# Palmer Archipelago Penguins Data in the palmerpenguins R Package - An Alternative to Anderson's Irises

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Abstract In 1935, Edgar Anderson collected size measurements for 150 flowers from three species of Iris on the Gaspé Peninsula in Quebec, Canada. Since then, Anderson's Iris observations have become a classic dataset in statistics, machine learning, and data science teaching materials. It is included in the base R datasets package as iris, making it easy for users to access without knowing much about it. However, the lack of data documentation, presence of non-intuitive variables (e.g. "sepal width"), and perfectly balanced groups with zero missing values make iris an inadequate and stale dataset for teaching and learning modern data science skills. Users would benefit from working with a more representative, real-world environmental dataset with a clear link to current scientific research. Importantly, Anderson's Iris dataset appeared in a 1936 publication by R. A. Fisher in the Annals of Eugenics (which is often the first-listed citation for the dataset), inextricably linking iris to racist research and theory. Thus, a modern alternative to the **iris** dataset is needed. In this paper, we introduce the palmerpenguins R package, which includes morphological measurements collected from 2007 - 2009 for three species of Pygoscelis penguins that breed on islands throughout the Palmer Archipelago, Antarctica. The penguins data provides an approachable, charismatic, and near drop-in replacement for iris with topical relevance global climate change and environmental impacts on marine predators. Since the release on CRAN in July 2020, the palmerpenguins package has been downloaded over 105,000 times, highlighting the demand and widespread adoption of this viable iris alternative. We directly compare the **iris** and **penguins** datasets for selected analyses to demonstrate that R users, in particular teachers and learners currently using iris, can switch to the Palmer Archipelago penguins for many use cases including data wrangling, visualization, linear modeling, multivariate analysis (e.g., PCA), cluster analysis and classification (e.g., by k-means).

#### Introduction

In 1935, American botanist Edgar Anderson measured petal and sepal structural dimensions (length and width) for 50 flowers from three *Iris* species: *Iris setosa, Iris versicolor*, and *Iris virginica* (Anderson, 1935). The manageable but non-trivial size (5 variables and 150 total observations) and characteristics of the dataset, including linear relationships and multivariate normality, have made the dataset amenable for introducing a wide range of statistical methods including data wrangling, visualization, linear modeling, multivariate analyses, and machine learning. Anderson's *Iris* data is built into a number of software packages including the auto-installed datasets package in R (R Core Team, 2019), Python's scikit-learn machine learning library (Pedregosa et al., 2011), and the SAS Sashelp library (SAS Institute, Cary NC), which have facilitated widespread use of the *Iris* data. As a result, eighty-six years after the data were initially published, Anderson's *Iris* measurements are ubiquitous in statistics, computational methods, software documentation, and data science courses and materials.

There are a number of reasons that modern data science practitioners and educators may want to move on from Anderson's Iris dataset. First, Anderson's Iris dataset lacks metadata almost entirely (Anderson, 1935), which does not reinforce best practices and limits meaningful interpretation and discussion of research methods, analyses, and outcomes. Of the five variables in the dataset, two (Sepal.Width and Sepal.Length) are not intuitive for most non-botanists. Even with explanation, the difference between Iris petal and sepal dimensions is not obvious. Second, the Iris dataset contains equal sample sizes for each of the three species (n = 50) with no missing values, which is cleaner than most real-world datasets that learners are likely to encounter "in the wild." Third, the single factor (Species) in the Iris data limits options for analyses. Finally, due to its publication in the Annals of Eugenics by eugenicist and statistician R.A. Fisher (Fisher, 1936), the Iris dataset is burdened by a history in scientific racism, which we are committed to addressing through the development of new data science education products as described below.

Given the growing need for fresh data science-ready datasets, we sought to identify an alternative dataset that could be made easily accessible for a broad audience. After evaluating the positive and negative features of the *Iris* dataset in data science and statistics materials, we established the following criteria for a suitable alternative:

- Available by appropriate license (ideally, CC0 "no rights reserved")
- Feature intuitive subjects and variables that are interesting and understandable to learners

across disciplines

- Complete metadata and documentation
- Manageable (but not trivial) in size
- Minimal data cleaning and pre-processing required for most analyses
- Real-world (not manufactured) modern data
- Provides similar opportunities for teaching and learning R, data science, and statistical skills
- Can easily replace *Iris* for most use cases

Here, we describe an alternative to Anderson's *Iris* data that largely satisfies these criteria: a refreshing, approachable, and charismatic dataset containing real-world morphological data for three *Pygoscelis* penguin species that breed throughout the Western Antarctic Peninsula region, made available through the United States Long-Term Ecological Research (US LTER) Network. By comparing data structure, size, and a range of analyses side-by-side for the two datasets, we demonstrate that the Palmer Archipelago penguin data are an ideal substitute for Anderson's *Iris* dataset for many use cases in statistics and data science education.

#### Data source

Body size measurements, clutch (i.e., egg laying) observations (e.g., date of first egg laid, and clutch completion), and carbon ( $^{13}$ C/ $^{12}$ C) and nitrogen ( $^{15}$ N/ $^{14}$ N,  $\delta^{15}$ N) stable isotope values of red blood cells for adult male and female Adélie (P. adeliae), chinstrap (P. antarcticus), and gentoo (P. papua) penguins on three islands (Biscoe, Dream and Torgersen) within the Palmer Archipelago were collected from 2007 - 2009 by Dr. Kristen Gorman in collaboration with the Palmer Station LTER, part of the US LTER Network. For complete data collection methods and published analyses, see Gorman et al. (2014). Throughout this paper, penguins species referred to as "Adélie", "Chinstrap", and "Gentoo."

The data in the **palmerpenguins** R package are available for use by CC0 license ("No Rights Reserved") in accordance with the Palmer Station LTER Data Policy and the LTER Data Access Policy, and were imported from the Environmental Data Initiative (EDI) Data Portal at the links below:

- Adélie penguin data (Palmer Station Antarctica LTER and Gorman, 2020a): KNB-LTER Data Package 219.5
- Gentoo penguin data (Palmer Station Antarctica LTER and Gorman, 2020b): KNB-LTER Data Package 220.5
- Chinstrap penguin data (Palmer Station Antarctica LTER and Gorman, 2020c): KNB-LTER Data Package 221.6

## R package: palmerpenguins

R users can install the palmerpenguins package from CRAN:

install.packages("palmerpenguins")

Information, examples, and links to community-contributed materials are available on the palmer-penguins package website: https://allisonhorst.github.io/palmerpenguins/. See the Appendix for how Python and Julia users can access the same data.

The palmerpenguins R package contains two data objects: penguins\_raw and penguins. The penguins\_raw data consists of all raw data for 17 variables, recorded completely or in part for 344 individual penguins, accessed directly from EDI (penguins\_raw properties are summarized in Appendix B). We generally recommend using the curated data in penguins, which is a subset of penguins\_raw retaining all 344 observations, minimally updated (Appendix A) and reduced to the following eight variables:

- species: a factor denoting the penguin species (Adélie, Chinstrap, or Gentoo)
- *island:* a factor denoting the Palmer Archipelago island in Antarctica where each penguin was observed (Biscoe Point, Dream Island, or Torgersen Island)
- bill\_length\_mm: a number denoting length of the dorsal ridge of a penguin bill (millimeters)
- bill\_depth\_mm: a number denoting the depth of a penguin bill (millimeters)
- flipper\_length\_mm: an integer denoting the length of a penguin flipper (millimeters)
- body\_mass\_g: an integer denoting the weight of a penguin's body (grams)
- sex: a factor denoting the sex of a penguin sex (male, female) based on molecular data
- year: an integer denoting the year of study (2007, 2008, or 2009)

**Table 1:** Overview comparison of penguins and iris dataset features and characteristics.

Feature	iris	penguins
Year(s) collected	1935	2007 - 2009
Dimensions (col x row)	5 x 150	8 x 344
Documentation	minimal	complete metadata
Variable classes	double (4), factor (1)	double (2), int (3), factor (3)
Missing values?	no $(n = 0; 0.0\%)$	yes (n = 19; 0.7%)

**Table 2:** Grouped sample size for iris (by species; n = 150 total) and penguins (by species and sex; n = 344 total). Penguins can be further grouped by variables for island and study year.

Iris sample size (by species)		Penguin sample size (by species and sex)			
Iris species	Sample size	Penguin species	Female	Male	NA
setosa	50	Adélie	73	73	6
versicolor	50	Chinstrap	34	34	0
virginica	50	Gentoo	58	61	5

The same data exist as comma-separated value (CSV) files in the package ("penguins\_raw.csv" and "penguins.csv"), and can be read in using the built-in path\_to\_file() function in palmerpenguins. For example,

```
library(palmerpenguins)
df <- read.csv(path_to_file("penguins.csv"))</pre>
```

will read in "penguins.csv" as if from an external file, thus automatically parsing *species*, *island*, and *sex* variables as characters instead of factors. This option allows users opportunities to practice or demonstrate reading in data from a CSV, then updating variable class (e.g., characters to factors).

## Comparing iris and penguins

The **penguins** data in **palmerpenguins** is useful and approachable for data science and statistics education, and is uniquely well-suited to replace the **iris** dataset. Comparisons presented are selected examples for common **iris** uses, and are not exhaustive.

## Data structure and sample size

Both **iris** and **penguins** are in tidy format (Wickham, 2014) with each column denoting a single variable and each row containing measurements for a single *Iris* flower or penguin, respectively. The two datasets are comparable in size: dimensions (columns  $\times$  rows) are  $5 \times 150$  and  $8 \times 344$  for **iris** and **penguins**, respectively, and sample sizes within species are similar (Table 1).

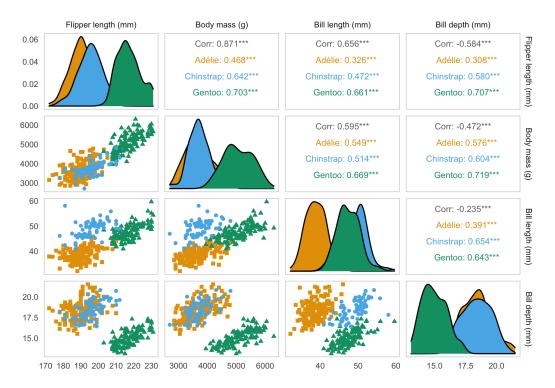
Notably, while sample sizes in **iris** across species are all the same, sample sizes in **penguins** differ across the three species. The inclusion of three factor variables in **penguins** (*species*, *island*, and *sex*), along with *year*, create additional opportunities for grouping, faceting, and analysis compared to the single factor (*Species*) in **iris**.

Unlike **iris**, which contains only complete cases, the **penguins** dataset contains a small number of missing values ( $n_{missing} = 19$ , out of 2752 total values). Missing values and unequal sample sizes are common in real-world data, and create added learning opportunity to the **penguins** dataset.

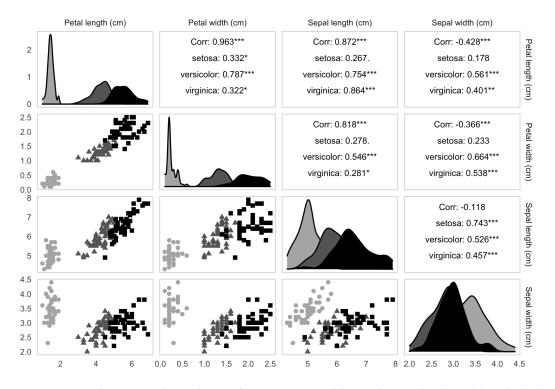
## Continuous quantitative variables

Distributions, relationships between variables, and clustering can be visually explored between species for the four structural size measurements in **penguins** (flipper length, body mass, bill length and depth; Figure 1) and **iris** (sepal width and length, petal width and length; Figure 2).

Both **penguins** and **iris** offer numerous opportunities to explore linear relationships and correlations, within and across species (Figures 1 & 2). A bivariate scatterplot made with the **iris** dataset reveals a clear linear relationship between petal length and petal width. Using **penguins** (3), we can

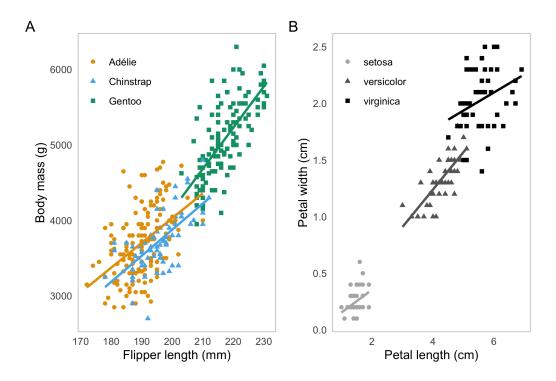


**Figure 1:** Distributions and correlations for numeric variables in the penguins data (flipper length (mm), body mass (g), bill length (mm) and bill depth (mm)) for the three observed species: Gentoo (green, triangles); Chinstrap (blue, circles); and Adélie (orange, squares). Significance levels indicated for bivariate correlations are: \*p < 0.05; \*p < 0.01; \*p < 0.01.



**Figure 2:** Distributions and correlations for numeric variables in the iris data (petal length (cm), petal width (cm), sepal length (cm) and sepal width (cm)) for the three included Iris species: setosa (light gray, circles); versicolor (dark gray, triangles); and virginica (black, squares). Significance levels indicated for bivariate correlations are: \*p < 0.05; \*\*p < 0.01; \*\*\*p < 0.001.

create a uniquely similar scatterplot with flipper length and body mass. The overall trend across all three species is approximately linear for both **iris** and **penguins**. Teachers may encourage students to explore how simple linear regression results and predictions differ when the species variable is omitted, compared to, for example, multiple linear regression with species included (3).



**Figure 3:** Representative linear relationships for (A): penguin flipper length (mm) and body mass (g) for Adélie (orange circles), Chinstrap (blue triangles), and Gentoo (green squares) penguins; (B): iris petal length (cm) and width (cm) for setosa (light gray circles), versicolor (dark gray triangles) and virginica (black squares) irises. Within-species linear model is visualized for each penguin or iris species.

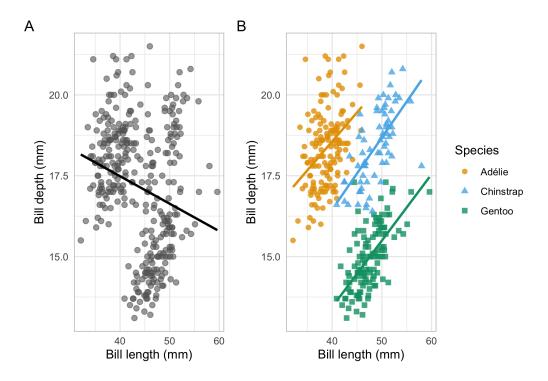
Notably, distinctions between species are clearer for iris petals - particularly, the much smaller petals for *Iris setosa* - compared to penguins, in which Adélie and Chinstrap penguins are largely overlapping in body size (body mass and flipper length), and are both generally smaller than Gentoo penguins.

Simpson's Paradox is a data phenomenon in which a trend observed between variables is reversed when data are pooled, omitting a meaningful variable. While often taught and discussed in statistics courses, finding a real-world and approachable example of Simpson's Paradox can be a challenge. Here, we show one (of several possible - see Figure 1) Simpson's Paradox examples in **penguins**: exploring bill dimensions with and without species included (Figure 4). When penguin species is omitted (Figure 4 A), bill length and depth appear negatively correlated overall. The trend is reversed when species is included, revealing an obviously positive correlation between bill length and bill depth within species (Figure 4 B).

# Principal component analysis

Principal component analysis (PCA) is a dimensional reduction method commonly used to explore patterns in multivariate data. The **iris** dataset frequently appears in PCA tutorials due to multivariate normality and clear interpretation of variable loadings and clustering.

A comparison of PCA with the four variables of structural size measurements in **penguins** and **iris** (both normalized prior to PCA) reveals highly similar results (Figure 5). For both datasets, one species is distinct (Gentoo penguins, and setosa irises) while the other two species (Chinstrap/Adélie and versicolor/virginica) appear somewhat overlapping in the first two principal components (Figure 5 A,B). Screeplots reveal that the variance explained by each principal component (PC) is very similar across the two datasets, particularly for PC1 and PC2: for **penguins**, 88.15% of total variance is captured by the first two PCs, compared to 95.81% for **iris**, with a similarly large percentage of variance captured by PC1 and PC2 in each (Figure 5 C,D).



**Figure 4:** Trends for penguin bill dimensions (bill length and bill depth, millimeters) if the species variable is excluded (A) or included (B), illustrating Simpson's Paradox. Note: linear regression for bill dimensions without including species in (A) is ill-advised; the linear trendline is only included to visualize trend reversal for Simpson's Paradox when compared to (B).

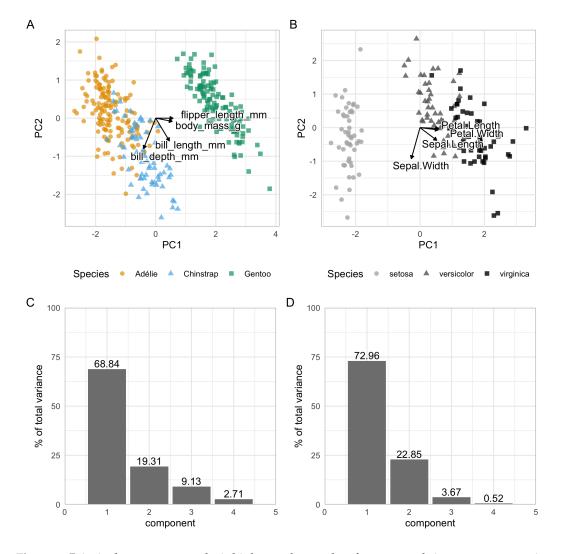
Table 3: K-means cluster assignments by species based on penguin bill length (mm) and depth (mm), and iris petal length (cm) and width (cm).

Penguins cluster assignments				Iris cluste	er assignmen	its	
Cluster	Adélie	Chinstrap	Gentoo	Cluster	setosa	versicolor	virginica
1	0	9	116	1	0	2	46
2	4	54	6	2	0	48	4
3	147	5	1	3	50	0	0

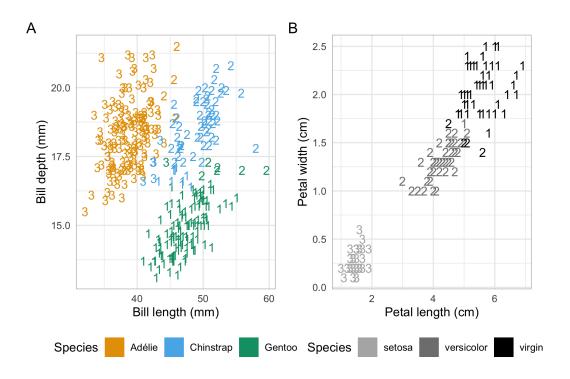
# K-means clustering

Unsupervised clustering by k-means is a common and popular entryway to machine learning and classification, and again, the **iris** dataset is frequently used in introductory examples. The **penguins** data provides similar opportunities for introducing k-means clustering. For simplicity, we compare k-means clustering using only two variables for each dataset: for **iris**, petal width and petal length, and for **penguins**, bill length and bill depth. All variables are scaled prior to k-means. Three clusters (k = 3) are specified for each, since there are three species of both *Iris* (*setosa*, *versicolor*, and *virginica*) and penguins (Adélie, Chinstrap and Gentoo).

K-means clustering with penguin bill dimensions and iris petal dimensions yields largely distinct clusters, each dominated by one species (6). For iris petal dimensions, k-means yields a perfectly separated cluster (Cluster 3) containing all 50 setosa iris observations and zero misclassified virginica or versicolor irises (Table 2). While clustering is not perfectly distinct for any penguin species, each species is largely contained within a single cluster, with little overlap from the other two species. For example, considering Adélie penguins (orange observations in Figure 6 A): 147 (out of 151) Adélie penguins are assigned to Cluster 3, zero are assigned to Cluster 1, and 4 are assigned to the Chinstrap-dominated Cluster 2 (Table 2). Only 5 (of 68) Chinstrap penguins and 1 (of 123) Gentoo penguins are assigned to the Adélie-dominated Cluster 3 (Table 2).



**Figure 5:** Principal component analysis biplots and screeplots for structural size measurements in penguins (A,C) and iris (B,D), revealing similarities in multivariate patterns, variable loadings, and variance explained by each component. For penguins, variables are flipper length (mm), body mass (g), bill length (mm) and bill depth (mm); groups are visualized by species (Adélie = orange circles, Chinstrap = blue triangles, Gentoo = green squares). For iris, variables are petal length (cm), petal width (cm), sepal length (cm) and sepal width (cm); groups are visualized by species (setosa = light gray circles, versicolor = dark gray triangles, virginica = black squares). Values above screeplot columns (C,D) indicate percent of total variance explained by each of the four principal components.



**Figure 6:** K-means clustering outcomes for penguin bill dimensions (A) and iris petal dimensions (B). Numbers indicate the cluster to which an observation was assigned, revealing a high degree of separation between species for both penguins and iris.

#### Conclusion

Here, we have shown that structural size measurements for Palmer Archipelago *Pygoscelis* penguins, available as **penguins** in the **palmerpenguins** R package, offer a near drop-in replacement for **iris** in a number of common use cases for data science and statistics education including exploratory data visualization, linear correlation and regression, PCA, and clustering by k-means. In addition, teaching and learning opportunities in **penguins** are increased due to a greater number of variables, missing values, unequal sample sizes, and Simpson's Paradox examples. Importantly, the **penguins** dataset encompasses real-world information derived from several charismatic marine predator species with regional breeding populations notably responding to environmental change occurring throughout the Western Antarctic Peninsula region of the Southern Ocean (see Bestelmeyer et al. (2011), Gorman et al. (2014), Gorman (2015), Gorman et al. (2017)). Thus, the **penguins** dataset can facilitate discussions more broadly on biodiversity responses to global change - a contemporary and critical topic in ecology, evolution, and the environmental sciences.

# **Appendix**

#### Penguins data processing

Data in the **penguins** object have been minimally updated from **penguins\_raw** as follows:

- All variable names are converted to lower snake case (e.g. from Flipper Length (mm) to flipper\_length\_mm)
- Entries in *species* are truncated to only include the common name (e.g. "Gentoo", instead of "gentoo penguin (Pygoscelis papua)")
- Recorded sex for penguin N36A1, originally recorded as ".", is updated to NA
- *culmen\_length\_mm* and *culmen\_depth\_mm* variable names are updated to *bill\_length\_mm* and *bill\_depth\_mm*, respectively
- Class for categorical variables (species, island, sex) is updated to factor
- Variable year was pulled from clutch observations

#### Summary of the penguins\_raw dataset

Feature	penguins_raw
Year(s) collected	2007 - 2009
Dimensions (col x row)	17 x 344
Documentation	complete metadata
Variable classes	character (9), Date (1), numeric (7)
Missing values?	yes (n = 336; 5.7%)

# palmerpenguins for other programming languages

Python: Python users can load the palmerpenguins datasets into their Python environment using the following code to install and access data in the palmerpenguins Python package:

```
pip install palmerpenguins
from palmerpenguins import load_penguins
penguins = load_penguins()
```

Julia: Julia users can access the penguins data in the **PalmerPenguins.jl** package. Example code to import the penguins data through **PalmerPenguins.jl** (more information on **PalmerPenguins.jl** from David Widmann can be found here):

```
julia> using PalmerPenguins
julia> table = PalmerPenguins.load()
```

## Acknowledgements

All analyses were performed in the R language environment using version 3.6.2 (R Core Team, 2019). Complete code for this paper is shared in the Supplemental Material. We acknowledge the following R packages used in analyses, with gratitude to developers and contributors:

- tidyverse (Wickham et al., 2019): for data import and cleaning
- ggplot2 (Wickham, 2016): for data visualizations
- kableExtra (Zhu, 2019): for finalized tables
- GGally (Schloerke et al., 2020): for pairs plots
- patchwork (Pedersen, 2019): for compound figures
- shadowtext (Yu, 2019): to add a background color to text labels
- recipes (Kuhn and Wickham, 2020): for data pre-processing
- pkgdown (Wickham and Hesselberth, 2020): to build the package website
- base and stats (R Core Team, 2019): for various analyses throughout

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