIRT1 protects the heart against oxidative  
83 stress by regulating many substrates, such as tumor necrosis factor (TNF- $\alpha$ ), nuclear factor- $\kappa$ B  
84 (NF- $\kappa$ B) and p53 [25-27]. Thus, we assumed that in diabetic patients with CHD, resveratrol may  
85 have beneficial effects on metabolic profile. Few studies investigated the effects of resveratrol  
86 supplementation in T2DM patients, but data on patients with diabetes who have CHD are scarce.  
87 The purpose of this study was to investigate the effects of resveratrol supplementation during 4  
88 weeks on glycemic control, serum lipids, biomarkers of inflammation and oxidative stress  
89 patients with diabetes and CHD.

90

## 91 **Methods**

### 92 **Study population**

93 This study was a randomized, double-blind, placebo-controlled trial, registered in the Iranian  
94 registry of clinical trials (<http://www.irct.ir>: IRCT20181029041490N1). It was performed at a  
95 cardiology clinic affiliated to Kashan University of Medical Sciences (KAUMS), Kashan, Iran,  
96 between September 2018 and March 2019. All experiments were performed in accordance with  
97 the Guidelines Helsinki. Experiments were approved by the ethics committee at KAUMS, and  
98 Iran. Informed consents were obtained from human participants of this study. Inclusion criteria  
99 were as follows: patients with T2DM, aged 40-85 years old with proven 2- and 3-vessel CHD.  
100 Diagnosis of T2DM was made according to the criteria of American Diabetes Association [28].  
101 Exclusion criteria were: consuming resveratrol three months prior to the intervention, taking  
102 antioxidant and/or anti-inflammatory supplements such as vitamin E and vitamin C, having an  
103 acute myocardial infarction or cardiac surgery in the past three months, or having renal or hepatic  
104 failure.

105

106 **Study design**

107 Participants were randomly allocated (balanced block randomization) into two groups to take  
108 either 500 mg/day resveratrol (Nutrissence, Las Vegas, USA) or placebo (Barij Essence, Kashan,  
109 Iran) (n=28 in each group) for 4 weeks. Due to the lack of evidence about the appropriate dosage  
110 of resveratrol for patients with diabetes and CHD, we used duration and the above-mentioned  
111 dose of resveratrol based on previous studies in chronic kidney disease patients [29], healthy  
112 young adults [30] and patients with ulcerative colitis [31]. Color, shape, size, and package of  
113 placebo and resveratrol were identical. Randomization assignment was conducted using  
114 computer-generated random numbers. Randomization and allocation were blinded for the  
115 investigators and patients until the final analyses were completed. The randomization and  
116 enrolling participants were performed by trained staff at the cardiology clinic. Compliance with

117 the consumption of placebo and resveratrol was performed by counting the remaining capsules in  
118 the capsule containers. All patients completed 3-day dietary intake records at week 1 and 4 of  
119 intervention. For obtaining patients' nutrient intakes according to 3-day food records, Nutritionist  
120 IV software (First Databank, San Bruno, CA), which was adapted for Iranian food pattern was  
121 applied. Anthropometric measures (Seca balances, Hamburg, Germany) were recorded at the  
122 beginning of the study and after the 4-week intervention in the cardiology clinic by a trained  
123 nutritionist.

124

## 125 **Outcomes**

126 Insulin resistance was considered as the primary outcome, but fasting plasma glucose (FPG),  
127 insulin, the quantitative insulin sensitivity check index (QUICKI), serum lipids, gene expression  
128 of interleukin-1 (IL-1), peroxisome proliferator-activated receptor gamma (PPAR- $\gamma$ ), tumor  
129 necrosis factor alpha (TNF- $\alpha$ ), transforming growth factor beta (TGF- $\beta$ ) and CIRT1, and  
130 biomarkers of inflammation and oxidative stress were considered as secondary outcomes. Fasting  
131 blood (15 mL) samples were taken at baseline and after the 4-week intervention at Kashan  
132 Reference Laboratory. Insulin levels were determined by ELISA kit (DiaMetra, Milano, Italy)  
133 with inter-assay and intra-assay coefficient variances (CVs) below 5%. The homeostasis model of  
134 assessment-insulin resistance (HOMA-IR) and QUICKI were calculated in accordance with the  
135 standard formula [32]. Enzymatic kits (Pars Azmun, Tehran, Iran) were used to evaluate FPG and  
136 serum lipids with inter- and intra-assay CVs below 5%. CRP levels were assessed by an ELISA  
137 kit (LDN, Nordhorn, Germany) with inter- and intra-assay CVs below 7%. Plasma total  
138 antioxidant capacity (TAC) was determined using the method reported by Benzie and Strain [33],  
139 total glutathione (GSH) by using the method described by Beutler et al. [34], and MDA

140 concentrations were determined by the spectrophotometric test [35] with inter- and intra-assay  
141 CVs below 5%.

142

### 143 **Isolation of lymphocytes, RNA extraction and cDNA synthesis**

144 Lymphocytes were isolated using 50% percoll solution (Sigma-Aldrich, Dorset, UK) gradient by  
145 centrifugation for 20 min and 3000 rpm at 4°C [36]. Total RNA was extracted based on acid  
146 guanidinium-phenol-chloroform procedure using RNX™-plus reagent (Cinnacolon, Tehran, Iran)  
147 according to the manufacturer's instructions. RNAs was treated with DNAase I (Fermentas,  
148 Lithuania) for the elimination of any genomic DNA contamination. Three micrograms of RNA  
149 were used for cDNA synthesis with random hexamer and oligo (dT) 18 primers through  
150 RevertAid™ Reverse Transcriptase (Fermantase, Canada) in total 20 µl reaction mixture [36].

151

### 152 **Real-time PCR analysis**

153 Appropriate primers for IL-1, PPAR- $\gamma$ , TNF- $\alpha$ , TGF- $\beta$ , SIRT1 and glyceraldehyde-3 phosphate  
154 dehydrogenase were designed (**Table 1**). Quantitative Real-time PCR was performed by the  
155 LightCycler® 96 sequence detection systems (Roche Diagnostics, Rotkreuz, Switzerland) using 4  
156 µl of 5× EVA GREEN I master mix (Salise Biodyne, Japan), 10 ng cDNA, 200 nM of each  
157 forward and reverse primers in final volume of 20 µl.

158

### 159 **Statistical methods and sample size**

160 Sample size formula for randomized clinical trial were used, where type one ( $\alpha$ ) and type two  
161 errors ( $\beta$ ) were 0.05, and 0.20 (power=80%), respectively. In a recent study [37], 0.97 as the SD  
162 and 0.78 as the change in mean (d) of HOMA-IR were used. According to the power calculation,

163 in each group 25 patients were needed; after allowing for 5 dropouts in each group, the final  
164 sample size was 30 patients in each group.

165  
166 The Kolmogorov-Smirnov test was used for checking the normality of data. To determine the  
167 differences in anthropometric measures and dietary intakes between two groups, the independent-  
168 samples *t*-test was used. Multiple linear regression models were used to evaluate treatment  
169 impacts on study outcomes after adjusting for confounding parameters. The effect sizes were  
170 presented as the mean differences with 95% confidence intervals. Pearson Chi-square test was  
171 applied for comparison of categorical variables. P values <0.05 were considered significant. The  
172 Statistical Package for Social Science version 18 (SPSS Inc., Chicago, Illinois, USA) were used  
173 for statistical analyses.

174  
175 **Results**  
176 Fifty-six patients [resveratrol (n=28) and placebo (n=28)] completed the trial (**Fig.1**). The  
177 compliance rate was high; both groups took more than 90% of capsules during the trial. No  
178 adverse effects were reported in T2DM patients with CHD while consuming resveratrol.

179  
180 No significant differences were seen between the groups regarding mean age, height, and weight  
181 and BMI at baseline and end-of-trial (**Table 2**). Mean smoking rate, treatment with antidiabetic  
182 (65-70% monotherapy and 30-35% combination therapy) and lipid-lowering drugs (100%  
183 statins), hypertension rate (70-78%), treatment with angiotensin converting enzymes inhibitors  
184 (ACEI) (100%), aldosterone receptor blockers (ARB) (100%), β-blockers (90-95%) and calcium  
185 channel blockers (5-10%) were not different between the two groups (Data not shown).

186

187 Macronutrient and micronutrient intake as calculated based on 3-days food record was not  
188 significantly different between the resveratrol and control group (**Supplemental file 1**).

189  
190 After the 4-week intervention, resveratrol significantly reduced FPG ( $\beta$  -10.04 mg/dL; 95% CI, -  
191 18.23, -1.86; P=0.01), insulin ( $\beta$  -1.09  $\mu$ IU/mL; 95% CI, -1.93, -0.24; P=0.01) and HOMA-IR ( $\beta$   
192 -0.48; 95% CI, -0.76, -0.21; P=0.001), and significantly increased QUICKI ( $\beta$  0.006; 95% CI,  
193 0.001, 0.01; P=0.02) when compared with the placebo (**Table 3**). In addition, resveratrol  
194 administration significantly increased serum HDL-cholesterol levels ( $\beta$  3.38 mg/dL; 95% CI,  
195 1.72, 5.05; P<0.001) and significantly decreased total-/HDL-cholesterol ratio ( $\beta$  -0.36; 95% CI, -  
196 0.59, -0.13; P=0.002) when compared with placebo. Resveratrol also caused a significant  
197 elevation in TAC ( $\beta$  58.88 mmol/L; 95% CI, 17.33, 100.44; P=0.006) and a significant reduction  
198 in plasma MDA levels ( $\beta$  -0.21  $\mu$ mol/L; 95% CI, -0.41, -0.005; P=0.04) when compared with the  
199 placebo. Resveratrol supplementation did not affect other metabolic parameters.

200  
201 Resveratrol upregulated PPAR- $\gamma$  (P=0.01) and SIRT1 (P=0.01) in peripheral blood mononuclear  
202 cell (PBMC) of T2DM patients with CHD, but it did not affect gene expression for IL-1, TNF- $\alpha$   
203 and TGF- $\beta$  (**Fig.2 &3**).

204  
205 **Discussion**  
206 In this study, which to the best of our knowledge is the first of its kind, 4-week supplementation  
207 with resveratrol in diabetic patients with CHD had beneficial effects on glycemic control, HDL-  
208 cholesterol, total-/HDL-cholesterol ratio, TAC and MDA. In addition, resveratrol upregulated  
209 PPAR- $\gamma$  and SIRT1 in PBMC of T2DM patients with CHD.

210

## 211 Effects on glycemic control and serum lipids

212 Our study showed that resveratrol significantly decreased FPG, insulin levels, HOMA-IR score  
213 and total-/HDL-cholesterol ratio, and increased QUICKI and HDL-cholesterol levels without  
214 affecting other lipids and lipoproteins in patients with T2DM and CHD. In addition, resveratrol  
215 upregulated PPAR- $\gamma$  and SIRT1 in PBMC of T2DM patients with CHD. T2DM and CHD are  
216 frequently associated with dyslipidemia and insulin resistance [38, 39]. Similar to our findings, in  
217 another recent study resveratrol at a dosage of 480 mg/day after 4 weeks in T2DM patients  
218 significantly decreased HOMA-IR, insulin levels, but did not influence triglycerides, total- and  
219 LDL-cholesterol levels [37]. Resveratrol supplementation in overweight pregnant women after 60  
220 days improved glucose homeostasis parameters, but also total cholesterol, LDL-cholesterol and  
221 triglycerides [40]. Moreover, a 12-week resveratrol administration (500 mg) three times per day  
222 in patients with metabolic syndrome decreased insulin concentrations [41]. In contrast to our  
223 findings, in another study a 12-week resveratrol supplementation (100 mg/day) did not affect  
224 FPG and HDL-cholesterol in T2DM patients [42]. Also, 6-month resveratrol supplementation at  
225 dosage 40 mg/day or 500 mg/day did not improve fasting glucose, glycated hemoglobin, insulin,  
226 C-peptide, free fatty acids, adiponectin nor interleukin-6 in T2DM patients when compared with  
227 placebo [43]. Resveratrol (200 mg/day) did not influence postprandial gene expression of PPAR-  
228  $\gamma$  and NF- $\kappa$ B, neither did it modify postprandial variations in circulating inflammatory markers  
229 (C-reactive protein, IL-6, IL-8, monocyte chemoattractant protein-1) and adhesion molecules  
230 (soluble E-selectin, soluble vascular cell adhesion molecule-1, soluble intercellular adhesion  
231 molecule-1) when compared to placebo [44]. The beneficial effect of resveratrol on serum lipids  
232 may be mediated by its phenolic hydroxyls which cause oxidation of unsaturated fatty acids and  
233 decrease of circulating total and LDL-cholesterol, triglycerides and apo AI as shown in  
234 hyperlipidemic mice [45]. As mentioned earlier, anti-atherosclerotic activity of resveratrol

235 probably involves also enhanced activity of PPAR- $\alpha$  [46], suppression of platelet aggregation  
236 [47], very moderate decrease of systolic blood pressure [48], and improvement of endothelial  
237 activity [49] which is all important particularly in T2DM patients with CVD who have very high  
238 risk for a CVD event. The novelty of our approach was that our patients were exactly those with  
239 very high risk having both T2DM and proven CVD. These patients were not analyzed previously  
240 concerning the outcomes which we have analyzed.

241

#### 242 **Effects on biomarkers of inflammation and oxidative stress**

243 This study showed that taking resveratrol supplements during 4 weeks causes a significant  
244 increase in plasma TAC and a significant reduction in MDA levels in T2DM patients with CHD,  
245 but did not influence other biomarkers of inflammation and oxidative damage. The novelty of our  
246 study is that it was performed on patients with both T2DM and CVD which has not been done  
247 before. In accordance with our findings, several experimental studies on animals have shown that  
248 resveratrol causes a significant decrease in MDA levels [50, 51]. 6 weeks of resveratrol at dosage  
249 500 mg/day in patients with ulcerative colitis significantly increased TAC and reduced MDA  
250 levels [52]. The results of a trial with 12-weeks resveratrol supplementation (600 mg/day)  
251 suggested that, unlike the results of our study, resveratrol did not modify antioxidative status in  
252 patients with nonalcoholic fatty liver disease [53]. Some researchers failed to detect any  
253 significant effects of resveratrol on biomarkers of inflammation and oxidative damage in obese  
254 men and non-obese women with normal glucose tolerance [54, 55]. In diabetic patients, oxidative  
255 damage and elevated inflammatory mediators are associated with endothelial dysfunction and the  
256 progression of macrovascular disease [56]. As already stated, resveratrol can protect endothelial  
257 cells from lipid damage, promote vasodilation via modulation of NO synthesis, and can inhibit  
258 platelet aggregation [57]. Resveratrol can also inhibit kappa B inhibitor kinase, which

259 subsequently prevents the translocation of NF- $\kappa$ B into the nucleus and the activation of the  
260 respective genes encoding cytokines, chemoattractant and adhesion molecules, and proliferating  
261 signaling molecules [58] as well as reduce oxidative damage through activation of sirtuin-  
262 activating proteins, in particular SIRT-1 [59].

263

264 It has been reported that deletion of SIRT1 in hepatocytes results in increased local  
265 inflammation [60]. In an *in vivo* study, overexpression of SIRT1 significantly reduced hepatic  
266 expression of TNF- $\alpha$  and IL-6 after chronic high-fat feeding [61]. The antioxidant and/or anti-  
267 inflammatory effects of resveratrol play an important role in control of NF- $\kappa$ B activation  
268 or chromatin remodeling by modulation of histone deacetylase (sirtuins) activity and  
269 subsequently inflammatory gene expression in lung epithelial cells [62]. SIRT1 and its activators  
270 might regulate the efficiency of the NF- $\kappa$ B signaling [63]. Several potential mechanisms by  
271 which resveratrol might inhibit TNF- $\alpha$ -mediated inflammation or insulin resistance in adipocytes  
272 include: 1) suppressing TNF- $\alpha$ -TNF receptor (TNFR) signaling, 2) interfering with TNF- $\alpha$   
273 receptor (TNFR) binding, or 3) altering the activity of proteins involved in inflammation or lipid  
274 and insulin metabolism [64]. Some studies have documented that polyphenols increase PPAR- $\gamma$   
275 expression or activity [65, 66]. Therefore, resveratrol may reduce inflammation or insulin  
276 resistance by directly interfering with TNF- $\alpha$  signaling or indirectly by activating PPAR- $\gamma$  [64].  
277 Another potential mechanism by which resveratrol might inhibit inflammation and insulin  
278 resistance could be via inhibition of TNF- $\alpha$ -mediated reactive oxygen species (ROS) production  
279 [67]. ROS increase inflammatory gene expression by activating redox-sensitive proteins,  
280 including redox-sensitive transcription factors such as NF- $\kappa$ B, activator protein 1 and NF-E2-  
281 related factor-2 [68] and apoptosis signal-regulating kinase-1/thioredoxin [69]. Increased ROS  
282 production by NAPDH oxidase and reduced antioxidative enzymes such as glutathione

283 peroxidase (GPx) and superoxide dismutase (SOD) contribute to inflammation and insulin  
284 resistance [70]. Resveratrol seems to attenuate inflammation by inducing gene expression of  
285 antioxidative enzymes such as SOD, heme oxygenase-1 and GPx [68]. Sakurai et al.[71]  
286 indicated that oligomerized grape seed polyphenols significantly decreased inflammation by  
287 suppressing ROS production and NF- $\kappa$ B activation. All these effects of resveratrol might explain  
288 its beneficial activity on glycemic control, HDL-cholesterol, TAC and MDA levels in T2DM  
289 patients, particularly those with CVD.

290

291 This study has some limitations. We did not measure resveratrol levels before and after the  
292 intervention. The next limitation was that gene expression related to lipid and oxidative damage  
293 in patients with T2DM and CHD were not evaluated. Resveratrol and placebo were provided two  
294 various companies. This should be considered in the interpretation of our findings.

295

296 **Conclusions**

297 The 4-week supplementation with resveratrol in patients with T2DM and CHD had beneficial  
298 impacts on glycemic control, HDL-cholesterol, total-/HDL-cholesterol ratio, TAC and MDA  
299 levels. Resveratrol also upregulated PPAR- $\gamma$  and SIRT1 in PBMC of T2DM patients with CHD.

300

**302 Abbreviations**

303 FPG, fasting plasma glucose; GSH, total glutathione; HOMA-IR, homeostasis model of  
304 assessment-insulin resistance; HDL-cholesterol, high density lipoprotein-cholesterol; CRP, C-  
305 reactive protein; LDL-cholesterol, low density lipoprotein-cholesterol; MDA, malondialdehyde;  
306 QUICKI, quantitative insulin sensitivity check index; VLDL-cholesterol, very low density  
307 lipoprotein-cholesterol; TAC, total antioxidant capacity; GAPDH, glyceraldehyde-3-Phosphate  
308 dehydrogenase; IL-1, interleukin-1; PPAR- $\gamma$ , peroxisome proliferator-activated receptor gamma;  
309 TNF- $\alpha$ , tumor necrosis factor alpha; TGF- $\beta$ , transforming growth factor beta; T2DM, type 2  
310 diabetes mellitus; CHD, coronary heart disease; NF- $\kappa$ B, nuclear factor kappa B; PBMC,  
311 peripheral blood mononuclear cell; eNOS, endothelial nitric oxide synthase; SIRT1, sirtuin 1.

312

**313 Declarations:****314 Ethics approval and consent to participate**

315 All procedures performed in studies involving human participants were in accordance with the  
316 ethical standards of the institutional and national research committee and with the 1964 Helsinki  
317 declaration and its later amendments.

318

**319 Consent for publication**

320 Not applicable.

321

**322 Availability of data and material**

323 The primary data for this study is available from the authors on direct request.

324

**325 Competing interests**

326 The authors declare no conflict of interest.

327

328 **Funding**

329 The research grant provided by Research Deputy of Kashan University of Medical Sciences  
330 (KAUMS).

331

332 **Author contributions**

333 ZA and AH: Conception, design, and statistical analysis, drafting of the manuscript and  
334 supervised the study.

335 GN, AF, ZR, EA and FB: data collection and manuscript drafting.

336

337 **Acknowledgements**

338 The present study was supported by a grant from the Vice-chancellor for Research, KAUMS,  
339 Kashan, and Iran.

340

341 **Clinical trial registration number**

342 www.irct.ir: <http://www.irct.ir/> IRCT20181029041490N1.

## References

1. Ghaffari MA, Askari Sede S, Rashtchizadeh N, Mohammadzadeh G, Majidi S. Association of CRP gene polymorphism with CRP levels and Coronary Artery Disease in Type 2 Diabetes in Ahvaz, southwest of Iran. *BioImpacts* : BI. 2014;4:133-9.
2. Whiting DR, Guariguata L, Weil C, Shaw J. IDF diabetes atlas: global estimates of the prevalence of diabetes for 2011 and 2030. *Diabetes research and clinical practice*. 2011;94:311-21.
3. Ali MK, Bullard KM, Saaddine JB, Cowie CC, Imperatore G, Gregg EW. Achievement of goals in U.S. diabetes care, 1999-2010. *The New England journal of medicine*. 2013;368:1613-24.
4. Sacks FM, Hermans MP, Fioretto P, Valensi P, Davis T, Horton E, et al. Association between plasma triglycerides and high-density lipoprotein cholesterol and microvascular kidney disease and retinopathy in type 2 diabetes mellitus: a global case-control study in 13 countries. *Circulation*. 2014;129:999-1008.
5. Peix A, Cabrera LO, Heres F, Rodriguez L, Valdes A, Valiente J, et al. Interrelationship between myocardial perfusion imaging, coronary calcium score, and endothelial function in asymptomatic diabetics and controls. *Journal of nuclear cardiology : official publication of the American Society of Nuclear Cardiology*. 2011;18:398-406.
6. Gyberg V, De Bacquer D, De Backer G, Jennings C, Kotseva K, Mellbin L, et al. Patients with coronary artery disease and diabetes need improved management: a report from the EUROASPIRE IV survey: a registry from the EuroObservational Research Programme of the European Society of Cardiology. *Cardiovasc Diabetol*. 2015;14:133.
7. Yahagi K, Kolodgie FD, Lutter C, Mori H, Romero ME, Finn AV, et al. Pathology of Human Coronary and Carotid Artery Atherosclerosis and Vascular Calcification in Diabetes Mellitus. *Arteriosclerosis, thrombosis, and vascular biology*. 2017;37:191-204.
8. Alikhah A, Pahlevan Kakhki M, Ahmadi A, Dehghanad R, Boroumand MA, Behmanesh M. The role of lnc-DC long non-coding RNA and SOCS1 in the regulation of STAT3 in coronary artery disease and type 2 diabetes mellitus. *Journal of diabetes and its complications*. 2018;32:258-65.
9. Merkler M, Reiner Z. The burden of hyperlipidaemia and diabetes in cardiovascular diseases. *Fundamental & clinical pharmacology*. 2007;21 Suppl 2:1-3.

- Published on 08 August 2019. Downloaded by Nottingham Trent University on 8/11/2019 11:14:58 AM.
10. Murgia D, Mauceri R, Campisi G, De Caro V. Advance on Resveratrol Application in Bone Regeneration: Progress and Perspectives for Use in Oral and Maxillofacial Surgery. *Biomolecules*. 2019;9.
  11. Sahebkar A. Effects of resveratrol supplementation on plasma lipids: a systematic review and meta-analysis of randomized controlled trials. *Nutr Rev*. 2013;71:822-35.
  12. Chu LM, Lassaletta AD, Robich MP, Sellke FW. Resveratrol in the prevention and treatment of coronary artery disease. *Current atherosclerosis reports*. 2011;13:439-46.
  13. Magyar K, Halmosi R, Palfi A, Feher G, Czopf L, Fulop A, et al. Cardioprotection by resveratrol: A human clinical trial in patients with stable coronary artery disease. *Clinical hemorheology and microcirculation*. 2012;50:179-87.
  14. Tome-Carneiro J, Gonzalvez M, Larrosa M, Yanez-Gascon MJ, Garcia-Almagro FJ, Ruiz-Ros JA, et al. Grape resveratrol increases serum adiponectin and downregulates inflammatory genes in peripheral blood mononuclear cells: a triple-blind, placebo-controlled, one-year clinical trial in patients with stable coronary artery disease. *Cardiovasc Drugs Ther*. 2013;27:37-48.
  15. Bagul PK, Deepthi N, Sultana R, Banerjee SK. Resveratrol ameliorates cardiac oxidative stress in diabetes through deacetylation of NFkB-p65 and histone 3. *The Journal of nutritional biochemistry*. 2015;26:1298-307.
  16. Malaguarnera L. Influence of Resveratrol on the Immune Response. *Nutrients*. 2019;11.
  17. Tome-Carneiro J, Gonzalvez M, Larrosa M, Garcia-Almagro FJ, Aviles-Plaza F, Parra S, et al. Consumption of a grape extract supplement containing resveratrol decreases oxidized LDL and ApoB in patients undergoing primary prevention of cardiovascular disease: a triple-blind, 6-month follow-up, placebo-controlled, randomized trial. *Molecular nutrition & food research*. 2012;56:810-21.
  18. Xia N, Forstermann U, Li H. Resveratrol and endothelial nitric oxide. *Molecules*. 2014;19:16102-21.
  19. Li H, Xia N, Hasselwander S, Daiber A. Resveratrol and Vascular Function. *Int J Mol Sci*. 2019;20.
  20. Tome-Carneiro J, Larrosa M, Yanez-Gascon MJ, Davalos A, Gil-Zamorano J, Gonzalvez M, et al. One-year supplementation with a grape extract containing resveratrol modulates inflammatory-related microRNAs and cytokines expression in peripheral blood mononuclear

cells of type 2 diabetes and hypertensive patients with coronary artery disease. *Pharmacol Res.* 2013;72:69-82.

21. Das SK, Patel VB, Oudit GY. Beneficial effects of grape resveratrol on serum adiponectin and inflammation: clinical trial in patients with stable coronary artery disease: editorial to: "Grape resveratrol increases serum adiponectin and downregulates inflammatory genes in peripheral blood mononuclear cells: a triple-blind, placebo-controlled, one-year clinical trial in patients with stable coronary artery disease" by J. Tome-Carneiro et al. *Cardiovascular drugs and therapy.* 2013;27:1-4.
22. Szkudelski T, Szkudelska K. Resveratrol and diabetes: from animal to human studies. *Biochimica et biophysica acta.* 2015;1852:1145-54.
23. Szkudelska K, Szkudelski T. Resveratrol, obesity and diabetes. *Eur J Pharmacol.* 2010;635:1-8.
24. Raj P, Louis XL, Thandapilly SJ, Movahed A, Zieroth S, Netticadan T. Potential of resveratrol in the treatment of heart failure. *Life sciences.* 2014;95:63-71.
25. Cattelan A, Ceolotto G, Bova S, Albiero M, Kuppusamy M, De Martin S, et al. NAD(+) -dependent SIRT1 deactivation has a key role on ischemia-reperfusion-induced apoptosis. *Vascular pharmacology.* 2015;70:35-44.
26. Shimoyama Y, Mitsuda Y, Tsuruta Y, Suzuki K, Hamajima N, Niwa T. SIRTUIN 1 gene polymorphisms are associated with cholesterol metabolism and coronary artery calcification in Japanese hemodialysis patients. *Journal of renal nutrition : the official journal of the Council on Renal Nutrition of the National Kidney Foundation.* 2012;22:114-9.
27. Wang P, Du B, Yin W, Wang X, Zhu W. Resveratrol attenuates CoCl<sub>2</sub>-induced cochlear hair cell damage through upregulation of Sirtuin1 and NF-kappaB deacetylation. *PloS one.* 2013;8:e80854.
28. Diagnosis and classification of diabetes mellitus. *Diabetes Care.* 2014;37 Suppl 1:S81-90.
29. Saldanha JF, Leal VO, Rizzetto F, Grimmer GH, Ribeiro-Alves M, Daleprane JB, et al. Effects of Resveratrol Supplementation in Nrf2 and NF-kappaB Expressions in Nondialyzed Chronic Kidney Disease Patients: A Randomized, Double-Blind, Placebo-Controlled, Crossover Clinical Trial. *J Ren Nutr.* 2016;26:401-6.

30. Polley KR, Jenkins N, O'Connor P, McCully K. Influence of exercise training with resveratrol supplementation on skeletal muscle mitochondrial capacity. *Appl Physiol Nutr Metab.* 2016;41:26-32.
31. Samsami-Kor M, Daryani NE, Asl PR, Hekmatdoost A. Anti-Inflammatory Effects of Resveratrol in Patients with Ulcerative Colitis: A Randomized, Double-Blind, Placebo-controlled Pilot Study. *Arch Med Res.* 2015;46:280-5.
32. Pisprasert V, Ingram KH, Lopez-Davila MF, Munoz AJ, Garvey WT. Limitations in the use of indices using glucose and insulin levels to predict insulin sensitivity: impact of race and gender and superiority of the indices derived from oral glucose tolerance test in African Americans. *Diabetes Care.* 2013;36:845-53.
33. Benzie IF, Strain JJ. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay. *Anal Biochem.* 1996;239:70-6.
34. Beutler E, Gelbart T. Plasma glutathione in health and in patients with malignant disease. *J Lab Clin Med.* 1985;105:581-4.
35. Janero DR. Malondialdehyde and thiobarbituric acid-reactivity as diagnostic indices of lipid peroxidation and peroxidative tissue injury. *Free Radic Biol Med.* 1990;9:515-40.
36. Gmelig-Meyling F, Waldmann TA. Separation of human blood monocytes and lymphocytes on a continuous Percoll gradient. *J Immunol Methods.* 1980;33:1-9.
37. Zare Javid A, Hormoznejad R, Yousefimanesh HA, Zakerkish M, Haghghi-Zadeh MH, Dehghan P, et al. The Impact of Resveratrol Supplementation on Blood Glucose, Insulin, Insulin Resistance, Triglyceride, and Periodontal Markers in Type 2 Diabetic Patients with Chronic Periodontitis. *Phytotherapy research : PTR.* 2017;31:108-14.
38. Raygan F, Rezavandi Z, Dadkhah Tehrani S, Farrokhanian A, Asemi Z. The effects of coenzyme Q10 administration on glucose homeostasis parameters, lipid profiles, biomarkers of inflammation and oxidative stress in patients with metabolic syndrome. *Eur J Nutr.* 2016;55:2357-64.
39. Razzaghi R, Pourbagheri H, Momen-Heravi M, Bahmani F, Shadi J, Soleimani Z, et al. The effects of vitamin D supplementation on wound healing and metabolic status in patients with diabetic foot ulcer: A randomized, double-blind, placebo-controlled trial. *J Diabetes Complications.* 2017;31:766-72.

- Published on 08 August 2019. Downloaded by Nottingham Trent University on 8/11/2019 11:14:58 AM.
40. Malvasi A, Kosmas I, Mynbaev OA, Sparic R, Gustapane S, Guido M, et al. Can trans resveratrol plus d-chiro-inositol and myo-inositol improve maternal metabolic profile in overweight pregnant patients? *Clin Ter.* 2017;168:e240-e7.
41. Mendez-del Villar M, Gonzalez-Ortiz M, Martinez-Abundis E, Perez-Rubio KG, Lizarraga-Valdez R. Effect of resveratrol administration on metabolic syndrome, insulin sensitivity, and insulin secretion. *Metab Syndr Relat Disord.* 2014;12:497-501.
42. Imamura H, Yamaguchi T, Nagayama D, Saiki A, Shirai K, Tatsuno I. Resveratrol Ameliorates Arterial Stiffness Assessed by Cardio-Ankle Vascular Index in Patients With Type 2 Diabetes Mellitus. *International heart journal.* 2017;58:577-83.
43. Bo S, Ponzo V, Ciccone G, Evangelista A, Saba F, Goitre I, et al. Six months of resveratrol supplementation has no measurable effect in type 2 diabetic patients. A randomized, double blind, placebo-controlled trial. *Pharmacol Res.* 2016;111:896-905.
44. Vors C, Couillard C, Paradis ME, Gigleux I, Marin J, Vohl MC, et al. Supplementation with Resveratrol and Curcumin Does Not Affect the Inflammatory Response to a High-Fat Meal in Older Adults with Abdominal Obesity: A Randomized, Placebo-Controlled Crossover Trial. *J Nutr.* 2018;148:379-88.
45. Xie HC, Han HP, Chen Z, He JP. A study on the effect of resveratrol on lipid metabolism in hyperlipidemic mice. *African journal of traditional, complementary, and alternative medicines : AJTCAM.* 2014;11:209-12.
46. Iannelli P, Zarrilli V, Varricchio E, Tramontano D, Mancini FP. The dietary antioxidant resveratrol affects redox changes of PPARalpha activity. *Nutr Metab Cardiovasc Dis.* 2007;17:247-56.
47. Shen MY, Hsiao G, Liu CL, Fong TH, Lin KH, Chou DS, et al. Inhibitory mechanisms of resveratrol in platelet activation: pivotal roles of p38 MAPK and NO/cyclic GMP. *British journal of haematology.* 2007;139:475-85.
48. Timmers S, Konings E, Bilet L, Houtkooper RH, van de Weijer T, Goossens GH, et al. Calorie restriction-like effects of 30 days of resveratrol supplementation on energy metabolism and metabolic profile in obese humans. *Cell metabolism.* 2011;14:612-22.
49. Zhang H, Zhang J, Ungvari Z, Zhang C. Resveratrol improves endothelial function: role of TNF{alpha} and vascular oxidative stress. *Arterioscler Thromb Vasc Biol.* 2009;29:1164-71.

50. Wang XL, Li T, Li JH, Miao SY, Xiao XZ. The Effects of Resveratrol on Inflammation and Oxidative Stress in a Rat Model of Chronic Obstructive Pulmonary Disease. *Molecules* (Basel, Switzerland). 2017;22.
51. Guo Y, Wang A, Liu X, Li E. Effects of resveratrol on reducing spermatogenic dysfunction caused by high-intensity exercise. *Reproductive biology and endocrinology : RB&E*. 2019;17:42.
52. Samsamikor M, Daryani NE, Asl PR, Hekmatdoost A. Resveratrol Supplementation and Oxidative/Anti-Oxidative Status in Patients with Ulcerative Colitis: A Randomized, Double-Blind, Placebo-controlled Pilot Study. *Archives of medical research*. 2016;47:304-9.
53. Asghari S, Rafraf M, Farzin L, Asghari-Jafarabadi M, Ghavami SM, Somi MH. Effects of Pharmacologic Dose of Resveratrol Supplementation on Oxidative/Antioxidative Status Biomarkers in Nonalcoholic Fatty Liver Disease Patients: A Randomized, Double-Blind, Placebo-Controlled Trial. *Adv Pharm Bull*. 2018;8:307-17.
54. Poulsen MM, Vestergaard PF, Clasen BF, Radko Y, Christensen LP, Stokilde-Jorgensen H, et al. High-dose resveratrol supplementation in obese men: an investigator-initiated, randomized, placebo-controlled clinical trial of substrate metabolism, insulin sensitivity, and body composition. *Diabetes*. 2013;62:1186-95.
55. Yoshino J, Conte C, Fontana L, Mittendorfer B, Imai S, Schechtman KB, et al. Resveratrol supplementation does not improve metabolic function in nonobese women with normal glucose tolerance. *Cell Metab*. 2012;16:658-64.
56. Kaur R, Kaur M, Singh J. Endothelial dysfunction and platelet hyperactivity in type 2 diabetes mellitus: molecular insights and therapeutic strategies. *Cardiovascular diabetology*. 2018;17:121.
57. Mohar DS, Malik S. The Sirtuin System: The Holy Grail of Resveratrol? *Journal of clinical & experimental cardiology*. 2012;3.
58. Ungvari Z, Bagi Z, Feher A, Recchia FA, Sonntag WE, Pearson K, et al. Resveratrol confers endothelial protection via activation of the antioxidant transcription factor Nrf2. *Am J Physiol Heart Circ Physiol*. 2010;299:H18-24.
59. Orlandi I, Stamerra G, Strippoli M, Vai M. During yeast chronological aging resveratrol supplementation results in a short-lived phenotype Sir2-dependent. *Redox biology*. 2017;12:745-54.

- Published on 08 August 2019. Downloaded by Nottingham Trent University on 8/11/2019 11:14:58 AM.
60. Purushotham A, Schug TT, Xu Q, Surapureddi S, Guo X, Li X. Hepatocyte-specific deletion of SIRT1 alters fatty acid metabolism and results in hepatic steatosis and inflammation. *Cell Metab.* 2009;9:327-38.
61. Pfluger PT, Herranz D, Velasco-Miguel S, Serrano M, Tschoop MH. Sirt1 protects against high-fat diet-induced metabolic damage. *Proc Natl Acad Sci U S A.* 2008;105:9793-8.
62. Rahman I, Marwick J, Kirkham P. Redox modulation of chromatin remodeling: impact on histone acetylation and deacetylation, NF-kappaB and pro-inflammatory gene expression. *Biochem Pharmacol.* 2004;68:1255-67.
63. Salminen A, Kauppinen A, Suuronen T, Kaarniranta K. SIRT1 longevity factor suppresses NF-kappaB -driven immune responses: regulation of aging via NF-kappaB acetylation? *BioEssays : news and reviews in molecular, cellular and developmental biology.* 2008;30:939-42.
64. Chuang CC, Martinez K, Xie G, Kennedy A, Bumrungpert A, Overman A, et al. Quercetin is equally or more effective than resveratrol in attenuating tumor necrosis factor- $\alpha$ -mediated inflammation and insulin resistance in primary human adipocytes. *Am J Clin Nutr.* 2010;92:1511-21.
65. Saito T, Abe D, Sekiya K. Sakuranetin induces adipogenesis of 3T3-L1 cells through enhanced expression of PPARgamma2. *Biochem Biophys Res Commun.* 2008;372:835-9.
66. Shin DW, Kim SN, Lee SM, Lee W, Song MJ, Park SM, et al. (-)-Catechin promotes adipocyte differentiation in human bone marrow mesenchymal stem cells through PPAR gamma transactivation. *Biochem Pharmacol.* 2009;77:125-33.
67. Goossens V, De Vos K, Vercammen D, Steemans M, Vancompernolle K, Fiers W, et al. Redox regulation of TNF signaling. *Biofactors.* 1999;10:145-56.
68. Rahman I, Biswas SK, Kirkham PA. Regulation of inflammation and redox signaling by dietary polyphenols. *Biochem Pharmacol.* 2006;72:1439-52.
69. Matsukawa J, Matsuzawa A, Takeda K, Ichijo H. The ASK1-MAP kinase cascades in mammalian stress response. *Journal of biochemistry.* 2004;136:261-5.
70. Furukawa S, Fujita T, Shimabukuro M, Iwaki M, Yamada Y, Nakajima Y, et al. Increased oxidative stress in obesity and its impact on metabolic syndrome. *J Clin Invest.* 2004;114:1752-61.

71. Sakurai T, Kitadate K, Nishioka H, Fujii H, Kizaki T, Kondoh Y, et al. Oligomerized grape seed polyphenols attenuate inflammatory changes due to antioxidative properties in coculture of adipocytes and macrophages. *J Nutr Biochem.* 2010;21:47-54.

**Table 1.** Specific primers used for real-time quantitative PCR

Gene	Primer	Product size (bp)	Annealing temperature (C)
GAPDH	F: AAGCTCATTCCCTGGTATGACAACG R: TCTTCCTTTGTGCTCTTGCTGG	126	61.3
PPAR- $\gamma$	F: ATGACAGACCTCAGACAGATTG R: AATGTTGGCAGTGGCTCAG	210	54
IL-1	F: GCTTCTCTCTGGTCCTTGG R: AGGGCAGGGTAGAGAAAGAG	174	56
TNF- $\alpha$	F: GTCAACCTCCTCTGCCAT R: CCAAAGTAGACCTGCCAGA	188	52
TGF- $\beta$	F: TTGAGACTTTCCGTTGCCG R: CGAGGTCTGGGGAAAAGTCT	227	56
SIRT1	F: GGCAAAGGAGCAGATTAGTAGG R: CTCTGGAACATCAGGCTCATC	22	58

GAPDH, glyceraldehyde-3-Phosphate dehydrogenase; IL-1, interleukin-1; PPAR- $\gamma$ , peroxisome proliferator-activated receptor gamma; SIRT1, sirtuin 1; TNF- $\alpha$ , tumor necrosis factor alpha; TGF- $\beta$ , transforming growth factor beta.

**Table 2.** General characteristics of study participants at baseline study

	Placebo group (n=28)	Resveratrol group (n=28)
Age (y)	63.3±10.1	61.0±8.6
Height (cm)	168.6±5.0	170.0±5.8
Weight at study baseline (kg)	79.9±11.0	82.9±10.9
Weight at end-of-trial (kg)	79.9±11.1	82.7±10.8
Weight change (kg)	-0.01±0.9	-0.2±1.2
BMI at study baseline (kg/m <sup>2</sup> )	28.1±3.4	28.6±3.1
BMI at end-of-trial (kg/m <sup>2</sup> )	28.1±3.5	28.5±3.0
BMI change (kg/m <sup>2</sup> )	0.001±0.3	-0.1±0.4

Data are means± SDs.

**Table 3.** The effect of resveratrol supplementation on metabolic status in type 2 diabetic patients with coronary heart disease

Variables	Placebo group (n=28)		Resveratrol group (n=28)		Difference in outcome measures between resveratrol and placebo treatment groups <sup>1</sup>	
	Baseline	Week 4	Baseline	Week 4	$\beta$ (95% CI)	P <sup>2</sup>
FPG (mg/dL)	133.9±52.2	136.6±50.0	129.5±56.2	122.5±53.1	-10.04 (-18.23, -1.86)	0.01
Insulin ( $\mu$ IU/mL)	13.1±4.2	12.7±4.3	12.4±2.7	11.0±2.7	-1.09 (-1.93, -0.24)	0.01
HOMA-IR	4.3±2.5	4.1±2.5	4.0±2.0	3.4±1.7	-0.48 (-0.76, -0.21)	0.001
QUICKI	0.31±0.01	0.31±0.02	0.31±0.01	0.32±0.01	0.006 (0.001, 0.01)	0.02
Triglycerides (mg/dL)	147.5±60.7	149.9±52.9	155.7±62.9	148.3±57.8	-8.47 (-18.78, 1.84)	0.10
VLDL-cholesterol (mg/dL)	29.5±12.1	30.0±10.6	31.1±12.6	29.7±11.6	-1.69 (-3.75, 0.36)	0.10
Total cholesterol (mg/dL)	137.7±26.7	137.6±23.2	145.9±29.4	140.1±28.6	-3.77 (-11.74, 4.18)	0.34
LDL-cholesterol (mg/dL)	68.7±21.0	68.6±21.1	78.3±21.1	70.9±20.3	-4.83 (-12.20, 2.54)	0.19
HDL-cholesterol (mg/dL)	39.5±5.9	39.1±5.4	36.4±6.5	39.5±7.7	3.38 (1.72, 5.05)	<0.001
Total-/HDL-cholesterol ratio	3.5±0.8	3.6±0.7	4.1±1.1	3.7±1.0	-0.36 (-0.59, -0.13)	0.002
CRP (mg/L)	3.9±1.9	4.1±1.5	4.2±1.5	4.2±1.0	0.10 (-0.46, 0.26)	0.58
TAC (mmol/L)	924.2±118.9	919.3±146.1	906.9±103.7	960.9±123.9	58.88 (17.23, 100.44)	0.006
GSH ( $\mu$ mol/L)	568.4±82.3	614.3±106.6	520.9±101.5	555.4±119.9	-28.56 (-82.81, 25.68)	0.29
MDA ( $\mu$ mol/L)	2.9±0.5	2.8±0.4	3.1±0.9	2.8±0.8	-0.21 (-0.41, -0.005)	0.04

Data are mean  $\pm$ SDs.

<sup>1</sup>"Outcome measures" refers to the change in values of measures of interest between baseline and week 4.  $\beta$  [difference in the mean outcomes measures between treatment groups (resveratrol group = 1 and placebo group = 0)].

<sup>2</sup> Obtained from multiple regression model (adjusted for baseline values of each biochemical variables).

CRP, C-reactive protein; FPG, fasting plasma glucose; GSH, glutathione; HOMA-IR, homeostasis model of assessment-estimated insulin resistance; MDA, malondialdehyde; QUICKI, quantitative insulin sensitivity check index; TAC, total antioxidant capacity.

**Legend to figure:**

**Fig.1.** Summary of patient flow diagram.

**Fig.2.** Fold change (means $\pm$  SDs) in gene expression levels of PPAR- $\gamma$ , IL-1 and TNF- $\alpha$  in study participants who were received resveratrol supplements and placebo  
P value was obtained from independent *t*-test. N=28 in each group.

**Fig.3.** Change (means $\pm$  SDs) in gene expression levels of TGF- $\beta$  and cirtuin 1 in study participants who were received resveratrol supplements and placebo  
P value was obtained from independent *t*-test. N=28 in each group.

IL-1, interleukin-1; PPAR- $\gamma$ , peroxisome proliferator-activated receptor gamma; TNF- $\alpha$ , tumor necrosis factor alpha; TGF- $\beta$ , transforming growth factor beta.

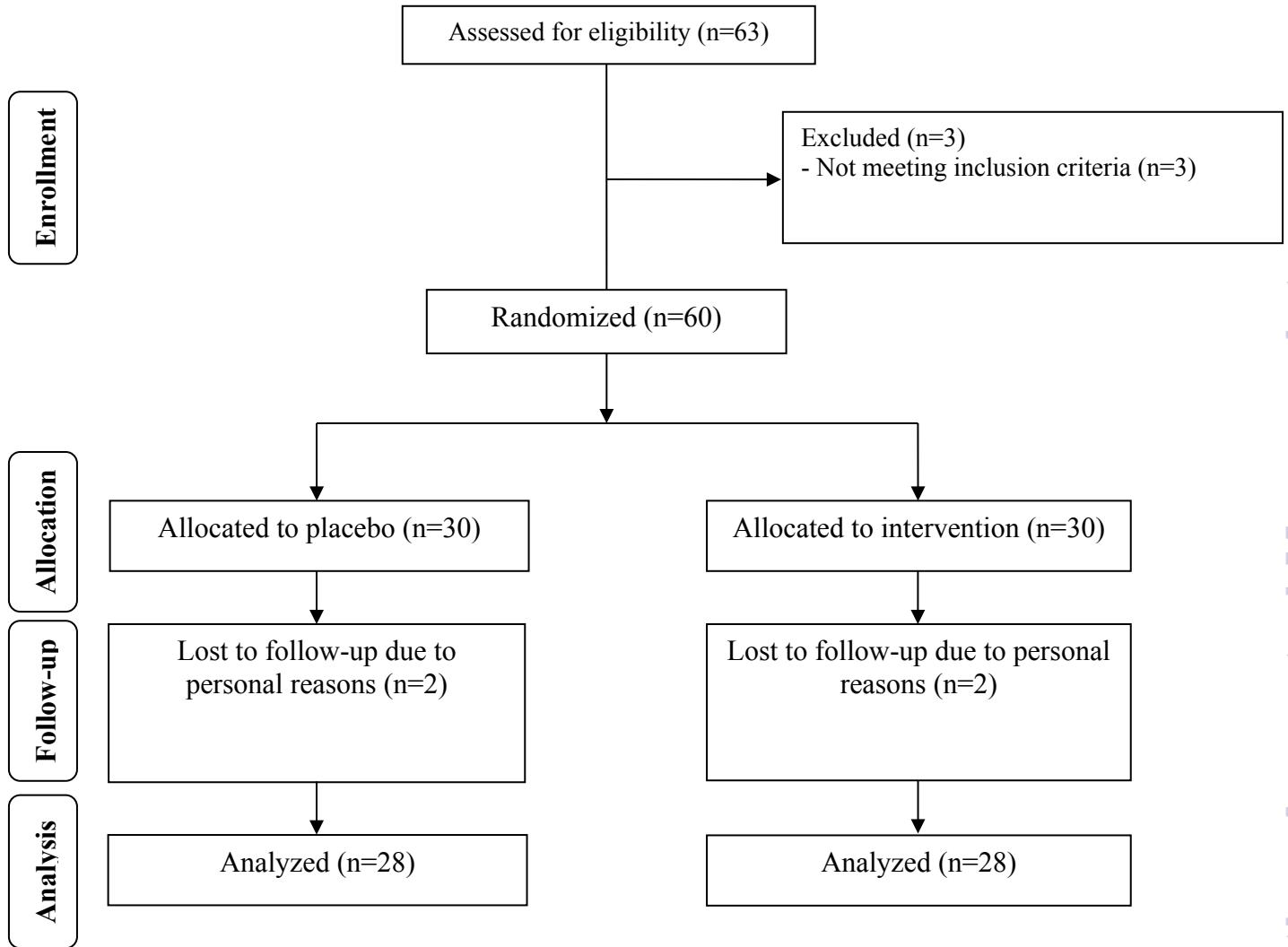


Fig.1

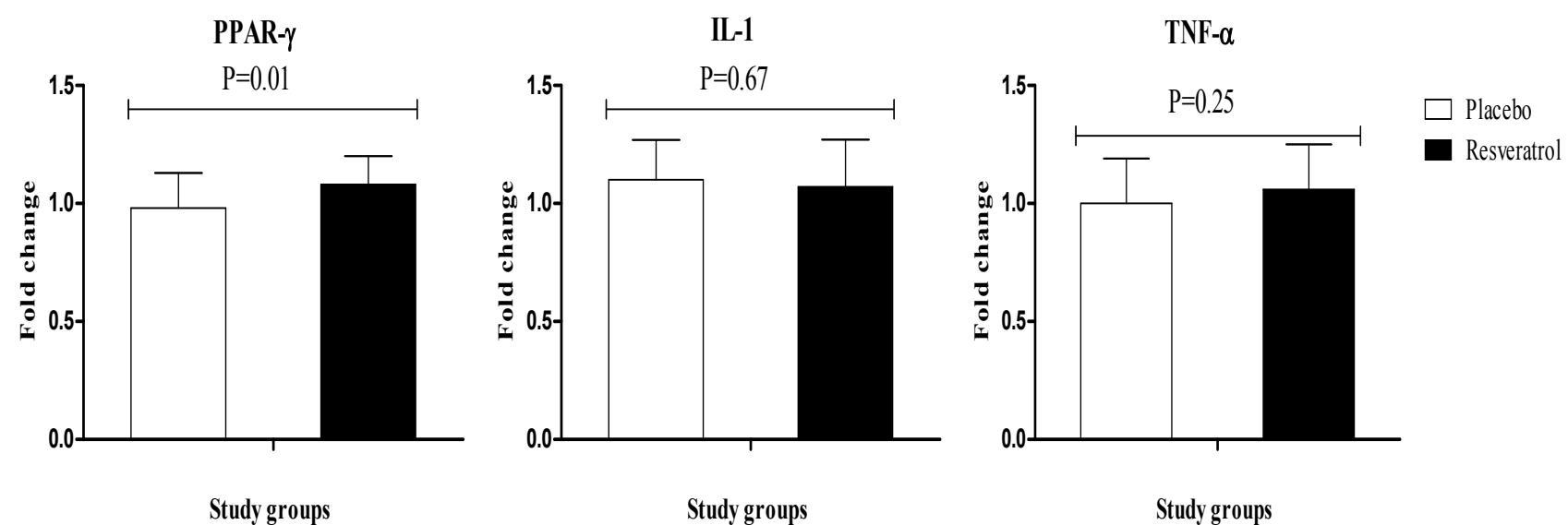


Fig.2

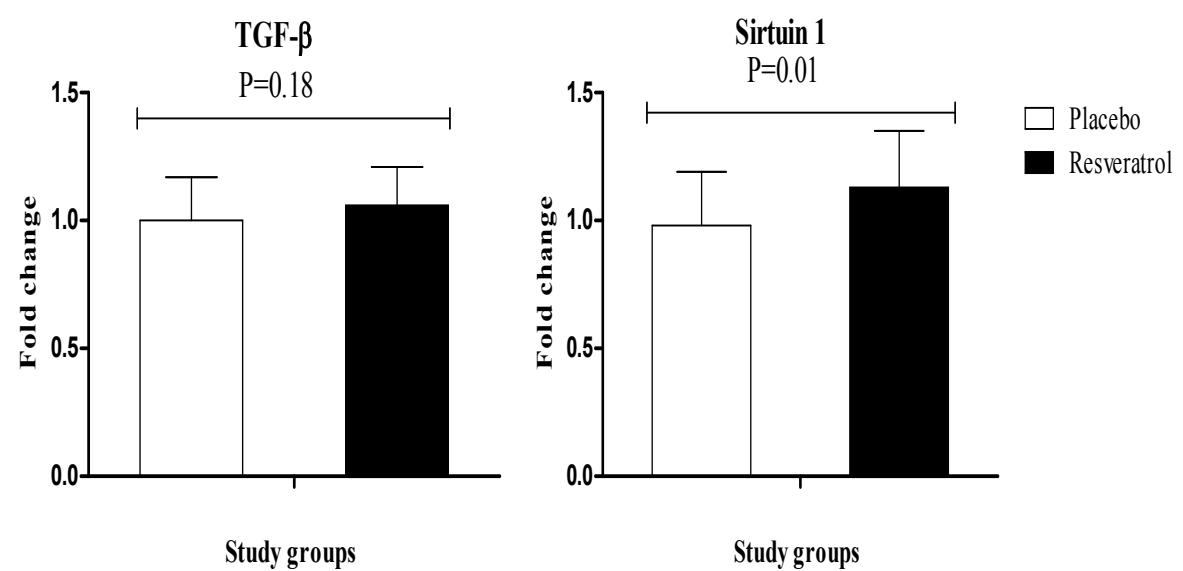


Fig.3