

10/29/2013

Instrument settings

Sheath fluid MQ

Nozzle 120 micron

1 micron beads (Invitrogen FluoSpheres). Pure stock was diluted 100 times in 1 ml MQ; we then added 100 ul of diluted stock in 1 ml of sample

Green channel in linear mode

Config file: Thaps_Nucleus3.cfg

Low Pressure - Flow rate and set to ~ 30 ul / min

Pure SYBRGreen stock was diluted 100 times in MQ; we then added 20 ul of diluted stock in 2 ml of sample

Cultures

RCTP (Italian Thaps)

1013 (Whales Thaps)

1335 (NY Thaps, reference)

The three cultures were grown either acclimated to Light:Dark cycle in the 13 degC incubator -> SYNCHRONIZED culture

Samples were collected around 2 pm so synchronized cells still start to divide, therefore some cells should be expected to be in G2 phase.

Based on the cell concentration of 10/24/13 experiment, we decided to dilute 3 times the cultures in the 20 degC incubator in F/2 medium (100 ul in 200 ul F/2 medium)

Experiment

Start at 2 pm

Volume analyzed for each sample (microliter):

0001.fcs - 36.17

0002.fcs - 76.30

0003.fcs - 80.84

0004.fcs - 48.66

0005.fcs - 37.98

0006.fcs - 56.54

Sample LOG

Plots STAINED SAMPLES

First column, cytograms showing the gate used to cluster the beads (log amplification). Mode of the Beads chlorophyll signal is indicated on the top side of the cytogram

Second column, cytograms showing the gate used to cluster THAPS (log amplification)

Third column, Green fluorescence of Thaps (linear amplification)

Fourth column, Green Fluorescence distribution in Thaps, with red line indicating the mode of the distribution. Value is indicated on the top side of the cytogram

