

PRACTICAL 6

Question 1. Using AL2CO server (<http://prodata.swmed.edu/al2co/al2co.php>), obtain the positional conservation scores from multiple sequence alignment (MSA) of given set of protein sequences (set1 and set2) using the methods given below:

- (i) Unweighted frequency and entropy-based measure
- (ii) Unweighted frequency and variance-based measure
- (iii) Unweighted frequency and sum of pairs measure
- (iv) Weighted frequency and variance-based measure
- (v) Normalize the scores obtained with (i)

Sequences:

Set 1: P69905, P01946, P01942, P01966, P01958, P01959, P01965, P06635, P60529, P80043, P01980

Set 2: TPIS_HUMAN, TPIS_YEAST, TPIS_GRAGA, TPIS_TRYCR, TPIS_MAIZE, TPIS_MOUSE, TPIS_DROME, TPIS_RABIT, TPIS_CAEEL

Solution. Firstly, I need to compute the Multiple Sequence Alignment of the given sets of sequences. It is because it is the aligned sequences itself that is passed as input to the **AL2CO** server, for it to compute conservation scores. So, below is an image of the input given to the **Clustal Omega** to compute the Multiple Sequence Alignment of set 1 and set 2. The outputs of the Multiple Sequence Alignment have been enclosed ahead of these input images.

The screenshot shows the Clustal Omega web interface. Under 'Input sequence', the 'Sequence Type' is set to 'Protein'. The text area contains the following sequences for Set 1:

```
>HBA_HUMAN
MVLSPADKTNVKAAGWGKGAHAGEYGAEALERMFSLFPTTKTYFPHFDLSHGSAQVKGHGKKVADALTNA
VAHYDDMPNALSALSDLHAHKLKRVDPVNFKLLSHCLLVTLAAHLPAEFTPAVHASLDKFLASVSTVLTSKYR
>HBA_RAT
MVLSDDKTNIKNCWGKIGGHGGEGYEEALQRMFAAFPTTKTYFSDIDVSPGSAQVKAGKKVADALAKA
ADHVEDLPGLSTLSDLHAHKLKRVDPVNFKFLSHCLLVTLACHHPGDFTPAMHASLDKFLASVSTVLTSKYR
>HBA_MOUSE
MVLSPADKTNVKAAGWGKGAHAGEYGAEALERMFSLFPTTKTYFPHFDLSHGSAQVKGHGKKVADALTNA
VAHYDDMPNALSALSDLHAHKLKRVDPVNFKLLSHCLLVTLAAHLPAEFTPAVHASLDKFLASVSTVLTSKYR
```

Buttons for 'Choose File', 'No file chosen', 'Use the example', and 'Clear sequence' are visible. The 'Parameters' section shows 'OUTPUT FORMAT' set to 'ClustalW with character counts'.

FIGURE 1. Input sequences to the CLUSTAL OMEGA from set 1

The screenshot shows the Clustal Omega web interface. Under 'Input sequence', the 'Sequence Type' is set to 'Protein'. The text area contains the following sequences for Set 2:

```
>TPIS_HUMAN
MAPSRKFFVGGNWKMNKRKQSLGELIGTLNAAKVPADTEVVCAPPTAYIDFARQKLDPKIAVAQAQNCYKVT
NGAFTGEISPGMIKDCGATWVVLGHSERRHVFGEDELIGQKVAHALAEGLVIAICIGEKLDEREAGITEKV
VFEQTKVIADNVKDWKSVVLAYEPVWAIGTGKTATPQQAQEVHEKLRGLKSNVSDAVAQSTRIIYGGSVT
GATCKELASQPDVDGFLVGGASLKPEFVDIINAKQ
>TPIS_YEAST
MARTFFVGGNFKLNGSKQSIKEIVERLNTASIPENVEVICPPATYLDYSVSLVKKPQVTVGAQNAYLKASGA
```

Buttons for 'Choose File', 'No file chosen', 'Use the example', and 'Clear sequence' are visible. The 'Parameters' section shows 'OUTPUT FORMAT' set to 'ClustalW with character counts'.

FIGURE 2. Input sequences to the CLUSTAL OMEGA from set 2

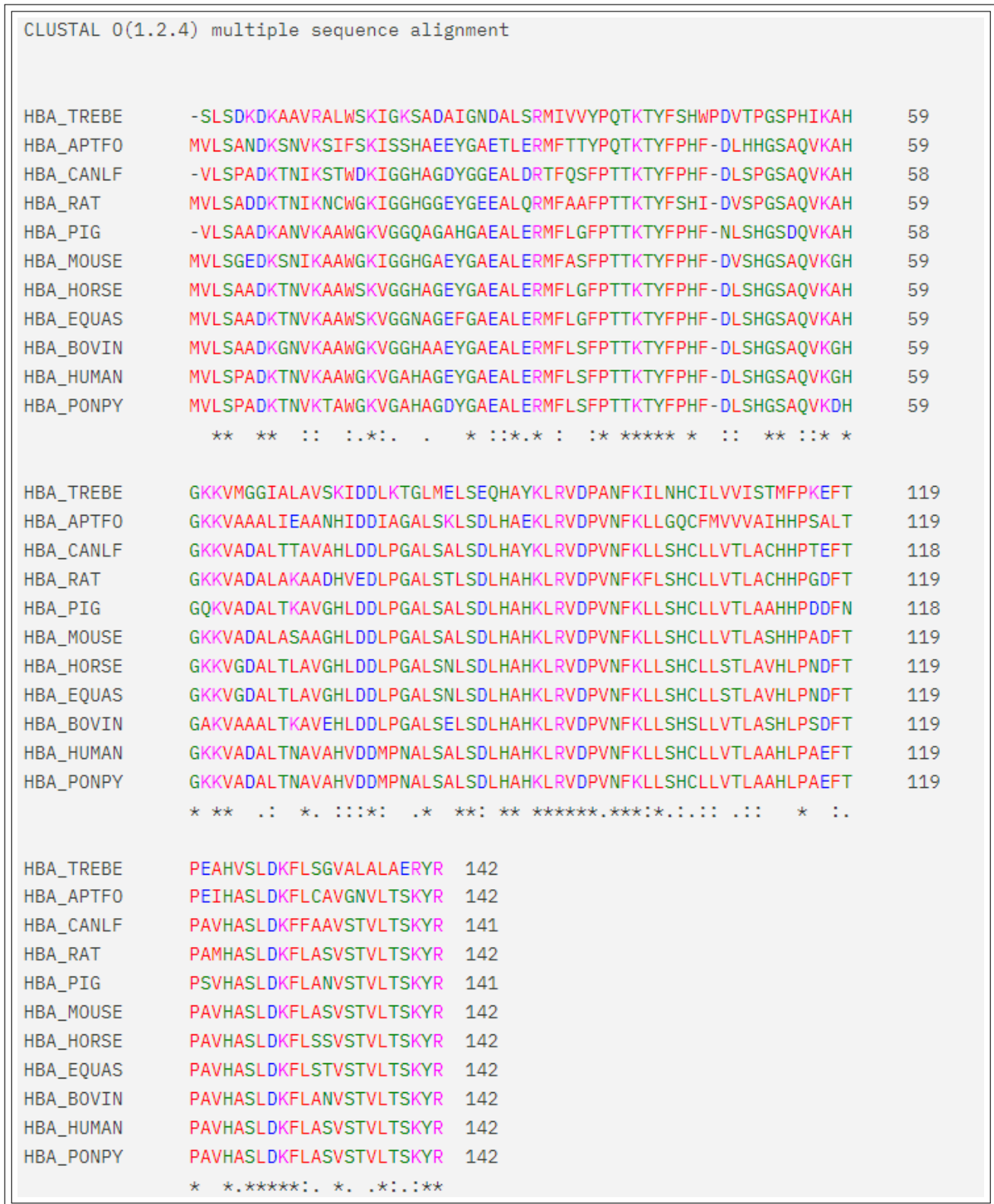


FIGURE 3. Output MSA of the CLUSTAL OMEGA from set 1

- The above image shows the multiple sequence alignment for the proteins enclosed in set 1. They are a total of 11 in number.
- The below image shows the multiple sequence alignment for the proteins enclosed in set 2. They are a total of 9 in number.
- These aligned sequences are those that will be passed to the AL2CO server tool to compute the conservation scores for these sets of proteins.

CLUSTAL O(1.2.4) multiple sequence alignment

```

TPIS_TRYCR      MASKPQPIAAANWKCNGSESLVPLIETLNAATFDHD--VQCVVAPTFLHIPMTKARLTN      58
TPIS_GRAGA      -----NWKCNLKADIAELVSAFNAAPPIDAAHVQVVVAPPVYLDSTRQAL-R      48
TPIS_YEAST      --MARTFFVGGNFKLNGSKQSIKEIVERLNTASIPEN--VEVVICPPATYLDYSVSLVKK      56
TPIS_MAIZE      --MGRKFFVGGNWKCNGTTDQVEKIVKTLNEGQVPPSDVVEVVVSPYVFLPVVKSQ-L-R      57
TPIS_CAEEL      --MTRKFFVGGNWKMNGDYASVDGIVTFLNASADNSS--VDVVVAPPAPYLAYAKSKL-K      55
TPIS_DROME      --MSRKFCVGGNWKMNGDQKSIKIAKTLSAALDPN--TEVVIGCPAIYLMYARNLL-P      55
TPIS_HUMAN      MAPSRKFFVGGNWKMNGRQKSLGELIGTLNAAKVPAD--TEVVCAPPTAYIDFARQKL-D      57
TPIS_RABIT      MAPSRKFFVGGNWKMNGRKKNLGELITTLNAAKVPAD--TEVVCAPPTAYIDFARQKL-D      57
TPIS_MOUSE      MAPTRKFFVGGNWKMNGRKKCLGELICTLNAANVPAG--TEVVCAPPTAYIDFARQKL-D      57

                *: *      : :      . .      . : *      . :      :

TPIS_TRYCR      PKFQIAAQNAI-TRSGAFTGEVSLQLIKDYGISWVVLGHSERRLY--YGETNEIVA EKVA      115
TPIS_GRAGA      ADFDTSAQNAWISKGAFTGELDAAMVKDVGAEWVILGHSERRHIAQLKESDHTIAMKAA      108
TPIS_YEAST      PQVTGAQNAYLKASGAFTGENSDQIKDVGAKWVILGHSERRSY--FHEDDKFIADKTK      114
TPIS_MAIZE      QEFHVAQNCWVKKGGAFTGEVSAEMLN LGVPWVILGHSERRAL--LGESNEFVGDKVA      115
TPIS_CAEEL      AGVLVAAQNCYKVPKGFTGEISPAMIKDLGLEWVILGHSERRHV--FGESDALIAEKT      113
TPIS_DROME      CELGLAGQNA YKAKGAFTGEISPAMIKDIGADWVILGHSERRAI--FGESDALIAEKAE      113
TPIS_HUMAN      PKIAVAAQNCYKVTNGAFTGEISPGMIKDCGATWVVLGHSERRHV--FGESDELIQKVA      115
TPIS_RABIT      PKIAVAAQNCYKVTNGAFTGEISPGMIKDCGATWVVLGHSERRHV--FGESDELIQKVA      115
TPIS_MOUSE      PKIAVAAQNCYKVTNGAFTGEISPGMIKDLGATWVVLGHSERRHV--FGESDELIQKVS      115

                . .*.      *****      : : *      **:*****      * :      . : *.

TPIS_TRYCR      QACA-AGFHVIVCVGETNEEREAGRTAAVVLTLAAVAQKLSKEAWSRVVIAYEPVWAIG      174
TPIS_GRAGA      YALQHASLGVICYIGELLEERESGQTIAVCERQLQALSDAI--SDWSDVVIAYEPVWAIG      166
TPIS_YEAST      FALG-QGVGVILCIGETLEEKKAGKTLDVVERQLNAVLEEV--KDWTNVVIAYEPVWAIG      171
TPIS_MAIZE      YALS-QGLKVIACVGETLEQREAGSTMDVAAQTKAIAEKI--KDWSNVVIAYEPVWAIG      172
TPIS_CAEEL      HALE-AGIKVVCIGETLEEREAGHTKDVNFRQLQAIVDKG--VSWENIVIAYEPVWAIG      170
TPIS_DROME      HALA-EGLVIAICIGETLEEREAGKTNEVVARQMCAYAQKI--KDWKNVVIAYEPVWAIG      170
TPIS_HUMAN      HALA-EGLVIAICIGETLEEREAGITEKVVFEQTQVIADNV--KDWKVVVLAYEPVWAIG      172
TPIS_RABIT      HALS-EGLVIAICIGETLEEREAGITEKVVFEQTQVIADNV--KDWKVVVLAYEPVWAIG      172
TPIS_MOUSE      HALA-EGLVIAICIGETLEEREAGITEKVVFEQTQVIADNV--KDWKVVVLAYEPVWAIG      172

                *      . . * : * : * : : : * : *      . :      * : * : *****

TPIS_TRYCR      TGKVATPQQAQEVHELLRRWVRSKLGTDIAAQLRILYGGSVTAKNARTLYQMRDINGFLV      234
TPIS_GRAGA      TGKVATPEQAQEVHEAVRAWLANNVSPQVAASTRILYGGSVSPANCESLAKQPNIDGFLV      226
TPIS_YEAST      TGLAATPEDAQDIHASIRKFLASKLGDKAASELRILYGGSSANGSNAVTFKDKADV DGLV      231
TPIS_MAIZE      TGKVATPQAQEVHASLRDWLKTNASPEVAESTRIIYGGSVTAANCKELAAQPDVDGFLV      232
TPIS_CAEEL      TGKTASGEQAQEVHEWIRAFLEKVPSPAVADATRIIYGGSVTADNAELGKKPDIDGFLV      230
TPIS_DROME      TGQTATPDQAQEVHAFRLQWLSDNISKEVSASLRIQYGGSVTAANAKELAKKPDIDGFLV      230
TPIS_HUMAN      TGKTATPQAQEVHEKLRGWLKSNVSDAVAQSTRIIYGGSVTGATCKELASQPDVDGFLV      232
TPIS_RABIT      TGKTATPQAQEVHEKLRGWLKSNVSDAVAQSTRIIYGGSVTGATCKELASQPDVDGFLV      232
TPIS_MOUSE      TGKTATPQAQEVHEKLRGWLKSNVNDGVAQSTRIIYGGSVTGATCKELASQPDVDGFLV      232

                ** . :      : * : : * : * : :      :      ** *****      . . :      : : *****

TPIS_TRYCR      GGASLKPEFVEIIEATK-----      251
TPIS_GRAGA      GGASMKPTFL EIVDSYKATLAEAV      250
TPIS_YEAST      GGASLKPEFVDIINSRN-----      248
TPIS_MAIZE      GGASLKPEFIDIINAATVKSA---      253
TPIS_CAEEL      GGASLKPDFVKIINARS-----      247
TPIS_DROME      GGASLKPEFVDIINARQ-----      247
TPIS_HUMAN      GGASLKPEFVDIINAKQ-----      249
TPIS_RABIT      GGASLKPEFVDIINAKQ-----      249
TPIS_MOUSE      GGASLKPEFVDIINAKQ-----      249

                ****:* * : . * : :

```

FIGURE 4. Output MSA of the CLUSTAL OMEGA from set 2

The general format of providing the input to the **AL2CO** server is given below. It includes the syntax in which the input must be provided to the AL2CO. The aligned sequences should only be passed to the AL2Co server. The parameters involved in submitting the input have been enclosed later for each category mentioned above:

AL2CO sequence conservation analysis server

The AL2CO program calculates positional conservation for a multiple sequence alignment.
[\[Documentation\]](#)

DATA INPUT

Enter protein sequence alignment in [CLUSTAL](#) format:

Clear sequences

HBA_HUMAN	MVLSPADKTNVKAANGKVGGAHAGEYGAEALERMFLSFPTTKTYFPHF-DLSHGSAQVKGH	59
HBA_RAT	MVLSADDKTNKNCWGKIGGHGGEYGEEALQRMFAAFPTTKTYFSHI-DVSPGSAQVKAH	59
HBA_MOUSE	MVLSGEDKSNIAANGKIGGHGAEYGAEALERMFASFPTTKTYFPHF-DVSHGSAQVKGH	59
HBA_BOVIN	MVLSAADKGNVKAANGKVGGAHAEYGAEALERMFLSFPTTKTYFPHF-DLSHGSAQVKGH	59
HBA_HORSE	MVLSAADKTNVKAANSKVGGHAGEYGAEALERMFLGFPTTKTYFPHF-DLSHGSAQVKAH	59
HBA_EQUAS	MVLSAADKTNVKAANSKVGGNAGEFGAEALERMFLGFPTTKTYFPHF-DLSHGSAQVKAH	59
HBA_PIG	-VLSAADKANVKAANGKVGGAHAGEYGAEALERMFLGFPTTKTYFPHF-NLSHGSDQVKAH	58
HBA_PONPY	MVLSPADKTNVKTAWGKVGGAHAGDYGAEALERMFLSFPTTKTYFPHF-DLSHGSAQVKDH	59
HBA_CANLF	-VLSPADKTNIKSTWDKIGGHAGDYGGEALDRTFQSFPTTKTYFPHF-DLSPGSAQVKAH	58
HBA_TREBE	-SLSDKKA AVRALWSKIGKSADAIGNDALSRMIVVYPQTKYFSHWPDVTPGSPHIKAH	59

Or upload a file

Choose File

No file chosen

DATA SUBMIT

Enter email to receive the result (optional):

Enter a job name (optional):

Submit

Reset

FIGURE 5. Input sequences to the AL2CO server from set 1

AL2CO sequence conservation analysis server

The AL2CO program calculates positional conservation for a multiple sequence alignment.
[\[Documentation\]](#)

DATA INPUT

Enter protein sequence alignment in [CLUSTAL](#) format:

Clear sequences

TPIS_HUMAN	MAPSRKFFVGGNWKMNGRKQSLGELIGTLNAAKVPAD--TEVVCAPPTAYIDFARQKL-D	57
TPIS_YEAST	--MARTFFVGGNFKLNGSKQSIKEIVERLNTASIPEN--VEVICPPATYLDYSVSLVKK	56
TPIS_GRAA	-----NWKCNLSKADIAELVSFAFNAAPPIDAAHVQVVVAPPVYLDSTRQAL-R	48
TPIS_TRYCR	MASKQPPIAAANWKCNGSESLVPLIETLNAATFDHD--VQCQVAPTFLLHIPMTKARLTN	58
TPIS_MAIZE	--MGRKFFVGGNWKCNGTDDQVEKIVKTLNEGQVPPSDVVEVVVSPPYVFLPVVKSQ-L-R	57
TPIS_MOUSE	MAPTRKFFVGGNWKMNGRKKCLGELICTLNAANVPAG--TEVVCAPPTAYIDFARQKL-D	57
TPIS_DROME	--MSRKFCVGGNWKMNQDQKSIAEIAKTLSSAALDPN--TEVIGCPAIYLYMYARNLL-P	55
TPIS_RABIT	MAPSRKFFVGGNWKMNGRKKNLGELITTLNAAKVPAD--TEVVCAPPTAYIDFARQKL-D	57
TPIS_CAEEEL	--MTRKFFVGGNWKMNQDYASVDGIVTFLNASADNSS--VDVVVAPPAPYLAYAKSKL-K	55

Or upload a file

Choose File

No file chosen

DATA SUBMIT

Enter email to receive the result (optional):

Enter a job name (optional):

Submit

Reset

FIGURE 6. Input sequences to the AL2CO server from set 2

(i) Unweighted frequency and entropy-based measure

The parameters for this scenario are enclosed in the image below. The important parameters are as follows:

- sequence weighting scheme
- conservation calculation method
- scoring matrix (for sum of pairs method only)
- scoring matrix transformation (for sum of pairs method only)
- normalize conservation values

The image shows a window titled "PARAMETERS" with a list of configuration options. The options include radio buttons for selecting between different schemes and methods, and input fields for numerical values. The settings shown are: sequence weighting scheme set to "unweighted", conservation calculation method set to "entropy", scoring matrix set to "BLOSUM62 matrix", scoring matrix transformation set to "no transformation", normalize conservation values set to "False", window size set to "1", gap fraction set to "0.5", exclude the first sequence set to "False", output alignment block size set to "70", and a field for a PDB file which is currently empty.

PARAMETERS

- [sequence weighting scheme](#): ☐ henikoff-henikoff ☐ independent count ☒ unweighted
- [conservation calculation method](#): ☒ entropy ☐ variance ☐ sum-of-pairs
- For sum-of-pairs method only:
[scoring matrix](#):
☒ BLOSUM62 matrix ☐ identity matrix
- [scoring matrix transformation](#):
☒ no transformation ☐ normalization ☐ adjustment
- [normalize conservation values](#): ☐ True ☒ False
- [window size used for averaging conservation \(for smoothing purpose\)](#):
- [gap fraction above which conservation calculation is not performed](#):
- [exclude the first sequence from calculation](#): ☐ True ☒ False
- [output alignment block size](#):
- [pdb file for which b-factor field is replaced with conservation \(optional\)](#):
 No file chosen

FIGURE 7. Parameters for the given scenario (i)

The AL2CO gives the list of positional conservation values and the alignment with integer conservation indices. The question asks to only calculate the positional conservation values.

The window of positional conservation values generates the following set of parameters which are the ones displayed in the above image.

It also displays some parameters taken into consideration to compute the desired positional conservation scores.

The image shows a window displaying the parameters used for the AL2CO calculation. The text is color-coded, with asterisks for important notes and different colors for various parameter names and values. The parameters listed are: gap fraction no less than 0.50, conservation set to M-S, M: mean, S: standard deviation, AL2CO parameters are: Input alignment file: QUERY_qfIljn, Output conservation file: QUERY_qfIljn.csv.txt, Output alignment file with index: QUERY_qfIljn.csv.aln; Block size: 70, Weighting scheme: unweighted, Conservation calculation method: entropy-based, Window size: 1, Conservation not normalized, and Gap fraction to suppress calculation: 0.50.

```
* gap fraction no less than 0.50; conservation set to M-S
M: mean; S: standard deviation

AL2CO parameters are:

Input alignment file: QUERY_qfIljn
Output conservation file: QUERY_qfIljn.csv.txt
Output alignment file with index: QUERY_qfIljn.csv.aln; Block size: 70
Weighting scheme: unweighted
Conservation calculation method: entropy-based
Window size: 1
Conservation not normalized
Gap fraction to suppress calculation: 0.50
```

FIGURE 8. Parameter output display in window

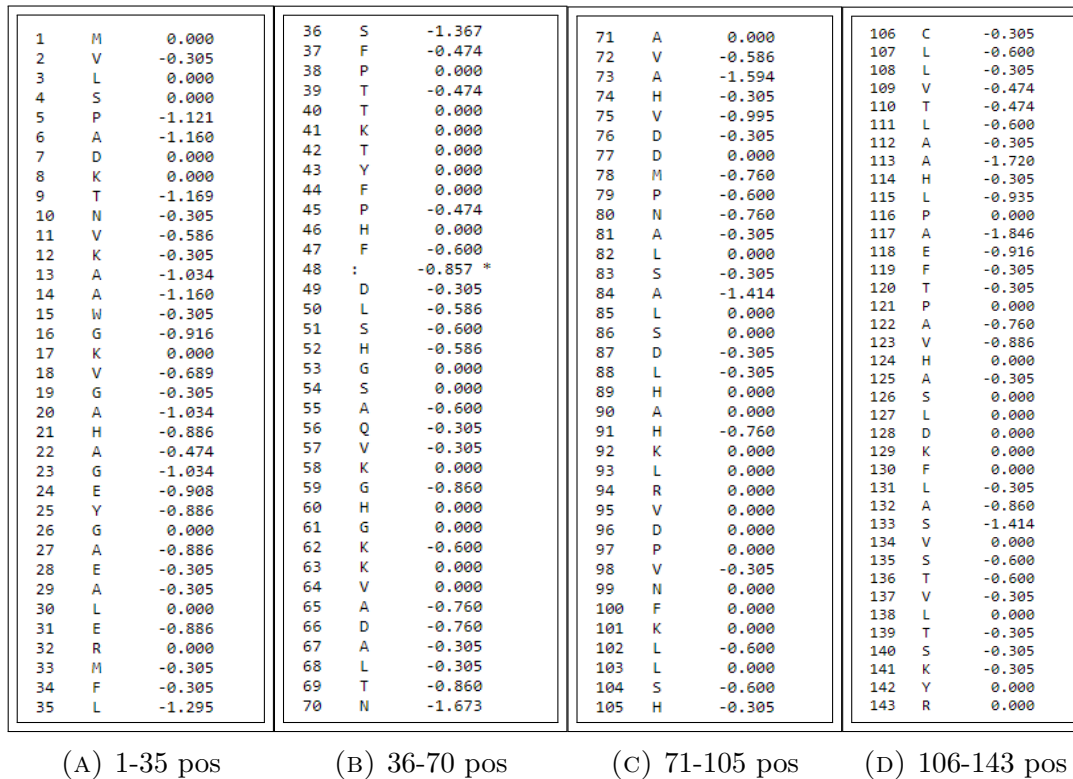


FIGURE 9. Positional conservation scores for set 1

Above is the image of the positional conservation scores for the set 1 proteins. Below is the image of the positional conservation scores for the set 2 proteins.

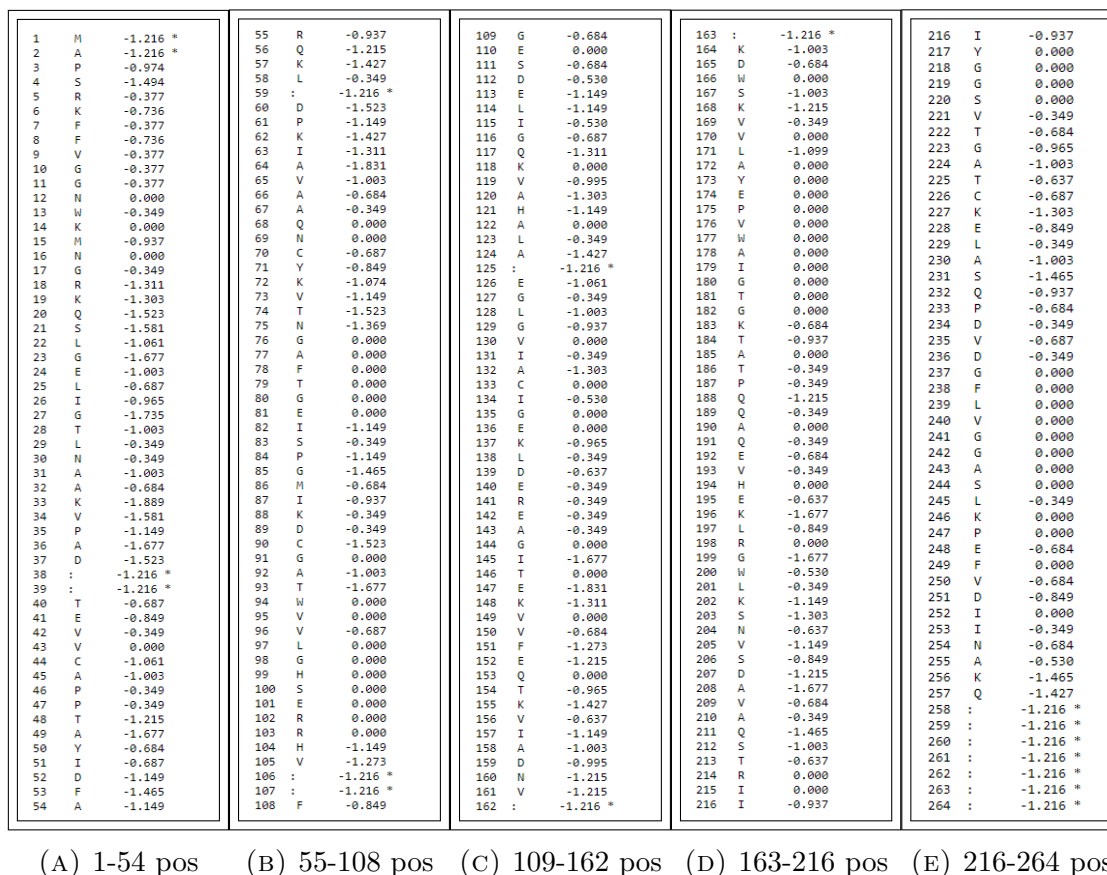


FIGURE 10. Positional conservation scores for set 2

(ii) Unweighted frequency and variance-based measure

The parameters for this scenario are enclosed in the image below. The important parameters are as follows:

- sequence weighting scheme
- conservation calculation method
- scoring matrix (for sum of pairs method only)
- scoring matrix transformation (for sum of pairs method only)
- normalize conservation values

The image shows a window titled "PARAMETERS" with a list of settings for a conservation calculation. The settings are as follows:

- sequence weighting scheme:** ☐ henikoff-henikoff ☐ independent count ☒ unweighted
- conservation calculation method:** ☐ entropy ☒ variance ☐ sum-of-pairs
- For sum-of-pairs method only:**
 - scoring matrix:** ☒ BLOSUM62 matrix ☐ identity matrix
 - scoring matrix transformation:** ☒ no transformation ☐ normalization ☐ adjustment
- normalize conservation values:** ☐ True ☒ False
- window size used for averaging conservation (for smoothing purpose):**
- gap fraction above which conservation calculation is not performed:**
- exclude the first sequence from calculation:** ☐ True ☒ False
- output alignment block size:**
- pdb file for which b-factor field is replaced with conservation (optional):**
 No file chosen

FIGURE 11. Parameters for the given scenario (ii)

The AL2CO gives the list of positional conservation values and the alignment with integer conservation indices. The question asks to only calculate the positional conservation values. The window of positional conservation values generates the following set of parameters which are the ones displayed in the above image. It also displays some parameters taken into consideration to compute the desired positional conservation scores.

The image shows a window displaying the following text:

```
* gap fraction no less than 0.50; conservation set to M-S
M: mean; S: standard deviation

AL2CO parameters are:

Input alignment file: QUERY_kUrrxa
Output conservation file: QUERY_kUrrxa.csv.txt
Output alignment file with index: QUERY_kUrrxa.csv.aln; Block size: 70
Weighting scheme: unweighted
Conservation calculation method: variance-based
Window size: 1
Conservation not normalized
Gap fraction to suppress calculation: 0.50
```

FIGURE 12. Parameter output display in window

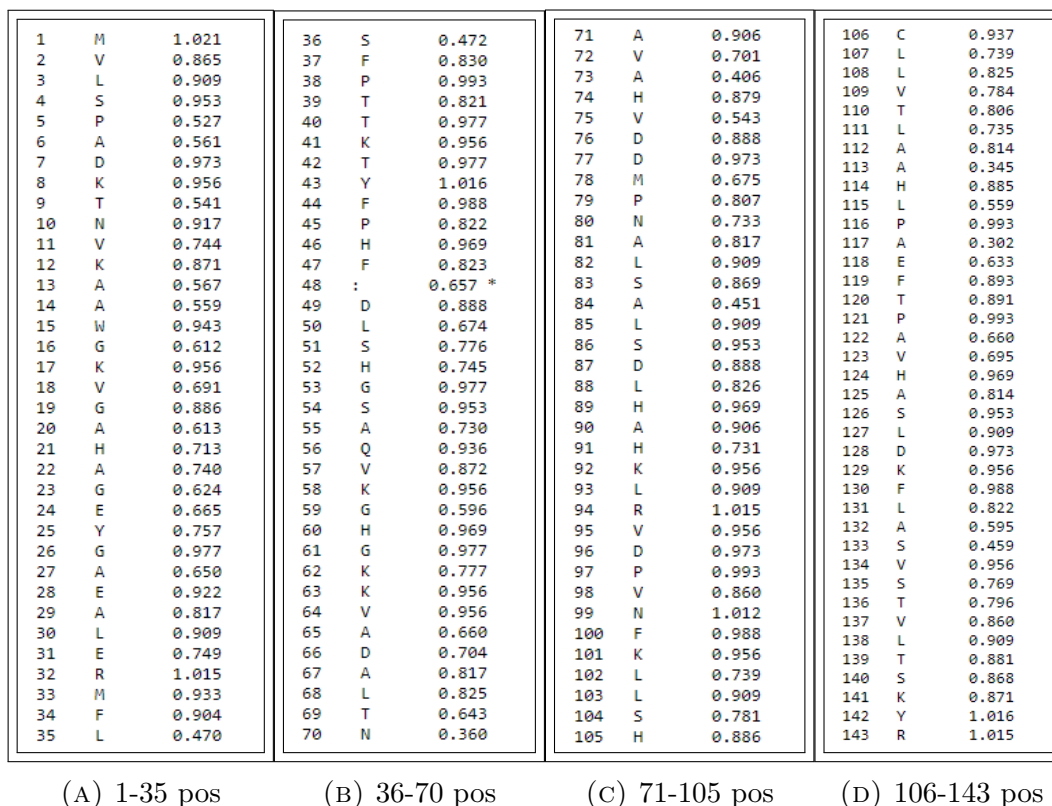


FIGURE 13. Positional conservation scores for set 1

Above is the image of the positional conservation scores for the set 1 proteins. Below is the image of the positional conservation scores for the set 2 proteins.

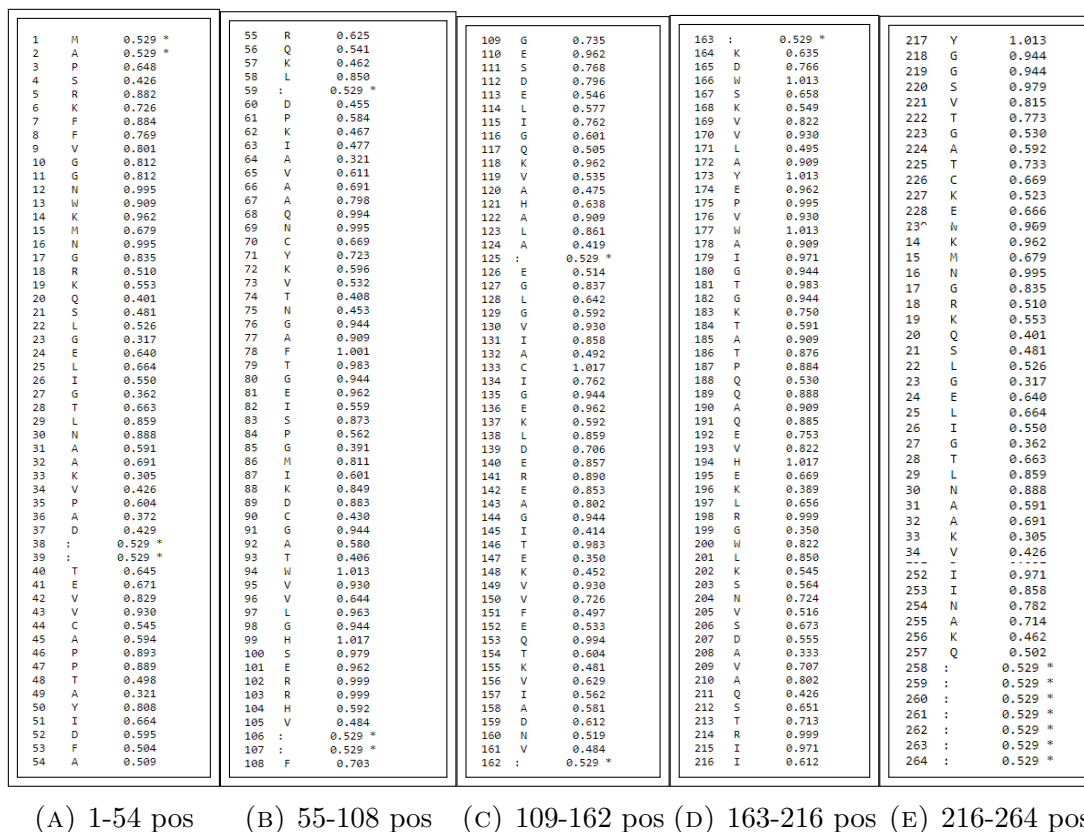


FIGURE 14. Positional conservation scores for set 2

(iii) Unweighted frequency and sum of pairs measure

The parameters for this scenario are enclosed in the image below. The important parameters are as follows:

- sequence weighting scheme
- conservation calculation method
- scoring matrix (for sum of pairs method only)
- scoring matrix transformation (for sum of pairs method only)
- normalize conservation values

The image shows a window titled "PARAMETERS" with a list of configuration options. The options are as follows:

- sequence weighting scheme:** ☐ henikoff-henikoff ☐ independent count ☒ unweighted
- conservation calculation method:** ☐ entropy ☐ variance ☒ sum-of-pairs
- For sum-of-pairs method only:**
 - scoring matrix:** ☒ BLOSUM62 matrix ☐ identity matrix
 - scoring matrix transformation:** ☒ no transformation ☐ normalization ☐ adjustment
- normalize conservation values:** ☐ True ☒ False
- window size used for averaging conservation (for smoothing purpose):**
- gap fraction above which conservation calculation is not performed:**
- exclude the first sequence from calculation:** ☐ True ☒ False
- output alignment block size:**
- pdb file for which b-factor field is replaced with conservation (optional):**
 No file chosen

FIGURE 15. Parameters for the given scenario (iii)

The AL2CO gives the list of positional conservation values and the alignment with integer conservation indices. The question asks to only calculate the positional conservation values. The window of positional conservation values generates the following set of parameters which are the ones displayed in the above image.

It also displays some parameters taken into consideration to compute the desired positional conservation scores.

The image shows a window displaying the following text:

```
* gap fraction no less than 0.50; conservation set to M-S
M: mean; S: standard deviation

AL2CO parameters are:

Input alignment file: QUERY_phGSQi
Output conservation file: QUERY_phGSQi.csv.txt
Output alignment file with index: QUERY_phGSQi.csv.aln; Block size: 70
Input matrix file: BLOSUM62
Matrix transformation: no transformation
Weighting scheme: unweighted
Conservation calculation method: sum-of-pairs measure
Window size: 1
Conservation not normalized
Gap fraction to suppress calculation: 0.50
```

FIGURE 16. Parameter output display in window

(iv) Weighted frequency and variance-based measure

The parameters for this scenario are enclosed in the image below. The important parameters are as follows:

- sequence weighting scheme
- conservation calculation method
- scoring matrix (for sum of pairs method only)
- scoring matrix transformation (for sum of pairs method only)
- normalize conservation values

The screenshot shows a window titled "PARAMETERS" with the following settings:

- sequence weighting scheme:** ☒ henikoff-henikoff ☐ independent count ☐ unweighted
- conservation calculation method:** ☐ entropy ☒ variance ☐ sum-of-pairs
- For sum-of-pairs method only:**
 - scoring matrix:** ☒ BLOSUM62 matrix ☐ identity matrix
 - scoring matrix transformation:** ☒ no transformation ☐ normalization ☐ adjustment
- normalize conservation values:** ☐ True ☒ False
- window size used for averaging conservation (for smoothing purpose):** 1
- gap fraction above which conservation calculation is not performed:** 0.5
- exclude the first sequence from calculation:** ☐ True ☒ False
- output alignment block size:** 70
- pdb file for which b-factor field is replaced with conservation (optional):**
Choose File No file chosen

FIGURE 19. Parameters for the given scenario (iv)

The AL2CO gives the list of positional conservation values and the alignment with integer conservation indices. The question asks to only calculate the positional conservation values. The window of positional conservation values generates the following set of parameters which are the ones displayed in the above image.

It also displays some parameters taken into consideration to compute the desired positional conservation scores.

The screenshot shows the following text in a window:

```
* gap fraction no less than 0.50; conservation set to M-S
M: mean; S: standard deviation

AL2CO parameters are:

Input alignment file: QUERY_EPrHPz
Output conservation file: QUERY_EPrHPz.csv.txt
Output alignment file with index: QUERY_EPrHPz.csv.aln; Block size: 70
Weighting scheme: weighted by the modified method of Henikoff & Henikoff
Conservation calculation method: variance-based
Window size: 1
Conservation not normalized
Gap fraction to suppress calculation: 0.50
```

FIGURE 20. Parameter output display in window

1	M	1.019	36	S	0.386	71	A	0.909	106	C	0.962
2	V	0.729	37	F	0.718	72	V	0.682	107	L	0.588
3	L	0.916	38	P	0.993	73	A	0.359	108	L	0.783
4	S	0.951	39	T	0.715	74	H	0.745	109	V	0.844
5	P	0.492	40	T	0.979	75	V	0.534	110	T	0.677
6	A	0.439	41	K	0.950	76	D	0.888	111	L	0.580
7	D	0.972	42	T	0.979	77	D	0.972	112	A	0.683
8	K	0.950	43	Y	1.012	78	M	0.683	113	A	0.355
9	T	0.465	44	F	0.989	79	P	0.618	114	H	0.769
10	N	0.770	45	P	0.712	80	N	0.664	115	L	0.529
11	V	0.763	46	H	0.972	81	A	0.692	116	P	0.993
12	K	0.744	47	F	0.711	82	L	0.916	117	A	0.299
13	A	0.566	48	:	0.582 *	83	S	0.749	118	E	0.584
14	A	0.415	49	D	0.901	84	A	0.428	119	F	0.837
15	W	0.888	50	L	0.621	85	L	0.916	120	T	0.907
16	G	0.604	51	S	0.605	86	S	0.951	121	P	0.993
17	K	0.950	52	H	0.689	87	D	0.763	122	A	0.589
18	V	0.719	53	G	0.977	88	L	0.715	123	V	0.519
19	G	0.831	54	S	0.951	89	H	0.972	124	H	0.972
20	A	0.502	55	A	0.628	90	A	0.909	125	A	0.685
21	H	0.627	56	Q	0.800	91	H	0.618	126	S	0.951
22	A	0.767	57	V	0.748	92	K	0.950	127	L	0.916
23	G	0.517	58	K	0.950	93	L	0.916	128	D	0.972
24	E	0.593	59	G	0.687	94	R	1.011	129	K	0.950
25	Y	0.679	60	H	0.972	95	V	0.956	130	F	0.989
26	G	0.977	61	G	0.977	96	D	0.972	131	L	0.839
27	A	0.561	62	K	0.812	97	P	0.993	132	A	0.528
28	E	0.784	63	K	0.950	98	V	0.715	133	S	0.400
29	A	0.768	64	V	0.956	99	N	1.011	134	V	0.956
30	L	0.916	65	A	0.612	100	F	0.989	135	S	0.577
31	E	0.628	66	D	0.561	101	K	0.950	136	T	0.615
32	R	1.011	67	A	0.692	102	L	0.632	137	V	0.715
33	M	0.944	68	L	0.710	103	L	0.916	138	L	0.916
34	F	0.780	69	T	0.545	104	S	0.619	139	T	0.738
35	L	0.373	70	N	0.382	105	H	0.839	140	S	0.743
									141	K	0.744
									142	Y	1.012
									143	R	1.011

(A) 1-35 pos

(B) 36-70 pos

(C) 71-105 pos

(D) 106-143 pos

FIGURE 21. Positional conservation scores for set 1

Above is the image of the positional conservation scores for the set 1 proteins. Below is the image of the positional conservation scores for the set 2 proteins.

1	M	0.494 *	55	R	0.582	109	G	0.682	163	:	0.494 *	217	Y	1.011
2	A	0.494 *	56	Q	0.505	110	E	0.960	164	K	0.568	218	G	0.951
3	P	0.666	57	K	0.399	111	S	0.700	165	D	0.718	219	G	0.951
4	S	0.365	58	L	0.809	112	D	0.765	166	W	1.013	220	S	0.977
5	R	0.827	59	:	0.494 *	113	E	0.484	167	S	0.617	221	V	0.776
6	K	0.641	60	D	0.440	114	L	0.508	168	K	0.558	222	T	0.711
7	F	0.830	61	P	0.551	115	I	0.726	169	V	0.815	223	G	0.521
8	F	0.713	62	K	0.421	116	G	0.643	170	V	0.930	224	A	0.518
9	V	0.739	63	I	0.497	117	Q	0.499	171	L	0.518	225	T	0.814
10	G	0.754	64	A	0.292	118	K	0.966	172	A	0.905	226	C	0.650
11	G	0.754	65	V	0.549	119	V	0.492	173	Y	1.011	227	K	0.429
12	N	0.994	66	A	0.623	120	A	0.465	174	E	0.960	228	E	0.596
13	W	0.876	67	A	0.793	121	H	0.569	175	P	0.997	229	L	0.823
14	K	0.966	68	Q	0.993	122	A	0.905	176	V	0.930	230	A	0.520
15	H	0.639	69	N	0.994	123	L	0.824	177	W	1.013	231	S	0.427
16	N	0.594	70	C	0.649	124	A	0.381	178	A	0.905	232	Q	0.601
17	G	0.810	71	Y	0.666	125	:	0.494 *	179	I	0.970	233	P	0.707
18	R	0.530	72	K	0.506	126	E	0.491	180	G	0.951	234	D	0.852
19	K	0.515	73	V	0.465	127	G	0.813	181	T	0.984	235	V	0.638
20	Q	0.372	74	T	0.380	128	L	0.566	182	G	0.951	236	D	0.847
21	S	0.482	75	N	0.446	129	G	0.573	183	K	0.718	237	G	0.951
22	L	0.514	76	G	0.951	130	V	0.930	184	T	0.535	238	F	1.006
23	G	0.270	77	A	0.905	131	I	0.849	185	A	0.905	239	L	0.959
24	E	0.597	78	F	1.006	132	A	0.401	186	T	0.870	240	V	0.930
25	L	0.656	79	T	0.984	133	C	1.018	187	P	0.880	241	G	0.951
26	I	0.557	80	G	0.951	134	I	0.726	188	Q	0.514	242	G	0.951
27	G	0.393	81	E	0.960	135	G	0.951	189	Q	0.853	243	A	0.905
28	T	0.595	82	I	0.478	136	E	0.960	190	A	0.905	244	S	0.977
29	L	0.826	83	S	0.840	137	K	0.598	191	Q	0.852	245	L	0.829
30	N	0.885	84	P	0.469	138	L	0.819	192	E	0.689	246	K	0.966
31	A	0.549	85	G	0.375	139	D	0.782	193	V	0.788	247	P	0.997
32	A	0.676	86	M	0.744	140	E	0.850	194	H	1.017	248	E	0.713
33	K	0.302	87	I	0.557	141	R	0.856	195	E	0.641	249	F	1.006
34	V	0.364	88	K	0.848	142	E	0.816	196	K	0.351	250	V	0.678
35	P	0.553	89	D	0.878	143	A	0.766	197	L	0.592	251	D	0.628
36	A	0.348	90	C	0.415	144	G	0.951	198	R	1.000	252	I	0.970
37	D	0.421	91	G	0.951	145	I	0.387	199	G	0.319	253	I	0.824
38	:	0.494 *	92	A	0.529	146	T	0.984	200	W	0.792	254	N	0.714
39	:	0.494 *	93	T	0.368	147	E	0.283	201	L	0.808	255	A	0.663
40	T	0.684	94	W	1.013	148	K	0.454	202	K	0.476	256	K	0.457
41	E	0.623	95	V	0.930	149	V	0.712	203	S	0.535	257	Q	0.451
42	V	0.795	96	V	0.676	150	V	0.692	204	N	0.690	258	:	0.494 *
43	V	0.930	97	L	0.959	151	F	0.440	205	V	0.471	259	:	0.494 *
44	C	0.563	98	G	0.951	152	E	0.562	206	S	0.656	260	:	0.494 *
45	A	0.557	99	H	1.017	153	Q	0.993	207	D	0.532	261	:	0.494 *
46	P	0.894	100	S	0.977	154	T	0.623	208	A	0.319	262	:	0.494 *
47	P	0.855	101	E	0.960	155	K	0.424	209	V	0.633	263	:	0.494 *
48	T	0.510	102	R	1.000	156	V	0.712	210	A	0.796	264	:	0.494 *
49	A	0.294	103	R	1.000	157	I	0.491	211	Q	0.405			
50	Y	0.771	104	H	0.522	158	A	0.507	212	S	0.575			
51	I	0.686	105	V	0.463	159	D	0.569	213	T	0.680			
52	D	0.564	106	:	0.494 *	160	N	0.511	214	R	1.000			
53	F	0.492	107	:	0.494 *	161	V	0.458	215	I	0.970			
54	A	0.450	108	F	0.650	162	:	0.494 *	216	I	0.586			

(A) 1-54 pos

(B) 55-108 pos

(C) 109-162 pos

(D) 163-216 pos

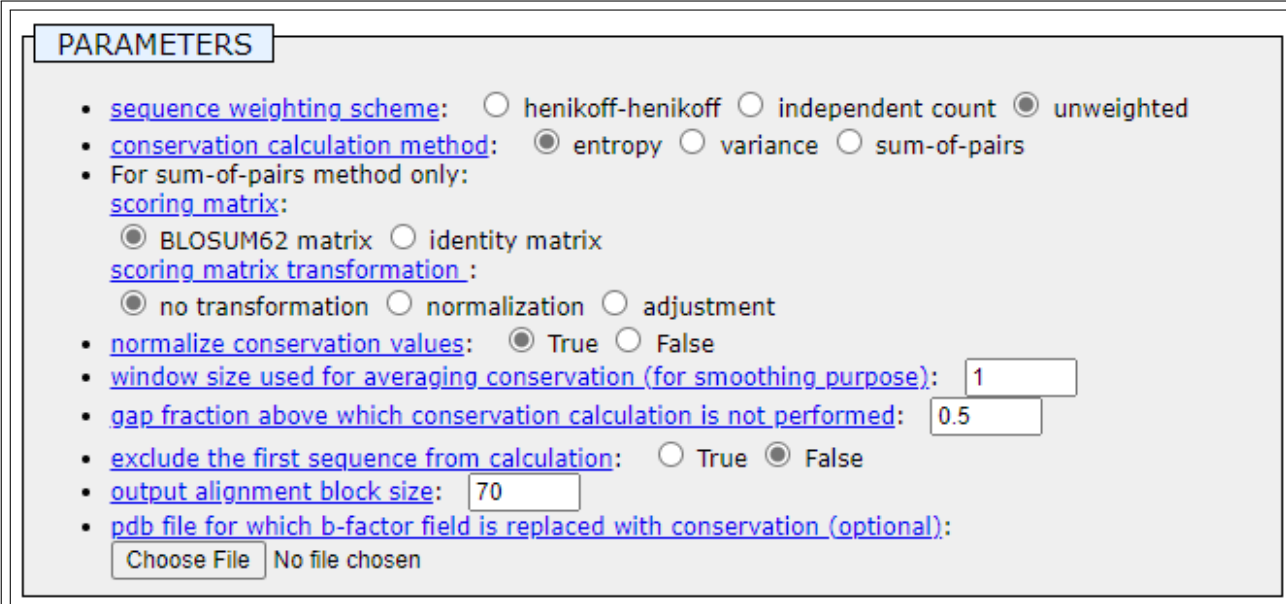
(E) 216-264 pos

FIGURE 22. Positional conservation scores for set 2

(v) Normalize the scores obtained with (i)

The parameters for this scenario are enclosed in the image below. The important parameters are as follows:

- sequence weighting scheme
- conservation calculation method
- scoring matrix (for sum of pairs method only)
- scoring matrix transformation (for sum of pairs method only)
- normalize conservation values

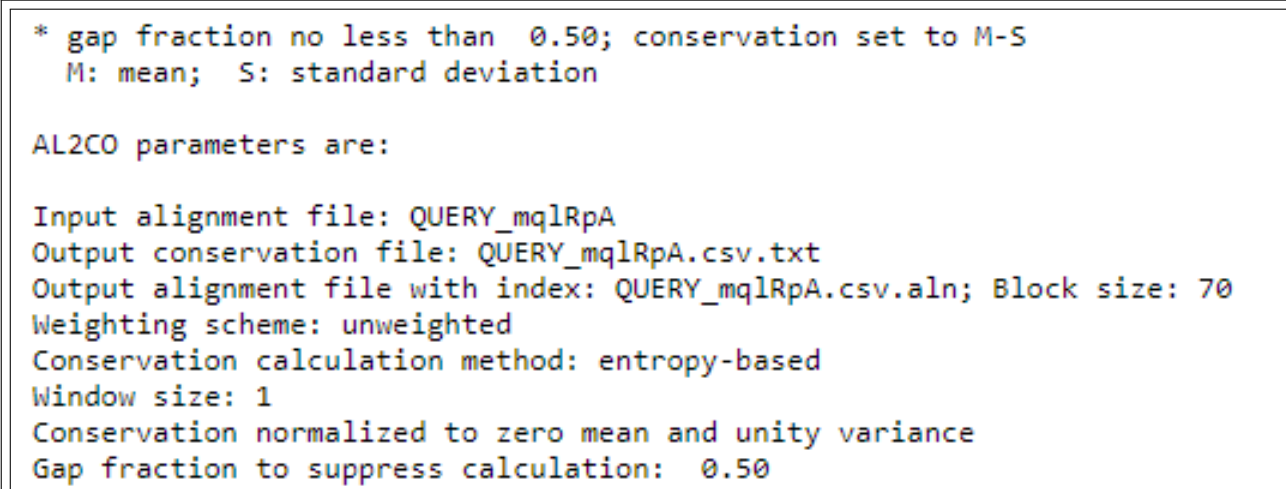


The screenshot shows a window titled "PARAMETERS" with the following settings:

- sequence weighting scheme:** ☐ henikoff-henikoff ☐ independent count ☒ unweighted
- conservation calculation method:** ☒ entropy ☐ variance ☐ sum-of-pairs
- For sum-of-pairs method only:**
 - scoring matrix:** ☒ BLOSUM62 matrix ☐ identity matrix
 - scoring matrix transformation:** ☒ no transformation ☐ normalization ☐ adjustment
- normalize conservation values:** ☒ True ☐ False
- window size used for averaging conservation (for smoothing purpose):** 1
- gap fraction above which conservation calculation is not performed:** 0.5
- exclude the first sequence from calculation:** ☐ True ☒ False
- output alignment block size:** 70
- pdb file for which b-factor field is replaced with conservation (optional):**
 No file chosen

FIGURE 23. Parameters for the given scenario (v)

The AL2CO gives the list of positional conservation values and the alignment with integer conservation indices. The question asks to only calculate the positional conservation values. The window of positional conservation values generates the following set of parameters which are the ones displayed in the above image. It also displays some parameters taken into consideration to compute the desired positional conservation scores.



```
* gap fraction no less than 0.50; conservation set to M-S
M: mean; S: standard deviation

AL2CO parameters are:

Input alignment file: QUERY_mqlRpA
Output conservation file: QUERY_mqlRpA.csv.txt
Output alignment file with index: QUERY_mqlRpA.csv.aln; Block size: 70
Weighting scheme: unweighted
Conservation calculation method: entropy-based
Window size: 1
Conservation normalized to zero mean and unity variance
Gap fraction to suppress calculation: 0.50
```

FIGURE 24. Parameter output display in window

1	M	0.943	36	S	-2.154	71	A	0.943	106	C	0.253
2	V	0.253	37	F	-0.131	72	V	-0.385	107	L	-0.417
3	L	0.943	38	P	0.943	73	A	-2.670	108	L	0.253
4	S	0.943	39	T	-0.131	74	H	0.253	109	V	-0.131
5	P	-1.597	40	T	0.943	75	V	-1.312	110	T	-0.131
6	A	-1.685	41	K	0.943	76	D	0.253	111	L	-0.417
7	D	0.943	42	T	0.943	77	D	0.943	112	A	0.253
8	K	0.943	43	Y	0.943	78	M	-0.778	113	A	-2.955
9	T	-1.705	44	F	0.943	79	P	-0.417	114	H	0.253
10	N	0.253	45	P	-0.131	80	N	-0.778	115	L	-1.175
11	V	-0.385	46	H	0.943	81	A	0.253	116	P	0.943
12	K	0.253	47	F	-0.417	82	L	0.943	117	A	-3.241
13	A	-1.399	48	:	-1.000 *	83	S	0.253	118	E	-1.134
14	A	-1.685	49	D	0.253	84	A	-2.262	119	F	0.253
15	W	0.253	50	L	-0.385	85	L	0.943	120	T	0.253
16	G	-1.134	51	S	-0.417	86	S	0.943	121	P	0.943
17	K	0.943	52	H	-0.385	87	D	0.253	122	A	-0.778
18	V	-0.618	53	G	0.943	88	L	0.253	123	V	-1.064
19	G	0.253	54	S	0.943	89	H	0.943	124	H	0.943
20	A	-1.399	55	A	-0.417	90	A	0.943	125	A	0.253
21	H	-1.064	56	Q	0.253	91	H	-0.778	126	S	0.943
22	A	-0.131	57	V	0.253	92	K	0.943	127	L	0.943
23	G	-1.399	58	K	0.943	93	L	0.943	128	D	0.943
24	E	-1.114	59	G	-1.006	94	R	0.943	129	K	0.943
25	Y	-1.064	60	H	0.943	95	V	0.943	130	F	0.943
26	G	0.943	61	G	0.943	96	D	0.943	131	L	0.253
27	A	-1.064	62	K	-0.417	97	P	0.943	132	A	-1.006
28	E	0.253	63	K	0.943	98	V	0.253	133	S	-2.262
29	A	0.253	64	V	0.943	99	N	0.943	134	V	0.943
30	L	0.943	65	A	-0.778	100	F	0.943	135	S	-0.417
31	E	-1.064	66	D	-0.778	101	K	0.943	136	T	-0.417
32	R	0.943	67	A	0.253	102	L	-0.417	137	V	0.253
33	M	0.253	68	L	0.253	103	L	0.943	138	L	0.943
34	F	0.253	69	T	-1.006	104	S	-0.417	139	T	0.253
35	L	-1.991	70	N	-2.848	105	H	0.253	140	S	0.253
									141	K	0.253
									142	Y	0.943
									143	R	0.943

(A) 1-35 pos

(B) 36-70 pos

(C) 71-105 pos

(D) 106-143 pos

FIGURE 25. Positional conservation scores for set 1

Above is the image of the positional conservation scores for the set 1 proteins. Below is the image of the positional conservation scores for the set 2 proteins.

1	M	-1.000 *	55	R	-0.492	109	G	-0.031	163	:	-1.000 *	217	Y	1.215
2	A	-1.000 *	56	Q	-0.998	110	E	1.215	164	K	-0.612	218	G	1.215
3	P	-0.560	57	K	-1.385	111	S	-0.031	165	D	-0.031	219	G	1.215
4	S	-1.507	58	L	0.580	112	D	0.250	166	W	1.215	220	S	1.215
5	R	0.529	59	:	-1.000 *	113	E	-0.879	167	S	-0.612	221	V	0.580
6	K	-0.125	60	D	-1.560	114	L	-0.879	168	K	-0.998	222	T	-0.031
7	F	0.529	61	P	-0.879	115	I	0.250	169	V	0.580	223	G	-0.543
8	F	-0.125	62	K	-1.385	116	G	-0.036	170	V	1.215	224	A	-0.612
9	V	0.529	63	I	-1.173	117	Q	-1.173	171	L	-0.787	225	T	0.056
10	G	0.529	64	A	-2.121	118	K	1.215	172	A	1.215	226	C	-0.036
11	G	0.529	65	V	-0.612	119	V	-0.598	173	Y	1.215	227	K	-1.159
12	N	1.215	66	A	-0.031	120	A	-1.159	174	E	1.215	228	E	-0.331
13	W	0.580	67	A	0.580	121	H	-0.879	175	P	1.215	229	L	0.580
14	K	1.215	68	Q	1.215	122	A	1.215	176	V	1.215	230	A	-0.612
15	M	-0.492	69	N	1.215	123	L	0.580	177	W	1.215	231	S	-1.454
16	N	1.215	70	C	-0.036	124	A	-1.385	178	A	1.215	232	Q	-0.492
17	G	0.580	71	Y	-0.331	125	:	-1.000 *	179	I	1.215	233	P	-0.031
18	R	-1.173	72	K	-0.741	126	E	-0.718	180	G	1.215	234	D	0.580
19	K	-1.159	73	V	-0.879	127	G	0.580	181	T	1.215	235	V	-0.036
20	Q	-1.560	74	T	-1.560	128	L	-0.612	182	G	1.215	236	D	0.580
21	S	-1.666	75	N	-1.279	129	G	-0.492	183	K	-0.031	237	G	1.215
22	L	-0.718	76	G	1.215	130	V	1.215	184	T	-0.492	238	F	1.215
23	G	-1.841	77	A	1.215	131	I	0.580	185	A	1.215	239	L	1.215
24	E	-0.612	78	F	1.215	132	A	-1.159	186	T	0.580	240	V	1.215
25	L	-0.036	79	T	1.215	133	C	1.215	187	P	0.580	241	G	1.215
26	I	-0.543	80	G	1.215	134	I	0.250	188	Q	-0.998	242	G	1.215
27	G	-1.946	81	E	1.215	135	G	1.215	189	Q	0.580	243	A	1.215
28	T	-0.612	82	I	-0.879	136	E	1.215	190	A	1.215	244	S	1.215
29	L	0.580	83	S	0.580	137	K	-0.543	191	Q	0.580	245	L	0.580
30	N	0.580	84	P	-0.879	138	L	0.580	192	E	-0.031	246	K	1.215
31	A	-0.612	85	G	-1.454	139	D	0.056	193	V	0.580	247	P	1.215
32	A	-0.031	86	M	-0.031	140	E	0.580	194	H	1.215	248	E	-0.031
33	K	-2.227	87	I	-0.492	141	R	0.580	195	E	0.056	249	F	1.215
34	V	-1.666	88	H	0.580	142	E	0.580	196	K	-1.841	250	V	-0.031
35	P	-0.879	89	D	0.580	143	A	0.580	197	L	-0.331	251	D	-0.331
36	A	-1.841	90	C	-1.560	144	G	1.215	198	R	1.215	252	I	1.215
37	D	-1.560	91	G	1.215	145	I	-1.841	199	G	-1.841	253	I	0.580
38	:	-1.000 *	92	A	-0.612	146	T	1.215	200	W	0.250	254	N	-0.031
39	:	-1.000 *	93	T	-1.841	147	E	-2.121	201	L	0.580	255	A	0.250
40	T	-0.036	94	W	1.215	148	K	-1.173	202	K	-0.879	256	K	-1.454
41	E	-0.331	95	V	1.215	149	V	1.215	203	S	-1.159	257	Q	-1.385
42	V	0.580	96	V	-0.036	150	V	-0.031	204	N	0.056	258	:	-1.000 *
43	V	1.215	97	L	1.215	151	F	-1.104	205	V	-0.879	259	:	-1.000 *
44	C	-0.718	98	G	1.215	152	E	-0.998	206	S	-0.331	260	:	-1.000 *
45	A	-0.612	99	H	1.215	153	Q	1.215	207	D	-0.998	261	:	-1.000 *
46	P	0.580	100	S	1.215	154	T	-0.543	208	A	-1.841	262	:	-1.000 *
47	P	0.580	101	E	1.215	155	K	-1.385	209	V	-0.031	263	:	-1.000 *
48	T	-0.998	102	R	1.215	156	V	0.056	210	A	0.580	264	:	-1.000 *
49	A	-1.841	103	R	1.215	157	I	-0.879	211	Q	-1.454			
50	Y	-0.031	104	H	-0.879	158	A	-0.612	212	S	-0.612			
51	I	-0.036	105	V	-1.104	159	D	-0.598	213	T	0.056			
52	D	-0.879	106	:	-1.000 *	160	N	-0.998	214	R	1.215			
53	F	-1.454	107	:	-1.000 *	161	V	-0.998	215	I	1.215			
54	A	-0.879	108	F	-0.331	162	:	-1.000 *	216	I	-0.492			

(A) 1-54 pos

(B) 55-108 pos

(C) 109-162 pos

(D) 163-216 pos

(E) 216-264 pos

FIGURE 26. Positional conservation scores for set 2

Question 2. Tabulate the topmost 10 residues with highest and lowest conservation scores (in both Set1 and Set 2) obtained with method (i).

Solution. First the output obtained in the Q1 (i) is taken. The output has three columns as seen from its image. The position on sequence alignment, the residue, and its position conservation score.

Now, I have copied the above text from the AL2CO server's positional conservation score output to a text file. I have now written a code to read the file and then convert it into a dataframe, which has then been sorted to extract the top 10 and bottom 10 values as desired. The code is given below:

```
1 import pandas as pd
2
3 new_line_list = []
4
5 # file.txt has the the positive conservation values the way it is shown in
  AL2CO server
6
7 with open("file.txt", 'r') as f:
8
9     # Read the text file and store in list
10
11     line = f.read()
12     line_list = line.split("\n")
13
14     # Remove the additional spaces and positions with gap alignments
15
16     for i in range(len(line_list)):
17         line_i = line_list[i].split(" ")
18         line_i = [str(value) for value in line_i if value != ""]
19         if line_i[1] != ":":
20             new_line_list.append(line_i)
21
22 # The above obtained list is then converted into a dataframe for easy
  readability and easier to perform sorting techniques and observe
23
24 df = pd.DataFrame(new_line_list)
25
26 # The first and third columns are converted to numeric data type because they
  represent the alignment position and position conservation scores
  respectively
27
28 df[2] = pd.to_numeric(df[2])
29 df[0] = pd.to_numeric(df[0])
30
31 # Sort the values in the descending order as per the last column, which is
  the column for positional conservation scores
32 final_df = df.sort_values(by=[2], ascending=False)
33
34 # Display the top 10 and bottom 10 residues
35 # Top 10 will be the highest scores
36 # Bottom 10 will be the lowest scores
37 final_df.head(10)
38 final_df.tail(10)
```

The last two lines will print the top 10 residues with highest and lowest positional conservation scores.

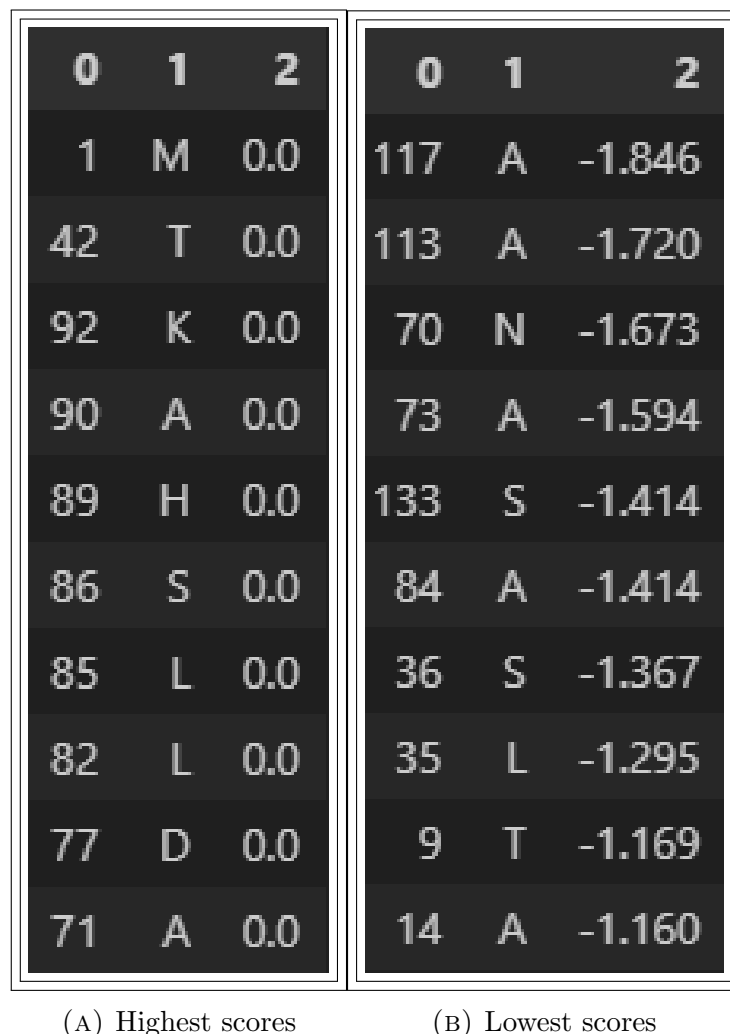


FIGURE 27. Top 10 residues with highest and lowest scores in set 1

The topmost 10 residues with highest and lowest conservation scores in set 1

In the following table, I have enclosed the readings from the above image with the top 10 residues with the highest and the lowest residues.

The results in this table are with respect to set 1 only.

Top 10 residues with the highest conservation scores			Top 10 residues with the lowest conservation scores		
Position	Residue	Score	Position	Residue	Score
1	Methionine	0.0	117	Alanine	-1.846
42	Threonine	0.0	113	Alanine	-1.720
92	Lysine	0.0	70	Asparagine	-1.673
90	Alanine	0.0	73	Alanine	-1.594
89	Histidine	0.0	133	Serine	-1.414
86	Serine	0.0	84	Alanine	-1.414
85	Leucine	0.0	36	Serine	-1.367
82	Leucine	0.0	35	Leucine	-1.295
77	Aspartate	0.0	9	Threonine	-1.169
71	Alanine	0.0	14	Alanine	-1.160

0	1	2	0	1	2
240	V	0.0	33	K	-1.889
172	A	0.0	147	E	-1.831
78	F	0.0	64	A	-1.831
77	A	0.0	27	G	-1.735
76	G	0.0	49	A	-1.677
237	G	0.0	93	T	-1.677
220	S	0.0	145	I	-1.677
166	W	0.0	199	G	-1.677
69	N	0.0	196	K	-1.677
68	Q	0.0	208	A	-1.677

(A) Highest scores

(B) Lowest scores

FIGURE 28. Top 10 residues with highest and lowest scores in set 2

The topmost 10 residues with highest and lowest conservation scores in set 2

In the following table, I have enclosed the readings from the above image with the top 10 residues with the highest and the lowest residues.

The results in this table are with respect to set 2 only.

Top 10 residues with the highest conservation scores			Top 10 residues with the lowest conservation scores		
Position	Residue	Score	Position	Residue	Score
240	Valine	0.0	33	Lysine	-1.889
172	Alanine	0.0	147	Glutamate	-1.831
78	Phenylalanine	0.0	64	Alanine	-1.831
77	Alanine	0.0	27	Glycine	-1.735
76	Glycine	0.0	49	Alanine	-1.677
237	Glycine	0.0	93	Threonine	-1.677
220	Serine	0.0	145	Isoleucine	-1.677
166	Tryptophan	0.0	199	Glycine	-1.677
69	Asparagine	0.0	196	Lysine	-1.677
68	Glutamine	0.0	208	Alanine	-1.677

Question 3. Write a program to compute the conservation score from MSA using unweighted frequency, and entropy, variance and sum of pairs-based measures.

Solution. The code for computing the conservation score from the MSA using the above mentioned techniques is given below:

```
1 # Creating a blosum62 matrix for sum of pairs measure
2 # The blosum matrix is read from a text file stored locally
3 # blosum_dict initializes the index for each amino acid in matrix
4 blosum_dict = {"A":0,"R":1,"N":2,"D":3,"C":4,"Q":5,"E":6,"G":7,"H":8,"I":9,
5 "L":10,"K":11,"M":12,"F":13,"P":14,"S":15,"T":16,"W":17,"Y":18,"V":19,"-":20}
6
7 blosum_matrix = []
8 with open("blosum62.txt", 'r') as f:
9     line = f.read()
10    line_list = line.split("\n")
11
12    for i in range(len(line_list)):
13        line_i = line_list[i].split(" ")
14        line_i = [int(value) for value in line_i if value != ""]
15        blosum_matrix.append(line_i)
```

LISTING 1. Blosum62 Matrix

```
1 import math
2
3 amino_acids_track =
4 { "0": "G", "1": "A", "2": "V", "3": "L", "4": "I",
5  "5": "T", "6": "S", "7": "M", "8": "C", "9": "P",
6  "10": "F", "11": "Y", "12": "W", "13": "H", "14": "K",
7  "15": "R", "16": "D", "17": "E", "18": "N", "19": "Q",
8  "20": "-"}
9
10 amino_acids =
11 {"G": 0, "A": 1, "V": 2, "L": 3, "I": 4, "T": 5,
12  "S": 6, "M": 7, "C": 8, "P": 9, "F": 10, "Y": 11,
13  "W": 12, "H": 13, "K": 14, "R": 15, "D": 16, "E": 17,
14  "N": 18, "Q": 19, "-": 20}
15
16 # Creating lists to store the conservation scores using the weighing scheme
17   of unweighted amino acid frequencies with the three conservation
18   calculation methods given in the question, entropy, variance and sum-of-
19   pairs
20
21 entropy_score_list = []
22 variance_score_list = []
23 sum_pairs_score_list = []
24
25 # Iterating through entire aligned sequence
26 for i in range(len(seqs[0])):
27     entropy_score = 0
28     variance_score = 0
29     sum_pairs_score = 0
30
31     each_freq = [0 for k in range(21)]
32     overall_freq = [0 for k in range(21)]
33     total_non_aligns = 0
```



```

31
32 # Iterating through each column of sequences
33 for j in range(len(seqs)):
34     each_freq[amino_acids[seqs[j][i]]] += 1
35
36 for j in range(len(each_freq)):
37     each_freq[j] /= (len(seqs)-each_freq[20])
38
39 for j in range(len(seqs)):
40     for k in range(len(seqs[0])):
41         overall_freq[amino_acids[seqs[j][k]]] += 1
42         if seqs[j][k] == "-":
43             total_non_aligns += 1
44
45 # For the sake of variance based conservation score
46 for j in range(len(overall_freq)):
47     overall_freq[j] /= (len(seqs) * len(seqs[0]) - total_non_aligns)
48
49 # Calculation of conservation score via the entropy method
50 for j in range(len(each_freq)-1):
51     if each_freq[j] != 0:
52         entropy_score += each_freq[j] * math.log(each_freq[j])
53
54 # Calculation of conservation score via the variance method
55 for j in range(len(each_freq)-1):
56     variance_score += abs(each_freq[j] - overall_freq[j])**2
57 variance_score = variance_score**0.5
58
59 # Calculation of conservation score via the sum-of-pairs method
60 for j in range(len(each_freq)-1):
61     for k in range(len(each_freq)-1):
62         sum_pairs_score += (each_freq[j]*each_freq[k]*
63                             blosum_matrix[blosum_dict[amino_acids_track[str(j)]]]
64                             [blosum_dict[amino_acids_track[str(k)]]])
65
66 # Rounding off the values to 4 decimal places (just like AL2CO)
67 entropy_score = round(entropy_score, 4)
68 variance_score = round(variance_score, 4)
69 sum_pairs_score = round(sum_pairs_score, 4)
70
71 # Appending the scores for each column to the list
72 entropy_score_list.append(entropy_score)
73 variance_score_list.append(variance_score)
74 sum_pairs_score_list.append(sum_pairs_score)
75
76 # Print the output lists
77 print(entropy_score_list)
78 print(variance_score_list)
79 print(sum_pairs_score_list)

```

LISTING 2. Computing the different metric conservation scores

The above code generates the lists of entropy based, variance based, and sum-of-pairs based scores. The input sequences used were same as the one that is used in the Set 1 of the first question. The value of output generated by the code is compared against the ones generated by the AL2CO server. They match.

The input passed to the above code is the list of 11 sequences that were passed as input in the first question. They are: P69905, P01946, P01942, P01966, P01958, P01959, P01965, P06635, P60529, P80043 and P01980. These correspond to HBA_HUMAN, HBA_RAT, HBA_MOUSE, HBA_BOVIN, HBA_HORSE, HBA_EQUAS, HBA_PIG, HBA_PONPY, HBA_CANLF, HBA_TREBE, HBA_APTFO respectively. The inputs and their outputs are given below:

Input given to the above code

```

1 seqs[0] =
2 "MVLSPADKTNVKAAWGKVGGAHAGEYGAEALERMFSLFPTTKTYFPHF -
3 DLSHGSAQVKGHGKKVADALTNVAHVDDMPNALSALSDLHAHKLRVDP
4 VNFKLLSHCLLVTLAAHLPAEFTPAVHASLSDKFLASVSTVLTSKYR "
5 seqs[1] =
6 "MVLSADDKTNKNCWGKIGGHGGEYGEEALQRMFAAFPTTKTYFSHI -
7 DVSPGSAQVKAHGGKVADALAKAADHVEDLPGALSTLSDLHAHKLRVDP
8 VNFKFLSHCLLVTLACHHPGDFTPAVHASLSDKFLASVSTVLTSKYR "
9 seqs[2] =
10 "MVLSGEDKSNIKAAWGKIGGHGAEYGAEALERMFASFPTTKTYFPHF -
11 DVSHGSAQVKGHGKKVADALASAAGHLDDLPGALSALSDLHAHKLRVDP
12 VNFKLLSHCLLVTLASHHPADFTPAVHASLSDKFLASVSTVLTSKYR "
13 seqs[3] =
14 "MVLSAADKGNVKAAWGKVGGAHAEYGAEALERMFSLFPTTKTYFPHF -
15 DLSHGSAQVKGHGAKVAAALTKAVEHLDDLPGALSELSDLHAHKLRVDP
16 VNFKLLSHSLLVTLASHLPDFTPAVHASLSDKFLANVSTVLTSKYR "
17 seqs[4] =
18 "MVLSAADKTNVKAAWSKVGGAHAGEYGAEALERMFLGFPTTKTYFPHF -
19 DLSHGSAQVKAHGGKVGDALTLAVGHLDDLPGALSNLSDLHAHKLRVDP
20 VNFKLLSHCLLVTLAVHLPNDFTPAVHASLSDKFLSSVSTVLTSKYR "
21 seqs[5] =
22 "MVLSAADKTNVKAAWSKVGGNAGEFGAEALERMFLGFPTTKTYFPHF -
23 DLSHGSAQVKAHGGKVGDALTLAVGHLDDLPGALSNLSDLHAHKLRVDP
24 VNFKLLSHCLLVTLAVHLPNDFTPAVHASLSDKFLSTVSTVLTSKYR "
25 seqs[6] =
26 "-VLSAADKANVKAAWGKVGGAHAGEYGAEALERMFLGFPTTKTYFPHF -
27 NLSHGSDQVKAHGGKVADALTKAVGHLDDLPGALSALSDLHAHKLRVDP
28 VNFKLLSHCLLVTLAAHPPDDFNPSVHASLSDKFLANVSTVLTSKYR "
29 seqs[7] =
30 "MVLSPADKTNVKTAWGKVGGAHAGDYGAEALERMFSLFPTTKTYFPHF -
31 DLSHGSAQVKDHGKKVADALTNVAHVDDMPNALSALSDLHAHKLRVDP
32 VNFKLLSHCLLVTLAAHLPAEFTPAVHASLSDKFLASVSTVLTSKYR "
33 seqs[8] =
34 "-VLSPADKTNKSTWDKIGGHAGDYGGEALDRTFQSFPPTTKTYFPHF -
35 DLSPGSAQVKAHGGKVADALTTAVAHDDLPGALSALSDLHAYKLRVDP
36 VNFKLLSHCLLVTLACHHPTEFTPAVHASLSDKFFAAVSTVLTSKYR "
37 seqs[9] =
38 "-SLSDKDKAAVRALWSKIGKSADAIGNDALSRMIVVYPQTKTYFSHP
39 DVTPGSPHIKAHGGKVMGGIALAVSKIDDLKTGLMELSEQHAYKLRVDP
40 ANFKILNHCILVVISTMFPEFTPEAHVSLDKFLSGVALALAERYR "
41 seqs[10] =
42 "MVLSANDKSNVKSIFSKISSHAEYGAETLERMFTTYPQTKTYFPHF -
43 DLHHGSAQVKAHGGKVAAALIEAANHIDDIAGALSKLSDLHAEKLRVDP
44 VNFKLLGQCFMVVVAIHHPALTPEIHASLSDKFLCAVGNVLTISKYR "

```

LISTING 3. Input given to the above code

Output lists generated by the above code

```
1 # Entropy based conservation score
2 [0.0, -0.305, 0.0, 0.0, -1.121, -1.16, 0.0, 0.0, -1.169, -0.305,
3 -0.586, -0.305, -1.034, -1.16, -0.305, -0.916, 0.0, -0.689, -0.305,
4 -1.034, -0.886, -0.474, -1.034, -0.908, -0.886, 0.0, -0.886,
5 -0.305, -0.305, 0.0, -0.886, 0.0, -0.305, -0.305, -1.295, -1.367,
6 -0.474, 0.0, -0.474, 0.0, 0.0, 0.0, 0.0, 0.0, -0.474, 0.0, -0.6,
7 0.0, -0.305, -0.586, -0.6, -0.586, 0.0, 0.0, -0.6, -0.305, -0.305,
8 0.0, -0.86, 0.0, 0.0, -0.6, 0.0, 0.0, -0.76, -0.76, -0.305, -0.305,
9 -0.86, -1.673, 0.0, -0.586, -1.594, -0.305, -0.995, -0.305, 0.0,
10 -0.76, -0.6, -0.76, -0.305, 0.0, -0.305, -1.414, 0.0, 0.0, -0.305,
11 -0.305, 0.0, 0.0, -0.76, 0.0, 0.0, 0.0, 0.0, 0.0, 0.0, -0.305, 0.0,
12 0.0, 0.0, -0.6, 0.0, -0.6, -0.305, -0.305, -0.6, -0.305, -0.474,
13 -0.474, -0.6, -0.305, -1.72, -0.305, -0.935, 0.0, -1.846, -0.916,
14 -0.305, -0.305, 0.0, -0.76, -0.886, 0.0, -0.305, 0.0, 0.0, 0.0,
15 0.0, 0.0, -0.305, -0.86, -1.414, 0.0, -0.6, -0.6, -0.305, 0.0,
16 -0.305, -0.305, -0.305, 0.0, 0.0]
17
18 # Variance based conservation score
19 [1.021, 0.865, 0.909, 0.953, 0.527, 0.561, 0.973, 0.956, 0.541,
20 0.917, 0.744, 0.871, 0.567, 0.56, 0.943, 0.612, 0.956, 0.691,
21 0.886, 0.613, 0.713, 0.74, 0.624, 0.665, 0.757, 0.977, 0.65, 0.922,
22 0.817, 0.909, 0.749, 1.015, 0.933, 0.904, 0.47, 0.473, 0.83, 0.993,
23 0.821, 0.977, 0.956, 0.977, 1.016, 0.988, 0.821, 0.969, 0.823,
24 0.993, 0.888, 0.674, 0.776, 0.745, 0.977, 0.953, 0.73, 0.936,
25 0.872, 0.956, 0.596, 0.969, 0.977, 0.777, 0.956, 0.956, 0.66,
26 0.704, 0.817, 0.825, 0.643, 0.36, 0.906, 0.701, 0.406, 0.879,
27 0.543, 0.888, 0.973, 0.675, 0.806, 0.733, 0.817, 0.909, 0.869,
28 0.451, 0.909, 0.953, 0.888, 0.826, 0.969, 0.906, 0.731, 0.956,
29 0.909, 1.015, 0.956, 0.973, 0.993, 0.86, 1.012, 0.988, 0.956,
30 0.739, 0.909, 0.781, 0.886, 0.937, 0.739, 0.825, 0.784, 0.806,
31 0.735, 0.814, 0.345, 0.885, 0.56, 0.993, 0.302, 0.633, 0.893,
32 0.891, 0.993, 0.66, 0.695, 0.969, 0.815, 0.953, 0.909, 0.973,
33 0.956, 0.988, 0.822, 0.595, 0.459, 0.956, 0.769, 0.797, 0.86,
34 0.909, 0.881, 0.868, 0.871, 1.016, 1.015]
35
36 # Sum-of-pairs based conservation score
37 [5.0, 3.008, 4.0, 4.0, 1.149, 1.157, 6.0, 5.0, 1.868, 4.661, 3.603,
38 4.504, 1.909, 1.521, 9.306, 2.264, 5.0, 3.504, 4.992, 2.405, 4.372,
39 2.876, 2.24, 2.455, 4.314, 6.0, 1.826, 4.512, 3.347, 4.0, 3.298,
40 5.0, 4.008, 4.992, 0.959, 1.132, 5.14, 7.0, 3.215, 5.0, 5.0, 5.0,
41 7.0, 6.0, 4.521, 8.0, 4.24, 7.0, 5.174, 2.81, 2.752, 3.959, 6.0,
42 4.0, 2.322, 4.198, 3.835, 5.0, 1.835, 8.0, 6.0, 3.405, 5.0, 4.0,
43 2.124, 2.694, 3.355, 3.669, 2.19, 0.124, 4.0, 2.413, 0.934, 6.488,
44 2.612, 5.331, 6.0, 3.14, 4.446, 3.149, 3.355, 4.0, 3.182, 0.678,
45 4.0, 4.0, 5.331, 3.017, 8.0, 4.0, 4.967, 5.0, 4.0, 5.0, 4.0, 6.0,
46 7.0, 3.339, 6.0, 6.0, 5.0, 3.058, 4.0, 2.926, 6.653, 7.306, 3.058,
47 3.678, 2.215, 3.479, 3.24, 3.504, 0.702, 6.322, 1.207, 7.0, 0.562,
48 3.008, 4.992, 4.182, 7.0, 2.182, 2.736, 8.0, 3.339, 4.0, 4.0, 6.0,
49 5.0, 6.0, 3.355, 2.289, 1.496, 4.0, 2.909, 3.231, 3.339, 4.0,
50 4.165, 3.347, 4.504, 7.0, 5.0]
```

LISTING 4. The conversation scores output generated by the above code

Question 4. Using the program written in Q3 (unweighted frequency and entropy-based measure), compare the MSA from Clustal Omega, MAFFT, and MUSCLE. Identify the residues with (i) similar and (ii) different conservation scores among the three alignment methods.

Solution. Below are the MSA obtained from the Clustal Omega, MAFFT, and MUSCLE for the protein sequences in set 1.

CLUSTAL O(1.2.4) multiple sequence alignment		
HBA_TREBE	-SLSDKDKA AVRALWSKIGKSADAIGNDALSRMIVVYPQTKTYFSHWPDVTPGSPHIKAH	59
HBA_APTFO	MVLSANDKSNVKSIFSKISSHAE EYGAETLERMFTTYPQTKTYFPHF-DLHHGSAQVKAH	59
HBA_CANLF	-VLSPADKTNIKSTWDKIGGHAGDYGGEALDRTFQSFPPTKTYFPHF-DLSPGSAQVKAH	58
HBA_RAT	MVLSADDKTNIKNCWKGKIGGHGGEYGEALQRMFAAFPTTKTYFSHI-DVSPGSAQVKAH	59
HBA_PIG	-VLSAADKANVKA AWGKVGGQAGAHGA EALERMFLGFPTTKTYFPHF-NLSHGSDQVKAH	58
HBA_MOUSE	MVLSGEDKSNIKAAWGKIGGHGA EYGA EALERMFA SFPTTKTYFPHF-DVSHGSAQVKGH	59
HBA_HORSE	MVLSAADKTNVKA AWSKVGGHAGEYGA EALERMFLGFPTTKTYFPHF-DLSHGSAQVKAH	59
HBA_EQUAS	MVLSAADKTNVKA AWSKVGGNAGEFGA EALERMFLGFPTTKTYFPHF-DLSHGSAQVKAH	59
HBA_BOVIN	MVLSAADKGNVKA AWGKVGGHAA EYGA EALERMFLSFPTTKTYFPHF-DLSHGSAQVKGH	59
HBA_HUMAN	MVLSPADKTNVKA AWGKVGAHAGEYGA EALERMFLSFPTTKTYFPHF-DLSHGSAQVKGH	59
HBA_PONPY	MVLSPADKTNVKTAWGKVGAHAGDYGAEALERMFLSFPTTKTYFPHF-DLSHGSAQVKDH	59
	** ** :: :.*:. . * :*. * : :* ***** * :: ** :.* *	
HBA_TREBE	GKKVMGGIALAVSKIDDLKTGLMELSEQHAYKLRVDPANFKILNHCILVVI STMFPKEFT	119
HBA_APTFO	GKKVAAALIEAANHIDDIAGALSKLSDLHAEKLRVDPVNFKLLGQC FMVVVAIHHP SALT	119
HBA_CANLF	GKKVADALTTAVAHLDLPGALSALSDLHAYKLRVDPVNFKLLSHCLLVTLACHHPTEFT	118
HBA_RAT	GKKVADALAKAADHVEDLPGALSTLSDLHAHKLRVDPVNFKFLSHCLLVTLACHHPGDFT	119
HBA_PIG	GQKVADALT KAVGHLDDLPGALSALSDLHAHKLRVDPVNFKLLSHCLLVTLAAHHPDDFN	118
HBA_MOUSE	GKKVADALASAAGHLDDLPGALSALSDLHAHKLRVDPVNFKLLSHCLLVTLASHHPADFT	119
HBA_HORSE	GKKVGDA LTLAVGHLDDLPGALSNLSDLHAHKLRVDPVNFKLLSHCLLVTLAVHLPNDFT	119
HBA_EQUAS	GKKVGDA LTLAVGHLDDLPGALSNLSDLHAHKLRVDPVNFKLLSHCLLVTLAVHLPNDFT	119
HBA_BOVIN	GAKVAAALTKAVEHLDDLPGALSELSDLHAHKLRVDPVNFKLLSHCLLVTLASHLP S DFT	119
HBA_HUMAN	GKKVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPVNFKLLSHCLLVTLAAHLP AEFT	119
HBA_PONPY	GKKVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPVNFKLLSHCLLVTLAAHLP AEFT	119
	* ** .: *. :*: : . * **: * *****.***:*. :. :. :. :. :. :. *	
HBA_TREBE	PEAHVSLDKFLSGVALALAE RYR	142
HBA_APTFO	PEIHASLDKFLCAVGNVLT SKYR	142
HBA_CANLF	PAVHASLDKFFAAVSTVLTSKYR	141
HBA_RAT	PAMHASLDKFLASVSTVLTSKYR	142
HBA_PIG	PSVHASLDKFLANVSTVLTSKYR	141
HBA_MOUSE	PAVHASLDKFLASVSTVLTSKYR	142
HBA_HORSE	PAVHASLDKFLSSVSTVLTSKYR	142
HBA_EQUAS	PAVHASLDKFLSTVSTVLTSKYR	142
HBA_BOVIN	PAVHASLDKFLANVSTVLTSKYR	142
HBA_HUMAN	PAVHASLDKFLASVSTVLTSKYR	142
HBA_PONPY	PAVHASLDKFLASVSTVLTSKYR	142
	* *.*****:. *. :*.:**	

FIGURE 29. Output MSA by Clustal Omega for set 1

CLUSTAL format alignment by MAFFT FFT-NS-i (v7.487)

```

HBA_HUMAN      MVLSPADKTNVKAAWGKVGAGHAGEYGAELERMFLSFPTTKTYFPHF-DLSHGSAQVKGH
HBA_PONPY      MVLSPADKTNVKTAWGKVGAGHAGDYGAELERMFLSFPTTKTYFPHF-DLSHGSAQVKDH
HBA_BOVIN      MVLSAADKGNVKAAWGKVGGAHAEYGAELERMFLSFPTTKTYFPHF-DLSHGSAQVKGH
HBA_HORSE      MVLSAADKTNVKAAWSKVGGHAGEYGAELERMFLGFPTTKTYFPHF-DLSHGSAQVKAH
HBA_EQUAS      MVLSAADKTNVKAAWSKVGGNAGEFGAEALERMFLGFPTTKTYFPHF-DLSHGSAQVKAH
HBA_MOUSE      MVLSGEDKSNIAAWGKIGGHGAEEYGAELERMFLSFPTTKTYFPHF-DVSHGSAQVKGH
HBA_RAT        MVLSADDKTNIKNCWGKIGGHGGEYGEELQRMFAAFPTTKTYFSHI-DVSPGSAQVKAH
HBA_PIG        -VLSAADKANVKAAWGKVGGAAGAHGAELERMFLGFPTTKTYFPHF-NLSHGSDQVKAH
HBA_CANLF      -VLSPADKTNIKSTWDKIGGHAGDYGGAEALDRTFQSPTTKTYFPHF-DLSPGSAQVKAH
HBA_APTFO      MVLSANDKSNVKSIFSKISSHAEYGAETLERMFTTYPQTKTYFPHF-DLHHGSAQVKAH
HBA_TREBE      -SLSDKDKAAVRALWSKIGKSADAIGNDALSRMIVVYPQTKTYFSHWPDVTPGSPHIKAH
                **  **  ::  :.*:.  .  *  :.*.*  :  :*  *****.*  ::  **  :.*  *

HBA_HUMAN      GKKVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPVNFKLLSHCLLVTLAAHLP AEFT
HBA_PONPY      GKKVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPVNFKLLSHCLLVTLAAHLP AEFT
HBA_BOVIN      GAKVAAALTKAVEHLDDLPGALSELSDLHAHKLRVDPVNFKLLSHSLLVTLASHLP SDFT
HBA_HORSE      GKKVGDALTLAVGHLDDLPGALSNLSDLHAHKLRVDPVNFKLLSHCLLSTLAVHLP NDFT
HBA_EQUAS      GKKVGDALTLAVGHLDDLPGALSNLSDLHAHKLRVDPVNFKLLSHCLLSTLAVHLP NDFT
HBA_MOUSE      GKKVADALASAAGHLDDLPGALSALSDLHAHKLRVDPVNFKLLSHCLLVTLASHHP ADFT
HBA_RAT        GKKVADALAKAADHVEDLPGALSTLSDLHAHKLRVDPVNFKFLSHCLLVTLACHHP GDFT
HBA_PIG        GQKVADALTKA VGHLDDLPGALSALSDLHAHKLRVDPVNFKLLSHCLLVTLAAHHP DDFN
HBA_CANLF      GKKVADALT TAVAH LDDLPGALSALSDLHAYKLRVDPVNFKLLSHCLLVTLACHHP TEFT
HBA_APTFO      GKKVAAALIEAANHIDDIAGALSKLSDLHAEKLRVDPVNFKLLGQCFMVVVAIHHP SALT
HBA_TREBE      GKKVMGGIALAVSKIDDLKTGLMELSEQHAYKLRVDPANFKILNHCILVVIISTMF PKFT
                *  **  .:  *.  ::*:  .*  **:  **  *****.**:*.:::  :::  *  :.

HBA_HUMAN      PAVHASLDKFLASVSTVLTSKYR
HBA_PONPY      PAVHASLDKFLASVSTVLTSKYR
HBA_BOVIN      PAVHASLDKFLANVSTVLTSKYR
HBA_HORSE      PAVHASLDKFLSSVSTVLTSKYR
HBA_EQUAS      PAVHASLDKFLSTVSTVLTSKYR
HBA_MOUSE      PAVHASLDKFLASVSTVLTSKYR
HBA_RAT        PAMHASLDKFLASVSTVLTSKYR
HBA_PIG        PSVHASLDKFLANVSTVLTSKYR
HBA_CANLF      PAVHASLDKFFAAVSTVLTSKYR
HBA_APTFO      PEIHASLDKFLCAVGNVLT SKYR
HBA_TREBE      PEAHVSLDKFLSGVALALAEYR
                *  *.*****:.  *.  .*:::**

```

FIGURE 30. Output MSA by MAFFT for set 1

CLUSTAL multiple sequence alignment by MUSCLE (3.8)

```

HBA_TREBE      -SLSDKDKAAVRALWSKIGKSADAIGNDALSRMIVVYPQTPTYFSHPDVTGPSPIKAH
HBA_APTFO      MVLSANDKSNVKSIFSKISSHAEYGAETLERMFTTYPQTPTYFPHF-DLHHGSAQVKAH
HBA_CANLF      -VLSPADKTNIKSTWDKIGGHAGDYGGAEALDRTFQSFPPTKTYFPHF-DLSPGSAQVKAH
HBA_RAT        MVLSADDKTNIKNCWGKIGGHGGEYGEALQRMFAAFPTTKTYFSHI-DVSPGSAQVKAH
HBA_PIG        -VLSAADKANVKAAGKVGQAGAHGAELERMFLGFPTTKTYFPHF-NLSHGSDQVKAH
HBA_MOUSE      MVLSGEDKSNIKAAGWKIGGHGAEGAEALERMFASFPTTKTYFPHF-DVSHGSAQVKGH
HBA_HORSE      MVLSAADKTNVKAAGSKVGGHAGYGAELERMFLGFPTTKTYFPHF-DLSHGSAQVKAH
HBA_EQUAS      MVLSAADKTNVKAAGSKVGGNAGEFGAEALERMFLGFPTTKTYFPHF-DLSHGSAQVKAH
HBA_BOVIN      MVLSAADKGNVKAAGKVGGHAAEGAEALERMFLSFPTTKTYFPHF-DLSHGSAQVKGH
HBA_HUMAN      MVLSPADKTNVKAAGKVGGAHAGYGAELERMFLSFPTTKTYFPHF-DLSHGSAQVKGH
HBA_PONPY      MVLSPADKTNVKTAWGKVGGAHAGDYGAELERMFLSFPTTKTYFPHF-DLSHGSAQVKDH
                **  **  .:  .:*.  .  *  :*:.*  :  :*  *****.*  ::  **  :*: *

HBA_TREBE      GKKVMGGIALAVSKIDDLKTGLMELSEQHAYKL RVD PANFKILNHCILVVI STMFPKEFT
HBA_APTFO      GKKVAAALIEAANHIDDIAGALSKLSDLHA EKL RVD PVNFKLLGQCFMVVVAIHHPSALT
HBA_CANLF      GKKVADALTTAVAHLDLPGALSALS D LHA YKL RVD PVNFKLLSHCLLVTLACHHPTEFT
HBA_RAT        GKKVADALAKAADHVEDLPGALSTLSDLHAHKL RVD PVNFKFLSHCLLVTLACHHPGDFT
HBA_PIG        GQKVADALTKAVGHLDDLPGALSALS D LHA HKL RVD PVNFKLLSHCLLVTLAAHHPDDFN
HBA_MOUSE      GKKVADALASAAGHLDDLPGALSALS D LHA HKL RVD PVNFKLLSHCLLVTLASHHPADFT
HBA_HORSE      GKKVGDA LTLAVGHLDDLPGALS N L S D LHA HKL RVD PVNFKLLSHCLLVTLAVHLPNDFT
HBA_EQUAS      GKKVGDA LTLAVGHLDDLPGALS N L S D LHA HKL RVD PVNFKLLSHCLLVTLAVHLPNDFT
HBA_BOVIN      GAKVAAALTKAVEHLDDLPGALSELSDLHAHKL RVD PVNFKLLSHSLLVTLASHLPDFT
HBA_HUMAN      GKKVADALTNAVAHVDDMPNALSALS D LHA HKL RVD PVNFKLLSHCLLVTLAAHLP AEFT
HBA_PONPY      GKKVADALTNAVAHVDDMPNALSALS D LHA HKL RVD PVNFKLLSHCLLVTLAAHLP AEFT
                *  **  .:  *.  :*:.*  .*  **:  **  *****.**:*.:.:.  .:  *  .:

HBA_TREBE      PEAHVSLDKFLSGVALALAERYR
HBA_APTFO      PEIHASLDKFLCAVGNVLT SKYR
HBA_CANLF      PAVHASLDKFFAAVSTVLTSKYR
HBA_RAT        PAMHASLDKFLASVSTVLTSKYR
HBA_PIG        PSVHASLDKFLANVSTVLTSKYR
HBA_MOUSE      PAVHASLDKFLASVSTVLTSKYR
HBA_HORSE      PAVHASLDKFLSSVSTVLTSKYR
HBA_EQUAS      PAVHASLDKFLSTVSTVLTSKYR
HBA_BOVIN      PAVHASLDKFLANVSTVLTSKYR
HBA_HUMAN      PAVHASLDKFLASVSTVLTSKYR
HBA_PONPY      PAVHASLDKFLASVSTVLTSKYR
                *  *.*****:.  *.  .*:..**

```

FIGURE 31. Output MSA by MUSCLE for set 1

Now, I will be putting these MSA aligned sequences (in the CLUSTAL format) to the code from question 3 to compute the given three score for each of these MSA alignments. I used the following code to check for covert the MSA obtained by CLUSTAL OMEGA, MAFFT, and MUSCLE into their respective dataframes.

```
1 import pandas as pd
2
3 # Reading the CLUSTAL Omega MSA file
4 clustal = []
5 with open("clustal.txt", 'r') as f:
6     line = f.read()
7     line_list = line.split("\n")
8
9     for i in range(len(line_list)):
10         line_i = line_list[i].split(" ")
11         line_i = [str(value) for value in line_i if value != ""]
12         clustal.append(line_i)
13
14 df1 = pd.DataFrame(clustal)
15 df1[2] = pd.to_numeric(df1[2])
16 df1[0] = pd.to_numeric(df1[0])
17
18 # Reading the MAFFT MSA file
19 mafft = []
20 with open("mafft.txt", 'r') as f:
21     line = f.read()
22     line_list = line.split("\n")
23
24     for i in range(len(line_list)):
25         line_i = line_list[i].split(" ")
26         line_i = [str(value) for value in line_i if value != ""]
27         mafft.append(line_i)
28
29 df2 = pd.DataFrame(mafft)
30 df2[2] = pd.to_numeric(df2[2])
31 df2[0] = pd.to_numeric(df2[0])
32
33 # Reading the MUSCLE MSA file
34 muscle = []
35 with open("muscle.txt", 'r') as f:
36     line = f.read()
37     line_list = line.split("\n")
38
39     for i in range(len(line_list)):
40         line_i = line_list[i].split(" ")
41         line_i = [str(value) for value in line_i if value != ""]
42         muscle.append(line_i)
43
44 df3 = pd.DataFrame(muscle)
45 df3[2] = pd.to_numeric(df3[2])
46 df3[0] = pd.to_numeric(df3[0])
```

LISTING 5. Code to create the dataframes for different MSAs

Now that I have the three dataframes for the MSA obtained by each of the alignment algorithms CLUSTAL OMEGA, MAFFT, and MUSCLE, I use the following code to list the similarities and differences.

```

1 similarities_3 = [] # All three same residue same position
2 differences_2 = [] # Any two different residue same position
3 differences_3 = [] # All three different residue same position
4
5 for i in range(len(df1)):
6     # Condition for checking any two sequences with different residues at same
        position
7     if df1[2][i]!=df2[2][i] or df1[2][i]!=df3[2][i] or df2[2][i]!=df3[2][i]:
8         print("Differences_2")
9         print(df1[2][i],df1[3][i],df2[2][i],df2[3][i],df3[2][i],df3[3][i],i)
10        differences_2.append([df1[2][i], df2[2][i], df3[2][i], i+1])
11
12    # Condition for checking all three sequences with different residues at
        same position
13    if df1[2][i]!=df2[2][i] and df1[2][i]!=df3[2][i] and df2[2][i]!=df3[2][i]:
14        print("Differences_3")
15        print(df1[2][i],df1[3][i],df2[2][i],df2[3][i],df3[2][i],df3[3][i],i)
16        differences_3.append([df1[2][i], df2[2][i], df3[2][i], i+1])
17
18    # Condition for checking all three sequences with similar residues at same
        position
19    if df1[2][i] == df2[2][i] and df1[2][i] == df3[2][i]:
20        print("Similarities_3")
21        print(df1[2][i],df1[3][i],df2[2][i],df2[3][i],df3[2][i],df3[3][i],i)
22        similarities_3.append([df1[2][i], df2[2][i], df3[2][i], i+1])
23
24 # Converting all of these lists into dataframes for easy visualization
25 df1 = pd.DataFrame(similarities_3, columns=["Clustal", "Mafft", "Muscle", "
        Position"])
26 df2 = pd.DataFrame(differences_3, columns=["Clustal", "Mafft", "Muscle", "
        Position"])
27 df3 = pd.DataFrame(differences_2, columns=["Clustal", "Mafft", "Muscle", "
        Position"])

```

LISTING 6. Code to find the similarities and differences in MSA

When I passed the above set 1 MSA, obtained by all three methods, to the code, unfortunately, all the differences dataframes were empty. This is indicative of the fact that all three MSA algorithms of CLUSTAL OMEGA, MAFFT, and MUSCLE aligned the given 11 sequences in set 1 in the exact same manner.



Clustal	Mafft	Muscle	Position

FIGURE 32. Output of differences_3 and differences_2

This is indicative of the fact that all the positions are same in all the three alignment techniques for the proteins in set 1. The similarities are:

Position	Clustal Residue	Mafft Residue	Muscle Residue	Score
3	Leucine	Leucine	Leucine	0.0
5	Proline	Proline	Proline	-1.121
143	Argenine	Argenine	Argenine	0.0

When I passed the above set 2 MSA, obtained by all three methods, to the code, unfortunately, the differences_3 dataframe was empty, however, the dataframes differences_2 and similarities_3 were not empty, indicating that differences and similarities exist upto some extent. This is indicative of the fact that all three MSA algorithms of CLUSTAL OMEGA, MAFFT, and MUSCLE do not align the given 9 sequences in set 2 in the exact same manner.

Clustal	Mafft	Muscle	Position
0	-0.974	-0.974	3
1	-1.494	-1.494	4
2	-0.377	-0.377	5
3	-0.736	-0.736	6
4	-0.377	-0.377	7
...
238	-0.349	-0.349	253
239	-0.684	-0.684	254
240	-0.530	-0.530	255
241	-1.465	-1.465	256
242	-1.427	-1.427	257

(A) similarities_3

Clustal	Mafft	Muscle	Position
0	-1.216	-1.214	1
1	-1.216	-1.214	2
2	-1.216	-1.214	38
3	-1.216	-1.214	39
4	-1.216	-1.214	59
5	-0.849	-0.562	71
6	-1.074	-1.149	72
7	-1.216	-1.214	106
8	-1.216	-1.214	107
9	-1.216	-1.214	125
10	-1.216	-0.849	162
11	-1.216	-0.684	163
12	-1.003	-1.214	164
13	-0.684	-1.214	165
14	-1.216	-1.214	258
15	-1.216	-1.214	259
16	-1.216	-1.214	260
17	-1.216	-1.214	261
18	-1.216	-1.214	262
19	-1.216	-1.214	263
20	-1.216	-1.214	264

(B) differences_2

FIGURE 33. The two non empty dataframes in MSA comparison for set 2

Another observation is that the differences observed in differences_2 is only the differences between either **mafft** or **muscle** and **clustal**. The mafft and muscle scores are exactly same throughout indicating their same alignment.

The similarities_3 are:

Position	Clustal Residue	Mafft Residue	Muscle Residue	Score
6	Lysine	Lysine	Lysine	-0.736
7	Phenylalanine	Phenylalanine	Phenylalanine	-0.377
255	Alanine	Alanine	Alanine	-0.530

The differences_2 are:

Position	Clustal	Score	Mafft	Score	Muscle	Score
71	Tyrosine	-0.849	Tyrosine	-0.562	Tyrosine	-0.562
72	Lysine	-1.074	Lysine	-1.149	Lysine	-1.149
162	Gap	*	Lysine	-0.849	Lysine	-0.849

The differences_2 are a total of 20 in number. The remaining are basically due to gap scores of clustal technique not aligning perfectly with the gap score of mafft and muscle techniques. Also, there are no occurrences of differences_3. The dataframe is empty.

Clustal	Mafft	Muscle	Position
---------	-------	--------	----------

FIGURE 34. Output of differences_3 for seq 2

Question 5. Check the scores manually at positions 9, 11, 20, 22 and 30 (use MSA from Clustal Omega)

Solution. Below are the images of manual calculation of scores. The scores calculated are unweighted frequencies, with scoring computational methods, entropy based, variance based, and sum-of-pairs based measures.

Also, in the sum-of-pairs method, after the entire summation, it needs to be squared and then square root should to be taken. This measure is to ensure that the value is positive.

Also, I saw a research paper on AL2CO that also had omitted the squaring and square rooting component in the sum-of-pairs method. Along with that, none of the values I computed showed a negative sign, hence I have dropped the squaring and square rooting component and computed the values. Below are the images of the manual calculations.

Set 1

9th position: [T, T, S, G, T, T, A, T, T, A, S]

$n[\text{Total}] = 11$	$n[T] = 6$	$n[S] = 2$	$n[A] = 2$	$n[G] = 1$
------------------------	------------	------------	------------	------------

Unweighted frequency: $f_a(i) = n_a(i)/n(i)$

$f_T(9) = [6/11]$ $f_S(9) = [2/11]$ $f_A(9) = [2/11]$ $f_G(9) = [1/11]$

Entropy based: $C^e(i) = \sum f_a(i) \cdot \ln(f_a(i))$, $a=1,20$

For all $a \notin \{T, A, G, S\}$ $f_a(i) = 0 \Rightarrow$ No impact on summation

For $a \in \{T, A, G, S\}$

$$C^e(9) = (6/11) \log(6/11) + (2/11) \log(2/11) + (2/11) \log(2/11) + (1/11) \log(1/11)$$

$$= -1.16851 \approx \boxed{-1.169}$$

$C^e(9) = -1.169$

Variance based: $C^v(i) = \{\sum [f_a(i) - f(i)]^2\}^{0.5}$, $a=1,20$

α	$f_a(i)$	$f(i)$
G	0.0909	0.0608
A	0.1818	0.1275
V	0.0	0.0807
L	0.0	0.1243
I	0.0	0.0147
T	0.5454	0.06089
S	0.1818	0.0833

α	$f_a(i)$	$f(i)$
M	0.0	0.025
C	0.0	0.0089
P	0.0	0.0448
F	0.0	0.0493
Y	0.0	0.0217
W	0.0	0.0071
H	0.0	0.0679

α	$f_a(i)$	$f(i)$
K	0.0	0.0807
R	0.0	0.0224
D	0.0	0.0641
E	0.0	0.0269
N	0.0	0.0256
Q	0.0	0.0115

This table contains the $f(i)$ which is same throughout this set of MSA. So, will be used ahead too.

$$\Rightarrow C^v(i) = [(0.0909 - 0.0608)^2 + \dots + (0.0 - 0.0115)^2]^{0.5} \approx \boxed{0.541}$$

Sum-of-pairs based: $C^p(i) = \{\sum_a \sum_b f_a(i) f_b(i) S_{ab}\}$ $a, b=1,20$

For all $b, a \notin \{T, A, G, S\}$ $f_a(i)$ or $f_b(i) = 0 \Rightarrow$ No impact on summation

T	0.545	-	-	-
A	-	0.1818	-	-
G	-	-	0.0909	-
S	-	-	-	0.1818

$f_a(i)$

T	5	-	-	-
A	0	4	-	-
G	-2	0	6	-
S	1	1	0	4

Blosum62 Matrix

T	A	G	S
---	---	---	---

$$C^p(i) = [0.545 * 0.545 * 5 + 0.545 * 0.1818 * 0 + \dots + 0.1818 * 0.1818 * 4] \approx \boxed{1.868}$$

FIGURE 35. Calculations for 9th position of set 1

11th position: [V, I, I, V, V, V, V, V, I, V, V]

$$n(\text{Total}) = 11 \quad n(V) = 8 \quad n(I) = 3$$

Unweighted frequency: $f_a(i) = n_a(i) / n(i)$

$$f_V(11) = [8/11] \quad f_I(11) = [3/11]$$

Entropy based: $C^e(i) = \sum f_a(i) \cdot \ln(f_a(i))$, $a=1, 20$

For all $a \notin \{V, I\}$, $f_a(i) = 0 \Rightarrow$ No impact on summation

For $a \in \{V, I\}$

$$C^e(11) = (8/11) \cdot \ln(8/11) + (3/11) \cdot \ln(3/11)$$

$$\approx -0.586$$

$$\Rightarrow C^e(11) \approx -0.586$$

Variance based: $C^v(i) = \{\sum [f_a(i) - f(i)]^2\}^{0.5}$, $a=1, 20$

The $f_a(i)$ is given above and $f(i)$ previous question

$$\Rightarrow C^v(11) = [(0.7272 - 0.0807)^2 + \dots + (0.2727 - 0.014)^2]$$

$$\approx 0.744$$

$$C^v(11) \approx 0.744$$

Sum-of-pairs based: $C^p(i) = \{\sum_a \sum_b f_a(i) \cdot f_b(i) \cdot S_{ab}\}$, $a, b=1, 20$

For all $a, b \notin \{V, I\}$, $f_a(i)$ or $f_b(i) = 0 \Rightarrow$ No impact on sum.

	V	I
V	0.727	—
I	—	0.272

$f_a(i)$

	V	I
V	4	—
I	3	4

Blosum 62
matrix

$$\Rightarrow C^p(11) = [0.727 \cdot 0.727 \cdot 4 + \dots + 0.272 \cdot 0.272 \cdot 4]$$

$$\approx 3.603$$

$$C^p(11) \approx 3.603$$

FIGURE 36. Calculations for 11th position of set 1

20th position: [A, G, G, G, G, G, A, G, K, S]

$$n(\text{Total}) = 11 \quad n(A) = 2 \quad n(G) = 7 \quad n(K) = 1 \quad n(S) = 1$$

Unweighted frequency: $f_a(i) = n_a(i)/n(i)$

$$f_A(20) = [2/11] \quad f_G(20) = [7/11] \quad f_K(20) = [1/11] \quad f_S(20) = [1/11]$$

Entropy based: $C^e(i) = \sum f_a(i) \cdot \ln(f_a(i))$, $a=1,20$.

For all $a \notin \{A, G, K, S\}$, $f_a(i) = 0 \Rightarrow$ No impact on sum

For $a \in \{A, G, K, S\}$

$$C^e(20) = (2/11) \cdot \ln(2/11) + (7/11) \cdot \ln(7/11) + (1/11) \cdot \ln(1/11) + (1/11) \cdot \ln(1/11)$$

$$\approx -1.034$$

$$C^e(20) \approx -1.034$$

Variance based: $C^v(i) = \{ \sum [f_a(i) - p(i)]^2 \}^{0.5}$, $a=1,20$

The $f_a(i)$ is above and $p(i)$ in first part.

$$\Rightarrow C^v(20) = [(0.6363 - 0.0608)^2 + \dots + (0.1818 - 0.0833)^2 + \dots + (0.09 - 0.0807)^2]^{0.5}$$

$$\approx 0.613 \Rightarrow C^v(20) \approx 0.613$$

Sum-of-pairs based: $C^p(i) = \{ \sum_a \sum_b f_a(i) \cdot f_b(i) \cdot S_{ab} \}$ $a, b=1,20$

For all $a, b \notin \{A, G, K, S\}$, $f_a(i)$ or $f_b(i) = 0 \Rightarrow$ No impact on sum

$f_a(i)$

	G	A	K	S
G	0.636	-	-	-
A	-	0.1818	-	-
K	-	-	0.0909	-
S	-	-	-	0.0909

	G	A	K	S
G	G	-	-	-
A	0	4	-	-
K	-2	-1	5	-
S	0	1	0	4

Blasum62
matrix

$$C^p(20) = [(0.636 * 0.636 * 6) + (0.636 * 0.0909 * 2) + \dots + (0.0909 * 0.0909 * 4)]$$

$$\approx 2.405$$

$$C^p(20) \approx 2.405$$

FIGURE 37. Calculations for 20th position of set 1

22nd position : [A, G, G, A, A, A, A, A, A, A]

$$n(\text{Total}) = 11 \quad n(A) = 9 \quad n(G) = 2$$

Unweighted frequency : $f_a(i) = n_a(i) / n(i)$

$$f_A(22) = [9/11] \quad f_G(22) = [2/11]$$

Entropy based : $C^e(i) = \sum f_a(i) \cdot \ln f_a(i)$, $a = 1, 20$.

For all $a \notin \{A, G\}$, $f_a(i) = 0 \Rightarrow$ No impact on sum.

For $a \in \{A, G\}$

$$C^e(22) = (2/11) \cdot \ln(2/11) + (9/11) \cdot \ln(9/11) \approx -0.474$$

$$C^e(22) \approx -0.474$$

Variance based : $C^v(i) = \{ \sum [f_a(i) - f(i)]^2 \}^{0.5}$, $a = 1, 20$.

The $f_a(i)$ is above and $f(i)$ was computed for first 9.

$$\Rightarrow C^v(22) = [(0.1818 - 0.0608)^2 + (0.818 - 0.1275)^2 + \dots + (0 - 0.0802)^2]^{0.5}$$

$$\approx 0.74$$

$$C^v(22) \approx 0.74$$

Sum-of-pairs based : $C^p(i) = \{ \sum_a \sum_b f_a(i) \cdot f_b(i) \cdot S_{ab} \}$, $a, b = 1, 20$

For all $b, a \notin \{A, G\}$, $f_a(i) = 0$ or $f_b(i) = 0 \Rightarrow$ No impact on sum

	A	G
A	0.8181	-
G	-	0.1818

$f_a(i)$

	A	G
A	4	-
G	0	6

Blosum62
matrix

$$\Rightarrow C^p(22) = [(0.8181 * 0.8181 * 4) + \dots + (0.1818 * 0.1818 * 6)]^{0.5}$$

$$\approx 2.876$$

$$C^p(22) \approx 2.876$$

FIGURE 38. Calculations for 22th position of set 1

30th position: [L, L, L, L, L, L, L, L, L, L, L]

$$n(\text{Total}) = 11 \quad n(L) = 11$$

Unweighted frequency: $f_a^u(i) = n_a(i) / n(i)$

$$f_L(30) = [11 / 11] = 1$$

Entropy based: $C^e(i) = \sum f_a(i) \cdot \ln(f_a(i))$, $a = 1, 20$.

For all $a \notin \{L\}$, $f_a(i) = 0 \Rightarrow$ No impact on sum.

For $a \in \{L\}$

$$C^e(30) = (1/11) \cdot \ln(1/11) = 0$$

$$\boxed{C^e(30) = 0}$$

Variance based: $C^v(i) = \left\{ \sum [f_a(i) - f(i)]^2 \right\}^{0.5}$

The $f_a(i)$ is above and $f(i)$ was computed for 1st Q.

$$\Rightarrow C^v(30) = [(1 - 0.12435)^2 + (0 - 0.0807)^2 + \dots + (0 - 0.0115)^2]^{0.5}$$

$$\approx 0.909$$

$$\boxed{C^v(30) \approx 0.909}$$

Sum-of-pairs method: $C^p(i) = \left\{ \sum_a \sum_b f_a(i) \cdot f_b(i) \cdot S_{ab} \right\}$ $a = 1, 20$

For all $a, b \notin \{L\}$, $f_a(i)$ or $f_b(i) = 0 \Rightarrow$ No impact on sum.

	L
L	1

$f_a(i)$

	L
L	4

Blosum62 matrix

$$\Rightarrow C^p(30) = [1 * 1 * 4]$$

$$\Rightarrow \boxed{C^p(30) = 4}$$

FIGURE 39. Calculations for 30th position of set 1

Question 6. Obtain the conservation score of 1BTM, A-chain using Consurf server <https://consurf.tau.ac.il/>

Solution. First thing I need to find is the UniProt ID. I searched for the 1BTM, A chain. The below is the structure I found along with the accession number of the protein. The accession number of the protein is **P00943**. I searched for the same on the UniProtKB and found the similarity in structure, and other features and structural elements. Below is the structure I found initially.

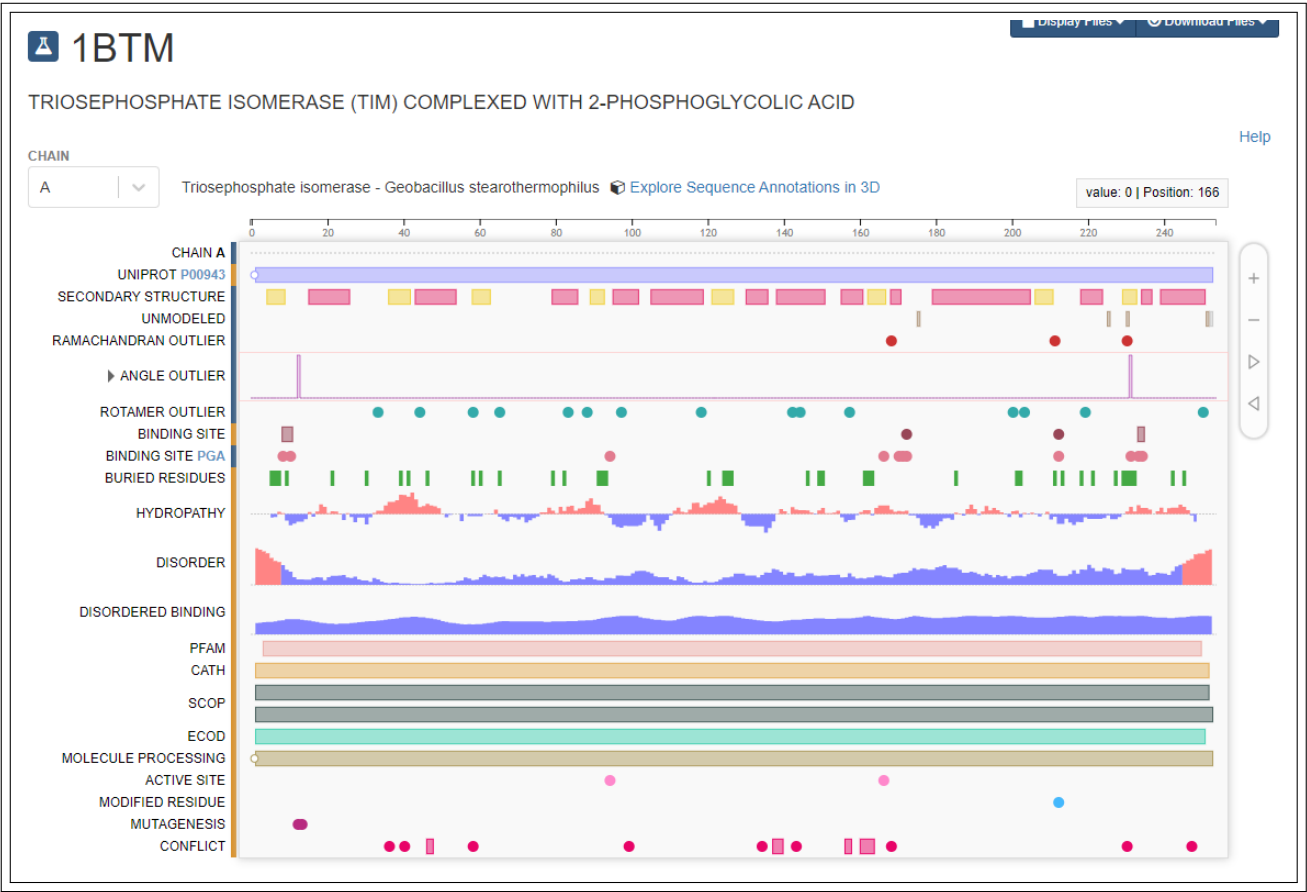


FIGURE 40. Finding the protein sequence UniProt ID

Protein sequence of 1BTM A-chain obtained from UniProtKB

```
>P00943
MRKPIIAGNWKMHKTLAEAVQFVEDVKGHVPPADEVISVVCAPFLFLDRLVQAAD
GTDLKIGAQTMMHFADQGAYTGEVSPVMLKDLGVTYVILGHSERRQMFAETDETVN
KKVLAAFTRLGPIIICCGESLEEREAGQTNAVVASQVEKALAGLTPEQVKQAVIA
YEPIWAIGTGKSSTPEDANSVCGHIRSVVSRLFGPEAAEAIRIQYGGSVKPDNIR
DFLAQQQIDGPLVGASLEPASFLQLVEAGRHE
```

LISTING 7. The protein sequence of the protein mentioned in the question

The above FASTA file is given as input to the **Consurf** server. There also exists an alternative approach to execute the above given task. In the input page of the Consurf server, there is an option to enter the **PDB or UniProt ID**. Enter the term **1BTM** there. Then under the section of **Select the chain identifier**, choose **Chain A**. Now enter the job title and email ID and **Run with default parameters**.

Below are two images, one for the input screen on the Consurf server, the other is the running parameters on Consurf server.

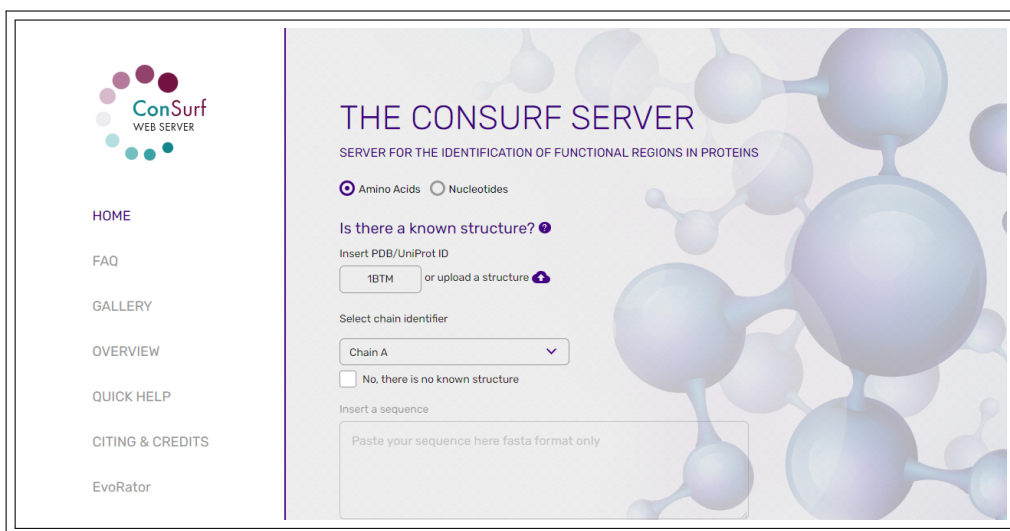


FIGURE 41. Input screen on the Consurf server

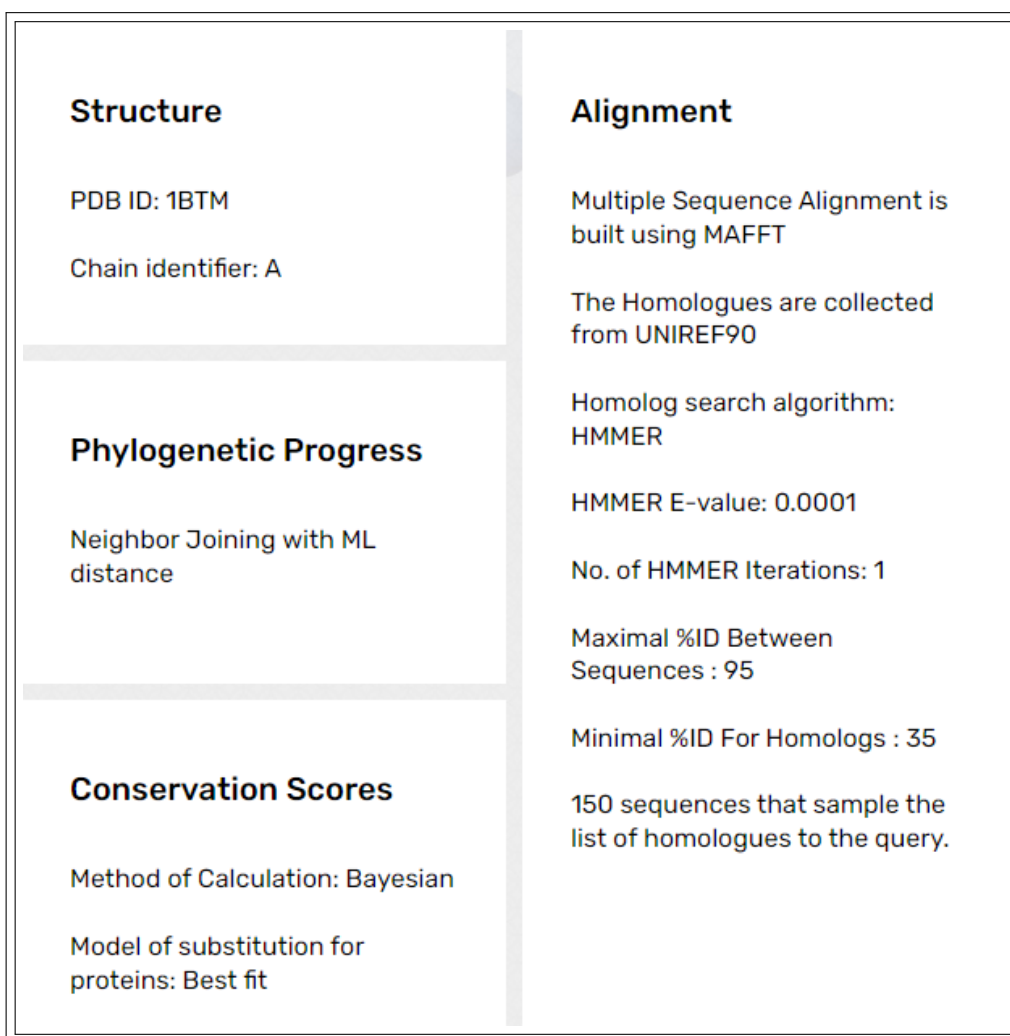


FIGURE 42. Running parameters of Consurf server

The results page for the above run on the Consurf server shows up as below. It consists of the structure of the protein sequence, along with its sequence and highlighted are highly and lowly conserved regions:

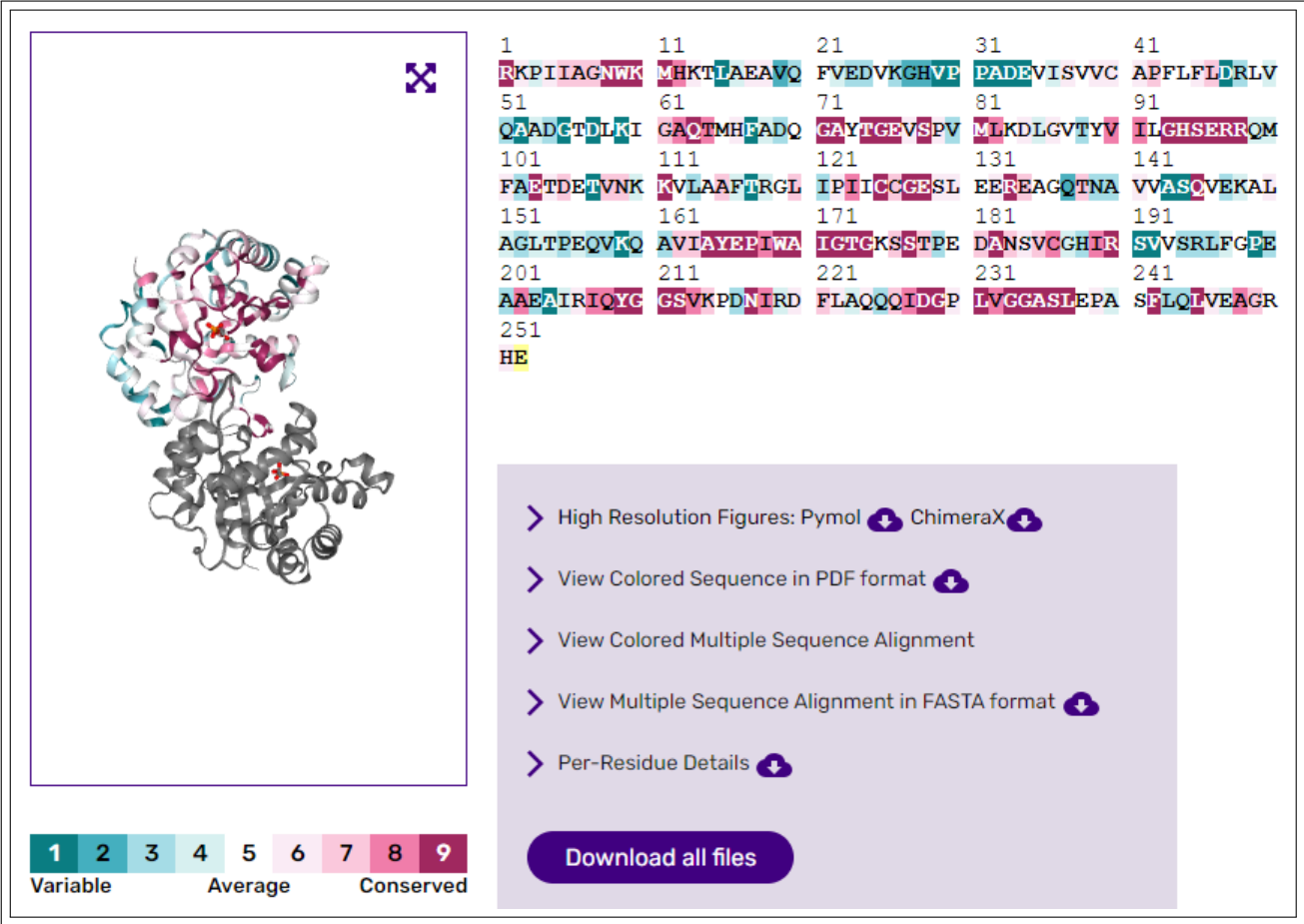


FIGURE 43. Results page

Given below are some of the parameters that play a role in assigning conservation scores, like, layers for assigning grades, confidence interval colors, residue variety.

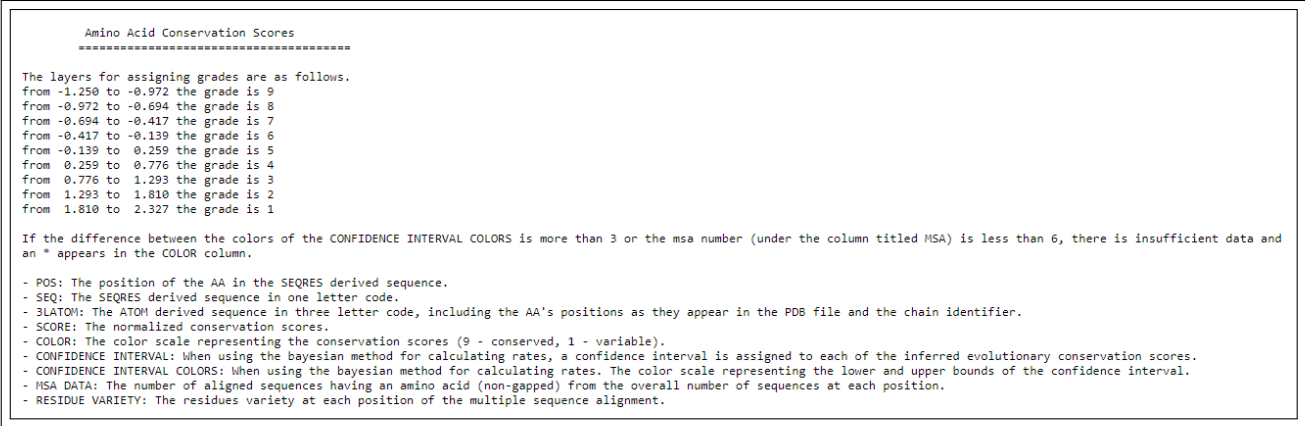
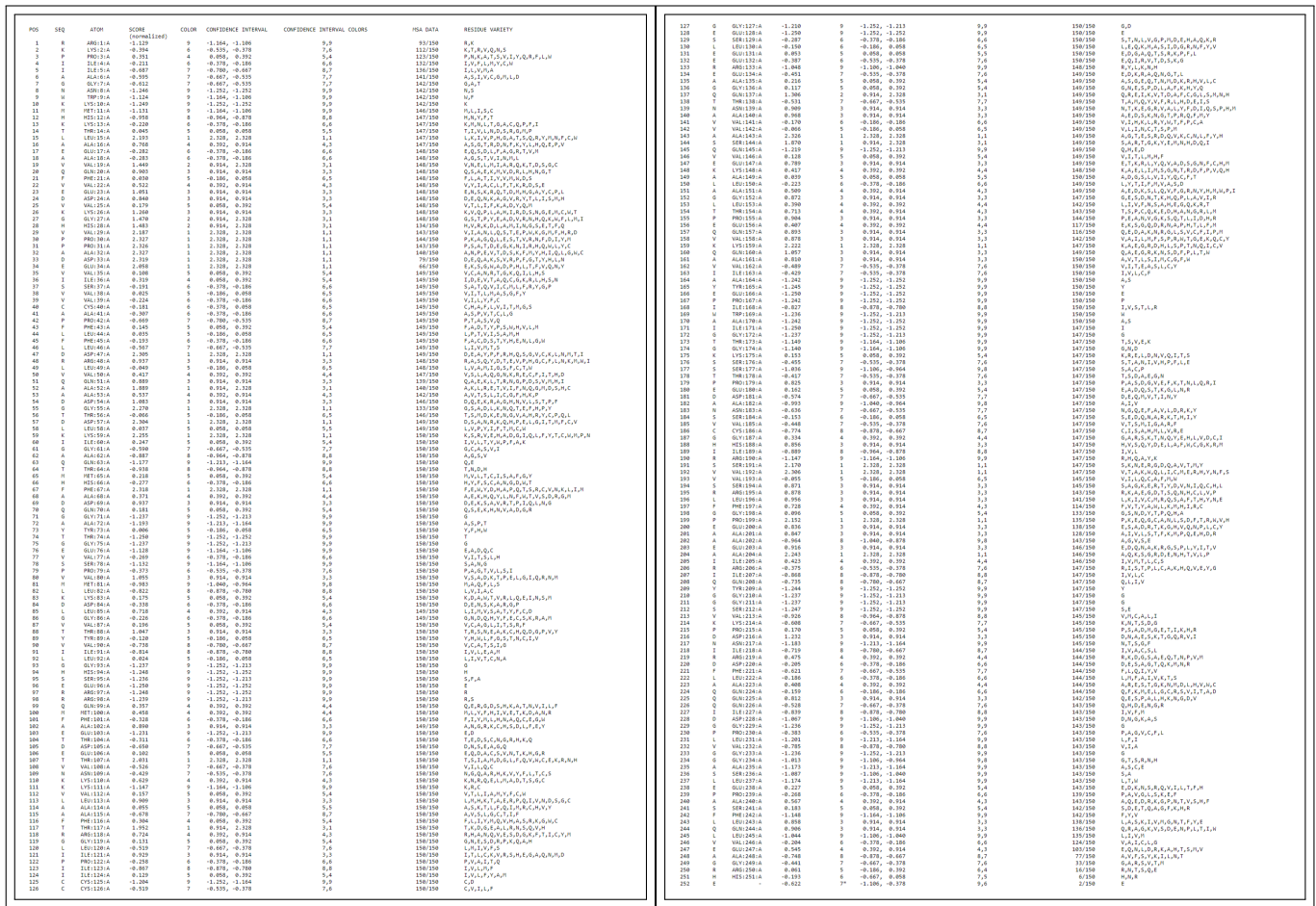


FIGURE 44. Parameters to compute the conservation scores

Now, given below is the set of all 252 residues in the given protein and also all properties related to each residue position, like, SEQ (amino acid), COLOR (indicating conservation), CONFIDENCE INTERVAL (indicate the range of conservation values for COLOR), RESIDUE VARIETY (presence of multiple residues at same position), etc.



(A) 1-126 positions

(B) 126-252 positions

FIGURE 45. Each residue-wise properties for the given protein

Names and Taxonomy related to protein from UniProtKB:

- (1) Recommended name is **Triosephosphate isomerase**
- (2) EC number is **EC:5.3.1.1**
- (3) Its optimal temperature is 60 degrees Celsius. It is thermostable
- (4) It plays a role in the following pathways: Carbohydrate biosynthesis; gluconeogenesis; Carbohydrate degradation; glycolysis; D-glyceraldehyde 3-phosphate from glycerone phosphate

The color scale in all of the above images are indicative of the conservation scores, with 9 indicating **conserved**, while 1 indicating **variable**. These numeric values have been assigned to different confidence interval ranges for the conservation scores. The split up is like this:

- from -1.250 to -0.972 the grade is 9
- from -0.972 to -0.694 the grade is 8
- from -0.694 to -0.417 the grade is 7
- from -0.417 to -0.139 the grade is 6
- from -0.139 to 0.259 the grade is 5
- from 0.259 to 0.776 the grade is 4
- from 0.776 to 1.293 the grade is 3
- from 1.293 to 1.810 the grade is 2
- from 1.810 to 2.327 the grade is 1

The ConSurf can also perform multiple sequence alignment with some predefined sequences based on their closeness to the given sequence. Below image is an example of one such MSA.

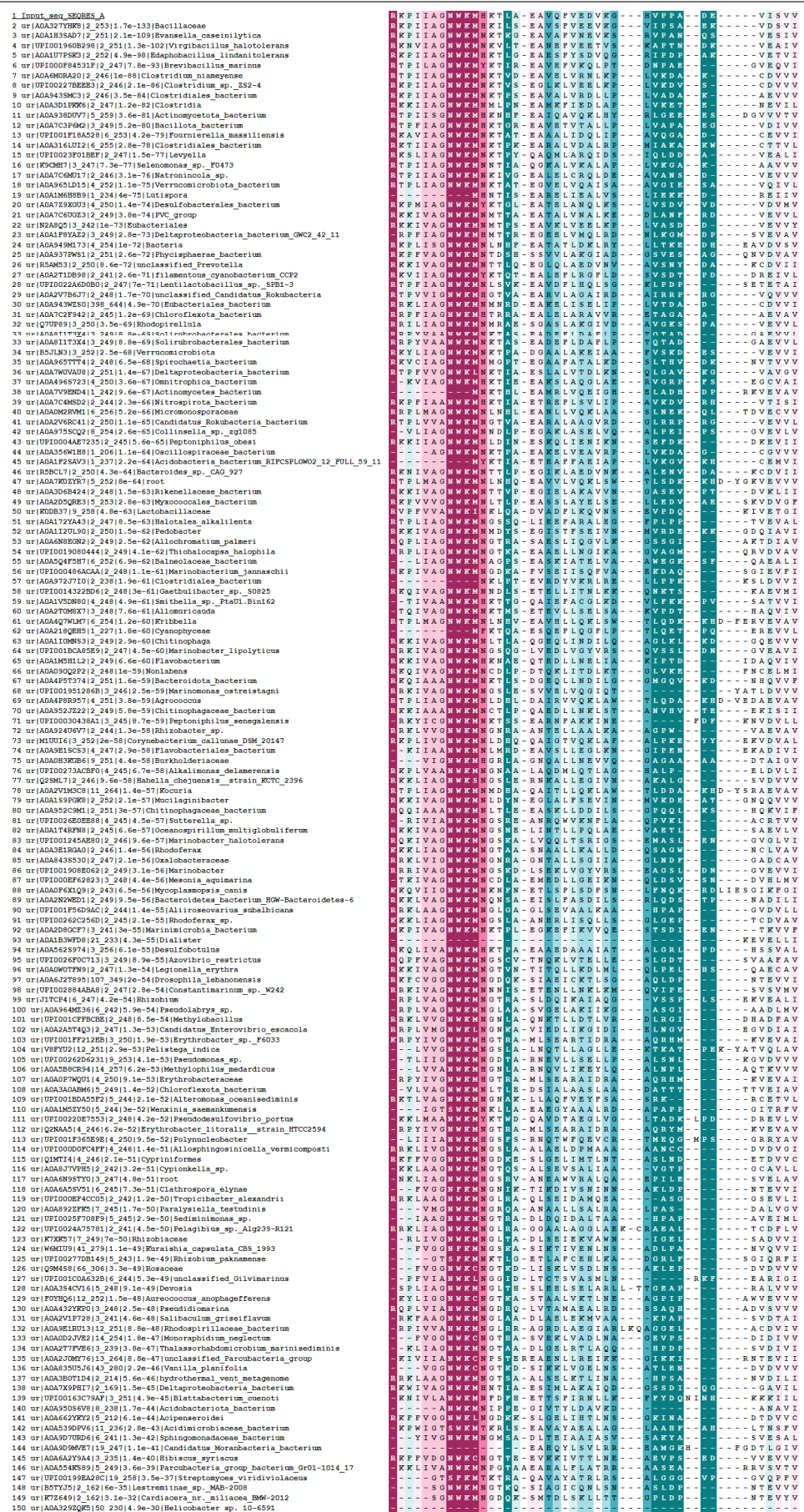


FIGURE 46. MSA on Consurf Server

Some additional results obtained from the Consurf server are given below in the image:

There are [28566 HMMER hits](#). 27844 of them are unique, including the query.

The calculation is performed on a sample of [150 sequences](#) that represent the list of homologues to the query.

Here is the [list of sequences](#) that produced significant alignments, but were not chosen as hits.

The best evolutionary model was selected to be: WAG. ([details](#)).

FIGURE 47. Additional Homology related results