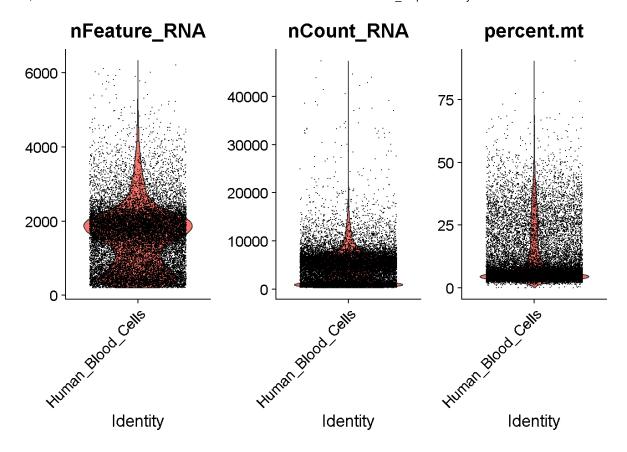
RNA_SequenceAnalysis

Antonio Pano 2022-11-22

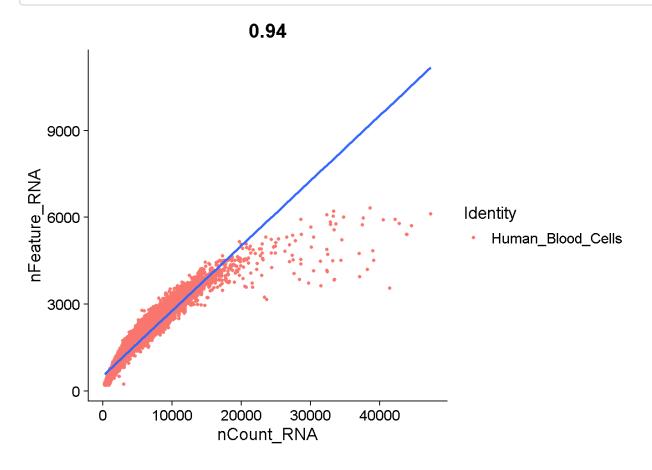
https://www.10xgenomics.com/resources/datasets/20-k-human-pbm-cs-5-ht-v-2-0-2-high-6-1-0 (https://www.10xgenomics.com/resources/datasets/20-k-human-pbm-cs-5-ht-v-2-0-2-high-6-1-0)

Quality Control

```
human_obj[["percent.mt"]] <- PercentageFeatureSet(human_obj, pattern = "^MT")
human_obj@meta.data %>% View
VlnPlot(human_obj, features = c("nFeature_RNA", "nCount_RNA", "percent.mt"), ncol = 3)
```



FeatureScatter(human_obj, feature1 = "nCount_RNA", feature2 = "nFeature_RNA") +
 geom_smooth(method = "lm")



Filtering

```
human_obj <- subset(human_obj, subset = nFeature_RNA > 400 & nFeature_RNA < 2650 & percent.mt < 7)
```

Normalization

```
human_obj <- NormalizeData(human_obj, normalization.method = "LogNormalize", scale.factor = 1000)</pre>
```

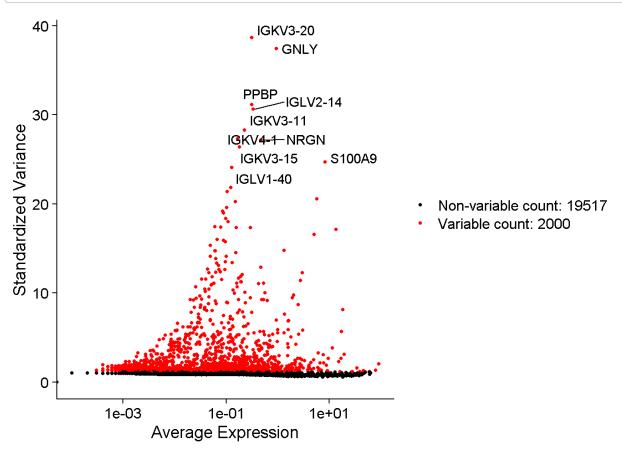
Identifying Highly Variable Features

```
human_obj <- FindVariableFeatures(human_obj, selection.method = "vst", nfeatures = 2000)</pre>
```

Top 10 most highly variable genes

```
top10 <- head(VariableFeatures(human_obj), 10)

plot1 <- VariableFeaturePlot(human_obj)
LabelPoints(plot = plot1, points = top10, repel = TRUE)</pre>
```



Scaling

 Scaling is done to make sure that cell clustering occurs because of natural causes and not axis "weight" similar to KNN.

```
all_genes <- rownames(human_obj)
human_obj <- ScaleData(human_obj, features = all_genes)</pre>
```

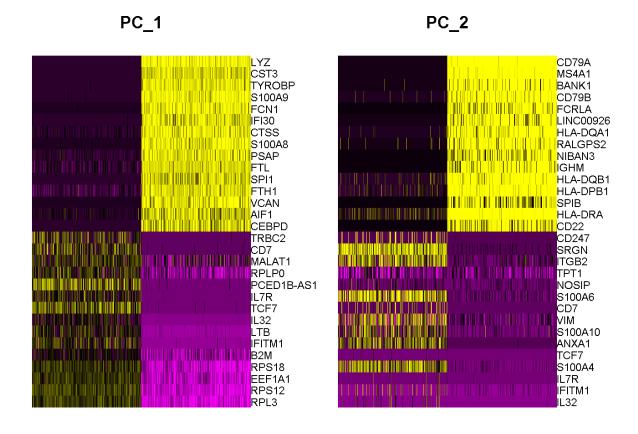
Dimensionality Reduction (Principal Component Analysis)

· RunPCA will throw an error if the data hasn't been scaled.

```
human_obj <- RunPCA(human_obj, features = VariableFeatures(object = human_obj))
# Visualizing the PCA plot
print(human_obj[["pca"]], dims = 1:5, nfeatures = TRUE)</pre>
```

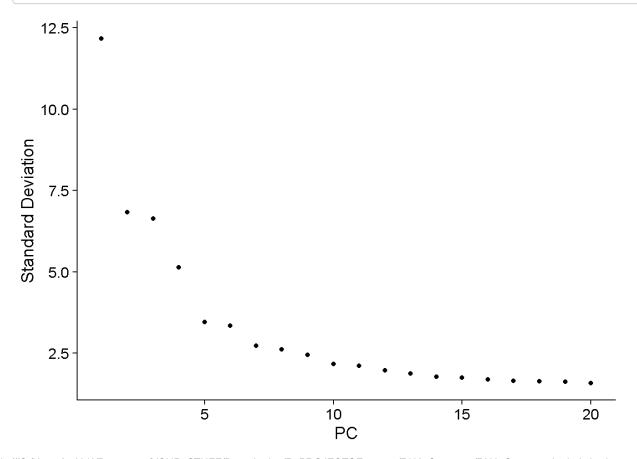
```
## PC_ 1
## Positive: RPL3
## Negative: LYZ
## PC_ 2
## Positive: IL32
## Negative: CD79A
## PC_ 3
## Positive: PPBP
## Negative: EEF1A1
## PC_ 4
## Positive: TPT1
## Negative: NKG7
## PC_ 5
## Positive: CRIP1
## Negative: CD7
```

```
# dims = "How many Principal Components?"
# cells = num of cells to plot
Seurat::DimHeatmap(human_obj, dims = 1:2, cells = 500, balanced = TRUE)
```



Determining the dimensionality of the data to use only the PC's that capture the majority of the signal in this downstream analysis. Better to use elbow on the higher side of the x-axis bc the effects could be larg e.

ElbowPlot(human obj)



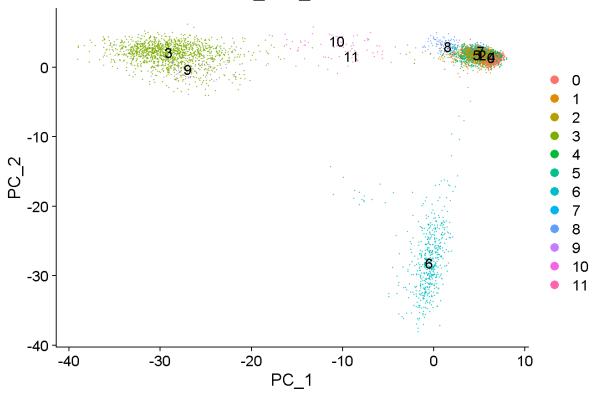
Clustering

```
human_obj <- FindNeighbors(human_obj, dims = 1:15)
human_obj <- FindClusters(human_obj, resolution = c(0, 0.3, 0.5, 0.7, 1))</pre>
```

```
## Modularity Optimizer version 1.3.0 by Ludo Waltman and Nees Jan van Eck
##
## Number of nodes: 9780
## Number of edges: 339344
## Running Louvain algorithm...
## Maximum modularity in 10 random starts: 1.0000
## Number of communities: 1
## Elapsed time: 2 seconds
## Modularity Optimizer version 1.3.0 by Ludo Waltman and Nees Jan van Eck
## Number of nodes: 9780
## Number of edges: 339344
## Running Louvain algorithm...
## Maximum modularity in 10 random starts: 0.9222
## Number of communities: 10
## Elapsed time: 2 seconds
## Modularity Optimizer version 1.3.0 by Ludo Waltman and Nees Jan van Eck
## Number of nodes: 9780
## Number of edges: 339344
##
## Running Louvain algorithm...
## Maximum modularity in 10 random starts: 0.8904
## Number of communities: 12
## Elapsed time: 3 seconds
## Modularity Optimizer version 1.3.0 by Ludo Waltman and Nees Jan van Eck
## Number of nodes: 9780
## Number of edges: 339344
##
## Running Louvain algorithm...
## Maximum modularity in 10 random starts: 0.8669
## Number of communities: 15
## Elapsed time: 3 seconds
## Modularity Optimizer version 1.3.0 by Ludo Waltman and Nees Jan van Eck
## Number of nodes: 9780
## Number of edges: 339344
## Running Louvain algorithm...
## Maximum modularity in 10 random starts: 0.8377
## Number of communities: 18
## Elapsed time: 3 seconds
```

```
View(human_obj@meta.data)
DimPlot(human_obj, group.by = "RNA_snn_res.0.5", label = TRUE)
```

RNA_snn_res.0.5



```
### Setting identity clusters

#Idents(human_obj)
Idents(human_obj) <- "RNA_snn_res.0.1"

#Idents(human_obj)</pre>
```

Non-Linear Dimensionality Reduction (UMAP)

```
library(reticulate)
py_install(packages = "umap-learn")
human_obj <- RunUMAP(human_obj, dims = 1:15)
DimPlot(human_obj, reduction = "umap")</pre>
```

