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NOAA/AOML Water Sampling Protocol using Sterivex with Zirconia Beads

1 PROTOCOL INFORMATION

1.1 Minimum Information about an Omics Protocol (MIOP)

- MIOP terms are listed in the YAML frontmatter of this page.
- See <https://github.com/BeBOP-OBON/miop/blob/main/model/schema/terms.yaml> for list and definitions.

1.2 Making eDNA FAIR (FAIRE)

- FAIRE terms are listed in the YAML frontmatter of this page.
- See <https://fair-edna.github.io/download.html> for the FAIRE checklist and more information.
- See <https://fair-edna.github.io/guidelines.html#missing-values> for guidelines on missing values that can be used for missing FAIRE or MIOP terms.

1.3 Authors

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1.4 Related Protocols

- This section contains protocols that should be known to users of this protocol.
- Internal Protocols: Derivative or altered protocols, or other protocols in this workflow.
- External Protocols: Protocols from manufacturers or other groups.
- Include the link to each protocol.
- Include the version number (internal) or access date (external) of the protocol when it was accessed.

1.4.1 Internal Protocols

PROTOCOL NAME	LINK	VERSION	RELEASE DATE
AOML 'omics Protocols	https://github.com/aomlomics/protocols		ongoing

1.4.2 External Protocols

PROTOCOL NAME	LINK	ISSUER / AUTHOR	ACCESS DATE
Not applicable			

1.5 Protocol Revision Record

- Version numbers start at 1.0.0 when the protocol is first completed and will increase when changes that impact the outcome of the procedure are made (patches: 1.0.1; minor changes: 1.1.0; major changes: 2.0.0).
- Release date is the date when a given protocol version was finalised.
- Description of revisions includes a brief description of what was changed relative to the previous version.

VERSION	RELEASE DATE	DESCRIPTION OF REVISIONS
1.0.0	2023-11-27	Initial release
1.0.1	2024-10-23	Formatting edits
1.1.0	2024-11-16	Addition of FAIR eDNA terms in YAML front matter
1.2.0	2025-01-08	Clarified the concentration of bleach throughout
1.2.1	2025-04-22	Corrected storage temperature
1.2.2	2025-12-15	Updated YAML front matter
1.2.3	2026-01-16	Updated YAML front matter

1.6 Acronyms and Abbreviations

ACRONYM / ABBREVIATION	DEFINITION
GO-SHIP	Global Ocean Ship-based Hydrographic Investigations Program

ACRONYM / ABBREVIATION	DEFINITION
NOAA	National Oceanic and Atmospheric Administration
AOML	Atlantic Oceanographic and Meteorological Laboratory
MSU	Mississippi State University
NGI	Northern Gulf Institute
eDNA	environmental DNA

1.7 Glossary

SPECIALISED TERM	DEFINITION
Field negative control	Negative control created during sampling. Usually distilled (DI) water run through a Sterivex filter in place of a seawater eDNA sample. This will act as a control for contamination during field sampling.
Niskin bottle	Plastic cylindrical bottle used for collecting water samples at different depths. Comes in a variety of volumes.

2 BACKGROUND

2.1 Summary

This protocol describes collection and filtration of marine environmental DNA samples using Sterivex cartridge filters and can be adapted to collect water samples from individual Niskin bottle, CTD rosette, or flow through systems. This protocol is used by NOAA's Atlantic Oceanographic and Meteorological Laboratory GO-SHIP collaborators.

2.2 Method description and rationale

This protocol is used to pump sea water samples collected via Niskin bottle, CTD rosette, or flow through systems and pump it through a 0.22 μm Sterivex filter using a peristaltic pump. Sterivex filters are loaded with Zymo ZR BashingBeads prior to pumping and DNA/RNA preservative is immediately added post-pumping to better preserve environmental DNA and expedite DNA extraction upon processing in the lab. The recommended filtration volume for GO-SHIP samples is ~8 liters, which takes ~2 hours. Precautions are taken to minimize contamination of samples by thoroughly sterilizing all equipment prior to use.

2.3 Spatial coverage and environment(s) of relevance

This protocol can be used across any marine environment to effectively collect water samples for biodiversity monitoring. This protocol can tolerate a wide range of depths for sampling - NOAA's AOML samples from 1m up to 1000m.

3 PERSONNEL REQUIRED

One person with pipetting experience. Research vessel experience is recommended but not required.

3.1 Safety

There are no major safety concerns with this protocol. Standard precautions should be taken such as wearing PPE at all times to avoid skin and eye exposure especially when working with bleach.

3.2 Training requirements

Standard molecular biology training including sterile technique and pipetting technique is required to properly conduct this protocol. Research vessel experience is recommended. Personnel should be trained in filtering protocol prior to conducting on ship.

3.3 Time needed to execute the procedure

The process of setting up sampling equipment and filtering seawater will take ~2 hours (120 minutes) depending on number of samples.

4 EQUIPMENT

DESCRIPTION	PRODUCT NAME AND MODEL	MANUFACTURER	QUANTITY	REMARK
Durable equipment				
20 L carboy	20 L Nalgene carboy	Generic brand	1	
8 L carboy	8 L Nalgene carboy	Generic brand	1	Used to hold bleach solution
Peristaltic pump	Masterflex peristaltic pump	Cole Parmer	1	
Pump heads	Masterflex L/S Easy-Load II Pump Heads for Precision Tubing	Avantor	3	The greater the # of pump heads, the faster the sampling process
Pump tubing	Masterflex Precision Pump Tubing, Peroxide-Cured Silicone	EW-96400-24	Cole Parmer	3
Hose-barb adapter	Masteflex fitting, Male Luer Lock to Hose Barb Adapter	Cole-Parmer	3	Depends on # of pump tubes
Serological pipette	10 mL Serological Pipette	Generic brand	3	Depends on # of pump tubes
2 L graduated cylinders	Graduated cylinder - 2 L	Generic brand	3	Depends on # of samples being pumped at once, can be substituted with carboys which tubing will be directly attached to (no serological pipets) using adapters
-20 °C freezer	-20 °C commercial chest freezer	Generic brand	1	
Consumable equipment				

DESCRIPTION	PRODUCT NAME AND MODEL	MANUFACTURER	QUANTITY	REMARK
Sterivex filter	Millipore Sterivex-GP Pressure Filter Unit, 0.22µm pore size	Millipore Sigma	175	Quantity depends on number of samples desired, remember to account for negative control field blanks
Inlet (male) luer-lock cap	MasterFlex Male Luer Lock Plug	VWR	175	Depends on # of samples, one per Sterivex
Outlet (female) luer-lock cap	MasterFlex Female Luer Thread Style Cap	VWR	175	Depends on # of samples, one per Sterivex
Pre-printed Cryo-Babies labels	Cryo-Babies LCRY-1700	Diversified Biotech	175	Depends on # of samples, one per Sterivex
60 mL syringe with male luer-lock outlet	Disposable syringe with luer lock - 60 mL	Generic brand	1	Used for sterilizing tubing
Sterile collection bags	Whil-Pak collection bags	Cole Parmer	1 box	Various sizes can be used for water collection
DNA/RNA Shield	Zymo DNA/RNA Shield	Zymo Research	175	(mL) 1mL required per Sterivex
Zymo ZR BashingBead Lysis Tubes	Zymo ZR BashingBead Lysis Tubes (0.1 & 0.5 mm)	Zymo Research	175	1 tube per Sterivex
Gloves	Powder-free nitrile gloves	Generic brand	1	(box) Can be any generic brand of gloves
Field notebook	Hard cover notebook	Generic brand	1	Encouraged to keep a digital sample log in addition to written notes
Chemicals				
5-9% Sodium hypochlorite	Household bleach	Generic brand	1	(bottle) Dilute 1:20 for lab use
Deionized or Milli-Q water	DI water	Generic brand	8	(L) Can use ship's DI water

- Description: E.g., “filter”.
- Product Name and Model: Provide the official name of the product.
- Manufacturer: Provide the name of the manufacturer of the product.
- Quantity: Provide quantities necessary for one application of the standard operating procedure (e.g., number of filters).
- Remark: For example, some of the consumables may need to be sterilized, some commercial solutions may need to be diluted or shielded from light during the operating procedure.

5 STANDARD OPERATING PROCEDURE

5.1 Sampling Protocol

5.1.1 Preparing Sterivex Filters

1. Wearing gloves, prepare the Sterivex filters by carefully adding 1 tube of Zymo ZR BashingBeads. It is recommended to cut off the top half of a 1000 uL tip and place it into the top of the Sterivex filter to act as a funnel while pouring beads into the Sterivex.
2. Label each Sterivex filter with a pre-printed sticker.

5.1.2 Preparing Sampling Equipment

1. Prepare a 1:20 dilution bleach solution by mixing 1 part household bleach (5-9% sodium hypochlorite) with DI water and storing in an 8 L carboy.
2. Attach hose-barb adapter to one end of peristaltic pump tubing.
3. Attach serological pipette to other end of peristaltic pump tubing.
4. Sterilize 60 mL syringe with 1:20 dilution bleach solution.
5. Rinse 20-L carboy 2 times with ~100 mL of sample water (200 mL total)
6. Wearing gloves, collect water from the flow-through system (or Niskin bottles) into 20-L carboy.
7. Place serological pipettes into bag(s) or bottle(s), ensuring the other end of the tubing (with hose-barb) is flowing into the graduated cylinders.
8. Turn on the pump and run ~100 mL of seawater to prime the tubing.
9. Run the pump at 100-150 rpm.
10. Pause the pump and discard water.

5.1.3 Sample Filtration

1. Attach the Sterivex filter input to the hose-barb that is already attached to the tubing outflow. The filter should screw on tightly. Avoid handling Sterivex filter input or output ends.
2. Run the pump and filter seawater until ~8 L has been filtered, measured using the graduated cylinders or another container.
3. On the log sheet, record the date, time, latitude, longitude, volume filtered and any notes about the sample. This information should be entered into an Excel spreadsheet every few days to maintain a digital copy in addition to the paper copy.
4. Pause pump and open pump head valves to release pressure.
5. Unscrew and remove Sterivex filters from the hose-barb adapters.
6. Gently remove any excess seawater from the filters using a sterilized 60 mL syringe.
7. Cap the bottom end (outlet) of the Sterivex filter with a female luer-lock cap.
8. In between filtering different water samples, rinse out the tubing with 1:20 dilution bleach solution, followed by DI water. Then proceed with the next volume of seawater.

5.1.4 Sample Preservation

1. Using a P1000 (1000 uL pipette), gently add 1000 uL of DNA/RNA Shield preservative into the Sterivex.
2. Cap the top end (inlet) of the Sterivex with a male luer-lock cap.

5.1.5 Storage

1. Freeze at -20 °C until extraction.

5.1.6 Post-Sampling

1. Return the bottle of DNA/RNA Shield to a refrigerator to reduce contamination.
2. Run ~1 L of 5% bleach through the lines to clean pump tubing. Repeat with ~1 L of deionized water.
3. Rinse the 20 L collection carboy 3 times with ~0.5 L DI water (1.5 L total) after sampling.

5.2 Quality control

Negative field controls are included with every research cruise. After the addition of Zymo ZR BashingBeads to a new Sterivex filter, DI water is filtered in place of sea water and then DNA/RNA Shield is added. These Sterivex are stored the same as other sample Sterivex.

5.3 Basic troubleshooting guide

Leaks

- If there is a leak present in the pump setup, you will notice trouble pulling water through the pump system. Check all seals and re-attach tubing.

Clogged Filter

- If a filter is clogged, turn valves and connections off and attempt to clear obstructions (i.e. large chunks of sediment or algae). Make note of any abnormal conditions and try to pump the full volume of seawater through the filter. In more productive areas, especially surface samples, we would frequently have filters that could not take the full volume. Since there are only two pumps and multiple sample depths at one time, it was common for us to use a cut-off time before starting the next sample (~45 min).

6 REFERENCES

Not applicable.

7 APPENDIX A: DATASHEETS

Not applicable.

8 APPENDIX B: VIDEO & IMAGE FILES

Filtering Video (Youtube): eDNA Sampling for Bio-GO-SHIP