Immune repertoire forensics

A RepSeq data analysis tutorial

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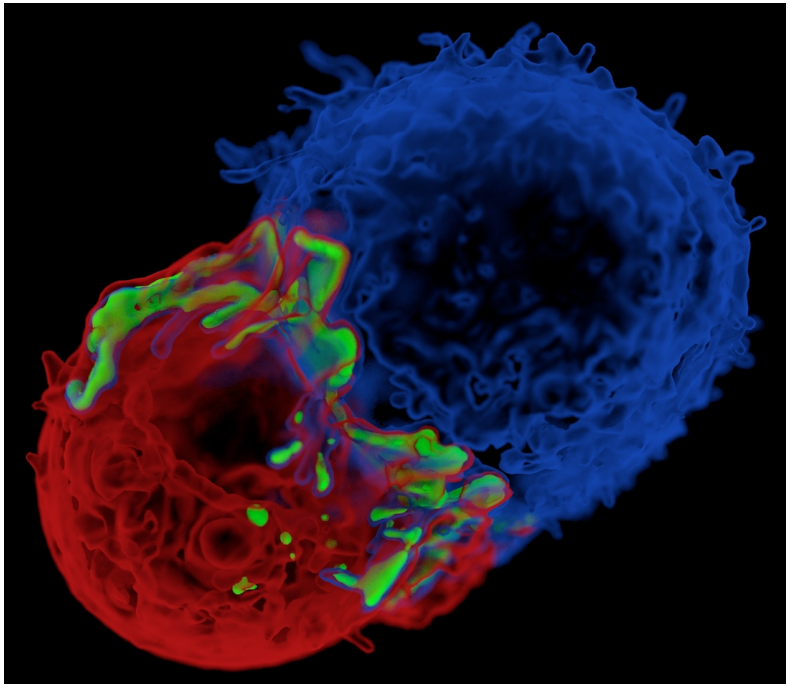
Skoltech, MA03172 course [Term 2, 2017-2018]

December 1, 2017

Outline

T-cell receptor

T-cell:APC contact



From James and Vale, Nature 2012, https://valelab.ucsf.edu/images/

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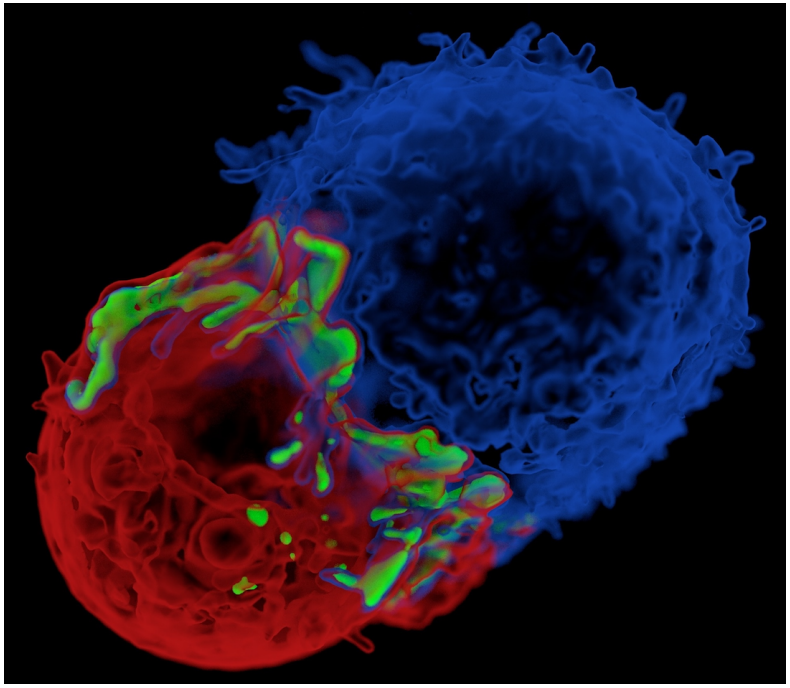
T-cell:APC contact



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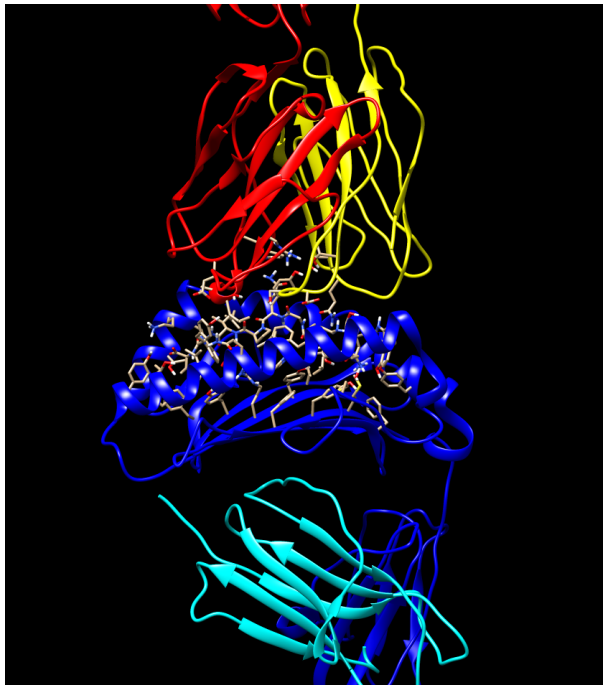
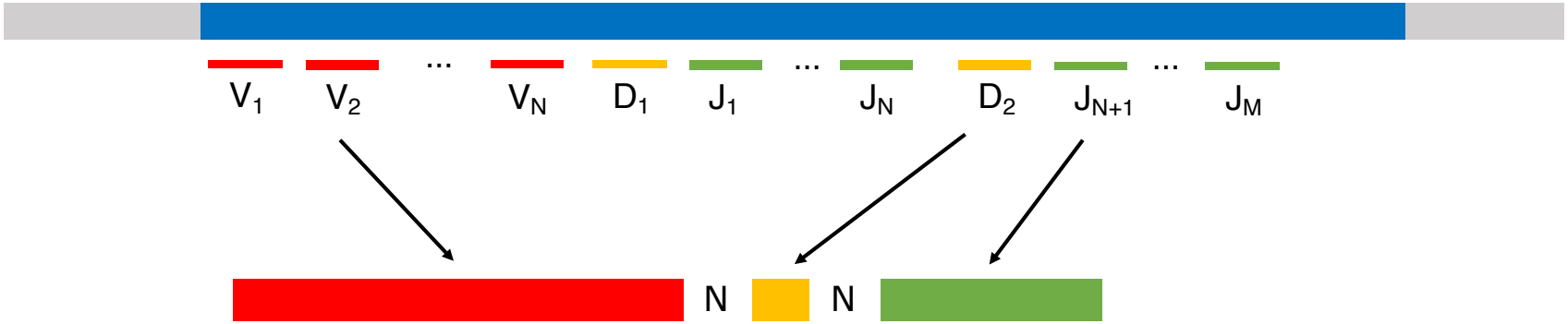
TCR:pMHC structure



PDB:1ao7, rendered using UCSF chimera, colored by chain

VDJ rearrangement

An example schema for TCRβ locus



Variable, Diversity and Joining are chosen at random, V-D and D-J junctions are filled with non-template N bases.

VDJ rearrangement

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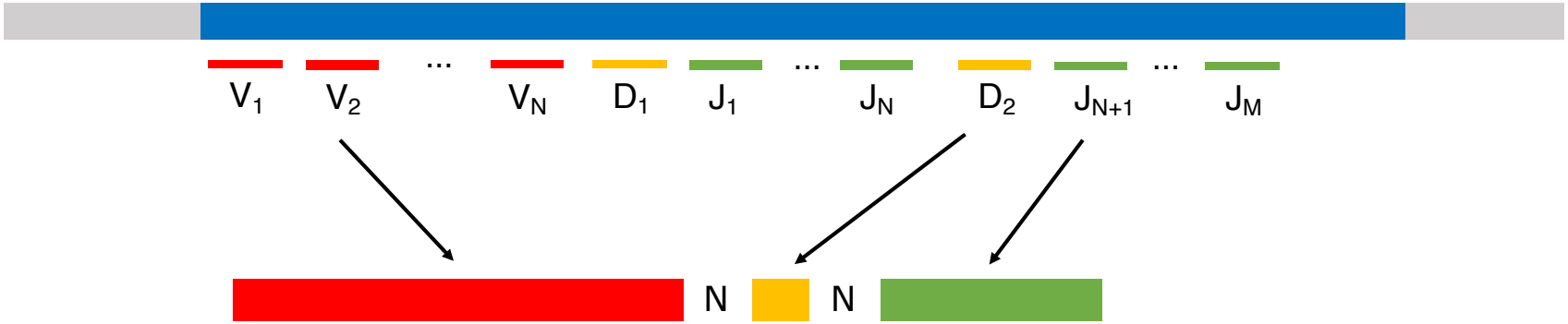

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VDJ rearrangement mechanism can be efficiently recaptured with a probabilistic model [Murugan et al. PNAS 2012]

$$
\begin{aligned}
P(\sigma) = {} & P(V)P(D,J) \\
& \times P(\#del_V|V)P(\#del_J|J)P(\#del_{D5},\#del_{D3}|D) \\
& \times P(\#ins_{VD})P(\#ins_{DJ}) \prod_{i \in ins_{VD}} P(b_i|b_{i-1}) \prod_{i \in ins_{DJ}} P(b_i|b_{i+1})
\end{aligned}
$$

TCR regions

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In total there are four framework (FRs) and three complementarity determining regions/loops (CDRs).

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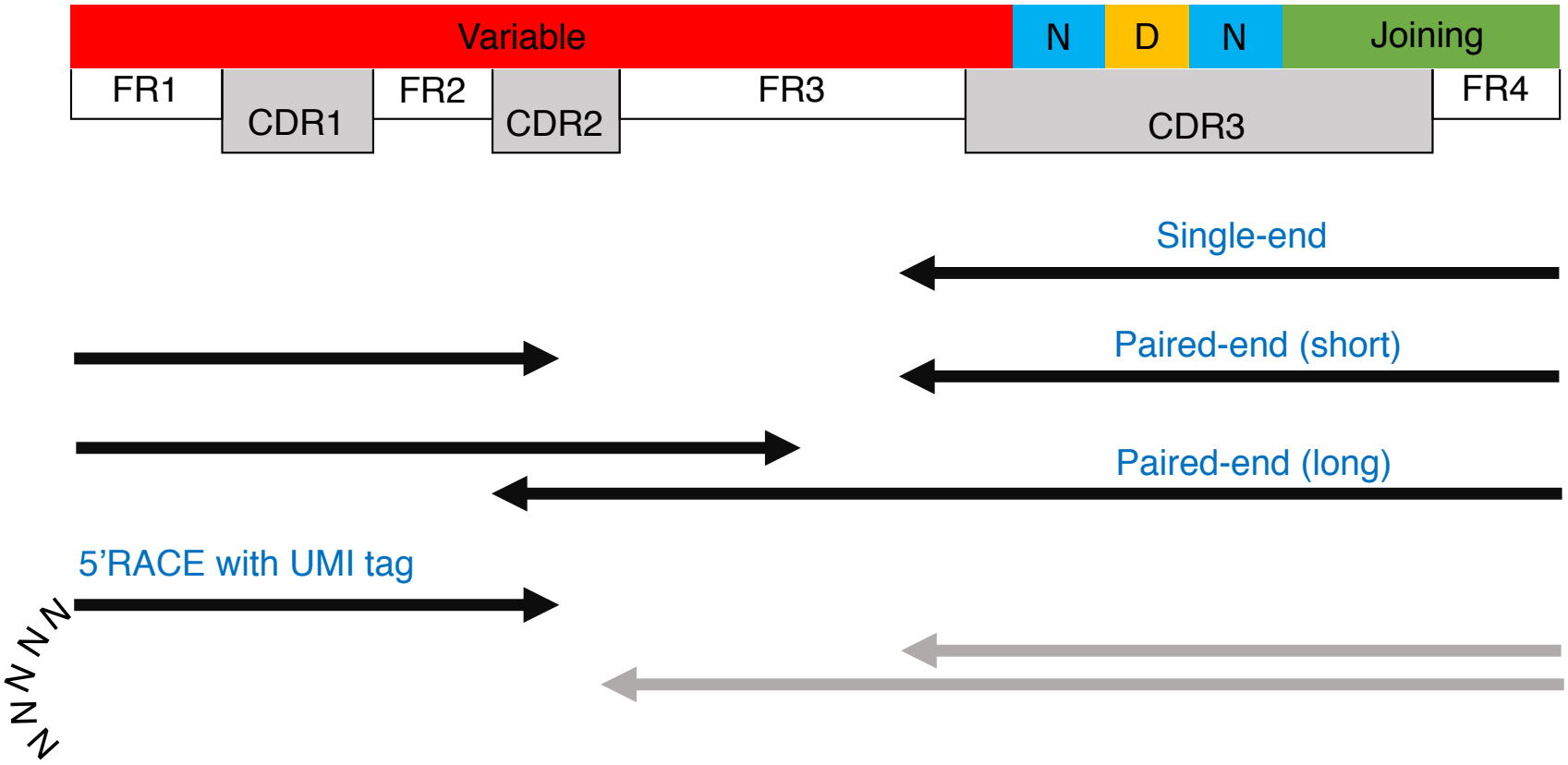
The likely functions of these regions are:

- FR regions maintain TCR secondary structure and (possibly) play role in MHC binding
- CDR1,2 are germline encoded and play role in antigen recognition, as well as (possibly) MHC binding
- CDR3 plays a major role in antigen recognition and is extremely variable

TCR repertoire sequencing



An example of a RepSeq dataset

After all pre-processing steps:

- Read grooming (filtering, etc)
- UMI-based assembly (for molecular barcoded data)
- V-D-J mapping and clonotype assembly

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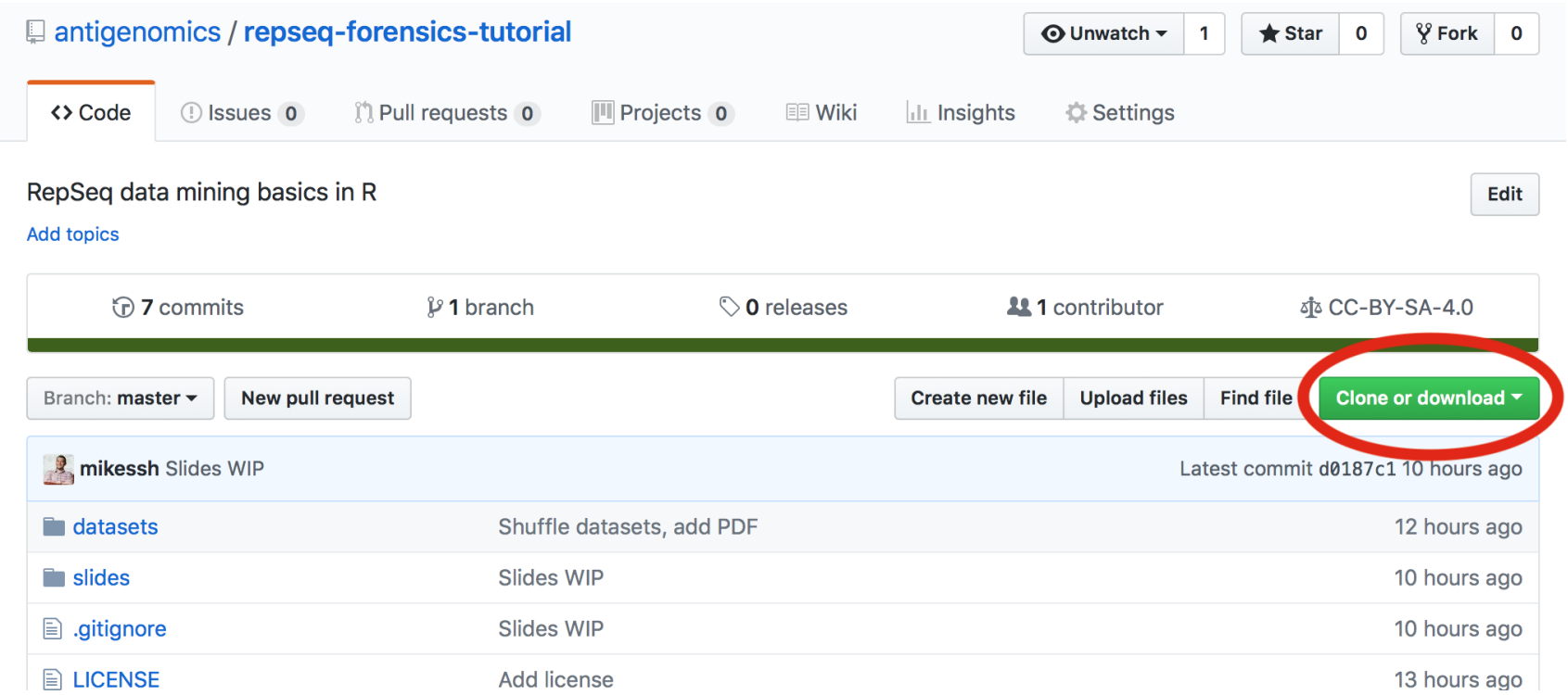
We finally get clonotype frequency tables that look like

Index	Frequency	Count	CDR3AA	V	D	J	CDR3NT
1	1.0%	3913	CSAGGLGSTDTQYF	TRBV20-1	TRBD1	TRBJ2-3	TGCAGTGCTGGGGGGCTCGGTAGCACAGATACGCAGTATTTT
2	0.90%	3440	CASNSGSSYNEQFF	TRBV5-1	TRBD2	TRBJ2-1	TGCGCCAGCAATAGCGGGAGCTCCTACAATGAGCAGTTCTTC
3	0.79%	3021	CSARQGNQPQHF	TRBV20-1	TRBD1	TRBJ1-5	TGCAGTGCGCGACAGGGGAATCAGCCCCAGCATTTT
4	0.65%	2490	CASSQEPGGEQFF	TRBV4-1	TRBD2	TRBJ2-1	TGCGCCAGCAGCCAAGAGCCGGGCGGGGAGCAGTTCTTC
5	0.61%	2336	CASSYGMNTEAFF	TRBV6-6	TRBD2	TRBJ1-1	TGTGCCAGCAGTTACGGGATGAACACTGAAGCTTTCTTT
6	0.52%	1992	CASSQGGRAPHTQYF	TRBV4-3	TRBD2	TRBJ2-3	TGCGCCAGCAGCCAAGGGGGGAGGGCCCCCCATACGCAGTATTTT
7	0.49%	1871	CASSQSQGGSYEQYF	TRBV5-1	TRBD1	TRBJ2-7	TGCGCCAGCAGCCAAAGTCAAGGGGGGTCCTACGAGCAGTACTTC
8	0.48%	1847	CASSRPKSGRSGELFF	TRBV11-2	TRBD2	TRBJ2-2	TGTGCCAGCAGCCGACCCAAGAGCGGGAGAAGTGGGGAGCTGTTTTTT

Outline

Downloading data

Navigate to `https://github.com/antigenomics/repseq-forensics-tutorial` and download the data + code bundle as zip



Executing R code

Open the `tutorial.Rmd` in RStudio, it can be found in the root folder of the bundle.

```
Plot diversity values

```{r message=FALSE}
diversity %>%
  melt %>%
  # set what values we are going to plot
  # fct_reorder reorders sample id by value
  ggplot(aes(x=fct_reorder2(sample_id, variable, value), y=value)) +
  # we'll make a bar plot
  geom_bar(stat = "identity", fill = "#0570b0") +
  # show each index on different subplot
  facet_wrap(~variable, scales = "free_y", ncol = 1) +
  xlab("") + ylab("Diversity index") +
  theme_bw()
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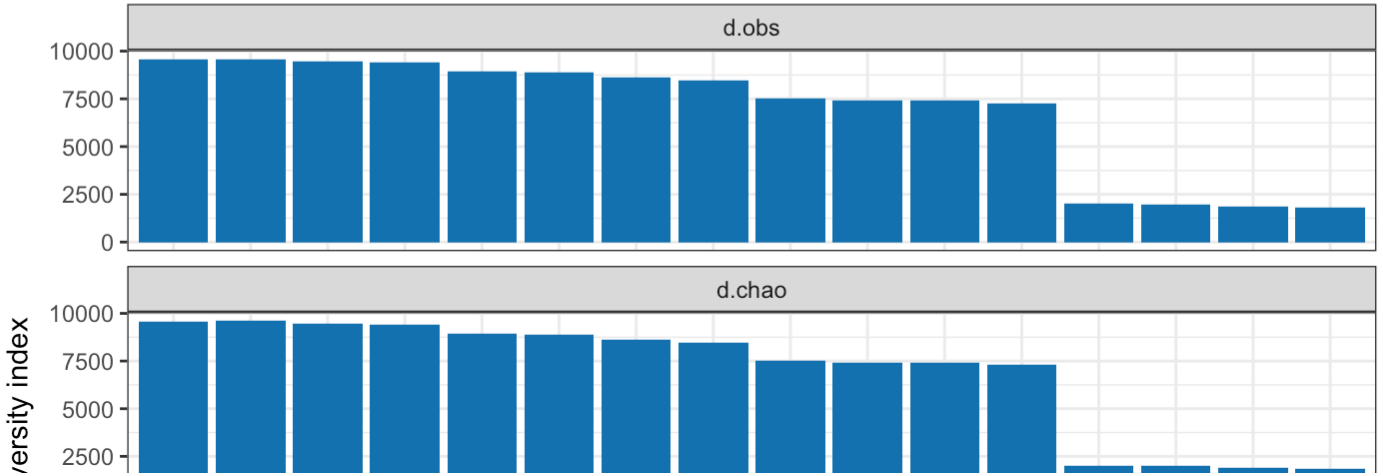

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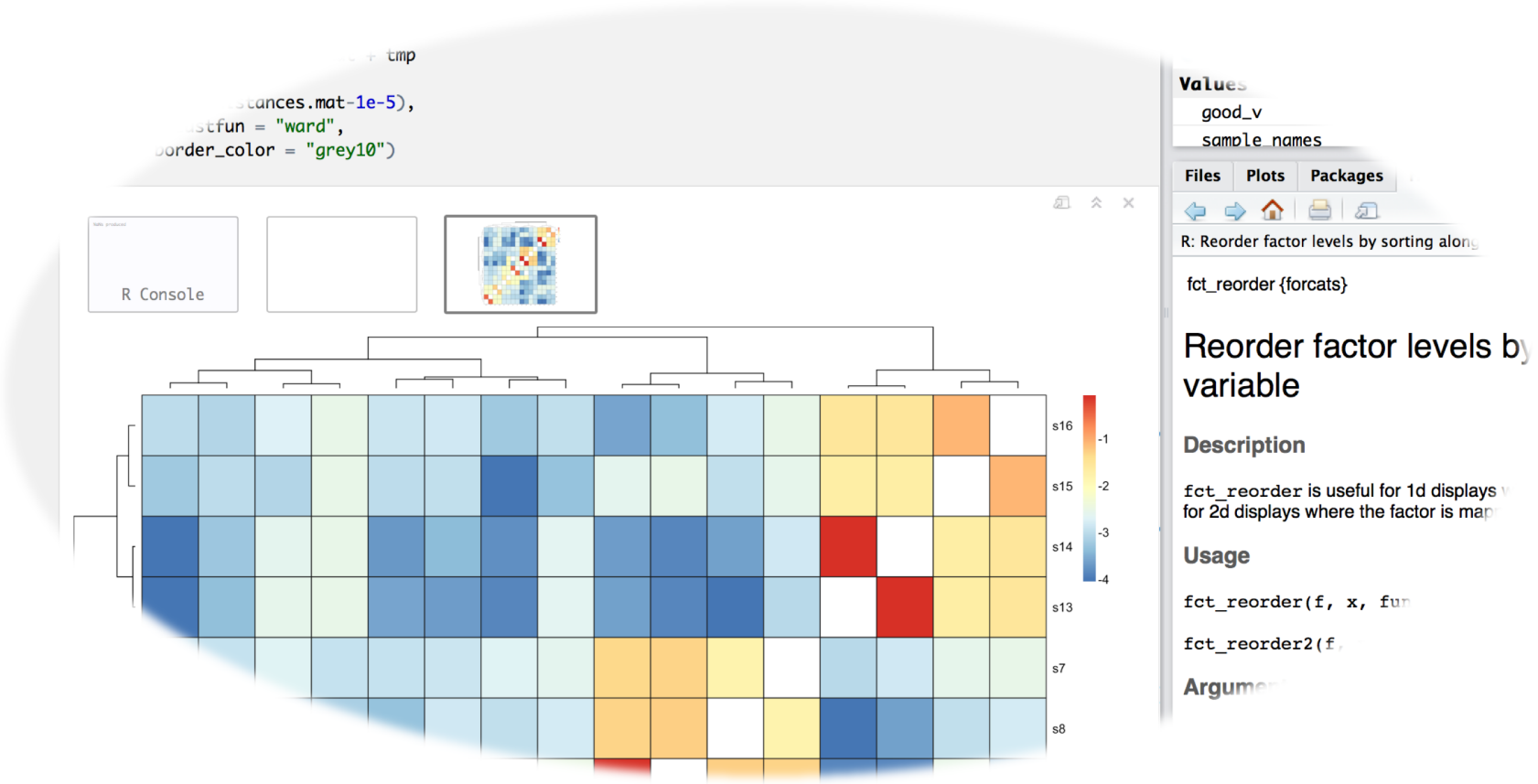
Interactive part



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