

Exploration of Shared SNPs in Thaps

trunc-qfiltered

June 29, 2017

Rambling exploration of SNP positions shared between two or more of the isolates. Code is included to document it thoroughly, (even if largely uninteresting to anyone else), and I will summarize it as I go.

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1 History

This was added to SVN 1/26/2014; not sure when it was started, but earliest related emails I see are from 1/21/14.

```
r413 | ruzzo | 2014-01-26 08:22:37 -0800 (Sun, 26 Jan 2014) | 2 lines
adding shared-snp analysis.
```

2 Preliminaries

NOTE: Some comments in code and some parts of the text, especially specific numbers and general conclusions, are based on Unqfiltered, Chr1, Medium stringency (i.e., “[2]” below) analysis. The broad picture does not appear to change with other choices, but details do, and the text is neither fully parameterized nor fully updated, so proceed with caution.

Load utility R code; do setup:

```
source('.././../R/wlr.R') # load util code; path relative this folder or sibling in scripts/larrys

## Running as: ruzzo @ bicycle.cs.washington.edu; SVN Id, I miss you. $Id: wlr.R 2017-06-26 or later $

setup.my.wd('shared-snps') # set working dir; UPDATE if this file moves, or if COPY/PASTE to new file
setup.my.knitr('f-knitr/')
generic.setup('figs-mine/')
```

3 Major Analysis/Performance Parameters.

Choices here control how this file is processed, what data is analyzed, speed, etc. Set them carefully before running “make.” Major choices are:

1. WHICH SNP TABLES ARE LOADED??? The logical vector `load.tb` selects the desired combination of SNP tables to load, in the order `full.unfiltered`, `chr1.unfiltered`, `full.qfiltered`, `chr1.qfiltered`. E.g., `load.tb=(T, F, T, F)` loads *full* tables for *both* q- and un-qfiltered data. Primary analysis is only performed on one of them, but the others are retained for comparison/debugging.
2. WHICH MAIN ANALYSIS??? If multiple tables are loaded, which is used for the main analysis? Parameter `pri` is a permutation of 1:4, corresponding to `load.tb`; the first loaded table in that order becomes the analysis focus. The default `pri=c(1, 2, 3, 4)` looks at un-q-filtered data in preference to q-filtered, and full tables in preference to Chr1 within each group.
(Choice of data for the “Table 1” coverage summary in section 5 is independent of this; full genome data is preferred over Chr 1 for both q- and unq-filtered reads; change `tset.picker` calls near the end of that section to modify this.)
3. CLEAR CACHE??? `clear.cache=T` forces Knitr cache removal at the start of the run; especially important if the previous parameters have changed since the last run.
4. HOW MANY BOOTSTRAP REPLICATES??? The variable `nboot` is a major performance factor; 1000 reps takes several hours. Set to 5 for debug and quick look; 100 or more for final run.
5. TRUNCATE TABLES TO Chrs ONLY??? I.e., remove mitochondrial-, plastid-, and BD- contigs.

The following code chunk sets the first four parameters based on where it’s run. To prototype/debug on a laptop, faster is better—run on Chr1 with small `nboot`; when run on the linux servers, I typically do full genomes, more replicates. Just override them if these defaults don’t work for you.

```

# for Makefile, params can be command line args, else base on system; see wlr.r for details.
# load.tb order: full.un, chr1.un, full.qfil, chr1.qfil

params <- pick.params(
  mac = list(load.tb=c(F,T,F,F), pri=1:4, clear.cache=F, nboot= 1, trunc.tables=T), # quick on lap
  linux = list(load.tb=c(F,F,F,T), pri=1:4, clear.cache=F, nboot= 5, trunc.tables=T), # quick qfil on server
  linux = list(load.tb=c(T,F,T,F), pri=1:4, clear.cache=T, nboot=101, trunc.tables=T) # full on server
)

# Alternatively, edit/uncomment the following to override the above as needed
#params<-pick.params(default=list(load.tb=c(T,T,T,T),pri=1:4,clear.cache=T,nboot=1000,trunc.tables=T))
print(params)

# $load.tb
# full.unf chr1.unf full.qf chr1.qf
# TRUE FALSE TRUE FALSE
#
# $pri
# [1] 3 4 1 2
#
# $clear.cache
# [1] TRUE
#
# $nboot
# [1] 101
#
# $trunc.tables
# [1] TRUE

```

CLEAR CACHE??!! Some code chunks use the knitr cache, but extent of cache consistency checks unknown. If in doubt, delete “cache/” (knitr’s) directory to force rebuild. T/F set in params above will/won’t force removal (actually, rename):

```

decache(params$clear.cache)

# Rename of 'cache' to 'cache80604' returned TRUE .

```

If still in doubt, also manually remove “00common/mycache/” (mine).

Load the main SNP data file(s) based on the parameters set in section 3.

```

# short names to keep the following chunk compact
tb <- params$load.tb
tset <- list(NULL, NULL, NULL, NULL) # tset = 'table set'

# see wlr.R for load paths
if(tb[1]){tset[[1]] <- load.snp.tables(use.chr1.tables = FALSE, data.name='full.tables.01.26.14')}

# Loading full tables from ../../../../data/ungit-data/full.tables.01.26.14.rda ...Loaded.
# ../00common/mycache/snp.tables.chr1.unqfiltered.rda saved.

if(tb[2]){tset[[2]] <- load.snp.tables(use.chr1.tables = TRUE, data.name='full.tables.01.26.14')}
if(tb[3]){tset[[3]] <- load.snp.tables(use.chr1.tables = FALSE, data.name='full.tables.02.25.15')}

# Loading full tables from ../../../../data/ungit-data/full.tables.02.25.15.rda ...Loaded.
# ../00common/mycache/snp.tables.chr1.qfiltered.rda saved.
# Bandaiding qfiltered tables...

if(tb[4]){tset[[4]] <- load.snp.tables(use.chr1.tables = TRUE, data.name='full.tables.02.25.15')}

```

Grrr! I should have excluded non-Chr contigs from full genome runs. Rather than change tons of code below to add mask params, I’m just going to truncate the tables, as follows. (See notes in wlr.r::make.mask for assumptions.)

```

if(params$trunc.tables){
  for(i in 1:4){
    if(!is.null(tset[[i]])){
      first.mito <- match("mitochondria.fasta", tset[[i]][[7]]$Chr)
      if(!is.na(first.mito)){ # will be NA for Chr1 tables
        for(j in 1:7){
          # hmmm... slow; wonder whether head(tset[[i]][[j]],first.mito-1) is faster;
          # ok, simple tests suggest not: system.time(head(data.frame(1:1e7,1:1e7),5e6))
          tset[[i]][[j]] <- tset[[i]][[j]][1:(first.mito-1),]
        }
      }
    }
  }
} else {
  cat('***\n*** DID YOU *REALLY* WANT UNTRUNCATED TABLES???\n***\n')
}

```

The tersely-named `tset` list is sometimes convenient, but give them more descriptive names, too.

```

snp.tables.full.unfiltered <- tset[[1]]; names(tset)[1] <- 'snp.tables.full.unfiltered'
snp.tables.chr1.unfiltered <- tset[[2]]; names(tset)[2] <- 'snp.tables.chr1.unfiltered'
snp.tables.full.qfiltered <- tset[[3]]; names(tset)[3] <- 'snp.tables.full.qfiltered'
snp.tables.chr1.qfiltered <- tset[[4]]; names(tset)[4] <- 'snp.tables.chr1.qfiltered'

```

Main analysis may just use one of the potentially 4 table sets. Pick it according to the priority specified in section 3, using the shorter name 'snp.tables' for this default choice.

```
snp.tables <- tset.picker(priority=params$pri, table.set=tset)
```

```

# Sanity check: unlike unfiltered tables, bug in early code gave qfiltered ones different numbers
# of rows per strain, which breaks much code. Verify this is no longer happening.
check.eq.nrows <- function(tables){
  if(!is.null(tables)){
    nrow.snp.tables <- unlist(lapply(tables,nrow))
    print(nrow.snp.tables)
    if(all(nrow.snp.tables == nrow.snp.tables[1])){
      cat('OK, all strains have same number of rows.\n')
    } else {
      cat('***\n*** Warning: Different strains have different numbers of rows! ***\n***\n')
    }
  }
}

dummy<-lapply(tset, check.eq.nrows)

#      1007      1012      1013      1014      1015      3367      1335
# 31301782 31301782 31301782 31301782 31301782 31301782 31301782
# OK, all strains have same number of rows.
#      1007      1012      1013      1014      1015      3367      1335
# 31301782 31301782 31301782 31301782 31301782 31301782 31301782
# OK, all strains have same number of rows.

```

Which tables have we got?:

```

# 'which.snp.tables' return summary of which tables, either as a char string (default), e.g.
# "Chr1-qfiltered", or as vector of 2 strings, e.g. c("full","unfiltered").
cat('This analysis uses: (', paste(unlist(lapply(tset,which.snp.tables)),collapse=', '), ') SNP tables.\n')

# This analysis uses: ( trunc-unfiltered, NULL, trunc-qfiltered, NULL ) SNP tables.

cat('Main shared SNP analysis focuses on', which.snp.tables(snp.tables), '\n')

# Main shared SNP analysis focuses on trunc-qfiltered

```

A L^AT_EX hack: I want which.snp.tables info in doc title/page headers, but it is unknown until now, so the following writes a command definition `\whichsnptables` into the .aux file, which is read during the *next* L^AT_EX run, when `\begin{document}` is processed:

```
\makeatletter
\immediate\write\@auxout{\noexpand\gdef\noexpand\whichsnptables{trunc-qfiltered}}
\makeatother
```

Subsequent analysis was initially all directed at Chr1. In general, I have *not* updated the discussion to reflect genome-wide analysis.

```
if(exists('snp.tables.chr1.qfiltered') && exists('snp.tables.chr1.unqfiltered')){
  # If have both, where is new unequal to old?
  uneq <- snp.tables.chr1.qfiltered[[1]]$Ref[1:chr1.len] != snp.tables.chr1.unqfiltered[[1]]$Ref[1:chr1.len]
  cat('Sum uneq:', sum(uneq, na.rm=T), '\n')
  cat('Sum NA: ', sum(is.na(uneq)), '\n')
  print(which(is.na(uneq)) [1:10])
  seecounts(which(is.na(uneq)) [1:4], snp.tables=snp.tables.qfiltered, debug=F)
}
```

In brief, “snp.tables” will be a list of 7 data frames, one per strain, giving read counts for each nucleotide at each position, SNP calls, etc.:

```
names(snp.tables)

# [1] "1007" "1012" "1013" "1014" "1015" "3367" "1335"

str(snp.tables[[1]])

# 'data.frame': 31301782 obs. of 16 variables:
# $ snp : int 0 0 0 0 0 0 0 0 0 0 ...
# $ chr : chr "Chr1" "Chr1" "Chr1" "Chr1" ...
# $ Pos : int 1 2 3 4 5 6 7 8 9 10 ...
# $ Ref : chr "T" "C" "C" "A" ...
# $ Cov : num 0 2 3 4 4 4 7 8 9 10 ...
# $ a : num 0 0 0 0 0 0 0 0 0 0 ...
# $ g : num 0 0 0 0 0 0 0 0 0 0 ...
# $ c : num 0 0 0 0 0 0 0 0 0 0 ...
# $ t : num 0 0 0 0 0 0 0 0 0 0 ...
# $ n : num 0 0 0 0 0 0 0 0 0 0 ...
# $ .match: num 0 2 3 4 4 4 7 8 9 10 ...
# $ exon : logi FALSE FALSE FALSE FALSE FALSE ...
# $ indel : logi FALSE FALSE FALSE FALSE FALSE ...
# $ chr : Factor w/ 66 levels "BD1_7","BD10_65",...: 39 39 39 39 39 39 39 39 39 39 ...
# $ pos : int 1 2 3 4 5 6 7 8 9 10 ...
# $ rawCov: num 1 3 4 5 7 7 10 12 13 15 ...
```

Just for background, also load the desert tables:

```
# from svn+ssh://cegl.ocean.washington.edu/var/svn/7_strains/trunk/code/snpNB/data
#load('../.../data/ungit-data/des.rda')
load('../.../data/des.rda')
```

What’s the total length of all deserts in each strain? Big deserts (defined as “big.threshold” or longer)?

```
some.desert.stats <- function(big.threshold=0){
  desert.len <- unlist(lapply(des, function(x){sum(unlist(lapply(x, function(y){sum(y[, 'Length'])))})))
  bigdes.len <- unlist(lapply(des, function(x){sum(unlist(lapply(x, function(y){
    sum(y[y[, 'Length']>=big.threshold, 'Length'])))})))
  rbind(desert.len, desert.pct=round( desert.len / genome.length.constants()$genome.length.trunc * 100),
    bigdes.len, bigdes.pct=round( bigdes.len / genome.length.constants()$genome.length.trunc * 100))
}
some.desert.stats(big.threshold=50000)
```

#	tp1007	tp1012	tp1013	tp1014	tp1015	thapsIT	tp1335
# desert.len	11146526	11332566	5801763	9464213	11251426	6780300	10883723
# desert.pct	36	36	19	30	36	22	35
# bigdes.len	3495805	3936973	55365	3627235	3727061	57119	4046934
# bigdes.pct	11	13	0	12	12	0	13

I.e., looking at all deserts, about 1/3 of L-clade, 1/5 of H-clade are in deserts, whereas, looking at the largest deserts ($> 50k$), only about 12% in L-clade (and none in H-clade). Note that the rough stats above include artifactual “deserts” created by gaps in the reference sequence, large genomic deletions, etc. A more careful analysis of this is found in nc-snps.rnw.

4 Refined SNP Calls

4.1 Method

It is appropriate that SNP calls should be conservative, to avoid many false positives, but, when a position is called a SNP in one isolate, we often see a significant number of reads for the same non-reference nucleotide at that position in other isolates, even if they are not called as SNPs. On the other hand, we sometimes see a position called a SNP in two or more isolates, but with *different* pairs of nucleotides, potentially suggesting technical errors. Analysis in this section attempts to refine the SNP calls by looking for issues such as these by looking at all 7 isolates jointly, at each position called a SNP in any of them.

For a given strain, the following function returns a vector of 0:4 to indicate which nonreference nucleotide has the maximum read count at the corresponding position. The values 1..4 indicate that the max count occurred at A, G, C, T, resp. (Ties are resolved arbitrarily ($a < g < c < t$), which possibly deserves further attention.) The value 0 means all nonreference counts are below threshold, based *either* on absolute count *or* as a fraction of coverage. Default only excludes 0 counts.

```
nref.nuc.new <- function(strain=1, mask=T, thresh.count=0, thresh.rate=0.0){
  # get read count for max nonref nuc
  nref <- apply(snp.tables[[strain]][mask, c('a', 'g', 'c', 't')], 1, max)
  # where does nref count match a (g,c,t, resp) count
  as <- ifelse(nref == snp.tables[[strain]][mask, 'a'], 1, 0)
  gs <- ifelse(nref == snp.tables[[strain]][mask, 'g'], 2, 0)
  cs <- ifelse(nref == snp.tables[[strain]][mask, 'c'], 3, 0)
  ts <- ifelse(nref == snp.tables[[strain]][mask, 't'], 4, 0)
  # most positions will show 3 zeros and one of 1:4, so max identifies max nonref count;
  # ties broken arbitrarily (a<g<c<t)
  merge <- pmax(as, gs, cs, ts)
  # but if max nonref count is zero or below threshold, return 0
  merge[nref == 0 | nref < thresh.count] <- 0
  merge[nref/snp.tables[[strain]][mask, 'Cov'] < thresh.rate] <- 0
  return(merge)
}
```

Get union and intersection of the sets of called SNPs. (“\$snp” is 0/1.) Also, 5-way (L-clade) and 4-way (L-excluding Gyre).

```
# 4-way union/intersection
u4.snps <- snp.tables[[1]]$snp
i4.snps <- snp.tables[[1]]$snp
for(i in c(2,5,7)) {
  u4.snps <- pmax(u4.snps, snp.tables[[i]]$snp)
  i4.snps <- pmin(i4.snps, snp.tables[[i]]$snp)
}
# 5-way: add gyre
u5.snps <- pmax(u4.snps, snp.tables[[4]]$snp)
i5.snps <- pmin(i4.snps, snp.tables[[4]]$snp)
# 7-way
union.snps <- pmax(u5.snps, snp.tables[[3]]$snp, snp.tables[[6]]$snp)
intersect.snps <- pmin(i5.snps, snp.tables[[3]]$snp, snp.tables[[6]]$snp)
nu4snps <- sum(u4.snps)
```

```

nu5snps <- sum(u5.snps)
ni4snps <- sum(i4.snps)
ni5snps <- sum(i5.snps)
nusnps <- sum(union.snps)
nisnps <- sum(intersect.snps)
c(n4u=nu4snps, n5u=nu5snps, n7u=nusnps, n4i=ni4snps, n5i=ni5snps, n7i=nisnps)

#      n4u      n5u      n7u      n4i      n5i      n7i
# 196296 197799 474613 128683  70687  15186

```

There are nusnps=474613 positions called as SNPs in one or more strains (but only nisnps=15186 that are shared among all 7). Note that the 4-way union is only modestly larger (1.5254229 times larger) than the 4-way intersection, emphasizing the inherent similarities among these SNP sets. The corresponding 5-way numbers show that Gyre adds relatively little to the 5-way union vs the 4-way union, whereas it removes a fair bit from the 5-way intersection. However, much of that loss is simply because Gyre has fewer called SNPs: only 89184 vs 128683 in the 4-way intersection, and they are highly concordant:

```

sum(snp.tables[[4]]$snp*i4.snps)/sum(snp.tables[[4]]$snp)

# [1] 0.7925973

```

So, a likely source of the Gyre's difference in called SNPs is technical (lower read coverage, higher read error rate) rather than biological.

Inclusion of the 2 H-clade members, however, causes more dramatic changes in both union and intersection numbers. I examine all these relationships in more detail below, but first I examine what I believe to be a significant source of technical error in these comparisons—erroneous SNP calls, especially false negative calls.

It is appropriate that SNP calls should be conservative, to avoid many false positives, but, if a position is called a SNP in one strain, we often see a significant number of reads for the same non-reference nucleotide at that position in other strains, even if they are not called as SNPs. For my purposes below, these will be considered “shared SNPs,” based on three different levels of permissiveness. Note that, e.g., $\geq 84\%$ of all positions have zero reads for any non-reference nucleotide, and only a small fraction have 2 or more non-reference reads:

```

nonmatch <- rbind(
  unlist(lapply(snp.tables,function(x){sum(x$Cov-x$.match == 0)})),
  unlist(lapply(snp.tables,function(x){sum(x$Cov-x$.match == 1)})),
  unlist(lapply(snp.tables,function(x){sum(x$Cov-x$.match == 2)})),
  unlist(lapply(snp.tables,function(x){sum(x$Cov-x$.match == 3)})),
  unlist(lapply(snp.tables,function(x){sum(x$Cov-x$.match >= 4)})),
  unlist(lapply(snp.tables,function(x){sum((x$Cov-x$.match)[union.snps==0] >= 4)}))
)/nrow(snp.tables[[1]])*100
rownames(nonmatch) <- c('% ==0', '% ==1', '% ==2', '% ==3', '% >=4', '% >=4, nonSNP')
nonmatch

#           1007           1012           1013           1014           1015           3367           1335
# % ==0      97.71716831  97.35791400  95.45329400  97.29079003  97.18569697  95.89943474  96.48734376
# % ==1       1.48140448   1.75279158   3.01610304   2.08080805   1.88427930   2.58814338   2.54747477
# % ==2       0.11277633   0.12101867   0.22209918   0.18711714   0.13599226   0.21703876   0.19842640
# % ==3       0.05885927   0.05629072   0.09496264   0.10621121   0.06021063   0.09526295   0.06375675
# % >=4       0.62979162   0.71198502   1.21354113   0.33507358   0.73382084   1.20012017   0.70299831
# % >=4, nonSNP 0.08015518  0.12493857  0.25313575  0.04763946  0.13208513  0.28212771  0.13240460

```

Build a table of max non-reference nucleotides at each position in the union.snps set. The three criteria are

- [[1]]: any non-zero count at any coverage is considered significant
- [[2]]: (count ≥ 2 and count/coverage ≥ 0.05) is considered significant
- [[3]]: (count ≥ 4 and count/coverage ≥ 0.10) is considered significant

In all three cases, the nonref nucleotide must also be consistent across all strains passing that threshold; see below.

```

non.refs <- vector('list',4)
for(i in 1:4){
  non.refs[[i]] <- matrix(0, nrow=nusnps, ncol=7)
  colnames(non.refs[[i]]) <- names(snp.tables)
  rownames(non.refs[[i]]) <-
    paste(snp.tables[[1]]$chr[union.snps==1], ': ', snp.tables[[1]]$pos[union.snps==1], sep='')
}
for(j in 1:7){
  non.refs[[1]][,j] <- nref.nuc.new(j, mask=union.snps==1, thresh.count=0, thresh.rate=0.00)
  non.refs[[2]][,j] <- nref.nuc.new(j, mask=union.snps==1, thresh.count=2, thresh.rate=0.05)
  non.refs[[3]][,j] <- nref.nuc.new(j, mask=union.snps==1, thresh.count=4, thresh.rate=0.10)
}

```

For comparison, I want to look at unfiltered SAMTools SNP calls. In complete opposition to the measures of consistency imposed above, I'm going to simply force this into the “non.refs” structure constructed above by imagining that any position called a SNP in any strain has its max nonref count on “A”, so any given position called a SNP in any strain will automatically be declared “consistent.” This will allow the tree-code, etc. given below to work in a uniform way (even though interpretation of the results is different.) Results will be jammed into a 4th component of the “non.refs” list; i.e., we have a 4th criterion:

- [[4]]: all called SNPs at a given position are considered “consistent.”

As this case was a late addition to the analysis, the commentary throughout this document has not necessarily been updated to reflect that this case is distinct from the first three.

```

for(j in 1:7){
  non.refs[[4]][,j] <- snp.tables[[j]]$snp[union.snps==1]
}

```

```

str(non.refs[[4]])

# num [1:474613, 1:7] 0 0 0 0 0 0 0 1 0 ...
# - attr(*, "dimnames")=List of 2
# ..$ : chr [1:474613] "Chr1:333" "Chr1:417" "Chr1:435" "Chr1:438" ...
# ..$ : chr [1:7] "1007" "1012" "1013" "1014" ...

```

“non.refs” indicates, among those positions in the union of all called SNPS having any non-reference read count above the thresholds listed above, the non-ref nucleotide having the highest read count in each strain. If, for a given position, the max of this code is the same as the min (among non-zero values), then every strain having over-threshold nonref reads in that position, in fact has most non-reference reads on the *same* nucleotide. These are defined as the “consistent” SNPs.

```

find.consistent <- function(nr){
  nr.max <- apply(nr,1,max)
  nr.min <- apply(nr,1,function(x){ifelse(max(x)==0,0,min(x[x>0]))})
  return(nr.min == nr.max)
}
consistent <- lapply(non.refs, find.consistent)

```

4.2 Save them

```

# wrap this in a data structure to be cached:
Description <- [2757 chars quoted with '']

filtered.snps <-
  list(Description=Description,

        Data=list(
          based.on.which.snp.tables=which.snp.tables(),
          number.union.snps=nusnps,
          number.intersection.snps=nisnps,

```



```

non.ref.nucleotide=non.refs,
consistent.snps=consistent),

Code=list(
  get.snps = function(strain, stringency=2){
    # return nusnps x 1 Bool vector of consistent SNPs @ specified stringency & strain
    return(filtered.snps$Data$consistent.snps[[stringency]] &
           filtered.snps$Data$non.ref.nucleotide[[stringency]][,strain] > 0)
  },
  get.snp.locs.char = function(strain, stringency=2){
    # return char vector of locations of consistent SNPs @ specified stringency & strain
    snps <- filtered.snps$Code$get.snps(strain, stringency)
    return(names(snps)[snps])
  },
  get.snp.locs.df = function(strain, stringency=2){
    # return data frame (Chr/Pos) of locations of consistent SNPs @ specified stringency & strain
    snplist <- strsplit(filtered.snps$Code$get.snp.locs.char(strain, stringency), ':', fixed=TRUE)
    # strsplit returns long list of 2-vectors, 1st=chr, 2nd=char position
    df <- data.frame(Chr=unlist(lapply(snplist,function(x){return(x[1])})),
                     Pos=as.integer(unlist(lapply(snplist,function(x){return(x[2])}))),
                     stringsAsFactors = FALSE)

    return(df)
  }
)

# dont't clobber existing .rda, but save if absent. (delete to re-save)
rda.filtered <- paste('../00common/mycache/filtered.snps', which.snp.tables(), 'rda', sep='.')
if(file.exists(rda.filtered)){
  cat('Pre-existing file', rda.filtered, 'unchanged.\n')
} else {
  cat('Saving', rda.filtered, '...\n')
  save(filtered.snps, file=rda.filtered, compress=TRUE)
  cat('Saved.\n')
}

# Pre-existing file ../00common/mycache/filtered.snps.trunc-qfiltered.rda unchanged.

```

Knitr seems to be failing to format the long char string above, which says:

```

cat(filtered.snps$Description)

# Contents of this .rda file:
#
# * Description: this text
#
# * Data -- 5 items defining filtered SNPs, at 4 different stringency levels, as defined
#   in shared-snps.rnw:
#
# * based.on.which.snp.tables: {"Chr1","full","trunc"}-{"unfiltered","qfiltered"},
#   depending on which snp tables were used to build this data. ("trunc" = all Chrs.)
#
# * number.union.snps: the total number of SNPs (SAMtools calls) in the union of SNPs
#   across all 7 strains.
#
# * number.intersection.snps: similar, for the 7-way intersection.
#
# nusnps/nisnps are easily recalculated from the data below, but their inclusion
# may be convenient, e.g., to quickly see if the .rda represents the full genome
# (nusnps=488848), or the chr 1 subset (nusnps=47499); (redundant with "based.on...";
# numbers above are for unfiltered, perhaps slightly different if qfiltered)
#
# * non.ref.nucleotide: 4 arrays, each nusnps x 7, of values 0..4 (0..1 in the 4th
#   array). In the 1st 3 arrays, 0 means the given position in the given strain did
#   not have nonreference read counts above the corresponding filtering threshold,

```

```
# i.e., is NOT a filtered SNP in that strain, whereas 1.4 mean that it did pass
# threshold, for A,C,G,T resp. In the 4th array, this value is just 1/0,
# indicating is/is not a called SNP in that strain.
#
# * consistent.snps: 4 Bool vectors of length nusnps flagging positions whose nonref
# nucs (wrt to the 4 filtering criteria) are deemed *consistent* across
# all 7 strains. For the 1st 3, this means all nonzero entries of non.ref.nuc
# are equal, i.e., nonref read counts passing threshold are on the SAME nonref
# nucleotide in all strains having over-threshold counts. Just for comparison
# and uniformity of data structures, the 4th is all TRUE, i.e., union of SNPs
# across all strains, without any regard for thresholds or consistency.
#
# In short, the filtered SNPs according to our medium filtering criteria are
# strains/positions where consistent.snps[[2]]==TRUE and non.ref.nucleotide[[2]]>0.
#
# Rownames in both non.ref.nucs and consistent define location, e.g. "Chr1:333".
#
# * Code -- simple routines to extract filtered SNPs in (potentially) convenient formats:
#
# * get.snps(strain, stringency=2)
# returns nusnps x 1 Bool vector of consistent SNPs @ specified stringency in
# given strain
#
# * get.snp.locs.char(strain, stringency=2)
# returns n x 1 char vector of locations of consistent SNPs @ specified stringency
# in given strain, e.g. "Chr1:1234", where n == sum(get.snps(...))
#
# * get.snp.locs.df(strain, stringency=2){
# As above, but returns data frame (char vector Chr, int vector Pos) with the same info.
```

```
str(consistent[[1]])

# Named logi [1:474613] TRUE FALSE TRUE TRUE TRUE TRUE ...
# - attr(*, "names")= chr [1:474613] "Chr1:333" "Chr1:417" "Chr1:435" "Chr1:438" ...
```

```
consistent.count <- unlist(lapply(consistent, sum)) ; consistent.count

# [1] 447177 469906 471171 474613

inconsistent.count <- consistent.count[4] - consistent.count; inconsistent.count

# [1] 27436 4707 3442 0

inconsistent.percent <- inconsistent.count/consistent.count[4]*100; inconsistent.percent

# [1] 5.7807098 0.9917554 0.7252224 0.0000000
```

I.e., of the 474613 positions in which a SNP is called, 447177 are consistent by my loose definition, and 471171 are consistent by my tightest definition. The increase in concordance supports the view that the loose definition is too loose. Perhaps misleadingly, these counts include positions that are “consistent SNPs” in only one strain; more below. (*TODO* I suspect, but have not yet systematically checked, that most of the rest are positions with low coverage and/or very low read counts on the mixture of non-reference nucleotides.)

4.3 Examples: Consistent

Here are a few (nonrandomly selected) prototypical consistent SNPs:

```
esnps <- names(consistent[[2]][consistent[[2]]])
esnps2 <- as.integer(unlist(lapply(strsplit(esnps[c(7,11:13,92)], ':', fixed=TRUE), function(x){x[2]})))
seecounts(esnps2, snp.tables=snp.tables)

# chr pos Ref Strain A G C T SNP exon indel nrf rat
# 1 Chr1 567 T
# 2 1007 0 0 1 25 0 TRUE FALSE
```

```

# 3      1012 0 0 14 39 1 TRUE FALSE
# 4      1013 0 0 13 87 0 TRUE FALSE
# 5      1014 0 1 0 23 0 TRUE FALSE
# 6      1015 0 0 8 40 1 TRUE FALSE
# 7      3367 0 0 16 38 1 TRUE FALSE
# 8      1335 0 0 2 99 0 TRUE FALSE
# 9 Chr1 1053 A
# 10     1007 25 0 0 4 0 TRUE FALSE
# 11     1012 35 0 0 12 0 TRUE FALSE
# 12     1013 2 1 0 32 0 TRUE FALSE
# 13     1014 5 0 0 5 0 TRUE FALSE
# 14     1015 29 0 0 15 1 TRUE FALSE
# 15     3367 2 0 0 7 0 TRUE FALSE
# 16     1335 56 0 0 39 1 TRUE FALSE
# 17 Chr1 1055 G
# 18     1007 0 23 0 1 0 TRUE FALSE
# 19     1012 0 37 0 6 0 TRUE FALSE
# 20     1013 1 39 0 6 0 TRUE FALSE
# 21     1014 0 6 0 2 1 TRUE FALSE
# 22     1015 0 26 0 14 0 TRUE FALSE
# 23     3367 0 12 0 0 0 TRUE FALSE
# 24     1335 0 54 0 32 1 TRUE FALSE
# 25 Chr1 1176 G
# 26     1007 1 53 0 0 0 FALSE FALSE
# 27     1012 0 54 0 0 0 FALSE FALSE
# 28     1013 19 56 0 0 0 FALSE FALSE
# 29     1014 0 28 0 0 0 FALSE FALSE
# 30     1015 3 85 0 0 0 FALSE FALSE
# 31     3367 9 2 0 0 1 FALSE FALSE
# 32     1335 0 156 0 0 0 FALSE FALSE
# 33 Chr1 8685 G
# 34     1007 6 15 0 0 0 TRUE FALSE
# 35     1012 10 23 0 0 0 TRUE FALSE
# 36     1013 18 21 0 0 1 TRUE FALSE
# 37     1014 4 8 0 0 0 TRUE FALSE
# 38     1015 10 24 0 0 1 TRUE FALSE
# 39     3367 0 4 0 0 0 TRUE FALSE
# 40     1335 5 32 0 0 0 TRUE FALSE

```

4.4 Examples: Inconsistent

Here is a brief look at some *in*-consistent positions. E.g., Chr1:2013 shows nontrivial counts on 3 alleles in Wales, as do 2319, 3286, 5002, 5433, whereas 7878 shows a different alternate allele in Italy than in Wales.

```

unc <- names(consistent[[2]][!consistent[[2]]])
unc2 <- as.integer(unlist(lapply(strsplit(unc[1:10], ':', fixed=TRUE), function(x){x[2]})))
seecounts(unc2, snp.tables=snp.tables)

```

```

#      chr   pos Ref Strain  A   G   C   T SNP  exon indel nrf rat
# 1  Chr1  2013   T
# 2      1007 4 0 0 15 0 TRUE FALSE
# 3      1012 6 0 0 21 0 TRUE FALSE
# 4      1013 7 10 0 6 1 TRUE FALSE
# 5      1014 1 0 0 6 0 TRUE FALSE
# 6      1015 13 0 0 13 1 TRUE FALSE
# 7      3367 7 0 0 25 0 TRUE FALSE
# 8      1335 16 0 0 42 1 TRUE FALSE
# 9  Chr1  2319   C
# 10     1007 0 28 10 0 1 TRUE FALSE
# 11     1012 0 43 17 0 1 TRUE FALSE
# 12     1013 13 15 9 0 1 TRUE FALSE
# 13     1014 0 18 6 0 1 TRUE FALSE
# 14     1015 0 53 20 0 1 TRUE FALSE
# 15     3367 4 0 24 0 0 TRUE FALSE
# 16     1335 0 118 28 0 1 TRUE FALSE

```

# 17	Chr1	3286	T								
# 18				1007	4	0	1	10	0	TRUE	FALSE
# 19				1012	7	0	3	32	0	TRUE	FALSE
# 20				1013	34	0	30	1	1	TRUE	FALSE
# 21				1014	4	0	4	10	0	TRUE	FALSE
# 22				1015	11	0	6	31	0	TRUE	FALSE
# 23				3367	5	0	29	0	0	TRUE	FALSE
# 24				1335	14	0	3	55	0	TRUE	FALSE
# 25	Chr1	5002	T								
# 26				1007	0	14	0	7	0	TRUE	FALSE
# 27				1012	0	20	0	19	1	TRUE	FALSE
# 28				1013	18	10	0	22	0	TRUE	FALSE
# 29				1014	0	5	0	2	0	TRUE	FALSE
# 30				1015	0	18	0	12	1	TRUE	FALSE
# 31				3367	0	0	0	31	0	TRUE	FALSE
# 32				1335	0	46	0	44	0	TRUE	FALSE
# 33	Chr1	5178	G								
# 34				1007	0	20	0	0	0	TRUE	FALSE
# 35				1012	0	32	0	0	0	TRUE	FALSE
# 36				1013	47	9	0	0	1	TRUE	FALSE
# 37				1014	0	13	0	0	0	TRUE	FALSE
# 38				1015	0	30	0	0	0	TRUE	FALSE
# 39				3367	32	19	0	0	1	TRUE	FALSE
# 40				1335	0	38	0	2	0	TRUE	FALSE
# 41	Chr1	5433	G								
# 42				1007	0	40	0	3	0	TRUE	FALSE
# 43				1012	0	53	0	5	0	TRUE	FALSE
# 44				1013	16	29	0	7	1	TRUE	FALSE
# 45				1014	9	8	0	0	1	TRUE	FALSE
# 46				1015	6	53	0	2	0	TRUE	FALSE
# 47				3367	8	37	0	0	0	TRUE	FALSE
# 48				1335	6	72	0	2	0	TRUE	FALSE
# 49	Chr1	7858	C								
# 50				1007	0	0	42	0	0	TRUE	FALSE
# 51				1012	0	0	35	0	0	TRUE	FALSE
# 52				1013	0	0	81	8	0	TRUE	FALSE
# 53				1014	0	0	12	0	0	TRUE	FALSE
# 54				1015	0	0	71	0	0	TRUE	FALSE
# 55				3367	20	0	2	0	1	TRUE	FALSE
# 56				1335	0	0	83	0	0	TRUE	FALSE
# 57	Chr1	8974	C								
# 58				1007	0	1	5	0	0	TRUE	FALSE
# 59				1012	0	2	13	0	0	TRUE	FALSE
# 60				1013	9	15	2	0	1	TRUE	FALSE
# 61				1014	0	1	1	0	0	TRUE	FALSE
# 62				1015	0	1	9	0	0	TRUE	FALSE
# 63				3367	2	0	1	0	0	TRUE	FALSE
# 64				1335	0	11	30	0	0	TRUE	FALSE
# 65	Chr1	10099	T								
# 66				1007	16	0	0	24	0	TRUE	FALSE
# 67				1012	45	0	0	26	0	TRUE	FALSE
# 68				1013	0	2	6	55	0	TRUE	FALSE
# 69				1014	32	0	0	11	0	TRUE	FALSE
# 70				1015	38	0	0	37	0	TRUE	FALSE
# 71				3367	0	1	0	7			

4.5 Examples: Homozygous nonref

And at some *homozygous nonreference* positions (defined to be those with nonref fraction > 0.75):

```
hnr <- lapply(snp.tables, function(x){x$.match/x$.Cov < 0.25}) # find them
hnr <- lapply(hnr, function(x){ifelse(is.na(x), FALSE, x)}) # remove NA
unlist(lapply(hnr, sum)) # count per strain

# 1007 1012 1013 1014 1015 3367 1335
# 16069 14356 120037 11436 6862 142515 1854
```

Hmm, in L-clade, excluding the ref isolate (1335) this tracks time-in culture to some degree; Maybe many of these are in hemizygous regions. Next two chunks lifted from nc-snp to get tables for hemi-deletion.

```
cnv.chrononly <- load.cnv.tables('.../data/cnv.txt', chrs.only=TRUE)

str(cnv.chrononly)

# 'data.frame': 1956 obs. of 11 variables:
# $ strain : Factor w/ 7 levels "IT","tp1007",...: 3 3 3 3 3 3 3 3 3 ...
# $ chr : Factor w/ 65 levels "BD1_7","BD10_65",...: 38 38 38 38 38 38 38 38 38 ...
# $ start : int 10601 112001 215001 358901 536501 554801 673401 781801 806901 853201 ...
# $ end : int 13500 116500 221100 370300 538600 559300 685000 787400 811100 855600 ...
# $ length : int 2900 4500 6100 11400 2100 4500 11600 5600 4200 2400 ...
# $ filtered : logi FALSE FALSE FALSE TRUE FALSE FALSE ...
# $ type : Factor w/ 1 level "CNVnator": 1 1 1 1 1 1 1 1 1 1 ...
# $ cov_ratio: num 0.63738 1.54893 1.65381 0.00204 0.68486 ...
# $ dup_frac : num 0.41188 0.00908 0.01178 0.97997 0.0211 ...
# $ iStart : num 10601 112001 215001 358901 536501 ...
# $ iEnd : num 13500 116500 221100 370300 538600 ...

cnv.chrononly[c(1:4,nrow(cnv.chrononly)+c(-1,0)),] ## first/last few rows

# strain chr start end length filtered type cov_ratio dup_frac iStart iEnd
# 1 tp1012 Chr1 10601 13500 2900 FALSE CNVnator 0.63738000 0.41187900 10601 13500
# 2 tp1012 Chr1 112001 116500 4500 FALSE CNVnator 1.54893000 0.00907677 112001 116500
# 3 tp1012 Chr1 215001 221100 6100 FALSE CNVnator 1.65381000 0.01178470 215001 221100
# 4 tp1012 Chr1 358901 370300 11400 TRUE CNVnator 0.00204431 0.97997300 358901 370300
# 1955 tp1335 Chr24 259901 278000 18100 FALSE CNVnator 1.41458000 0.38091100 31264334 31282433
# 1956 tp1335 Chr24 286901 289800 2900 FALSE CNVnator 1.74941000 0.74228100 31291334 31294233
```

```
get.cnv.dels <- function(cov.thresh.lo = 0.0,
                        cov.thresh.hi = 0.8,
                        cnv,
                        snp.tables = NULL,
                        DEBUG = FALSE)
){
  # build list of 7 Bool vectors of genome length, with i-th == T iff
  # * i-th pos is 'NA' in genome seq (if snp.tables are provided), or
  # * in CNVnator call for coverage in half-open [cov.thresh.lo, hi), and
  # * not marked 'filtered' by CNVnator
  cnv.deletions <- vector(mode='list', 7) # make list of bool vectors
  if(is.null(snp.tables)){
    # if no tables, assume full
    t.len <- genome.length.constants()$genome.length.trunc
  } else {
    t.len <- nrow(snp.tables[[1]])
  }
  for(st in 1:7){
    if(is.null(snp.tables)){
      cnv.deletions[[st]] <- logical(t.len) # all F
    } else {
      cnv.deletions[[st]] <- is.na(snp.tables[[st]]$Pos[1:t.len]) # NA positions in genome
    }
  }
  strain.names <- c(paste('tp10', c('07', 12:15), sep=''), 'IT', 'tp1335')
```

```

names(cnv.deletions) <- strain.names
for(i in 1:nrow(cnv)){
  if(!cnv$filtered[i] &&
      cnv$cov_ratio[i] >= cov.thresh.lo &&
      cnv$cov_ratio[i] < cov.thresh.hi)
  {
    if(DEBUG){
      print(cnv[i,])
      print(as.character(cnv$strain[i]))
    }
    # following ASSUMES no CNVnator call crosses a chromosome bdry, & that
    # t.len ends at chr end (typically chr1 or chr24)
    if(cnv$iEnd[i] <= t.len){
      cnv.deletions[[as.character(cnv$strain[i])][cnv$iStart[i]:cnv$iEnd[i]] <- TRUE
    }
  }
}
return(cnv.deletions)
}

# sanity check:
cnv.dels.38 <- get.cnv.dels(0.3, 0.8, cnv.chrononly, snp.tables = NULL)
unlist(lapply(cnv.dels.38, sum)) # does it match low.length.38 in tic ?

# tp1007 tp1012 tp1013 tp1014 tp1015 IT tp1335
# 1672500 1781500 1383600 1313700 988400 320900 1453000

# 1672500 1781500 1399400 1313700 988400 336500 1453000 <== low.length.38 from tic (circa page 8)
# 1672500 1781500 1399400 1313700 988400 336500 1453000 <== low.length.38 from tic (pg9, 6/28/17)
rm(cnv.dels.38)

```

Slight discrepancy in H-clade that I should hunt down, but basically OK. (hmm; maybe untrunc tbls.)

```

# the ones we want for the current analysis:
hemi.masks <- get.cnv.dels(0.3, 0.8, cnv.chrononly, snp.tables=snp.tables)

rbind(
  homnr      = unlist(lapply(hnr, sum)),
  hemi       = unlist(lapply(hemi.masks, sum)),
  homnr.unhemi = unlist(lapply(list(1,2,3,4,5,6,7), function(i){sum(hnr[[i]] & !hemi.masks[[i]]))}))
)

#           1007      1012      1013      1014      1015      3367      1335
# homnr      16069     14356     120037    11436      6862     142515     1854
# hemi       1834990   1940024   1527725   1472095   1134652   480817    1596965
# homnr.unhemi 9650      7347     111674     10091      5113     140185     1829

# based on the thought that hnr in 1335 may reflect errors in the ref seq,
# are they shared with others?
unlist(lapply(hnr, function(x){sum(x & hnr[[7]])})) # hnr shared with 1335

# 1007 1012 1013 1014 1015 3367 1335
# 517 592 748 362 617 793 1854

# answer: around 300 in each strain, of 558 in NY, genomewide,
# so that seems like a plausibly important factor.

hnr.lclade <- hnr[[1]] | hnr[[2]] | hnr[[4]] | hnr[[5]] | hnr[[7]] # union over L-clade
sum(hnr.lclade) # count all in L-clade

# [1] 31723

sum(hnr[[3]] | hnr[[6]]) # present in H-clade

# [1] 188637

```

```

sum(hnr[[3]] & hnr[[6]]) # shared in H-clade

# [1] 73915

# look at a few in L-clade
w.hnr.l <- which(hnr.lclade)
seecounts(w.hnr.l[1:10], snp.tables=snp.tables)

#   chr  pos Ref Strain  A  G  C  T SNP  exon indel nrf rat
# 1  Chr1 1559  A      1007 7  0  0 24  0 TRUE FALSE
# 2      1012 11  0  0 37  0 TRUE FALSE
# 3      1013 9  0  0 5  0 TRUE FALSE
# 4      1014 4  0  0 16  0 TRUE FALSE
# 5      1015 47 0  0 35  0 TRUE FALSE
# 6      3367 0  0  0 0  0 TRUE FALSE
# 7      1335 60 0  0 50  0 TRUE FALSE
# 9  Chr1 1575  G      1007 24 7  0 0  0 TRUE FALSE
# 10     1012 42 13  0 0  0 TRUE FALSE
# 11     1013 17 16  0 0  0 TRUE FALSE
# 12     1014 15 4  0 0  0 TRUE FALSE
# 13     1015 43 31  0 0  1 TRUE FALSE
# 14     3367 0 2  0 0  0 TRUE FALSE
# 15     1335 34 74  0 0  0 TRUE FALSE
# 17 Chr1 1893  C      1007 0  0 14 32  0 TRUE FALSE
# 18     1012 0  0 38 52  0 TRUE FALSE
# 19     1013 0  0 95 14  0 TRUE FALSE
# 20     1014 0  0 5 31  0 TRUE FALSE
# 21     1015 0  0 47 44  0 TRUE FALSE
# 22     3367 0  0 29 0  0 TRUE FALSE
# 23     1335 0  0 68 85  0 TRUE FALSE
# 25 Chr1 2223  A      1007 25 13  0 0  0 TRUE FALSE
# 26     1012 13 12  1 0  0 TRUE FALSE
# 27     1013 5 24  0 0  0 TRUE FALSE
# 28     1014 0 4  0 0  0 TRUE FALSE
# 29     1015 19 22  0 0  1 TRUE FALSE
# 30     3367 15 3  0 0  0 TRUE FALSE
# 31     1335 33 22  0 0  0 TRUE FALSE
# 33 Chr1 2319  C      1007 0 28 10 0  1 TRUE FALSE
# 34     1012 0 43 17 0  1 TRUE FALSE
# 35     1013 13 15 9 0  1 TRUE FALSE
# 36     1014 0 18 6 0  1 TRUE FALSE
# 37     1015 0 53 20 0  1 TRUE FALSE
# 38     3367 4 0 24 0  0 TRUE FALSE
# 39     1335 0 118 28 0  1 TRUE FALSE
# 41 Chr1 2502  A      1007 14 2  0 0  0 FALSE FALSE
# 42     1012 17 6  0 0  0 FALSE FALSE
# 43     1013 6 13  0 0  0 FALSE FALSE
# 44     1014 1 6  0 0  0 FALSE FALSE
# 45     1015 20 7  0 0  0 FALSE FALSE
# 46     3367 3 3  0 0  0 FALSE FALSE
# 47     1335 29 17  0 0  0 FALSE FALSE
# 49 Chr1 2573  C      1007 0  0 11 28  1 TRUE FALSE
# 50     1012 0  0 30 50  1 TRUE FALSE
# 51     1013 0  0 231 12  0 TRUE FALSE
# 52     1014 0  0 4 18  1 TRUE FALSE
# 53     1015 0  0 50 38  1 TRUE FALSE
# 54     3367 0  0 71 0  0 TRUE FALSE
# 55     1335 0  0 62 75  1 TRUE FALSE
# 57 Chr1 3938  G      1007 12 20  0 0  0 TRUE FALSE
# 58     1012 9 22  0 0  0 TRUE FALSE
# 59

```

```
# 60      1013 35 19 0 0 0 TRUE FALSE
# 61      1014 8 2 0 0 0 TRUE FALSE
# 62      1015 25 53 0 0 0 TRUE FALSE
# 63      3367 14 13 0 0 0 TRUE FALSE
# 64      1335 59 42 0 0 0 TRUE FALSE
# 65 Chr1 4876 G
# 66      1007 0 1 0 0 0 FALSE FALSE
# 67      1012 1 4 0 0 0 FALSE FALSE
# 68      1013 0 0 0 0 0 FALSE FALSE
# 69      1014 1 0 0 0 0 FALSE FALSE
# 70      1015 0 3 0 0 0 FALSE FALSE
# 71      3367 4 4 0 0 0 FALSE FALSE
# 72      1335 2 2 0 0 0 FALSE FALSE
# 73 Chr1 4938 T
# 74      1007 0 43 0 23 1 FALSE FALSE
# 75      1012 0 63 0 48 1 FALSE FALSE
# 76      1013 0 83 0 2 0 FALSE FALSE
# 77      1014 0 27 0 4 1 FALSE FALSE
# 78      1015 0 75 0 47 1 FALSE FALSE
# 79      3367 0 19 0 12 1 FALSE FALSE
# 80      1335 0 57 0 59 1 FALSE FALSE

# one of those is a little weird:
xx<-snp.tables[[1]][149457,]
for (i in 2:7){xx <- rbind(xx,snp.tables[[i]][149457,])}
row.names(xx)<-names(snp.tables)
# My guess is that Chr/Pos/Ref are left as NA if coverage is zero.
xx

#      snp Chr Pos Ref Cov a g c t n .match exon indel chr pos rawCov
# 1007 0 <NA> NA <NA> 0 0 0 0 0 0 0 FALSE FALSE <NA> NA 0
# 1012 0 <NA> NA <NA> 0 0 0 0 0 0 0 FALSE FALSE <NA> NA 0
# 1013 0 <NA> NA <NA> 0 0 0 0 0 0 0 FALSE FALSE <NA> NA 0
# 1014 0 Chr1 149457 G 0 0 0 0 0 0 0 FALSE FALSE Chr1 149457 1
# 1015 0 <NA> NA <NA> 0 0 0 0 0 0 0 FALSE FALSE <NA> NA 0
# 3367 0 <NA> NA <NA> 0 0 0 0 0 0 0 FALSE FALSE <NA> NA 0
# 1335 0 Chr1 149457 G 0 0 0 0 0 0 0 FALSE FALSE Chr1 149457 1
```

5 Table 1 stats

Here is a brief summary of per-strain SNP counts, pairwise overlaps, and other conveniently available stats, such as those shown in Table 1 of the paper.

```
snp.counts <- matrix(NA,7,4)
snp.pctofny <- matrix(NA,7,4)
snp.pctofself <- matrix(NA,7,4)
snp.inter <- matrix(NA,7,7)
snp.union <- matrix(NA,7,7)
rownames(snp.counts) <- names(snp.tables)
rownames(snp.pctofny) <- names(snp.tables)
rownames(snp.pctofself) <- names(snp.tables)
rownames(snp.inter) <- names(snp.tables)
colnames(snp.inter) <- names(snp.tables)
rownames(snp.union) <- names(snp.tables)
colnames(snp.union) <- names(snp.tables)
for(stringency in 1:4){
  cat('\nStringency', stringency, ifelse(stringency==4,'(i.e. raw SAMTools SNP calls)',''),
      '\n-----\n')
  for(i in 1:7){
    f.snps.i <- filtered.snps$Code$get.snps(i, stringency)
    snp.counts[i,stringency] <- sum(f.snps.i)
    for(j in i:7){
      f.snps.j <- filtered.snps$Code$get.snps(j, stringency)
      snp.inter[i,j] <- sum(f.snps.i & f.snps.j)
```



```

    snp.union[i,j] <- sum(f.snps.i | f.snps.j)
  }
}
snp.pctofny [,stringency] <- snp.inter[,7]/snp.counts[7,stringency]
snp.pctofself[,stringency] <- snp.inter[,7]/snp.counts[,stringency]
cat('Union Counts:\n'); print(snp.union)
cat('Intersect Counts:\n'); print(snp.inter)
cat('Intersect as percent of union:\n'); print(snp.inter/snp.union*100,digits=3)
}

#
# Stringency 1 :
# -----
# Union Counts:
#      1007    1012    1013    1014    1015    3367    1335
# 1007 184621 190979 363297 196256 197762 354191 199128
# 1012      NA 187793 364751 198002 198919 355526 200266
# 1013      NA      NA 296795 356666 366717 391621 364222
# 1014      NA      NA      NA 165741 196847 347035 195294
# 1015      NA      NA      NA      NA 191668 357845 198939
# 3367      NA      NA      NA      NA      NA 283086 355107
# 1335      NA      NA      NA      NA      NA      NA 187044
# Intersect Counts:
#      1007    1012    1013    1014    1015    3367    1335
# 1007 184621 181435 118119 154106 178527 113516 172537
# 1012      NA 187793 119837 155532 180542 115353 174571
# 1013      NA      NA 296795 105870 121746 188260 119617
# 1014      NA      NA      NA 165741 160562 101792 157491
# 1015      NA      NA      NA      NA 191668 116909 179773
# 3367      NA      NA      NA      NA      NA 283086 115023
# 1335      NA      NA      NA      NA      NA      NA 187044
# Intersect as percent of union:
#      1007 1012 1013 1014 1015 3367 1335
# 1007 100 95 32.5 78.5 90.3 32.0 86.6
# 1012  NA 100 32.9 78.6 90.8 32.4 87.2
# 1013  NA  NA 100.0 29.7 33.2 48.1 32.8
# 1014  NA  NA  NA 100.0 81.6 29.3 80.6
# 1015  NA  NA  NA  NA 100.0 32.7 90.4
# 3367  NA  NA  NA  NA  NA 100.0 32.4
# 1335  NA  NA  NA  NA  NA  NA 100.0
#
# Stringency 2 :
# -----
# Union Counts:
#      1007    1012    1013    1014    1015    3367    1335
# 1007 181682 189049 374070 191223 196135 364611 195371
# 1012      NA 186199 376272 194060 197053 366680 196729
# 1013      NA      NA 304444 356961 378452 408556 373836
# 1014      NA      NA      NA 137771 193813 346571 187896
# 1015      NA      NA      NA      NA 190384 369360 195610
# 3367      NA      NA      NA      NA      NA 290844 364209
# 1335      NA      NA      NA      NA      NA      NA 180709
# Intersect Counts:
#      1007    1012    1013    1014    1015    3367    1335
# 1007 181682 178832 112056 128230 175931 107915 167020
# 1012      NA 186199 114371 129910 179530 110363 170179
# 1013      NA      NA 304444 85254 116376 186732 111317
# 1014      NA      NA      NA 137771 134342 82044 130584
# 1015      NA      NA      NA      NA 190384 111868 175483
# 3367      NA      NA      NA      NA      NA 290844 107344
# 1335      NA      NA      NA      NA      NA      NA 180709
# Intersect as percent of union:
#      1007 1012 1013 1014 1015 3367 1335
# 1007 100 94.6 30.0 67.1 89.7 29.6 85.5
# 1012  NA 100.0 30.4 66.9 91.1 30.1 86.5
# 1013  NA  NA 100.0 23.9 30.8 45.7 29.8
# 1014  NA  NA  NA 100.0 69.3 23.7 69.5
# 1015  NA  NA  NA  NA 100.0 30.3 89.7

```

```

# 3367 NA NA NA NA NA 100.0 29.5
# 1335 NA NA NA NA NA NA 100.0
#
# Stringency 3 :
# -----
# Union Counts:
#      1007 1012 1013 1014 1015 3367 1335
# 1007 169346 183708 363625 176590 190884 354428 188236
# 1012 NA 179828 368821 185220 192949 359446 191771
# 1013 NA NA 296287 330697 371321 403242 364437
# 1014 NA NA NA 88328 187158 319824 176609
# 1015 NA NA NA NA 184704 362666 191220
# 3367 NA NA NA NA NA 283373 355133
# 1335 NA NA NA NA NA NA 171199
# Intersect Counts:
#      1007 1012 1013 1014 1015 3367 1335
# 1007 169346 165466 102008 81084 163166 98291 152309
# 1012 NA 179828 107294 82936 171583 103755 159256
# 1013 NA NA 296287 53918 109670 176418 103049
# 1014 NA NA NA 88328 85874 51877 82918
# 1015 NA NA NA NA 184704 105411 164683
# 3367 NA NA NA NA NA 283373 99439
# 1335 NA NA NA NA NA NA 171199
# Intersect as percent of union:
#      1007 1012 1013 1014 1015 3367 1335
# 1007 100 90.1 28.1 45.9 85.5 27.7 80.9
# 1012 NA 100.0 29.1 44.8 88.9 28.9 83.0
# 1013 NA NA 100.0 16.3 29.5 43.7 28.3
# 1014 NA NA NA 100.0 45.9 16.2 47.0
# 1015 NA NA NA NA 100.0 29.1 86.1
# 3367 NA NA NA NA NA 100.0 28.0
# 1335 NA NA NA NA NA NA 100.0
#
# Stringency 4 (i.e. raw SAMTools SNP calls) :
# -----
# Union Counts:
#      1007 1012 1013 1014 1015 3367 1335
# 1007 161103 176738 343873 171675 185741 336599 180313
# 1012 NA 166089 346766 176177 186459 339458 182312
# 1013 NA NA 247737 302322 352586 386037 339669
# 1014 NA NA NA 89184 179976 295574 162912
# 1015 NA NA NA NA 174701 345396 184068
# 3367 NA NA NA NA NA 240413 331982
# 1335 NA NA NA NA NA NA 153901
# Intersect Counts:
#      1007 1012 1013 1014 1015 3367 1335
# 1007 161103 150454 64967 78612 150063 64917 134691
# 1012 NA 166089 67060 79096 154331 67044 137678
# 1013 NA NA 247737 34599 69852 102113 61969
# 1014 NA NA NA 89184 83909 34023 80173
# 1015 NA NA NA NA 174701 69718 144534
# 3367 NA NA NA NA NA 240413 62332
# 1335 NA NA NA NA NA NA 153901
# Intersect as percent of union:
#      1007 1012 1013 1014 1015 3367 1335
# 1007 100 85.1 18.9 45.8 80.8 19.3 74.7
# 1012 NA 100.0 19.3 44.9 82.8 19.8 75.5
# 1013 NA NA 100.0 11.4 19.8 26.5 18.2
# 1014 NA NA NA 100.0 46.6 11.5 49.2
# 1015 NA NA NA NA 100.0 20.2 78.5
# 3367 NA NA NA NA NA 100.0 18.8
# 1335 NA NA NA NA NA NA 100.0

vs.stringency <- cbind(snp.counts, matrix(NA,7,1), round(snp.counts[,1:3]/snp.counts[,4]*100,1))
colnames(vs.stringency) <- c('[[1]]', '[[2]]', '[[3]]', '[[4]]', '----', '[[1]]%', '[[2]]%', '[[3]]%')

# SNPs vs filtering stringency (raw counts and as % of [[4]]). Medium filter
# adds 10-20% in most cases. Big exception is Gyre, where low coverage,

```

```
# high err rate and SAMTools conservatism seemed to seriously undercall:
print(vs.stringency)

#      [[1]]  [[2]]  [[3]]  [[4]]  ----  [[1]]%  [[2]]%  [[3]]%
# 1007 184621 181682 169346 161103   NA  114.6  112.8  105.1
# 1012 187793 186199 179828 166089   NA  113.1  112.1  108.3
# 1013 296795 304444 296287 247737   NA  119.8  122.9  119.6
# 1014 165741 137771  88328  89184   NA  185.8  154.5   99.0
# 1015 191668 190384 184704 174701   NA  109.7  109.0  105.7
# 3367 283086 290844 283373 240413   NA  117.7  121.0  117.9
# 1335 187044 180709 171199 153901   NA  121.5  117.4  111.2

# Intersect NY as % of self (vs stringency):
print(snp.pctofself*100, digits=3)

#      [,1]  [,2]  [,3]  [,4]
# 1007  93.5  91.9  89.9  83.6
# 1012  93.0  91.4  88.6  82.9
# 1013  40.3  36.6  34.8  25.0
# 1014  95.0  94.8  93.9  89.9
# 1015  93.8  92.2  89.2  82.7
# 3367  40.6  36.9  35.1  25.9
# 1335 100.0 100.0 100.0 100.0

# Intersect NY as % of NY (vs stringency):
print(snp.pctofny*100, digits=3)

#      [,1]  [,2]  [,3]  [,4]
# 1007  92.2  92.4  89.0  87.5
# 1012  93.3  94.2  93.0  89.5
# 1013  64.0  61.6  60.2  40.3
# 1014  84.2  72.3  48.4  52.1
# 1015  96.1  97.1  96.2  93.9
# 3367  61.5  59.4  58.1  40.5
# 1335 100.0 100.0 100.0 100.0
```

Quick look at coverage. Are there any NA?:

```
nacount <- NULL
for(i in 1:4){
  if(!is.null(tset[[i]])){
    nacount <- rbind(nacount,
                     unlist(lapply(tset[[i]], function(x){sum(is.na(x$Cov))}))
  )
  rownames(nacount)[nrow(nacount)] <- names(tset)[i]
}
}
nacount

#      1007 1012 1013 1014 1015 3367 1335
# snp.tables.full.unfiltered    0    0    0    0    0    0    0
# snp.tables.full.qfiltered     0    0    0    0    0    0    0
```

Seemingly no. What's average in unq- vs q-filtered:

```
snp.tables.unqfil <- tset.picker(c(1,2), table.set = tset)
snp.tables.qfil   <- tset.picker(c(3,4), table.set = tset)
cov.unqfil <- unlist(lapply(snp.tables.unqfil, function(x){mean(x$Cov)}))
cov.qfil   <- unlist(lapply(snp.tables.qfil,   function(x){mean(x$Cov, na.rm=T)}))
cov.both <- rbind(cov.unqfil, cov.qfil, cov.qfil/cov.unqfil)
i <- 1
if(!is.null(snp.tables.unqfil)){
  rownames(cov.both)[i] <- which.snp.tables(snp.tables.unqfil)
  i <- i+1
}
if(!is.null(snp.tables.qfil)){
  rownames(cov.both)[i] <- which.snp.tables(snp.tables.qfil)
```

```

i <- i+1
}
if(i==3){
  rownames(cov.both)[i] <- 'Ratio'
}
cat('Mean Coverage:\n'); cov.both

# Mean Coverage:
#
#           1007           1012           1013           1014           1015           3367           1335
# trunc-unfiltered 37.0555484 70.8060724 69.6610432 33.1009373 61.5365159 64.0284488 107.7425968
# trunc-qfiltered  28.2750286 51.3249686 45.4036337 13.7261052 48.7880005 44.8042054 81.8823765
# Ratio           0.7630444  0.7248668  0.6517794  0.4146742  0.7928301  0.6997547  0.7599815

```

5.1 Table 1 Data

Throw together the conveniently-available Table 1 data, in Table 1 row order:

```

# if coverage unavailable, build NA vector
if(!is.null(cov.unqfil)){cov.unqfilv <- cov.unqfil} else {cov.unqfilv <- rep(NA,times=7)}
if(!is.null(cov.qfil )){cov.qfilv  <- cov.qfil } else {cov.qfilv  <- rep(NA,times=7)}
tldata.df <- data.frame(
  id      = st.locs(1:7, id=T, loc=F, date=F),
  loc     = st.locs(1:7, id=F, loc=T, date=F),
  date    = st.locs(1:7, id=F, loc=F, date=T),
  cov.unq = cov.unqfilv,
  cov.q    = cov.qfilv,
  SNPs.4   = snp.counts[,4],
  SNPs.2   = snp.counts[,2],
  olap.ny.4 = snp.pctofny[,4]*100,
  olap.ny.2 = snp.pctofny[,2]*100
)
tlrow.order <- c(7,1,2,5,3,6,4)
print(tldata.df[tlrow.order,],digits=3)

#           id           loc date cov.unq cov.q SNPs.4 SNPs.2 olap.ny.4 olap.ny.2
# 1335 CCMP1335      New York 1958  107.7  81.9 153901 180709    100.0    100.0
# 1007 CCMP1007      Virginia 1964   37.1  28.3 161103 181682     87.5     92.4
# 1012 CCMP1012      W. Australia 1965  70.8  51.3 166089 186199     89.5     94.2
# 1015 CCMP1015      Puget Sound 1985  61.5  48.8 174701 190384     93.9     97.1
# 1013 CCMP1013      Wales 1973   69.7  45.4 247737 304444     40.3     61.6
# 3367 CCMP3367      Italy 2007   64.0  44.8 240413 290844     40.5     59.4
# 1014 CCMP1014 N. Pacific Gyre 1971  33.1  13.7  89184 137771     52.1     72.3

```

6 Shared-SNPs P-Value

Text of the main paper quotes a “p-value” for the observed degree of SNP sharing in L-clade (and/or L-clade excluding Gyre) under a null model that these isolates were sampled from a population globally in Hardy-Weinberg equilibrium. Details of this analysis are as follows.

6.1 SNP Concordance

Arbitrarily pick one isolate, say, A , as the “template”. Arbitrarily pick a heterozygous (aka “SNP”) position in A . Let p_1 , and $q_1 = 1 - p_1$ be the frequencies in the overall population of the two nucleotides observed at that position in A . (Positions having 3 or 4 nucleotide variants segregating in the population are assumed to be negligibly rare.) Under the HWE null model, a second isolate B will also be heterozygous at the same position with probability $2p_1q_1 \leq 1/2$. Similarly, this position will be heterozygous in a third isolate C with the same probability, *independently*, and so on for isolates D and E . Overall, (assuming HWE) the probability that a heterozygous position in A is simultaneously heterozygous in the other 4 isolates is at most $1/2^4 = 1/16$. Continuing, suppose we pick a second heterozygous position in A , on a different chromosome with allele frequencies $p_2, q_2 = 1 - p_2$, say. Again assuming HWE, this

position will be a SNP in all of B, C, D and E with probability $(2p_2q_2)^4 \leq 1/16$, and this is independent of the first position, since segregation on different chromosomes is unlinked. Repeat this at 24 heterozygous positions in A , one per chromosome. Then, the number of five-way concordant positions observed should be dominated by the number observed when sampling from a binomial distribution with parameters $n = 24$ and $p = 1/16$, i.e., we expect at most $1/16 = 6.25\%$ of positions to agree, or at most $24/16 = 1.5$ five-way concordant positions in total. In sharp contrast, choosing CCMP 1014 (North Pacific Gyre) as the template, we see many more five-way concordant positions than predicted under these assumptions:

```
gyre.count <- sum(snp.tables[[4]]$snp)
# 'unfil.' => unfiltered for consistency; see below.
unfil.fiveway.count <- sum(snp.tables[[4]]$snp * i4.snps)
unfil.fiveway.percent <- unfil.fiveway.count / gyre.count * 100
unfil.p.value <- pbinom(floor(unfil.fiveway.count/gyre.count*24)-1, 24, 1/16, lower.tail = FALSE)
consistency.comparison <-
  data.frame(
    fiveway.count = unfil.fiveway.count,
    fiveway.percent = unfil.fiveway.percent,
    p.value = unfil.p.value
  )
consistency.comparison

#   fiveway.count fiveway.percent      p.value
# 1          70687       79.25973 4.142632e-19
```

Namely, 89184 positions are called as SNPs in CCMP1014, of which 70687 or 79.2597327% are also called as SNPs in *all four* other L-clade isolates. 79.2597327% of 24 is 19.0223358, and the probability of seeing 19 or more “Heads” in 24 flips of a biased coin with $P(\text{Heads}) \leq 1/16$, i.e., our p-value under the HWE null hypothesis, is at most: $4.1426317 \times 10^{-19}$ based on this simple binomial model. This is obviously strong evidence against the null hypothesis.

This analysis is potentially overly-simplistic in four respects, addressed below.

1. “ $2pq \leq 1/2$ ” is conservative. Neutral theory predicts that most variant nucleotides are rare in the population, so $2pq \ll 1/2$ is to be expected. This should make our quoted p-value very conservative.
2. Effect of Erroneous SNP calls. We base our analysis on *predicted* (by SAMTOOLS) heterozygous positions, not absolute-truth, which may affect our conclusions. However,
 - False negatives in A are irrelevant, since we never examine those positions. (This is the motivation for using CCMP1014 as the template; it has the lowest predicted SNP rate, likely due to a high false negative rate in that sequencing run. As noted elsewhere, it had the lowest coverage and lowest sequence quality of the 7 isolates, both of which impair SNP calling.)
 - False negatives in $BCDE$ make such positions appear *non*-concordant. For our purpose, this makes our statistic more conservative since it can only deflate a statistic that we argue is nevertheless unexpectedly large.
 - False positive calls in A are conservatively treated, as well: barring simultaneous false-positive calls in all of $BCDE$, such a position will appear non-concordant, again deflating the statistic. The *false* positive rates in B, C, D and E are unknown, but cannot exceed SAMTOOLS *total* positive rate, which is below 1% in all 7 isolates, suggesting a simultaneous $BCDE$ false positive rate $< 10^{-8}$, which will have a negligible effect.
 - A potentially more serious issue is a true positive in A aligned to false positives in BCD and/or E . (I.e., a position that is polymorphic in the population and heterozygous in A , under the HWE null model is likely to be homozygous for one of the two alleles in one or more of $BCDE$; false positive SNP calls in all of those isolates would make the site appear concordant, i.e., provide evidence against the null model.) However, (a) my impression is that SAMTOOLS is more prone to false negative calls than to false positive calls (see Section 4), and (b) we would need a high rate of false positives to turn a truly heterozygous but non-concordant A call into a false “concordant” call—I’d expect at most half (especially given point 1 above) of $BCDE$ to be heterozygous, but all would need to be falsely declared heterozygous. Such a high false positive rate on $BCDE$ seems unlikely (see previous bullet), and would likely be counterbalanced

by a similarly increased rate of false positives on A , which, as noted, tend to deflate our statistic (previous bullet again).

- Systematic errors. If there were, say, a sequence-context-dependent bias in the DNA sequencing, mapping and/or SNP-calling that tended to suggest (or hide) a SNP at some position, we’re going to systematically over- (or under-) estimate concordant SNPs across isolates. The discordance of called SNPs between the L- and H-clades and within the H-clade suggests that this is not a major problem, but it is worth noting as a possibility.
3. Discordant nucleotides at “concordant” SNP positions. A “shared” SNP at a given position might be, say, G/C in one isolate vs T/C in another, reflecting an unexpected tri-allelic position in the population or a technical sequencing error. It is inappropriate to count such a “shared” SNP position as evidence against the null hypothesis, since it isn’t clear that it is truly shared. Instead, I will identify such inconsistent positions, based on the “stringency [[2]]” criteria established above, and treat each as non-concordant. I.e., a position will be considered to be a “5-way concordant SNP” if and only if it was called as a SNP by SAMTOOLS (independently) in all 5 L-clade isolates, *and* shows the same dominant non-reference nucleotide in all 5, according to criteria [[2]] above. As it turns out, this correction has a very minor effect on the resulting p-value:

```
# 'unfil.' => Ignoring "consistency"; 'fil.' => Filtering for "consistency":
fil.fiveway.count <- sum((snp.tables[[4]]$snp * i4.snps)[union.snps == 1] & consistent[[2]])
fil.fiveway.percent <- fil.fiveway.count / gyre.count * 100
fil.p.value <- pbinom(floor(fil.fiveway.count/gyre.count*24)-1, 24, 1/16, lower.tail = FALSE)
# append new stats to previous table for easy comparison
consistency.comparison <-
  rbind(consistency.comparison,
        data.frame(
          fiveway.count = fil.fiveway.count,
          fiveway.percent = fil.fiveway.percent,
          p.value = fil.p.value
        )
  )
rownames(consistency.comparison) <- c('unfiltered', 'consistency.filtered')
consistency.comparison

#               fiveway.count fiveway.percent      p.value
# unfiltered           70687          79.25973 4.142632e-19
# consistency.filtered    69941          78.42326 1.976512e-17
```

In particular, it removes 0.8% of five-way consistent positions (only 746 of 70687 positions), and still shows a highly significant p-value.

4. “ $P(E[X]) \neq E[P(X)]$ ”. I’m expressing this poorly, but finding the p-value based on the *expected* number of concordant positions is somewhat non-standard. A more typical set-up would use the *actual* value of some statistic, then calculate the probability of observing a value that extreme (or more extreme) under the null model. The fundamental problem is that we have thousands of SNPs, but I don’t see an easy way to use more than 24 of them at a time, because potential genetic linkage seemingly destroys statistical independence, which is key to most simple analyses. A somewhat more formal, but still non-standard, approach is the following. Suppose we randomly sample one SNP per chromosome and count the number X of them that are 5-way concordant. What I outlined above calculated the p-value based on $E[X]$, the expected value of X , i.e., $P(E[X])$. Alternatively, we can calculate $E[P(X)]$, the expected p-value. (They are not the same.) In effect, this averages the p-values that would be seen over many different randomly-sampled sets of 24 SNPs. This is not difficult to calculate. First, the probability that we would observe $0 \leq i \leq 24$ concordant positions in a sample of 24, given that 78.42% of positions are concordant follows this binomial distribution:

```
x.equals.i.distribution <- dbinom(0:24, 24, fil.fiveway.percent/100)
print(x.equals.i.distribution, digits=3)

# [1] 1.04e-16 9.04e-15 3.78e-13 1.01e-11 1.92e-10 2.80e-09 3.22e-08 3.01e-07 2.32e-06 1.50e-05
# [11] 8.18e-05 3.78e-04 1.49e-03 5.00e-03 1.43e-02 3.46e-02 7.07e-02 1.21e-01 1.71e-01 1.96e-01
# [21] 1.78e-01 1.23e-01 6.12e-02 1.93e-02 2.93e-03
```

Second, the p-value corresponding to $0 \leq i \leq 24$ observed concordant positions also follows a different binomial distribution:

```
p.val.of.x.equals.i <- c(1, pbinom(0:23, 24, 1/16, lower.tail = F))
print(p.val.of.x.equals.i, digits=3)

# [1] 1.00e+00 7.88e-01 4.48e-01 1.87e-01 5.95e-02 1.49e-02 3.01e-03 4.99e-04 6.90e-05 8.02e-06
# [11] 7.89e-07 6.60e-08 4.72e-09 2.87e-10 1.49e-11 6.59e-13 2.46e-14 7.66e-16 1.98e-17 4.14e-19
# [21] 6.88e-21 8.70e-23 7.88e-25 4.56e-27 1.26e-29
```

Finally, the expected (or “average”) p-value is just the weighted average of the latter values, weighted by the former:

```
e.of.p.of.x <- sum(x.equals.i.distribution * p.val.of.x.equals.i)
e.of.p.of.x

# [1] 6.808209e-10
```

This is still highly significant, but weaker than the $P(E[X])$ analysis, basically because $X < E[X]$ has a fair probability of occurring, and the corresponding p-value $P(X)$ rises rapidly as X declines.

Another way to look at the numbers:

```
pvdof <- data.frame(x.density=x.equals.i.distribution,
                    x.cdf=cumsum(x.equals.i.distribution),
                    pval.of.x=p.val.of.x.equals.i)
print(pvdof, digits=4)

#   x.density    x.cdf pval.of.x
# 1 1.037e-16 1.037e-16 1.000e+00
# 2 9.043e-15 9.147e-15 7.875e-01
# 3 3.780e-13 3.871e-13 4.476e-01
# 4 1.008e-11 1.046e-11 1.869e-01
# 5 1.922e-10 2.027e-10 5.950e-02
# 6 2.795e-09 2.998e-09 1.490e-02
# 7 3.217e-08 3.517e-08 3.010e-03
# 8 3.007e-07 3.358e-07 4.994e-04
# 9 2.322e-06 2.658e-06 6.899e-05
# 10 1.500e-05 1.766e-05 8.015e-06
# 11 8.181e-05 9.947e-05 7.887e-07
# 12 3.784e-04 4.779e-04 6.603e-08
# 13 1.490e-03 1.968e-03 4.716e-09
# 14 4.999e-03 6.967e-03 2.875e-10
# 15 1.428e-02 2.124e-02 1.493e-11
# 16 3.459e-02 5.584e-02 6.590e-13
# 17 7.072e-02 1.266e-01 2.456e-14
# 18 1.210e-01 2.475e-01 7.662e-16
# 19 1.710e-01 4.185e-01 1.977e-17
# 20 1.963e-01 6.148e-01 4.143e-19
# 21 1.783e-01 7.931e-01 6.877e-21
# 22 1.235e-01 9.165e-01 8.701e-23
# 23 6.119e-02 9.777e-01 7.884e-25
# 24 1.934e-02 9.971e-01 4.556e-27
# 25 2.929e-03 1.000e+00 1.262e-29
```

E.g., row 9 in that table says that the concordance rate (78%) is so high that a sample of 24 SNPs will almost always have 9 or more five-way concordant positions (probability of fewer is only 2.658e-06), while under the null model, seeing 9 or more is very unlikely (probability at most 6.899e-05). ***AM I OFF-BY-ONE INTERPRETING ROW 9 HERE??***

6.2 Notes

In earlier drafts, an analog of the above analysis was based on the concordance of *refined* SNPs. This now seems to me to be questionable, since the “refined” SNP calling makes SNPs called across L-clade non-independent. OTOH,

the above analysis seems valid: SAMTOOLS was run on each isolate independently, and likewise “criterion [[2]]” is evaluated independently in each strain, and is being used here solely to remove SNP predictions, not to add them. “Systematic errors” as outlined above remain a potential problem, but again discordance with/within H-clade suggests that this is of limited concern.

For completeness, I did a similar analysis including a sample of H-clade comparisons: Gyre vs Italy, NY vs Italy, NY vs Italy+Wales, and of Italy vs Wales. As expected, none of these show a statistically significant p-value, although the $\approx 40\%$ concordance in the 2-way comparisons, while $< 1/2$ as predicted, is a bit higher than I expected based on “neutral theory implies many rare variants.” (I did not bother to include “criterion[[2]] filtering” in these calculations.)

```
# 'gi.twoway' => gyre vs italy 2-way concordance;
# 'ni.twoway' => new york vs italy 2-way concordance;
# not bothering with criterion[[2]] filtering
gi.twoway.count <- sum(snp.tables[[4]]$snp * snp.tables[[6]]$snp)
gi.twoway.percent <- gi.twoway.count / gyre.count * 100
gi.p.value <- pbinom(floor(gi.twoway.count/gyre.count*24)-1, 24, 1/2, lower.tail = FALSE)
ny.count <- sum(snp.tables[[7]]$snp)
ni.twoway.count <- sum(snp.tables[[7]]$snp * snp.tables[[6]]$snp)
ni.twoway.percent <- ni.twoway.count / ny.count * 100
ni.p.value <- pbinom(floor(ni.twoway.count/ny.count*24)-1, 24, 1/2, lower.tail = FALSE)
niw.threeway.count <- sum(snp.tables[[7]]$snp * snp.tables[[6]]$snp * snp.tables[[3]]$snp)
niw.threeway.percent <- niw.threeway.count / ny.count * 100
niw.p.value <- pbinom(floor(niw.threeway.count/ny.count*24)-1, 24, 1/4, lower.tail = FALSE)
it.count <- sum(snp.tables[[6]]$snp)
iw.twoway.count <- sum(snp.tables[[6]]$snp * snp.tables[[3]]$snp)
iw.twoway.percent <- iw.twoway.count / it.count * 100
iw.p.value <- pbinom(floor(iw.twoway.count/it.count*24)-1, 24, 1/2, lower.tail = FALSE)
consistency.comparison <-
  rbind(consistency.comparison,
    data.frame(
      fiveway.count = c(gi.twoway.count, ni.twoway.count, niw.threeway.count, iw.twoway.count),
      fiveway.percent = c(gi.twoway.percent, ni.twoway.percent, niw.threeway.percent, iw.twoway.percent),
      p.value = c(gi.p.value, ni.p.value, niw.p.value, iw.p.value)
    )
  )
colnames(consistency.comparison)[1:2] <- c('552232way.count', '552232way.percent') # old col names misleading
rownames(consistency.comparison)[3:6] <- c('gyre.vs.italy', 'new.york.vs.italy', # new rows
      'ny.vs.it.plus.wales', 'it.vs.wales')

consistency.comparison

#           552232way.count 552232way.percent      p.value
# unfiltered              70687          79.25973 4.142632e-19
# consistency.filtered      69941          78.42326 1.976512e-17
# gyre.vs.italy             34023          38.14922 9.242052e-01
# new.york.vs.italy         62332          40.50136 9.242052e-01
# ny.vs.it.plus.wales       35796          23.25911 7.533516e-01
# it.vs.wales              102113          42.47399 8.462719e-01
```

6.3 P-Value: The Bottom Line

So, what to say in the body of the paper? $E[P(X)]$ is highly significant, and conservative, but complex to explain. $P(E[X])$ is simpler to explain, but may be criticized as misleading if we aren’t very careful in that explanation. I’m slightly leaning towards the last option, but want to sleep on it and draft the key sentence or two before settling.

7 Sharing

The following analysis looks at the sharing patterns among the consistent SNPs. I assume that shared SNPs reflect shared ancestry, and that SNPs accumulate slowly over time. Then, in outline, the story is consistent with what we have seen in other analyses—there seem to be 3 groups: 1013 (Wales) in one, 3367 (Italy) in another, and the other 5 in a third, with some hints as to the order of divergence. A caveat is that in a sexual population, non-shared SNPs do not immediately imply non-shared ancestry; they may merely reflect Hardy-Weinberg capturing a homozygous state

in one isolate vs the other. (Or read errors, etc.) Thus, if we are right that the H-isolates retain sex, then the large number of “private” SNPs in H may be at least partially due to HWE.

Analysis is broken into cases based on how many strains share a particular SNP.

7.1 Code

To categorize SNPs by sharing patterns, first convert the 7-way consistent sharing pattern into a 7-bit binary number, and tabulate based on that:

```
# convert (n x 7) 0-1 matrix to n vector of 0-127
tobin <- function(x){
  bin <- integer(nrow(x)) # initialized to 0
  for(i in 1:7){
    bin <- bin*2 + as.integer(x[,i]>0)
  }
  return(bin)
}

# get full set of patterns
snp.pattern.all <- lapply(non.refs,tobin)
# prune to just the consistent ones
snp.pattern <- snp.pattern.all
for(i in 1:3){
  snp.pattern[[i]][!consistent[[i]]] <- NA
}

# analogous to built-in ``table'' but simpler. Count entries in an integer
# vector sharing values in a (smallish) range. Result is a 2-column matrix with
# the shared values in col 1 and count of occurrences of that value in col 2.
# Out-of-range values cause subscript error.
mytable <- function(vec, therange=range(vec,na.rm=T)){
  counts <- matrix(0,nrow=therange[2]-therange[1]+1,ncol=2,dimnames=list(NULL,c('val','count')))
  counts[1:nrow(counts),1] <- therange[1]:therange[2]
  for(i in 1:length(vec)){
    if(!is.na(vec[i])){
      counts[vec[i]-therange[1]+1,2] <- counts[vec[i]-therange[1]+1,2] + 1
    }
  }
  return(counts)
}

pattern.counts <- lapply(snp.pattern, function(x){mytable(x,c(0,127))})
```

To display the results, build a data frame whose i -th row, $0 \leq i \leq 127$ shows one of the 128 possible sharing patterns, with counts of the numbers of consistent, shared SNPs with that pattern according to criteria c1-c3.

```
tobitvec <- function(x){
  bitvec <- integer(7)
  for(i in 7:1){
    bitvec[i] <- x %% 2
    x <- x %% 2
  }
  return(bitvec)
}

flg <- function(x){
  return(ifelse(x==1, 'X', ''))
}

pat.summary <- function(listOfTbls){
  mydf <- data.frame(pat=0:127,sharedBy=NA,
    tp1007='',tp1012='',tp1013='',tp1014='',tp1015='',tp3367='',tp1335='',
    count1=NA,count2=NA,count3=NA,count4=NA,stringsAsFactors=F)

  for(i in 1:128){
```

```

bvec <- tobitvec(i-1)
mydf[i, 'sharedBy'] = sum(bvec)
mydf[i, 'tp1007'] = flg(bvec[1])
mydf[i, 'tp1012'] = flg(bvec[2])
mydf[i, 'tp1013'] = flg(bvec[3])
mydf[i, 'tp1014'] = flg(bvec[4])
mydf[i, 'tp1015'] = flg(bvec[5])
mydf[i, 'tp3367'] = flg(bvec[6])
mydf[i, 'tp1335'] = flg(bvec[7])
}

for(i in 1:length(listOfTbls)){
  tbl <- listOfTbls[[i]]
  if(!is.null(tbl)){
    mydf[,9+i] <- tbl[,2] ## count1/2/3/4 are columns 10/11/12/13 in mydf
    #for(j in 1:length(tbl)){
    #  k <- as.integer(rownames(tbl)[j]);
    #  mydf[k+1,9+i] <- tbl[j] ## count1/2/3 are columns 10/11/12
    #}
  }
}

mydf$pat <- as.octmode(mydf$pat) # display bit pattern in octal
return(mydf)
}

pat.summaries <- pat.summary(pattern.counts)

```

7.2 Sanity Checks

Some sanity checking: table sums equal to number of consistent positions?

```

all(consistent.count == apply(pat.summaries[,10:13],2,sum))

# [1] TRUE

```

More sanity checking: visually inspect a pattern with small counts, specifically pattern 12, i.e., consistent SNPs shared by only strains 1014 and 1015 (2nd and 3 rows from bottom, binary code $12 = 2^3 + 2^2$). There are only 10 such positions on Chr1. Chr1 2524239 has pattern 12 under criteria c1 and c2 but not c3; Chr1 1088766 has in c2 only. Both look good. Neither position is a *called* SNP except in 1015. However, all but 1 nonreference read agree with the called SNP (the exception being one read in Wales). Both 1014 and 1015 have at least 2 non-reference reads, comprising at least 5% of coverage, and in both strains, those reads are on the same non-reference base, satisfying criterion c2. The other strains have higher coverage and/or lower non-reference counts, so they do not satisfy c2. Position 2524239 also satisfies c1, but not c3, since 2 reads out of 35 is below the 10% threshold. (It is pattern 4 under c3, i.e., a SNP private to 1015.) Position 1088766 is also pattern 4 under c3 (2 reads out of 56 in 1335 is below both thresholds), and it is not consistent under c1, since the single A read in 1013 is discordant with the other non-reference reads.

```

unlist(lapply(snp.pattern,function(x){sum(x==12,na.rm=T)}))

# [1] 133 139 132 417

sp1 <- snp.pattern[[1]]==12
sp2 <- snp.pattern[[2]]==12
sp3 <- snp.pattern[[3]]==12
sp4 <- snp.pattern[[4]]==12
c(sum(sp1,na.rm=T), sum(sp2,na.rm=T), sum(sp3,na.rm=T), sum(sp4,na.rm=T))

# [1] 133 139 132 417

r1 <- rownames(non.refs[[1]])[which(sp1)]
r2 <- rownames(non.refs[[2]])[which(sp2)]
r3 <- rownames(non.refs[[3]])[which(sp3)]
r4 <- rownames(non.refs[[4]])[which(sp4)]

r2

```

```

# [1] "Chr1:1799155" "Chr2:713075" "Chr2:1464209" "Chr2:2406031"
# [5] "Chr2:2480466" "Chr2:2480532" "Chr2:2480838" "Chr2:2483322"
# [9] "Chr2:2488863" "Chr2:2489189" "Chr2:2490933" "Chr2:2492886"
# [13] "Chr2:2492887" "Chr2:2497794" "Chr2:2500122" "Chr2:2503000"
# [17] "Chr2:2507585" "Chr2:2507680" "Chr2:2510117" "Chr2:2513923"
# [21] "Chr2:2515103" "Chr2:2516669" "Chr2:2516751" "Chr2:2518558"
# [25] "Chr2:2518653" "Chr2:2518980" "Chr2:2519285" "Chr2:2519288"
# [29] "Chr2:2519718" "Chr2:2520984" "Chr2:2521271" "Chr2:2522648"
# [33] "Chr2:2524223" "Chr2:2524439" "Chr2:2525160" "Chr2:2525463"
# [37] "Chr2:2527281" "Chr2:2527916" "Chr2:2528472" "Chr2:2528769"
# [41] "Chr2:2529076" "Chr2:2529140" "Chr2:2529684" "Chr2:2530064"
# [45] "Chr2:2530216" "Chr2:2530239" "Chr2:2530294" "Chr2:2530768"
# [49] "Chr2:2530896" "Chr2:2531114" "Chr2:2531498" "Chr2:2531567"
# [53] "Chr2:2532173" "Chr2:2532365" "Chr2:2533028" "Chr2:2533171"
# [57] "Chr2:2533440" "Chr2:2534441" "Chr2:2535121" "Chr2:2535122"
# [61] "Chr2:2535314" "Chr2:2535493" "Chr2:2535503" "Chr2:2535509"
# [65] "Chr2:2535862" "Chr2:2536242" "Chr2:2537201" "Chr2:2537864"
# [69] "Chr2:2537917" "Chr2:2538072" "Chr2:2538498" "Chr2:2539318"
# [73] "Chr2:2543595" "Chr2:2545615" "Chr2:2545798" "Chr2:2546865"
# [77] "Chr2:2546991" "Chr2:2547055" "Chr2:2547086" "Chr2:2547120"
# [81] "Chr2:2547155" "Chr2:2547212" "Chr2:2547248" "Chr2:2547318"
# [85] "Chr2:2547554" "Chr2:2547938" "Chr2:2547944" "Chr2:2548131"
# [89] "Chr2:2549281" "Chr2:2551574" "Chr2:2551930" "Chr2:2554708"
# [93] "Chr2:2554860" "Chr2:2555005" "Chr2:2555203" "Chr2:2555820"
# [97] "Chr3:192441" "Chr3:496665" "Chr4:1086589" "Chr4:1393682"
# [101] "Chr4:1747983" "Chr4:2314475" "Chr5:7509" "Chr5:141375"
# [105] "Chr5:1071721" "Chr6:1330532" "Chr7:399475" "Chr7:1736991"
# [109] "Chr7:1813303" "Chr8:556556" "Chr10:54351" "Chr10:95217"
# [113] "Chr10:947088" "Chr11a:344258" "Chr11b:75778" "Chr12:214112"
# [117] "Chr12:458461" "Chr12:507608" "Chr13:96361" "Chr13:375598"
# [121] "Chr14:284131" "Chr15:417704" "Chr16a:39914" "Chr16a:177501"
# [125] "Chr16a:206719" "Chr16a:394030" "Chr17:461465" "Chr19a_19:300076"
# [129] "Chr19a_19:303090" "Chr19c_29:64170" "Chr19c_29:64811" "Chr19c_29:65720"
# [133] "Chr20:230994" "Chr20:486431" "Chr20:519835" "Chr22:380816"
# [137] "Chr23:190382" "Chr23:274291" "Chr24:114599"

c1 <- as.integer(unlist(lapply(strsplit(r1[1:min(20,length(r1))],':',fixed=TRUE),function(x){x[2]})))
c2 <- as.integer(unlist(lapply(strsplit(r2[1:min(20,length(r2))],':',fixed=TRUE),function(x){x[2]})))
c3 <- as.integer(unlist(lapply(strsplit(r3[1:min(20,length(r3))],':',fixed=TRUE),function(x){x[2]})))
c4 <- as.integer(unlist(lapply(strsplit(r4[1:min(20,length(r4))],':',fixed=TRUE),function(x){x[2]})))

c1
# [1] 614335 914018 1317406 2388286 62676 713075 2406031 2480466 2480838 2481998 2483322
# [12] 2488863 2489189 2490933 2492887 2497794 2500122 2503000 2507585 2507680

c2
# [1] 1799155 713075 1464209 2406031 2480466 2480532 2480838 2483322 2488863 2489189 2490933
# [12] 2492886 2492887 2497794 2500122 2503000 2507585 2507680 2510117 2513923

c3
# [1] 371484 518347 1210354 2209068 2264683 2898352 1276745 1464904 1464905 1766966 2347253
# [12] 2406031 2480532 2480838 2483322 2488863 2489189 2490933 2497794 2507585

c4
# [1] 518347 691730 767408 1049906 1390437 2072951 2254059 2254789 2264683 2823796 2898352
# [12] 2998868 77394 77407 155680 761325 968120 1182096 1222176 1264023

seecounts(c2,snp.tables=snp.tables)

# chr pos Ref Strain A G C T SNP exon indel nrf rat
# 1 Chr1 1799155 C
# 2 1007 0 0 10 1 0 TRUE FALSE
# 3 1012 0 0 16 1 0 TRUE FALSE
# 4 1013 0 0 10 0 0 TRUE FALSE

```

# 5			1014	0	0	8	2	0	TRUE	FALSE
# 6			1015	0	0	12	3	1	TRUE	FALSE
# 7			3367	1	0	1	1	1	TRUE	FALSE
# 8			1335	0	0	7	1	0	TRUE	FALSE
# 9	Chr1	713075	T							
# 10			1007	0	0	0	37	0	TRUE	FALSE
# 11			1012	0	0	0	90	0	TRUE	FALSE
# 12			1013	0	0	0	65	0	TRUE	FALSE
# 13			1014	0	0	0	32	0	TRUE	FALSE
# 14			1015	0	0	0	84	0	TRUE	FALSE
# 15			3367	0	0	0	53	0	TRUE	FALSE
# 16			1335	0	0	0	109	0	TRUE	FALSE
# 17	Chr1	1464209	T							
# 18			1007	0	0	0	22	0	FALSE	FALSE
# 19			1012	0	0	0	38	0	FALSE	FALSE
# 20			1013	0	0	0	22	0	FALSE	FALSE
# 21			1014	0	0	0	12	0	FALSE	FALSE
# 22			1015	0	0	0	30	0	FALSE	FALSE
# 23			3367	0	0	0	39	0	FALSE	FALSE
# 24			1335	0	0	0	81	0	FALSE	FALSE
# 25	Chr1	2406031	C							
# 26			1007	0	0	18	0	0	TRUE	FALSE
# 27			1012	0	0	23	0	0	TRUE	FALSE
# 28			1013	0	0	46	0	0	TRUE	FALSE
# 29			1014	0	0	13	0	0	TRUE	FALSE
# 30			1015	0	0	34	0	0	TRUE	FALSE
# 31			3367	0	0	29	0	0	TRUE	FALSE
# 32			1335	0	0	68	0	0	TRUE	FALSE
# 33	Chr1	2480466	A							
# 34			1007	26	0	0	0	0	TRUE	FALSE
# 35			1012	42	0	0	0	0	TRUE	FALSE
# 36			1013	39	0	0	0	0	TRUE	FALSE
# 37			1014	9	0	0	0	0	TRUE	FALSE
# 38			1015	49	0	0	0	0	TRUE	FALSE
# 39			3367	32	0	0	0	0	TRUE	FALSE
# 40			1335	77	0	0	0	0	TRUE	FALSE
# 41	Chr1	2480532	G							
# 42			1007	0	25	0	0	0	TRUE	FALSE
# 43			1012	0	27	0	0	0	TRUE	FALSE
# 44			1013	0	43	0	0	0	TRUE	FALSE
# 45			1014	0	1	0	0	0	TRUE	FALSE
# 46			1015	0	23	0	0	0	TRUE	FALSE
# 47			3367	0	23	0	0	0	TRUE	FALSE
# 48			1335	0	71	0	0	0	TRUE	FALSE
# 49	Chr1	2480838	T							
# 50			1007	0	0	0	8	0	TRUE	FALSE
# 51			1012	0	0	0	12	0	TRUE	FALSE
# 52			1013	0	0	0	24	0	TRUE	FALSE
# 53			1014	0	0	0	6	0	TRUE	FALSE
# 54			1015	0	0	0	15	0	TRUE	FALSE
# 55			3367	0	0	0	9	0	TRUE	FALSE
# 56			1335	0	0	0	81	0	TRUE	FALSE
# 57	Chr1	2483322	A							
# 58			1007	22	0	0	0	0	TRUE	FALSE
# 59			1012	23	0	0	0	0	TRUE	FALSE
# 60			1013	52	0	0	0	0	TRUE	FALSE
# 61			1014	24	0	0	0	0	TRUE	FALSE
# 62			1015	55	0	0	0	0	TRUE	FALSE
# 63			3367	37	0	0	0	0	TRUE	FALSE
# 64			1335	82	0	0	0	0	TRUE	FALSE
# 65	Chr1	2488863	C							
# 66			1007	0	0	26	0	0	FALSE	FALSE
# 67			1012	0	0	34	0	0	FALSE	FALSE
# 68			1013	0	0	27	0	0	FALSE	FALSE
# 69			1014	0	0	11	0	0	FALSE	FALSE
# 70			1015	0	0	34	0	0	FALSE	FALSE
# 71			3367	0	0	43	0	0	FALSE	FALSE

# 72			1335	0	0	71	0	0	FALSE	FALSE
# 73	Chr1 2489189	C								
# 74			1007	0	0	32	0	0	FALSE	FALSE
# 75			1012	0	0	63	0	0	FALSE	FALSE
# 76			1013	0	0	44	0	0	FALSE	FALSE
# 77			1014	0	0	26	0	0	FALSE	FALSE
# 78			1015	0	0	59	0	0	FALSE	FALSE
# 79			3367	0	0	24	0	0	FALSE	FALSE
# 80			1335	0	0	110	0	0	FALSE	FALSE
# 81	Chr1 2490933	G								
# 82			1007	0	25	0	0	0	FALSE	FALSE
# 83			1012	0	57	0	0	0	FALSE	FALSE
# 84			1013	0	40	0	0	0	FALSE	FALSE
# 85			1014	0	9	0	0	0	FALSE	FALSE
# 86			1015	0	36	0	0	0	FALSE	FALSE
# 87			3367	0	37	0	1	0	FALSE	FALSE
# 88			1335	0	57	0	0	0	FALSE	FALSE
# 89	Chr1 2492886	T								
# 90			1007	0	0	0	27	0	FALSE	FALSE
# 91			1012	0	0	0	61	0	FALSE	FALSE
# 92			1013	0	0	0	41	0	FALSE	FALSE
# 93			1014	0	0	0	18	0	FALSE	FALSE
# 94			1015	0	0	0	53	0	FALSE	FALSE
# 95			3367	0	0	0	48	0	FALSE	FALSE
# 96			1335	0	0	0	80	0	FALSE	FALSE
# 97	Chr1 2492887	G								
# 98			1007	0	22	0	0	0	FALSE	FALSE
# 99			1012	0	61	0	0	0	FALSE	FALSE
# 100			1013	0	35	0	0	0	FALSE	FALSE
# 101			1014	0	17	0	0	0	FALSE	FALSE
# 102			1015	0	55	0	0	0	FALSE	FALSE
# 103			3367	0	50	0	0	0	FALSE	FALSE
# 104			1335	0	85	0	0	0	FALSE	FALSE
# 105	Chr1 2497794	T								
# 106			1007	0	0	0	35	0	TRUE	FALSE
# 107			1012	0	0	0	60	0	TRUE	FALSE
# 108			1013	0	0	0	58	0	TRUE	FALSE
# 109			1014	0	0	0	12	0	TRUE	FALSE
# 110			1015	0	0	0	64	0	TRUE	FALSE
# 111			3367	0	0	0	43	0	TRUE	FALSE
# 112			1335	0	0	0	107	0	TRUE	FALSE
# 113	Chr1 2500122	A								
# 114			1007	18	0	0	0	0	FALSE	FALSE
# 115			1012	47	0	0	0	0	FALSE	FALSE
# 116			1013	34	0	0	0	0	FALSE	FALSE
# 117			1014	6	0	0	0	0	FALSE	FALSE
# 118			1015	35	0	0	0	0	FALSE	FALSE
# 119			3367	27	0	0	0	0	FALSE	FALSE
# 120			1335	51	0	0	0	0	FALSE	FALSE
# 121	Chr1 2503000	T								
# 122			1007	0	0	0	29	0	FALSE	FALSE
# 123			1012	0	0	0	35	0	FALSE	FALSE
# 124			1013	0	0	0	57	0	FALSE	FALSE
# 125			1014	0	0	0	10	0	FALSE	FALSE
# 126			1015	0	0	0	34	0	FALSE	FALSE
# 127			3367	0	0	0	41	0	FALSE	FALSE
# 128			1335	0	0	0	28	0	FALSE	FALSE
# 129	Chr1 2507585	A								
# 130			1007	34	0	0	0	0	TRUE	FALSE
# 131			1012	55	0	0	0	0	TRUE	FALSE
# 132			1013	32	0	0	0	0	TRUE	FALSE
# 133			1014	13	0	0	0	0	TRUE	FALSE
# 134			1015	41	0	0	0	0	TRUE	FALSE
# 135			3367	61	0	0	0	0	TRUE	FALSE
# 136			1335	104	0	0	0	0	TRUE	FALSE
# 137	Chr1 2507680	A								
# 138			1007	26	0	0	0	0	FALSE	FALSE

```

# 139      1012  46  0  0  0  0  0 FALSE FALSE
# 140      1013  32  0  0  0  0  0 FALSE FALSE
# 141      1014  15  0  0  0  0  0 FALSE FALSE
# 142      1015  54  0  0  0  0  0 FALSE FALSE
# 143      3367  51  0  0  0  0  0 FALSE FALSE
# 144      1335  78  0  0  0  0  0 FALSE FALSE
# 145 Chr1 2510117  C
# 146      1007   0  0  19  0  0  0 TRUE  FALSE
# 147      1012   0  0  56  1  0  0 TRUE  FALSE
# 148      1013   0  0  42  0  0  0 TRUE  FALSE
# 149      1014   0  0  13  0  0  0 TRUE  FALSE
# 150      1015   0  0  39  0  0  0 TRUE  FALSE
# 151      3367   0  0  36  0  0  0 TRUE  FALSE
# 152      1335   0  0  92  0  0  0 TRUE  FALSE
# 153 Chr1 2513923  A
# 154      1007  39  0  0  0  0  0 FALSE FALSE
# 155      1012  57  0  0  0  0  0 FALSE FALSE
# 156      1013  23  0  0  0  0  0 FALSE FALSE
# 157      1014   4  0  0  0  0  0 FALSE FALSE
# 158      1015  39  0  0  0  0  0 FALSE FALSE
# 159      3367  53  0  0  0  0  0 FALSE FALSE
# 160      1335  53  0  0  0  0  0 FALSE FALSE

```

Position 1088766, however, in a good example of the situation that motivated this analysis—one strain has a G/C SNP and 5 of the other 6 strains have nonreference reads consistent with that SNP. Although, excluding 1015, the nonreference read counts are not high enough to justify a SNP call in any strain considered in isolation, the fact that they *consistently* agree with the 1015 SNP suggests that they are real. One alternative hypothesis is that there is some sequence-dependent bias at this locus that favors misreading a G as a C. On the other hand, one could equally well posit a shared SNP, and a locus-dependant bias that *supresses* C reads, explaining the unbalanced readout that we observe. However, it is hard to reconcile either view with the significant strain-specific patterns that we see in the shared SNPs (as seen below). I think a more likely explanation is that (a) there are some number of relatively rare SNPs present in each of the sampled populations, (b) some of these SNPs happened to be present in one or two cells of the roughly 5-10 cells that we believe constituted the founding population of the culture grown for sequencing, and (c) stochastic effects during culture growth and during sequencing may have further perturbed the apparent frequency of each variant, but the bottom line is that the above-threshold presence of consistent non-reference reads is evidence for shared SNPs at the population level (and the proportions of such reads represent estimates of the population-level frequencies of the variants, albeit a noisy estimate at any specific position).

An aside: I was curious to see whether there is any consistent pattern to positions that are called consistent SNPs in all but Italy, so I repeated the above, basically. My summary is that coverage in Italy tends to be below average in these positions, but otherwise they don't stand out. For the record:

```

abit <- snp.pattern[[2]]==125
abit[is.na(abit)]<-F
sum(abit)

# [1] 13630

rabit <- rownames(non.refs[[2]])[which(abit)]
rabits <- rabit[1:20]
cabit <- as.integer(unlist(lapply(strsplit(rabits,','),function(x){x[2]})))
cabit

# [1] 1244 1575 6485 7181 7220 7661 8144 8208 8518 8552 8567 8670 8685 14361 15254
# [16] 15280 16103 25546 30784 33852

seecounts(cabit,snp.tables=snp.tables)

#      chr   pos Ref Strain  A  G  C  T SNP  exon indel nrf rat
# 1  Chr1 1244  G
# 2      1007  2 25  0  0  0  TRUE FALSE
# 3      1012  3 32  0  0  0  TRUE FALSE
# 4      1013 10 24  0  0  1  TRUE FALSE
# 5      1014  3 17  0  0  0  TRUE FALSE
# 6      1015 15 43  0  0  1  TRUE FALSE
# 7      3367  0  1  0  0  0  TRUE FALSE
# 8      1335 82 65  0  0  1  TRUE FALSE
# 9  Chr1 1575  G

```

# 10				1007	24	7	0	0	0	TRUE	FALSE
# 11				1012	42	13	0	0	0	TRUE	FALSE
# 12				1013	17	16	0	0	0	TRUE	FALSE
# 13				1014	15	4	0	0	0	TRUE	FALSE
# 14				1015	43	31	0	0	1	TRUE	FALSE
# 15				3367	0	2	0	0	0	TRUE	FALSE
# 16				1335	34	74	0	0	0	TRUE	FALSE
# 17	Chr1	6485	G								
# 18				1007	24	19	0	0	0	TRUE	FALSE
# 19				1012	29	29	0	0	0	TRUE	FALSE
# 20				1013	49	33	0	0	0	TRUE	FALSE
# 21				1014	6	5	0	0	0	TRUE	FALSE
# 22				1015	31	32	0	0	1	TRUE	FALSE
# 23				3367	0	37	0	0	0	TRUE	FALSE
# 24				1335	62	52	0	0	0	TRUE	FALSE
# 25	Chr1	7181	G								
# 26				1007	0	30	29	0	0	TRUE	FALSE
# 27				1012	0	52	34	0	0	TRUE	FALSE
# 28				1013	0	19	72	0	0	TRUE	FALSE
# 29				1014	0	13	7	0	0	TRUE	FALSE
# 30				1015	0	40	33	0	1	TRUE	FALSE
# 31				3367	0	29	0	0	0	TRUE	FALSE
# 32				1335	0	78	73	0	0	TRUE	FALSE
# 33	Chr1	7220	C								
# 34				1007	16	0	19	6	0	TRUE	FALSE
# 35				1012	38	0	22	11	0	TRUE	FALSE
# 36				1013	82	1	30	9	0	TRUE	FALSE
# 37				1014	12	0	6	2	0	TRUE	FALSE
# 38				1015	55	0	22	5	1	TRUE	FALSE
# 39				3367	0	0	8	0	0	TRUE	FALSE
# 40				1335	55	0	32	20	0	TRUE	FALSE
# 41	Chr1	7661	T								
# 42				1007	0	0	9	9	0	TRUE	FALSE
# 43				1012	0	0	5	19	0	TRUE	FALSE
# 44				1013	0	0	24	14	1	TRUE	FALSE
# 45				1014	0	0	6	3	0	TRUE	FALSE
# 46				1015	0	0	5	34	0	TRUE	FALSE
# 47				3367	0	0	0	4	0	TRUE	FALSE
# 48				1335	0	0	4	24	0	TRUE	FALSE
# 49	Chr1	8144	G								
# 50				1007	8	9	0	0	0	TRUE	FALSE
# 51				1012	12	10	0	0	1	TRUE	FALSE
# 52				1013	38	29	0	0	0	TRUE	FALSE
# 53				1014	5	4	0	0	0	TRUE	FALSE
# 54				1015	15	16	0	0	0	TRUE	FALSE
# 55				3367	0	0	0	0	0	TRUE	FALSE
# 56				1335	12	15	0	0	1	TRUE	FALSE
# 57	Chr1	8208	G								
# 58				1007	0	6	0	7	1	TRUE	FALSE
# 59				1012	0	19	0	11	0	TRUE	FALSE
# 60				1013	0	1	0	48	0	TRUE	FALSE
# 61				1014	0	5	0	3	0	TRUE	FALSE
# 62				1015	0	19	0	11	1	TRUE	FALSE
# 63				3367	0	1	0	0	0	TRUE	FALSE
# 64				1335	0	2					

```

# 90      1007  7  0  0  5  0  TRUE FALSE
# 91      1012 16  0  0 10  0  TRUE FALSE
# 92      1013 16  0  0 11  0  TRUE FALSE
# 93      1014  2  0  0  4  0  TRUE FALSE
# 94      1015 14  0  0 10  1  TRUE FALSE
# 95      3367  5  0  0  0  0  TRUE FALSE
# 96      1335  7  0  0  6  0  TRUE FALSE
# 97 Chr1  8685  G
# 98      1007  6 15  0  0  0  TRUE FALSE
# 99      1012 10 23  0  0  0  TRUE FALSE
# 100     1013 18 21  0  0  1  TRUE FALSE
# 101     1014  4  8  0  0  0  TRUE FALSE
# 102     1015 10 24  0  0  1  TRUE FALSE
# 103     3367  0  4  0  0  0  TRUE FALSE
# 104     1335  5 32  0  0  0  TRUE FALSE
# 105 Chr1 14361  A
# 106     1007 20  7  0  0  0  FALSE FALSE
# 107     1012 35  5  0  0  0  FALSE FALSE
# 108     1013  1 11  0  0  1  FALSE FALSE
# 109     1014  6  2  0  0  0  FALSE FALSE
# 110     1015 35  7  0  0  0  FALSE FALSE
# 111     3367  2  1  0  0  0  FALSE FALSE
# 112     1335 50  8  0  0  0  FALSE FALSE
# 113 Chr1 15254  T
# 114     1007 11  0  0 16  1  FALSE FALSE
# 115     1012 26  0  0 38  1  FALSE FALSE
# 116     1013 37  0  0 48  1  FALSE FALSE
# 117     1014  3  0  0  8  1  FALSE FALSE
# 118     1015 18  0  0 32  1  FALSE FALSE
# 119     3367  0  0  0 73  0  FALSE FALSE
# 120     1335 13  0  0 32  1  FALSE FALSE
# 121 Chr1 15280  T
# 122     1007  0 13  0 20  1  FALSE FALSE
# 123     1012  0 27  0 28  1  FALSE FALSE
# 124     1013  0  5  0 64  0  FALSE FALSE
# 125     1014  0  2  0  8  0  FALSE FALSE
# 126     1015  0 19  0 29  1  FALSE FALSE
# 127     3367  0  0  0 42  0  FALSE FALSE
# 128     1335  0 21  0 70  1  FALSE FALSE
# 129 Chr1 16103  A
# 130     1007 10  0 11  0  1  FALSE FALSE
# 131     1012 44  0 19  0  1  FALSE FALSE
# 132     1013 21  0 13  0  1  FALSE FALSE
# 133     1014 14  0  2  0  0  FALSE FALSE
# 134     1015 29  0 10  0  1  FALSE FALSE
# 135     3367 33  0  0  0  0  FALSE FALSE
# 136     1335 47  0 11  0  0  FALSE FALSE
# 137 Chr1 25546  A
# 138     1007 23  0  0 14  1  FALSE FALSE
# 139     1012 46  0  0 19  1  FALSE FALSE
# 140     1013  6  0  0 42  1  FALSE FALSE
# 141     1014  7  0  0 15  1  FALSE FALSE
# 142     1015 52  0  0 17  1  FALSE FALSE
# 143     3367 60  0  0  0  0  FALSE FALSE
# 144     1335 67  0  0  5  0  FALSE FALSE
# 145 Chr1 30784  C
# 146     1007 16  0 13  0  1  TRUE FALSE
# 147     1012 33  0 32  0  1  TRUE FALSE
# 148     1013 19  0 33  0  1  TRUE FALSE
# 149     1014  4  0 11  0  1  TRUE FALSE
# 150     1015 39  0 29  0  1  TRUE FALSE
# 151     3367  0  0 55  0  0  TRUE FALSE
# 152     1335 46  0 50  0  1  TRUE FALSE
# 153 Chr1 33852  C
# 154     1007  0 24 25  0  1  FALSE FALSE
# 155     1012  0 18 26  0  1  FALSE FALSE
# 156     1013  0 28 33  0  1  FALSE FALSE
# 157     1014  0  9  4  0  1  FALSE FALSE
# 158     1015  0 19 28  0  1  FALSE FALSE
# 159     3367  0  0 26  0  0  FALSE FALSE
# 160     1335  0 30 53  0  1  FALSE FALSE

```

More sanity: there are 83 sites on Chr1 shared by zero strains in the tightest condition. (I.e., SAMTOOLS called it a SNP, but the read counts/proportions fall below our 3rd threshold). Are they due to low coverage? Seemingly yes:

```

zp3 <- snp.pattern[[3]] == 0
zr3 <- rownames(non.refs[[3]])[which(zp3)]
zc3 <- as.integer(unlist(lapply(strsplit(zr3[1:min(100,length(zr3))],':',fixed=TRUE),function(x){x[2]})))

```



```

zc3

# [1] 16115 16615 19117 25748 43500 55857 56591 65787 66879 68328 80862 81001 90622
# [14] 90721 91284 110754 116443 116453 120183 126702 127986 129056 147698 153874 159756 160912
# [27] 161271 170686 180314 181477 182139 196862 196864 199166 206132 206143 221888 234931 242276
# [40] 242914 244505 268954 274655 282391 282511 283646 289363 311952 312625 314132 326217 371008
# [53] 376784 387078 387091 389263 395153 406158 410771 431788 438958 438976 443898 447253 448223
# [66] 452774 488812 495476 498133 501830 501975 504462 506422 515441 515595 530113 530114 532320
# [79] 534149 541667 543095 575081 585297 586276 612732 622585 651159 652889 655373 655380 657704
# [92] 657955 658216 685697 687653 692115 692139 700484 700845 701061

seecounts(zc3[1:5], snp.tables=snp.tables)

# chr pos Ref Strain A G C T SNP exon indel nrf rat
# 1 Chr1 16115 T
# 2 1007 0 0 0 5 0 FALSE FALSE
# 3 1012 0 0 0 9 0 FALSE FALSE
# 4 1013 0 0 0 6 0 FALSE FALSE
# 5 1014 0 0 0 3 0 FALSE FALSE
# 6 1015 0 0 0 10 0 FALSE FALSE
# 7 3367 0 0 3 3 1 FALSE FALSE
# 8 1335 0 0 0 6 0 FALSE FALSE
# 9 Chr1 16615 C
# 10 1007 0 0 39 0 0 FALSE FALSE
# 11 1012 0 0 54 0 0 FALSE FALSE
# 12 1013 0 0 4 2 1 FALSE FALSE
# 13 1014 0 0 19 0 0 FALSE FALSE
# 14 1015 0 0 46 0 0 FALSE FALSE
# 15 3367 0 0 13 0 0 FALSE FALSE
# 16 1335 0 0 40 0 0 FALSE FALSE
# 17 Chr1 19117 A
# 18 1007 16 0 0 0 0 TRUE FALSE
# 19 1012 21 0 0 0 0 TRUE FALSE
# 20 1013 1 0 0 1 0 TRUE FALSE
# 21 1014 6 0 0 0 0 TRUE FALSE
# 22 1015 21 0 0 0 0 TRUE FALSE
# 23 3367 0 0 0 1 1 TRUE FALSE
# 24 1335 24 0 0 0 0 TRUE FALSE
# 25 Chr1 25748 C
# 26 1007 0 0 17 0 0 FALSE FALSE
# 27 1012 0 0 36 0 0 FALSE FALSE
# 28 1013 3 0 7 0 1 FALSE FALSE
# 29 1014 1 0 4 0 0 FALSE FALSE
# 30 1015 0 0 32 0 0 FALSE FALSE
# 31 3367 0 0 1 0 0 FALSE FALSE
# 32 1335 1 0 34 0 0 FALSE FALSE
# 33 Chr1 43500 A
# 34 1007 10 0 0 3 1 FALSE FALSE
# 35 1012 10 0 0 3 1 FALSE FALSE
# 36 1013 10 0 1 1 0 FALSE FALSE
# 37 1014 5 0 0 0 0 FALSE FALSE
# 38 1015 11 0 0 2 0 FALSE FALSE
# 39 3367 6 0 0 3 0 FALSE FALSE
# 40 1335 13 0 0 1 0 FALSE FALSE

```

7.3 Main Analysis

Turning to the main analysis, there is a large increase in the number of consistent positions between the loose and medium stringency levels; medium and tight are similar in most respects. The likely interpretation is that the loose criterion is including many “SNPs” induced by read errors, and that either of the tighter criteria are successfully filtering them out. In the interest of simplicity, the narrative below will focus on the shared SNPs at the medium stringency level (the “count2” column in the data frame), although the numbers for all three (sometimes all 4) are displayed. Also note that the prose and some comments in the code were based on the Chr1 analysis, and so may occasionally be off-target for the whole-genome data.

```

# Show a subset of pat.summaries, optionally with totals of count_i in last row, and optionally
# aggregating low-count rows as ``Other''
#
#   sharedBy=c(2,4) selects SNPs shared by 2 or 4 strains,
#   subset=as.octmode('35') select those with sharing pattern a subset (optionally proper) of this
#   split=as.octmode('14') additionally restricts to patterns straddling split/subset minus split
#   c2.thresh=42 suppresses printout of rows with count2 < 42
#   restrict.to=c(0,42,127) restrict to these 3 rows
showgroup <- function(p.summ=pat.summaries, sharedBy=0:7, subset=127, split=NULL, proper.subset=F,
                      total=T, c2.thresh=0, fourteenth=F, restrict.to=NULL){
  # pick just those bit patterns that are subsets of 'subset'
  pick <- bitwAnd(0:127,bitwNot(subset))==0
  if(proper.subset){
    pick[subset+1] <- F
  }
  if(!is.null(split)){ # AND that straddle left/right subtrees
    cosplit <- bitwAnd(subset,bitwNot(split))
    pick <- pick & bitwAnd(0:127,split)!=0 & bitwAnd(0:127,cosplit)!=0
  }
  # and have desired shareBy counts
  pick <- pick & (p.summ$sharedBy %in% sharedBy)
  # and are among the set of interest
  if(!is.null(restrict.to)){
    pick <- pick & (0:127 %in% restrict.to)
  }
  # find rows with low counts
  pick.low <- pick & (p.summ$count2 < c2.thresh)
  # now show them
  show <- p.summ[pick & !pick.low,]
  # rename columns just to narrow the printouts
  colnames(show) <- c('Pat','ShrBy','1007','1012','1013','1014','1015','3367','1335',
                     'count1','count2','count3','count4')
  show[,1] <- format(show[,1]) # convert octal col to char so can override in last row(2)
  nlow <- sum(pick.low)
  if(nlow > 0){
    n <- nrow(show)+1
    lows <- apply(p.summ[pick.low,10:13],2,sum)
    show[n,10:13] <- lows
    show[n,1:9] <- ''
    row.names(show)[n] <- 'Other'
    if(fourteenth){
      # do this: add 14th col just to hold this comment:
      show <- cbind(show, ' ', stringsAsFactors=F)
      show[n,14] <- paste('(', nlow, 'rows w/ c2 < ', c2.thresh, ')')
    } else {
      ## or this (looks a bit funky, but fits across page without line-wrap):
      show[n,1:8] <- c('(', nlow, 'rows', 'w/', 'c2', '<', c2.thresh, ')')
    }
  }
  if(total){
    n <- nrow(show)+1
    tots <- apply(show[,10:13],2,sum)
    show[n,10:13] <- tots
    show[n,1:9] <- ''
    row.names(show)[n] <- 'Total'
    if(ncol(show)==14){show[n,14] <- ''}
  }
  return(show)
}

```

First, are there any SNPs that are not “consistent SNPs?” Yes, a few in c3. As noted above, they seem to be mainly low-coverage positions.

```

showgroup(pat.summaries,0,total=F) # chr1 totals: 0 0 83

# Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 1 0 0 111 578 4755 0

```

Next, look at completely shared SNPs, those found in all 7 strains.

```
showgroup(pat.summaries,7,total=F) # Chr1 count1 = 8593, count2 = 7054, count3 = 4790 c4=1641
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 128	177	7	X	X	X	X	X	X	X	77690	62182	38744	15186

I.e., of the 469906 consistent positions, 62182 or 13.2% are shared by all 7 strains.

Next look at singletons, aka private SNPs—SNPs that are called in one strain and no other strain has a significant number of non-ref reads at that position. Presumably these are variants that arose in a given population after it separated from the others.

```
showgroup(pat.summaries,1) # chr1 totals: 9669 18865 19670 23574
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 2	001	1							X	449	632	1129	2260
# 3	002	1						X		73721	85117	87494	105614
# 5	004	1					X			1720	2156	2729	4608
# 9	010	1				X				383	525	485	1231
# 17	020	1			X					82364	94364	96464	113191
# 33	040	1		X						502	655	1102	2450
# 65	100	1	X							231	339	496	2005
# Total										159370	183788	189899	231359

The import of shared/private SNPs changes between sexual and asexual populations. Presumably asexuals slowly gain and rarely lose private SNPs; shared ones predate separation of the lineages. In sexual lineages, however, SNPs may be rather freely “gained” or “lost,” merely by recombination (converting between homo- and heterozygous in the sample we sequenced). Thus, the low private counts for the 5 L-isolates compared to the large count of het positions overall suggest that (a) they are asexual, and (b) none of them has been isolated from the others for very long (if at all). Conversely, the high counts for Italy and Wales suggest that (a) if asexual, they have been separated from each other and from the rest for a long time, but (b) if sexual, there is little surprise: we have $\approx 160\text{K}$ SNPs shared between the two (90K just in those two (below), plus 70K shared by all 7), and $\approx 90\text{K}$ additional positions that are het in one but not the other. These are close to, but not exactly equal to, the 1:2:1 ratios we would naively expect from two samples of a single HWE population. The most parsimonious explanation seems to be that the H-clade is sexual, but perhaps some het positions private to each population separates them.

Aside: counts of “consistent” SNPs minus these singletons yeilds count of shared SNPs:

```
singlets <- apply(pat.summaries[pat.summaries$sharedBy==1,10:13],2,sum)
rbind(consistent=consistent.count,singlets=singlets,shared=consistent.count-singlets)
```

#	count1	count2	count3	count4
# consistent	447177	469906	471171	474613
# singlets	159370	183788	189899	231359
# shared	287807	286118	281272	243254

The slightly higher count of shared positions in the medium case further supports this choice for subsequent analysis.

Next look at consistent SNPs shared between just a pair of isolates.

```
showgroup(pat.summaries,2) # chr 1 counts: 7641 9549 9472 6924
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 4	003	2						X	X	994	298	532	587
# 6	005	2					X		X	287	523	1088	1407
# 7	006	2					X	X		624	138	282	590
# 10	011	2				X			X	515	486	317	827
# 11	012	2				X		X		565	49	32	93
# 13	014	2				X	X			133	139	132	417
# 18	021	2		X					X	998	167	337	402
# 19	022	2		X				X		82160	87499	83482	58009
# 21	024	2		X			X			686	195	410	625
# 25	030	2		X	X					609	69	47	93

# 34	041	2		X					X	42	92	313	368
# 35	042	2		X				X		503	119	254	394
# 37	044	2		X			X			69	279	1001	1809
# 41	050	2		X		X				13	24	53	105
# 49	060	2		X	X					627	116	237	388
# 66	101	2	X						X	29	47	73	314
# 67	102	2	X					X		351	67	96	351
# 69	104	2	X				X			39	122	329	1196
# 73	110	2	X			X				12	11	29	150
# 81	120	2	X		X					432	76	98	309
# 97	140	2	X	X						955	1144	1235	2144
# Total										90643	91660	90377	70578

I.e., of the 91660 paired SNPs, 87499 or 95.5% are found between Italy and Wales, with comparatively few shared between any other pairs (only).

SNPs shared among exactly 3 isolates are relatively rare. (The 5 trios containing both Italy and Wales predominate in the loose set, probably because they share many pairs that become triples with the addition of a few read errors.)

```
showgroup(pat.summaries,3) # chr 1 counts: 1438 294 671 1034
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 8	007	3					X	X	X	104	197	371	557
# 12	013	3				X		X	X	257	226	146	338
# 14	015	3				X	X		X	1041	984	660	1389
# 15	016	3				X	X	X		46	60	49	152
# 20	023	3			X			X	X	1274	558	1020	533
# 22	025	3			X		X		X	135	216	466	522
# 23	026	3			X		X	X		793	431	763	789
# 26	031	3			X	X			X	268	233	151	361
# 27	032	3			X	X		X		698	131	74	86
# 29	034	3			X	X	X			103	119	91	219
# 36	043	3		X				X	X	56	86	151	133
# 38	045	3		X			X		X	202	593	1970	1656
# 39	046	3		X			X	X		58	154	425	604
# 42	051	3		X		X			X	52	57	74	126
# 43	052	3		X		X		X		8	13	17	22
# 45	054	3		X		X	X			20	78	133	292
# 50	061	3		X	X				X	52	80	131	115
# 51	062	3		X	X			X		703	269	454	469
# 53	064	3		X	X		X			53	184	458	601
# 57	070	3		X	X	X				22	9	14	24
# 68	103	3	X					X	X	24	34	46	143
# 70	105	3	X				X		X	78	181	396	805
# 71	106	3	X				X	X		32	66	109	377
# 74	111	3	X			X			X	6	11	8	139
# 75	112	3	X			X		X		10	11	8	26
# 77	114	3	X			X	X			12	36	56	365
# 82	121	3	X	X					X	22	22	34	73
# 83	122	3	X		X			X		501	162	165	354
# 85	124	3	X		X		X			43	88	152	400
# 89	130	3	X		X	X				9	9	9	27
# 98	141	3	X	X					X	78	149	258	519
# 99	142	3	X	X				X		386	409	463	755
# 101	144	3	X	X			X			383	1176	2395	4432
# 105	150	3	X	X		X				28	51	55	238
# 113	160	3	X	X	X					337	375	399	712
# Total										7894	7458	12171	18353

Four-way sharing is more common, but dominated by the coastal (i.e., non-Gyre) L-clade isolates. This is likely a reflection of the strong 5-way sharing among the L-clade, from which the Gyre commonly drops out due to the lower coverage/higher error rate in that sequencing run.

```
showgroup(pat.summaries,4) # chr 1 counts: 564 1346 2552 3479
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
---	-----	-------	------	------	------	------	------	------	------	--------	--------	--------	--------

# 16	017	4				X	X	X	X	390	329	211	564
# 24	027	4			X		X	X	X	461	759	1423	771
# 28	033	4			X	X		X	X	1139	973	574	306
# 30	035	4			X	X	X		X	578	509	329	503
# 31	036	4			X	X	X	X		345	320	227	211
# 40	047	4		X				X	X	127	256	708	708
# 44	053	4		X		X		X	X	35	34	26	56
# 46	055	4		X		X	X		X	606	696	668	971
# 47	056	4		X		X	X	X		26	44	50	88
# 52	063	4		X	X			X	X	151	184	332	194
# 54	065	4		X	X			X	X	122	284	731	582
# 55	066	4		X	X		X	X		217	489	1025	851
# 58	071	4		X	X	X			X	9	20	7	28
# 59	072	4		X	X	X		X		41	36	21	31
# 61	074	4		X	X	X	X			20	46	51	116
# 72	107	4	X					X	X	58	84	129	330
# 76	113	4	X			X		X	X	7	9	5	66
# 78	115	4	X			X	X		X	141	139	122	604
# 79	116	4	X			X	X	X		8	8	11	101
# 84	123	4	X		X			X	X	63	98	91	124
# 86	125	4	X		X		X		X	67	113	223	283
# 87	126	4	X		X		X	X		99	198	268	425
# 90	131	4	X		X	X			X	6	3	0	52
# 91	132	4	X		X	X		X		19	21	10	38
# 93	134	4	X		X	X	X			18	17	22	143
# 100	143	4	X	X				X	X	37	58	103	190
# 102	145	4	X	X			X		X	5992	12644	22332	23189
# 103	146	4	X	X			X	X		196	510	969	1795
# 106	151	4	X	X		X			X	43	69	54	220
# 107	152	4	X	X		X		X		18	27	15	67
# 109	154	4	X	X		X	X			1227	1390	1065	1738
# 114	161	4	X	X	X				X	74	96	113	207
# 115	162	4	X	X	X			X		1848	1932	1828	1014
# 117	164	4	X	X	X		X			237	627	1053	1752
# 121	170	4	X	X	X	X				13	18	15	69
# Total										14438	23040	34811	38387

Five-way sharing is much more common, and is strongly dominated by the 5 L-clade isolates.

showgroup(pat.summaries,5) # chr 1 counts: 3969 5047 4624 6125													
#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 32	037	5			X	X	X	X	X	2247	1877	1193	620
# 48	057	5		X		X	X	X	X	221	219	189	324
# 56	067	5		X	X		X	X	X	556	1015	2544	1151
# 60	073	5		X	X	X		X	X	94	72	62	38
# 62	075	5		X	X	X	X		X	195	187	210	328
# 63	076	5		X	X	X	X	X		106	130	124	128
# 80	117	5	X			X	X	X	X	48	32	25	241
# 88	127	5	X		X		X	X	X	225	321	501	482
# 92	133	5	X		X	X		X	X	31	28	15	52
# 94	135	5	X		X	X	X		X	126	117	88	235
# 95	136	5	X		X	X	X	X		34	56	26	106
# 104	147	5	X	X			X	X	X	2073	4042	6741	10001
# 108	153	5	X	X		X		X	X	39	27	26	96
# 110	155	5	X	X		X	X		X	40157	35344	22417	30602
# 111	156	5	X	X		X	X	X		565	575	410	735
# 116	163	5	X	X	X			X	X	255	271	328	316
# 118	165	5	X	X	X		X		X	2726	5022	8440	9715
# 119	166	5	X	X	X		X	X		902	1976	3172	2688
# 122	171	5	X	X	X	X			X	41	19	13	70
# 123	172	5	X	X	X	X		X		58	71	45	86
# 125	174	5	X	X	X	X	X			659	682	468	782
# Total										51358	52083	47037	58796

Six-way sharing is also common, with the sets *excluding* Gyre, Italy, or Wales having the most mutually-shared SNPs.

```
showgroup(pat.summaries,6) # chr 1 counts: 4166 4741 5312 4722
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 64	077	6		X	X	X	X	X	X	850	847	827	485
# 96	137	6	X		X	X	X	X	X	405	324	240	333
# 112	157	6	X	X			X	X	X	13239	10814	6862	12202
# 120	167	6	X	X	X		X	X	X	11742	21003	35227	15091
# 124	173	6	X	X	X	X		X	X	131	87	47	114
# 126	175	6	X	X	X	X	X		X	16884	13630	8608	12697
# 127	176	6	X	X	X	X	X	X		2422	2412	1566	1032
# Total										45673	49117	53377	41954

8 Trees

So, overall, the picture looks like a long shared history (62182 7-way shared positions), followed by a split of the 5 L-isolates from the 2 H-isolates, then a long shared history in the 5 (35344 quintuples), in parallel with a long shared history in H- (87499 pairs), then separate histories in Italy and Wales (>85117 “private” SNPs in each, although again if they are sexual, many of these just reflect HWE), and very limited differentiation among the 5 L-isolates.

Branch lengths of course depend on filtering criteria used (and, of course, full vs Chr1 differ by about a factor of 10), but the tree *topology* appears to be fairly stable. Various versions are drawn below, exactly to explore how robust this story is. I think we should go with “medium stringency” SNP filtering (based on un-qfiltered reads).

NOTE: Much of this analysis make less sense for q-filtered read data, since (a) the point of the SNP filtering was to try to correct for noise in the raw reads, which may (or may not; haven’t looked closely, yet) be largely fixed by qfiltering (e.g., “loose” or no SNP filtering may be more appropriate, post-q-filtering, esp. if we had re-run SAMTools to call SNPs based on the q-filtered reads), and (b) tree topology *does* appear to change, in that Gyre’s coverage has been so sharply reduced by qfiltering that it clearly stands aside from the others (and that’s confirmed by bootstrap), but this also seems to be clearly a technical rather than a biological artifact. SO, code below will run on q-filtered data, but *is not tuned to it*. Likewise, most comments in the prose below were made to describe the un-q-filtered data, and *are misleading and in some cases flatly wrong* for qfiltered data, but it doesn’t seem worthwhile to bother with a rewrite...

Trees are coded in newick format, which doesn’t seem to tolerate line-breaks; print with line-wrap:.

```
# wrap a long char string across multiple lines in printout
cat.hardwrap <- function(str,width=80){
  while(nchar(str)>width){
    cat(substr(str,1,width),'\n')
    str <- substr(str,width+1,nchar(str))
  }
  cat(str,'\n')
}
```

Trees are built as follows. Code for drawing, especially, is specific to the topology of the medium tree, and placement of some of the figure elements have been hand-optimized for this case; drawings for the other variants will not be as pretty.

```
# set up for tree figs

# the newick parser in ape seems to be confused by commas and parens in
# tip names, and blanks are not allowed, so replace by *, <, >, _, resp.
newick.name <- function(name){
  name <- gsub(' ', '_', name, fixed=TRUE)
  name <- gsub(' ', '*', name, fixed=TRUE)
  name <- gsub('(', '<', name, fixed=TRUE)
  name <- gsub(')', '>', name, fixed=TRUE)
  return(name)
}

# undo above changes
newick.name.undo <- function(name){
  #name <- gsub('_', ' ', name, fixed=TRUE) # unnecessary; ape plot routine handles this one
  name <- gsub('*', ' ', name, fixed=TRUE)
  name <- gsub('<', '(', name, fixed=TRUE)
  name <- gsub('>', ')', name, fixed=TRUE)
}
```

```

return(name)
}

# make a newick string from tree; see it below
# 'pre' is prefixed to ccmpid; 'nb' optionally included;
# 'alt' can be used instead of pre/ccmp/nb/where for less formal labeling
# 'newstyle'==T => new node label: [nb_]where[(pre-less-id)]
# 'newstyle'==F => old node label: [nb_] [pre id]where
newickize <- function(tree,pre='CCMP',nb=TRUE,alt=F,newstyle=TRUE){
  if(is.null(tree$where)){
    # not a leaf; paste together newick from subtrees
    sub1 <- newickize(tree$sub1,pre=pre,nb=nb,alt=alt,newstyle=newstyle)
    sub2 <- newickize(tree$sub2,pre=pre,nb=nb,alt=alt,newstyle=newstyle)
    new <- paste('(', sub1, ',', sub2, ')', sep='')
    if(!is.null(tree$length)){
      # internal node, add length
      return(paste(new, ':', tree$length, sep=''))
    } else {
      # top level; escape blanks and add trailing ';'
      return(paste(gsub(' ', '_', new), ';', sep=''))
    }
  } else {
    # a leaf; build label and branch length
    if(alt){
      # label is just alt; if alt omitted, default to where
      new <- newick.name(ifelse(is.null(tree$alt), tree$where, tree$alt))
    } else {
      if(newstyle){
        # new node label = [nb_]where[(pre-less-id)]
        new <- ifelse(nb && !is.null(tree$nb), paste(tree$nb, '_', sep=''), '')
        new <- newick.name(paste(new, tree$where, sep=''))
        new <- ifelse(is.null(tree$id), new, paste(new, '(', tree$id, ')', sep=''))
        new <- newick.name(new)
      } else {
        # old style node label = [nb_] [pre id]where
        new <- ifelse(nb && !is.null(tree$nb), paste(tree$nb, '_', sep=''), '')
        new <- ifelse(is.null(tree$id), new, paste(new, pre, tree$id, '_', sep=''))
        new <- newick.name(paste(new, tree$where, sep=''))
      }
    }
    #add length to either
    new <- paste(new, ':', tree$length, sep='')
  }
  return(new)
}

# Make a tree as nested lists, **based on the chr1, count2 topology**, but using any of the counts.
# Internal nodes have subtrees sub1/2 and length
# Root has sub1/2, but no length
# Leaves have where, length, optionally, id, alt, nb. (Omit id for 'outgroup'. Use 'alt' for less formal
# labeling in cartoon version; it defaults to 'where'. Use 'nb' to add abcde annotations for legend.)
# The single parameter v is any of the 4 count vectors contained in pat.summaries (most conveniently
# indexed in octal). E.g., make.tree(pat.summaries[, 'count2']) reproduces the count2 tree.
# (This was previously built by hand-pasting the edge lengths; tree.by.hand is retained in appendix
# for comparison, & its counts are in comments below).
#
make.tree <- function(v){
  pat.count <- function(pat, pat.counts=v){return(pat.counts[1+strtoi(pat,8)])}
  thetree <-
    list(
      sub1 = list(
        sub1 = list(
          sub1 = list(id=3367, length=pat.count('002'), where='Venice, Italy', alt='Venice'), #8813
          sub2 = list(id=1013, length=pat.count('020'), where='Wales, UK'), #9652
          length=pat.count('022')), #9365
        sub2 = list(
          sub1 = list(
            sub1 = list(
              sub1 = list(id=1007, length=pat.count('100'), nb='e', where='Virginia, USA'), #30
              sub2 = list(id=1012, length=pat.count('040'), nb='d', where='Perth, W. Australia', alt='Perth'), #61
              length=pat.count('140')), #19
            sub2 = list(
              sub1 = list(id=1015, length=pat.count('004'), nb='c', where='Washington, USA', alt='Puget Sound'), #207
              sub2 = list(id=1335, length=pat.count('001'), nb='b', where='New York, USA', alt='NY'), #41
              length=pat.count('005')), #18
              length=pat.count('145')), #1005
            sub2 = list(id=1014, length=pat.count('010'), nb='a', where='N. Pacific Gyre'), #61
              length=pat.count('155')), #3912
              length=pat.count('177')), #7054
          sub2 = list(length=0, where='outgroup')
        )
      )
    )
}

```

```

    )
    return(thetree)
}

```

Code to plot a tree given newick description. Again, code is somewhat general, but has some specializations tied to the medium-stringency, full-genome, un-filtered data.

```

# run following 2 lines after an R upgrade
# update.packages()
# install.packages("ape")
library(ape)
show.tree <- function(newick.str=newick.medium,
  col.edge = 'darkblue', lwd.edge = 2,
  col.elabel='darkblue', cex.elabel=0.8, font.elabel=3,
  col.arrow = 'red', lwd.arrow=1.5, cex.arrow = 0.9, font.arrow = 4,
  col.clade = 'black', lwd.clade=1, cex.clade = 1.0, font.clade = 3,
  col.legend='beige', cex.legend=0.8,
  col.tip = 'darkblue', font.tip = 4,
  plusx=FALSE, pltdebug=FALSE, total.snps=consistent.count[2]){

  ####
  #
  # ADJUST NEWICK & GET LENGTHS, COORDINATES
  #
  newick.str.noout <- sub('outgroup','_',newick.str) # Hide outgroup ('_' prints as blank)
  the.tree <- read.tree(text=newick.str.noout)

  ## nasty hack: ape's newick parser seems to be confused by commas, () in tip labels, so
  ## newickize replaced them by '*<>'; before plotting, I want to convert them back, and hope
  ## this doesn't break anything else... And if a revised version of ape changes the internal
  ## representation of a tree, this may need to be redone.
  the.tree$tip.label <- newick.name.undo(the.tree$tip.label)

  # extract branch lengths as char string of comma-separated numbers via pattern matching hack:
  # lengths always preceded by colon
  lengths.ch <- strsplit(paste(newick.str,':'),'^0-9[^:]*:')[[1]]

  # then convert to ints, dropping empty string at front
  lengths.int <- scan(what=integer(),quiet=T,sep=',',text=lengths.ch[-1])

  # then to data frame with named rows; a..g are terminal branches; others are internal.
  # a..e match legend in plot; f/g = wales/italy. lengths appear in postfix order of
  # newick tree, and ape draws the 1st of them at the bottom of the plot.
  lmed <- data.frame(lengths=lengths.int,
    row.names=c('g','f','fg','e','d','de','c','b','bc','bcde','a','abcde','all','out'))

  # extract counts needed for legend:
  #leg.counts <- c( 61, 41,207, 61, 30, 1005, 18, 19) #by hand, medium chr1
  leg.counts <- lmed[c('a','b','c','d','e','bcde','bc','de'),1]
  discord <- total.snps - sum(lmed$lengths)

  #tree.labels <- list( ## x,y,text; coords are all picked by eye
  # 3000, 3.62, paste(lmed['all',1], 'shared by 7', sep='\n'), # 7054
  # 8900, 5.75, paste(lmed['abcde',1], 'by 5', sep='\n'), # 3912
  # 12000, 1.50, paste(lmed['fg',1], 'shared by 2', sep='\n'), # 9365
  # 21000, 2.00, paste(lmed['f',1], 'only\nin Wales'), # 9652
  # 21000, 1.00, paste(lmed['g',1], 'only\nin Italy'), # 8813
  # 11500, 4.50, '*')
  # automating x-placement, below; retain above for comparison...
  tip <- integer(7) # x coords of tree tips
  tip[1] <- sum(lmed[c('all','fg','g'),1])
  tip[2] <- sum(lmed[c('all','fg','f'),1])
  tip[3] <- sum(lmed[c('all','abcde','bcde','de','e'),1])
  tip[4] <- sum(lmed[c('all','abcde','bcde','de','d'),1])
  tip[5] <- sum(lmed[c('all','abcde','bcde','bc','c'),1])
  tip[6] <- sum(lmed[c('all','abcde','bcde','bc','b'),1])
  tip[7] <- sum(lmed[c('all','abcde','a'),1])

  inode <- integer(5) # x coords of (some) internal nodes
  inode[1] <- 0 # root
  inode[2] <- lmed['all',1] # lca of all
  inode[3] <- sum(lmed[c('all','fg'),1]) # lca H-clade
  inode[4] <- sum(lmed[c('all','abcde'),1]) # lca L-clade
  inode[5] <- sum(lmed[c('all','abcde','bcde'),1]) # lca L-clade, nonGyre
  tree.labels <- list( ## x,y,text; y coords partially picked by eye
    sum(inode[c(1,2)])/2, 3.62, paste(lmed['all',1], 'shared by 7', sep='\n'), # 7054
    sum(inode[c(2,4)])/2, 5.75, paste(lmed['abcde',1], 'by 5', sep='\n'), # 3912
    sum(inode[c(2,3)])/2, 1.50, paste(lmed['fg',1], 'shared by 2', sep='\n'), # 9365
    (inode[3]+tip[2])/2, 2.00, paste(lmed['f',1], 'only\nin 1013'), # 9652

```



```

(inode[3]+tip[1])/2, 1.00, paste(lmed['g' ,1], 'only\nin 3367'), # 8813
sum(inode[c(4,5)]/2, 4.35, '*' )

tree.labels <- list( ## x,y,text; y coords partially picked by eye
  sum(inode[c(1,2)]/2, 3.62, paste(lmed['all' ,1], 'in 7', sep='\n'), # 7054
  sum(inode[c(2,4)]/2, 5.75, paste(lmed['abcde',1], 'in 5', sep='\n'), # 3912
  sum(inode[c(2,3)]/2, 1.50, paste(lmed['fg' ,1], 'in 2', sep='\n'), # 9365
  (inode[3]+tip[2])/2, 2.00, paste(lmed['f' ,1], 'only\nin 1013'), # 9652
  (inode[3]+tip[1])/2, 1.00, paste(lmed['g' ,1], 'only\nin 3367'), # 8813
  sum(inode[c(4,5)]/2, 4.35, '*' )

####
#
# BOGUS PLOT
#
# a messy bit: need string widths to set xlim; but strwidth needs x-scale so must plot first.
# M plot completely invisible, overlay 2nd plot via par(new=F...) .
#
# PROVISIONALLY set x.lim here at about 30% wider than tree; fine tune it for the real plot
# based on strwidth(tip labels) below.
#
provisional.tree.x.lim <- 1.3 * max(tip) # <= PROVISIONAL plot width
plot(0,0, type='n', bty='n', xaxt='n', yaxt='n', xlab='', ylab='', xlim=c(0,provisional.tree.x.lim), ylim=c(0,7))

tiplabel.x <- integer(7)
for(i in 1:7){
  # see warning above about internals of the tree; labels have '_', printed as ' '.
  tiplabel.x[i] <- tip[i]+strwidth(gsub('_', ' ', the.tree$tip.label[i], fixed=T), font=font.tip)
}

# visually show tip coords & max x to debug placement issues
plt.debug <- function(tree.x.lim, tip, tiplabel.x, spx=NULL, spy=NULL){
  if(pltdebug){ # F to hide/T to show debug
    cat('Tip labels:', paste(the.tree$tip.label, sep=', collapse='/'), '\n')
    axis(2) # useful only for placing labels
    for(i in 1:7){
      points(c(tip[i], tiplabel.x[i]), c(i,i)) # debug: do I have right tip coordinates?
    }
    lines(rep(tree.x.lim, 2), c(0,7)) # where is right edge?
    if(!is.null(spx)){
      points(spx, spy) # show spline control points, for tweaking
    }
  }
}

plt.debug(provisional.tree.x.lim, tip, tiplabel.x)

label.end.H <- max(tiplabel.x[1:2])
label.end.L <- max(tiplabel.x[3:7])
clade.dx <- strwidth('x') # space between clade marker line and its label
xdel <- 3*clade.dx # space between labeled clade tips and marker line

tree.x.lim <- 1.03*(max(tiplabel.x)+xdel) # <= FINAL plot width
if(pltdebug){cat('Plot width hacking:', provisional.tree.x.lim, tree.x.lim, tree.x.lim/1.03/max(tip), clade.dx)}

par(new=T) # I.e., NOT starting a new plot

####
#
# REAL PLOT
#
plot(the.tree,
  x.lim = tree.x.lim,
  y.lim = c(0,7),
  font=font.tip, label.offset=100, # bold-italic, nudged slightly right
  tip.color=col.tip, edge.color=col.edge,
  edge.width=lwd.edge,
  edge.lty=c(1,1,1,1, 1 ,1,1,1,1,1,1,1,0) # 5th is bottleneck edge; 14th is outgroup
)
lines(00+c(0,0), c(3.5,6), col='white', lwd=6) # Hide vertical line to outgroup
axis(1, pos=0.25, at=seq(0,25,by=5)*10^round(log10(max(tip)/25)))

if(pltdebug){text(tip[1]+100, 1.0, 'Venice, Italy (3367)', adj=0, font=font.tip)}

####
#
# BOTTLENECK ANNOTATION
#

```

```

# spline/ellipse control points (spy/y) & tweaks thereto (dx/y)
dx <- 0.01 * tree.x.lim
dy <- .04
spx <- c(7400, 7400, 9900, 10500) # by eye, chr1, for comparison
spx <- c(inode[2]+dx, inode[2]+dx, inode[4]-3*dx, inode[4]-dx)
spy <- c( 3.8,  3.9,  5.6-dy,  5.6-dy)

plt.debug(tree.x.lim, tip, tiplabel.x, spx, spy)

if(T){
  #ellipse version, defined by rect thru 2 middle pts of spx/y
  spf<-function(x){
    ifelse(x <= spx[2], spy[1],
           ifelse(x >= spx[3], spy[4],
                  spy[2]+(spy[3]-spy[2])*sqrt(pmax(0,1-((x-spx[3])/(spx[3]-spx[2]))^2))))
  }
} else {
  # spline version
  spf <- splinefun(spx,spy,method='hyman')
}
serx <- seq(spx[1],spx[length(spx)],length.out=50)
sery <- spf(serx)
tailx <- spx[1]
taily <- spy[1]
headx <- spx[4]
heady <- spy[4]
arrows(headx,heady,headx+tree.x.lim*1e-3,heady, length=.1,col=col.arrow,lwd=lwd.arrow)
lines(rev(serx), rev(sery), lty=c(5,1),col=col.arrow, lwd=lwd.arrow)
bottle.txt <- "inbreeding\nLoH / LoS"
if(T){
  text((headx+tailx)/2+(headx-tailx)*(-.01), (heady+taily)/2+(heady-taily)*(-.10),
       bottle.txt, srt=66, font=font.arrow, cex=cex.arrow, col=col.arrow)
} else {
  # experiment at wrapping text along curved path; not too pretty, but retain for now, maybe revisit
  bottlec <- strsplit(bottle,split=NULL)[[1]]
  for(i in 1:length(bottlec)){
    text(xser[i],yser[i],bottlec[i], srt=65, font=4, cex=.7, col=col.arrow)
  }
}

####
#
# CLADE ANNOTATION
#
clade.L.x <- label.end.L + xdel
clade.H.x <- label.end.H + xdel
dy <- .33
lines(rep(clade.L.x,2),c(3-dy,7+dy),lwd=lwd.clade,col=col.clade)
lines(rep(clade.H.x,2),c(1-dy,2+dy),lwd=lwd.clade,col=col.clade)
text(clade.L.x+clade.dx,5.0,'L-clade',srt=90,font=font.clade,cex=cex.clade,col=col.clade)
text(clade.H.x+clade.dx,1.5,'H-clade',srt=90,font=font.clade,cex=cex.clade,col=col.clade)

####
#
# LEGEND
#
# parameter plusx controls whether we try to annotate b/c (+) and d/e (x) sharing in tree; I think
# it looks cluttered, rather than adding clarity, so I vote no, but code is here, in case. "Logic,"
# if any, for my symbol choice is that + overlaid on x looks like the * at the next level; this
# analogy is more visible if we use pch 3/4/8 rather than Courier or Helvetica chars, but probably
# should use same in both tree & legend, which will take a modicum of additional work.
legend.text <- c('a: only in 1014 ',
                'b: only in 1335 ',
                'c: only in 1015 ',
                'd: only in 1012 ',
                'e: only in 1007 ',
                '*: shared by bcde',
                paste(ifelse(plusx,'+:',' '), 'shared by b/c '),
                paste(ifelse(plusx,'x:', ' '), 'shared by d/e ')
)

legend.text <- c('a: only in 1014 ',
                'b: only in 1335 ',
                'c: only in 1015 ',
                'd: only in 1012 ',
                'e: only in 1007 ',
                '*: in bcde ',
                paste(ifelse(plusx,'+:',' '), 'in bc '),
                paste(ifelse(plusx,'x:', ' '), 'in de '),
                'Discordant SNPs '
)

```

```

)
legend.text <- paste(legend.text, format(c(leg.counts, discord), width=4), sep=' - ')
legend.text <- paste(legend.text, ' ') # add a little more right margin in box
opar <- par(family='mono', cex=cex.legend)
legend('topright', legend=legend.text, cex=cex.legend, inset=c(0.05, 0), bg=col.legbox, box.col=col.legbox)
par(opar)
if(plusx){
  points(tree.labels[[16]], tree.labels[[17]]+.14, pch=8, col=col.elabel)
  points(tree.labels[[16]]+200, tree.labels[[17]]+1, pch=3, col=col.elabel)
  points(tree.labels[[16]]+200, tree.labels[[17]]-1, pch=4, col=col.elabel)
}

####
#
# EDGE LENGTHS
#
for(i in seq(1, length(tree.labels)-1, by=3)){
  if(F){ # T for \n in edge labels; F to remove (except "by 5")
    text(tree.labels[[i]], tree.labels[[i+1]], tree.labels[[i+2]])
  } else {
    # points(tree.labels[[i]], tree.labels[[i+1]], pch=3, col='green') # for debugging
    text(tree.labels[[i]], tree.labels[[i+1]], sub('\n([~z])', '\\1', tree.labels[[i+2]]),
         pos=3, offset=.4, font=font.elabel, col=col.elabel, cex=cex.elabel)
  }
}
}

caption <- function(stringency, which.tables=which.snp.tables(string.val=F)){
  caption.where <- '(UNKNOWN genome subset).'
  if(which.tables[1]=='Chr1') {caption.where <- 'on Chr1.'}
  if(which.tables[1]=='full') {caption.where <- 'genome-wide.'}
  if(which.tables[1]=='trunc') {caption.where <- 'all Chrs.'}
  cap.stringency <- c(
    'loose SNP filters.',
    'medium SNP filters.',
    'strict SNP filters.',
    'unfiltered SNPs.')
  cap <- paste('Tree based on', which.tables[2], 'reads and', cap.stringency[stringency],
    'Lengths\\' are numbers of shared/private SNPs', caption.where)
  return(cap)
}

```

Trees based on all four SNP filtering criteria are shown below. Their topologies are exactly the same, although the branch lengths are different. In all four, the length of the branch labeled “*” is probably inflated by lower coverage and higher error rate in 1014, which may mask further legitimate sharing between it and the other L-isolates. The branch lengths among the other 4 are too short for their topology to be convincing without a more rigorous analysis (e.g., a bootstrap test), but detail there is irrelevant to the story.

My sense is that the “medium” version is the best for the paper, made here and shown in Fig 1. In theory, this should look exactly like Fig 3, but something is apparently different between Knitr and direct-to-pdf. (Increasing fig.width in Knitr’s chunk headers from 8 (as in the pdf call below) to 9 helps somewhat, but probably still best to make the paper fig directly rather than via Knitr.)

```

###
#
# MAKE PDF FOR PAPER
#
if(which.snp.tables() == 'trunc-unfiltered'){
  paperfig.path <- paste('figs-mine/fig3-paperfig-medium-tree-', which.snp.tables(), '.pdf', sep='')
} else {
  paperfig.path <- paste('figs-mine/paperfig-medium-tree-', which.snp.tables(), '.pdf', sep='')
}
pdf(paperfig.path, width=8, height=5, onefile=TRUE, family='Helvetica', fonts='Courier', pointsize=10)
newick.medium <- newickize(make.tree(pat.summaries[, 'count2']))
show.tree(newick.medium, total.snps=consistent.count[2], pltdebug=F)
dev.off()

# pdf
# 2

# fig.paths for knitr chunks below; .h for "hand-made" trees; plain for automatic chr1/full versions
myfigpath <- paste(getwd(), '/figs-knitr/newick-', which.snp.tables(), '-', sep='')
myfigpath.h <- paste(getwd(), '/figs-knitr/newick-', sep='')

```

Figure 2, i.e., criteria [[1]]:

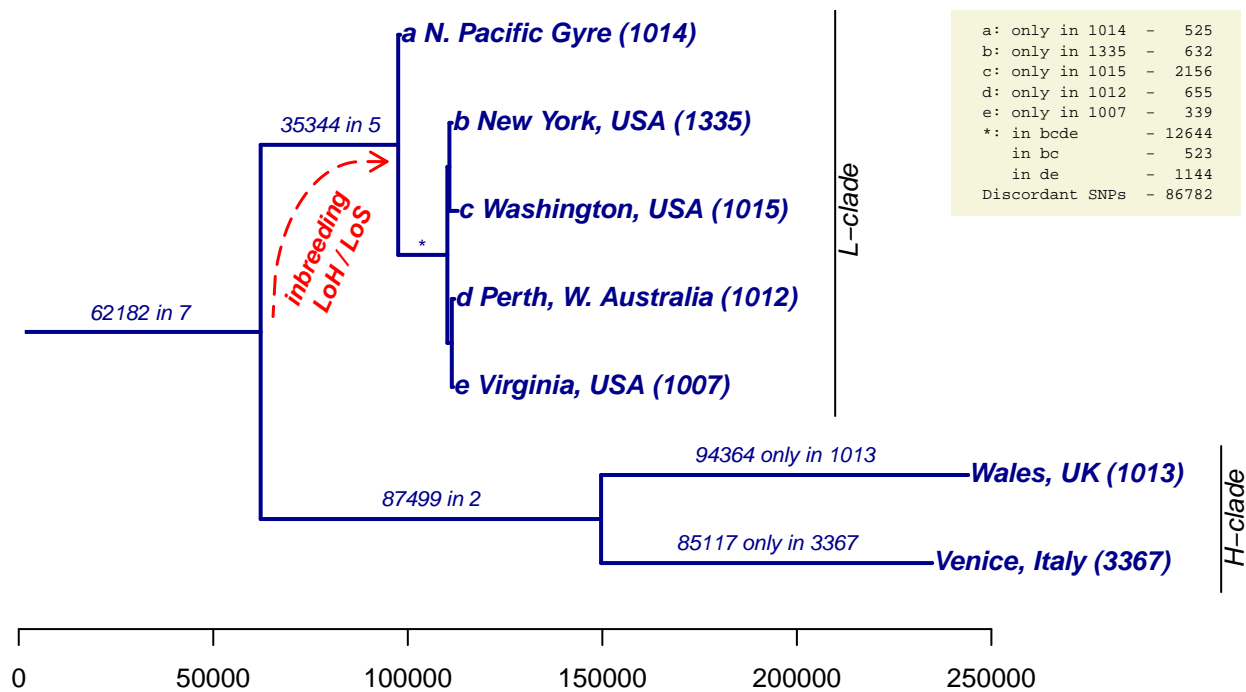


Figure 1: Proposed fig. for paper: Tree based on qfiltered reads and medium SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs.

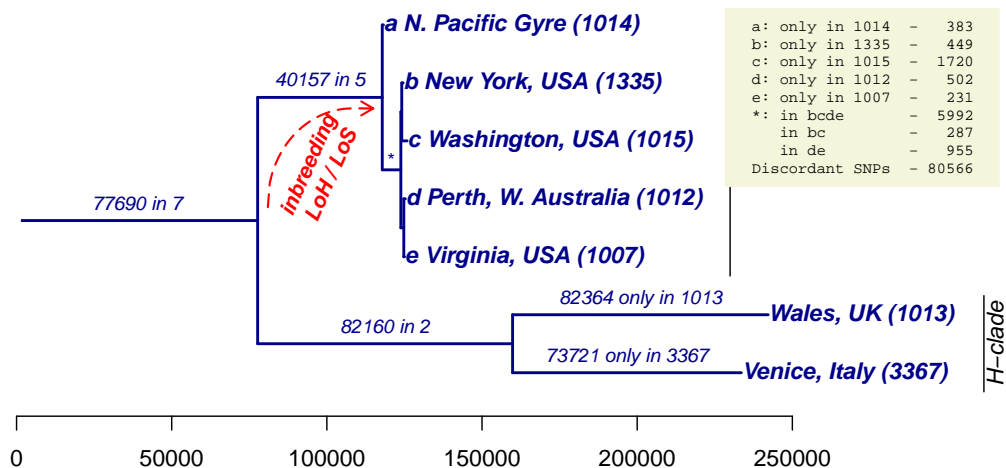


Figure 2: Tree based on qfiltered reads and loose SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs.

```
newick.loose <- newickize(make.tree(pat.summaries[, 'count1']))
show.tree(newick.loose, total.snps=consistent.count[1])
```

Figure 3, i.e. [[2]]:

```
# newick.medium <- newickize(tree.by.hand)
# simple.newick.medium <- newickize(tree.by.hand, alt=TRUE)
newick.medium <- newickize(make.tree(pat.summaries[, 'count2']))
simple.newick.medium <- newickize(make.tree(pat.summaries[, 'count2']), alt=TRUE)
show.tree(newick.medium, total.snps=consistent.count[2])
```

Figure 4, i.e. [[3]]:

```
newick.strict <- newickize(make.tree(pat.summaries[, 'count3']))
show.tree(newick.strict, total.snps=consistent.count[3])
```

Figure 5, i.e. [[4]]:

```
newick.unfiltered <- newickize(make.tree(pat.summaries[, 'count4']))
show.tree(newick.unfiltered, total.snps=consistent.count[4])
```

Some other versions of the trees are included in the appendix.

Counts for all tree edges in the medium tree:

```
#pat.summaries[c(128,110,102,6,97,19,9,2,5,33,65,17,3),]
tree.edges <- c(128,110,102,6,97,19,9,2,5,33,65,17,3)-1
non.edges <- setdiff(0:127, tree.edges)
sg.edges <- showgroup(restrict.to=tree.edges) ; sg.edges
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 2	001	1							X	449	632	1129	2260
# 3	002	1						X		73721	85117	87494	105614
# 5	004	1					X			1720	2156	2729	4608
# 6	005	2					X		X	287	523	1088	1407
# 9	010	1				X				383	525	485	1231
# 17	020	1			X					82364	94364	96464	113191

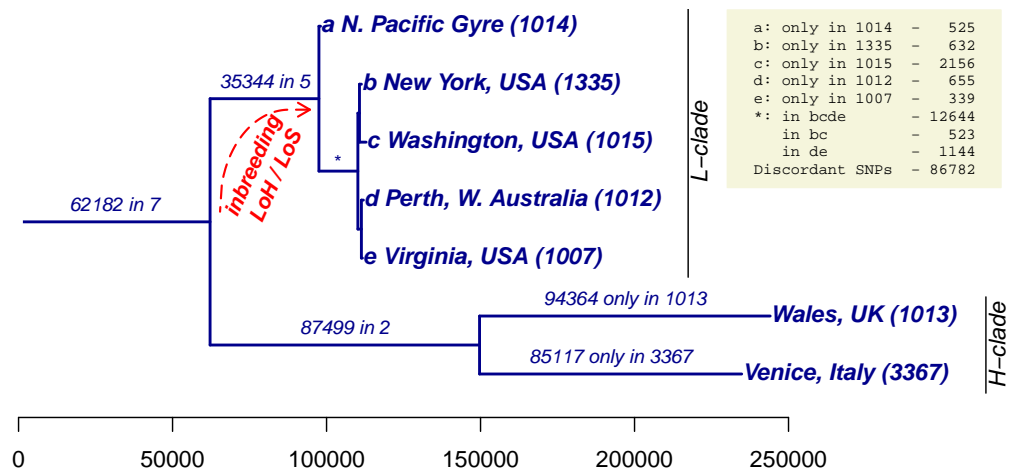


Figure 3: Tree based on qfiltered reads and medium SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs.

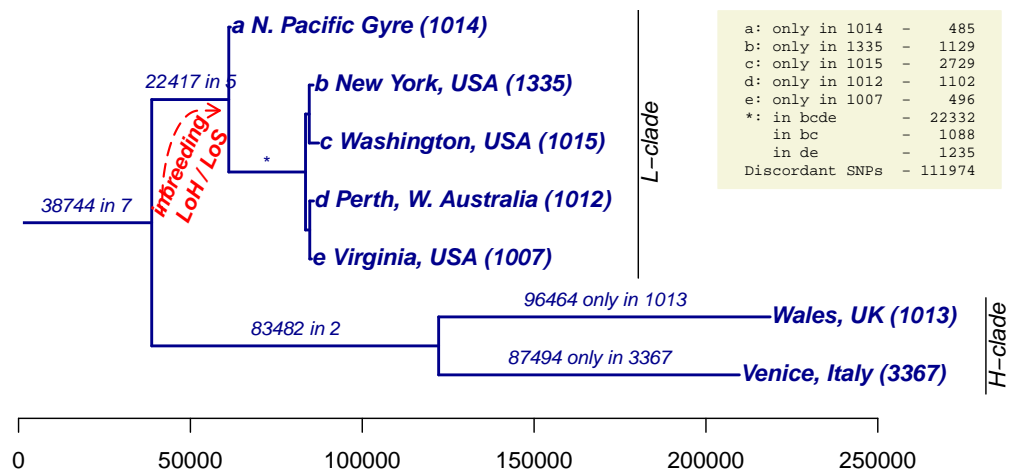


Figure 4: Tree based on qfiltered reads and strict SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs.

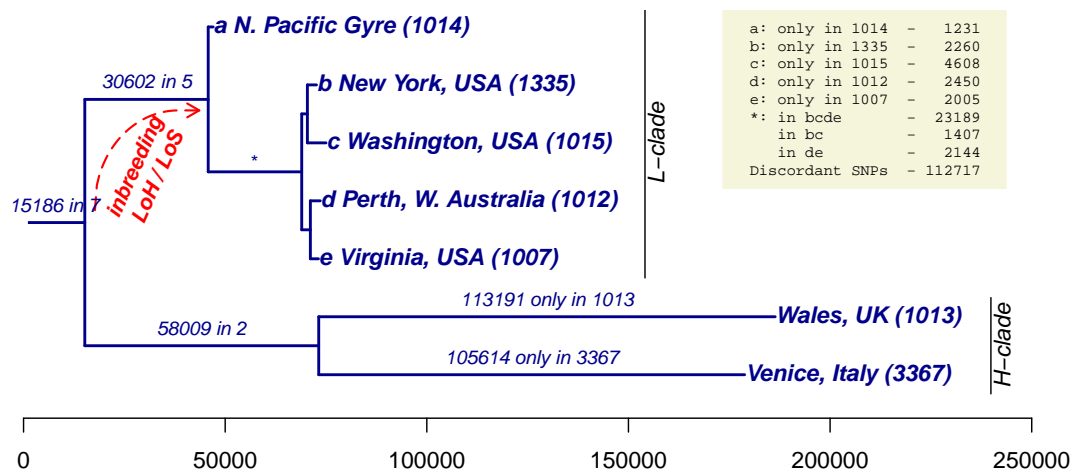


Figure 5: Tree based on qfiltered reads and unfiltered SNPs. “Lengths” are numbers of shared/private SNPs all Chrs.

# 19	022	2			X			X	82160	87499	83482	58009
# 33	040	1		X					502	655	1102	2450
# 65	100	1	X						231	339	496	2005
# 97	140	2	X	X					955	1144	1235	2144
# 102	145	4	X	X			X	X	5992	12644	22332	23189
# 110	155	5	X	X		X	X	X	40157	35344	22417	30602
# 128	177	7	X	X	X	X	X	X	77690	62182	38744	15186
# Total									366611	383124	359197	361896

Counts for the top 10 discordant patterns, i.e., SNPs whose sharing pattern does not match any of the bifurcations in the tree:

```
tenth <- sort(showgroup(restrict.to=non.edges)[-length(non.edges)+1, 'count2'], decreasing=T)[10]
sg.non.edges <- showgroup(restrict.to=non.edges, c2.thresh = tenth) ; sg.non.edges
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 32	037	5			X	X	X	X	X	2247	1877	1193	620
# 104	147	5	X	X			X	X	X	2073	4042	6741	10001
# 109	154	4	X	X		X	X			1227	1390	1065	1738
# 112	157	6	X	X		X	X	X	X	13239	10814	6862	12202
# 115	162	4	X	X	X			X		1848	1932	1828	1014
# 118	165	5	X	X	X		X		X	2726	5022	8440	9715
# 119	166	5	X	X	X		X	X		902	1976	3172	2688
# 120	167	6	X	X	X		X	X	X	11742	21003	35227	15091
# 126	175	6	X	X	X	X	X		X	16884	13630	8608	12697
# 127	176	6	X	X	X	X	X	X		2422	2412	1566	1032
# Other	(105 rows	w/	c2	< 1390)				25256	22684	37272	45919
# Total										80566	86782	111974	112717

And percent of discordant SNPs:

```
nsge <- nrow(sg.edges)
discordv <- consistent.count - sg.edges[nsge, c('count1', 'count2', 'count3', 'count4')] ; discordv
```

#	count1	count2	count3	count4
# Total	80566	86782	111974	112717

```
discordv.pct <- round(discordv/consistent.count*100,1) ; discordv.pct
```

#	count1	count2	count3	count4
# Total	18	18.5	23.8	23.7

In short, the sharing pattern observed at 86782 or 18.5% of the 469906 medium-stringency consistent SNPs positions observed across all 7 isolates are discordant with the medium tree. (The strict tree has slightly more.)

A majority of the discordant SNPs fall into one of three patterns: 6-way sharing excluding Gyre (likely a technical artifact since the low coverage in Gyre reduces our power to detect SNPs there), or 6-way sharing excluding one of the two H-isolates (likely a reflection of sexuality in the H-clade—SNP positions in a population in Hardy-Weinberg equilibrium are fairly likely to be homozygous for the reference allele in a given individual).

```
third.biggest <- sort(showgroup(pat.summaries,6)[-8,'count2'],decreasing=T)[3]
big.three <- showgroup(pat.summaries,6,c2.thresh = third.biggest); big.three

#      Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 112 157    6    X    X    X    X    X    X 13239 10814  6862 12202
# 120 167    6    X    X    X    X    X    X 11742 21003 35227 15091
# 126 175    6    X    X    X    X    X    X 16884 13630  8608 12697
# Other (      4 rows w/  c2  < 10814 )      3808  3670  2680  1964
# Total                                45673 49117 53377 41954

big.three.frac <- sum(big.three[1:3,'count2'])/discordv$count2; big.three.frac

# [1] 0.5236915
```

I.e., 52.4% of discordant SNPs fall into one of these three categories.

Out of curiosity: what is the ratio of full genome to Chr 1 branch lengths. Except for the shortest few, generally $\approx 10x$, as expected given the length of Chr 1:

```
# (vectors derived by editing Newick strings, and in that order)
print(
  c(Italy=86155, Wales=95697, IW=89598, Virg=330, Aust=632, VA=1296,
    Puget=2113, NY=658, PNY=480, four=10059, Gyre=568, five=39517, all=69526) /
  c(Italy=8813, Wales=9652, IW=9365, Virg=30, Aust=61, VA=19,
    Puget=207, NY=41, PNY=18, four=1005, Gyre=61, five=3912, all= 7054),
  digits=3)

# Italy Wales IW Virg Aust VA Puget NY PNY four Gyre five all
# 9.78 9.91 9.57 11.00 10.36 68.21 10.21 16.05 26.67 10.01 9.31 10.10 9.86

round(genome.length.constants()$genome.length.trunc / genome.length.constants()$chr1.length, digits=4)

# [1] 10.2879
```

9 Semi-Automated Tree-Building

Slightly formalizing the process above: Look for the bifurcation of the 7 strains that maximizes the number of shared SNPs *within* each side of the partition while minimizing the number and fraction of SNPs that are shared by subsets that include at least one strain on each side of the partition. The 2/5 split is the winner, with 6418 SNPs in conflict with that partition (16% of the 39842 SNPs not shared by all 7; Chr1 data). The runner-up places the Gyre in a group by itself (7079 = 18% in conflict).

```
treepart <- function(p.summ=pat.summaries, root=127, verbose=T, stringency='count2'){
  root.shared <- p.summ[root+1,stringency]
  df<-NULL
  for(i in 1:floor(root/2)){
    if(bitwAnd(i,root)==i && i < root-i){
      l1 <- showgroup(p.summ,subset=i,split=NULL,proper.subset=F,total=T)
      l <- l1[nrow(l1),stringency]
      r1 <- showgroup(p.summ,subset=root-i,split=NULL,proper.subset=F,total=T)
      r <- r1[nrow(r1),stringency]
      c1 <- showgroup(p.summ,subset=root,split=i,proper.subset=T,total=T)
      c <- c1[nrow(c1),stringency]
      df <- rbind(df, data.frame(pat=i,left=l,right=r,both=l+r,cross=c,all=l+r+c,ratio=c/(l+r+c),
                                best=' ',stringsAsFactors=F))
    }
  }
  df$pat<-as.octmode(df$pat)
```



```

maxl <- which.max(df$left)
maxr <- which.max(df$right)
maxb <- which.max(df$both)
minc <- which.min(df$cross)
minr <- which.min(df$ratio)
df$best[c(maxl,maxr,maxb,minc,minr)] <- '<'
df$best[maxl] <- paste(df$best[maxl], 'L') # max Left
df$best[maxr] <- paste(df$best[maxr], 'R') # max Right
df$best[maxb] <- paste(df$best[maxb], 'B') # max Both (L+R)
df$best[minc] <- paste(df$best[minc], 'C') # min Cross
df$best[minr] <- paste(df$best[minr], 'O') # min ratio (Cross/(Left+Right+Cross))
if(verbose){
  same <- all(maxl==c(maxr,maxb,minc,minr))
  cat('root:',      format(as.octmode(root),width=3),
      '; shared:',  root.shared,
      '\n  max l',   format(as.octmode(df$pat[maxl]),width=3),
      '\n  max r',   format(as.octmode(df$pat[maxr]),width=3),
      '\n  max both', format(as.octmode(df$pat[maxb]),width=3),
      '\n  min cross', format(as.octmode(df$pat[minc]),width=3),
      '\n  min ratio', format(as.octmode(df$pat[minr]),width=3),
      '\n\nAll the same?:', same,
      '\n')
  cat('\n')
}
return(df)
}

```

```
treepart()
```

```

# root: 177 ; shared: 62182 . max l 077 , max r 010 , max both 010 , min cross 010 , min ratio 010 .
# All the same?: FALSE
#   pat  left  right  both  cross  all    ratio    best
# 1   01   1210 289197 290407 117895 408302 0.2887446
# 2   02   85695 179062 264757 143545 408302 0.3515658
# 3   03   86625 105697 192322 215980 408302 0.5289712
# 4   04   2734 279522 282256 126046 408302 0.3087078
# 5   05   3889 274296 278185 130117 408302 0.3186783
# 6   06   87989 100546 188535 219767 408302 0.5382462
# 7   07   89639 98363 188002 220300 408302 0.5395516
# 8   10   1103 332135 333238 75064 408302 0.1838443 < R B C O
# 9   11   2221 282010 284231 124071 408302 0.3038707
# 10  12   86269 123335 209604 198698 408302 0.4866447
# 11  13   87911 102474 190385 217917 408302 0.5337152
# 12  14   3398 276093 279491 128811 408302 0.3154797
# 13  15   6023 273221 279244 129058 408302 0.3160847
# 14  16   88762 98932 187694 220608 408302 0.5403060
# 15  17   92437 97647 190084 218218 408302 0.5344524
# 16  20   94942 165462 260404 147898 408302 0.3622270
# 17  21   95741 96070 191811 216491 408302 0.5302227
# 18  22  267558 61350 328908 79394 408302 0.1944492
# 19  23  269213 8703 277916 130386 408302 0.3193372
# 20  24   97293 91454 188747 219555 408302 0.5377270
# 21  25   98831 89139 187970 220332 408302 0.5396300
# 22  26  270478 4870 275348 132954 408302 0.3256266
# 23  27  273828 3327 277155 131147 408302 0.3212010
# 24  30   95536 112945 208481 199821 408302 0.4893951
# 25  31   97054 93029 190083 218219 408302 0.5344549
# 26  32  268332 21310 289642 118660 408302 0.2906182
# 27  33  271905 6449 278354 129948 408302 0.3182644
# 28  34   98145 89824 187969 220333 408302 0.5396324
# 29  35  101895 88428 190323 217979 408302 0.5338671
# 30  36  271890 3636 275526 132776 408302 0.3251907
# 31  37  280857 2716 283573 124729 408302 0.3054822
# 32  40   1233 283707 284940 123362 408302 0.3021342
# 33  41   1957 273177 275134 133168 408302 0.3261507
# 34  42   86469 103226 189695 218607 408302 0.5354052

```

```
# 35 43 87577 98843 186420 221882 408302 0.5434262
# 36 44 3668 272853 276521 131781 408302 0.3227537
# 37 45 5508 269028 274536 133766 408302 0.3276153
# 38 46 89196 97572 186768 221534 408302 0.5425739
# 39 47 91873 95971 187844 220458 408302 0.5399386
# 40 50 1782 275846 277628 130674 408302 0.3200425
# 41 51 3049 271596 274645 133657 408302 0.3273484
# 42 52 87080 99819 186899 221403 408302 0.5422530
# 43 53 88991 97918 186909 221393 408302 0.5422285
# 44 54 4434 270058 274492 133810 408302 0.3277231
# 45 55 8497 268202 276699 131603 408302 0.3223178
# 46 56 90128 96225 186353 221949 408302 0.5435903
# 47 57 95836 95357 191193 217109 408302 0.5317363
# 48 60 95713 93634 189347 218955 408302 0.5362575
# 49 61 96684 89422 186106 222196 408302 0.5441952
# 50 62 268717 6909 275626 132676 408302 0.3249458
# 51 63 270814 3906 274720 133582 408302 0.3271647
# 52 64 98527 88440 186967 221335 408302 0.5420865
# 53 65 101114 86697 187811 220491 408302 0.5400194
# 54 66 272743 2629 275372 132930 408302 0.3255678
# 55 67 278683 1453 280136 128166 408302 0.3139000
# 56 70 96340 90579 186919 221383 408302 0.5422041
# 57 71 98107 88583 186690 221612 408302 0.5427649
# 58 72 269573 4578 274151 134151 408302 0.3285583
# 59 73 273771 3195 276966 131336 408302 0.3216639
# 60 74 99536 87112 186648 221654 408302 0.5428678
# 61 75 105295 86101 191396 216906 408302 0.5312391
# 62 76 274535 1596 276131 132171 408302 0.3237089
# 63 77 288224 917 289141 119161 408302 0.2918453 < L
```

Comparing the 5/2 split to the second-place NPG/rest split (below), the former has fewer pattern instances in conflict with the split (6418 vs 7079), as well as somewhat more random distribution of the conflicting patterns (92 vs 62 rows), whereas the 1/6 split has the majority of its conflicts (3912 of 7079, or 55%) concentrated in one pattern—the 5 NE strains. Collectively, these seem to favor the 5/2 split as the correct “history.”

```
showgroup(pat.summaries,split=strtoi('022'), subset=127, proper.subset=T, c2.thresh=100)
```

```
# Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 4 003 2 X X 994 298 532 587
# 7 006 2 X X 624 138 282 590
# 8 007 3 X X 104 197 371 557
# 12 013 3 X X 257 226 146 338
# 16 017 4 X X 390 329 211 564
# 18 021 2 X X 998 167 337 402
# 20 023 3 X X 1274 558 1020 533
# 21 024 2 X X 686 195 410 625
# 22 025 3 X X 135 216 466 522
# 23 026 3 X X 793 431 763 789
# 24 027 4 X X 461 759 1423 771
# 26 031 3 X X 268 233 151 361
# 27 032 3 X X 698 131 74 86
# 28 033 4 X X 1139 973 574 306
# 29 034 3 X X 103 119 91 219
# 30 035 4 X X 578 509 329 503
# 31 036 4 X X 345 320 227 211
# 32 037 5 X X 2247 1877 1193 620
# 35 042 2 X X 503 119 254 394
# 39 046 3 X X 58 154 425 604
# 40 047 4 X X 127 256 708 708
# 48 057 5 X X 221 219 189 324
# 49 060 2 X X 627 116 237 388
# 51 062 3 X X 703 269 454 469
# 52 063 4 X X 151 184 332 194
# 53 064 3 X X 53 184 458 601
# 54 065 4 X X 122 284 731 582
# 55 066 4 X X 217 489 1025 851
# 56 067 5 X X 556 1015 2544 1151
```

```

# 62 075 5 X X X X X 195 187 210 328
# 63 076 5 X X X X X 106 130 124 128
# 64 077 6 X X X X X 850 847 827 485
# 83 122 3 X X X X 501 162 165 354
# 86 125 4 X X X X 67 113 223 283
# 87 126 4 X X X X 99 198 268 425
# 88 127 5 X X X X 225 321 501 482
# 94 135 5 X X X X 126 117 88 235
# 96 137 6 X X X X 405 324 240 333
# 99 142 3 X X X X 386 409 463 755
# 103 146 4 X X X X 196 510 969 1795
# 104 147 5 X X X X 2073 4042 6741 10001
# 111 156 5 X X X X 565 575 410 735
# 112 157 6 X X X X 13239 10814 6862 12202
# 113 160 3 X X X 337 375 399 712
# 115 162 4 X X X X 1848 1932 1828 1014
# 116 163 5 X X X X 255 271 328 316
# 117 164 4 X X X X 237 627 1053 1752
# 118 165 5 X X X X 2726 5022 8440 9715
# 119 166 5 X X X X 902 1976 3172 2688
# 120 167 6 X X X X 11742 21003 35227 15091
# 125 174 5 X X X X 659 682 468 782
# 126 175 6 X X X X 16884 13630 8608 12697
# 127 176 6 X X X X 2422 2412 1566 1032
# Other ( 39 rows w/ c2 < 100 ) 3209 1750 1921 4847
# Total 75686 79394 97058 94037

showgroup(pat.summaries,split=strtoi('010'), subset=127, proper.subset=T, c2.thresh=100)

# Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 10 011 2 X 515 486 317 827
# 12 013 3 X X 257 226 146 338
# 13 014 2 X X 133 139 132 417
# 14 015 3 X X 1041 984 660 1389
# 16 017 4 X X X 390 329 211 564
# 26 031 3 X X X 268 233 151 361
# 27 032 3 X X X 698 131 74 86
# 28 033 4 X X X 1139 973 574 306
# 29 034 3 X X X 103 119 91 219
# 30 035 4 X X X 578 509 329 503
# 31 036 4 X X X X 345 320 227 211
# 32 037 5 X X X X 2247 1877 1193 620
# 46 055 4 X X X X 606 696 668 971
# 48 057 5 X X X X 221 219 189 324
# 62 075 5 X X X X 195 187 210 328
# 63 076 5 X X X X 106 130 124 128
# 64 077 6 X X X X 850 847 827 485
# 78 115 4 X X X X 141 139 122 604
# 94 135 5 X X X X 126 117 88 235
# 96 137 6 X X X X 405 324 240 333
# 109 154 4 X X X X 1227 1390 1065 1738
# 110 155 5 X X X X 40157 35344 22417 30602
# 111 156 5 X X X X 565 575 410 735
# 112 157 6 X X X X 13239 10814 6862 12202
# 125 174 5 X X X X 659 682 468 782
# 126 175 6 X X X X 16884 13630 8608 12697
# 127 176 6 X X X X 2422 2412 1566 1032
# Other ( 35 rows w/ c2 < 100 ) 2151 1232 1130 3730
# Total 87668 75064 49099 72767

```

Below is the full summary of shared SNPs that do *not* directly correspond to tree splits, e.g. deep coalescence, independent coincident mutations, false positives/false negatives in the shared SNP calls, loss of SNPs in hemizygous regions, etc. (Additionally, SAMTools' SNP calls exclude positions it judges to be homozygous, and I think it operates without regard to the reference sequence, so homozygous nonreference positions, while rare except in IT/Wales, often are not called SNPs by SAMTools, but are relevant for this analysis. Provided the position is called a SNP in some other isolate, the consistency filtering we've done above should recover it, but this is still worth keeping in mind when

examining the data.)

First, here are SNPs that “coalesce” on the branch from the LCA of bcde, i.e., shared among some nonempty, proper subset of bcde other than bc or de. There are 8 such patterns: any of the 4 choose 3 trios plus any of the 4 pairs having exactly one of bc.

```
sg4 <- showgroup(pat.summaries, subset=strtoi('0145'), split=5, proper.subset = F)
sg4
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 34	041	2		X					X	42	92	313	368
# 37	044	2		X			X			69	279	1001	1809
# 38	045	3		X			X		X	202	593	1970	1656
# 66	101	2	X						X	29	47	73	314
# 69	104	2	X				X			39	122	329	1196
# 70	105	3	X				X		X	78	181	396	805
# 98	141	3	X	X					X	78	149	258	519
# 101	144	3	X	X			X			383	1176	2395	4432
# 102	145	4	X	X			X		X	5992	12644	22332	23189
# Total										6912	15283	29067	34288

```
sg4n <- nrow(sg4)
sg4pct <- round(sg4$count2[sg4n-1]/sg4$count2[sg4n]*100,1)
sg4pct
```

```
# [1] 82.7
```

So, of the 15283 SNPs found only in bcde, 82.7% have a sharing pattern consistent with the given tree structure.

Similarly, we analyze patterns relative to the root of the L-clade (14 patterns—any nonempty proper subset of bcde together with a):

```
sg5 <- showgroup(pat.summaries, subset=strtoi('0155'), split=8, proper.subset = F)
sg5
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 10	011	2				X			X	515	486	317	827
# 13	014	2				X	X			133	139	132	417
# 14	015	3				X	X		X	1041	984	660	1389
# 41	050	2		X		X				13	24	53	105
# 42	051	3		X		X			X	52	57	74	126
# 45	054	3		X		X	X			20	78	133	292
# 46	055	4		X		X	X		X	606	696	668	971
# 73	110	2	X			X				12	11	29	150
# 74	111	3	X			X			X	6	11	8	139
# 77	114	3	X			X	X			12	36	56	365
# 78	115	4	X			X	X		X	141	139	122	604
# 105	150	3	X	X		X				28	51	55	238
# 106	151	4	X	X		X			X	43	69	54	220
# 109	154	4	X	X		X	X			1227	1390	1065	1738
# 110	155	5	X	X		X	X		X	40157	35344	22417	30602
# Total										44006	39515	25843	38183

```
sg5n <- nrow(sg5)
sg5pct <- round(sg5$count2[sg5n-1]/sg5$count2[sg5n]*100,1)
```

I.e., of the 39515 SNPs found only in abcde, 89.4% have a sharing pattern consistent with the given tree structure.

Finally, how many SNPs have patterns inconsistent with the 5-2 split, i.e., include at least one strain on each side of the 5-2 split, but not shared by all 7?

```
sg7 <- showgroup(pat.summaries, subset=127, split=strtoi('022'), proper.subset=F)
sg7
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 4	003	2						X	X	994	298	532	587
# 7	006	2					X	X		624	138	282	590
# 8	007	3					X	X	X	104	197	371	557

# 11	012	2				X		X		565	49	32	93
# 12	013	3				X		X	X	257	226	146	338
# 15	016	3				X	X	X		46	60	49	152
# 16	017	4				X	X	X	X	390	329	211	564
# 18	021	2			X			X		998	167	337	402
# 20	023	3			X			X	X	1274	558	1020	533
# 21	024	2			X		X			686	195	410	625
# 22	025	3			X		X		X	135	216	466	522
# 23	026	3			X		X	X		793	431	763	789
# 24	027	4			X		X	X	X	461	759	1423	771
# 25	030	2			X	X				609	69	47	93
# 26	031	3			X	X			X	268	233	151	361
# 27	032	3			X	X		X		698	131	74	86
# 28	033	4			X	X		X	X	1139	973	574	306
# 29	034	3			X	X	X			103	119	91	219
# 30	035	4			X	X	X		X	578	509	329	503
# 31	036	4			X	X	X	X		345	320	227	211
# 32	037	5			X	X	X	X	X	2247	1877	1193	620
# 35	042	2		X				X		503	119	254	394
# 36	043	3		X				X	X	56	86	151	133
# 39	046	3		X			X	X		58	154	425	604
# 40	047	4		X			X	X	X	127	256	708	708
# 43	052	3		X		X		X		8	13	17	22
# 44	053	4		X		X		X	X	35	34	26	56
# 47	056	4		X		X	X	X		26	44	50	88
# 48	057	5		X		X	X	X	X	221	219	189	324
# 49	060	2		X	X					627	116	237	388
# 50	061	3		X	X				X	52	80	131	115
# 51	062	3		X	X			X		703	269	454	469
# 52	063	4		X	X			X	X	151	184	332	194
# 53	064	3		X	X		X			53	184	458	601
# 54	065	4		X	X		X		X	122	284	731	582
# 55	066	4		X	X		X	X		217	489	1025	851
# 56	067	5		X	X		X	X	X	556	1015	2544	1151
# 57	070	3		X	X	X				22	9	14	24
# 58	071	4		X	X	X			X	9	20	7	28
# 59	072	4		X	X	X		X		41	36	21	31
# 60	073	5		X	X	X		X	X	94	72	62	38
# 61	074	4		X	X	X	X			20	46	51	116
# 62	075	5		X	X	X	X		X	195	187	210	328
# 63	076	5		X	X	X	X	X		106	130	124	128
# 64	077	6		X	X	X	X	X	X	850	847	827	485
# 67	102	2	X					X		351	67	96	351
# 68	103	3	X					X	X	24	34	46	143
# 71	106	3	X				X	X		32	66	109	377
# 72	107	4	X				X	X	X	58	84	129	330
# 75	112	3	X			X		X		10	11	8	26
# 76	113	4	X			X		X	X	7	9	5	66
# 79	116	4	X			X	X	X		8	8	11	101
# 80	117	5	X			X	X	X	X	48	32	25	241
# 81	120	2	X		X					432	76	98	309
# 82	121	3	X		X				X	22	22	34	73
# 83	122	3	X		X			X		501	162	165	354
# 84	123	4	X		X			X	X	63	98	91	124
# 85	124	3	X		X		X			43	88	152	400
# 86	125	4	X		X		X		X	67	113	223	283
# 87	126	4	X		X		X	X		99	198	268	425
# 88	127	5	X		X		X	X	X	225	321	501	482
# 89	130	3	X		X	X				9	9	9	27
# 90	131	4	X		X	X			X	6	3	0	52
# 91	132	4	X		X	X		X		19	21	10	38
# 92	133	5	X		X	X		X	X	31	28	15	52
# 93	134	4	X		X	X	X			18	17	22	143
# 94	135	5	X		X	X	X		X	126	117	88	235
# 95	136	5	X		X	X	X	X		34	56	26	106
# 96	137	6	X		X	X	X	X	X	405	324	240	333
# 99	142	3	X	X				X		386	409	463	755

```

# 100 143 4 X X X X 37 58 103 190
# 103 146 4 X X X X 196 510 969 1795
# 104 147 5 X X X X 2073 4042 6741 10001
# 107 152 4 X X X X 18 27 15 67
# 108 153 5 X X X X 39 27 26 96
# 111 156 5 X X X X 565 575 410 735
# 112 157 6 X X X X 13239 10814 6862 12202
# 113 160 3 X X X X 337 375 399 712
# 114 161 4 X X X X 74 96 113 207
# 115 162 4 X X X X 1848 1932 1828 1014
# 116 163 5 X X X X 255 271 328 316
# 117 164 4 X X X X 237 627 1053 1752
# 118 165 5 X X X X 2726 5022 8440 9715
# 119 166 5 X X X X 902 1976 3172 2688
# 120 167 6 X X X X 11742 21003 35227 15091
# 121 170 4 X X X X 13 18 15 69
# 122 171 5 X X X X 41 19 13 70
# 123 172 5 X X X X 58 71 45 86
# 124 173 6 X X X X 131 87 47 114
# 125 174 5 X X X X 659 682 468 782
# 126 175 6 X X X X 16884 13630 8608 12697
# 127 176 6 X X X X 2422 2412 1566 1032
# 128 177 7 X X X X 77690 62182 38744 15186
# Total 153376 141576 135802 109223

sg7n <- nrow(sg7)
sg7pct <- round(sg7$count2[sg7n-1]/sg7$count2[sg7n]*100,1)
sg7pct

# [1] 43.9

```

A more compact version of that table, showing only the larger counts:

```

thresh <- signif(.02 * sg7$count2[sg7n],1)
thresh

# [1] 3000

showgroup(pat.summaries, subset=127, split=strtoi('022'), proper.subset=F, c2.thresh = thresh)

#      Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 104 147 5 X X X X X X X 2073 4042 6741 10001
# 112 157 6 X X X X X X 13239 10814 6862 12202
# 118 165 5 X X X X X X 2726 5022 8440 9715
# 120 167 6 X X X X X X 11742 21003 35227 15091
# 126 175 6 X X X X X X 16884 13630 8608 12697
# 128 177 7 X X X X X X 77690 62182 38744 15186
# Other ( 87 rows w/ c2 < 3000 ) 29022 24883 31180 34331
# Total 153376 141576 135802 109223

```

So, of the 141576 SNPs found both in the L- and H-clades, 43.9% have a sharing pattern consistent with the given tree structure, i.e., are found in all 7 isolates. Among the others, three patterns dominate—(i) the 6-way pattern excluding the Gyre is the largest, plausibly explained by 7-way sharing from which the Gyre drops out due to low coverage/high error rate, (ii) the 6-way excluding Italy, and (iii) ditto for Wales. Origin of the later two cases is unclear, but may partly reflect Hardy-Weinberg—some positions that are *population-level* SNPs in those isolates will be homozygous-reference in the CCMP founder cell for IT or Wales. If I take the 7-way shared SNP count (69526) as a surrogate approximating the number of population-level SNPs in either IT or Wales that are shared with the L-clade, then I might expect, based on HWE, roughly half that number to be lost (become homozygous) in IT, and a similar number in Wales. However, the observed counts of these positions are lower by $\approx 20K$ than I might have guessed from HWE, perhaps suggesting that IT and Wales are distinct populations, each with a pool of many thousand private polymorphisms.

In aggregate:

```

untreelike <-
  sg7$count2[sg7n]-sg7$count2[sg7n-1] +
  sg5$count2[sg5n]-sg5$count2[sg5n-1] +
  sg4$count2[sg4n]-sg4$count2[sg4n-1]
untreelike

# [1] 86204

consistent.count[2]

# [1] 469906

unpct <- round(untreelike/consistent.count[2]*100,1)
unpct

# [1] 18.3

```

I.e., 86204 or 18.3% of the 469906 consistent SNPs identified (by criterion 2) across all 7 isolates are discordant with the assumed tree.

Overall, based on this data, I take the following to be obvious: (a) separation of the the H-isolates from the L-isolates (and from each other??), and (b) near-identity of the L-isolates. Due to the small counts, the exact topology among the L-isolates (esp. bcde) is uncertain, but *any* topology there is consistent with the asexual/clonal/global-expansion hypothesis, so there is little point in examining this subtree more carefully. Again, we believe the (apparent) slight separation of the Gyre from the other L-isolates is largely driven by technical artifacts (lower coverage/higher error rates) in the sequencing rather than by biological effects. However, the discord between Gyre SNPs and others is the major substantive ambiguity in the offered tree. Nevertheless, in the next section we show by a bootstrap analysis that the offered placement of Gyre with respect to the other 4 L-isolates is strongly supported by the data.

9.1 Bootstrap

How robust is the inferred tree? Italy/Wales seem clearly related to each other but separate from the other 5. Likewise, the 4 coastal L-isolates seem to be closely related, with little data to separate them (and perhaps little sense in trying). So, the key question here is whether the top level bifurcation is 2/5 or NPG/6. Here, we do a simple bootstrap test (on c2 numbers only) to see whether the 2/5 split is consistently the most parsimonious.

```

n2 <- sum(pattern.counts[[2]][,2]); n2

# [1] 469906

```

Conceptually, we sample, with replacement, n2=469906 SNP positions from among the 469906 positions declared consistent SNPs according to criterion c2, and recalculate the statistics examined above to see whether the 2/5 split again minimizes conflicting sharing patterns. This resampling/calculation is repeated nboot times (set near front of file). Since all that matters is the sharing pattern, this procedure is expedited by actually sampling 469906 independent integers in the range 0:127 with probabilities proportional to the pattern counts given in column 2 of pattern.counts[[2]]. The sample is then tabulated in a 128 row table analogous to pattern.summaries, for analysis by showgroups/treepart, as above.

```

boot.sample <- sample(0:127,n2,replace=T,prob=pattern.counts[[2]][,2])
str(boot.sample)

# int [1:469906] 127 109 16 18 18 2 16 109 18 2 ...

boot.count <- mytable(boot.sample,c(0,127))
boot.count[c(1:4,125:128),] # show a few rows

#      val count
# [1,]    0   625
# [2,]    1   631
# [3,]    2 85325
# [4,]    3   292
# [5,]   124   722

```

```
# [6,] 125 13882
# [7,] 126 2292
# [8,] 127 62392

boot.counts <- list(NULL,boot.count,NULL) # dummy list with just c2 summaries
cor(pattern.counts[[2]][,2],boot.counts[[2]][,2]) # just curious - how correlated are they?

# [1] 0.9999865

boot.summaries <- pat.summary(boot.counts)
showgroup(boot.summaries,c2.thresh=400) #show a few rows
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 1	000	0								NA	625	NA	NA
# 2	001	1							X	NA	631	NA	NA
# 3	002	1						X		NA	85325	NA	NA
# 5	004	1					X			NA	2121	NA	NA
# 6	005	2					X		X	NA	550	NA	NA
# 9	010	1				X				NA	523	NA	NA
# 10	011	2				X			X	NA	484	NA	NA
# 14	015	3				X	X		X	NA	1022	NA	NA
# 17	020	1			X					NA	93622	NA	NA
# 19	022	2			X			X		NA	87425	NA	NA
# 20	023	3			X			X	X	NA	549	NA	NA
# 23	026	3			X		X	X		NA	458	NA	NA
# 24	027	4			X		X	X	X	NA	736	NA	NA
# 28	033	4			X	X		X	X	NA	1042	NA	NA
# 30	035	4			X	X	X		X	NA	528	NA	NA
# 32	037	5			X	X	X	X	X	NA	1832	NA	NA
# 33	040	1		X						NA	659	NA	NA
# 38	045	3		X			X		X	NA	612	NA	NA
# 46	055	4		X		X	X		X	NA	721	NA	NA
# 55	066	4		X	X		X	X		NA	478	NA	NA
# 56	067	5		X	X		X	X	X	NA	992	NA	NA
# 64	077	6		X	X	X	X	X	X	NA	751	NA	NA
# 97	140	2	X	X						NA	1107	NA	NA
# 101	144	3	X	X			X			NA	1184	NA	NA
# 102	145	4	X	X			X		X	NA	12647	NA	NA
# 103	146	4	X	X			X	X		NA	515	NA	NA
# 104	147	5	X	X			X	X	X	NA	4052	NA	NA
# 109	154	4	X	X		X	X			NA	1408	NA	NA
# 110	155	5	X	X		X	X		X	NA	35461	NA	NA
# 111	156	5	X	X		X	X	X		NA	575	NA	NA
# 112	157	6	X	X		X	X	X	X	NA	10806	NA	NA
# 115	162	4	X	X	X			X		NA	1904	NA	NA
# 117	164	4	X	X	X		X			NA	653	NA	NA
# 118	165	5	X	X	X		X		X	NA	5063	NA	NA
# 119	166	5	X	X	X		X	X		NA	1925	NA	NA
# 120	167	6	X	X	X		X	X	X	NA	21159	NA	NA
# 125	174	5	X	X	X	X	X			NA	722	NA	NA
# 126	175	6	X	X	X	X	X		X	NA	13882	NA	NA
# 127	176	6	X	X	X	X	X	X		NA	2292	NA	NA
# 128	177	7	X	X	X	X	X	X	X	NA	62392	NA	NA
# Other	(88 rows	w/	c2	<	400)			NA	10473	NA	NA
# Total										NA	469906	NA	NA

Tree partition analysis (and how to pluck out only the best rows based on 3 smallest cross counts and “best” criteria):

```
tp <- treepart(boot.summaries,root=127) ; tp

# root: 177 ; shared: 62392 . max l 077 , max r 010 , max both 010 , min cross 010 , min ratio 010 .
# All the same?: FALSE
# pat left right both cross all ratio best
# 1 01 1256 288443 289699 118440 408139 0.2901953
# 2 02 85950 178885 264835 143304 408139 0.3511157
# 3 03 86873 104979 191852 216287 408139 0.5299347
# 4 04 2746 278944 281690 126449 408139 0.3098185
```



```

# 5 05 3927 273725 277652 130487 408139 0.3197122
# 6 06 88232 99824 188056 220083 408139 0.5392354
# 7 07 89887 97655 187542 220597 408139 0.5404948
# 8 10 1148 331649 332797 75342 408139 0.1845989 < R B C O
# 9 11 2263 281358 283621 124518 408139 0.3050872
# 10 12 86520 122657 209177 198962 408139 0.4874859
# 11 13 88143 101715 189858 218281 408139 0.5348202
# 12 14 3402 275412 278814 129325 408139 0.3168651
# 13 15 6089 272628 278717 129422 408139 0.3171028
# 14 16 88996 98179 187175 220964 408139 0.5413940
# 15 17 92728 96929 189657 218482 408139 0.5353127
# 16 20 94247 165944 260191 147948 408139 0.3624942
# 17 21 95027 96301 191328 216811 408139 0.5312185
# 18 22 266997 61606 328603 79536 408139 0.1948748
# 19 23 268618 8714 277332 130807 408139 0.3204962
# 20 24 96546 91668 188214 219925 408139 0.5388483
# 21 25 98115 89369 187484 220655 408139 0.5406369
# 22 26 269915 4918 274833 133306 408139 0.3266191
# 23 27 273243 3368 276611 131528 408139 0.3222628
# 24 30 94831 113202 208033 200106 408139 0.4902888
# 25 31 96325 93227 189552 218587 408139 0.5355700
# 26 32 267778 21354 289132 119007 408139 0.2915845
# 27 33 271371 6442 277813 130326 408139 0.3193177
# 28 34 97375 90017 187392 220747 408139 0.5408623
# 29 35 101208 88637 189845 218294 408139 0.5348521
# 30 36 271299 3653 274952 133187 408139 0.3263276
# 31 37 280336 2740 283076 125063 408139 0.3064226
# 32 40 1284 283167 284451 123688 408139 0.3030536
# 33 41 2006 272572 274578 133561 408139 0.3272439
# 34 42 86730 102533 189263 218876 408139 0.5362781
# 35 43 87827 98064 185891 222248 408139 0.5445400
# 36 44 3686 272305 275991 132148 408139 0.3237818
# 37 45 5570 268498 274068 134071 408139 0.3284935
# 38 46 89436 96842 186278 221861 408139 0.5435918
# 39 47 92135 95276 187411 220728 408139 0.5408158
# 40 50 1841 275289 277130 131009 408139 0.3209911
# 41 51 3109 271039 274148 133991 408139 0.3282975
# 42 52 87348 99102 186450 221689 408139 0.5431703
# 43 53 89239 97163 186402 221737 408139 0.5432879
# 44 54 4449 269457 273906 134233 408139 0.3288904
# 45 55 8622 267664 276286 131853 408139 0.3230591
# 46 56 90373 95507 185880 222259 408139 0.5445669
# 47 57 96176 94671 190847 217292 408139 0.5323971
# 48 60 95036 93947 188983 219156 408139 0.5369641
# 49 61 95981 89660 185641 222498 408139 0.5451525
# 50 62 268207 6970 275177 132962 408139 0.3257763
# 51 63 270256 3906 274162 133977 408139 0.3282632
# 52 64 97792 88671 186463 221676 408139 0.5431385
# 53 65 100405 86959 187364 220775 408139 0.5409309
# 54 66 272203 2669 274872 133267 408139 0.3265236
# 55 67 278088 1506 279594 128545 408139 0.3149540
# 56 70 95662 90856 186518 221621 408139 0.5430037
# 57 71 97409 88836 186245 221894 408139 0.5436726
# 58 72 269072 4619 273691 134448 408139 0.3294172
# 59 73 273297 3211 276508 131631 408139 0.3225151
# 60 74 98775 87363 186138 222001 408139 0.5439348
# 61 75 104642 86369 191011 217128 408139 0.5319952
# 62 76 273963 1642 275605 132534 408139 0.3247276
# 63 77 287626 974 288600 119539 408139 0.2928880 < L

```

```

otp <- order(tp[, 'cross'])[1:3] # 3 smallest 'cross' counts
btp <- which(tp[, 'best'] != '') # 'best' by Left/Right/Both/Cross/ratio
toptp <- unique(c(otp, btp, 18, 8)) # above, plus 5/2, 6/1 splits
print(tp[toptp,]) # show the winners

# pat left right both cross all ratio best
# 8 10 1148 331649 332797 75342 408139 0.1845989 < R B C O
# 18 22 266997 61606 328603 79536 408139 0.1948748
# 1 01 1256 288443 289699 118440 408139 0.2901953
# 63 77 287626 974 288600 119539 408139 0.2928880 < L

```

Now repeat the above nboot times, and summarize results:

```

nboot <- params$nboot # default from params set in section 2
nboot <- ((nboot+2) %/% 4) * 4 + 1 # summary is cleaner if n mod 4 == 1, so int median/quartiles
cat('***\n*** Doing', nboot, 'bootstrap replicates.\n***\n')

```

```

# ***
# *** Doing 101 bootstrap replicates.
# ***

bcor <- numeric(nboot)
b52cross <- integer(nboot)
b61cross <- integer(nboot)
brev <- logical(nboot)
for(i in 1:nboot){
  boot.sample <- sample(0:127,n2,replace=T,prob=pattern.counts[[2]][,2])
  boot.count <- mtable(boot.sample,c(0,127))
  boot.counts <- list(NULL,boot.count,NULL) # dummy list with just c2 summaries
  boot.summaries <- pat.summary(boot.counts)
  tp <- treepart(boot.summaries,root=127, verbose=F)
  bcor[i] <- cor(pattern.counts[[2]][,2],boot.counts[[2]][,2]) # just curious - how correlated are they?
  b52cross[i] <- tp[18,'cross']
  b61cross[i] <- tp[ 8,'cross']
  brev[i] <- (b52cross[i] > b61cross[i])
  if(brev[i]){
    # show the unexpected ones; probably breaks w/ cache
    otp <- order(tp[, 'cross'])[1:3]
    btp <- which(tp[, 'best'] != '')
    totp <- unique(c(otp,btp,18,8))
    print(tp[top,])
  }
}

#   pat  left  right  both  cross  all  ratio  best
# 8   10  1088 332249 333337 75187 408524 0.1840455 < R B C O
# 18  22 267474 61282 328756 79768 408524 0.1952590
# 1   01  1196 289143 290339 118185 408524 0.2892976
# 63  77 288371   896 289267 119257 408524 0.2919216 < L
#   pat  left  right  both  cross  all  ratio  best
# 8   10  1057 332177 333234 74775 408009 0.1832680 < R B C O
# 18  22 267508 61021 328529 79480 408009 0.1947996
# 1   01  1160 289146 290306 117703 408009 0.2884814
# 63  77 288256 873 289129 118880 408009 0.2913661 < L
#   pat  left  right  both  cross  all  ratio  best
# 8   10  1209 332226 333435 74716 408151 0.1830597 < R B C O
# 18  22 267615 61098 328713 79438 408151 0.1946289
# 1   01  1252 289364 290616 117535 408151 0.2879694
# 63  77 288335 981 289316 118835 408151 0.2911545 < L
#   pat  left  right  both  cross  all  ratio  best
# 8   10  1115 332625 333740 74945 408685 0.1833808 < R B C O
# 18  22 267324 61658 328982 79703 408685 0.1950231
# 1   01  1226 289045 290271 118414 408685 0.2897439
# 63  77 288062 967 289029 119656 408685 0.2927830 < L
#   pat  left  right  both  cross  all  ratio  best
# 8   10  1147 332068 333215 75372 408587 0.1844699 < R B C O
# 18  22 267629 61237 328866 79721 408587 0.1951139
# 1   01  1220 289521 290741 117846 408587 0.2884233
# 63  77 288475 921 289396 119191 408587 0.2917151 < L
#   pat  left  right  both  cross  all  ratio  best
# 8   10  1094 332239 333333 75144 408477 0.1839614 < R B C O
# 18  22 267590 61319 328909 79568 408477 0.1947919
# 1   01  1209 289230 290439 118038 408477 0.2889710
# 63  77 288291 936 289227 119250 408477 0.2919381 < L
#   pat  left  right  both  cross  all  ratio  best
# 8   10  1129 331922 333051 75091 408142 0.1839825 < R B C O
# 18  22 267142 61339 328481 79661 408142 0.1951796
# 1   01  1183 288932 290115 118027 408142 0.2891812
# 63  77 287829 914 288743 119399 408142 0.2925428 < L
#   pat  left  right  both  cross  all  ratio  best
# 8   10  1078 331642 332720 75177 407897 0.1843039 < R B C O
# 18  22 267228 61500 328728 79169 407897 0.1940907
# 1   01  1221 289013 290234 117663 407897 0.2884625
# 63  77 287876 933 288809 119088 407897 0.2919561 < L
#   pat  left  right  both  cross  all  ratio  best

```

```

# 8 10 1108 332043 333151 75154 408305 0.1840634 < R B C O
# 18 22 267364 61672 329036 79269 408305 0.1941416
# 1 01 1177 289100 290277 118028 408305 0.2890682
# 63 77 288049 914 288963 119342 408305 0.2922864 < L
# pat left right both cross all ratio best
# 8 10 1134 331690 332824 75413 408237 0.1847285 < R B C O
# 18 22 267249 61601 328850 79387 408237 0.1944630
# 1 01 1204 288856 290060 118177 408237 0.2894814
# 63 77 287661 938 288599 119638 408237 0.2930602 < L
# pat left right both cross all ratio best
# 8 10 1169 331836 333005 75157 408162 0.1841352 < R B C O
# 18 22 267443 61137 328580 79582 408162 0.1949765
# 1 01 1246 289175 290421 117741 408162 0.2884663
# 63 77 288198 960 289158 119004 408162 0.2915607 < L
# pat left right both cross all ratio best
# 8 10 1104 332220 333324 75231 408555 0.1841392 < R B C O
# 18 22 267058 61409 328467 80088 408555 0.1960275
# 1 01 1253 289103 290356 118199 408555 0.2893099
# 63 77 287999 960 288959 119596 408555 0.2927293 < L
# pat left right both cross all ratio best
# 8 10 1102 332340 333442 74746 408188 0.1831166 < R B C O
# 18 22 267721 61287 329008 79180 408188 0.1939792
# 1 01 1142 289604 290746 117442 408188 0.2877155
# 63 77 288502 885 289387 118801 408188 0.2910448 < L
# pat left right both cross all ratio best
# 8 10 1096 332522 333618 74810 408428 0.1831657 < R B C O
# 18 22 267564 61396 328960 79468 408428 0.1945704
# 1 01 1231 289156 290387 118041 408428 0.2890130
# 63 77 288348 926 289274 119154 408428 0.2917381 < L
# pat left right both cross all ratio best
# 8 10 1099 332217 333316 74923 408239 0.1835273 < R B C O
# 18 22 267385 61375 328760 79479 408239 0.1946874
# 1 01 1197 289117 290314 117925 408239 0.2888627
# 63 77 288070 946 289016 119223 408239 0.2920422 < L
# pat left right both cross all ratio best
# 8 10 1151 332282 333433 74768 408201 0.1831647 < R B C O
# 18 22 267648 61608 329256 78945 408201 0.1933974
# 1 01 1229 289252 290481 117720 408201 0.2883873
# 63 77 288379 948 289327 118874 408201 0.2912144 < L
# pat left right both cross all ratio best
# 8 10 1118 331932 333050 75132 408182 0.1840650 < R B C O
# 18 22 267582 61339 328921 79261 408182 0.1941805
# 1 01 1225 289232 290457 117725 408182 0.2884130
# 63 77 288235 956 289191 118991 408182 0.2915146 < L
# pat left right both cross all ratio best
# 8 10 1045 332077 333122 75027 408149 0.1838226 < R B C O
# 18 22 267395 61457 328852 79297 408149 0.1942844
# 1 01 1193 288978 290171 117978 408149 0.2890562
# 63 77 288010 891 288901 119248 408149 0.2921678 < L
# pat left right both cross all ratio best
# 8 10 1178 332105 333283 74818 408101 0.1833321 < R B C O
# 18 22 267564 61296 328860 79241 408101 0.1941701
# 1 01 1292 289112 290404 117697 408101 0.2884016
# 63 77 288323 930 289253 118848 408101 0.2912220 < L
# pat left right both cross all ratio best
# 8 10 1123 332123 333246 75316 408562 0.1843441 < R B C O
# 18 22 267365 61401 328766 79796 408562 0.1953094
# 1 01 1228 289097 290325 118237 408562 0.2893979
# 63 77 288070 965 289035 119527 408562 0.2925554 < L
# pat left right both cross all ratio best
# 8 10 1109 331845 332954 74932 407886 0.1837082 < R B C O
# 18 22 267420 61249 328669 79217 407886 0.1942136
# 1 01 1211 288896 290107 117779 407886 0.2887547
# 63 77 287997 930 288927 118959 407886 0.2916477 < L
# pat left right both cross all ratio best
# 8 10 1152 331872 333024 75339 408363 0.1844903 < R B C O
# 18 22 267153 61616 328769 79594 408363 0.1949099

```

```

# 1 01 1232 288949 290181 118182 408363 0.2894043
# 63 77 287763 957 288720 119643 408363 0.2929820 < L
# pat left right both cross all ratio best
# 8 10 1067 332072 333139 75272 408411 0.1843045 < R B C O
# 18 22 267734 61335 329069 79342 408411 0.1942700
# 1 01 1188 289283 290471 117940 408411 0.2887777
# 63 77 288273 931 289204 119207 408411 0.2918800 < L
# pat left right both cross all ratio best
# 8 10 1107 332585 333692 74967 408659 0.1834463 < R B C O
# 18 22 267848 61315 329163 79496 408659 0.1945289
# 1 01 1163 289634 290797 117862 408659 0.2884116
# 63 77 288339 917 289256 119403 408659 0.2921825 < L
# pat left right both cross all ratio best
# 8 10 1111 331964 333075 75043 408118 0.1838757 < R B C O
# 18 22 267208 61212 328420 79698 408118 0.1952818
# 1 01 1245 288957 290202 117916 408118 0.2889262
# 63 77 287952 902 288854 119264 408118 0.2922292 < L
# pat left right both cross all ratio best
# 8 10 1117 331600 332717 75224 407941 0.1843992 < R B C O
# 18 22 267370 61480 328850 79091 407941 0.1938785
# 1 01 1211 289091 290302 117639 407941 0.2883726
# 63 77 287962 927 288889 119052 407941 0.2918363 < L
# pat left right both cross all ratio best
# 8 10 1110 332218 333328 74679 408007 0.1830336 < R B C O
# 18 22 267669 61266 328935 79072 408007 0.1938006
# 1 01 1252 289330 290582 117425 408007 0.2878014
# 63 77 288511 920 289431 118576 408007 0.2906225 < L
# pat left right both cross all ratio best
# 8 10 1156 332267 333423 74685 408108 0.1830030 < R B C O
# 18 22 267426 61251 328677 79431 408108 0.1946323
# 1 01 1209 289277 290486 117622 408108 0.2882129
# 63 77 287916 924 288840 119268 408108 0.2922462 < L
# pat left right both cross all ratio best
# 8 10 1077 331827 332904 75204 408108 0.1842748 < R B C O
# 18 22 267213 60935 328148 79960 408108 0.1959285
# 1 01 1171 288869 290040 118068 408108 0.2893058
# 63 77 288131 886 289017 119091 408108 0.2918125 < L
# pat left right both cross all ratio best
# 8 10 1099 332443 333542 75194 408736 0.1839672 < R B C O
# 18 22 267643 61262 328905 79831 408736 0.1953119
# 1 01 1207 289344 290551 118185 408736 0.2891475
# 63 77 288192 920 289112 119624 408736 0.2926681 < L
# pat left right both cross all ratio best
# 8 10 1076 332146 333222 74863 408085 0.1834495 < R B C O
# 18 22 267763 61240 329003 79082 408085 0.1937881
# 1 01 1267 289142 290409 117676 408085 0.2883615
# 63 77 288460 909 289369 118716 408085 0.2909100 < L
# pat left right both cross all ratio best
# 8 10 1077 332176 333253 75044 408297 0.1837976 < R B C O
# 18 22 267789 61603 329392 78905 408297 0.1932539
# 1 01 1164 289199 290363 117934 408297 0.2888437
# 63 77 288326 907 289233 119064 408297 0.2916113 < L
# pat left right both cross all ratio best
# 8 10 1098 332186 333284 75189 408473 0.1840734 < R B C O
# 18 22 267816 61180 328996 79477 408473 0.1945710
# 1 01 1163 289497 290660 117813 408473 0.2884230
# 63 77 288547 855 289402 119071 408473 0.2915027 < L
# pat left right both cross all ratio best
# 8 10 1098 332046 333144 75150 408294 0.1840585 < R B C O
# 18 22 267518 61353 328871 79423 408294 0.1945240
# 1 01 1228 288934 290162 118132 408294 0.2893307
# 63 77 288237 902 289139 119155 408294 0.2918363 < L
# pat left right both cross all ratio best
# 8 10 1066 332545 333611 75075 408686 0.1836985 < R B C O
# 18 22 267809 61506 329315 79371 408686 0.1942102
# 1 01 1236 289204 290440 118246 408686 0.2893322
# 63 77 288457 886 289343 119343 408686 0.2920164 < L

```

#	pat	left	right	both	cross	all	ratio	best
# 8	10	1048	332753	333801	74630	408431	0.1827236	< R B C O
# 18	22	268561	61022	329583	78848	408431	0.1930510	
# 1	01	1187	289914	291101	117330	408431	0.2872701	
# 63	77	289021	844	289865	118566	408431	0.2902963	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1080	332679	333759	74430	408189	0.1823420	< R B C O
# 18	22	267689	61137	328826	79363	408189	0.1944271	
# 1	01	1174	289272	290446	117743	408189	0.2884522	
# 63	77	288140	971	289111	119078	408189	0.2917227	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1081	332018	333099	75086	408185	0.1839509	< R B C O
# 18	22	267605	61006	328611	79574	408185	0.1949459	
# 1	01	1166	289258	290424	117761	408185	0.2884991	
# 63	77	288192	904	289096	119089	408185	0.2917525	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1114	331620	332734	75019	407753	0.1839815	< R B C O
# 18	22	266972	61317	328289	79464	407753	0.1948827	
# 1	01	1189	288513	289702	118051	407753	0.2895160	
# 63	77	287622	912	288534	119219	407753	0.2923804	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1130	332138	333268	75042	408310	0.1837868	< R B C O
# 18	22	267820	61231	329051	79259	408310	0.1941148	
# 1	01	1193	289360	290553	117757	408310	0.2884010	
# 63	77	288265	926	289191	119119	408310	0.2917367	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1113	332374	333487	75242	408729	0.1840877	< R B C O
# 18	22	267641	61564	329205	79524	408729	0.1945641	
# 1	01	1236	289184	290420	118309	408729	0.2894558	
# 63	77	288468	898	289366	119363	408729	0.2920346	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1176	332239	333415	74775	408190	0.1831868	< R B C O
# 18	22	267737	61470	329207	78983	408190	0.1934957	
# 1	01	1229	289220	290449	117741	408190	0.2884466	
# 63	77	288216	934	289150	119040	408190	0.2916289	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1088	332082	333170	75305	408475	0.1843564	< R B C O
# 18	22	267763	61011	328774	79701	408475	0.1951184	
# 1	01	1202	289255	290457	118018	408475	0.2889234	
# 63	77	288539	876	289415	119060	408475	0.2914744	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1081	332029	333110	75249	408359	0.1842717	< R B C O
# 18	22	267378	61184	328562	79797	408359	0.1954089	
# 1	01	1196	289257	290453	117906	408359	0.2887312	
# 63	77	288070	937	289007	119352	408359	0.2922722	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1105	331970	333075	75010	408085	0.1838097	< R B C O
# 18	22	267456	61316	328772	79313	408085	0.1943541	
# 1	01	1198	289263	290461	117624	408085	0.2882341	
# 63	77	288142	933	289075	119010	408085	0.2916304	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1117	331732	332849	75407	408256	0.1847052	< R B C O
# 18	22	267113	61663	328776	79480	408256	0.1946818	
# 1	01	1224	288880	290104	118152	408256	0.2894066	
# 63	77	287705	959	288664	119592	408256	0.2929338	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1137	332017	333154	75210	408364	0.1841739	< R B C O
# 18	22	267440	61708	329148	79216	408364	0.1939838	
# 1	01	1189	289133	290322	118042	408364	0.2890607	
# 63	77	287958	957	288915	119449	408364	0.2925062	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1060	332127	333187	74769	407956	0.1832771	< R B C O
# 18	22	267320	61196	328516	79440	407956	0.1947269	
# 1	01	1201	288758	289959	117997	407956	0.2892395	
# 63	77	287988	894	288882	119074	407956	0.2918795	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1091	331836	332927	75518	408445	0.1848915	< R B C O

```

# 18 22 266893 61841 328734 79711 408445 0.1951572
# 1 01 1206 288799 290005 118440 408445 0.2899778
# 63 77 287810 926 288736 119709 408445 0.2930847 < L
# pat left right both cross all ratio best
# 8 10 1096 331558 332654 75098 407752 0.1841757 < R B C O
# 18 22 267263 61350 328613 79139 407752 0.1940861
# 1 01 1194 288884 290078 117674 407752 0.2885921
# 63 77 287955 913 288868 118884 407752 0.2915596 < L
# pat left right both cross all ratio best
# 8 10 1092 332374 333466 74958 408424 0.1835299 < R B C O
# 18 22 267481 61191 328672 79752 408424 0.1952677
# 1 01 1179 289344 290523 117901 408424 0.2886730
# 63 77 288481 896 289377 119047 408424 0.2914790 < L
# pat left right both cross all ratio best
# 8 10 1137 332326 333463 74819 408282 0.1832532 < R B C O
# 18 22 267629 61063 328692 79590 408282 0.1949388
# 1 01 1221 289539 290760 117522 408282 0.2878452
# 63 77 288445 935 289380 118902 408282 0.2912252 < L
# pat left right both cross all ratio best
# 8 10 1082 332312 333394 74641 408035 0.1829279 < R B C O
# 18 22 267054 61549 328603 79432 408035 0.1946696
# 1 01 1164 288868 290032 118003 408035 0.2891982
# 63 77 287738 896 288634 119401 408035 0.2926244 < L
# pat left right both cross all ratio best
# 8 10 1098 332518 333616 74739 408355 0.1830246 < R B C O
# 18 22 267725 61551 329276 79079 408355 0.1936526
# 1 01 1218 289516 290734 117621 408355 0.2880361
# 63 77 288406 910 289316 119039 408355 0.2915086 < L
# pat left right both cross all ratio best
# 8 10 1071 331552 332623 75435 408058 0.1848634 < R B C O
# 18 22 267017 61378 328395 79663 408058 0.1952247
# 1 01 1133 288711 289844 118214 408058 0.2896990
# 63 77 287646 882 288528 119530 408058 0.2929240 < L
# pat left right both cross all ratio best
# 8 10 1098 332105 333203 74825 408028 0.1833820 < R B C O
# 18 22 268130 61091 329221 78807 408028 0.1931412
# 1 01 1209 289421 290630 117398 408028 0.2877205
# 63 77 288769 901 289670 118358 408028 0.2900732 < L
# pat left right both cross all ratio best
# 8 10 1056 332349 333405 74768 408173 0.1831772 < R B C O
# 18 22 267617 61250 328867 79306 408173 0.1942951
# 1 01 1198 289261 290459 117714 408173 0.2883924
# 63 77 288234 911 289145 119028 408173 0.2916116 < L
# pat left right both cross all ratio best
# 8 10 1134 331631 332765 75151 407916 0.1842316 < R B C O
# 18 22 266934 61563 328497 79419 407916 0.1946945
# 1 01 1224 288614 289838 118078 407916 0.2894665
# 63 77 287736 967 288703 119213 407916 0.2922489 < L
# pat left right both cross all ratio best
# 8 10 1077 332415 333492 74725 408217 0.1830522 < R B C O
# 18 22 267611 61114 328725 79492 408217 0.1947298
# 1 01 1247 289414 290661 117556 408217 0.2879743
# 63 77 288339 947 289286 118931 408217 0.2913426 < L
# pat left right both cross all ratio best
# 8 10 1091 332027 333118 75453 408571 0.1846754 < R B C O
# 18 22 267451 61612 329063 79508 408571 0.1946002
# 1 01 1165 289098 290263 118308 408571 0.2895653
# 63 77 288092 895 288987 119584 408571 0.2926884 < L
# pat left right both cross all ratio best
# 8 10 1057 332387 333444 74868 408312 0.1833598 < R B C O
# 18 22 267732 61419 329151 79161 408312 0.1938738
# 1 01 1207 289431 290638 117674 408312 0.2881963
# 63 77 288514 905 289419 118893 408312 0.2911817 < L
# pat left right both cross all ratio best
# 8 10 1101 332116 333217 75086 408303 0.1838977 < R B C O
# 18 22 267164 61255 328419 79884 408303 0.1956488
# 1 01 1200 288871 290071 118232 408303 0.2895693

```

```

# 63 77 287998 953 288951 119352 408303 0.2923123 < L
# pat left right both cross all ratio best
# 8 10 1069 332419 333488 74856 408344 0.1833160 < R B C O
# 18 22 267621 61374 328995 79349 408344 0.1943190
# 1 01 1159 289124 290283 118061 408344 0.2891214
# 63 77 288327 889 289216 119128 408344 0.2917344 < L
# pat left right both cross all ratio best
# 8 10 1148 332556 333704 74841 408545 0.1831891 < R B C O
# 18 22 267952 61122 329074 79471 408545 0.1945220
# 1 01 1240 289552 290792 117753 408545 0.2882253
# 63 77 288516 955 289471 119074 408545 0.2914587 < L
# pat left right both cross all ratio best
# 8 10 1101 332137 333238 74929 408167 0.1835744 < R B C O
# 18 22 267922 61656 329578 78589 408167 0.1925413
# 1 01 1223 289571 290794 117373 408167 0.2875612
# 63 77 288715 895 289610 118557 408167 0.2904620 < L
# pat left right both cross all ratio best
# 8 10 1133 332067 333200 75143 408343 0.1840193 < R B C O
# 18 22 267756 61238 328994 79349 408343 0.1943195
# 1 01 1204 289116 290320 118023 408343 0.2890291
# 63 77 288490 931 289421 118922 408343 0.2912307 < L
# pat left right both cross all ratio best
# 8 10 1072 332726 333798 74937 408735 0.1833388 < R B C O
# 18 22 268007 61152 329159 79576 408735 0.1946885
# 1 01 1214 289526 290740 117995 408735 0.2886834
# 63 77 288542 897 289439 119296 408735 0.2918664 < L
# pat left right both cross all ratio best
# 8 10 1060 331873 332933 75398 408331 0.1846492 < R B C O
# 18 22 266840 61684 328524 79807 408331 0.1954468
# 1 01 1207 288474 289681 118650 408331 0.2905731
# 63 77 287491 844 288335 119996 408331 0.2938694 < L
# pat left right both cross all ratio best
# 8 10 1131 332016 333147 75348 408495 0.1844527 < R B C O
# 18 22 267402 61661 329063 79432 408495 0.1944504
# 1 01 1187 289224 290411 118084 408495 0.2890709
# 63 77 288210 877 289087 119408 408495 0.2923120 < L
# pat left right both cross all ratio best
# 8 10 1121 332634 333755 75084 408839 0.1836518 < R B C O
# 18 22 267858 61407 329265 79574 408839 0.1946341
# 1 01 1224 289747 290971 117868 408839 0.2882993
# 63 77 288530 902 289432 119407 408839 0.2920636 < L
# pat left right both cross all ratio best
# 8 10 1057 332011 333068 75162 408230 0.1841168 < R B C O
# 18 22 267047 61710 328757 79473 408230 0.1946770
# 1 01 1211 288611 289822 118408 408230 0.2900522
# 63 77 287751 897 288648 119582 408230 0.2929280 < L
# pat left right both cross all ratio best
# 8 10 1005 332163 333168 74892 408060 0.1835318 < R B C O
# 18 22 267603 61192 328795 79265 408060 0.1942484
# 1 01 1114 289175 290289 117771 408060 0.2886120
# 63 77 288165 866 289031 119029 408060 0.2916948 < L
# pat left right both cross all ratio best
# 8 10 1128 331715 332843 75146 407989 0.1841863 < R B C O
# 18 22 266872 61518 328390 79599 407989 0.1951008
# 1 01 1239 288474 289713 118276 407989 0.2899000
# 63 77 287614 902 288516 119473 407989 0.2928339 < L
# pat left right both cross all ratio best
# 8 10 1153 331873 333026 75528 408554 0.1848666 < R B C O
# 18 22 267239 61693 328932 79622 408554 0.1948873
# 1 01 1218 289133 290351 118203 408554 0.2893204
# 63 77 288098 926 289024 119530 408554 0.2925684 < L
# pat left right both cross all ratio best
# 8 10 1181 332057 333238 75164 408402 0.1840442 < R B C O
# 18 22 267262 61364 328626 79776 408402 0.1953369
# 1 01 1232 289008 290240 118162 408402 0.2893277
# 63 77 287991 913 288904 119498 408402 0.2925990 < L
# pat left right both cross all ratio best

```

```

# 8 10 1085 332114 333199 75384 408583 0.1845011 < R B C O
# 18 22 267795 61501 329296 79287 408583 0.1940536
# 1 01 1189 289403 290592 117991 408583 0.2887810
# 63 77 288337 904 289241 119342 408583 0.2920875 < L
# pat left right both cross all ratio best
# 8 10 1108 332117 333225 74967 408192 0.1836562 < R B C O
# 18 22 268019 60927 328946 79246 408192 0.1941390
# 1 01 1173 289434 290607 117585 408192 0.2880630
# 63 77 288403 929 289332 118860 408192 0.2911865 < L
# pat left right both cross all ratio best
# 8 10 1074 332681 333755 74654 408409 0.1827922 < R B C O
# 18 22 268204 60934 329138 79271 408409 0.1940971
# 1 01 1233 289838 291071 117338 408409 0.2873051
# 63 77 288804 935 289739 118670 408409 0.2905666 < L
# pat left right both cross all ratio best
# 8 10 1081 331931 333012 75016 408028 0.1838501 < R B C O
# 18 22 267248 61314 328562 79466 408028 0.1947562
# 1 01 1192 288850 290042 117986 408028 0.2891615
# 63 77 287800 898 288698 119330 408028 0.2924554 < L
# pat left right both cross all ratio best
# 8 10 1098 332250 333348 74978 408326 0.1836229 < R B C O
# 18 22 267508 61115 328623 79703 408326 0.1951945
# 1 01 1210 289169 290379 117947 408326 0.2888550
# 63 77 288035 885 288920 119406 408326 0.2924281 < L
# pat left right both cross all ratio best
# 8 10 1062 331758 332820 75472 408292 0.1848481 < R B C O
# 18 22 267169 61442 328611 79681 408292 0.1951569
# 1 01 1197 288874 290071 118221 408292 0.2895501
# 63 77 287845 897 288742 119550 408292 0.2928051 < L
# pat left right both cross all ratio best
# 8 10 1126 332248 333374 75028 408402 0.1837111 < R B C O
# 18 22 267470 61188 328658 79744 408402 0.1952586
# 1 01 1150 289188 290338 118064 408402 0.2890877
# 63 77 288350 892 289242 119160 408402 0.2917713 < L
# pat left right both cross all ratio best
# 8 10 1102 331714 332816 75144 407960 0.1841945 < R B C O
# 18 22 267284 61325 328609 79351 407960 0.1945068
# 1 01 1216 288884 290100 117860 407960 0.2889009
# 63 77 287923 948 288871 119089 407960 0.2919134 < L
# pat left right both cross all ratio best
# 8 10 1073 332654 333727 74606 408333 0.1827087 < R B C O
# 18 22 267923 61313 329236 79097 408333 0.1937071
# 1 01 1194 289413 290607 117726 408333 0.2883088
# 63 77 288537 905 289442 118891 408333 0.2911619 < L
# pat left right both cross all ratio best
# 8 10 1115 331827 332942 75207 408149 0.1842636 < R B C O
# 18 22 267373 61532 328905 79244 408149 0.1941546
# 1 01 1239 289020 290259 117890 408149 0.2888406
# 63 77 287967 943 288910 119239 408149 0.2921458 < L
# pat left right both cross all ratio best
# 8 10 1096 331728 332824 75524 408348 0.1849501 < R B C O
# 18 22 267499 61620 329119 79229 408348 0.1940232
# 1 01 1190 289150 290340 118008 408348 0.2889888
# 63 77 288115 904 289019 119329 408348 0.2922238 < L
# pat left right both cross all ratio best
# 8 10 1126 331817 332943 75278 408221 0.1844050 < R B C O
# 18 22 267225 61408 328633 79588 408221 0.1949630
# 1 01 1224 288880 290104 118117 408221 0.2893457
# 63 77 287983 901 288884 119337 408221 0.2923343 < L
# pat left right both cross all ratio best
# 8 10 1104 332480 333584 74763 408347 0.1830869 < R B C O
# 18 22 267561 61384 328945 79402 408347 0.1944474
# 1 01 1232 289180 290412 117935 408347 0.2888107
# 63 77 288409 862 289271 119076 408347 0.2916049 < L
# pat left right both cross all ratio best
# 8 10 1114 332099 333213 75182 408395 0.1840914 < R B C O
# 18 22 267363 61708 329071 79324 408395 0.1942335

```



```

# 1 01 1204 289092 290296 118099 408395 0.2891784
# 63 77 288290 869 289159 119236 408395 0.2919624 < L
# pat left right both cross all ratio best
# 8 10 1125 332695 333820 74685 408505 0.1828252 < R B C O
# 18 22 267604 61633 329237 79268 408505 0.1940441
# 1 01 1269 289222 290491 118014 408505 0.2888924
# 63 77 288106 947 289053 119452 408505 0.2924126 < L
# pat left right both cross all ratio best
# 8 10 1066 332944 334010 75159 409169 0.1836869 < R B C O
# 18 22 268341 61323 329664 79505 409169 0.1943085
# 1 01 1205 289938 291143 118026 409169 0.2884529
# 63 77 288912 951 289863 119306 409169 0.2915812 < L
# pat left right both cross all ratio best
# 8 10 1117 332286 333403 74972 408375 0.1835862 < R B C O
# 18 22 267625 61636 329261 79114 408375 0.1937288
# 1 01 1217 289401 290618 117757 408375 0.2883551
# 63 77 288278 916 289194 119181 408375 0.2918421 < L
# pat left right both cross all ratio best
# 8 10 1115 332162 333277 74871 408148 0.1834408 < R B C O
# 18 22 267735 61234 328969 79179 408148 0.1939958
# 1 01 1268 289409 290677 117471 408148 0.2878147
# 63 77 288359 921 289280 118868 408148 0.2912375 < L
# pat left right both cross all ratio best
# 8 10 1155 331780 332935 75515 408450 0.1848819 < R B C O
# 18 22 267313 61278 328591 79859 408450 0.1955172
# 1 01 1236 288846 290082 118368 408450 0.2897980
# 63 77 288241 937 289178 119272 408450 0.2920113 < L
# pat left right both cross all ratio best
# 8 10 1082 332301 333383 75049 408432 0.1837491 < R B C O
# 18 22 268080 61236 329316 79116 408432 0.1937067
# 1 01 1218 289711 290929 117503 408432 0.2876929
# 63 77 288710 950 289660 118772 408432 0.2907999 < L
# pat left right both cross all ratio best
# 8 10 1115 331740 332855 75429 408284 0.1847464 < R B C O
# 18 22 267114 61686 328800 79484 408284 0.1946782
# 1 01 1156 288920 290076 118208 408284 0.2895240
# 63 77 287972 945 288917 119367 408284 0.2923627 < L
# pat left right both cross all ratio best
# 8 10 1116 332395 333511 74763 408274 0.1831197 < R B C O
# 18 22 267226 61297 328523 79751 408274 0.1953370
# 1 01 1286 288827 290113 118161 408274 0.2894159
# 63 77 287756 942 288698 119576 408274 0.2928817 < L
# pat left right both cross all ratio best
# 8 10 1083 331731 332814 75098 407912 0.1841034 < R B C O
# 18 22 267237 61655 328892 79020 407912 0.1937183
# 1 01 1216 288726 289942 117970 407912 0.2892045
# 63 77 287693 917 288610 119302 407912 0.2924699 < L
# pat left right both cross all ratio best
# 8 10 1132 332497 333629 74856 408485 0.1832528 < R B C O
# 18 22 267630 61654 329284 79201 408485 0.1938896
# 1 01 1225 289078 290303 118182 408485 0.2893178
# 63 77 288228 933 289161 119324 408485 0.2921135 < L
# pat left right both cross all ratio best
# 8 10 1081 331712 332793 74993 407786 0.1839028 < R B C O
# 18 22 267312 61105 328417 79369 407786 0.1946340
# 1 01 1209 288948 290157 117629 407786 0.2884577
# 63 77 287747 864 288611 119175 407786 0.2922489 < L
# pat left right both cross all ratio best
# 8 10 1090 332121 333211 74760 407971 0.1832483 < R B C O
# 18 22 267570 61236 328806 79165 407971 0.1940457
# 1 01 1303 289219 290522 117449 407971 0.2878857
# 63 77 288124 940 289064 118907 407971 0.2914594 < L

# summarize:
corsummary <- t(as.matrix(c(summary(bcor),sd=sd(bcor))))
row.names(corsummary) <- 'bcor'
bdelta <- b61cross-b52cross
brevp <- 100*brev # make it percent reversed instead of logical

```

```
thesummary <- rbind(summary(b52cross), summary(b61cross), summary(c(bdelta)), summary(brevp))
row.names(thesummary) <- c('b52cross', 'b61cross', 'b61-b52', '% rev')
thesummary <- cbind(thesummary, sd=c(sd(b52cross), sd(b61cross), sd(bdelta), sd(brevp)))
```

SUMMARY: In 101 bootstrap replicates, we saw 101 samples with the 6/1 split having fewer conflicts than the 5/2 split, and the minimum separation between them was ≈ -18 sigma, hence highly statistically significant.

```
# 'opt' hacking is trying to force knitr to show more digits of bcor in summary, as Rstudio does, but
# it still fails... Bottom line is the correlation seems to be > .999 in all samples, rounds to 1.0,
# as seen in this run of 1001 samples cut/paste from Rstudio:
#      Min.      1st Qu.      Median      Mean      3rd Qu.      Max.      sd
# bcor   " 0.9998" " 0.9999" " 0.9999" " 0.9999" " 1" " 1" " 0.00003462"
# > max(bcor)
# [1] 0.9999915
o.opts <- options(digits=7,width=127)
format(rbind(corsummary,thesummary), scientific=F, digits=4, drop0trailing=T)

#      Min.      1st Qu.      Median      Mean      3rd Qu.      Max.
# bcor   " 0.999973432" " 0.999991997" " 0.99999417" " 0.999993603" " 0.999995979" " 0.999998
# b52cross "78589"      "79244"      "79432"      "79415.267326733" "79590"      "80088"
# b61cross "74430"      "74856"      "75049"      "75047.534653465" "75204"      "75528"
# b61-b52  "-4988"      "-4581"      "-4346"      "-4367.732673267" "-4206"      "-3660"
# % rev    " 100"      " 100"      " 100"      " 100"      " 100"      " 100"
#      sd
# bcor   " 0.000003499"
# b52cross " 268.69067312"
# b61cross " 246.249327486"
# b61-b52  " 276.221754795"
# % rev    " 0"

options(o.opts)
```

Based on this, it is reasonable to claim that we are confident that the tree topology is as shown in the earlier figures, with the exception of the exact order of the splits with the 4 NE coastal isolates.

10 Notes

This section is a random brain dump of limitations of the current analysis, ideas for improvements, etc. In the main, these may not be worth doing, unless we see significant holes or get pushed by reviewers, etc, but I wanted to catalog before we forget them.

Noise: Various sources of “noise” in the data:

1. Read errors, low read depth — perhaps fixed by medium/strict thresholding
2. Deep coalescence
3. Skew because 1335 is the reference. (Julie notes we could partially fix this by remapping based on discovered SNPs, tho that wouldn't fix gross misassembly in 1335, e.g. collapsed or misordered tandem duplicates, or segments missing in 1335 that are present in one or more other strains, etc.; much harder to fix those, let's just hope they are rare...)
4. Varying error rates and sequencing depth among the 7. E.g., plausibly the 1000 SNPs shared by 4 but not by Gyre are a result of lower read depth (we missed a SNP that is actually present) and/or higher error rates (causing a position to appear inconsistent in gyre) in the gyre data. I can't think of a way to correct for this effect. It might be possible, perhaps by simulation, to estimate the size of the effect and see whether it could explain ≈ 1000 SNPs.
5. Varying numbers of founder cells in the sequencing cultures. (Again, I made some attempts at modeling this, but nothing very satisfactory yet.)

6. Tri-allelic positions where stochastic fluctuation in sequence sampling promotes the rare allele to prominence. (Julie replies: “isn’t this the same as more than one founder cell? If they are diploid there should only ever be two alleles, unless there were random and very rare, thus unlikely, trisomy events?” I agree, but it is a concrete example of an effect of multiple founders that might be important. Not sure this is the most important such effect...)
7. Gaps/indels - alignments are likely to be of lower quality in the vicinity of an indel, so, maybe lower coverage/more SNPs. We ignored them. Does this add any systematic bias? e.g. if one strain had more indels than another, would this confound other analyses? unclear. Julie suggested a paper titled “Barking up the wrong tree-length: yada yada yada gap penalties”; maybe relevant?

Other Items/Potential To Dos:

1. any spacial structure to various sub-classes?
2. after top level split, should I reanalyze halves of partition in isolation? said another way, I think the tree-building is sensible, but not sure it’s optimal.
3. if we believe no sex, then I think gain of SNP should be more common than loss of SNP, since the later can only happen by (a) mutation reverting to reference, (b) second mutation matching nonreference, (c) homologous repair (look for blocks of LOH), or (d) false negative e.g. from low read depth. Does tree-building appropriately weight the gain vs loss cases? (Does it even care?)
4. should we weight coding and/or nonsynonymous SNPs more heavily? Julie says “you do not want to weight the coding or nonsynonymous/coding SNPs because for time you want the more clock-like neutral mutations.” I.e., I got this backwards. Maybe should redo tree based on noncoding SNPs only.
5. We could also do an actual parsimony analysis based on 2-state model (homozygous-ref vs not), but I’m not quite sure how to handle this in a mixed sex/nosex case.
6. Might be interesting to look at sharing just within (shared?) deserts. Given tree model above and expectation that bottleneck followed split of H- from L-clades, I would expect little or no sharing of L-clade desert SNPs with H-clade; sharing between It/Wales might suggest “desert” is actually a region under strong purifying selection (e.g. a gene); sharing/non-sharing within L-clade deserts might suggest more about evo history of the 5.

11 Appendix: Old Trees, etc.

Tangents, old stuff of historical interest at best, etc..

11.1 HWE Sharing

Tangent: As a function of nonref allele freq, assuming HWE, what is probability that nonref allele will be seen in k strains, $0 \leq k \leq 4$ (Fig 6).

```
myfigpath.h <- paste(getwd(), '/figs-knitr/', sep='')

```

```
p <- (0:20)/20
q <- 1-p
r <- 2*p*q+p^2
plot(p, 1*q^0*r^4, type='b', pch='4', ylab="share prob")
points(p, 4*q^2*r^3, type='b', pch='3')
points(p, 6*q^4*r^2, type='b', pch='2')
points(p, 4*q^6*r^1, type='b', pch='1')
points(p, 1*q^8*r^0, type='b', pch='0')
```

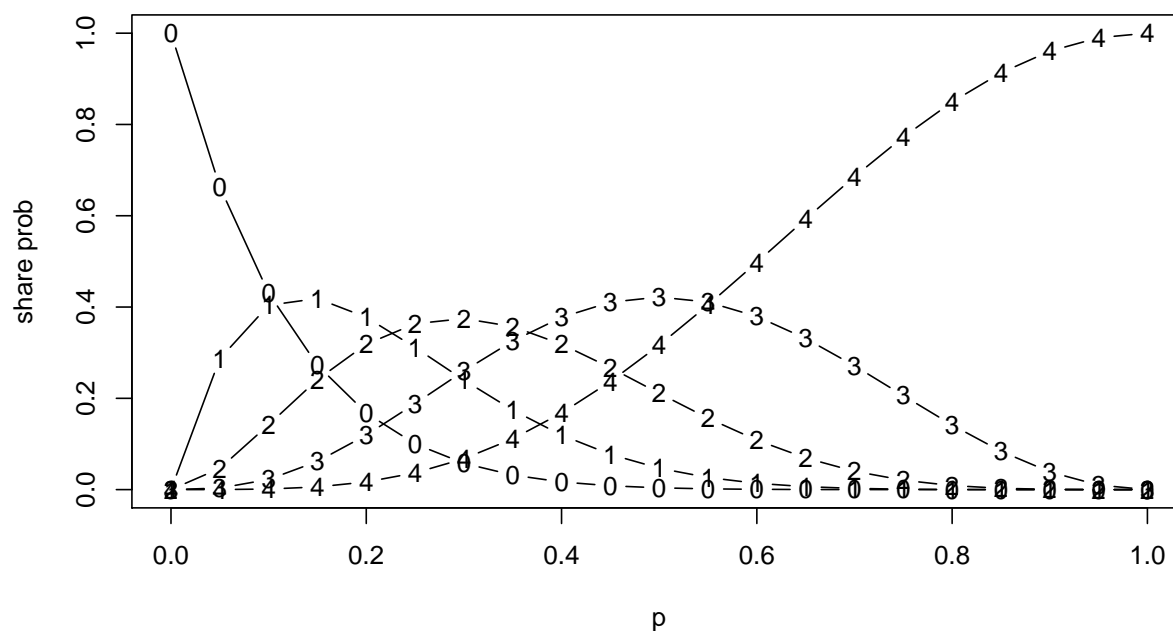


Figure 6: Sharing Probability

11.2 Old Tree Stuff

All based on un-q-filtered reads.

The first pass at the tree analysis was the Chr1 tree, *loose criteria* (c1); it is rendered via <http://iubio.bio.indiana.edu/treeapp/treeprint-form.html> as Fig 7, and in newick format is:

```
newick.chr1.loose <- '(((tp3367_Italy:4551,tp1013_Wales:4954):5920,((tp1007_Virginia:10,tp1012_Australia:29):9,
cat.hardwrap(newick.chr1.loose)

# (((tp3367_Italy:4551,tp1013_Wales:4954):5920,((tp1007_Virginia:10,tp1012_Austra
# lia:29):9,(tp1015_Puget_Sound:90,tp1335_NY:13):11):320,tp1014_Gyre:22):3484):859
# 3,outgroup:0);
```

Chr 1 tree based on *medium criteria* (c2) has exactly the same topology is, although the branch lengths are different. As noted earlier, the length of the branch labeled “*” is probably inflated by lower coverage and higher error rate in 1014, which may mask further legitimate sharing between it and the other L-isolates. The branch lengths among the other 4 are too short for its topology to be convincing without a more rigorous analysis (e.g., a bootstrap test).

Chr1 tree, medium criteria, in newick format:

```
newick.chr1.med <- '(((tp3367_Italy:8813,tp1013_Wales:9652):9365,((e_tp1007_Virginia:30,d_tp1012_Australia:61):1
cat.hardwrap(newick.chr1.med)

# (((tp3367_Italy:8813,tp1013_Wales:9652):9365,((e_tp1007_Virginia:30,d_tp1012_Au
# stralia:61):19,(c_tp1015_Puget_Sound:207,b_tp1335_NY:41):18):1005,a_tp1014_Gyre:
# 61):3912):7054,outgroup:0);
```

NOTE: In early code, tree was not being recalculated; it was defined by constants in the following code chunk, hand-copied from the analysis above.

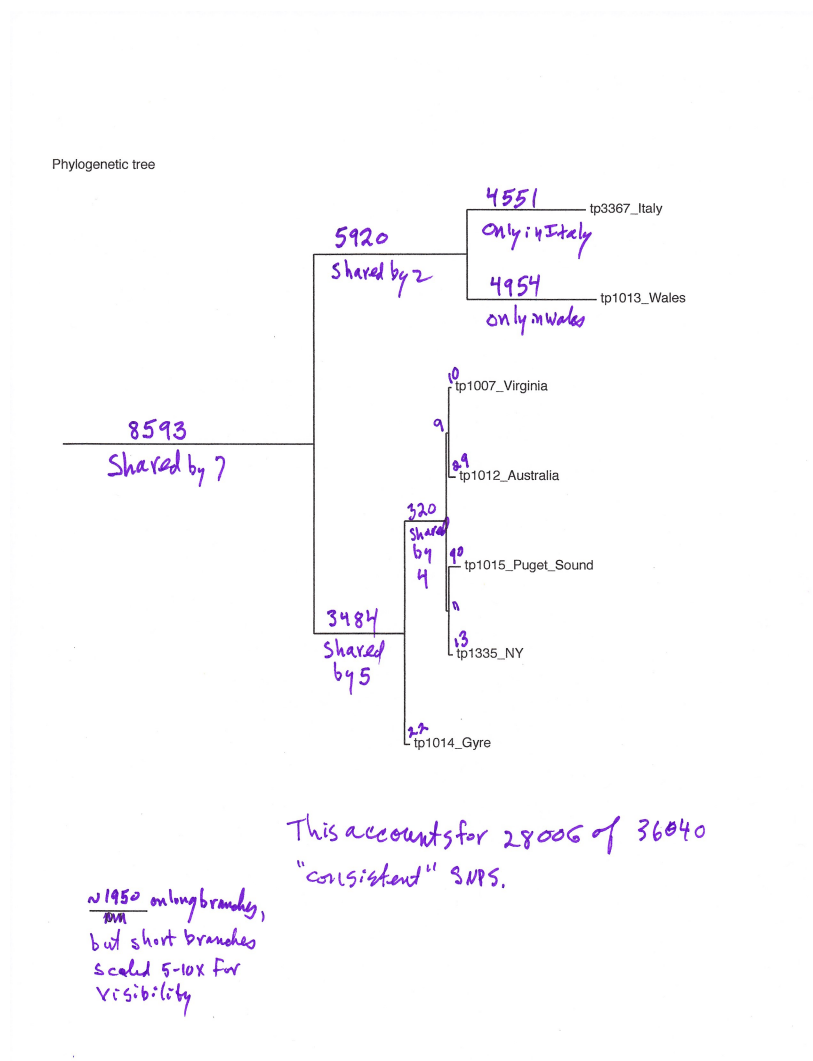


Figure 7: Inferred Tree, based on Chr1, un-q-filtered reads, loose criteria. (Note: to visually resolve the edges among the 5, their lengths were scaled by 5x – 10x in this figure, but not in the newick description shown in the text.)

```
# tree parameters as nested lists
# Internal nodes have subtrees sub1/2 and length
# Root has sub1/2, but no length
# Leaves have where, length, optionally, id, alt, nb. (Omit id for 'outgroup'. Use 'alt' for less formal
# labeling in cartoon version; it defaults to 'where'. Use 'nb' to add abcde annotations for legend.)
# This hand-made version is now subsumed by make.tree; retained for comparison
tree.by.hand <-
  list(
    sub1 = list(
      sub1 = list(
        sub1 = list(id=3367, length=8813, where='Venice, Italy', alt='Venice'),
        sub2 = list(id=1013, length=9652, where='Wales, UK'),
        length=9365),
      sub2 = list(
        sub1 = list(
          sub1 = list(
            sub1 = list(id=1007, length=30, nb='e', where='Virginia, USA'),
            sub2 = list(id=1012, length=61, nb='d', where='Perth, W. Australia', alt='Perth'),
            length=19),
          sub2 = list(
            sub1 = list(id=1015, length=207, nb='c', where='Washington, USA', alt='Puget Sound'),
            sub2 = list(id=1335, length=41, nb='b', where='New York, USA', alt='NY'),
            length=18),
          length=1005),
        sub2 = list(id=1014, length=61, nb='a', where='N. Pacific Gyre'),
        length=3912),
      length=7054),
    sub2 = list(length=0, where='outgroup')
  )

# historical, format example, and debug help:
oldwick.medium <- '(((CCMP3367_Italy:8813,CCMP1013_Wales:9652):9365,((e_CCMP1007_Virginia:30,d_CCMP1012_Australia:61):19,(c_CCMP
# with simpler labeling for cartoon
simple.oldwick.medium <- '(((Italy:8813,Wales:9652):9365,((Virginia:30,Australia:61):19,(Puget:207,NY:41):18):1005,Gyre:61):3912
cat.hardwrap(oldwick.medium)

# (((CCMP3367_Italy:8813,CCMP1013_Wales:9652):9365,((e_CCMP1007_Virginia:30,d_CCM
# P1012_Australia:61):19,(c_CCMP1015_Puget_Sound:207,b_CCMP1335_NY:41):18):1005,a_
# CCMP1014_NPG:61):3912):7054,outgroup:0);

cat.hardwrap(simple.oldwick.medium)

# (((Italy:8813,Wales:9652):9365,((Virginia:30,Australia:61):19,(Puget:207,NY:41)
# :18):1005,Gyre:61):3912):7054,outgroup:0);
```

Two other versions of the tree, for possible use in figs in the main paper.

Figure 8: `[[* as of 10/4/2015, this fig and next have stray stars on virginia, wales labels; probably due to hacking with commas in newick; not worth fixing unless we resurrect these trees for some purpose, but if so, see use of newick.name.undo in show.tree as probable fix. *]]`

```
tree.scale <- ifelse(which.snp.tables(string.val=F)[1]=='Chr1', 1, 10)
tree.x.lim <- 3e4 * tree.scale
the.simple.tree <- read.tree(text=simple.newick.medium)
plot(the.simple.tree, x.lim = tree.x.lim)
axis(1)
```

Figure 9:

```
plot(the.simple.tree, x.lim = tree.x.lim)
axis(1, (0:4)*7000*tree.scale, (0:4)*7000*tree.scale)
```

At some much earlier point, Tony ran the whole-genome version of the then-current code above, and manually entered tree branch lengths/legend for the resulting tree, shown in Fig 10. Code above can now automatically generate such a tree, but retain the following for comparison. The basic story seems clear—same topology and branch lengths scaled by about 10x, which is completely reasonable given that Chr1 is about 10% of the genome. Note that this tree is not being recalculated; it is defined by constants in the following code chunk.

```
fullgenome.newick.medium <- '(((3367_Italy:86155,1013_Wales:95697):89598,((e_1007_VA:330,d_1012_Australia:632):1296,(c_1015_WA:2
cat.hardwrap(fullgenome.newick.medium)

# (((3367_Italy:86155,1013_Wales:95697):89598,((e_1007_VA:330,d_1012_Australia:63
# 2):1296,(c_1015_WA:2113,b_1335_NY:658):480):10059,a_1014_NPG:568):39517):69526,o
# utgroup:0);
```

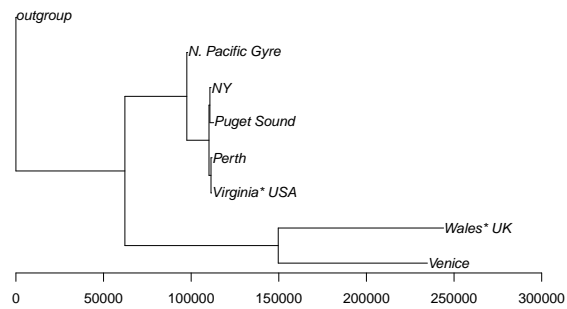


Figure 8: Tree based on qfiltered reads and medium SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs. (no edge labels, nolegend)

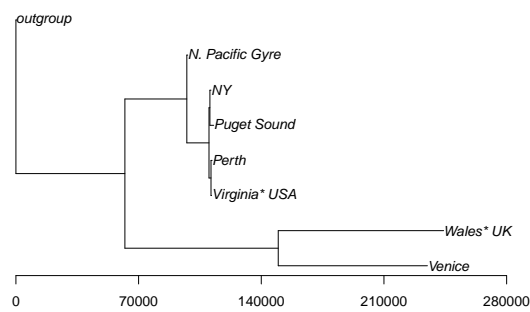


Figure 9: Tree based on qfiltered reads and medium SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs. (no edge labels, no legend, short scale bar)

```

legend.text <- c('a: only in 1014 ',
                'b: only in 1335 ',
                'c: only in 1015 ',
                'd: only in 1012 ',
                'e: only in 1007 ',
                '*: shared by bcde',
                '  shared by b/c ',
                '  shared by d/e ')
)
fullgenome.tree.x.lim <- 300000
fullgenome.counts <- c( 568, 658, 2113, 632, 330, 10059, 480, 1296 )
fullgenome.legend.text <- paste(legend.text, format(fullgenome.counts, width=5), sep=' - ')
fullgenome.tree.labels <- list( ## x,y,text
  41000, 3.63, '69526\nshared by 7',
  90000, 5.75, '39517\nby 5 (**) ',
  115000, 1.5, '89598\nshared by 2',
  210000, 2.0, '95697 only\nin Wales',
  210000, 1.0, '86155 only\nin Italy',
  113500, 4.6, '*')

```

Figure 10:

```

library(ape)
the.fullgenome.tree <- read.tree(text=fullgenome.newick.medium)
plot(the.fullgenome.tree, x.lim = fullgenome.tree.x.lim)
axis(1) # ; axis(2) useful only for placing labels
opar <- par(family='mono', cex=.8)
legend('topright', legend=fullgenome.legend.text)
par(opar)
for(i in seq(1, length(fullgenome.tree.labels)-2, by=3)){
  text(fullgenome.tree.labels[[i]], fullgenome.tree.labels[[i+1]], fullgenome.tree.labels[[i+2]])
}

```

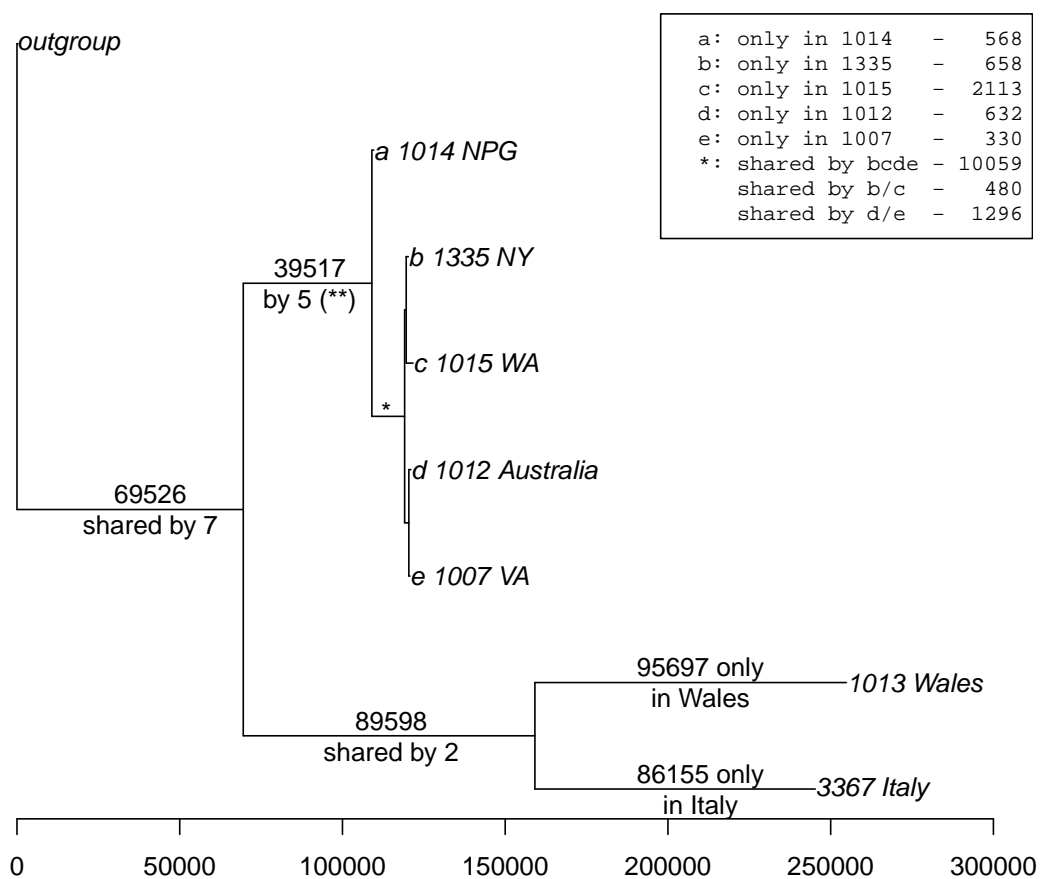



Figure 10: Tree based on unfiltered reads and medium SNP filters. “Lengths” are numbers of shared/private SNPs genome-wide. (By-hand legacy version)

