annotate

2022-10-17

```
Load data
```

```
data <- read_tsv("sample.txt.gz")</pre>
## Rows: 4540462 Columns: 7
## -- Column specification -----
## Delimiter: "\t"
## chr (5): sample.id, replica, nt.seq, best.V.gene, aa.seq
## dbl (2): time.point, clone.count
##
## i Use `spec()` to retrieve the full column specification for this data.
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.
database <- read_tsv("database.txt.gz")</pre>
## Rows: 405 Columns: 2
## -- Column specification -------
## Delimiter: "\t"
## chr (2): aa.seq.db, epitope
## i Use `spec()` to retrieve the full column specification for this data.
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.
Data structure is better be kept the way specified below
glimpse(data)
## Rows: 4,540,462
## Columns: 7
## $ sample.id <chr> "S2", "S2
                                                         <chr> "F1", "F1", "F1", "F1", "F1", "F1", "F1", "F1", "F1", "F1"~
## $ replica
## $ clone.count <dbl> 5160, 2317, 2154, 2073, 1583, 915, 845, 811, 673, 673, 666~
## $ nt.seq
                                                         <chr> "CGTGCCAGCAGCGCCCGGACTAGCGGGAGTAGGGACAATGAGCAGTTCTTC", "TG~
## $ best.V.gene <chr> "TRBV7-3*00", "TRBV6-2*00", "TRBV27*00", "TRBV20-1*00", "T~
                                                         <chr> "RASSARTSGSRDNEQFF", "CASSYRGTAWETQYF", "CASRPLLDRNNEQFF",~
## $ aa.seq
glimpse(database)
## Rows: 405
## Columns: 2
## $ aa.seq.db <chr> "CSVVDAAPGANVLTF", "CAWSPGPVNEQFF", "CSARASYEQYF", "CASSDSGT~
                                                 <chr> "LLWNGPMAV", "LLWNGPMAV, "
## $ epitope
Compute distances between strings
```

```
get_distances <- function(aa.seq.1, aa.seq.2, threshold = 1,</pre>
                           method = "hamming", ...) {
  stringdistmatrix(unique(aa.seq.1), unique(aa.seq.2),
                    method = method,
                    useNames = T, ...) %>%
    melt %>%
    filter(value <= threshold) %>%
    rename(aa.seq = Var1, aa.seq.db = Var2, dist = value) %>%
    mutate(aa.seq = as.character(aa.seq), aa.seq.db = as.character(aa.seq.db))
}
with(database, get_distances(aa.seq.db, aa.seq.db)) %>% head
##
                                aa.seq.db dist
                aa.seq
## 1
       CSVVDAAPGANVLTF
                          CSVVDAAPGANVLTF
## 2
         CAWSPGPVNEQFF
                            CAWSPGPVNEQFF
## 3
           CSARASYEQYF
                              CSARASYEQYF
                                              0
## 4
         CASSDSGTDTQYF
                            CASSDSGTDTQYF
## 5
        CASSFGTGRAGYTF
                           CASSFGTGRAGYTF
                                              0
## 6 CASSDWGGTGRGPEAFF CASSDWGGTGRGPEAFF
An optimized routine that splits by length and processes in chunks(hamming only)
get_1mm_pairs <- function(aa.seq, aa.seq.db, chunks = 64) {</pre>
  d <- tibble(aa.seq = unique(aa.seq)) %>%
    mutate(len = nchar(aa.seq),
           chunk.id = rep(1:chunks, length.out = length(unique(aa.seq))))
  db <- tibble(aa.seq.db = unique(aa.seq.db)) %>%
    mutate(len.db = nchar(aa.seq.db))
  d %>%
    group_by(chunk.id, len) %>%
    group_modify(~ get_distances(.x$aa.seq, db %>%
                                    filter(len.db == .y$len) %>%
                                     .$aa.seq.db))
}
with(database, get_1mm_pairs(aa.seq.db, aa.seq.db)) %>% head
## # A tibble: 6 x 5
## # Groups:
               chunk.id, len [4]
##
     chunk.id
                len aa.seq
                                      aa.seq.db
                                                       dist
##
        <int> <int> <chr>
                                      <chr>
                                                       <dbl>
                                     CASSLMYEQYF
## 1
            1
                 11 CASSLMYEQYF
                                                           0
## 2
            1
                 12 CASSEGIYGYTF
                                     CASSEGIYGYTF
                                                           0
## 3
            1
                 13 CATTGGSGYEQYF
                                     CATTGGSGYEQYF
                                                           0
## 4
            1
                 13 CASSGSSGYEQYF
                                     CASSGSSGYEQYF
                                                           0
## 5
                 13 CSASHRAGNEQYF
                                     CSASHRAGNEQYF
                                                           0
                 15 CSVVDAAPGANVLTF CSVVDAAPGANVLTF
Now the general routine for tables in original format. Sample table should come first, database should come
```

Now the general routine for tables in original format. Sample table should come first, database should come second.

```
get_1mm_annot <- function(d, db) {
  pairs <- get_1mm_pairs(d$aa.seq, db$aa.seq.db) %>%
```

```
inner_join(db)
 d %>%
   left_join(pairs) %>%
   select(-chunk.id, -len)
}
get_1mm_annot(data %>% head(100000), database) %>%
 filter(!is.na(epitope)) %>% head
## Joining, by = "aa.seq.db"
## Joining, by = "aa.seq"
## # A tibble: 6 x 10
    sample.id replica time.point clone.count nt.seq
                                                best.V.gene aa.seq aa.seq.db
##
                        <dbl>
##
    <chr>>
             <chr>
                                   <dbl> <chr>
                                                <chr>
                                                           <chr> <chr>
## 1 S2
             F1
                                    257 TGTGCCA~ TRBV12-4*00 CASSL~ CASSLAGS~
## 2 S2
            F1
                            0
                                    257 TGTGCCA~ TRBV12-4*00 CASSL~ CASSLGGA~
## 3 S2
             F1
                            0
                                    106 TGTGCCA~ TRBV12-4*00 CASSL~ CASSLAGS~
## 4 S2
             F1
                            0
                                    106 TGTGCCA~ TRBV12-4*00 CASSL~ CASSLGGA~
## 5 S2
             F1
                            0
                                     61 TGTGCCA~ TRBV3-1*00 CASSQ~ CASSSAGA~
                                     57 TGTGCCA~ TRBV12-4*00 CASSP~ CASSPGPT~
## 6 S2
             F1
                            0
## # ... with 2 more variables: dist <dbl>, epitope <chr>
Compute final table
system.time({data.ann <- get_1mm_annot(data, database)})</pre>
## Joining, by = "aa.seq.db"
## Joining, by = "aa.seq"
     user system elapsed
## 113.929
          3.620 117.692
glimpse(data.ann)
## Rows: 4,548,994
## Columns: 10
               <chr> "S2", "S2"~
## $ sample.id
               <chr> "F1", "F1"~
## $ replica
## $ time.point
              ## $ clone.count <dbl> 5160, 2317, 2154, 2073, 1583, 915, 845, 811, 673, 673, 666~
               <chr> "CGTGCCAGCAGCGCCCGGACTAGCGGGAGTAGGGACAATGAGCAGTTCTTC", "TG~
## $ nt.seq
## $ best.V.gene <chr> "TRBV7-3*00", "TRBV6-2*00", "TRBV27*00", "TRBV20-1*00", "T~
               <chr> "RASSARTSGSRDNEQFF", "CASSYRGTAWETQYF", "CASRPLLDRNNEQFF",~
## $ aa.seq
## $ aa.seq.db
               ## $ dist
               ## $ epitope
```