# EE 542 – Laboratory Assignment

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The git repo for all the codes provided in this lab: https://github.com/yuesOctober/GDCproject/tree/yue

### Download the repo:

git clone https://github.com/yuesOctober/GDCproject.git

<u>Basic git command:</u> <a href="https://confluence.atlassian.com/bitbucketserver/basic-git-commands-776639767.html">https://confluence.atlassian.com/bitbucketserver/basic-git-commands-776639767.html</a>

## GDC Data Lab:

In this lab, you will learn:

- 1. How to download, integrate, and preprocess files related to a particular disease type, and how to use the data obtained.
- 2. As an example, you will go through the entire process to get the miRNA files, and the related file metadata, case metadata to the disease Liver Hepatocellular Carcinoma
- 3. You will apply the machine learning package to the miRNA matrix extracted to detect normal/cancer samples.

Steps you need to do with this lab:

- 1. Go through the entire tutorial and do the Part 1 and Part 2 with miRNA expression data for all cancer types and do a multiclass classification. You can treat all the samples from normal tissues from different cancer types as one single class.
- 2. In Part2, try a different model other than the one provided in the sample code.
- 3. Visualize the features after feature selection with t-SNE and PCA method [3]
- 4. Plot the evaluation metrics and ROC curve for the model.

## Submission guideline:

Each team should create a github repo and provide the link to your repo for in your slide. You need have a readme file explaining how to run your source codes. For video demo submission, you need show the steps to run your code and explain. Only one submission per team is needed.

### Notes:

In this lab, the miRNA expression quantification data is used, however in your final project, you could use different types or combination of different types of biomarkers, e.g. somatic mutation, copy number variation, DNA methylation etc for cancer type idenfication.

Part 1: Data download, integration and preprocess.

## 1. Introduction to GDC data:

Read the document below to get a sense of GDC data.

https://gdc.cancer.gov/about-data

Biomarker Data:

Data Category	Data Type
DNA Methylation	Methylation Beta Value
Simple Nucleotide	Annotated Somatic Mutation
Variation	Raw Simple Somatic Mutation
	Aggregated Somatic Mutation
	Masked Somatic Mutation
Transcriptome Profile	Gene Expression Quantification
	Isoform Expression Quanfitication
	miRNA expression Quantification

# 2. Example: Downloading miRNA files of Disease: Liver Hepatocellular Carcinoma

miRNA Expression Quantification is a table that associates miRNA IDs with read count and a normalized count in reads-per-million-miRNA-mapped. <a href="https://docs.gdc.cancer.gov/Data/Bioinformatics-Pipelines/miRNA-Pipeline/">https://docs.gdc.cancer.gov/Data/Bioinformatics-Pipelines/miRNA-Pipeline/</a>

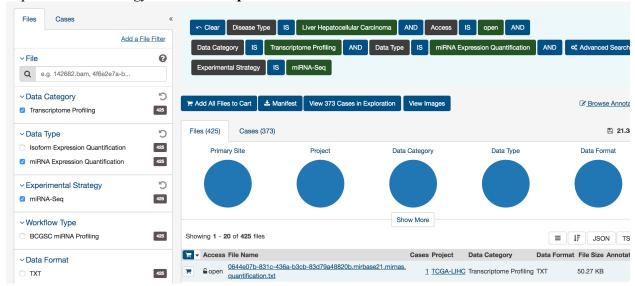
Download Expression Quantification data: miRNA sequence data

- 1. Go to the data portal <a href="https://portal.gdc.cancer.gov/repository">https://portal.gdc.cancer.gov/repository</a>, on the left side there are two tabs: Files and Cases
- 2. Click Cases and select a disease type: Liver Hepatocellular Carcinoma
- 3. Click Files and select

Data Category: Transcriptome Profiling

Data type: miRNA Expression Quantification

Experimental Strategy: miRNA-Seq



You will see 373 cases and 425 files. That means there are duplicates for some cases. Also in those cases, there are some normal cases without cancer.

4. Click on the **Manifest download.** This will download the manifest file for use with GDC data transfer tool.

The Manifest file contains the id, filename, md5, size and patient state.

id filename md5 size	state		
baa65cc1-acb7-46c0-b68b-ce11600b476d	0644e07b-831c-436a-b3cb-83d79a48820b.mirbase21.mirnas.quantification.txt	1a94e462dd73d4bbb8a2866e09d568e6	50268 live
593d4a08-a05e-42c8-9440-4176fbf177fe	455502e4-9e9b-48b5-a6be-1a722de47909.mirbase21.mirnas.quantification.txt	66e27e0fbbeb4fd0de482c0e1b7194d8	50261 live
9a7a5f6e-5b59-48d4-8ad6-608ddd5e739e	8c8e40d1-f62f-4d9d-9435-ef0bc4fcd64e.mirbase21.mirnas.quantification.txt	86650cd63bbd18211f37d4275f974877	50137 live
1e78c8a5-aed7-4cf7-8c99-ba2220053948	7bb700da-edf4-4831-80ef-8d252f590257.mirbase21.mirnas.quantification.txt	4ce6c76f6af6ff9684d4698346ea8895	50301 live
517095c1-a30d-4582-9f41-fcb9ddd251e1	f7f8clac-96cb-49bf-a485-dc8404105191.mirbase21.mirnas.quantification.txt	ab7e824cf406d4af257c95535c456b50	50046 live
9b23f8cb-6b59-4040-b7c8-ba4fa08eba55	466776cb-6906-4da2-b788-a05a154decf3.mirbase21.mirnas.quantification.txt	aec98f0de51afae776f88218f9c6676d	50206 live
0b74f41b-1771-4f42-8181-ca6fc7686b5c	6bee6719-9ee9-4561-8c59-1667f2632d52.mirbase21.mirnas.quantification.txt	e3eca02afce43633544f433cbdb4dfd3	50358 live
963ea266-2577-425f-b47d-168d6c8d7c72	820f4603-0895-4741-865c-fdd98fbcb4fb.mirbase21.mirnas.quantification.txt	06f2c8199f2ff690f6191d14e89bc6ac	50133 live
9a1850a4-7050-4c64-8d3d-50d040cb7c89	bb405508-0f6a-4353-8c87-887be99855d5.mirbase21.mirnas.quantification.txt	fd40688c8c81efa0c88cdb2b68d051de	50402 live
74026969-eab8-44f1-8746-d89d5a450ba1	45dalc01-0316-4dbf-939b-4a758fd7e5e7.mirbase21.mirnas.quantification.txt	f20d3488568b2446f558fd50e0f9e7c4	50294 live
772f0a50-c019-4d85-a1c9-6534ff57f482	f7332ald-bal6-44cd-b6c8-2639fdd568bf.mirbase21.mirnas.quantification.txt	a09f9536d861e0ae81ff46ec3597b2ed	50361 live

5. Data transfer tool Download:

https://gdc.cancer.gov/access-data/gdc-data-transfer-tool

Download the version according to your OS type.

Command line to **download** and **unzip** a **OSX** version:

## Download:

wget -c -t 0 <u>https://gdc.cancer.gov/files/public/file/gdc-client\_v1.3.0\_OSX\_x64.zip</u> Unzip:

Unzip gdc-client v1.3.0 OSX x64.zip

Note: For other versions, just replace with the corresponding OS version file name.

## **Binary Distributions**

Links to the binary distributions for supported platforms are provided below.

- gdc-client\_v1.3.0\_Windows\_x64.zip
- gdc-client\_v1.3.0\_Ubuntu14.04\_x64.zip
- #gdc-client\_v1.3.0\_OSX\_x64.zip
- Mgdc-client\_v1.3.0\_CentOS7\_x64\_Beta.zip
- Try out the new Beta GDC Data Transfer Tool User Interface!

# **System Recommendations**

The system recommendations for using the GDC Data Transfer Tool are as follows:

- OS: Linux (Ubuntu 14.x or later), OS X (10.9 Mavericks or later), or Windows (7 or later)
- CPU: At least eight 64-bit cores, Intel or AMD
- RAM: At least 8 GiB
- Storage: Enterprise-class storage system capable of at least 1 Gb/s (gigabit per second) write throughput and sufficient free space for BAM files.

Please use a AWS machine instance if your own laptop does not meet the requirement.

- 6. Download the files with gdc-client tool:
  - a. make a directory for the data: mkdir live miRNA

cd live miRNA

b. Download with gdc-client.

./<path-to-gdc-client>/gdc-client download –m <path-to-manifest-file> e.g.

./~/Downloads/gdc-client -m ~/Downloads/gdc\_manifest.2018-08-23.txt

Check the successful download:

Since large volumes of data are downloaded, it is important to check the file integrity. You could use the md5 checksum to check the integrity of downloaded files.

Run the code: python3 check.py

A sample python 3 code **check.py** is provided.

8. If some files fail download, use the following command:

```
./<path-to-gdc-client>/gdc-client download <id>e.g.
../gdc-client download fa63ce14-b9b5-4041-9df7-3b86ba9ede16
```

9. Once we get the biomarker files. We also need get the case ids related to the files.

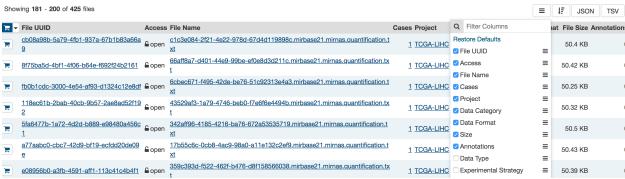
This is because we need correlate the biomarker files with the corresponding case clinical/biospecimen files.

Here we need to write some python codes to extract all the file\_ids and the corresponding case ids for future use.

Get the cases related to the files:

The code *parse\_file\_case\_id.py* is provided.

Click on the tab , and check all the following items, then click on the **JSON** tab. It will download the case ids for the files.



Screenshot of a downloaded file:

```
"file name": "0644e07b-831c-436a-b3cb-83d79a48820b.mirbase21.mirnas.quantification.txt",
"data_format": "TXT",
"access": "open",
"file_id": "baa65cc1-acb7-46c0-b68b-ce11600b476d",
"data_category": "Transcriptome Profiling",
"file_size": 50268,
"cases": [
    "project": {
      "project_id": "TCGA-LIHC"
    "case_id": "7bdc5f86-4d7d-4f1f-bc23-ab51fa9fb947"
"file_name": "455502e4-9e9b-48b5-a6be-1a722de47909.mirbase21.mirnas.quantification.txt",
"data_format": "TXT",
"access": "open",
"file_id": "593d4a08-a05e-42c8-9440-4176fbf177fe",
"data_category": "Transcriptome Profiling",
"file_size": 50261,
"cases": [
    "project": {
      "project_id": "TCGA-LIHC"
    "case id": "801b1d2c-eb6f-4eef-a00b-83da939d755a"
```

11. Get the meta data for the files and corresponding cases:

The source code: *request\_meta.py*The fields for the files and cases:

File fields:

https://docs.gdc.cancer.gov/API/Users\_Guide/Appendix\_A\_Available\_Fields/#file-fields case fields:

https://docs.gdc.cancer.gov/API/Users Guide/Appendix A Available Fields/#case-fields

Once we get the meta data for the miRNA files, we can see that some samples come from a normal solid tissue and some others come from tumor.

12. Now we could generate the miRNA matrix for all the files with labeled normal or tumor. The miRNA seq that comes from tumor is labeled with 1, and normal tissue is labeled with 0.

Part 2: Apply Machine Learning Package (sklearn) to the above data.

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Sample code provided: predict.pv

The steps are as following:

- 1. Data standardization.
- 2. Train and test data split.
- 3. Feature selection.
- 4. Model hyper-parameters tuning with cross validation
- **5.** Model prediction with the best hyper-parameters
- 6. Evaluation: Precision, Sensitivity, Accuracy, F1-score, Specificity

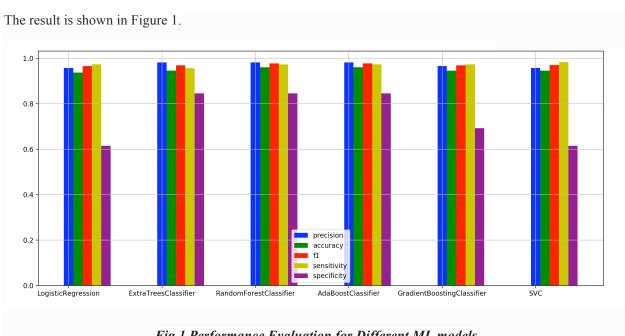


Fig.1 Performance Evaluation for Different ML models

Below are some good reference papers for your project.

# **Reference:**

- [1] Hyeongmin Kim & Yong-Min Kim, "Pan-cancer analysis of somatic mutations and transcriptomes reveals common functional gene clusters shared by multiple cancer types," Scientific Reports, volume 8, Article number: 6041 (2018) ,https://www.nature.com/articles/s41598-018-24379-y
- [2] Marieke Lydia Kuijjer, Joseph Nathaniel Paulson, Peter Salzman, Wei Ding & John Quackenbush, "Cancer subtype identification using somatic mutation data,", British Journal of Cancervolume 118, pages1492–1501 (2018).
- [3] https://medium.com/@luckylwk/visualising-high-dimensional-datasets-using-pca-and-t-snein-python-8ef87e7915b