



<CODE-ON>

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# Determination of **restriction sites** and their customized hiding within **DNA sequence**

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# Program's **main menu** and outputs

DNA sequence analyzer

DNA sequence

Row	Base sequences (5' – 3')	Restriction site name
0	GAATTC	EcoRI
1	TCTAGA	XbaI
2	ACTAGT	SpeI
3	CTGCAG	PstI
4	GCGGCCGC	NotI
5	GCTCTTC	SapI
6	GGTCTC	BsaI
7	ATCGAT	BavCI

Base  Name

Resulting DNA

The given DNA sequence in 5'-3' is ATCGATCCCGAATTCGAATTCAAA and in 3'-5' is TAGCTAGGGCTTAAGCTTAAGTTT

The selected restriction sites are GAATTC — EcoRI, TCTAGA — XbaI, ACTAGT — SpeI, CTGCAG — PstI, GCGGCCGC — NotI, GCTCTTC — SapI, GGTCTC — BsaI, ATCGAT — BavCI.

In the 3'-5' DNA sequence there are following restrictions:

- TAGCTA named as BavCI found at positions 0.
- CTTAAG named as EcoRI found at positions 9.

In the 5'-3' DNA sequence there are following restrictions:

- ATCGAT named as BavCI found at positions 0.
- GAATTC named as EcoRI found at positions 9.

The DNA sequences resulting after the clearing of the selected restriction:

(5' – 3'): ATAGATCCCGAGTTCGAGTTCAAA

(3' – 5'): TATCTAGGGCTCAAGCTCAAGTTT

# What is **special** about our program?

The **main feature** of our program is that it not only determines the precise location of prohibited restriction sites according to iGEM standards, but also allows the user to **hide these sites** by altering the minimum number of **nucleotides**.

The algorithms are **versatile and adaptable**, so the researcher may **add** new restriction sites into the database and/or **delete** previous ones, so we **are not limited** with those provided by default.

The program will detect and change the DNA sequence according to the user's preferences.

# What is **special** about our program?

An intuitive **interface** visually shows the restriction sites within the DNA sequence and **annotates the precise name** and **position** of the restriction site.

While the user enters DNA sequence for 5'-3' direction, they receive the **similar data output** for the second strand with reversed direction.

After determining the number and character of **restriction site**, the researcher can **hide** either all restriction sites or **customize** which restriction site they prefer to alter.

The program makes sure that **DNA alteration** is similar to silent mutation and **amino acids sequence** encoded in coding frame remains the same.

# What is **special** about our program?

- Works for **bacterial** genome
- Inputs: the **DNA** sequence and **restriction sites** sequences to be further removed
- Can be applied for **full gene sequence**
- Assumes that DNA sequence input = coding region

# How-to run the program

- In the raw “DNA sequence”, enter the DNA sequence for analysis. The input sequence should be a coding region in 5’-3’ direction.
- By default, to determine the prohibited restriction sites and their location, **no sites** from an initial database are chosen. So you may choose all of them.
- To run the program, press the “Run” button.
- The resulting output **identifies** prohibited restriction sites within the given DNA sequence, and names their precise position.
- Also, the program presents **similar data** for the reversed (3’-5’) strand without additional input.

# How-to run the program

- By default, the program alters the nucleotide sequence and hides **all found** prohibited restriction sites within the given DNA sequence.
- If you want to **customize** which restriction sites you would like to remove, you may change the initial settings.
- To do so, close the program window to return to the main menu.  
**Note:** in the menu's table, the light blue sites will remain unchanged; the dark blue sites will be hidden.
- Click on dark blue sites in the table of restriction sites to choose those to **keep** them within the DNA sequence.
- Press the "Run" button.

# Let's deal with some **algorithms!**

For further convenience, we created **supplementary dictionaries**, that can convert amino acids' name to codon name that this amino acid encodes.

```
acids_to_codons = {'A': ['GCT', 'GCC', 'GCA', 'GCG'],
                   'R': ['CGT', 'CGC', 'CGA', 'CGG', 'AGA', 'AGG'],
                   'N': ['AAT', 'AAC'],
                   'D': ['GAT', 'GAC'],
                   'B': ['AAT', 'AAC', 'GAT', 'GAC'],
                   'C': ['TGT', 'TGC'],
                   'Q': ['CAA', 'CAG'],
                   'E': ['GAA', 'GAG'],
                   'Z': ['CAA', 'CAG', 'GAA', 'GAG'],
                   'G': ['GGT', 'GGC', 'GGA', 'GGG'],
                   'H': ['CAT', 'CAC'],
                   'Met (start)': ['ATG'],
                   'I': ['ATT', 'ATC', 'ATA'],
                   'L': ['CTT', 'CTC', 'CTA', 'CTG', 'TTA', 'TTG'],
                   'K': ['AAA', 'AAG'],
                   'F': ['TTT', 'TTC'],
                   'P': ['CCT', 'CCC', 'CCA', 'CCG'],
                   'S': ['TCT', 'TCC', 'TCA', 'TCG', 'AGT', 'AGC'],
                   'T': ['ACT', 'ACC', 'ACA', 'ACG'],
                   'W': ['TGG'],
                   'Y': ['TAT', 'TAC'],
                   'V': ['GTT', 'GTC', 'GTA', 'GTG'],
                   'stop': ['TAA', 'TGA', 'TAG']}
```



# Let's deal with some **algorithms!**

Our first **supplementary script** is a “translator” of the **DNA sequence** to its complementary strand. That is the backbone of our program.

```
def complement(dna: str) -> str:
    '''
    Input: a DNA sequences
    Output: its complementary sequences
    '''
    map = {"C": "G", "G": "C", "A": "T", "T": "A"}
    complement_dna = ""
    for nucleotid in dna:
        complement_dna += map[nucleotid]
    return complement_dna
```

# Let's deal with some algorithms!

Our main script deals with the **DNA sequences** and **restriction sites** list. It starts with finding out restrictions in every single **nucleotide bases strand**, followed up by list “app” that includes the tuples of **restriction code**, name of restriction code, and its position in the sequence.

After that, the program goes through the given **sequence** again and creates a **new array** of an equal base length as the DNA itself. Our newly created **array** has the **same length** as the DNA. The numbered element demonstrates if the symbol with the same number belongs to the restriction or not. We will need this array a bit later!

```
def find_instances(dna: str, restrictions: str) -> list:
    '''
    Input: a DNA sequence and restriction site
    Output: a list containing tuples of position, base sequence, and names
    of the selected restriction sites.
    '''
    app = []
    for restriction in restrictions:
        base, name = restriction[0], restriction[1]
        for e in [m.start() for m in re.finditer(base, dna)]:
            app.append((e, base, name))
    app.sort()
    instances = [None] * len(dna)
    pos, j = 0, 0
    while pos < len(dna):
        while j < len(app) and pos > app[j][0]:
            j += 1
        if j >= len(app): break
        if pos == app[j][0]:
            for i in range(len(app[j][1])):
                instances[pos + i] = (app[j][1], app[j][2])
            pos += len(app[j][1]) - 1
        pos += 1
    return instances
```

# Let's deal with some **algorithms!**

The second **supplementary script** converts the DNA sequence to amino acid sequence. There we use the previously mentioned **dictionary**.

```
def convert_to_acids(dna: str) -> list:
    '''
    Input: a DNA sequence
    Output: a sequence of aminoacids
    '''
    acids = []
    for i in range(0, len(dna), 3):
        if i >= len(dna) - 2: break
        codon = dna[i: i+3]
        acids.append(codons_to_acids[codon])
    return acids
```

# Let's deal with some **algorithms!**

The next script hides the **restriction sites** after their **detection** in the given sequence.

The algorithm starts with the **codons detection** and if it finds out one with nucleotide from the restriction site list. Then the program looks for another codons that code the **same nucleotide**.

After that the program changes the **codon** and deletes the restriction from the **instances array**. After such reorganization, we receive the DNA without restriction sites.

```
def remove_instances(dna: str, instances: list) -> str:
    '''
    Input: a DNA sequence and instances of restriction sites in it
    Output: the DNA sequence after removing all restrictions sites from it
    '''

    result_dna = list(dna)
    acids = convert_to_acids(dna)
    for i in range(0, len(dna), 3):
        if instances[i] is not None:
            acid = acids[i // 3]
            for codon in acids_to_codons[acid]:
                if list(codon) != result_dna[i: i + 3]:
                    for j in range(3): result_dna[i + j] = codon[j]
                    if instances[i - 1] is not None:
                        for j in range(i - 3, i):
                            instances[j] = None
                    if i + 3 < len(dna) and instances[i + 3] is not None:
                        for j in range(i + 3, i + 6):
                            instances[j] = None
    result_dna = ''.join(result_dna)
    return result_dna
```

# Let's deal with some **algorithms!**

Next script is needed to put the **output data** in suitable format: found **restriction sites** and their positions.

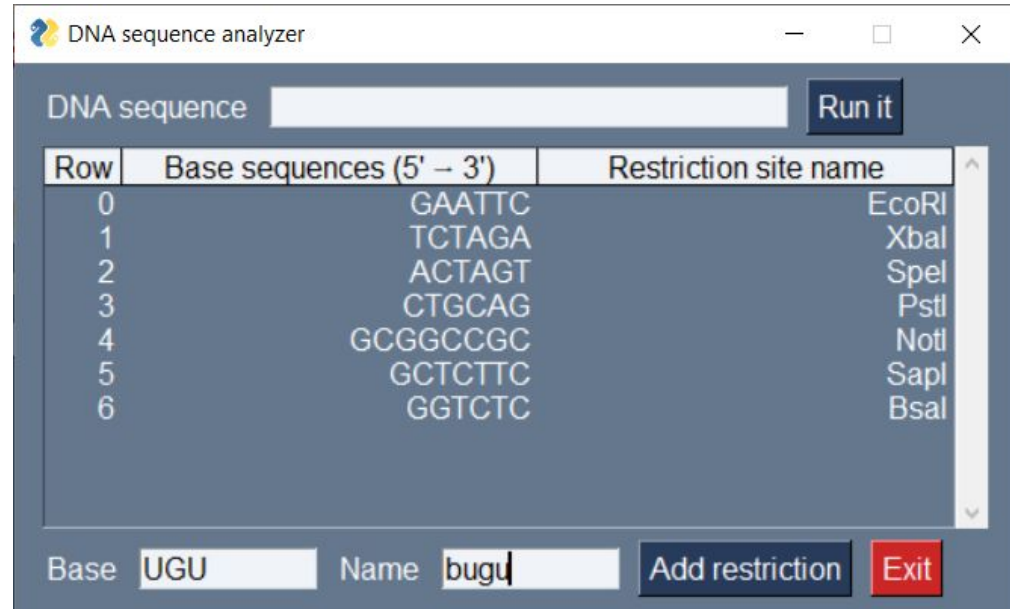
```
def find_positions(instances: list) -> dict:
    """
    Input: instances of restriction sites in the DNA
    Output: a dictionary which maps a restriction into positions where it
    was found in the given DNA
    """
    positions, i = dict(), 0
    while i < len(instances):
        if instances[i] is not None:
            rest = instances[i]
            if rest not in positions: positions[rest] = [i]
            else: positions[rest].append(i)
            i += len(rest[0])
        else: i += 1
    return positions
```

## ... and finally an **interface!**

To make an interface for our project, we decided to use open source module **PySimpleGUI**.

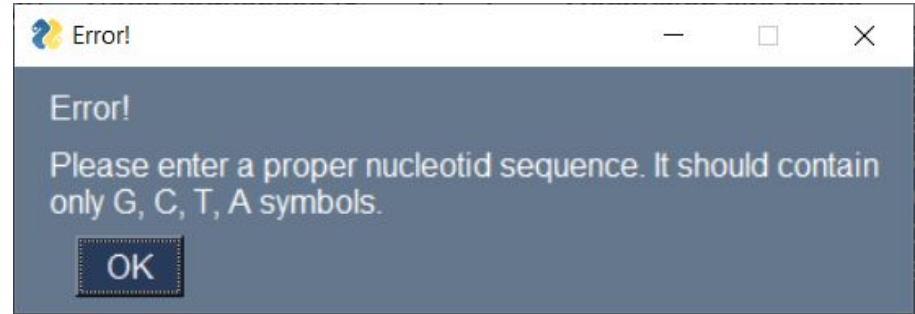
There is a field “DNA sequence” in which we will input the **sequence** of our choice, table of **restriction sites** that user could choose on his own, “Base” and “Name” fields that made for adding new restriction site, and “Exit” button.

Let's try to add a **new restriction** site in the table. For example, UGU and call it 'bugu'.



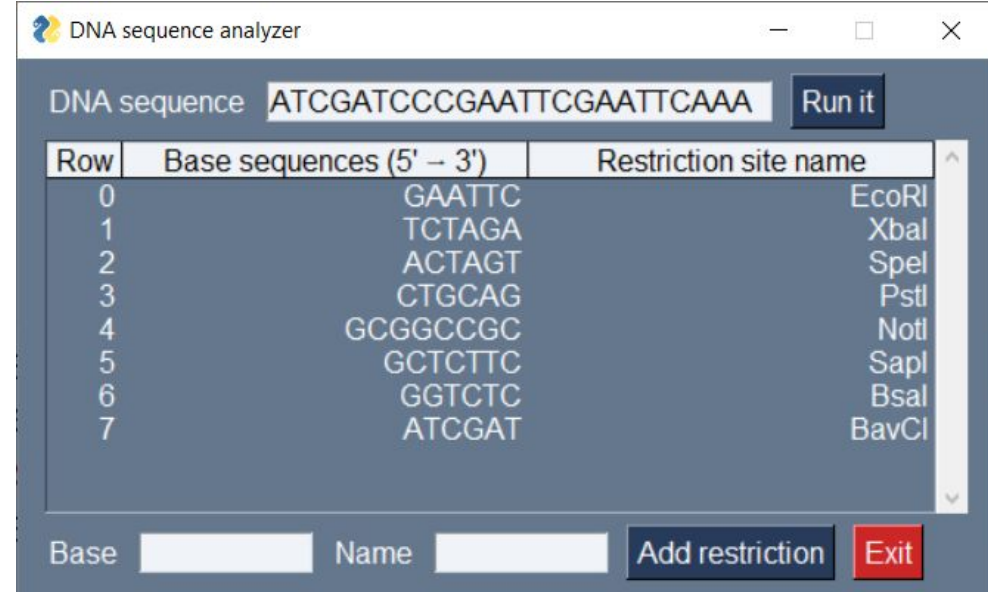
# ... and finally an **interface!**

Oops... We get an error message: The letter U should not be in the code. Let's input the **restriction site** ATCGAT and call it BavCl.



## ... and finally an **interface**!

As you can see, we successfully added a new restriction site. Now, let's test our program. Firstly, we need to choose **restriction sites** of our interest. We want to look at all **restrictions** that could be in the sequence, so we will choose everything.



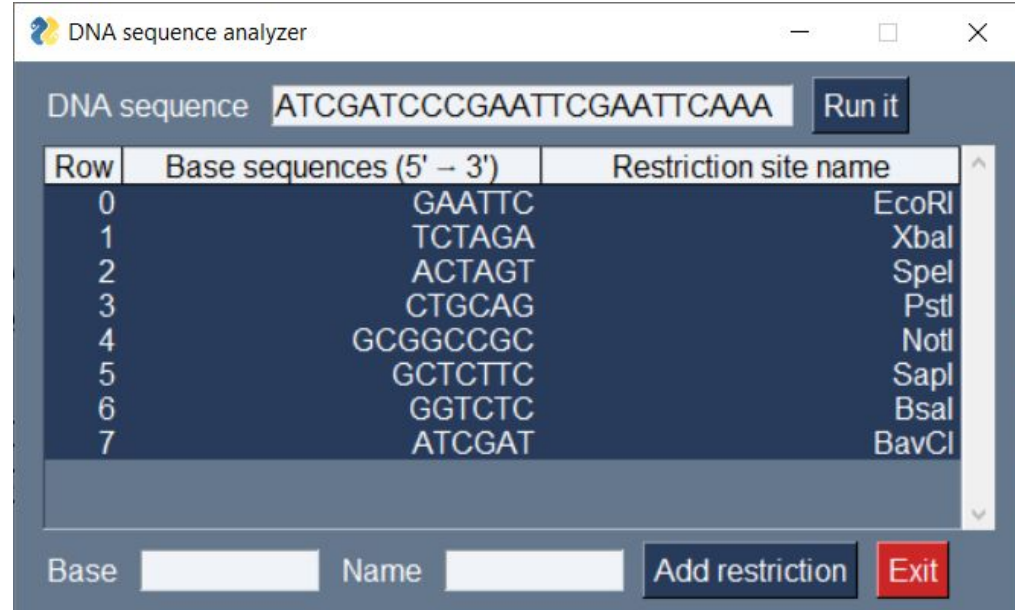


# ... and finally an **interface!**

Now we need to **input** some sequence.

Let it be “ATC GAT CCC GAA  
TTC GAA TTC AAA”

And click the **“Run”** button!

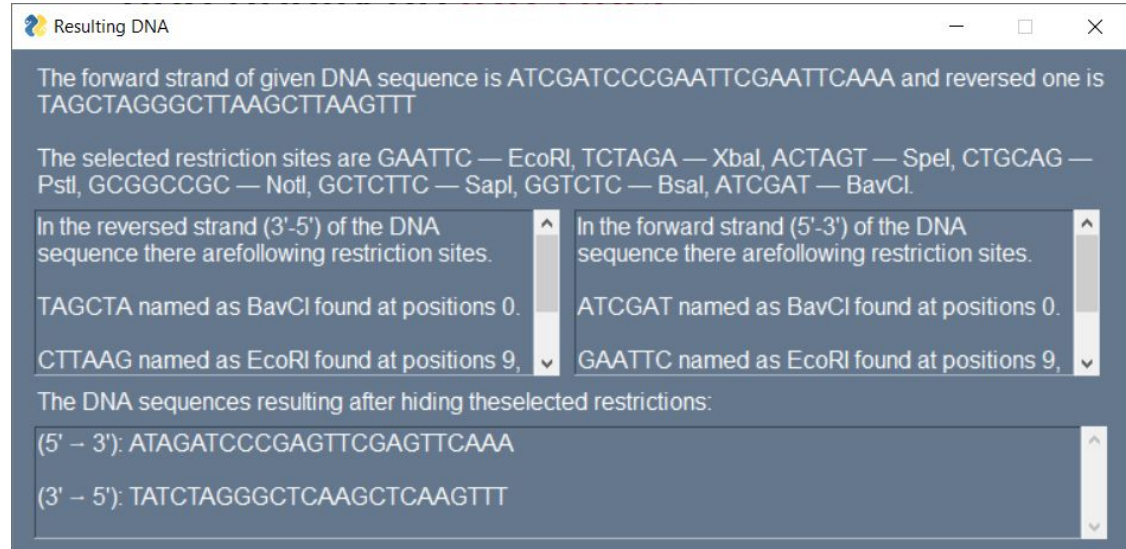


# ... and finally an **interface!**

At first, we see the given **sequence** in 5' → 3' and another complementary sequence. Below we can see the chosen restriction sites.

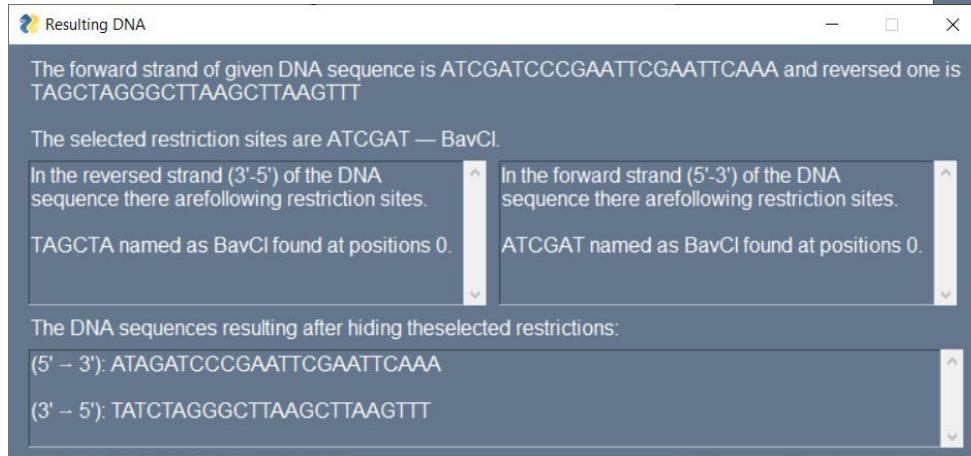
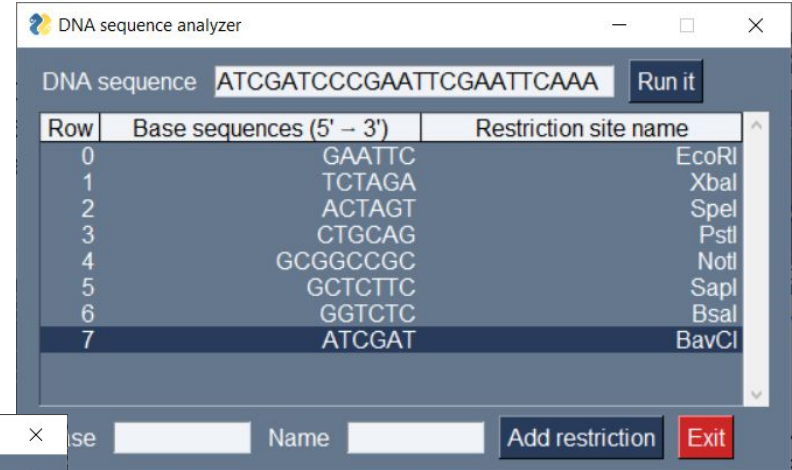
As it turns out, there is **ATCGAT/BavC** on 0 position, and **GAATTC /EcoRI** on 9 and 15 positions. (Reading (5' → 3'))

After that, we could see the DNA sequence after the restriction sites were **deleted** by changing the base sites.



## ... and finally an **interface!**

What if we do not want to **hide** restriction site EcoRI, but only BavCI? That is simple! We go back, pick BavCI only, and run the program again.




In our final answer, the third nucleotide was **altered**. We can prove using a table that both ATA and ATC code isoleucine, and that is why our result is correct.

# Examples of inputs and outputs

The input DNA sequence contains an unknown number of restriction sites.

The programs detect three prohibited restriction sites: EcoRI, PstI, BsaI.

Then, by default, the program hides all restriction sites by altering nucleotide sequence within these sites.



The screenshot shows a web application window titled "Resulting DNA". The content is as follows:

The forward strand of given DNA sequence is GAATTCAAACCTGTTCTGCAGAACTCCATATGTGGAGGGGGTCTCCAC and reversed one is CTTAAGTTTGGACAAGACGTCTTGAGGTATACACCTCCCCCAGAGGTG

The selected restriction sites are GAATTC — EcoRI, TCTAGA — XbaI, ACTAGT — SpeI, CTGCAG — PstI, GCGGCCGC — NotI, GCTCTC — SapI, GGTCTC — BsaI.

CTTAAG named as EcoRI found at positions 0.	GAATTC named as EcoRI found at positions 0.
GACGTC named as PstI found at positions 15.	CTGCAG named as PstI found at positions 15.
CCAGAG named as BsaI found at positions 39.	GGTCTC named as BsaI found at positions 39.

The DNA sequences resulting after hiding these selected restrictions:

(5' – 3'): GAGTTCAAACCTGTTTTCAGAACTCCATATGTGGAGGGGGTCTCCAC

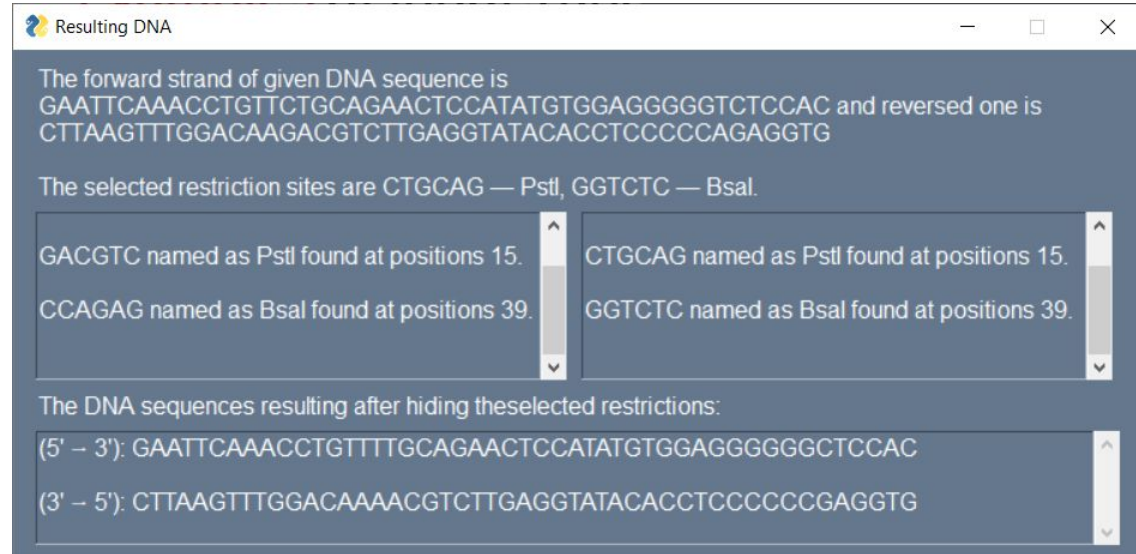
(3' – 5'): CTCAAGTTTGGACAAAACGTCTTGAGGTATACACCTCCCCCAGAGGTG

# Examples of inputs and outputs

The same DNA sequence contains three prohibited restrictions sites: EcoRI, PstI, BsaI (see the 20th slide).

This time, user prefers to hide only PstI, BsaI, but leave EcoRI unchanged. They select only PstI, BsaI from the main menu, willing to remove only those two. EcoRI is kept in the sequence (position 0).

The output contains the unchanged EcoRI site within the DNA sequence output.



# Examples of inputs and outputs

The given sequence of DNA does not contain any prohibited restriction site.

The program neither detects any nor changes the DNA sequence.



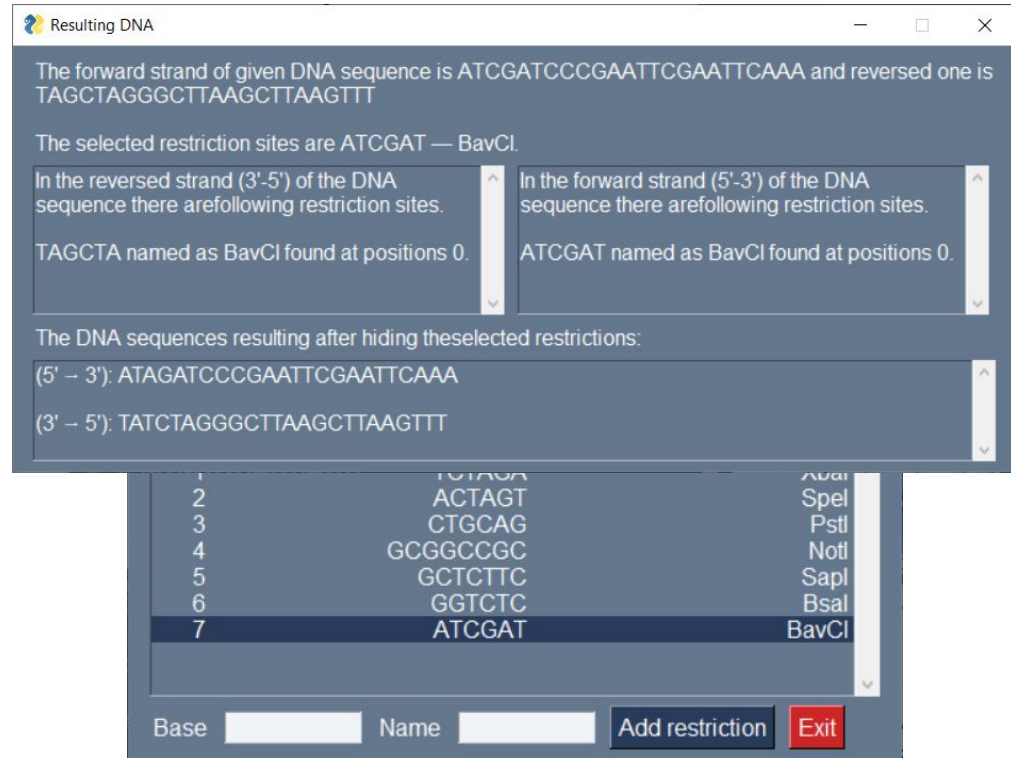


# Examples of inputs and outputs

The given sequence of DNA contains the restriction site not present in the main base of prohibited restriction sites.

The researcher may add any new restriction site (ATCGAT- BavCI) into the base, and the program will detect and alter it if that site is present within the sequence.

So its position is 0 in the DNA sequence and the output sequence has an altered site (ATAGAT), coding the same 2 amino acids (Ile,asp).



# Possible **modifications**

For future development of the program, we consider:

- Insert additional database, so user may also create their account to save preferable restriction sites to be removed
- Make visualization of restriction maps and demonstrate restriction bars within the strand
- Adapt algorithms for more complex genes organization similar to eukaryotes