```
set.seed(123)

# 1. Install and Load Required Packages ----

# This block installs and loads the necessary libraries.

# It first checks if the packages are installed, and if not, installs them.


required_packages <- c("NOISeq", "pheatmap", "ggplot2", "RColorBrewer", "cowplot")

for (pkg in required_packages) {

 if (!requireNamespace(pkg, quietly = TRUE)) {

  install.packages(pkg)

 }

 library(pkg, character.only = TRUE)

}


# 2. Load RNA-seq Data ----

# Reads the RNA-seq data from a local file, selecting only the count matrix columns.


rna_seq_data <-
read.table("C:/Users/talkt/Dropbox/PC/Downloads/GSE159717_rnaseq_deseq_5dpi_counts_raw.tsv",

                   header = TRUE, row.names = 1, sep = "\t")

count_matrix <- rna_seq_data[, 6:ncol(rna_seq_data)]

sample_names <- colnames(count_matrix)


# 3. Define Experimental Conditions ----

# Creates a 'coldata' data frame with sample names and corresponding conditions (Rem, SARS, mock).


conditions <- ifelse(grepl("Rem", sample_names), "Rem",

              ifelse(grepl("SARS", sample_names), "SARS", "mock"))

coldata <- data.frame(sample = sample_names, condition = factor(conditions))

rownames(coldata) <- sample_names
```

```
# 4. Create NOISeq Data Object ----

# Prepares the RNA-seq data for analysis with NOISeq.


noidata <- readData(data = count_matrix, factors = coldata)

print(noidata)


# 5. Differential Expression Analysis (SARS vs Mock) ----

# Runs the NOISeqbio function for SARS vs Mock comparison and writes the results to a CSV file.


noi_SARS_vs_Mock <- noiseqbio(noidata, k = 0.5, norm = "tmm", factor = "condition", conditions = c("SARS", "mock"))

write.csv(noi_SARS_vs_Mock@results[[1]], "sig_SARS_vs_Mock_NOISeq5.csv", row.names = TRUE)


# 6. Differential Expression Analysis (Rem vs Mock) ----

# Runs the NOISeqbio function for Rem vs Mock comparison and writes the results to a CSV file.


noi_Rem_vs_Mock <- noiseqbio(noidata, k = 0.5, norm = "tmm", factor = "condition", conditions = c("Rem", "mock"))

write.csv(noi_Rem_vs_Mock@results[[1]], "sig_Rem_vs_Mock_NOISeq5.csv", row.names = TRUE)


# 7. Volcano Plot Creation ----

# Create Volcano plots for SARS vs Mock and Rem vs Mock comparisons.


volcano_data_sars <- noi_SARS_vs_Mock@results[[1]]

volcano_data_rem <- noi_Rem_vs_Mock@results[[1]]


# Add significance criteria based on probability values.

volcano_data_sars$significant <- ifelse(volcano_data_sars$prob >= 0.95, "Significant", "Not Significant")
```

```
volcano_data_rem$significant <- ifelse(volcano_data_rem$prob >= 0.95, "Significant", "Not Significant")


# Define colors for significant and non-significant points.

colors <- c("Not Significant" = "grey", "Significant" = "red")


# Create Volcano plot for SARS vs Mock.

volcano_plot_sars <- ggplot(volcano_data_sars, aes(x = log2FC, y = -log10(prob), color = significant)) +

 geom_point(alpha = 0.6, na.rm = TRUE) +

 scale_color_manual(values = colors) +

 labs(title = "Volcano Plot: SARS vs Mock", x = "Log2 Fold Change", y = "-log10 Probability") +

 theme_minimal()


# Create Volcano plot for Rem vs Mock.

volcano_plot_rem <- ggplot(volcano_data_rem, aes(x = log2FC, y = -log10(prob), color = significant)) +

 geom_point(alpha = 0.6, na.rm = TRUE) +

 scale_color_manual(values = colors) +

 labs(title = "Volcano Plot: Rem vs Mock", x = "Log2 Fold Change", y = "-log10 Probability") +

 theme_minimal()


# Combine the two volcano plots side by side using cowplot.

combined_volcano <- plot_grid(volcano_plot_sars, volcano_plot_rem, labels = c("A", "B"), ncol = 2)

print(combined_volcano)


# 8. Principal Component Analysis (PCA) ----

# Performs PCA to visualize data variability across conditions (SARS, Rem, Mock).


pca_results <- dat(noidata, type = "PCA")

pca_scores <- as.data.frame(pca_results@dat$result$scores)
```

```
colnames(pca_scores) <- c("PC1", "PC2")
rownames(pca_scores) <- coldata$sample
pca_scores$sample <- rownames(pca_scores)

# Merge PCA Scores with Conditions.
pca_scores <- merge(pca_scores, coldata, by = "sample", all.x = TRUE)

# Calculate Explained Variance for PCA components.
explained_var <- pca_results@dat$result$var.exp[, 1] * 100
pc1_label <- paste0("PC1: ", round(explained_var[1], 1), "% variance")
pc2_label <- paste0("PC2: ", round(explained_var[2], 1), "% variance")

# Plot the PCA results.
ggplot(pca_scores, aes(x = PC1, y = PC2, color = condition)) +
 geom_point(size = 5, alpha = 0.8) +
 labs(title = "PCA across SARS, Rem, and Mock", x = pc1_label, y = pc2_label) +
 theme_minimal(base_size = 15) +
 scale_color_brewer(palette = "Set1", name = "Group")

# 9. Heatmap of Correlation Matrix ----
# Computes the correlation matrix of samples and generates a heatmap.

cor_matrix <- cor(count_matrix)
sample_labels <- colnames(count_matrix)

pheatmap(cor_matrix,
      color = colorRampPalette(c("blue", "white", "red"))(100),
      clustering_distance_rows = "euclidean",
      clustering_distance_cols = "euclidean",
      clustering_method = "complete",
      labels_row = sample_labels,
```

```
      labels_col = sample_labels,
      legend_breaks = seq(0.99, 1, by = 0.0025),
      legend_labels = round(seq(0.99, 1, by = 0.0025), 3),
      main = "Correlation Heatmap of RNA-seq Samples",
      fontsize = 10, fontsize_row = 8, fontsize_col = 8,
      display_numbers = TRUE, number_format = "%.2f", border_color = NA)

# 10. MA Plot Creation ----
# MA plots display mean expression vs log2 fold change for SARS vs Mock and Rem vs Mock.

# Extract data for SARS vs Mock (before shrinkage).
ma_data_before_shrink <- as.data.frame(noi_SARS_vs_Mock@results[[1]])

# Create MA Plot (Before Shrinkage).
ggplot(ma_data_before_shrink, aes(x = SARS_mean, y = log2FC)) +
 geom_point(alpha = 0.4) +
 labs(title = "MA Plot: SARS vs Mock (Before Shrinkage)", x = "Mean of Normalized Counts
(SARS)", y = "Log2 Fold Change") +
 theme_minimal() +
 scale_y_continuous(limits = c(-2, 2))

# Manual Shrinkage of log2 fold change for SARS vs Mock.
ma_data_after_shrink <- ma_data_before_shrink
ma_data_after_shrink$log2FC_shrunk <- ma_data_before_shrink$log2FC * 0.5

# Create MA Plot (After Shrinkage).
ggplot(ma_data_after_shrink, aes(x = SARS_mean, y = log2FC_shrunk)) +
 geom_point(alpha = 0.4) +
 labs(title = "MA Plot: SARS vs Mock (After Shrinkage)", x = "Mean of Normalized Counts (SARS)",
y = "Log2 Fold Change (Shrunk)") +
 theme_minimal() +
 scale_y_continuous(limits = c(-2, 2))
```

```
# Extract data for Rem vs Mock (before shrinkage) and generate similar plots.
ma_data_rem_before_shrink <- as.data.frame(noi_Rem_vs_Mock@results[[1]])

ggplot(ma_data_rem_before_shrink, aes(x = Rem_mean, y = log2FC)) +
  geom_point(alpha = 0.4) +
  labs(title = "MA Plot: Rem vs Mock (Before Shrinkage)", x = "Mean of Normalized Counts (Rem)",
y = "Log2 Fold Change") +
  theme_minimal() +
  scale_y_continuous(limits = c(-2, 2))

# Manually Apply Shrinkage to Rem vs Mock log2 fold change.
ma_data_rem_after_shrink <- ma_data_rem_before_shrink
ma_data_rem_after_shrink$log2FC_shrunk <- ma_data_rem_after_shrink$log
# 10. MA Plot Creation (continued) ----

# Manually Apply Shrinkage to Rem vs Mock log2 fold change.
ma_data_rem_after_shrink <- ma_data_rem_before_shrink
ma_data_rem_after_shrink$log2FC_shrunk <- ma_data_rem_before_shrink$log2FC * 0.5

# Create MA Plot (After Shrinkage) for Rem vs Mock.
ggplot(ma_data_rem_after_shrink, aes(x = Rem_mean, y = log2FC_shrunk)) +
  geom_point(alpha = 0.4) +
  labs(title = "MA Plot: Rem vs Mock (After Shrinkage)",
       x = "Mean of Normalized Counts (Rem)",
       y = "Log2 Fold Change (Shrunk)") +
  theme_minimal() +
  scale_y_continuous(limits = c(-2, 2))

# 11. Differential Expression Analysis: Rem vs SARS ----
# This section runs the differential expression analysis for the Rem vs SARS comparison.
```

```
noi_Rem_vs_SARS <- noiseqbio(noidata, k = 0.5, norm = "tmm", factor = "condition", conditions = c("Rem", "SARS"))

# Extract the results and save them to a CSV file.
rem_vs_sars_results <- noi_Rem_vs_SARS@results[[1]]
write.csv(rem_vs_sars_results, "rem_vs_sars_NOISeq.csv", row.names = TRUE)

# Create the MA plot for Rem vs SARS comparison (Before Shrinkage).
ma_data_before_rem_vs_sars <- as.data.frame(rem_vs_sars_results)

ggplot(ma_data_before_rem_vs_sars, aes(x = Rem_mean, y = log2FC)) +
  geom_point(alpha = 0.4, na.rm = TRUE) +
  labs(title = "MA Plot: Rem vs SARS (Before Shrinkage)",
       x = "Mean of Normalized Counts (Rem)",
       y = "Log2 Fold Change") +
  theme_minimal()

# Manually Apply Shrinkage to log2 fold change for Rem vs SARS.
ma_data_after_rem_vs_sars <- ma_data_before_rem_vs_sars
ma_data_after_rem_vs_sars$log2FC_shrunk <- ma_data_before_rem_vs_sars$log2FC * 0.5

# Create the MA Plot for Rem vs SARS (After Shrinkage).
ggplot(ma_data_after_rem_vs_sars, aes(x = Rem_mean, y = log2FC_shrunk)) +
  geom_point(alpha = 0.4, na.rm = TRUE) +
  labs(title = "MA Plot: Rem vs SARS (After Shrinkage)",
       x = "Mean of Normalized Counts (Rem)",
       y = "Log2 Fold Change (Shrunk)") +
  theme_minimal()

# 12. Summary of Differential Expression Results ----
```

```
# This section summarizes the number of significantly differentially expressed genes for each comparison.

# Subset SARS vs Mock results for significant genes (prob >= 0.95, log2FC > 0.6).
filtered_sars_mock <- subset(noi_SARS_vs_Mock@results[[1]],
                             prob >= 0.95 & abs(log2FC) > 0.6)
num_filtered_sars_mock <- nrow(filtered_sars_mock)
cat("Number of filtered genes for SARS vs Mock:", num_filtered_sars_mock, "\n")

# Subset Rem vs Mock results for significant genes (prob >= 0.95, log2FC > 0.6).
filtered_rem_mock <- subset(noi_Rem_vs_Mock@results[[1]],
                            prob >= 0.95 & abs(log2FC) > 0.6)
num_filtered_rem_mock <- nrow(filtered_rem_mock)
cat("Number of filtered genes for Rem vs Mock:", num_filtered_rem_mock, "\n")

# Subset Rem vs SARS results for significant genes (prob >= 0.95, log2FC > 0.6).
filtered_rem_vs_sars <- subset(noi_Rem_vs_SARS@results[[1]],
                               prob >= 0.95 & abs(log2FC) > 0.6)
num_filtered_rem_vs_sars <- nrow(filtered_rem_vs_sars)
cat("Number of filtered genes for Rem vs SARS:", num_filtered_rem_vs_sars, "\n")

> sessionInfo()
R version 4.4.1 (2024-06-14 ucrt)
Platform: x86_64-w64-mingw32/x64
Running under: Windows 11 x64 (build 22631)

Matrix products: default


locale:
[1] LC_COLLATE=English_India.utf8  LC_CTYPE=English_India.utf8
```

[3] LC_MONETARY=English_India.utf8 LC_NUMERIC=C

[5] LC_TIME=English_India.utf8

time zone: Australia/Brisbane

tzcode source: internal

attached base packages:

[1] splines stats graphics grDevices utils datasets methods

[8] base

other attached packages:

[1] cowplot_1.1.3 RColorBrewer_1.1-3 ggplot2_3.5.1

[4] pheatmap_1.0.12 NOISeq_2.48.0 Matrix_1.7-0

[7] Biobase_2.64.0 BiocGenerics_0.50.0

loaded via a namespace (and not attached):

[1] vctrs_0.6.5 cli_3.6.3 rlang_1.1.4 generics_0.1.3

[5] labeling_0.4.3 glue_1.8.0 colorspace_2.1-1 scales_1.3.0

[9] fansi_1.0.6 grid_4.4.1 munsell_0.5.1 tibble_3.2.1

[13] lifecycle_1.0.4 compiler_4.4.1 dplyr_1.1.4 pkgconfig_2.0.3

[17] farver_2.1.2 lattice_0.22-6 R6_2.5.1 tidyselect_1.2.1

[21] utf8_1.2.4 pillar_1.9.0 magrittr_2.0.3 withr_3.0.1

[25] gtable_0.3.5

>