# ModulOmics Manual

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**ModulOmics** is a method to identify cancer driver pathways de novo by integrating multiple data types (protein-protein interactions, mutual exclusivity of mutations or copy number alterations, transcriptional co-regulation, and RNA co-expression) into a single probabilistic model. ModulOmics uses patient data, and protein-protein interaction networks, based here on the Hippie database [1] and regulatory connections, based here on the Transcriptional Regulatory Relationships Unraveled by Sentence-based Text mining database (TRRUST) [2].

The following manual describes the R code for ModulOmics – including the input and output data, and how driver modules can be easily inferred. Example files are based on Her2 breast cancer subtype from TCGA [3].

## **Dependencies**

library("TiMEx") library("igraph") library("cplexAPI") library("gtools")

The R code uploaded here contains all the functions needed for ModulOmics and a wrapper function named *run ModulOmics*.

### Libraries

**Code** containing callPipeSP, funcsTiMExPPI.R, ilpSolver.R and librariesTiMExPPI.R, and an inner library data downloaded here.

**Input** mutation and expression input files **output** for result files

#### Input

mutInput location of the binary genetic alterations file

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zScoreInput location of the zScored expression file

PPIInput location of the shortest path score for each pair of proteins, computed based on the PPI network

**TRRUSTInput** full path for the regulatory network

**pathCode** full path for the library containing funcsTiMExPPI.R, ilpSolver.R and librariesTiMExPPI.R

pathOutput full path for the output directory

**quantNo** quantile to consider active transcription factor, default value = 3 **topResults** number of modules to retreive from each cluster, default value = 5 **noClusters** number of clusters to start to stochastic search in order to locate global maximum, default value = 10

**SETS** number of initial seed modules retreived from the ILP, default value = 200 **LOWER\_LIMIT** lower K value (smallest modules), default value = 2, cannot drop below 2

**UPPER\_LIMIT** upper K value (largest modules), default value = 5, cannot exceed 6

## Output

**Return value** A matrix of the detected modules and their single and set scores. **Files** The function saves (K-1) files named "ModuleOmicsK.txt" to the output library, containing the sets and scores of each K.

# Code example

# format: column names are gene followed by either "-Mut" or "-CNA, rownames are patients. Data as downloaded from TCGA mutInput<-"<ful>full path>/input/Her2.txt"

# format: column names are genes, rownames are patients. Data as downloaded from TCGA

zScoreInput<-"<full path>/input/Her2\_zScore.txt" pathOutput<-"<full path>/output/" pathCode<-"<full path>/code/"

source(paste(pathCode,"callPipeSP.R",sep=""))

# The function reads cancer cohort and computes ModuleOmics scores results<-run\_ModulOmics(mutInput, zScoreInput, pathCode, pathOutput)

# References

- [1] Martin H. Schaefer, Jean-Fred F. Fontaine, Arunachalam Vinayagam, Pablo Porras, Erich E. Wanker, and Miguel A. Andrade-Navarro. HIPPIE: Integrating protein interaction networks with experiment based quality scores. PLoS ONE, 7(2), 2012.
- [2] Heonjong Han, Hongseok Shim, Donghyun Shin, Jung Eun Shim, Yunhee Ko, Junha Shin, Hanhae Kim, Ara Cho, Eiru Kim, Tak Lee, et al. Trrust: a reference database of human transcriptional regulatory interactions. Scientific Reports, 5:11432, 2015.

[3] Cancer Genome Atlas Network. Comprehensive molecular portraits of human breast tumours. Nature, 490(7418):61–70, 2012.