

# R for Photobiology

*A handbook*

Pedro J. Aphalo,  
Andreas Albert  
and  
Titta Kotilainen

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## Preface

This is just a very early draft of a handbook that will accompany the release of the suite of R packages for photobiology (`r4photobiology`).

### Acknowledgements

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## List of abbreviations and symbols

For quantities and units used in photobiology we follow, as much as possible, the recommendations of the Commission Internationale de l'Éclairage as described by (Sliney 2007).

Symbol	Definition
$\alpha$	(%).
$\Delta e$	water vapour pressure difference (Pa).
$\epsilon$	emittance ( $\text{W m}^{-2}$ ).
$\lambda$	wavelength (nm).
$\theta$	solar zenith angle (degrees).
$\nu$	frequency (Hz or $\text{s}^{-1}$ ).
$\rho$	(%).
$\sigma$	Stefan-Boltzmann constant.
$\tau$	(%).
$\chi$	water vapour content in the air ( $\text{g m}^{-3}$ ).
$A$	(absorbance units).
ANCOVA	analysis of covariance.
ANOVA	analysis of variance.
BSWF	.
$c$	speed of light in a vacuum.
CCD	charge coupled device, a type of light detector.
CDOM	coloured dissolved organic matter.
CFC	chlorofluorocarbons.
c.i.	confidence interval.
CIE	Commission Internationale de l'Éclairage; or erythema action spectrum standardized by CIE.
CTC	closed-top chamber.
DAD	diode array detector, linear light detector based on photodiodes.
DBP	dibutylphthalate.
DC	direct current.
DIBP	diisobutylphthalate.
DNA(N)	UV action spectrum for 'naked' DNA.
DNA(P)	UV action spectrum for DNA in plants.
DOM	dissolved organic matter.
DU	Dobson units.
$e$	water vapour partial pressure (Pa).
$E$	(energy) irradiance ( $\text{W m}^{-2}$ ).
$E(\lambda)$	spectral (energy) irradiance ( $\text{W m}^{-2} \text{ nm}^{-1}$ ).

## LIST OF ABBREVIATIONS AND SYMBOLS

$E_0$	fluence rate, also called scalar irradiance ( $\text{W m}^{-2}$ ).
ESR	early stage researcher.
FACE	free air carbon-dioxide enhancement.
FEL	a certain type of 1000 W incandescent lamp.
FLAV	UV action spectrum for accumulation of flavonoids.
FWHM	full-width half-maximum.
GAW	Global Atmosphere Watch.
GEN	generalized plant action spectrum, also abbreviated as GPAS (Caldwell 1971).
GEN(G)	mathematical formulation of GEN by (Green et al. 1974) .
GEN(T)	mathematical formulation of GEN by (Thimijan et al. 1978).
$h$	Planck's constant.
$h'$	Planck's constant per mole of photons.
$H$	exposure, frequently called dose by biologists ( $\text{kJ m}^{-2} \text{d}^{-1}$ ).
$H^{\text{BE}}$	biologically effective (energy) exposure ( $\text{kJ m}^{-2} \text{d}^{-1}$ ).
$H_p^{\text{BE}}$	biologically effective photon exposure ( $\text{mol m}^{-2} \text{d}^{-1}$ ).
HPS	high pressure sodium, a type of discharge lamp.
HSD	honestly significant difference.
$k_B$	Boltzmann constant.
$L$	radiance ( $\text{W sr}^{-1} \text{m}^{-2}$ ).
LAI	leaf area index, the ratio of projected leaf area to the ground area.
LED	light emitting diode.
LME	linear mixed effects (type of statistical model).
LSD	least significant difference.
$n$	number of replicates (number of experimental units per treatment).
$N$	total number of experimental units in an experiment.
$N_A$	Avogadro constant (also called Avogadro's number).
NIST	National Institute of Standards and Technology (U.S.A.).
NLME	non-linear mixed effects (statistical model).
OTC	open-top chamber.
PAR	, 400–700 nm. measured as energy or photon irradiance.
PC	polycarbonate, a plastic.
PG	UV action spectrum for plant growth.
PHIN	UV action spectrum for photoinhibition of isolated chloroplasts.
PID	(control algorithm).
PMMA	polymethylmethacrylate.
PPFD	, another name for PAR photon irradiance ( $Q_{\text{PAR}}$ ).
PTFE	polytetrafluoroethylene.
PVC	polyvinylchloride.
$q$	energy in one photon ('energy of light').
$q'$	energy in one mole of photons.
$Q$	photon irradiance ( $\text{mol m}^{-2} \text{s}^{-1}$ or $\mu\text{mol m}^{-2} \text{s}^{-1}$ ).
$Q(\lambda)$	spectral photon irradiance ( $\text{mol m}^{-2} \text{s}^{-1} \text{nm}^{-1}$ or $\mu\text{mol m}^{-2} \text{s}^{-1} \text{nm}^{-1}$ ).
$r_0$	distance from sun to earth.
RAF	(nondimensional).
RH	relative humidity (%).
$s$	energy effectiveness (relative units).



$s(\lambda)$	spectral energy effectiveness (relative units).
$s^p$	quantum effectiveness (relative units).
$s^p(\lambda)$	spectral quantum effectiveness (relative units).
s.d.	standard deviation.
SDK	software development kit.
s.e.	standard error of the mean.
SR	spectroradiometer.
$t$	time.
$T$	temperature.
TUV	tropospheric UV.
$U$	electric potential difference or voltage (e.g. sensor output in V).
UV	ultraviolet radiation ( $\lambda = 100\text{--}400\text{ nm}$ ).
UV-A	ultraviolet-A radiation ( $\lambda = 315\text{--}400\text{ nm}$ ).
UV-B	ultraviolet-B radiation ( $\lambda = 280\text{--}315\text{ nm}$ ).
UV-C	ultraviolet-C radiation ( $\lambda = 100\text{--}280\text{ nm}$ ).
UV <sup>BE</sup>	biologically effective UV radiation.
UTC	coordinated universal time, replaces GMT in technical use.
VIS	radiation visible to the human eye ( $\approx 400\text{--}700\text{ nm}$ ).
WMO	World Meteorological Organization.
VPD	water vapour pressure deficit (Pa).
WOUDC	World Ozone and Ultraviolet Radiation Data Centre.



# **Part I**

## **Preliminaries**



## **Part II**

# **Cookbook of calculations**



## Simple summaries and spectral features

### Abstract

In this chapter we explain how to obtain different summaries common to all types of spectral data. In addition we describe how to extract spectral features from spectral data.

### 1.1 Packages used in this chapter

For executing the examples listed in this chapter you need first to load the following packages from the library:

```
library(photobiology)

## Loading required package: data.table

library(photobiologygg)

## Loading required package: photobiologyWavebands
## Loading required package: proto
## Loading required package: ggplot2
## Loading required package: methods
## Loading required package: scales

library(photobiologyLamps)
```

### 1.2 Task: Summaries related to wavelength

Functions `max`, `min`, `range`, `midpoint` when used with an object of class `generic_spct` (or a derived class) return the result of applying these functions to the `w.length` component of these objects, returning always values expressed in nanometres as long as the objects have been correctly created.

```

range(sun.spct)

## [1] 293 800

midpoint(sun.spct)

## [1] 546.5

max(sun.spct)

## [1] 800

min(sun.spct)

## [1] 293

```

Functions `spread` and `stepsize` are generics defined in package `photobiology`. `spread` returns maximum less minimum wavelengths values in nanometres, while `stepsize` returns a numeric vector of length two with the maximum and the minimum wavelength step between observations, also in nanometers.

```

spread(sun.spct)

## [1] 507

stepsize(sun.spct)

## [1] 1 1

```

In the case of the `summary` method, specializations for `source_spct` and `...` are provided. But for other spectral objects, the `summary` method for `data.table` is called. For the `summary` specializations defined, the corresponding `print` method specializations are also defined.

```

summary(sun.spct)

## wavelength ranges from 293 to 800 nm
## largest wavelength step size is 1 nm
## spectral irradiance ranges from 2.61e-06 to 0.8205 W m-2 nm-1
## energy irradiance is 269.1 W m-2
## photon irradiance is 1255 umol s-1 m-2

```

## 1.3 Task: Find peaks and valleys

### 1.3.1 Obtaining the location of peaks as an index into the spectral data

Function `find_peaks`, takes as argument a numeric vector, and returns a logical vector of the same length, with `TRUE` for local maxima and `FALSE` for all other observations. Infinite values are discarded.



### 1.3. TASK: FIND PEAKS AND VALLEYS

```
find_peaks(sun.spct$s.e.irrad)

## [1] FALSE FALSE FALSE FALSE FALSE FALSE FALSE
## [8] FALSE FALSE FALSE FALSE FALSE FALSE FALSE
## [15] FALSE FALSE FALSE FALSE FALSE FALSE FALSE
## [22] FALSE TRUE FALSE TRUE FALSE FALSE FALSE
## [29] TRUE FALSE FALSE FALSE FALSE FALSE TRUE
## [36] FALSE FALSE TRUE FALSE TRUE FALSE FALSE
## [43] TRUE FALSE FALSE FALSE FALSE TRUE FALSE
## [50] FALSE TRUE FALSE TRUE FALSE TRUE FALSE
## [57] FALSE TRUE FALSE FALSE FALSE TRUE FALSE
## [64] FALSE FALSE FALSE FALSE TRUE FALSE FALSE
## [71] FALSE TRUE FALSE TRUE FALSE FALSE FALSE
## [78] TRUE FALSE FALSE FALSE FALSE FALSE FALSE
## [85] FALSE TRUE FALSE FALSE TRUE FALSE FALSE
## [92] FALSE TRUE FALSE TRUE FALSE FALSE FALSE
## [99] TRUE FALSE FALSE FALSE TRUE FALSE FALSE
## [106] FALSE FALSE FALSE FALSE TRUE FALSE TRUE
## [113] FALSE FALSE FALSE FALSE TRUE FALSE FALSE
## [120] TRUE FALSE TRUE FALSE TRUE FALSE FALSE
## [127] TRUE FALSE TRUE FALSE FALSE FALSE FALSE
## [134] TRUE FALSE FALSE FALSE FALSE FALSE FALSE
## [141] TRUE FALSE FALSE TRUE FALSE FALSE FALSE
## [148] FALSE FALSE TRUE FALSE TRUE FALSE FALSE
## [155] FALSE TRUE FALSE FALSE TRUE FALSE FALSE
## [162] FALSE FALSE FALSE TRUE FALSE FALSE FALSE
## [169] TRUE FALSE FALSE TRUE FALSE TRUE FALSE
## [176] TRUE FALSE FALSE FALSE TRUE FALSE FALSE
## [183] TRUE FALSE FALSE TRUE FALSE TRUE FALSE
## [190] TRUE FALSE FALSE FALSE FALSE FALSE TRUE
## [197] FALSE TRUE FALSE FALSE TRUE FALSE TRUE
## [204] FALSE FALSE FALSE TRUE FALSE FALSE FALSE
## [211] TRUE FALSE FALSE TRUE FALSE FALSE TRUE
## [218] FALSE FALSE TRUE FALSE FALSE FALSE TRUE
## [225] FALSE TRUE FALSE FALSE FALSE TRUE FALSE
## [232] FALSE TRUE FALSE FALSE FALSE TRUE FALSE
## [239] TRUE FALSE FALSE FALSE FALSE TRUE FALSE
## [246] TRUE FALSE FALSE FALSE FALSE FALSE FALSE
## [253] TRUE FALSE TRUE FALSE FALSE TRUE FALSE
## [260] TRUE FALSE FALSE TRUE FALSE FALSE TRUE
## [267] FALSE FALSE TRUE FALSE TRUE FALSE TRUE
## [274] FALSE FALSE TRUE FALSE TRUE FALSE FALSE
## [281] TRUE FALSE FALSE FALSE TRUE FALSE FALSE
## [288] FALSE FALSE TRUE FALSE FALSE FALSE FALSE
## [295] TRUE FALSE FALSE FALSE TRUE FALSE FALSE
## [302] TRUE FALSE FALSE TRUE FALSE FALSE TRUE
## [309] FALSE FALSE FALSE FALSE TRUE FALSE FALSE
## [316] TRUE FALSE FALSE FALSE TRUE FALSE FALSE
## [323] TRUE FALSE FALSE FALSE TRUE FALSE TRUE
## [330] FALSE FALSE FALSE FALSE FALSE TRUE FALSE
## [337] TRUE FALSE FALSE FALSE TRUE FALSE TRUE
## [344] FALSE FALSE TRUE FALSE FALSE FALSE TRUE
## [351] FALSE FALSE FALSE FALSE FALSE FALSE FALSE
## [358] FALSE TRUE FALSE TRUE FALSE FALSE FALSE
## [365] FALSE FALSE FALSE FALSE FALSE TRUE FALSE
## [372] FALSE FALSE FALSE FALSE TRUE FALSE FALSE
## [379] FALSE FALSE FALSE TRUE FALSE FALSE FALSE
## [386] FALSE FALSE FALSE FALSE FALSE FALSE FALSE
## [393] FALSE FALSE FALSE TRUE FALSE FALSE FALSE
## [400] TRUE FALSE FALSE FALSE FALSE TRUE FALSE
```

```
## [407] FALSE FALSE FALSE FALSE FALSE TRUE FALSE
## [414] FALSE FALSE FALSE TRUE FALSE FALSE TRUE
## [421] FALSE FALSE FALSE FALSE FALSE FALSE FALSE
## [428] FALSE FALSE TRUE FALSE FALSE FALSE FALSE
## [435] TRUE FALSE FALSE TRUE FALSE FALSE FALSE
## [442] TRUE FALSE FALSE FALSE FALSE FALSE FALSE
## [449] TRUE FALSE FALSE TRUE FALSE FALSE TRUE
## [456] FALSE FALSE FALSE TRUE FALSE FALSE TRUE
## [463] FALSE FALSE FALSE FALSE FALSE FALSE FALSE
## [470] TRUE FALSE FALSE FALSE FALSE FALSE FALSE
## [477] FALSE FALSE TRUE FALSE FALSE TRUE FALSE
## [484] FALSE TRUE FALSE TRUE FALSE FALSE TRUE
## [491] FALSE FALSE TRUE FALSE FALSE FALSE FALSE
## [498] FALSE FALSE TRUE FALSE TRUE FALSE FALSE
## [505] FALSE TRUE FALSE FALSE
```

To obtain the indexes, one can use R's function which

```
which(find_peaks(sun.spct$s.e.irrad))

## [1] 23 25 29 35 38 40 43 48 51 53 55
## [12] 58 62 68 72 74 78 86 89 93 95 99
## [23] 103 110 112 117 120 122 124 127 129 134 141
## [34] 144 150 152 156 159 165 169 172 174 176 180
## [45] 183 186 188 190 196 198 201 203 207 211 214
## [56] 217 220 224 226 230 233 237 239 244 246 253
## [67] 255 258 260 263 266 269 271 273 276 278 281
## [78] 285 290 295 299 302 305 308 313 316 320 323
## [89] 327 329 335 337 341 343 346 350 359 361 370
## [100] 376 382 396 400 405 412 417 420 430 435 438
## [111] 442 449 452 455 459 462 470 479 482 485 487
## [122] 490 493 500 502 506
```

### 1.3.2 Obtaining the location of peaks as a wavelength in nanometres

Function `get_peaks` takes two numeric vectors as arguments, `x` is, for spectra assumed to be a vector of wavelengths, and `y` the spectral variable to search for local maxima.

```
with(sun.spct, get_peaks(w.length, s.e.irrad))

##      x      y label
## 1 321 0.1822031 321
## 2 330 0.3295190 330
## 3 335 0.3129253 335
## 4 340 0.3352353 340
## 5 343 0.3380052 343
## 6 347 0.3207918 347
## 7 350 0.3453572 350
## 8 354 0.3758625 354
## 9 360 0.3707068 360
## 10 366 0.4491898 366
## 11 370 0.4393233 370
## 12 378 0.4969714 378
## 13 381 0.4362110 381
## 14 385 0.3915446 385
## 15 391 0.4822105 391
```

### 1.3. TASK: FIND PEAKS AND VALLEYS

```
## 16 395 0.4699886 395
## 17 402 0.6497388 402
## 18 409 0.6615421 409
## 19 412 0.6742498 412
## 20 416 0.6761818 416
## 21 421 0.6701269 421
## 22 426 0.6388873 426
## 23 436 0.7336607 436
## 24 442 0.7188581 442
## 25 451 0.8204633 451
## 26 457 0.7984935 457
## 27 461 0.7711277 461
## 28 468 0.7665312 468
## 29 472 0.7693357 472
## 30 475 0.7724634 475
## 31 478 0.7869773 478
## 32 482 0.7832759 482
## 33 490 0.7728111 490
## 34 495 0.7899872 495
## 35 506 0.7701737 506
## 36 509 0.7466557 509
## 37 512 0.7510876 512
## 38 516 0.7302733 516
## 39 525 0.7376088 525
## 40 531 0.7603297 531
## 41 536 0.7429248 536
## 42 545 0.7272464 545
## 43 552 0.7178863 552
## 44 555 0.7117599 555
## 45 563 0.6991590 563
## 46 568 0.6750915 568
## 47 573 0.6713390 573
## 48 577 0.6644287 577
## 49 582 0.6853736 582
## 50 597 0.6256272 597
## 51 600 0.6372767 600
## 52 605 0.6614323 605
## 53 612 0.6468436 612
## 54 621 0.6464099 621
## 55 638 0.6233510 638
## 56 642 0.6138918 642
## 57 653 0.5819163 653
## 58 662 0.5995383 662
## 59 668 0.5977089 668
## 60 674 0.5879885 674
## 61 692 0.5029201 692
## 62 697 0.5164799 697
## 63 704 0.5013394 704
## 64 712 0.5070675 712
## 65 722 0.4345295 722
## 66 727 0.3855997 727
## 67 734 0.4676589 734
## 68 744 0.5006212 744
## 69 747 0.5025733 747
## 70 751 0.5000141 751
## 71 754 0.5007593 754
## 72 774 0.4746771 774
## 73 777 0.4716414 777
## 74 782 0.4680026 782
```

```
## 75 794 0.4213304 794
## 76 798 0.4236281 798
```

The returned value is a (shorter) data frame with two numeric vectors,  $x$  and  $y$ , and an optional character variable `label`, for each local maximum found in  $y$ .

Methods for spectral objects will be added.

## 1.4 Task: Refining the location of peaks and valleys

The functions described in the previous section locate the observation with the locally highest  $y$ -value. This is in most cases the true location of the peaks as they may fall in between two observations along the wavelength axis. By fitting a suitable model to describe the shape of the peak, which is the result of the true peak and the slit function of the spectrometer, the true location of a peak can be approximated more precisely. There is no universally useful model, so we show some examples of a possible method of peak-position refinement.

In this example, in the second statement we refine the location of the shortest-wavelength peak found by `get_peaks` in the first statement. For this approach to work, the peaks should be clearly visible, and not very close to each other. We use the spectral irradiance measured from a UV-B lamp as an example.

```
stepsize(philips.tl01.bentham.spct)

## [1] 0.5 0.5

peaks <- with(philips.tl01.bentham.spct, get_peaks(w.length, s.e.irrad, span = 51))
fit <- nls(s.e.irrad ~ d + a1*exp(-0.5*((w.length-c1)/b1)^2),
          start=list(a1=6, b1=1, c1=peaks[1, 1], d=0), data=philips.tl01.bentham.spct)
fit

## Nonlinear regression model
## model: s.e.irrad ~ d + a1 * exp(-0.5 * ((w.length - c1)/b1)^2)
## data: philips.tl01.bentham.spct
##      a1      b1      c1      d
## 6.08259 0.90542 311.64836 0.03093
## residual sum-of-squares: 7.684
##
## Number of iterations to convergence: 8
## Achieved convergence tolerance: 3.682e-06

fit$m$getPars()[["c1"]]

## [1] 311.6484

peaks[1, 1]

## [1] 311.5
```

### 1.5. TASK:

Because in this example the spectral resolution of the data is high, the improvement is small. We now subsample the spectral data, and repeat the calculations.

```
my.tl01.spct <- subset(philips.tl01.bentham.spct, w.length %% 2 == 0)
stepsize(my.tl01.spct)

## [1] 2 2

my.peaks <- with(my.tl01.spct, get_peaks(w.length, s.e.irrad, span = 51))
my.fit <- nls(s.e.irrad ~ d + a1*exp(-0.5*((w.length-c1)/b1)^2),
             start=list(a1=6, b1=1, c1=my.peaks[1, 1], d=0), data=my.tl01.spct)
my.fit

## Nonlinear regression model
##   model: s.e.irrad ~ d + a1 * exp(-0.5 * ((w.length - c1)/b1)^2)
##   data: my.tl01.spct
##      a1      b1      c1      d
## 4.36981 0.91409 311.85914 0.03175
## residual sum-of-squares: 0.1554
##
## Number of iterations to convergence: 4
## Achieved convergence tolerance: 1.956e-07

my.fit$m$getPars()[["c1"]]

## [1] 311.8591

my.peaks[1, 1]

## [1] 312
```

### 1.5 Task:



## **Part III**

# **Catalogue of data sources**





## **Part IV**

# **Data acquisition and modelling**



## CHAPTER 2

### Further reading about R

- 2.1 Introductory texts
- 2.2 Texts on specific aspects
- 2.3 Advanced texts
- 2.4 Application-specific texts



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## Build information

### **Sys.info()**

```
##               sysname
##             "Windows"
##             release
##             "7 x64"
##             version
## "build 7601, Service Pack 1"
##             nodename
##             "MUSTI"
##             machine
##             "x86-64"
##             login
##             "aphalo"
##             user
##             "aphalo"
##             effective_user
##             "aphalo"
```

### **sessionInfo()**

```
## R version 3.2.0 (2015-04-16)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 7 x64 (build 7601) Service Pack 1
##
## locale:
## [1] LC_COLLATE=English_United Kingdom.1252
## [2] LC_CTYPE=English_United Kingdom.1252
## [3] LC_MONETARY=English_United Kingdom.1252
## [4] LC_NUMERIC=C
## [5] LC_TIME=English_United Kingdom.1252
##
## attached base packages:
## [1] methods    tools      stats      graphics
```

## APPENDIX A. BUILD INFORMATION

```
## [5] grDevices utils      datasets base
##
## other attached packages:
## [1] photobiologyLamps_0.3.0
## [2] photobiologygg_0.3.3
## [3] scales_0.2.4
## [4] ggplot2_1.0.1
## [5] proto_0.3-10
## [6] photobiologyWavebands_0.3.1.9000
## [7] photobiology_0.6.7.90000
## [8] data.table_1.9.4
## [9] stringr_1.0.0
## [10] knitr_1.10.5
##
## loaded via a namespace (and not attached):
## [1] Rcpp_0.11.6      magrittr_1.5
## [3] MASS_7.3-40      munsell_0.4.2
## [5] colorspace_1.2-6 splus2R_1.2-0
## [7] highr_0.5        plyr_1.8.2
## [9] caTools_1.17.1   grid_3.2.0
## [11] gtable_0.1.2     digest_0.6.8
## [13] reshape2_1.4.1   formatR_1.2
## [15] bitops_1.0-6     memoise_0.2.1
## [17] evaluate_0.7     stringi_0.4-1
## [19] lubridate_1.3.3  chron_2.3-45
```