Bayesian determination of the effect of a deep euctectic solvent on the structure of lipid monolayers

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Deep eutectic solvents present a novel class of non-aqueous room temperature solvent with tunable properties, that are capable of promoting the self-assembly of surfactant molecules. However, the solvation model in these systems still challenges the classic understanding of amphiphilicity. In this work, we present the first example of the self-assembly of phospholipid monolayers at the interface between air and a non-aqueous solvent. Furthermore, we use novel, chemically-consistent Bayesian modelling of X-ray and neutron reflectometry measurements to show the ability of the deep eutectic solvent to interact with the phosphatidylglycerol lipid head, leading to an apparent increase in the volume compared to that observed in water. No such change was observed for the phosphocholine head, indicating that the interaction is head specific.

Deep eutectic solvents (DES) are a green, sustainable solvent system obtained through the complexation of naturally occurring compounds, such as sugars, alcohols, amines, and carboxylic acids[1, 2]. An extensive hydrogen bonding network is present, allowing the mixture to remain liquid at room temperature[3–5]. Through different combinations of the precursor materials, it is possible to tune the solvent's physicochemical properties, such as polarity[6], viscosity and surface tension[1], network charge[7], and hydrophobicity[8, 9]. Furthermore, recently DES have indicated the presence of a solvophobic effect through the promotion of surfactant micelle formation[10, 11], phospholipid bilayer formation[12–14], and the ability to stabilise non-ionic polymer conformations[15] and protein conformation[16].

Phospholipid monolayer formation at the air/liquid interface plays a key role in many biological and technological processes, where the solvent-specific solubility of different components of the phospholipid enables a stable monolayer to form at the interface[17]. Investigations at the air/salt-water interface have identified the importance that interactions between charged phospholipid heads and ions present in solution have on the structure, monomer packing and stability of the monolayer[17, 18]. Despite the broad interest in these systems, the presence of stable phospholipid monolayers at the interface between air and a non-aqueous media has not been previously reported, to the best of the authors' knowledge.

This work presents the first investigation of the structure of four phospholipid monolayers at the air-DES interface, as determined by chemically-consistent modelling of X-ray reflectometry (XRR) measurements. Evaluating the interaction of two, chemically distinct, phospholipid heads with this non-aqueous solvent, in addition to the effect of the tail chain length. The analysis was then extended to model complementary neutron reflectometry (NR) measurements for two contrasts of DMPC and DPPC at a single surface pressure.

Recent developments in computational resources and software have enabled powerful methodologies and algorithms to be harnessed by those from non-expert backgrounds. This has benefitted significantly from open-source software projects such as the Python language[19] and the Jupyter notebooks framework[20]. In the area of NR and XRR data-analysis, the landscape of open-source software is diverse, with a range of software packages available from a variety of sources; refnx[21], MOTOFIT[22], Aurore[23], and GenX[24].

The use of a Python library, such as refnx, enables the implementation custom models that contain chemically-relevant information as well as the application of probability distribution function (PDF) sampling techniques. The Python library emcee[25] offers refnx to access the Goodman & Weare Affine Invariant Markov chain Monte Carlo (MCMC) Ensemble method[26]. This allows the sampling of the high-dimensionality parameter space, relevant in reflectometry analysis, in a Bayesian fashion, where the new samples are generated with consideration of those sampled previously[27]. Bayesian inference gives an understanding of the PDF for the fitted parameters and therefore estimations of their inverse uncertainties and inter-parameter correlations.

We developed a chemically-consistent model (detail in the ESI) that allowed for the co-refinement of XRR and NR measurements at different surface concentration. The model was applied to the study of four phospholipids monolayers, namely 1,2-dipalmitoyl-sn-glycero-3-phosphocholine (DPPC, C_{16} tails), 1,2-dimyristoyl-sn-glycero-3-phosphocholine (DMPC, C_{14} tails), 1,2-dilauroyl-sn-glycero-3-phosphocholine (DLPC, C_{12} tails) and 1,2-dimyristoyl-sn-glycero-3-phospho-(1'-rac-glycerol) (DMPG, C_{14} tails), at the air-1:2 choline chloride:glycerol mixture interface, by XRR. Unlike monolayer models applied previously[17, 18, 28–32], our model made no assumption of the volume of the lipid head, V_h, or tail, V_t. Instead these parameters, along with the thickness of the phospholipid head layer, d_h, were allowed to vary for each lipid. However, their values were constained to be self-consistent for a single lipid over multiple measurements at different surface pressures.

This model was required, despite the general consensus that the volume of the phosphocholine (PC) head is $320\,\text{Å}^3$ to $360\,\text{Å}^3$, while the phosphatidylglycerol (PG) head is $289\,\text{Å}^3$ to $291\,\text{Å}^3$ (see Table SI), as it not clear if the presence of the DES would affect the head volume due to electrostatic interactions, which may differ from those between the head and water. Furthermore, the effect of pressure, which is not necessarily accounted for the in the literature, has been shown to compress the lipid tail volume[33, 34], and the need to account for this hydrocarbon compaction according to their phase in the modelling of NR data has recently been demonstrated[35].

The custom model for each lipid, fitted to the experimental XRR data, and the associated SLD profiles are shown in Figure 2. Table I presents the mean value, and asymmetric uncertainties that correspond to a 95 % confidence interval, for the PDF for each of the varying parameters; the tail tile angle, θ_t, and the interfacial

FIG. 2. The XRR profiles (left) and SLD profiles (right) for each of the four lipids; (a) DLPC, (b) DMPC, (c) DPPC, (d) DMPG, at the four measured surface pressures; lowest surface pressure at the bottom and increasing moving up. The different surface pressure XRR profiles have been offset in the y-axis by an order of magnitude and SLD profiles offset in the y-axis by $5 \times 10^{-6}\,\text{Å}^{-2}$, for clarity.

roughness, σ, which were free to vary independent of surface pressure. In addition to the head and tail volumes, V_h and V_t respectively, and the head layer thickness, d_h, for which a single value was fitted for each lipid across all surface pressures.

In keeping this previous literature, [17, 36], the tail layer thickness, d_t, increases with the number of hydrocarbon atoms present. Furthermore, the thickness of the tail layers agress well with values found for water-analogues; $13.72^{+0.01}_{-0.01}$ Å at $30\,\text{mN}\,\text{m}^{-1}$ in DES compared with $d_t = 15.8\,\text{Å}$ at $30\,\text{mN}\,\text{m}^{-1}$[29] in water for DMPC, and $16.91^{+0.01}_{-0.01}$ Å at $30\,\text{mN}\,\text{m}^{-1}$ in DES compared with $d_t = 16.7\,\text{Å}$ at $40\,\text{mN}\,\text{m}^{-1}$[31] in water for DPPC.

Figure 3 shows the tail layer thickness increasing with surface pressure, before plateuing; for DPPC this occurs at $20\,\text{mN}\,\text{m}^{-1}$, DMPC at $30\,\text{mN}\,\text{m}^{-1}$ and for DMPG and DLPC the plateu can be assumed to be at higher pressures than those studied. This phenomenon has been
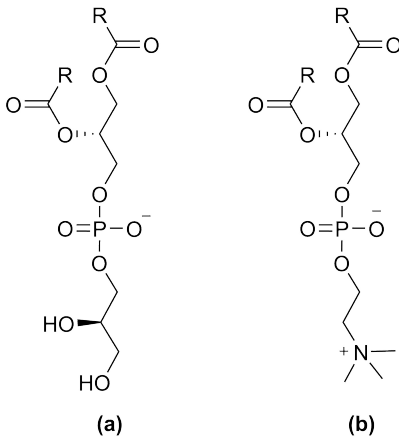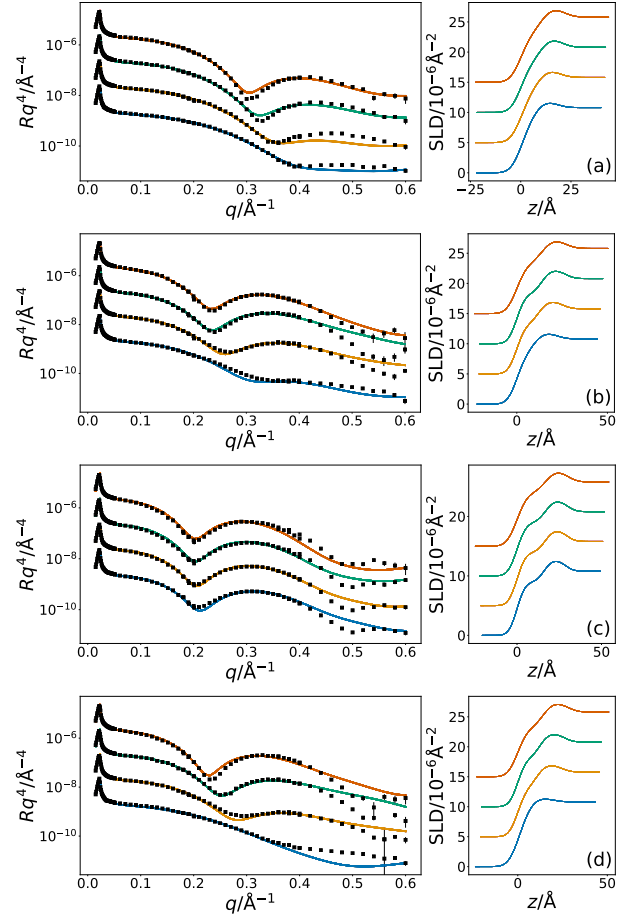

FIG. 1. The two lipid classes with different head groups compared in this study, where R indicates the hydrocarbon tail; (a) phosphatidylglycerol (PG), (b) phosphocholine (PC).

TABLE I. The best-fit values, and associated 95 % confidence intervals for the varying parameters in the XRR models, at the highest surface pressure (SP) measured. The values of d_t were found from the appropriate values of θ_t using Eqn. S1 and the values for ϕ_h were obtained from the appropriate use of Eqn. S3.

| Lipid	DLPC	DMPC	DPPC	DMPG
SP/mN m^{-1}	35	40	30	30
$\theta_t/°$	$48.22^{+0.20}_{-0.20}$	$39.78^{+0.06}_{-0.07}$	$34.47^{+0.07}_{-0.07}$	$38.92^{+0.06}_{-0.06}$
$\sigma/Å$	$4.34^{+0.02}_{-0.02}$	$4.53^{+0.01}_{-0.01}$	$4.90^{+0.00}_{-0.00}$	$4.44^{+0.01}_{-0.01}$
$V_t/Å^3$	$625.78^{+3.45}_{-3.66}$	$718.75^{+0.53}_{-0.53}$	$765.31^{+0.39}_{-0.37}$	$734.03^{+0.60}_{-0.59}$
$V_h/Å^3$	$331.30^{+0.60}_{-0.62}$	$339.54^{+0.28}_{-0.28}$	$321.99^{+0.25}_{-0.26}$	$329.94^{+0.34}_{-0.34}$
$d_h/Å$	$11.02^{+0.13}_{-0.12}$	$13.21^{+0.04}_{-0.04}$	$12.70^{+0.03}_{-0.03}$	$13.95^{+0.03}_{-0.04}$
$\phi_h/\times10^{-2}$	$50.53^{+1.12}_{-1.10}$	$50.57^{+0.24}_{-0.24}$	$43.96^{+0.24}_{-0.22}$	$54.93^{+0.20}_{-0.20}$
$d_t/Å$	$10.30^{+0.04}_{-0.04}$	$13.82^{+0.01}_{-0.01}$	$16.91^{+0.01}_{-0.01}$	$13.99^{+0.01}_{-0.01}$
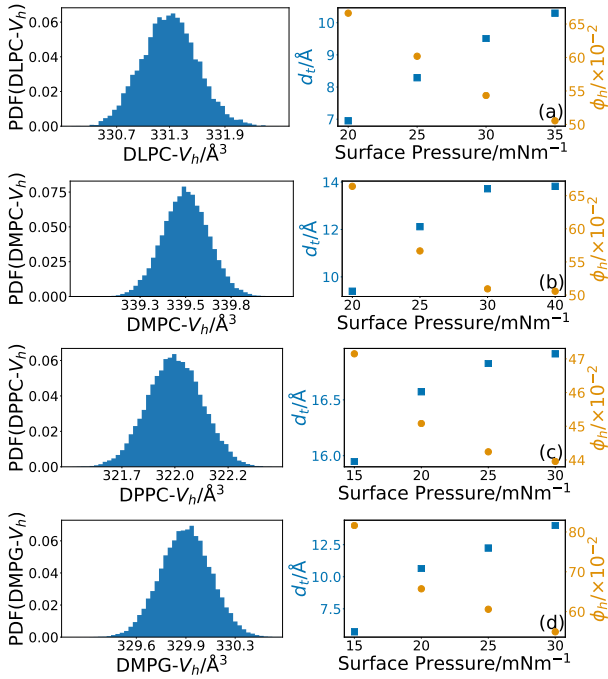
FIG. 3. The PDFs of the head volume (left) and variation of d_t (blue squares) and ϕ_h (yellow circles) with surface pressure for each of the four lipids; (a) DLPC, (b) DMPC, (c) DPPC, (d) DMPG. The values of d_t were found from the appropriate values of θ_t using Eqn. S1.

noted before for DMPC[28] and DPPC[35] at the air-water interface.

In Figure 3, it is clear that for all four lipids, as the surface pressure is increased there is a corresponding decrease in the percentage solvent, ϕ_h, present in the lipid head layer. This can be rationalised by considering when the surface pressure is increased, the free volume available between the lipid heads reduces forcing the solvent out of the lipid head layer and into the bulk. A similar effect has been observed when increasing the surface pressure from $11\,\text{mN m}^{-1}$ to $31\,\text{mN m}^{-1}$ for a DMPC/DMPG

monolayer at the air-water interface[28].

When comparing Tables SI and I it is clear that the volume of the lipid tails are significantly lower in the current measurements than found previously, by other techniques. It is unlikely that this is a result of the DES subphase, due to the hydrophobic nature of the lipid tails. Such a reduction has been shown previously[35], where it was rationalised by the compaction of the monolayer at elevated surface pressure. In that work, the optimal value of the tail volume for DPPC was found to be $772\,Å^3$ at a surface pressure of $35\,\text{mN m}^{-1}$, this agrees well with the value of $765.31^{+0.39}_{-0.37}\,Å^3$ found in this work at surface pressures of 15, 20, 25, and $30\,\text{mN m}^{-1}$.

In this work, a single tail volume was fitted to each lipid for all four surface pressures that were measured. This is based on the assumption, that at all four surface pressures, the lipids adopt the same liquid expanded phase (as discussed Section **??**) and therefore any variation in the structure with surface pressure would manifest only as a change in the tail thickness, via the tail tilt angle. It is clear when comparing Tables SI and I that some of the tail volumes are reduced in the current XRR measurements compared to those determined previously. The reduction was found to be 8 % to 12 % for DPPC, DMPC and DLPC when compared with literature sources at $24\,°\text{C}$ to $30\,°\text{C}$, this is in good agreement with the maximum compression percentage of 15 % noted by Small *et al.*[34]. DMPG shows a small increase in the tail volume relative to the literature value quoted at lower temperature. Notably our value is similar to that found for DMPC on DES, which has the same tail structure and suggests that our results are at least self-consistent.

Figure 3 shows the PDFs for the head volume for each of the four lipids. The three lipids with the PC head are consistent with values of around $330\,Å^3$, regardless of hydrocarbon tail. This agrees well with the values found for the same head in water (Table SI). Interestingly, the volume for the PG head is similar to that for the PC head with a value of $329.94^{+0.34}_{-0.34}\,Å^3$, whereas it is considered to be smaller in water when measured by either DMPG using differential vibrating tube densimetry[37] ($291\,Å^3$) or POPG using molecular dynamics simulations[38] ($289\,Å^3$). This indicates that there may be some effect arising from the solvation in DES causing an apparent increase in the PG head volume when compared with water.

The major difference between the two heads is the fact PG head is negatively charged whereas the PC head is zwitterionic (Figure 1). It has been shown previously that, due to the electrostatic interaction, the conformation of the PC head is folded in water[39]. A similar structure may occur for the PG head, with a weaker interaction between the partially postively-charged alcoholic hydrogen atoms and the negatively-charged phosphate group. Therefore, the observed increase found for the PG volume in DES when compared with water may be due to
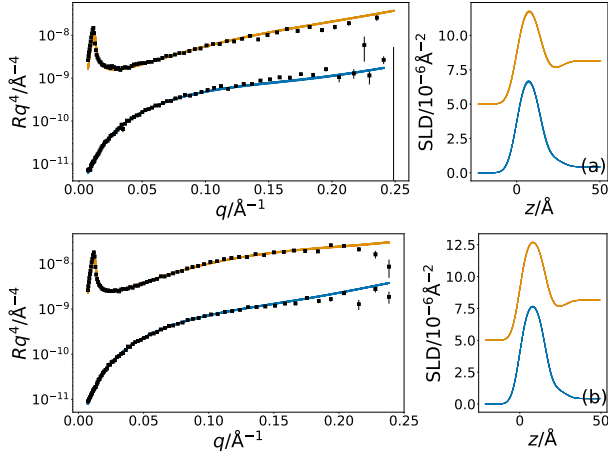
FIG. 4. The NR and SLD profiles at a surface pressure of $20\,\mathrm{mNm^{-1}}$, the h-DES contrast is shown in blue while the hd-DES in yellow; (a) DMPC, (b) DPPC. The NR profiles have been offset in the y-axis by an order of magnitude and SLD profiles offset in the y-axis by $5\times10^{-6}\,\mathrm{\AA^{-2}}$, for clarity.

TABLE II. he best-fit values, and associated 95 % confidence intervals for the varying parameters in the co-refined NR models. The values of d_t were found from the appropriate values of θ_t using Eqn. S1, and the values of ϕ_h were found using Eqn. S3.

Lipid	d$_{54}$-DMPC		d$_{62}$-DPPC	
SP/mNm^{-1}	20	25	15	20
$\theta_t/^\circ$	$38.98^{+0.75}_{-0.76}$	$24.65^{+0.06}_{-0.01}$	$53.11^{+0.46}_{-0.46}$	$40.66^{+0.42}_{-0.42}$
$\sigma_{t,h,s}/\mathrm{\AA}$	$4.41^{+0.16}_{-0.16}$	$2.51^{+0.03}_{-0.01}$	$4.27^{+0.16}_{-0.17}$	$3.98^{+0.10}_{-0.11}$
$\phi_h/\times10^{-2}$	$50.00^{+0.53}_{-0.54}$	$41.54^{+0.01}_{-0.03}$	$59.20^{+0.43}_{-0.43}$	$48.44^{+0.33}_{-0.33}$
$d_t/\mathrm{\AA}$	$13.98^{+0.15}_{-0.15}$	$16.35^{+0.00}_{-0.01}$	$12.31^{+0.13}_{-0.13}$	$15.56^{+0.10}_{-0.10}$

the unfolding of the PG head, where the solvent is capable of providing a greater screening effect to the PG head than that present in water. This may not be observed for the PC head due to the greater strength of the folding interaction arising from the formally-charged nature of the ammonium group. It would be anticipated that this unfolding would result in an increase in the thickness of the lipid head layer. Previously, DPPG has been reported to have a head layer thickness of $(10.3\pm0.4)\,\mathrm{\AA}$ at $22\,\mathrm{mN\,m^{-1}}$ from NR measurements[30], which it slightly less than the $13.95^{+0.03}_{-0.04}$ angstrom determined in the current work, further suggesting that the unfolding of the PG head. Previously, the presence of short range, electrostatic interactions has been noted to impact the solution structure adopted by zwitterionic surfactants in choline chloride:glycerol[40].

The determined volumes for the head and tails, and the thickness of the head layers, for DMPC and DPPC were then used as constraints in the co-refinement of the custom model against two contrasts NR data (Figure 4). In these analysis the only two varying parameters were the tail tilt angle, θ_t, and the interfacial roughness, σ. The quality of agreement with the NR measurements indicates that the values found for the head and tail volumes are consistent between the pair of measurements for the same system. It is clear, that again stable monolayers of the lipids are forming at the air-DES interface, and that the head and tail volumes determined from XRR measurements are robust-enough to be used in the modelling of NR data. Futhermore, the trends present with varying surface pressure in the XRR models are consistent with that found in the NR models.

For the first time, stable phosphocholine and phosphatidylglycerol lipid monolayers have been observed and characterised on a non-aqueous liquid surface. Until the emergence of ionic liquids and DES, only a limited number of molecular solvents exhibited the ability to promote self-assembly and, to the best of our knowledge, only water among those had demonstrated the formation of functional phospholipid monolayers at the air-liquid interface.

A physically and chemically constrained modelling approach and Bayesian analysis method was used to rationalise these measurements showing that the structures are remarkably similar at the air-DES interface to those previously observed at the air-water interface. This has the important implication that DES therefore offers the possibility of performing studies of model membranes in the absence of water. Such applications may include fundamental investigations of phospholipid monolayers in extreme environments (total or partial absence of water, cryogenic temperatures), protein membrane interactions and development of new technologies for drug delivery. However, the fact remains that the PG head containing lipid did show a significant difference; having a larger head volume than observed for the same system in water. This shows that the transfer of lipids to a DES is not just a simple substitution of the subphase. In this specific case we have proposed an explanation based on unfolding of the PG head that is enabled by electrostatic screening of the head charges by the charged solvent.

The ability to determine the head volume was facilitated by access to easy to use, and open-source software that allowed for the straightforward use a custom, chemically-consistent model within the analysis of the XRR and NR measurements. Futhermore, this work presents the first, to our knowledge, use of chemically-consistent parameterisation to co-refine XRR measurements at different surface concentrations.

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