

Exploration of Shared SNPs in Thaps

trunc-qfiltered

July 26, 2017

Rambling exploration of SNP positions shared between two or more of the isolates. Code is included to document it thoroughly, (even if largely uninteresting to anyone else), and I will summarize it as I go.

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1 History

This was added to SVN 1/26/2014; not sure when it was started, but earliest related emails I see are from 1/21/14.

```
r413 | ruzzo | 2014-01-26 08:22:37 -0800 (Sun, 26 Jan 2014) | 2 lines
adding shared-snp analysis.
```

2 Preliminaries

NOTE: Some comments in code and some parts of the text, especially specific numbers and general conclusions, are based on Unqfiltered, Chr1, Medium stringency (i.e., “[2]” below) analysis. The broad picture does not appear to change with other choices, but details do, and the text is neither fully parameterized nor fully updated, so proceed with caution.

Load utility R code; do setup:

```
source('.././../R/wlr.R') # load util code; path relative this folder or sibling in scripts/larrys

## Running as: ruzzo @ bicycle.cs.washington.edu; SVN Id, I miss you. $Id: wlr.R 2017-07-21 or later $

setup.my.wd('shared-snps') # set working dir; UPDATE if this file moves, or if COPY/PASTE to new file
setup.my.knitr('figs-knitr/')
generic.setup('figs-mine/')
```

3 Major Analysis/Performance Parameters.

Choices here control how this file is processed, what data is analyzed, speed, etc. Set them carefully before running “make.” Major choices are:

1. WHICH SNP TABLES ARE LOADED??? The logical vector `load.tb` selects the desired combination of SNP tables to load, in the order `full.unfiltered`, `chr1.unfiltered`, `full.qfiltered`, `chr1.qfiltered`. E.g., `load.tb=(T, F, T, F)` loads *full* tables for *both* q- and un-qfiltered data. Primary analysis is only performed on one of them, but the others are retained for comparison/debugging.
2. WHICH MAIN ANALYSIS??? If multiple tables are loaded, which is used for the main analysis? Parameter `pri` is a permutation of 1:4, corresponding to `load.tb`; the first loaded table in that order becomes the analysis focus. The default `pri=c(1, 2, 3, 4)` looks at un-q-filtered data in preference to q-filtered, and full tables in preference to Chr1 within each group.
(Choice of data for the “Table 1” coverage summary in section 5 is independent of this; full genome data is preferred over Chr 1 for both q- and unq-filtered reads; change `tset.picker` calls near the end of that section to modify this.)
3. CLEAR CACHE??? `clear.cache=T` forces Knitr cache removal at the start of the run; especially important if the previous parameters have changed since the last run.
4. HOW MANY BOOTSTRAP REPLICATES??? The variable `nboot` is a major performance factor; 1000 reps takes several hours. Set to 5 for debug and quick look; 100 or more for final run.
5. TRUNCATE TABLES TO Chrs ONLY??? I.e., remove mitochondrial-, plastid-, and BD- contigs.

The following code chunk sets the first four parameters based on where it’s run. To prototype/debug on a laptop, faster is better—run on Chr1 with small `nboot`; when run on the linux servers, I typically do full genomes, more replicates. Just override them if these defaults don’t work for you.

```

# for Makefile, params can be command line args, else base on system; see wlr.r for details.
# load.tb order: full.un, chr1.un, full.qfil, chr1.qfil

params <- pick.params(
  mac = list(load.tb=c(F,T,F,F), pri=1:4, clear.cache=F, nboot= 1, trunc.tables=T), # quick on lap
  linux = list(load.tb=c(F,F,F,T), pri=1:4, clear.cache=F, nboot= 5, trunc.tables=T), # quick qfil on server
  linux = list(load.tb=c(T,F,T,F), pri=1:4, clear.cache=T, nboot=101, trunc.tables=T) # full on server
)

# Alternatively, edit/uncomment the following to override the above as needed
#params<-pick.params(default=list(load.tb=c(T,T,T,T),pri=1:4,clear.cache=T,nboot=1000,trunc.tables=T))
print(params)

# $load.tb
# full.unf chr1.unf full.qf chr1.qf
# TRUE FALSE TRUE FALSE
#
# $pri
# [1] 3 4 1 2
#
# $clear.cache
# [1] TRUE
#
# $nboot
# [1] 101
#
# $trunc.tables
# [1] TRUE

```

CLEAR CACHE??!! Some code chunks use the knitr cache, but extent of cache consistency checks unknown. If in doubt, delete “cache/” (knitr’s) directory to force rebuild. T/F set in params above will/won’t force removal (actually, rename):

```

decache(params$clear.cache)

# Rename of 'cache' to 'cache91396' returned TRUE .

```

If still in doubt, also manually remove “00common/mycache/” (mine).

Load the main SNP data file(s) based on the parameters set in section 3.

```

# short names to keep the following chunk compact
tb <- params$load.tb
tset <- list(NULL, NULL, NULL, NULL) # tset = 'table set'

# see wlr.R for load paths
if(tb[1]){tset[[1]] <- load.snp.tables(use.chr1.tables = FALSE, data.name='full.tables.01.26.14')}

# Loading full tables from ../../../../data/ungit-data/full.tables.01.26.14.rda ...Loaded.
# ../00common/mycache/snp.tables.chr1.unqfiltered.rda saved.

if(tb[2]){tset[[2]] <- load.snp.tables(use.chr1.tables = TRUE , data.name='full.tables.01.26.14')}
if(tb[3]){tset[[3]] <- load.snp.tables(use.chr1.tables = FALSE, data.name='full.tables.02.25.15')}

# Loading full tables from ../../../../data/ungit-data/full.tables.02.25.15.rda ...Loaded.
# ../00common/mycache/snp.tables.chr1.qfiltered.rda saved.
# Bandaiding qfiltered tables...

if(tb[4]){tset[[4]] <- load.snp.tables(use.chr1.tables = TRUE , data.name='full.tables.02.25.15')}

```

Grrr! I should have excluded non-Chr contigs from full genome runs. Rather than change tons of code below to add mask params, I’m just going to truncate the tables, as follows. (See notes in wlr.r::make.mask for assumptions.)

```

if(params$trunc.tables){
  for(i in 1:4){
    if(!is.null(tset[[i]])){
      first.mito <- match("mitochondria.fasta", tset[[i]][[7]]$Chr)
      if(!is.na(first.mito)){ # will be NA for Chr1 tables
        for(j in 1:7){
          # hmmm... slow; wonder whether head(tset[[i]][[j]],first.mito-1) is faster;
          # ok, simple tests suggest not: system.time(head(data.frame(1:1e7,1:1e7),5e6))
          tset[[i]][[j]] <- tset[[i]][[j]][1:(first.mito-1),]
        }
      }
    }
  }
} else {
  cat('***\n*** DID YOU *REALLY* WANT UNTRUNCATED TABLES???\n***\n')
}

```

The tersely-named `tset` list is sometimes convenient, but give them more descriptive names, too.

```

snp.tables.full.unfiltered <- tset[[1]]; names(tset)[1] <- 'snp.tables.full.unfiltered'
snp.tables.chr1.unfiltered <- tset[[2]]; names(tset)[2] <- 'snp.tables.chr1.unfiltered'
snp.tables.full.qfiltered <- tset[[3]]; names(tset)[3] <- 'snp.tables.full.qfiltered'
snp.tables.chr1.qfiltered <- tset[[4]]; names(tset)[4] <- 'snp.tables.chr1.qfiltered'

```

Main analysis may just use one of the potentially 4 table sets. Pick it according to the priority specified in section 3, using the shorter name 'snp.tables' for this default choice.

```
snp.tables <- tset.picker(priority=params$pri, table.set=tset)
```

```

# Sanity check: unlike unfiltered tables, bug in early code gave qfiltered ones different numbers
# of rows per strain, which breaks much code. Verify this is no longer happening.
check.eq.nrows <- function(tables){
  if(!is.null(tables)){
    nrow.snp.tables <- unlist(lapply(tables,nrow))
    print(nrow.snp.tables)
    if(all(nrow.snp.tables == nrow.snp.tables[1])){
      cat('OK, all strains have same number of rows.\n')
    } else {
      cat('***\n*** Warning: Different strains have different numbers of rows! ***\n***\n')
    }
  }
}

dummy<-lapply(tset, check.eq.nrows)

#      1007      1012      1013      1014      1015      3367      1335
# 31301782 31301782 31301782 31301782 31301782 31301782 31301782
# OK, all strains have same number of rows.
#      1007      1012      1013      1014      1015      3367      1335
# 31301782 31301782 31301782 31301782 31301782 31301782 31301782
# OK, all strains have same number of rows.

```

Which tables have we got?:

```

# 'which.snp.tables' return summary of which tables, either as a char string (default), e.g.
# "Chr1-qfiltered", or as vector of 2 strings, e.g. c("full","unfiltered").
cat('This analysis uses: (', paste(unlist(lapply(tset,which.snp.tables)),collapse=', '), ') SNP tables.\n')

# This analysis uses: ( trunc-unfiltered, NULL, trunc-qfiltered, NULL ) SNP tables.

cat('Main shared SNP analysis focuses on', which.snp.tables(snp.tables), '\n')

# Main shared SNP analysis focuses on trunc-qfiltered

```

A L^AT_EX hack: I want which.snp.tables info in doc title/page headers, but it is unknown until now, so the following writes a command definition `\whichsnptables` into the .aux file, which is read during the *next* L^AT_EX run, when `\begin{document}` is processed:

```
\makeatletter
\immediate\write\@auxout{\noexpand\gdef\noexpand\whichsnptables{trunc-qfiltered}}
\makeatother
```

Subsequent analysis was initially all directed at Chr1. In general, I have *not* updated the discussion to reflect genome-wide analysis.

```
if(exists('snp.tables.chr1.qfiltered') && exists('snp.tables.chr1.unqfiltered')){
  # If have both, where is new unequal to old?
  uneq <- snp.tables.chr1.qfiltered[[1]]$Ref[1:chr1.len] != snp.tables.chr1.unqfiltered[[1]]$Ref[1:chr1.len]
  cat('Sum uneq:', sum(uneq, na.rm=T), '\n')
  cat('Sum NA: ', sum(is.na(uneq)), '\n')
  print(which(is.na(uneq))[1:10])
  seecounts(which(is.na(uneq))[1:4], snp.tables=snp.tables.qfiltered, debug=F)
}
```

In brief, “snp.tables” will be a list of 7 data frames, one per strain, giving read counts for each nucleotide at each position, SNP calls, etc.:

```
names(snp.tables)

# [1] "1007" "1012" "1013" "1014" "1015" "3367" "1335"

str(snp.tables[[1]])

# 'data.frame': 31301782 obs. of 16 variables:
# $ snp : int 0 0 0 0 0 0 0 0 0 0 ...
# $ chr : chr "Chr1" "Chr1" "Chr1" "Chr1" ...
# $ Pos : int 1 2 3 4 5 6 7 8 9 10 ...
# $ Ref : chr "T" "C" "C" "A" ...
# $ Cov : num 0 2 3 4 4 4 7 8 9 10 ...
# $ a : num 0 0 0 0 0 0 0 0 0 0 ...
# $ g : num 0 0 0 0 0 0 0 0 0 0 ...
# $ c : num 0 0 0 0 0 0 0 0 0 0 ...
# $ t : num 0 0 0 0 0 0 0 0 0 0 ...
# $ n : num 0 0 0 0 0 0 0 0 0 0 ...
# $ .match: num 0 2 3 4 4 4 7 8 9 10 ...
# $ exon : logi FALSE FALSE FALSE FALSE FALSE ...
# $ indel : logi FALSE FALSE FALSE FALSE FALSE ...
# $ chr : Factor w/ 66 levels "BD1_7","BD10_65",...: 39 39 39 39 39 39 39 39 39 39 ...
# $ pos : int 1 2 3 4 5 6 7 8 9 10 ...
# $ rawCov: num 1 3 4 5 7 7 10 12 13 15 ...
```

Just for background, also load the desert tables:

```
# from svn+ssh://cegl.ocean.washington.edu/var/svn/7_strains/trunk/code/snpNB/data
#load('../..../data/ungit-data/des.rda')
load('../..../data/des.rda')
```

What’s the total length of all deserts in each strain? Big deserts (defined as “big.threshold” or longer)?

```
some.desert.stats <- function(big.threshold=0){
  desert.len <- unlist(lapply(des, function(x){sum(unlist(lapply(x, function(y){sum(y[, 'Length'])))})))})
  bigdes.len <- unlist(lapply(des, function(x){sum(unlist(lapply(x, function(y){
    sum(y[y[, 'Length']>=big.threshold, 'Length'])))})))})
  rbind(desert.len, desert.pct=round( desert.len / genome.length.constants()$genome.length.trunc * 100),
        bigdes.len, bigdes.pct=round( bigdes.len / genome.length.constants()$genome.length.trunc * 100))
}
some.desert.stats(big.threshold=50000)
```

#	tp1007	tp1012	tp1013	tp1014	tp1015	thapsIT	tp1335
# desert.len	11146526	11332566	5801763	9464213	11251426	6780300	10883723
# desert.pct	36	36	19	30	36	22	35
# bigdes.len	3495805	3936973	55365	3627235	3727061	57119	4046934
# bigdes.pct	11	13	0	12	12	0	13

I.e., looking at all deserts, about 1/3 of L-clade, 1/5 of H-clade are in deserts, whereas, looking at the largest deserts ($> 50k$), only about 12% in L-clade (and none in H-clade). Note that the rough stats above include artifactual “deserts” created by gaps in the reference sequence, large genomic deletions, etc. A more careful analysis of this is found in nc-snps.rnw.

4 Refined SNP Calls

4.1 Method

It is appropriate that SNP calls should be conservative, to avoid many false positives, but, when a position is called a SNP in one isolate, we often see a significant number of reads for the same non-reference nucleotide at that position in other isolates, even if they are not called as SNPs. On the other hand, we sometimes see a position called a SNP in two or more isolates, but with *different* pairs of nucleotides, potentially suggesting technical errors. Analysis in this section attempts to refine the SNP calls by looking for issues such as these by looking at all 7 isolates jointly, at each position called a SNP in any of them.

For a given strain, the following function returns a vector of 0:4 to indicate which nonreference nucleotide has the maximum read count at the corresponding position. The values 1..4 indicate that the max count occurred at A, G, C, T, resp. (Ties are resolved arbitrarily ($a < g < c < t$), which possibly deserves further attention.) The value 0 means all nonreference counts are below threshold, based *either* on absolute count *or* as a fraction of coverage. Default only excludes 0 counts.

```
nref.nuc.new <- function(strain=1, mask=T, thresh.count=0, thresh.rate=0.0){
  # get read count for max nonref nuc
  nref <- apply(snp.tables[[strain]][mask, c('a', 'g', 'c', 't')], 1, max)
  # where does nref count match a (g,c,t, resp) count
  as <- ifelse(nref == snp.tables[[strain]][mask, 'a'], 1, 0)
  gs <- ifelse(nref == snp.tables[[strain]][mask, 'g'], 2, 0)
  cs <- ifelse(nref == snp.tables[[strain]][mask, 'c'], 3, 0)
  ts <- ifelse(nref == snp.tables[[strain]][mask, 't'], 4, 0)
  # most positions will show 3 zeros and one of 1:4, so max identifies max nonref count;
  # ties broken arbitrarily (a<g<c<t)
  merge <- pmax(as, gs, cs, ts)
  # but if max nonref count is zero or below threshold, return 0
  merge[nref == 0 | nref < thresh.count] <- 0
  merge[nref/snp.tables[[strain]][mask, 'Cov'] < thresh.rate] <- 0
  return(merge)
}
```

Get union and intersection of the sets of called SNPs. (“\$snp” is 0/1.) Also, 5-way (L-clade) and 4-way (L-excluding Gyre).

```
# 4-way union/intersection
u4.snps <- snp.tables[[1]]$snp
i4.snps <- snp.tables[[1]]$snp
for(i in c(2,5,7)) {
  u4.snps <- pmax(u4.snps, snp.tables[[i]]$snp)
  i4.snps <- pmin(i4.snps, snp.tables[[i]]$snp)
}
# 5-way: add gyre
u5.snps <- pmax(u4.snps, snp.tables[[4]]$snp)
i5.snps <- pmin(i4.snps, snp.tables[[4]]$snp)
# 7-way
union.snps <- pmax(u5.snps, snp.tables[[3]]$snp, snp.tables[[6]]$snp)
intersect.snps <- pmin(i5.snps, snp.tables[[3]]$snp, snp.tables[[6]]$snp)
nu4snps <- sum(u4.snps)
```

```

nu5snps <- sum(u5.snps)
ni4snps <- sum(i4.snps)
ni5snps <- sum(i5.snps)
nusnps <- sum(union.snps)
nisnps <- sum(intersect.snps)
c(n4u=nu4snps, n5u=nu5snps, n7u=nusnps, n4i=ni4snps, n5i=ni5snps, n7i=nisnps)

#      n4u      n5u      n7u      n4i      n5i      n7i
# 196296 197799 474613 128683  70687  15186

```

There are nusnps=474613 positions called as SNPs in one or more strains (but only nisnps=15186 that are shared among all 7). Note that the 4-way union is only modestly larger (1.5254229 times larger) than the 4-way intersection, emphasizing the inherent similarities among these SNP sets. The corresponding 5-way numbers show that Gyre adds relatively little to the 5-way union vs the 4-way union, whereas it removes a fair bit from the 5-way intersection. However, much of that loss is simply because Gyre has fewer called SNPs: only 89184 vs 128683 in the 4-way intersection, and they are highly concordant:

```

sum(snp.tables[[4]]$snp*i4.snps)/sum(snp.tables[[4]]$snp)

# [1] 0.7925973

```

So, a likely source of the Gyre's difference in called SNPs is technical (lower read coverage, higher read error rate) rather than biological.

Inclusion of the 2 H-clade members, however, causes more dramatic changes in both union and intersection numbers. I examine all these relationships in more detail below, but first I examine what I believe to be a significant source of technical error in these comparisons—erroneous SNP calls, especially false negative calls.

It is appropriate that SNP calls should be conservative, to avoid many false positives, but, if a position is called a SNP in one strain, we often see a significant number of reads for the same non-reference nucleotide at that position in other strains, even if they are not called as SNPs. For my purposes below, these will be considered “shared SNPs,” based on three different levels of permissiveness. Note that, e.g., $\geq 84\%$ of all positions have zero reads for any non-reference nucleotide, and only a small fraction have 2 or more non-reference reads:

```

nonmatch <- rbind(
  unlist(lapply(snp.tables,function(x){sum(x$Cov-x$.match == 0)})),
  unlist(lapply(snp.tables,function(x){sum(x$Cov-x$.match == 1)})),
  unlist(lapply(snp.tables,function(x){sum(x$Cov-x$.match == 2)})),
  unlist(lapply(snp.tables,function(x){sum(x$Cov-x$.match == 3)})),
  unlist(lapply(snp.tables,function(x){sum(x$Cov-x$.match >= 4)})),
  unlist(lapply(snp.tables,function(x){sum((x$Cov-x$.match)[union.snps==0] >= 4)}))
)/nrow(snp.tables[[1]])*100
rownames(nonmatch) <- c('% ==0', '% ==1', '% ==2', '% ==3', '% >=4', '% >=4, nonSNP')
nonmatch

#           1007           1012           1013           1014           1015           3367           1335
# % ==0      97.71716831  97.35791400  95.45329400  97.29079003  97.18569697  95.89943474  96.48734376
# % ==1       1.48140448   1.75279158   3.01610304   2.08080805   1.88427930   2.58814338   2.54747477
# % ==2       0.11277633   0.12101867   0.22209918   0.18711714   0.13599226   0.21703876   0.19842640
# % ==3       0.05885927   0.05629072   0.09496264   0.10621121   0.06021063   0.09526295   0.06375675
# % >=4       0.62979162   0.71198502   1.21354113   0.33507358   0.73382084   1.20012017   0.70299831
# % >=4, nonSNP 0.08015518  0.12493857  0.25313575  0.04763946  0.13208513  0.28212771  0.13240460

```

Build a table of max non-reference nucleotides at each position in the union.snps set. The three criteria are

- [[1]]: any non-zero count at any coverage is considered significant
- [[2]]: (count ≥ 2 and count/coverage ≥ 0.05) is considered significant
- [[3]]: (count ≥ 4 and count/coverage ≥ 0.10) is considered significant

In all three cases, the nonref nucleotide must also be consistent across all strains passing that threshold; see below.

```

non.refs <- vector('list',4)
for(i in 1:4){
  non.refs[[i]] <- matrix(0, nrow=nusnps, ncol=7)
  colnames(non.refs[[i]]) <- names(snp.tables)
  rownames(non.refs[[i]]) <-
    paste(snp.tables[[1]]$chr[union.snps==1], ':', snp.tables[[1]]$pos[union.snps==1], sep='')
}
for(j in 1:7){
  non.refs[[1]][,j] <- nref.nuc.new(j, mask=union.snps==1, thresh.count=0, thresh.rate=0.00)
  non.refs[[2]][,j] <- nref.nuc.new(j, mask=union.snps==1, thresh.count=2, thresh.rate=0.05)
  non.refs[[3]][,j] <- nref.nuc.new(j, mask=union.snps==1, thresh.count=4, thresh.rate=0.10)
}

```

For comparison, I want to look at unfiltered SAMTools SNP calls. In complete opposition to the measures of consistency imposed above, I'm going to simply force this into the “non.refs” structure constructed above by imagining that any position called a SNP in any strain has its max nonref count on “A”, so any given position called a SNP in any strain will automatically be declared “consistent.” This will allow the tree-code, etc. given below to work in a uniform way (even though interpretation of the results is different.) Results will be jammed into a 4th component of the “non.refs” list; i.e., we have a 4th criterion:

- [[4]]: all called SNPs at a given position are considered “consistent.”

As this case was a late addition to the analysis, the commentary throughout this document has not necessarily been updated to reflect that this case is distinct from the first three.

```

for(j in 1:7){
  non.refs[[4]][,j] <- snp.tables[[j]]$snp[union.snps==1]
}

```

```

str(non.refs[[4]])

# num [1:474613, 1:7] 0 0 0 0 0 0 0 1 0 ...
# - attr(*, "dimnames")=List of 2
# ..$ : chr [1:474613] "Chr1:333" "Chr1:417" "Chr1:435" "Chr1:438" ...
# ..$ : chr [1:7] "1007" "1012" "1013" "1014" ...

```

“non.refs” indicates, among those positions in the union of all called SNPS having any non-reference read count above the thresholds listed above, the non-ref nucleotide having the highest read count in each strain. If, for a given position, the max of this code is the same as the min (among non-zero values), then every strain having over-threshold nonref reads in that position, in fact has most non-reference reads on the *same* nucleotide. These are defined as the “consistent” SNPs.

```

find.consistent <- function(nr){
  nr.max <- apply(nr,1,max)
  nr.min <- apply(nr,1,function(x){ifelse(max(x)==0,0,min(x[x>0]))})
  return(nr.min == nr.max)
}
consistent <- lapply(non.refs, find.consistent)

```

4.2 Save them

```

# wrap this in a data structure to be cached:
Description <- [2753 chars quoted with '']

refined.snps <-
  list(Description=Description,

        Data=list(
          based.on.which.snp.tables=which.snp.tables(),
          number.union.snps=nusnps,
          number.intersection.snps=nisnps,

```



```

non.ref.nucleotide=non.refs,
consistent.snps=consistent),

Code=list(
  get.snps = function(strain, stringency=2){
    # return nusnps x 1 Bool vector of consistent SNPs @ specified stringency & strain
    return(refined.snps$Data$consistent.snps[[stringency]] &
           refined.snps$Data$non.ref.nucleotide[[stringency]][,strain] > 0)
  },
  get.snp.locs.char = function(strain, stringency=2){
    # return char vector of locations of consistent SNPs @ specified stringency & strain
    snps <- refined.snps$Code$get.snp(locs.char(strain, stringency)
    return(names(snps)[snps])
  },
  get.snp.locs.df = function(strain, stringency=2){
    # return data frame (Chr/Pos) of locations of consistent SNPs @ specified stringency & strain
    snplist <- strsplit(refined.snps$Code$get.snp(locs.char(strain, stringency), ':', fixed=TRUE)
    # strsplit returns long list of 2-vectors, 1st=chr, 2nd=char position
    df <- data.frame(Chr=unlist(lapply(snplist,function(x){return(x[1])})),
                     Pos=as.integer(unlist(lapply(snplist,function(x){return(x[2])}))),
                     stringsAsFactors = FALSE)

    return(df)
  }
)

# dont't clobber existing .rda, but save if absent. (delete to re-save)
# result for trunc, unfiltered tables saved to "data" else "mycache"
if(which.snp.tables() == 'trunc-unfiltered'){
  rda.refined <- '.././../data/refined.snps-trunc-unfiltered.rda'
} else {
  rda.refined <- paste('../00common/mycache/refined.snps', which.snp.tables(), 'rda', sep='.')
}
if(file.exists(rda.refined)){
  cat('Pre-existing file', rda.refined, 'unchanged.\n')
} else {
  cat('Saving', rda.refined, '...\n')
  save(refined.snps, file=rda.refined, compress=TRUE)
  cat('Saved.\n')
}

# Saving ../00common/mycache/refined.snps.trunc-qfiltered.rda ...Saved.

```

Knitr seems to be failing to format the long char string above, which says:

```

cat(refined.snps$Description)

# Contents of this .rda file:
#
# * Description: this text
#
# * Data -- 5 items defining refined SNPs, at 4 different stringency levels, as defined
#   in shared-snps.rnw:
#
# * based.on.which.snp.tables: {"Chr1","full","trunc"}-{"unfiltered","qfiltered"},
#   depending on which snp tables were used to build this data. ("trunc" = all Chrs.)
#
# * number.union.snps: the total number of SNPs (SAMtools calls) in the union of SNPs
#   across all 7 strains.
#
# * number.intersection.snps: similar, for the 7-way intersection.
#
# nusnps/nisnps are easily recalculated from the data below, but their inclusion
# may be convenient, e.g., to quickly see if the .rda represents the full genome
# (nusnps=488848), or the chr 1 subset (nusnps=47499); (redundant with "based.on...";

```

```
#      numbers above are for unfiltered, perhaps slightly different if qfiltered)
#
#      * non.ref.nucleotide: 4 arrays, each nusnps x 7, of values 0..4 (0..1 in the 4th
#      array). In the 1st 3 arrays, 0 means the given position in the given strain did
#      not have nonreference read counts above the corresponding filtering threshold,
#      i.e., is NOT a refined SNP in that strain, whereas 1..4 mean that it did pass
#      threshold, for A,C,G,T resp. In the 4th array, this value is just 1/0,
#      indicating is/is not a called SNP in that strain.
#
#      * consistent.snps: 4 Bool vectors of length nusnps flagging positions whose nonref
#      nucs (wrt to the 4 filtering criteria) are deemed *consistent* across
#      all 7 strains. For the 1st 3, this means all nonzero entries of non.ref.nuc
#      are equal, i.e., nonref read counts passing threshold are on the SAME nonref
#      nucleotide in all strains having over-threshold counts. Just for comparison
#      and uniformity of data structures, the 4th is all TRUE, i.e., union of SNPs
#      across all strains, without any regard for thresholds or consistency.
#
#      In short, the refined SNPs according to our medium filtering criteria are
#      strains/positions where consistent.snps[[2]]==TRUE and non.ref.nucleotide[[2]]>0.
#
#      Rownames in both non.ref.nucs and consistent define location, e.g. "Chr1:333".
#
#      * Code -- simple routines to extract refined SNPs in (potentially) convenient formats:
#
#      * get.snps(strain, stringency=2)
#      returns nusnps x 1 Bool vector of consistent SNPs @ specified stringency in
#      given strain
#
#      * get.snp.locs.char(strain, stringency=2)
#      returns n x 1 char vector of locations of consistent SNPs @ specified stringency
#      in given strain, e.g. "Chr1:1234", where n == sum(get.snps(...))
#
#      * get.snp.locs.df(strain, stringency=2){
#      As above, but returns data frame (char vector Chr, int vector Pos) with the same info.
```

```
str(consistent[[1]])

# Named logi [1:474613] TRUE FALSE TRUE TRUE TRUE TRUE ...
# - attr(*, "names")= chr [1:474613] "Chr1:333" "Chr1:417" "Chr1:435" "Chr1:438" ...
```

```
consistent.count <- unlist(lapply(consistent, sum)) ; consistent.count

# [1] 447177 469906 471171 474613

inconsistent.count <- consistent.count[4] - consistent.count; inconsistent.count

# [1] 27436 4707 3442 0

inconsistent.percent <- inconsistent.count/consistent.count[4]*100; inconsistent.percent

# [1] 5.7807098 0.9917554 0.7252224 0.0000000
```

I.e., of the 474613 positions in which a SNP is called, 447177 are consistent by my loose definition, and 471171 are consistent by my tightest definition. The increase in concordance supports the view that the loose definition is too loose. Perhaps misleadingly, these counts include positions that are “consistent SNPs” in only one strain; more below. (*TODO* I suspect, but have not yet systematically checked, that most of the rest are positions with low coverage and/or very low read counts on the mixture of non-reference nucleotides.)

4.3 Examples: Consistent

Here are a few (nonrandomly selected) prototypical consistent SNPs:

	chr	pos	Ref	Strain	A	G	C	T	SNP	exon	indel	nrf	rat
# 1	Chr1	567	T										
# 2				1007	0	0	1	25	0	TRUE	FALSE		
# 3				1012	0	0	14	39	1	TRUE	FALSE		
# 4				1013	0	0	13	87	0	TRUE	FALSE		
# 5				1014	0	1	0	23	0	TRUE	FALSE		
# 6				1015	0	0	8	40	1	TRUE	FALSE		
# 7				3367	0	0	16	38	1	TRUE	FALSE		
# 8				1335	0	0	2	99	0	TRUE	FALSE		
# 9	Chr1	1053	A										
# 10				1007	25	0	0	4	0	TRUE	FALSE		
# 11				1012	35	0	0	12	0	TRUE	FALSE		
# 12				1013	2	1	0	32	0	TRUE	FALSE		
# 13				1014	5	0	0	5	0	TRUE	FALSE		
# 14				1015	29	0	0	15	1	TRUE	FALSE		
# 15				3367	2	0	0	7	0	TRUE	FALSE		
# 16				1335	56	0	0	39	1	TRUE	FALSE		
# 17	Chr1	1055	G										
# 18				1007	0	23	0	1	0	TRUE	FALSE		
# 19				1012	0	37	0	6	0	TRUE	FALSE		
# 20				1013	1	39	0	6	0	TRUE	FALSE		
# 21				1014	0	6	0	2	1	TRUE	FALSE		
# 22				1015	0	26	0	14	0	TRUE	FALSE		
# 23				3367	0	12	0	0	0	TRUE	FALSE		
# 24				1335	0	54	0	32	1	TRUE	FALSE		
# 25	Chr1	1176	G										
# 26				1007	1	53	0	0	0	FALSE	FALSE		
# 27				1012	0	54	0	0	0	FALSE	FALSE		
# 28				1013	19	56	0	0	0	FALSE	FALSE		
# 29				1014	0	28	0	0	0	FALSE	FALSE		
# 30				1015	3	85	0	0	0	FALSE	FALSE		
# 31				3367	9	2	0	0	1	FALSE	FALSE		
# 32				1335	0	156	0	0	0	FALSE	FALSE		
# 33	Chr1	8685	G										
# 34				1007	6	15	0	0	0	TRUE	FALSE		
# 35				1012	10	23	0	0	0	TRUE	FALSE		
# 36				1013	18	21	0	0	1	TRUE	FALSE		
# 37				1014	4	8	0	0	0	TRUE	FALSE		
# 38				1015	10	24	0	0	1	TRUE	FALSE		
# 39				3367	0	4	0	0	0	TRUE	FALSE		
# 40				1335	5	32	0	0	0	TRUE	FALSE		

Here is a brief look at some *in*-consistent positions. E.g., Chr1:2013 shows nontrivial counts on 3 alleles in Wales, as do 2319, 3286, 5002, 5433, whereas 7878 shows a different alternate allele in Italy than in Wales.

[illegible]

# 10				1007	0	28	10	0	1	TRUE	FALSE
# 11				1012	0	43	17	0	1	TRUE	FALSE
# 12				1013	13	15	9	0	1	TRUE	FALSE
# 13				1014	0	18	6	0	1	TRUE	FALSE
# 14				1015	0	53	20	0	1	TRUE	FALSE
# 15				3367	4	0	24	0	0	TRUE	FALSE
# 16				1335	0	118	28	0	1	TRUE	FALSE
# 17	Chr1	3286	T								
# 18				1007	4	0	1	10	0	TRUE	FALSE
# 19				1012	7	0	3	32	0	TRUE	FALSE
# 20				1013	34	0	30	1	1	TRUE	FALSE
# 21				1014	4	0	4	10	0	TRUE	FALSE
# 22				1015	11	0	6	31	0	TRUE	FALSE
# 23				3367	5	0	29	0	0	TRUE	FALSE
# 24				1335	14	0	3	55	0	TRUE	FALSE
# 25	Chr1	5002	T								
# 26				1007	0	14	0	7	0	TRUE	FALSE
# 27				1012	0	20	0	19	1	TRUE	FALSE
# 28				1013	18	10	0	22	0	TRUE	FALSE
# 29				1014	0	5	0	2	0	TRUE	FALSE
# 30				1015	0	18	0	12	1	TRUE	FALSE
# 31				3367	0	0	0	31	0	TRUE	FALSE
# 32				1335	0	46	0	44	0	TRUE	FALSE
# 33	Chr1	5178	G								
# 34				1007	0	20	0	0	0	TRUE	FALSE
# 35				1012	0	32	0	0	0	TRUE	FALSE
# 36				1013	47	9	0	0	1	TRUE	FALSE
# 37				1014	0	13	0	0	0	TRUE	FALSE
# 38				1015	0	30	0	0	0	TRUE	FALSE
# 39				3367	32	19	0	0	1	TRUE	FALSE
# 40				1335	0	38	0	2	0	TRUE	FALSE
# 41	Chr1	5433	G								
# 42				1007	0	40	0	3	0	TRUE	FALSE
# 43				1012	0	53	0	5	0	TRUE	FALSE
# 44				1013	16	29	0	7	1	TRUE	FALSE
# 45				1014	9	8	0	0	1	TRUE	FALSE
# 46				1015	6	53	0	2	0	TRUE	FALSE
# 47				3367	8	37	0	0	0	TRUE	FALSE
# 48				1335	6	72	0	2	0	TRUE	FALSE
# 49	Chr1	7858	C								
# 50				1007	0	0	42	0	0	TRUE	FALSE
# 51				1012	0	0	35	0	0	TRUE	FALSE
# 52				1013	0	0	81	8	0	TRUE	FALSE
# 53				1014	0	0	12	0	0	TRUE	FALSE
# 54				1015	0	0	71	0	0	TRUE	FALSE
# 55				3367	20	0	2	0	1	TRUE	FALSE
# 56				1335	0	0	83	0	0	TRUE	FALSE
# 57	Chr1	8974	C								
# 58				1007	0	1	5	0	0	TRUE	FALSE
# 59				1012	0	2	13	0	0	TRUE	FALSE
# 60				1013	9	15	2	0	1	TRUE	FALSE
# 61				1014	0	1	1	0	0	TRUE	FALSE
# 62				1015	0	1	9	0	0	TRUE	FALSE
# 63				3367	2	0	1	0	0	TRUE	FALSE
# 64				1335	0	11	30	0	0	TRUE	FALSE
# 65	Chr1	10099	T								
# 66				1007	16	0	0	24	0	TRUE	FALSE
# 67				1012	45	0	0	26	0	TRUE	FALSE
# 68				1013	0	2	6	55	0	TRUE	FALSE
# 69				1014	32	0	0	11	0	TRUE	FALSE
# 70				1015	38	0	0	37	0	TRUE	FALSE
# 71				3367	0	1	0	7	0	TRUE	FALSE
# 72				1335	52	0	0	61	1	TRUE	FALSE
# 73	Chr1	15154	A								
# 74				1007	13	0	0	0	0	FALSE	FALSE
# 75				1012	37	0	0	1	0	FALSE	FALSE
# 76				1013	2	0	35	7	1	FALSE	FALSE

```
# 77      1014 10    0 0 0    0 FALSE FALSE
# 78      1015 24    0 0 0    0 FALSE FALSE
# 79      3367  3    0 0 12   1 FALSE FALSE
# 80      1335 47    0 0 3    0 FALSE FALSE
```

4.5 Examples: Homozygous nonref

And at some *homozygous nonreference* positions (defined to be those with nonref fraction > 0.75):

```
hnr <- lapply(snp.tables, function(x){x$.match/x$.Cov < 0.25}) # find them
hnr <- lapply(hnr, function(x){ifelse(is.na(x), FALSE, x)})      # remove NA
unlist(lapply(hnr, sum))                                       # count per strain

#   1007   1012   1013   1014   1015   3367   1335
# 16069 14356 120037 11436   6862 142515   1854
```

Hmm, in L-clade, excluding the ref isolate (1335) this tracks time-in culture to some degree; Maybe many of these are in hemizygous regions. Next two chunks lifted from nc-snp to get tables for hemi-deletion.

```
cnv.chrononly <- load.cnv.tables('.../.../data/cnv.txt', chrs.only=TRUE)

str(cnv.chrononly)

# 'data.frame': 1956 obs. of 11 variables:
# $ strain : Factor w/ 7 levels "IT","tp1007",...: 3 3 3 3 3 3 3 3 3 3 ...
# $ chr : Factor w/ 65 levels "BD1_7","BD10_65",...: 38 38 38 38 38 38 38 38 38 38 ...
# $ start : int 10601 112001 215001 358901 536501 554801 673401 781801 806901 853201 ...
# $ end : int 13500 116500 221100 370300 538600 559300 685000 787400 811100 855600 ...
# $ length : int 2900 4500 6100 11400 2100 4500 11600 5600 4200 2400 ...
# $ filtered : logi FALSE FALSE FALSE TRUE FALSE FALSE ...
# $ type : Factor w/ 1 level "CNVnator": 1 1 1 1 1 1 1 1 1 1 ...
# $ cov_ratio: num 0.63738 1.54893 1.65381 0.00204 0.68486 ...
# $ dup_frac : num 0.41188 0.00908 0.01178 0.97997 0.0211 ...
# $ iStart : num 10601 112001 215001 358901 536501 ...
# $ iEnd : num 13500 116500 221100 370300 538600 ...

cnv.chrononly[c(1:4,nrow(cnv.chrononly)+c(-1,0)),] ## first/last few rows

# strain chr start end length filtered type cov_ratio dup_frac iStart iEnd
# 1 tp1012 Chr1 10601 13500 2900 FALSE CNVnator 0.63738000 0.41187900 10601 13500
# 2 tp1012 Chr1 112001 116500 4500 FALSE CNVnator 1.54893000 0.00907677 112001 116500
# 3 tp1012 Chr1 215001 221100 6100 FALSE CNVnator 1.65381000 0.01178470 215001 221100
# 4 tp1012 Chr1 358901 370300 11400 TRUE CNVnator 0.00204431 0.97997300 358901 370300
# 1955 tp1335 Chr24 259901 278000 18100 FALSE CNVnator 1.41458000 0.38091100 31264334 31282433
# 1956 tp1335 Chr24 286901 289800 2900 FALSE CNVnator 1.74941000 0.74228100 31291334 31294233
```

```
get.cnv.dels <- function(cov.thresh.lo = 0.0,
                        cov.thresh.hi = 0.8,
                        cnv,
                        snp.tables = NULL,
                        DEBUG = FALSE)
){
  # build list of 7 Bool vectors of genome length, with i-th == T iff
  # * i-th pos is 'NA' in genome seq (if snp.tables are provided), or
  # * in CNVnator call for coverage in half-open [cov.thresh.lo, hi), and
  # * not marked 'filtered' by CNVnator
  cnv.deletions <- vector(mode='list',7) # make list of bool vectors
  if(is.null(snp.tables)){
    # if no tables, assume full
    t.len <- genome.length.constants()$genome.length.trunc
  } else {
    t.len <- nrow(snp.tables[[1]])
  }
}
```

```

for(st in 1:7){
  if(is.null(snp.tables)){
    cnv.deletions[[st]] <- logical(t.len) # all F
  } else {
    cnv.deletions[[st]] <- is.na(snp.tables[[st]]$Pos[1:t.len]) # NA positions in genome
  }
}
strain.names <- c(paste('tp10',c('07',12:15),sep=''),'IT','tp1335')
names(cnv.deletions) <- strain.names
for(i in 1:nrow(cnv)){
  if(!cnv$filtered[i] &&
    cnv$cov_ratio[i] >= cov.thresh.lo &&
    cnv$cov_ratio[i] < cov.thresh.hi)
  {
    if(DEBUG){
      print(cnv[i,])
      print(as.character(cnv$strain[i]))
    }
    # following ASSUMES no CNVnator call crosses a chromosome bdry, & that
    # t.len ends at chr end (typically chr1 or chr24)
    if(cnv$iEnd[i] <= t.len){
      cnv.deletions[[as.character(cnv$strain[i])]][cnv$iStart[i]:cnv$iEnd[i]] <- TRUE
    }
  }
}
return(cnv.deletions)
}

# sanity check:
cnv.dels.38 <- get.cnv.dels(0.3, 0.8, cnv.chronly, snp.tables = NULL)
unlist(lapply(cnv.dels.38,sum)) # does it match low.length.38 in tic ?

# tp1007 tp1012 tp1013 tp1014 tp1015 IT tp1335
# 1672500 1781500 1383600 1313700 988400 320900 1453000

# 1672500 1781500 1399400 1313700 988400 336500 1453000 <== low.length.38 from tic (circa page 8)
# 1672500 1781500 1399400 1313700 988400 336500 1453000 <== low.length.38 from tic (pg9, 6/28/17)
rm(cnv.dels.38)

```

Slight discrepancy in H-clade that I should hunt down, but basically OK. (hmm; maybe untrunc tbls.)

```

# the ones we want for the current analysis:
hemi.masks <- get.cnv.dels(0.3, 0.8, cnv.chronly, snp.tables=snp.tables)

rbind(
  homnr = unlist(lapply(hnr,sum)),
  hemi = unlist(lapply(hemi.masks, sum)),
  homnr.unhemi = unlist(lapply(list(1,2,3,4,5,6,7), function(i){sum(hnr[[i]] & !hemi.masks[[i]]))})
)

#           1007    1012    1013    1014    1015    3367    1335
# homnr      16069   14356  120037  11436   6862  142515   1854
# hemi      1834990 1940024 1527725 1472095 1134652 480817 1596965
# homnr.unhemi 9650    7347  111674  10091   5113  140185   1829

# based on the thought that hnr in 1335 may reflect errors in the ref seq,
# are they shared with others?
unlist(lapply(hnr, function(x){sum(x & hnr[[7]])})) # hnr shared with 1335

# 1007 1012 1013 1014 1015 3367 1335
# 517 592 748 362 617 793 1854

# answer: around 300 in each strain, of 558 in NY, genomewide,
# so that seems like a plausibly important factor.

hnr.lclade <- hnr[[1]] | hnr[[2]] | hnr[[4]] | hnr[[5]] | hnr[[7]] # union over L-clade
sum(hnr.lclade) # count all in L-clade

```

```

# [1] 31723

sum(hnr[[3]] | hnr[[6]]) # present in H-clade

# [1] 188637

sum(hnr[[3]] & hnr[[6]]) # shared in H-clade

# [1] 73915

# look at a few in L-clade
w.hnr.l <- which(hnr.lclade)
seecounts(w.hnr.l[1:10], snp.tables=snp.tables)

#   chr  pos Ref Strain  A  G  C  T SNP  exon indel nrf rat
# 1  Chr1 1559  A
# 2      1007 7  0  0 24  0 TRUE FALSE
# 3      1012 11 0  0 37  0 TRUE FALSE
# 4      1013 9  0  0 5  0 TRUE FALSE
# 5      1014 4  0  0 16  0 TRUE FALSE
# 6      1015 47 0  0 35  0 TRUE FALSE
# 7      3367 0  0  0 0  0 TRUE FALSE
# 8      1335 60 0  0 50  0 TRUE FALSE
# 9  Chr1 1575  G
# 10     1007 24 7  0 0  0 TRUE FALSE
# 11     1012 42 13 0 0  0 TRUE FALSE
# 12     1013 17 16 0 0  0 TRUE FALSE
# 13     1014 15 4  0 0  0 TRUE FALSE
# 14     1015 43 31 0 0  1 TRUE FALSE
# 15     3367 0  2  0 0  0 TRUE FALSE
# 16     1335 34 74 0 0  0 TRUE FALSE
# 17 Chr1 1893  C
# 18     1007 0  0 14 32  0 TRUE FALSE
# 19     1012 0  0 38 52  0 TRUE FALSE
# 20     1013 0  0 95 14  0 TRUE FALSE
# 21     1014 0  0 5 31  0 TRUE FALSE
# 22     1015 0  0 47 44  0 TRUE FALSE
# 23     3367 0  0 29 0  0 TRUE FALSE
# 24     1335 0  0 68 85  0 TRUE FALSE
# 25 Chr1 2223  A
# 26     1007 25 13  0 0  0 TRUE FALSE
# 27     1012 13 12  1 0  0 TRUE FALSE
# 28     1013 5 24  0 0  0 TRUE FALSE
# 29     1014 0  4  0 0  0 TRUE FALSE
# 30     1015 19 22  0 0  1 TRUE FALSE
# 31     3367 15 3  0 0  0 TRUE FALSE
# 32     1335 33 22  0 0  0 TRUE FALSE
# 33 Chr1 2319  C
# 34     1007 0 28 10 0  1 TRUE FALSE
# 35     1012 0 43 17 0  1 TRUE FALSE
# 36     1013 13 15  9 0  1 TRUE FALSE
# 37     1014 0 18  6 0  1 TRUE FALSE
# 38     1015 0 53 20 0  1 TRUE FALSE
# 39     3367 4  0 24 0  0 TRUE FALSE
# 40     1335 0 118 28 0  1 TRUE FALSE
# 41 Chr1 2502  A
# 42     1007 14 2  0 0  0 FALSE FALSE
# 43     1012 17 6  0 0  0 FALSE FALSE
# 44     1013 6 13  0 0  0 FALSE FALSE
# 45     1014 1  6  0 0  0 FALSE FALSE
# 46     1015 20 7  0 0  0 FALSE FALSE
# 47     3367 3  3  0 0  0 FALSE FALSE
# 48     1335 29 17  0 0  0 FALSE FALSE
# 49 Chr1 2573  C
# 50     1007 0  0 11 28  1 TRUE FALSE
# 51     1012 0  0 30 50  1 TRUE FALSE
# 52     1013 0  0 231 12  0 TRUE FALSE
# 53     1014 0  0  4 18  1 TRUE FALSE

```

```

# 54      1015  0  0  50 38  1 TRUE FALSE
# 55      3367  0  0  71  0  0 TRUE FALSE
# 56      1335  0  0  62 75  1 TRUE FALSE
# 57 Chr1 3938  G
# 58      1007 12 20  0  0  0 TRUE FALSE
# 59      1012  9 22  0  0  0 TRUE FALSE
# 60      1013 35 19  0  0  0 TRUE FALSE
# 61      1014  8  2  0  0  0 TRUE FALSE
# 62      1015 25 53  0  0  0 TRUE FALSE
# 63      3367 14 13  0  0  0 TRUE FALSE
# 64      1335 59 42  0  0  0 TRUE FALSE
# 65 Chr1 4876  G
# 66      1007  0  1  0  0  0 FALSE FALSE
# 67      1012  1  4  0  0  0 FALSE FALSE
# 68      1013  0  0  0  0  0 FALSE FALSE
# 69      1014  1  0  0  0  0 FALSE FALSE
# 70      1015  0  3  0  0  0 FALSE FALSE
# 71      3367  4  4  0  0  0 FALSE FALSE
# 72      1335  2  2  0  0  0 FALSE FALSE
# 73 Chr1 4938  T
# 74      1007  0 43  0 23  1 FALSE FALSE
# 75      1012  0 63  0 48  1 FALSE FALSE
# 76      1013  0 83  0  2  0 FALSE FALSE
# 77      1014  0 27  0  4  1 FALSE FALSE
# 78      1015  0 75  0 47  1 FALSE FALSE
# 79      3367  0 19  0 12  1 FALSE FALSE
# 80      1335  0 57  0 59  1 FALSE FALSE

# one of those is a little weird:
xx<-snp.tables[[1]][149457,]
for (i in 2:7){xx <- rbind(xx,snp.tables[[i]][149457,])}
row.names(xx)<-names(snp.tables)
# My guess is that Chr/Pos/Ref are left as NA if coverage is zero.
xx

#      snp  Chr  Pos  Ref  Cov  a  g  c  t  n .match  exon  indel  chr  pos  rawCov
# 1007  0 <NA>  NA <NA>  0 0 0 0 0 0  0 FALSE FALSE <NA>  NA  0
# 1012  0 <NA>  NA <NA>  0 0 0 0 0 0  0 FALSE FALSE <NA>  NA  0
# 1013  0 <NA>  NA <NA>  0 0 0 0 0 0  0 FALSE FALSE <NA>  NA  0
# 1014  0 Chr1 149457  G  0 0 0 0 0 0  0 FALSE FALSE Chr1 149457  1
# 1015  0 <NA>  NA <NA>  0 0 0 0 0 0  0 FALSE FALSE <NA>  NA  0
# 3367  0 <NA>  NA <NA>  0 0 0 0 0 0  0 FALSE FALSE <NA>  NA  0
# 1335  0 Chr1 149457  G  0 0 0 0 0 0  0 FALSE FALSE Chr1 149457  1

```

5 Table 1 stats

Here is a brief summary of per-strain SNP counts, pairwise overlaps, and other conveniently available stats, such as those shown in Table 1 of the paper.

```

snp.counts    <- matrix(NA,7,4)
snp.pctofny   <- matrix(NA,7,4)
snp.pctofself <- matrix(NA,7,4)
snp.inter     <- matrix(NA,7,7)
snp.union     <- matrix(NA,7,7)
rownames(snp.counts)    <- names(snp.tables)
rownames(snp.pctofny)   <- names(snp.tables)
rownames(snp.pctofself) <- names(snp.tables)
rownames(snp.inter)     <- names(snp.tables)
colnames(snp.inter)     <- names(snp.tables)
rownames(snp.union)     <- names(snp.tables)
colnames(snp.union)     <- names(snp.tables)
for(stringency in 1:4){
  cat('\nStringency', stringency, ifelse(stringency==4,'(i.e. raw SAMTools SNP calls)', ''),
      ':\n-----\n')
}

```



```

for(i in 1:7){
  f.snps.i <- refined.snps$Code$get.snps(i, stringency)
  snp.counts[i,stringency] <- sum(f.snps.i)
  for(j in i:7){
    f.snps.j <- refined.snps$Code$get.snps(j, stringency)
    snp.inter[i,j] <- sum(f.snps.i & f.snps.j)
    snp.union[i,j] <- sum(f.snps.i | f.snps.j)
  }
}
snp.pctofny [,stringency] <- snp.inter[,7]/snp.counts[7,stringency]
snp.pctofself[,stringency] <- snp.inter[,7]/snp.counts[,stringency]
cat('Union Counts:\n'); print(snp.union)
cat('Intersect Counts:\n'); print(snp.inter)
cat('Intersect as percent of union:\n'); print(snp.inter/snp.union*100,digits=3)
}

#
# Stringency 1 :
# -----
# Union Counts:
#      1007   1012   1013   1014   1015   3367   1335
# 1007 184621 190979 363297 196256 197762 354191 199128
# 1012      NA 187793 364751 198002 198919 355526 200266
# 1013      NA      NA 296795 356666 366717 391621 364222
# 1014      NA      NA      NA 165741 196847 347035 195294
# 1015      NA      NA      NA      NA 191668 357845 198939
# 3367      NA      NA      NA      NA      NA 283086 355107
# 1335      NA      NA      NA      NA      NA      NA 187044
# Intersect Counts:
#      1007   1012   1013   1014   1015   3367   1335
# 1007 184621 181435 118119 154106 178527 113516 172537
# 1012      NA 187793 119837 155532 180542 115353 174571
# 1013      NA      NA 296795 105870 121746 188260 119617
# 1014      NA      NA      NA 165741 160562 101792 157491
# 1015      NA      NA      NA      NA 191668 116909 179773
# 3367      NA      NA      NA      NA      NA 283086 115023
# 1335      NA      NA      NA      NA      NA      NA 187044
# Intersect as percent of union:
#      1007 1012 1013 1014 1015 3367 1335
# 1007 100 95 32.5 78.5 90.3 32.0 86.6
# 1012  NA 100 32.9 78.6 90.8 32.4 87.2
# 1013  NA NA 100.0 29.7 33.2 48.1 32.8
# 1014  NA NA      NA 100.0 81.6 29.3 80.6
# 1015  NA NA      NA      NA 100.0 32.7 90.4
# 3367  NA NA      NA      NA      NA 100.0 32.4
# 1335  NA NA      NA      NA      NA      NA 100.0
#
# Stringency 2 :
# -----
# Union Counts:
#      1007   1012   1013   1014   1015   3367   1335
# 1007 181682 189049 374070 191223 196135 364611 195371
# 1012      NA 186199 376272 194060 197053 366680 196729
# 1013      NA      NA 304444 356961 378452 408556 373836
# 1014      NA      NA      NA 137771 193813 346571 187896
# 1015      NA      NA      NA      NA 190384 369360 195610
# 3367      NA      NA      NA      NA      NA 290844 364209
# 1335      NA      NA      NA      NA      NA      NA 180709
# Intersect Counts:
#      1007   1012   1013   1014   1015   3367   1335
# 1007 181682 178832 112056 128230 175931 107915 167020
# 1012      NA 186199 114371 129910 179530 110363 170179
# 1013      NA      NA 304444 85254 116376 186732 111317
# 1014      NA      NA      NA 137771 134342 82044 130584
# 1015      NA      NA      NA      NA 190384 111868 175483
# 3367      NA      NA      NA      NA      NA 290844 107344
# 1335      NA      NA      NA      NA      NA      NA 180709
# Intersect as percent of union:

```

```

#      1007  1012  1013  1014  1015  3367  1335
# 1007  100  94.6  30.0  67.1  89.7  29.6  85.5
# 1012   NA 100.0  30.4  66.9  91.1  30.1  86.5
# 1013   NA   NA 100.0  23.9  30.8  45.7  29.8
# 1014   NA   NA   NA 100.0  69.3  23.7  69.5
# 1015   NA   NA   NA   NA 100.0  30.3  89.7
# 3367   NA   NA   NA   NA   NA 100.0  29.5
# 1335   NA   NA   NA   NA   NA   NA 100.0
#
# Stringency 3 :
# -----
# Union Counts:
#      1007  1012  1013  1014  1015  3367  1335
# 1007 169346 183708 363625 176590 190884 354428 188236
# 1012   NA 179828 368821 185220 192949 359446 191771
# 1013   NA   NA 296287 330697 371321 403242 364437
# 1014   NA   NA   NA 88328 187158 319824 176609
# 1015   NA   NA   NA   NA 184704 362666 191220
# 3367   NA   NA   NA   NA   NA 283373 355133
# 1335   NA   NA   NA   NA   NA   NA 171199
# Intersect Counts:
#      1007  1012  1013  1014  1015  3367  1335
# 1007 169346 165466 102008 81084 163166 98291 152309
# 1012   NA 179828 107294 82936 171583 103755 159256
# 1013   NA   NA 296287 53918 109670 176418 103049
# 1014   NA   NA   NA 88328 85874 51877 82918
# 1015   NA   NA   NA   NA 184704 105411 164683
# 3367   NA   NA   NA   NA   NA 283373 99439
# 1335   NA   NA   NA   NA   NA   NA 171199
# Intersect as percent of union:
#      1007  1012  1013  1014  1015  3367  1335
# 1007  100  90.1  28.1  45.9  85.5  27.7  80.9
# 1012   NA 100.0  29.1  44.8  88.9  28.9  83.0
# 1013   NA   NA 100.0  16.3  29.5  43.7  28.3
# 1014   NA   NA   NA 100.0  45.9  16.2  47.0
# 1015   NA   NA   NA   NA 100.0  29.1  86.1
# 3367   NA   NA   NA   NA   NA 100.0  28.0
# 1335   NA   NA   NA   NA   NA   NA 100.0
#
# Stringency 4 (i.e. raw SAMTools SNP calls) :
# -----
# Union Counts:
#      1007  1012  1013  1014  1015  3367  1335
# 1007 161103 176738 343873 171675 185741 336599 180313
# 1012   NA 166089 346766 176177 186459 339458 182312
# 1013   NA   NA 247737 302322 352586 386037 339669
# 1014   NA   NA   NA 89184 179976 295574 162912
# 1015   NA   NA   NA   NA 174701 345396 184068
# 3367   NA   NA   NA   NA   NA 240413 331982
# 1335   NA   NA   NA   NA   NA   NA 153901
# Intersect Counts:
#      1007  1012  1013  1014  1015  3367  1335
# 1007 161103 150454 64967 78612 150063 64917 134691
# 1012   NA 166089 67060 79096 154331 67044 137678
# 1013   NA   NA 247737 34599 69852 102113 61969
# 1014   NA   NA   NA 89184 83909 34023 80173
# 1015   NA   NA   NA   NA 174701 69718 144534
# 3367   NA   NA   NA   NA   NA 240413 62332
# 1335   NA   NA   NA   NA   NA   NA 153901
# Intersect as percent of union:
#      1007  1012  1013  1014  1015  3367  1335
# 1007  100  85.1  18.9  45.8  80.8  19.3  74.7
# 1012   NA 100.0  19.3  44.9  82.8  19.8  75.5
# 1013   NA   NA 100.0  11.4  19.8  26.5  18.2
# 1014   NA   NA   NA 100.0  46.6  11.5  49.2
# 1015   NA   NA   NA   NA 100.0  20.2  78.5
# 3367   NA   NA   NA   NA   NA 100.0  18.8

```

```
# 1335    NA    NA    NA    NA    NA    NA 100.0

vs.stringency <- cbind(snp.counts, matrix(NA,7,1), round(snp.counts[,1:3]/snp.counts[,4]*100,1))
colnames(vs.stringency) <- c('[[1]]', '[[2]]', '[[3]]', '[[4]]', '----', '[[1]]%', '[[2]]%', '[[3]]%')

# SNPs vs filtering stringency (raw counts and as % of [[4]]). Medium filter
# adds 10-20% in most cases. Big exception is Gyre, where low coverage,
# high err rate and SAMTools conservatism seemed to seriously undercall:
print(vs.stringency)

#      [[1]] [[2]] [[3]] [[4]] ---- [[1]]% [[2]]% [[3]]%
# 1007 184621 181682 169346 161103    NA 114.6 112.8 105.1
# 1012 187793 186199 179828 166089    NA 113.1 112.1 108.3
# 1013 296795 304444 296287 247737    NA 119.8 122.9 119.6
# 1014 165741 137771 88328 89184    NA 185.8 154.5 99.0
# 1015 191668 190384 184704 174701    NA 109.7 109.0 105.7
# 3367 283086 290844 283373 240413    NA 117.7 121.0 117.9
# 1335 187044 180709 171199 153901    NA 121.5 117.4 111.2

# Intersect NY as % of self (vs stringency):
print(snp.pctofself*100, digits=3)

#      [,1] [,2] [,3] [,4]
# 1007 93.5 91.9 89.9 83.6
# 1012 93.0 91.4 88.6 82.9
# 1013 40.3 36.6 34.8 25.0
# 1014 95.0 94.8 93.9 89.9
# 1015 93.8 92.2 89.2 82.7
# 3367 40.6 36.9 35.1 25.9
# 1335 100.0 100.0 100.0 100.0

# Intersect NY as % of NY (vs stringency):
print(snp.pctofny*100, digits=3)

#      [,1] [,2] [,3] [,4]
# 1007 92.2 92.4 89.0 87.5
# 1012 93.3 94.2 93.0 89.5
# 1013 64.0 61.6 60.2 40.3
# 1014 84.2 72.3 48.4 52.1
# 1015 96.1 97.1 96.2 93.9
# 3367 61.5 59.4 58.1 40.5
# 1335 100.0 100.0 100.0 100.0
```

Quick look at coverage. Are there any NA?:

```
nacount <- NULL
for(i in 1:4){
  if(!is.null(tset[[i]])){
    nacount <- rbind(nacount,
                     unlist(lapply(tset[[i]], function(x){sum(is.na(x$Cov))}))
  )
  rownames(nacount)[nrow(nacount)] <- names(tset)[i]
}
}
nacount

#      1007 1012 1013 1014 1015 3367 1335
# snp.tables.full.unfiltered    0    0    0    0    0    0    0
# snp.tables.full.qfiltered     0    0    0    0    0    0    0
```

Seemingly no. What's average in unq- vs q-filtered:

```
snp.tables.unqfil <- tset.picker(c(1,2), table.set = tset)
snp.tables.qfil   <- tset.picker(c(3,4), table.set = tset)
cov.unqfil <- unlist(lapply(snp.tables.unqfil, function(x){mean(x$Cov)}))
cov.qfil   <- unlist(lapply(snp.tables.qfil,   function(x){mean(x$Cov, na.rm=T)}))
```

```

cov.both <- rbind(cov.unqfil, cov.qfil, cov.qfil/cov.unqfil)
i <- 1
if(!is.null(snp.tables.unqfil)){
  rownames(cov.both)[i] <- which.snp.tables(snp.tables.unqfil)
  i <- i+1
}
if(!is.null(snp.tables.qfil)){
  rownames(cov.both)[i] <- which.snp.tables(snp.tables.qfil)
  i <- i+1
}
if(i==3){
  rownames(cov.both)[i] <- 'Ratio'
}
cat('Mean Coverage:\n'); cov.both

# Mean Coverage:
#
#          1007          1012          1013          1014          1015          3367          1335
# trunc-unfiltered 37.0555484 70.8060724 69.6610432 33.1009373 61.5365159 64.0284488 107.7425968
# trunc-qfiltered  28.2750286 51.3249686 45.4036337 13.7261052 48.7880005 44.8042054  81.8823765
# Ratio           0.7630444  0.7248668  0.6517794  0.4146742  0.7928301  0.6997547  0.7599815

```

5.1 Table 1 Data

Throw together the conveniently-available Table 1 data, in Table 1 row order:

```

# if coverage unavailable, build NA vector
if(!is.null(cov.unqfil)){cov.unqfilv <- cov.unqfil} else {cov.unqfilv <- rep(NA, times=7)}
if(!is.null(cov.qfil )){cov.qfilv <- cov.qfil } else {cov.qfilv <- rep(NA, times=7)}
tldata.df <- data.frame(
  id      = st.locs(1:7, id=T, loc=F, date=F),
  loc     = st.locs(1:7, id=F, loc=T, date=F),
  date    = st.locs(1:7, id=F, loc=F, date=T),
  cov.unq = cov.unqfilv,
  cov.q    = cov.qfilv,
  SNPs.4   = snp.counts[,4],
  SNPs.2   = snp.counts[,2],
  olap.ny.4 = snp.pctofny[,4]*100,
  olap.ny.2 = snp.pctofny[,2]*100
)
tlrow.order <- c(7,1,2,5,3,6,4)
print(tldata.df[tlrow.order,], digits=3)

#          id          loc date cov.unq cov.q SNPs.4 SNPs.2 olap.ny.4 olap.ny.2
# 1335 CCMP1335      New York 1958  107.7  81.9 153901 180709    100.0    100.0
# 1007 CCMP1007      Virginia 1964   37.1  28.3 161103 181682     87.5     92.4
# 1012 CCMP1012  W. Australia 1965   70.8  51.3 166089 186199     89.5     94.2
# 1015 CCMP1015    Puget Sound 1985   61.5  48.8 174701 190384     93.9     97.1
# 1013 CCMP1013        Wales 1973   69.7  45.4 247737 304444     40.3     61.6
# 3367 CCMP3367        Italy 2007   64.0  44.8 240413 290844     40.5     59.4
# 1014 CCMP1014 N. Pacific Gyre 1971  33.1  13.7  89184 137771     52.1     72.3

```

6 Shared-SNPs P-Value

Text of the main paper quotes a “p-value” for the observed degree of SNP sharing in L-clade (and/or L-clade excluding Gyre) under a null model that these isolates were sampled from a population globally in Hardy-Weinberg equilibrium. Details of this analysis are as follows.

6.1 SNP Concordance

Arbitrarily pick one isolate, say, A , as the “template”. Arbitrarily pick a heterozygous (aka “SNP”) position in A . Let p_1 , and $q_1 = 1 - p_1$ be the frequencies in the overall population of the two nucleotides observed at that position in A .

(Positions having 3 or 4 nucleotide variants segregating in the population are assumed to be negligibly rare.) Under the HWE null model, a second isolate *B* will also be heterozygous at the same position with probability $2p_1q_1 \leq 1/2$. Similarly, this position will be heterozygous in a third isolate *C* with the same probability, *independently*, and so on for isolates *D* and *E*. Overall, (assuming HWE) the probability that a heterozygous position in *A* is simultaneously heterozygous in the other 4 isolates is at most $1/2^4 = 1/16$. Continuing, suppose we pick a second heterozygous position in *A*, on a different chromosome with allele frequencies $p_2, q_2 = 1 - p_2$, say. Again assuming HWE, this position will be a SNP in all of *B, C, D* and *E* with probability $(2p_2q_2)^4 \leq 1/16$, and this is independent of the first position, since segregation on different chromosomes is unlinked. Repeat this at 24 heterozygous positions in *A*, one per chromosome. Then, the number of five-way concordant positions observed should be dominated by the number observed when sampling from a binomial distribution with parameters $n = 24$ and $p = 1/16$, i.e., we expect at most $1/16 = 6.25\%$ of positions to agree, or at most $24/16 = 1.5$ five-way concordant positions in total. In sharp contrast, choosing CCMP 1014 (North Pacific Gyre) as the template, we see many more five-way concordant positions than predicted under these assumptions:

```
gyre.count <- sum(snp.tables[[4]]$snp)
# NOTE: what we now call "refined" SNPs were once called "filtered" SNPs and I have NOT tried
# to update variable names and annotation in the code below to reflect the terminology change...
# 'unfil.' => unfiltered for consistency; see below.
unfil.fiveway.count <- sum(snp.tables[[4]]$snp * i4.snps)
unfil.fiveway.percent <- unfil.fiveway.count / gyre.count * 100
unfil.p.value <- pbinom(floor(unfil.fiveway.count/gyre.count*24)-1, 24, 1/16, lower.tail = FALSE)
consistency.comparison <-
  data.frame(
    fiveway.count = unfil.fiveway.count,
    fiveway.percent = unfil.fiveway.percent,
    p.value = unfil.p.value
  )
consistency.comparison

#   fiveway.count fiveway.percent      p.value
# 1          70687          79.25973 4.142632e-19
```

Namely, 89184 positions are called as SNPs in CCMP1014, of which 70687 or 79.2597327% are also called as SNPs in *all four* other L-clade isolates. 79.2597327% of 24 is 19.0223358, and the probability of seeing 19 or more “Heads” in 24 flips of a biased coin with $P(\text{Heads}) \leq 1/16$, i.e., our p-value under the HWE null hypothesis, is at most: $4.1426317 \times 10^{-19}$ based on this simple binomial model. This is obviously strong evidence against the null hypothesis.

This analysis is potentially overly-simplistic in four respects, addressed below.

1. “ $2pq \leq 1/2$ ” is conservative. Neutral theory predicts that most variant nucleotides are rare in the population, so $2pq \ll 1/2$ is to be expected. This should make our quoted p-value very conservative.
2. Effect of Erroneous SNP calls. We base our analysis on *predicted* (by SAMTOOLS) heterozygous positions, not absolute-truth, which may affect our conclusions. However,
 - False negatives in *A* are irrelevant, since we never examine those positions. (This is the motivation for using CCMP1014 as the template; it has the lowest predicted SNP rate, likely due to a high false negative rate in that sequencing run. As noted elsewhere, it had the lowest coverage and lowest sequence quality of the 7 isolates, both of which impare SNP calling.)
 - False negatives in *BCDE* make such positions appear *non*-concordant. For our purpose, this makes our statistic more conservative since it can only deflate a statistic that we argue is nevertheless unexpectedly large.
 - False positive calls in *A* are conservatively treated, as well: barring simultaneous false-positive calls in all of *BCDE*, such a position will appear non-concordant, again deflating the statistic. The *false* positive rates in *B, C, D* and *E* are unknown, but cannot exceed SAMTOOLS *total* positive rate, which is below 1% in all 7 isolates, suggesting a simultaneous *BCDE* false positive rate $< 10^{-8}$, which will have a negligible effect.

- A potentially more serious issue is a true positive in *A* aligned to false positives in *BCD* and/or *E*. (I.e., a position that is polymorphic in the population and heterozygous in *A*, under the HWE null model is likely to be homozygous for one of the two alleles in one or more of *BCDE*; false positive SNP calls in all of those isolates would make the site appear concordant, i.e., provide evidence against the null model.) However, (a) my impression is that SAMTOOLS is more prone to false negative calls than to false positive calls (see Section 4), and (b) we would need a high rate of false positives to turn a truly heterozygous but non-concordant *A* call into a false “concordant” call—I’d expect at most half (especially given point 1 above) of *BCDE* to be heterozygous, but all would need to be falsely declared heterozygous. Such a high false positive rate on *BCDE* seems unlikely (see previous bullet), and would likely be counterbalanced by a similarly increased rate of false positives on *A*, which, as noted, tend to deflate our statistic (previous bullet again).
 - Systematic errors. If there were, say, a sequence-context-dependent bias in the DNA sequencing, mapping and/or SNP-calling that tended to suggest (or hide) a SNP at some position, we’re going to systematically over- (or under-) estimate concordant SNPs across isolates. The discordance of called SNPs between the L- and H-clades and within the H-clade suggests that this is not a major problem, but it is worth noting as a possibility.
3. Discordant nucleotides at “concordant” SNP positions. A “shared” SNP at a given position might be, say, G/C in one isolate vs T/C in another, reflecting an unexpected tri-allelic position in the population or a technical sequencing error. It is inappropriate to count such a “shared” SNP position as evidence against the null hypothesis, since it isn’t clear that it is truly shared. Instead, I will identify such inconsistent positions, based on the “stringency [[2]]” criteria established above, and treat each as non-concordant. I.e., a position will be considered to be a “5-way concordant SNP” if and only if it was called as a SNP by SAMTOOLS (independently) in all 5 L-clade isolates, *and* shows the same dominant non-reference nucleotide in all 5, according to criteria [[2]] above. As it turns out, this correction has a very minor effect on the resulting p-value:

```
# 'unfil.' => Ignoring "consistency"; 'fil.' => Filtering for "consistency":
fil.fiveway.count <- sum((snp.tables[[4]]$snp * i4.snps)[union.snps == 1] & consistent[[2]])
fil.fiveway.percent <- fil.fiveway.count / gyre.count * 100
fil.p.value <- pbinom(floor(fil.fiveway.count/gyre.count*24)-1, 24, 1/16, lower.tail = FALSE)
# append new stats to previous table for easy comparison
consistency.comparison <-
  rbind(consistency.comparison,
        data.frame(
          fiveway.count = fil.fiveway.count,
          fiveway.percent = fil.fiveway.percent,
          p.value = fil.p.value
        )
  )
rownames(consistency.comparison) <- c('unfiltered', 'consistency.filtered')
consistency.comparison

#               fiveway.count fiveway.percent      p.value
# unfiltered           70687         79.25973 4.142632e-19
# consistency.filtered    69941         78.42326 1.976512e-17
```

In particular, it removes 0.8% of five-way consistent positions (only 746 of 70687 positions), and still shows a highly significant p-value.

4. “ $P(E[X]) \neq E[P(X)]$ ”. I’m expressing this poorly, but finding the p-value based on the *expected* number of concordant positions is somewhat non-standard. A more typical set-up would use the *actual* value of some statistic, then calculate the probability of observing a value that extreme (or more extreme) under the null model. The fundamental problem is that we have thousands of SNPs, but I don’t see an easy way to use more than 24 of them at a time, because potential genetic linkage seemingly destroys statistical independence, which is key to most simple analyses. A somewhat more formal, but still non-standard, approach is the following. Suppose we randomly sample one SNP per chromosome and count the number X of them that are 5-way concordant. What I outlined above calculated the p-value based on $E[X]$, the expected value of X , i.e., $P(E[X])$. Alternatively, we can calculate $E[P(X)]$, the expected p-value. (They are not the same.) In effect, this averages the p-values that

would be seen over many different randomly-sampled sets of 24 SNPs. This is not difficult to calculate. First, the probability that we would observe $0 \leq i \leq 24$ concordant positions in a sample of 24, given that 78.42% of positions are concordant follows this binomial distribution:

```
x.equals.i.distribution <- dbinom(0:24, 24, fil.fiveway.percent/100)
print(x.equals.i.distribution, digits=3)

# [1] 1.04e-16 9.04e-15 3.78e-13 1.01e-11 1.92e-10 2.80e-09 3.22e-08 3.01e-07 2.32e-06 1.50e-05
# [11] 8.18e-05 3.78e-04 1.49e-03 5.00e-03 1.43e-02 3.46e-02 7.07e-02 1.21e-01 1.71e-01 1.96e-01
# [21] 1.78e-01 1.23e-01 6.12e-02 1.93e-02 2.93e-03
```

Second, the p-value corresponding to $0 \leq i \leq 24$ observed concordant positions also follows a different binomial distribution:

```
p.val.of.x.equals.i <- c(1, pbinom(0:23, 24, 1/16, lower.tail = F))
print(p.val.of.x.equals.i, digits=3)

# [1] 1.00e+00 7.88e-01 4.48e-01 1.87e-01 5.95e-02 1.49e-02 3.01e-03 4.99e-04 6.90e-05 8.02e-06
# [11] 7.89e-07 6.60e-08 4.72e-09 2.87e-10 1.49e-11 6.59e-13 2.46e-14 7.66e-16 1.98e-17 4.14e-19
# [21] 6.88e-21 8.70e-23 7.88e-25 4.56e-27 1.26e-29
```

Finally, the expected (or “average”) p-value is just the weighted average of the latter values, weighted by the former:

```
e.of.p.of.x <- sum(x.equals.i.distribution * p.val.of.x.equals.i)
e.of.p.of.x

# [1] 6.808209e-10
```

This is still highly significant, but weaker than the $P(E[X])$ analysis, basically because $X < E[X]$ has a fair probability of occurring, and the corresponding p-value $P(X)$ rises rapidly as X declines.

Another way to look at the numbers:

```
pvdf <- data.frame(x.density=x.equals.i.distribution,
                   x.cdf=cumsum(x.equals.i.distribution),
                   pval.of.x=p.val.of.x.equals.i)
print(pvdf, digits=4)

#   x.density    x.cdf pval.of.x
# 1 1.037e-16 1.037e-16 1.000e+00
# 2 9.043e-15 9.147e-15 7.875e-01
# 3 3.780e-13 3.871e-13 4.476e-01
# 4 1.008e-11 1.046e-11 1.869e-01
# 5 1.922e-10 2.027e-10 5.950e-02
# 6 2.795e-09 2.998e-09 1.490e-02
# 7 3.217e-08 3.517e-08 3.010e-03
# 8 3.007e-07 3.358e-07 4.994e-04
# 9 2.322e-06 2.658e-06 6.899e-05
# 10 1.500e-05 1.766e-05 8.015e-06
# 11 8.181e-05 9.947e-05 7.887e-07
# 12 3.784e-04 4.779e-04 6.603e-08
# 13 1.490e-03 1.968e-03 4.716e-09
# 14 4.999e-03 6.967e-03 2.875e-10
# 15 1.428e-02 2.124e-02 1.493e-11
# 16 3.459e-02 5.584e-02 6.590e-13
# 17 7.072e-02 1.266e-01 2.456e-14
# 18 1.210e-01 2.475e-01 7.662e-16
# 19 1.710e-01 4.185e-01 1.977e-17
# 20 1.963e-01 6.148e-01 4.143e-19
# 21 1.783e-01 7.931e-01 6.877e-21
# 22 1.235e-01 9.165e-01 8.701e-23
# 23 6.119e-02 9.777e-01 7.884e-25
# 24 1.934e-02 9.971e-01 4.556e-27
# 25 2.929e-03 1.000e+00 1.262e-29
```

E.g., row 9 in that table says that the concordance rate (78%) is so high that a sample of 24 SNPs will almost always have 9 or more five-way concordant positions (probability of fewer is only $2.658\text{e-}06$), while under the null model, seeing 9 or more is very unlikely (probability at most $6.899\text{e-}05$). ***AM I OFF-BY-ONE INTERPRETING ROW 9 HERE??***

6.2 Notes

In earlier drafts, an analog of the above analysis was based on the concordance of *refined* SNPs. This now seems to me to be questionable, since the “refined” SNP calling makes SNPs called across L-clade non-independent. OTOH, the above analysis seems valid: SAMTOOLS was run on each isolate independently, and likewise “criterion [[2]]” is evaluated independently in each strain, and is being used here solely to remove SNP predictions, not to add them. “Systematic errors” as outlined above remain a potential problem, but again discordance with/within H-clade suggests that this is of limited concern.

For completeness, I did a similar analysis including a sample of H-clade comparisons: Gyre vs Italy, NY vs Italy, NY vs Italy+Wales, and of Italy vs Wales. As expected, none of these show a statistically significant p-value, although the $\approx 40\%$ concordance in the 2-way comparisons, while $< 1/2$ as predicted, is a bit higher than I expected based on “neutral theory implies many rare variants.” (I did not bother to include “criterion[[2]] filtering” in these calculations.)

```
# 'gi.twoway' => gyre vs italy 2-way concordance;
# 'ni.twoway' => new york vs italy 2-way concordance;
# not bothering with criterion[[2]] filtering
gi.twoway.count    <- sum(snp.tables[[4]]$snp * snp.tables[[6]]$snp)
gi.twoway.percent <- gi.twoway.count / gyre.count * 100
gi.p.value <- pbinom(floor(gi.twoway.count/gyre.count*24)-1, 24, 1/2, lower.tail = FALSE)
ny.count <- sum(snp.tables[[7]]$snp)
ni.twoway.count    <- sum(snp.tables[[7]]$snp * snp.tables[[6]]$snp)
ni.twoway.percent <- ni.twoway.count / ny.count * 100
ni.p.value <- pbinom(floor(ni.twoway.count/ny.count*24)-1, 24, 1/2, lower.tail = FALSE)
niw.threeway.count <- sum(snp.tables[[7]]$snp * snp.tables[[6]]$snp * snp.tables[[3]]$snp)
niw.threeway.percent <- niw.threeway.count / ny.count * 100
niw.p.value <- pbinom(floor(niw.threeway.count/ny.count*24)-1, 24, 1/4, lower.tail = FALSE)
it.count <- sum(snp.tables[[6]]$snp)
iw.twoway.count    <- sum(snp.tables[[6]]$snp * snp.tables[[3]]$snp)
iw.twoway.percent <- iw.twoway.count / it.count * 100
iw.p.value <- pbinom(floor(iw.twoway.count/it.count*24)-1, 24, 1/2, lower.tail = FALSE)
consistency.comparison <-
  rbind(consistency.comparison,
    data.frame(
      fiveway.count    = c(gi.twoway.count,    ni.twoway.count,    niw.threeway.count,    iw.twoway.count),
      fiveway.percent  = c(gi.twoway.percent,  ni.twoway.percent,  niw.threeway.percent, iw.twoway.percent),
      p.value          = c(gi.p.value,         ni.p.value,         niw.p.value,         iw.p.value)
    )
  )
colnames(consistency.comparison)[1:2] <- c('552232way.count', '552232way.percent') # old col names misleading
rownames(consistency.comparison)[3:6] <- c('gyre.vs.italy', 'new.york.vs.italy', 'ny.vs.it.plus.wales', 'it.vs.wales')
consistency.comparison

#           552232way.count 552232way.percent      p.value
# unfiltered              70687             79.25973 4.142632e-19
# consistency.filtered      69941             78.42326 1.976512e-17
# gyre.vs.italy             34023             38.14922 9.242052e-01
# new.york.vs.italy         62332             40.50136 9.242052e-01
# ny.vs.it.plus.wales       35796             23.25911 7.533516e-01
# it.vs.wales              102113            42.47399 8.462719e-01
```

6.3 P-Value: The Bottom Line

So, what to say in the body of the paper? $E[P(X)]$ is highly significant, and conservative, but complex to explain. $P(E[X])$ is simpler to explain, but may be criticized as misleading if we aren’t very careful in that explanation. I’m slightly leaning towards the last option, but want to sleep on it and draft the key sentence or two before settling.

7 Sharing

The following analysis looks at the sharing patterns among the consistent SNPs. I assume that shared SNPs reflect shared ancestry, and that SNPs accumulate slowly over time. Then, in outline, the story is consistent with what we have seen in other analyses—there seem to be 3 groups: 1013 (Wales) in one, 3367 (Italy) in another, and the other 5 in a third, with some hints as to the order of divergence. A caveat is that in a sexual population, non-shared SNPs do not immediately imply non-shared ancestry; they may merely reflect Hardy-Weinberg capturing a homozygous state in one isolate vs the other. (Or read errors, etc.) Thus, if we are right that the H-isolates retain sex, then the large number of “private” SNPs in H may be at least partially due to HWE.

Analysis is broken into cases based on how many strains share a particular SNP.

7.1 Code

To categorize SNPs by sharing patterns, first convert the 7-way consistent sharing pattern into a 7-bit binary number, and tabulate based on that:

```
# convert (n x 7) 0-1 matrix to n vector of 0-127
tobin <- function(x){
  bin <- integer(nrow(x)) # initialized to 0
  for(i in 1:7){
    bin <- bin*2 + as.integer(x[,i]>0)
  }
  return(bin)
}

# get full set of patterns
snp.pattern.all <- lapply(non.refs,tobin)
# prune to just the consistent ones
snp.pattern <- snp.pattern.all
for(i in 1:3){
  snp.pattern[[i]][!consistent[[i]]] <- NA
}

# analogous to built-in ``table'' but simpler. Count entries in an integer
# vector sharing values in a (smallish) range. Result is a 2-column matrix with
# the shared values in col 1 and count of occurrences of that value in col 2.
# Out-of-range values cause subscript error.
mytable <- function(vec, therange=range(vec,na.rm=T)){
  counts <- matrix(0,nrow=therange[2]-therange[1]+1,ncol=2,dimnames=list(NULL,c('val','count')))
  counts[1:nrow(counts),1] <- therange[1]:therange[2]
  for(i in 1:length(vec)){
    if(!is.na(vec[i])){
      counts[vec[i]-therange[1]+1,2] <- counts[vec[i]-therange[1]+1,2] + 1
    }
  }
  return(counts)
}

pattern.counts <- lapply(snp.pattern, function(x){mytable(x,c(0,127))})
```

To display the results, build a data frame whose i -th row, $0 \leq i \leq 127$ shows one of the 128 possible sharing patterns, with counts of the numbers of consistent, shared SNPs with that pattern according to criteria c1-c3.

```
tobitvec <- function(x){
  bitvec <- integer(7)
  for(i in 7:1){
    bitvec[i] <- x %% 2
    x <- x %/% 2
  }
  return(bitvec)
}

flg <- function(x){
```

```

    return(ifelse(x==1, 'X', ''))
  }

pat.summary <- function(listOfTbls){
  mydf <- data.frame(pat=0:127,sharedBy=NA,
    tp1007='',tp1012='',tp1013='',tp1014='',tp1015='',tp3367='',tp1335='',
    count1=NA,count2=NA,count3=NA,count4=NA,stringsAsFactors=F)

  for(i in 1:128){
    bvec <- tobitvec(i-1)
    mydf[i,'sharedBy']=sum(bvec)
    mydf[i,'tp1007']=flg(bvec[1])
    mydf[i,'tp1012']=flg(bvec[2])
    mydf[i,'tp1013']=flg(bvec[3])
    mydf[i,'tp1014']=flg(bvec[4])
    mydf[i,'tp1015']=flg(bvec[5])
    mydf[i,'tp3367']=flg(bvec[6])
    mydf[i,'tp1335']=flg(bvec[7])
  }

  for(i in 1:length(listOfTbls)){
    tbl <- listOfTbls[[i]]
    if(!is.null(tbl)){
      mydf[,9+i] <- tbl[,2] ## count1/2/3/4 are columns 10/11/12/13 in mydf
      #for(j in 1:length(tbl)){
      #  k <- as.integer(rownames(tbl)[j]);
      #  mydf[k+1,9+i] <- tbl[j] ## count1/2/3 are columns 10/11/12
      #}
    }
  }

  mydf$pat <-as.octmode(mydf$pat) # display bit pattern in octal
  return(mydf)
}

pat.summaries <- pat.summary(pattern.counts)

```

7.2 Sanity Checks

Some sanity checking: table sums equal to number of consistent positions?

```

all(consistent.count == apply(pat.summaries[,10:13],2,sum))

# [1] TRUE

```

More sanity checking: visually inspect a pattern with small counts, specifically pattern 12, i.e., consistent SNPs shared by only strains 1014 and 1015 (2nd and 3 rows from bottom, binary code $12 = 2^3 + 2^2$). There are only 10 such positions on Chr1. Chr1 2524239 has pattern 12 under criteria c1 and c2 but not c3; Chr1 1088766 has in c2 only. Both look good. Neither position is a *called* SNP except in 1015. However, all but 1 nonreference read agree with the called SNP (the exception being one read in Wales). Both 1014 and 1015 have at least 2 non-reference reads, comprising at least 5% of coverage, and in both strains, those reads are on the same non-reference base, satisfying criterion c2. The other strains have higher coverage and/or lower non-reference counts, so they do not satisfy c2. Position 2524239 also satisfies c1, but not c3, since 2 reads out of 35 is below the 10% threshold. (It is pattern 4 under c3, i.e., a SNP private to 1015.) Position 1088766 is also pattern 4 under c3 (2 reads out of 56 in 1335 is below both thresholds), and it is not consistent under c1, since the single A read in 1013 is discordant with the other non-reference reads.

```

unlist(lapply(snp.pattern,function(x){sum(x==12,na.rm=T)}))

# [1] 133 139 132 417

spl <- snp.pattern[[1]]==12
sp2 <- snp.pattern[[2]]==12

```

```

sp3 <- snp.pattern[[3]]==12
sp4 <- snp.pattern[[4]]==12
c(sum(sp1,na.rm=T), sum(sp2,na.rm=T), sum(sp3,na.rm=T), sum(sp4,na.rm=T))

# [1] 133 139 132 417

r1 <- rownames(non.refs[[1]])[which(sp1)]
r2 <- rownames(non.refs[[2]])[which(sp2)]
r3 <- rownames(non.refs[[3]])[which(sp3)]
r4 <- rownames(non.refs[[4]])[which(sp4)]

r2

# [1] "Chr1:1799155"      "Chr2:713075"      "Chr2:1464209"      "Chr2:2406031"
# [5] "Chr2:2480466"      "Chr2:2480532"      "Chr2:2480838"      "Chr2:2483322"
# [9] "Chr2:2488863"      "Chr2:2489189"      "Chr2:2490933"      "Chr2:2492886"
# [13] "Chr2:2492887"      "Chr2:2497794"      "Chr2:2500122"      "Chr2:2503000"
# [17] "Chr2:2507585"      "Chr2:2507680"      "Chr2:2510117"      "Chr2:2513923"
# [21] "Chr2:2515103"      "Chr2:2516669"      "Chr2:2516751"      "Chr2:2518558"
# [25] "Chr2:2518653"      "Chr2:2518980"      "Chr2:2519285"      "Chr2:2519288"
# [29] "Chr2:2519718"      "Chr2:2520984"      "Chr2:2521271"      "Chr2:2522648"
# [33] "Chr2:2524223"      "Chr2:2524439"      "Chr2:2525160"      "Chr2:2525463"
# [37] "Chr2:2527281"      "Chr2:2527916"      "Chr2:2528472"      "Chr2:2528769"
# [41] "Chr2:2529076"      "Chr2:2529140"      "Chr2:2529684"      "Chr2:2530064"
# [45] "Chr2:2530216"      "Chr2:2530239"      "Chr2:2530294"      "Chr2:2530768"
# [49] "Chr2:2530896"      "Chr2:2531114"      "Chr2:2531498"      "Chr2:2531567"
# [53] "Chr2:2532173"      "Chr2:2532365"      "Chr2:2533028"      "Chr2:2533171"
# [57] "Chr2:2533440"      "Chr2:2534441"      "Chr2:2535121"      "Chr2:2535122"
# [61] "Chr2:2535314"      "Chr2:2535493"      "Chr2:2535503"      "Chr2:2535509"
# [65] "Chr2:2535862"      "Chr2:2536242"      "Chr2:2537201"      "Chr2:2537864"
# [69] "Chr2:2537917"      "Chr2:2538072"      "Chr2:2538498"      "Chr2:2539318"
# [73] "Chr2:2543595"      "Chr2:2545615"      "Chr2:2545798"      "Chr2:2546865"
# [77] "Chr2:2546991"      "Chr2:2547055"      "Chr2:2547086"      "Chr2:2547120"
# [81] "Chr2:2547155"      "Chr2:2547212"      "Chr2:2547248"      "Chr2:2547318"
# [85] "Chr2:2547554"      "Chr2:2547938"      "Chr2:2547944"      "Chr2:2548131"
# [89] "Chr2:2549281"      "Chr2:2551574"      "Chr2:2551930"      "Chr2:2554708"
# [93] "Chr2:2554860"      "Chr2:2555005"      "Chr2:2555203"      "Chr2:2555820"
# [97] "Chr3:192441"      "Chr3:496665"      "Chr4:1086589"      "Chr4:1393682"
# [101] "Chr2:2547983"      "Chr4:2314475"      "Chr5:7509"         "Chr5:141375"
# [105] "Chr5:1071721"      "Chr6:1330532"      "Chr7:399475"       "Chr7:1736991"
# [109] "Chr7:1813303"      "Chr8:556556"      "Chr10:54351"       "Chr10:95217"
# [113] "Chr10:947088"      "Chr11a:344258"     "Chr11b:75778"      "Chr12:214112"
# [117] "Chr12:458461"      "Chr12:507608"      "Chr13:96361"       "Chr13:375598"
# [121] "Chr14:284131"      "Chr15:417704"      "Chr16a:39914"      "Chr16a:177501"
# [125] "Chr16a:206719"      "Chr16a:394030"      "Chr17:461465"      "Chr19a_19:300076"
# [129] "Chr19a_19:303090"  "Chr19c_29:64170"   "Chr19c_29:64811"   "Chr19c_29:65720"
# [133] "Chr20:230994"      "Chr20:486431"      "Chr20:519835"      "Chr22:380816"
# [137] "Chr23:190382"      "Chr23:274291"      "Chr24:114599"

c1 <- as.integer(unlist(lapply(strsplit(r1[1:min(20,length(r1))],':',fixed=TRUE),function(x){x[2]})))
c2 <- as.integer(unlist(lapply(strsplit(r2[1:min(20,length(r2))],':',fixed=TRUE),function(x){x[2]})))
c3 <- as.integer(unlist(lapply(strsplit(r3[1:min(20,length(r3))],':',fixed=TRUE),function(x){x[2]})))
c4 <- as.integer(unlist(lapply(strsplit(r4[1:min(20,length(r4))],':',fixed=TRUE),function(x){x[2]})))

c1

# [1] 614335 914018 1317406 2388286 62676 713075 2406031 2480466 2480838 2481998 2483322
# [12] 2488863 2489189 2490933 2492887 2497794 2500122 2503000 2507585 2507680

c2

# [1] 1799155 713075 1464209 2406031 2480466 2480532 2480838 2483322 2488863 2489189 2490933
# [12] 2492886 2492887 2497794 2500122 2503000 2507585 2507680 2510117 2513923

c3

# [1] 371484 518347 1210354 2209068 2264683 2898352 1276745 1464904 1464905 1766966 2347253
# [12] 2406031 2480532 2480838 2483322 2488863 2489189 2490933 2497794 2507585

```

c4

```
# [1] 518347 691730 767408 1049906 1390437 2072951 2254059 2254789 2264683 2823796 2898352
# [12] 2998868 77394 77407 155680 761325 968120 1182096 1222176 1264023
```

```
seecounts(c2,snp.tables=snp.tables)
```

#	chr	pos	Ref	Strain	A	G	C	T	SNP	exon	indel	nrf	rat
# 1	Chr1	1799155	C										
# 2				1007	0	0	10	1	0	TRUE	FALSE		
# 3				1012	0	0	16	1	0	TRUE	FALSE		
# 4				1013	0	0	10	0	0	TRUE	FALSE		
# 5				1014	0	0	8	2	0	TRUE	FALSE		
# 6				1015	0	0	12	3	1	TRUE	FALSE		
# 7				3367	1	0	1	1	1	TRUE	FALSE		
# 8				1335	0	0	7	1	0	TRUE	FALSE		
# 9	Chr1	713075	T										
# 10				1007	0	0	0	37	0	TRUE	FALSE		
# 11				1012	0	0	0	90	0	TRUE	FALSE		
# 12				1013	0	0	0	65	0	TRUE	FALSE		
# 13				1014	0	0	0	32	0	TRUE	FALSE		
# 14				1015	0	0	0	84	0	TRUE	FALSE		
# 15				3367	0	0	0	53	0	TRUE	FALSE		
# 16				1335	0	0	0	109	0	TRUE	FALSE		
# 17	Chr1	1464209	T										
# 18				1007	0	0	0	22	0	FALSE	FALSE		
# 19				1012	0	0	0	38	0	FALSE	FALSE		
# 20				1013	0	0	0	22	0	FALSE	FALSE		
# 21				1014	0	0	0	12	0	FALSE	FALSE		
# 22				1015	0	0	0	30	0	FALSE	FALSE		
# 23				3367	0	0	0	39	0	FALSE	FALSE		
# 24				1335	0	0	0	81	0	FALSE	FALSE		
# 25	Chr1	2406031	C										
# 26				1007	0	0	18	0	0	TRUE	FALSE		
# 27				1012	0	0	23	0	0	TRUE	FALSE		
# 28				1013	0	0	46	0	0	TRUE	FALSE		
# 29				1014	0	0	13	0	0	TRUE	FALSE		
# 30				1015	0	0	34	0	0	TRUE	FALSE		
# 31				3367	0	0	29	0	0	TRUE	FALSE		
# 32				1335	0	0	68	0	0	TRUE	FALSE		
# 33	Chr1	2480466	A										
# 34				1007	26	0	0	0	0	TRUE	FALSE		
# 35				1012	42	0	0	0	0	TRUE	FALSE		
# 36				1013	39	0	0	0	0	TRUE	FALSE		
# 37				1014	9	0	0	0	0	TRUE	FALSE		
# 38				1015	49	0	0	0	0	TRUE	FALSE		
# 39				3367	32	0	0	0	0	TRUE	FALSE		
# 40				1335	77	0	0	0	0	TRUE	FALSE		
# 41	Chr1	2480532	G										
# 42				1007	0	25	0	0	0	TRUE	FALSE		
# 43				1012	0	27	0	0	0	TRUE	FALSE		
# 44				1013	0	43	0	0	0	TRUE	FALSE		
# 45				1014	0	1	0	0	0	TRUE	FALSE		
# 46				1015	0	23	0	0	0	TRUE	FALSE		
# 47				3367	0	23	0	0	0	TRUE	FALSE		
# 48				1335	0	71	0	0	0	TRUE	FALSE		
# 49	Chr1	2480838	T										
# 50				1007	0	0	0	8	0	TRUE	FALSE		
# 51				1012	0	0	0	12	0	TRUE	FALSE		
# 52				1013	0	0	0	24	0	TRUE	FALSE		
# 53				1014	0	0	0	6	0	TRUE	FALSE		
# 54				1015	0	0	0	15	0	TRUE	FALSE		
# 55				3367	0	0	0	9	0	TRUE	FALSE		
# 56				1335	0	0	0	81	0	TRUE	FALSE		
# 57	Chr1	2483322	A										
# 58				1007	22	0	0	0	0	TRUE	FALSE		
# 59				1012	23	0	0	0	0	TRUE	FALSE		
# 60				1013	52	0	0	0	0	TRUE	FALSE		

# 61		1014	24	0	0	0	0	TRUE	FALSE
# 62		1015	55	0	0	0	0	TRUE	FALSE
# 63		3367	37	0	0	0	0	TRUE	FALSE
# 64		1335	82	0	0	0	0	TRUE	FALSE
# 65	Chr1 2488863	C							
# 66		1007	0	0	26	0	0	FALSE	FALSE
# 67		1012	0	0	34	0	0	FALSE	FALSE
# 68		1013	0	0	27	0	0	FALSE	FALSE
# 69		1014	0	0	11	0	0	FALSE	FALSE
# 70		1015	0	0	34	0	0	FALSE	FALSE
# 71		3367	0	0	43	0	0	FALSE	FALSE
# 72		1335	0	0	71	0	0	FALSE	FALSE
# 73	Chr1 2489189	C							
# 74		1007	0	0	32	0	0	FALSE	FALSE
# 75		1012	0	0	63	0	0	FALSE	FALSE
# 76		1013	0	0	44	0	0	FALSE	FALSE
# 77		1014	0	0	26	0	0	FALSE	FALSE
# 78		1015	0	0	59	0	0	FALSE	FALSE
# 79		3367	0	0	24	0	0	FALSE	FALSE
# 80		1335	0	0	110	0	0	FALSE	FALSE
# 81	Chr1 2490933	G							
# 82		1007	0	25	0	0	0	FALSE	FALSE
# 83		1012	0	57	0	0	0	FALSE	FALSE
# 84		1013	0	40	0	0	0	FALSE	FALSE
# 85		1014	0	9	0	0	0	FALSE	FALSE
# 86		1015	0	36	0	0	0	FALSE	FALSE
# 87		3367	0	37	0	1	0	FALSE	FALSE
# 88		1335	0	57	0	0	0	FALSE	FALSE
# 89	Chr1 2492886	T							
# 90		1007	0	0	0	27	0	FALSE	FALSE
# 91		1012	0	0	0	61	0	FALSE	FALSE
# 92		1013	0	0	0	41	0	FALSE	FALSE
# 93		1014	0	0	0	18	0	FALSE	FALSE
# 94		1015	0	0	0	53	0	FALSE	FALSE
# 95		3367	0	0	0	48	0	FALSE	FALSE
# 96		1335	0	0	0	80	0	FALSE	FALSE
# 97	Chr1 2492887	G							
# 98		1007	0	22	0	0	0	FALSE	FALSE
# 99		1012	0	61	0	0	0	FALSE	FALSE
# 100		1013	0	35	0	0	0	FALSE	FALSE
# 101		1014	0	17	0	0	0	FALSE	FALSE
# 102		1015	0	55	0	0	0	FALSE	FALSE
# 103		3367	0	50	0	0	0	FALSE	FALSE
# 104		1335	0	85	0	0	0	FALSE	FALSE
# 105	Chr1 2497794	T							
# 106		1007	0	0	0	35	0	TRUE	FALSE
# 107		1012	0	0	0	60	0	TRUE	FALSE
# 108		1013	0	0	0	58	0	TRUE	FALSE
# 109		1014	0	0	0	12	0	TRUE	FALSE
# 110		1015	0	0	0	64	0	TRUE	FALSE
# 111		3367	0	0	0	43	0	TRUE	FALSE
# 112		1335	0	0	0	107	0	TRUE	FALSE
# 113	Chr1 2500122	A							
# 114		1007	18	0	0	0	0	FALSE	FALSE
# 115		1012	47	0	0	0	0	FALSE	FALSE
# 116		1013	34	0	0	0	0	FALSE	FALSE
# 117		1014	6	0	0	0	0	FALSE	FALSE
# 118		1015	35	0	0	0	0	FALSE	FALSE
# 119		3367	27	0	0	0	0	FALSE	FALSE
# 120		1335	51	0	0	0	0	FALSE	FALSE
# 121	Chr1 2503000	T							
# 122		1007	0	0	0	29	0	FALSE	FALSE
# 123		1012	0	0	0	35	0	FALSE	FALSE
# 124		1013	0	0	0	57	0	FALSE	FALSE
# 125		1014	0	0	0	10	0	FALSE	FALSE
# 126		1015	0	0	0	34	0	FALSE	FALSE
# 127		3367	0	0	0	41	0	FALSE	FALSE

```

# 128      1335    0  0  0  28    0 FALSE FALSE
# 129 Chr1 2507585  A      1007  34  0  0  0    0 TRUE  FALSE
# 130      1012  55  0  0  0    0 TRUE  FALSE
# 131      1013  32  0  0  0    0 TRUE  FALSE
# 132      1014  13  0  0  0    0 TRUE  FALSE
# 133      1015  41  0  0  0    0 TRUE  FALSE
# 134      3367  61  0  0  0    0 TRUE  FALSE
# 135      1335 104  0  0  0    0 TRUE  FALSE
# 136 Chr1 2507680  A      1007  26  0  0  0    0 FALSE FALSE
# 137      1012  46  0  0  0    0 FALSE FALSE
# 138      1013  32  0  0  0    0 FALSE FALSE
# 139      1014  15  0  0  0    0 FALSE FALSE
# 140      1015  54  0  0  0    0 FALSE FALSE
# 141      3367  51  0  0  0    0 FALSE FALSE
# 142      1335  78  0  0  0    0 FALSE FALSE
# 143 Chr1 2510117  C      1007    0  0  19  0    0 TRUE  FALSE
# 144      1012    0  0  56  1    0 TRUE  FALSE
# 145      1013    0  0  42  0    0 TRUE  FALSE
# 146      1014    0  0  13  0    0 TRUE  FALSE
# 147      1015    0  0  39  0    0 TRUE  FALSE
# 148      3367    0  0  36  0    0 TRUE  FALSE
# 149      1335    0  0  92  0    0 TRUE  FALSE
# 150 Chr1 2513923  A      1007  39  0  0  0    0 FALSE FALSE
# 151      1012  57  0  0  0    0 FALSE FALSE
# 152      1013  23  0  0  0    0 FALSE FALSE
# 153      1014   4  0  0  0    0 FALSE FALSE
# 154      1015  39  0  0  0    0 FALSE FALSE
# 155      3367  53  0  0  0    0 FALSE FALSE
# 156      1335  53  0  0  0    0 FALSE FALSE
# 157
# 158
# 159
# 160

```

Position 1088766, however, is a good example of the situation that motivated this analysis—one strain has a G/C SNP and 5 of the other 6 strains have nonreference reads consistent with that SNP. Although, excluding 1015, the nonreference read counts are not high enough to justify a SNP call in any strain considered in isolation, the fact that they *consistently* agree with the 1015 SNP suggests that they are real. One alternative hypothesis is that there is some sequence-dependent bias at this locus that favors misreading a G as a C. On the other hand, one could equally well posit a shared SNP, and a locus-dependant bias that *supresses* C reads, explaining the unbalanced readout that we observe. However, it is hard to reconcile either view with the significant strain-specific patterns that we see in the shared SNPs (as seen below). I think a more likely explanation is that (a) there are some number of relatively rare SNPs present in each of the sampled populations, (b) some of these SNPs happened to be present in one or two cells of the roughly 5-10 cells that we believe constituted the founding population of the culture grown for sequencing, and (c) stochastic effects during culture growth and during sequencing may have further perturbed the apparent frequency of each variant, but the bottom line is that the above-threshold presence of consistent non-reference reads is evidence for shared SNPs at the population level (and the proportions of such reads represent estimates of the population-level frequencies of the variants, albeit a noisy estimate at any specific position).

An aside: I was curious to see whether there is any consistent pattern to positions that are called consistent SNPs in all but Italy, so I repeated the above, basically. My summary is that coverage in Italy tends to be below average in these positions, but otherwise they don't stand out. For the record:

```

abit <- snp.pattern[[2]]==125
abit[is.na(abit)]<-F
sum(abit)

# [1] 13630

rabit <- rownames(non.refs[[2]])[which(abit)]
rabits <- rabbit[1:20]
cabit <- as.integer(unlist(lapply(strsplit(rabits, ':', fixed=TRUE), function(x){x[2]})))
cabit

# [1] 1244 1575 6485 7181 7220 7661 8144 8208 8518 8552 8567 8670 8685 14361 15254
# [16] 15280 16103 25546 30784 33852

```

```
seecounts(cabit,snp.tables=snp.tables)
```

#	chr	pos	Ref	Strain	A	G	C	T	SNP	exon	indel	nrf	rat
# 1	Chr1	1244	G										
# 2				1007	2	25	0	0	0	TRUE	FALSE		
# 3				1012	3	32	0	0	0	TRUE	FALSE		
# 4				1013	10	24	0	0	1	TRUE	FALSE		
# 5				1014	3	17	0	0	0	TRUE	FALSE		
# 6				1015	15	43	0	0	1	TRUE	FALSE		
# 7				3367	0	1	0	0	0	TRUE	FALSE		
# 8				1335	82	65	0	0	1	TRUE	FALSE		
# 9	Chr1	1575	G										
# 10				1007	24	7	0	0	0	TRUE	FALSE		
# 11				1012	42	13	0	0	0	TRUE	FALSE		
# 12				1013	17	16	0	0	0	TRUE	FALSE		
# 13				1014	15	4	0	0	0	TRUE	FALSE		
# 14				1015	43	31	0	0	1	TRUE	FALSE		
# 15				3367	0	2	0	0	0	TRUE	FALSE		
# 16				1335	34	74	0	0	0	TRUE	FALSE		
# 17	Chr1	6485	G										
# 18				1007	24	19	0	0	0	TRUE	FALSE		
# 19				1012	29	29	0	0	0	TRUE	FALSE		
# 20				1013	49	33	0	0	0	TRUE	FALSE		
# 21				1014	6	5	0	0	0	TRUE	FALSE		
# 22				1015	31	32	0	0	1	TRUE	FALSE		
# 23				3367	0	37	0	0	0	TRUE	FALSE		
# 24				1335	62	52	0	0	0	TRUE	FALSE		
# 25	Chr1	7181	G										
# 26				1007	0	30	29	0	0	TRUE	FALSE		
# 27				1012	0	52	34	0	0	TRUE	FALSE		
# 28				1013	0	19	72	0	0	TRUE	FALSE		
# 29				1014	0	13	7	0	0	TRUE	FALSE		
# 30				1015	0	40	33	0	1	TRUE	FALSE		
# 31				3367	0	29	0	0	0	TRUE	FALSE		
# 32				1335	0	78	73	0	0	TRUE	FALSE		
# 33	Chr1	7220	C										
# 34				1007	16	0	19	6	0	TRUE	FALSE		
# 35				1012	38	0	22	11	0	TRUE	FALSE		
# 36				1013	82	1	30	9	0	TRUE	FALSE		
# 37				1014	12	0	6	2	0	TRUE	FALSE		
# 38				1015	55	0	22	5	1	TRUE	FALSE		
# 39				3367	0	0	8	0	0	TRUE	FALSE		
# 40				1335	55	0	32	20	0	TRUE	FALSE		
# 41	Chr1	7661	T										
# 42				1007	0	0	9	9	0	TRUE	FALSE		
# 43				1012	0	0	5	19	0	TRUE	FALSE		
# 44				1013	0	0	24	14	1	TRUE	FALSE		
# 45				1014	0	0	6	3	0	TRUE	FALSE		
# 46				1015	0	0	5	34	0	TRUE	FALSE		
# 47				3367	0	0	0	4	0	TRUE	FALSE		
# 48				1335	0	0	4	24	0	TRUE	FALSE		
# 49	Chr1	8144	G										
# 50				1007	8	9	0	0	0	TRUE	FALSE		
# 51				1012	12	10	0	0	1	TRUE	FALSE		
# 52				1013	38	29	0	0	0	TRUE	FALSE		
# 53				1014	5	4	0	0	0	TRUE	FALSE		
# 54				1015	15	16	0	0	0	TRUE	FALSE		
# 55				3367	0	0	0	0	0	TRUE	FALSE		
# 56				1335	12	15	0	0	1	TRUE	FALSE		
# 57	Chr1	8208	G										
# 58				1007	0	6	0	7	1	TRUE	FALSE		
# 59				1012	0	19	0	11	0	TRUE	FALSE		
# 60				1013	0	1	0	48	0	TRUE	FALSE		
# 61				1014	0	5	0	3	0	TRUE	FALSE		
# 62				1015	0	19	0	11	1	TRUE	FALSE		
# 63				3367	0	1	0	0	0	TRUE	FALSE		
# 64				1335	0	28	0	16	1	TRUE	FALSE		
# 65	Chr1	8518	T										
# 66				1007	0	0	20	15	1	FALSE	FALSE		
# 67				1012	0	0	40	20	1	FALSE	FALSE		
# 68				1013	0	0	45	56	1	FALSE	FALSE		
# 69				1014	0	0	10	16	0	FALSE	FALSE		
# 70				1015	0	0	36	13	1	FALSE	FALSE		
# 71				3367	0	0	0	2	0	FALSE	FALSE		
# 72				1335	0	0	113	53	1	FALSE	FALSE		
# 73	Chr1	8552	G										
# 74				1007	3	9	0	0	0	TRUE	FALSE		
# 75				1012	20	21	0	0	0	TRUE	FALSE		
# 76				1013	28	16	0	0	1	TRUE	FALSE		
# 77				1014	6	2	0	0	0	TRUE	FALSE		

# 78				1015	14	13	0	0	0	TRUE	FALSE
# 79				3367	0	12	0	0	0	TRUE	FALSE
# 80				1335	24	47	0	0	0	TRUE	FALSE
# 81	Chr1	8567	A								
# 82				1007	14	18	0	0	1	TRUE	FALSE
# 83				1012	26	30	0	0	1	TRUE	FALSE
# 84				1013	50	66	0	0	1	TRUE	FALSE
# 85				1014	1	3	0	0	0	TRUE	FALSE
# 86				1015	12	31	0	0	1	TRUE	FALSE
# 87				3367	22	0	0	0	0	TRUE	FALSE
# 88				1335	51	40	0	0	1	TRUE	FALSE
# 89	Chr1	8670	A								
# 90				1007	7	0	0	5	0	TRUE	FALSE
# 91				1012	16	0	0	10	0	TRUE	FALSE
# 92				1013	16	0	0	11	0	TRUE	FALSE
# 93				1014	2	0	0	4	0	TRUE	FALSE
# 94				1015	14	0	0	10	1	TRUE	FALSE
# 95				3367	5	0	0	0	0	TRUE	FALSE
# 96				1335	7	0	0	6	0	TRUE	FALSE
# 97	Chr1	8685	G								
# 98				1007	6	15	0	0	0	TRUE	FALSE
# 99				1012	10	23	0	0	0	TRUE	FALSE
# 100				1013	18	21	0	0	1	TRUE	FALSE
# 101				1014	4	8	0	0	0	TRUE	FALSE
# 102				1015	10	24	0	0	1	TRUE	FALSE
# 103				3367	0	4	0	0	0	TRUE	FALSE
# 104				1335	5	32	0	0	0	TRUE	FALSE
# 105	Chr1	14361	A								
# 106				1007	20	7	0	0	0	FALSE	FALSE
# 107				1012	35	5	0	0	0	FALSE	FALSE
# 108				1013	1	11	0	0	1	FALSE	FALSE
# 109				1014	6	2	0	0	0	FALSE	FALSE
# 110				1015	35	7	0	0	0	FALSE	FALSE
# 111				3367	2	1	0	0	0	FALSE	FALSE
# 112				1335	50	8	0	0	0	FALSE	FALSE
# 113	Chr1	15254	T								
# 114				1007	11	0	0	16	1	FALSE	FALSE
# 115				1012	26	0	0	38	1	FALSE	FALSE
# 116				1013	37	0	0	48	1	FALSE	FALSE
# 117				1014	3	0	0	8	1	FALSE	FALSE
# 118				1015	18	0	0	32	1	FALSE	FALSE
# 119				3367	0	0	0	73	0	FALSE	FALSE
# 120				1335	13	0	0	32	1	FALSE	FALSE
# 121	Chr1	15280	T								
# 122				1007	0	13	0	20	1	FALSE	FALSE
# 123				1012	0	27	0	28	1	FALSE	FALSE
# 124				1013	0	5	0	64	0	FALSE	FALSE
# 125				1014	0	2	0	8	0	FALSE	FALSE
# 126				1015	0	19	0	29	1	FALSE	FALSE
# 127				3367	0	0	0	42	0	FALSE	FALSE
# 128				1335	0	21	0	70	1	FALSE	FALSE
# 129	Chr1	16103	A								
# 130				1007	10	0	11	0	1	FALSE	FALSE
# 131				1012	44	0	19	0	1	FALSE	FALSE
# 132				1013	21	0	13	0	1	FALSE	FALSE
# 133				1014	14	0	2	0	0	FALSE	FALSE
# 134				1015	29	0	10	0	1	FALSE	FALSE
# 135				3367	33	0	0	0	0	FALSE	FALSE
# 136				1335	47	0	11	0	0	FALSE	FALSE
# 137	Chr1	25546	A								
# 138				1007	23	0	0	14	1	FALSE	FALSE
# 139				1012	46	0	0	19	1	FALSE	FALSE
# 140				1013	6	0	0	42	1	FALSE	FALSE
# 141				1014	7	0	0	15	1	FALSE	FALSE
# 142				1015	52	0	0	17	1	FALSE	FALSE
# 143				3367	60	0	0	0	0	FALSE	FALSE
# 144				1335	67	0	0	5	0	FALSE	FALSE
# 145	Chr1	30784	C								
# 146				1007	16	0	13	0	1	TRUE	FALSE
# 147				1012	33	0	32	0	1	TRUE	FALSE
# 148				1013	19	0	33	0	1	TRUE	FALSE
# 149				1014	4	0	11	0	1	TRUE	FALSE
# 150				1015	39	0	29	0	1	TRUE	FALSE
# 151				3367	0	0	55	0	0	TRUE	FALSE
# 152				1335	46	0	50	0	1	TRUE	FALSE
# 153	Chr1	33852	C								
# 154				1007	0	24	25	0	1	FALSE	FALSE
# 155				1012	0	18	26	0	1	FALSE	FALSE
# 156				1013	0	28	33	0	1	FALSE	FALSE
# 157				1014	0	9	4	0	1	FALSE	FALSE


```
# 158      1015  0 19  28  0  1 FALSE FALSE
# 159      3367  0  0  26  0  0 FALSE FALSE
# 160      1335  0 30  53  0  1 FALSE FALSE
```

More sanity: there are 83 sites on Chr1 shared by zero strains in the tightest condition. (I.e., SAMTOOLS called it a SNP, but the read counts/proportions fall below our 3rd threshold). Are they due to low coverage? Seemingly yes:

```
zp3 <- snp.pattern[[3]] == 0
zr3 <- rownames(non.refs[[3]])[which(zp3)]
zc3 <- as.integer(unlist(lapply(strsplit(zr3[1:min(100,length(zr3))],':',fixed=TRUE),function(x){x[2]})))
zc3
```

```
# [1] 16115 16615 19117 25748 43500 55857 56591 65787 66879 68328 80862 81001 90622
# [14] 90721 91284 110754 116443 116453 120183 126702 127986 129056 147698 153874 159756 160912
# [27] 161271 170686 180314 181477 182139 196862 196864 199166 206132 206143 221888 234931 242276
# [40] 242914 244505 268954 274655 282391 282511 283646 289363 311952 312625 314132 326217 371008
# [53] 376784 387078 387091 389263 395153 406158 410771 431788 438958 438976 443898 447253 448223
# [66] 452774 488812 495476 498133 501830 501975 504462 506422 515441 515595 530113 530114 532320
# [79] 534149 541667 543095 575081 585297 586276 612732 622585 651159 652889 655373 655380 657704
# [92] 657955 658216 685697 687653 692115 692139 700484 700845 701061
```

```
seecounts(zc3[1:5], snp.tables=snp.tables)
```

```
#      chr   pos Ref Strain  A G  C  T SNP  exon indel nrf rat
# 1  Chr1 16115   T
# 2      1007  0 0  0  5  0 FALSE FALSE
# 3      1012  0 0  0  9  0 FALSE FALSE
# 4      1013  0 0  0  6  0 FALSE FALSE
# 5      1014  0 0  0  3  0 FALSE FALSE
# 6      1015  0 0  0 10  0 FALSE FALSE
# 7      3367  0 0  3  3  1 FALSE FALSE
# 8      1335  0 0  0  6  0 FALSE FALSE
# 9  Chr1 16615   C
# 10     1007  0 0 39  0  0 FALSE FALSE
# 11     1012  0 0 54  0  0 FALSE FALSE
# 12     1013  0 0  4  2  1 FALSE FALSE
# 13     1014  0 0 19  0  0 FALSE FALSE
# 14     1015  0 0 46  0  0 FALSE FALSE
# 15     3367  0 0 13  0  0 FALSE FALSE
# 16     1335  0 0 40  0  0 FALSE FALSE
# 17 Chr1 19117   A
# 18     1007 16 0  0  0  0 TRUE FALSE
# 19     1012 21 0  0  0  0 TRUE FALSE
# 20     1013  1 0  0  1  0 TRUE FALSE
# 21     1014  6 0  0  0  0 TRUE FALSE
# 22     1015 21 0  0  0  0 TRUE FALSE
# 23     3367  0 0  0  1  1 TRUE FALSE
# 24     1335 24 0  0  0  0 TRUE FALSE
# 25 Chr1 25748   C
# 26     1007  0 0 17  0  0 FALSE FALSE
# 27     1012  0 0 36  0  0 FALSE FALSE
# 28     1013  3 0  7  0  1 FALSE FALSE
# 29     1014  1 0  4  0  0 FALSE FALSE
# 30     1015  0 0 32  0  0 FALSE FALSE
# 31     3367  0 0  1  0  0 FALSE FALSE
# 32     1335  1 0 34  0  0 FALSE FALSE
# 33 Chr1 43500   A
# 34     1007 10 0  0  3  1 FALSE FALSE
# 35     1012 10 0  0  3  1 FALSE FALSE
# 36     1013 10 0  1  1  0 FALSE FALSE
# 37     1014  5 0  0  0  0 FALSE FALSE
# 38     1015 11 0  0  2  0 FALSE FALSE
# 39     3367  6 0  0  3  0 FALSE FALSE
# 40     1335 13 0  0  1  0 FALSE FALSE
```

7.3 Main Analysis

Turning to the main analysis, there is a large increase in the number of consistent positions between the loose and medium stringency levels; medium and tight are similar in most respects. The likely interpretation is that the loose criterion is including many “SNPs” induced by read errors, and that either of the tighter criteria are successfully filtering them out. In the interest of simplicity, the narrative below will focus on the shared SNPs at the medium stringency level (the “count2” column in the data frame), although the numbers for all three (sometimes all 4) are displayed. Also note that the prose and some comments in the code were based on the Chr1 analysis, and so may occasionally be off-target for the whole-genome data.

```
# Show a subset of pat.summaries, optionally with totals of count_i in last row, and optionally
# aggregating low-count rows as ``Other``
#
#   sharedBy=c(2,4) selects SNPs shared by 2 or 4 strains,
#   subset=as.octmode('35') select those with sharing pattern a subset (optionally proper) of this
#   split=as.octmode('14') additionally restricts to patterns straddling split/subset minus split
#   c2.thresh=42 suppresses printout of rows with count2 < 42
#   restrict.to=c(0,42,127) restrict to these 3 rows
showgroup <- function(p.summ=pat.summaries, sharedBy=0:7, subset=127, split=NULL, proper.subset=F,
                      total=T, c2.thresh=0, fourteenth=F, restrict.to=NULL){
  # pick just those bit patterns that are subsets of 'subset'
  pick <- bitwAnd(0:127,bitwNot(subset))==0
  if(proper.subset){
    pick[subset+1] <- F
  }
  if(!is.null(split)){ # AND that straddle left/right subtrees
    cosplit <- bitwAnd(subset,bitwNot(split))
    pick <- pick & bitwAnd(0:127,split)!=0 & bitwAnd(0:127,cosplit)!=0
  }
  # and have desired shareBy counts
  pick <- pick & (p.summ$sharedBy %in% sharedBy)
  # and are among the set of interest
  if(!is.null(restrict.to)){
    pick <- pick & (0:127 %in% restrict.to)
  }
  # find rows with low counts
  pick.low <- pick & (p.summ$count2 < c2.thresh)
  # now show them
  show <- p.summ[pick & ! pick.low,]
  # rename columns just to narrow the printouts
  colnames(show) <- c('Pat','ShrBy','1007', '1012', '1013', '1014', '1015', '3367', '1335',
                      'count1', 'count2', 'count3','count4')
  show[,1] <- format(show[,1]) # convert octal col to char so can override in last row(2)
  nlow <- sum(pick.low)
  if(nlow > 0){
    n <- nrow(show)+1
    lows <- apply(p.summ[pick.low,10:13],2,sum)
    show[n,10:13] <- lows
    show[n,1:9] <- ''
    row.names(show)[n] <- 'Other'
    if(fourteenth){
      # do this: add 14th col just to hold this comment:
      show <- cbind(show, ' ', stringsAsFactors=F)
      show[n,14] <- paste('(', nlow, 'rows w/ c2 < ', c2.thresh, ')')
    } else {
      ## or this (looks a bit funky, but fits across page without line-wrap):
      show[n,1:8] <- c('(', nlow, 'rows', 'w/', 'c2', '<', c2.thresh, ')')
    }
  }
  if(total){
    n <- nrow(show)+1
    tots <- apply(show[,10:13],2,sum)
    show[n,10:13] <- tots
    show[n,1:9] <- ''
    row.names(show)[n] <- 'Total'
    if(ncol(show)==14){show[n,14]<-''}
  }
}
```

```
}
  return(show)
}
```

First, are there any SNPs that are not “consistent SNPs?” Yes, a few in c3. As noted above, they seem to be mainly low-coverage positions.

```
showgroup(pat.summaries,0,total=F) # chr1 totals: 0 0 83

#   Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 1    0     0                                111    578   4755     0
```

Next, look at completely shared SNPs, those found in all 7 strains.

```
showgroup(pat.summaries,7,total=F) # Chr1 count1 = 8593, count2 = 7054, count3 = 4790 c4=1641

#   Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 128 177    7    X    X    X    X    X    X    X  77690  62182  38744  15186
```

I.e., of the 469906 consistent positions, 62182 or 13.2% are shared by all 7 strains.

Next look at singletons, aka private SNPs—SNPs that are called in one strain and no other strain has a significant number of non-ref reads at that position. Presumably these are variants that arose in a given population after it separated from the others.

```
showgroup(pat.summaries,1) # chr1 totals: 9669 18865 19670 23574

#   Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 2    001    1                                X    449    632    1129    2260
# 3    002    1                                X  73721  85117  87494 105614
# 5    004    1                                X    1720   2156   2729   4608
# 9    010    1                                X    383     525     485   1231
# 17   020    1                                X  82364  94364  96464 113191
# 33   040    1                X    X    502     655    1102   2450
# 65   100    1    X                X    231     339     496   2005
# Total                                159370 183788 189899 231359
```

The import of shared/private SNPs changes between sexual and asexual populations. Presumably asexuals slowly gain and rarely lose private SNPs; shared ones predate separation of the lineages. In sexual lineages, however, SNPs may be rather freely “gained” or “lost,” merely by recombination (converting between homo- and heterozygous in the sample we sequenced). Thus, the low private counts for the 5 L-isolates compared to the large count of het positions overall suggest that (a) they are asexual, and (b) none of them has been isolated from the others for very long (if at all). Conversely, the high counts for Italy and Wales suggest that (a) if asexual, they have been separated from each other and from the rest for a long time, but (b) if sexual, there is little surprise: we have $\approx 160\text{K}$ SNPs shared between the two (90K just in those two (below), plus 70K shared by all 7), and $\approx 90\text{K}$ additional positions that are het in one but not the other. These are close to, but not exactly equal to, the 1:2:1 ratios we would naively expect from two samples of a single HWE population. The most parsimonious explanation seems to be that the H-clade is sexual, but perhaps some het positions private to each population separates them.

Aside: counts of “consistent” SNPs minus these singletons yeilds count of shared SNPs:

```
singlets <- apply(pat.summaries[pat.summaries$sharedBy==1,10:13],2,sum)
rbind(consistent=consistent.count,singlets=singlets,shared=consistent.count-singlets)

#           count1 count2 count3 count4
# consistent 447177 469906 471171 474613
# singlets   159370 183788 189899 231359
# shared     287807 286118 281272 243254
```

The slightly higher count of shared positions in the medium case further supports this choice for subsequent analysis.

Next look at consistent SNPs shared between just a pair of isolates.

```
showgroup(pat.summaries,2) # chr 1 counts: 7641 9549 9472 6924
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 4	003	2						X	X	994	298	532	587
# 6	005	2					X		X	287	523	1088	1407
# 7	006	2					X	X		624	138	282	590
# 10	011	2				X			X	515	486	317	827
# 11	012	2				X		X		565	49	32	93
# 13	014	2				X	X			133	139	132	417
# 18	021	2			X				X	998	167	337	402
# 19	022	2			X			X		82160	87499	83482	58009
# 21	024	2			X		X			686	195	410	625
# 25	030	2			X	X				609	69	47	93
# 34	041	2		X					X	42	92	313	368
# 35	042	2		X				X		503	119	254	394
# 37	044	2		X			X			69	279	1001	1809
# 41	050	2		X		X				13	24	53	105
# 49	060	2		X	X					627	116	237	388
# 66	101	2	X						X	29	47	73	314
# 67	102	2	X					X		351	67	96	351
# 69	104	2	X				X			39	122	329	1196
# 73	110	2	X			X				12	11	29	150
# 81	120	2	X		X					432	76	98	309
# 97	140	2	X	X						955	1144	1235	2144
# Total										90643	91660	90377	70578

I.e., of the 91660 paired SNPs, 87499 or 95.5% are found between Italy and Wales, with comparatively few shared between any other pairs (only).

SNPs shared among exactly 3 isolates are relatively rare. (The 5 trios containing both Italy and Wales predominate in the loose set, probably because they share many pairs that become triples with the addition of a few read errors.)

```
showgroup(pat.summaries,3) # chr 1 counts: 1438 294 671 1034
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 8	007	3					X	X	X	104	197	371	557
# 12	013	3				X		X	X	257	226	146	338
# 14	015	3				X	X		X	1041	984	660	1389
# 15	016	3				X	X	X		46	60	49	152
# 20	023	3			X			X	X	1274	558	1020	533
# 22	025	3			X		X		X	135	216	466	522
# 23	026	3			X		X	X		793	431	763	789
# 26	031	3			X	X			X	268	233	151	361
# 27	032	3			X	X		X		698	131	74	86
# 29	034	3			X	X	X			103	119	91	219
# 36	043	3		X				X	X	56	86	151	133
# 38	045	3		X			X		X	202	593	1970	1656
# 39	046	3		X			X	X		58	154	425	604
# 42	051	3		X		X			X	52	57	74	126
# 43	052	3		X		X		X		8	13	17	22
# 45	054	3		X		X	X			20	78	133	292
# 50	061	3		X	X				X	52	80	131	115
# 51	062	3		X	X			X		703	269	454	469
# 53	064	3		X	X		X			53	184	458	601
# 57	070	3		X	X	X				22	9	14	24
# 68	103	3	X					X	X	24	34	46	143
# 70	105	3	X				X		X	78	181	396	805
# 71	106	3	X				X	X		32	66	109	377
# 74	111	3	X			X			X	6	11	8	139
# 75	112	3	X			X		X		10	11	8	26
# 77	114	3	X			X	X			12	36	56	365
# 82	121	3	X		X				X	22	22	34	73
# 83	122	3	X		X			X		501	162	165	354
# 85	124	3	X		X		X			43	88	152	400
# 89	130	3	X		X	X				9	9	9	27
# 98	141	3	X	X					X	78	149	258	519
# 99	142	3	X	X				X		386	409	463	755

# 101	144	3	X	X		X		383	1176	2395	4432
# 105	150	3	X	X		X		28	51	55	238
# 113	160	3	X	X	X			337	375	399	712
# Total								7894	7458	12171	18353

Four-way sharing is more common, but dominated by the coastal (i.e., non-Gyre) L-clade isolates. This is likely a reflection of the strong 5-way sharing among the L-clade, from which the Gyre commonly drops out due to the lower coverage/higher error rate in that sequencing run.

showgroup(pat.summaries,4) # chr 1 counts: 564 1346 2552 3479

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 16	017	4				X	X	X	X	390	329	211	564
# 24	027	4			X		X	X	X	461	759	1423	771
# 28	033	4			X	X		X	X	1139	973	574	306
# 30	035	4			X	X	X		X	578	509	329	503
# 31	036	4			X	X	X	X		345	320	227	211
# 40	047	4		X			X	X	X	127	256	708	708
# 44	053	4		X		X		X	X	35	34	26	56
# 46	055	4		X		X	X		X	606	696	668	971
# 47	056	4		X		X	X	X		26	44	50	88
# 52	063	4		X	X			X	X	151	184	332	194
# 54	065	4		X	X		X		X	122	284	731	582
# 55	066	4		X	X		X	X		217	489	1025	851
# 58	071	4		X	X	X			X	9	20	7	28
# 59	072	4		X	X	X		X		41	36	21	31
# 61	074	4		X	X	X	X			20	46	51	116
# 72	107	4	X				X	X	X	58	84	129	330
# 76	113	4	X			X		X	X	7	9	5	66
# 78	115	4	X			X	X		X	141	139	122	604
# 79	116	4	X			X	X	X		8	8	11	101
# 84	123	4	X		X			X	X	63	98	91	124
# 86	125	4	X	X			X		X	67	113	223	283
# 87	126	4	X		X		X	X		99	198	268	425
# 90	131	4	X		X	X			X	6	3	0	52
# 91	132	4	X		X	X		X		19	21	10	38
# 93	134	4	X		X	X	X			18	17	22	143
# 100	143	4	X	X				X	X	37	58	103	190
# 102	145	4	X	X			X		X	5992	12644	22332	23189
# 103	146	4	X	X			X	X		196	510	969	1795
# 106	151	4	X	X		X			X	43	69	54	220
# 107	152	4	X	X		X		X		18	27	15	67
# 109	154	4	X	X		X	X			1227	1390	1065	1738
# 114	161	4	X	X	X				X	74	96	113	207
# 115	162	4	X	X	X			X		1848	1932	1828	1014
# 117	164	4	X	X	X		X			237	627	1053	1752
# 121	170	4	X	X	X	X				13	18	15	69
# Total										14438	23040	34811	38387

Five-way sharing is much more common, and is strongly dominated by the 5 L-clade isolates.

showgroup(pat.summaries,5) # chr 1 counts: 3969 5047 4624 6125

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 32	037	5			X	X	X	X	X	2247	1877	1193	620
# 48	057	5		X		X	X	X	X	221	219	189	324
# 56	067	5		X	X		X	X	X	556	1015	2544	1151
# 60	073	5		X	X	X		X	X	94	72	62	38
# 62	075	5		X	X	X	X		X	195	187	210	328
# 63	076	5		X	X	X	X	X		106	130	124	128
# 80	117	5	X			X	X	X	X	48	32	25	241
# 88	127	5	X		X		X	X	X	225	321	501	482
# 92	133	5	X		X	X		X	X	31	28	15	52
# 94	135	5	X		X	X	X		X	126	117	88	235
# 95	136	5	X		X	X	X	X		34	56	26	106
# 104	147	5	X	X			X	X	X	2073	4042	6741	10001

# 108	153	5	X	X		X		X	X	39	27	26	96
# 110	155	5	X	X		X	X		X	40157	35344	22417	30602
# 111	156	5	X	X		X	X	X		565	575	410	735
# 116	163	5	X	X	X			X	X	255	271	328	316
# 118	165	5	X	X	X		X		X	2726	5022	8440	9715
# 119	166	5	X	X	X		X	X		902	1976	3172	2688
# 122	171	5	X	X	X	X			X	41	19	13	70
# 123	172	5	X	X	X	X		X		58	71	45	86
# 125	174	5	X	X	X	X	X			659	682	468	782
# Total										51358	52083	47037	58796

Six-way sharing is also common, with the sets *excluding* Gyre, Italy, or Wales having the most mutually-shared SNPs.

```
showgroup(pat.summaries,6) # chr 1 counts: 4166 4741 5312 4722
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 64	077	6		X	X	X	X	X	X	850	847	827	485
# 96	137	6	X		X	X	X	X	X	405	324	240	333
# 112	157	6	X	X		X	X	X	X	13239	10814	6862	12202
# 120	167	6	X	X	X		X	X	X	11742	21003	35227	15091
# 124	173	6	X	X	X	X		X	X	131	87	47	114
# 126	175	6	X	X	X	X	X		X	16884	13630	8608	12697
# 127	176	6	X	X	X	X	X	X		2422	2412	1566	1032
# Total										45673	49117	53377	41954

8 Trees

So, overall, the picture looks like a long shared history (62182 7-way shared positions), followed by a split of the 5 L-isolates from the 2 H-isolates, then a long shared history in the 5 (35344 quintuples), in parallel with a long shared history in H- (87499 pairs), then separate histories in Italy and Wales (>85117 “private” SNPs in each, although again if they are sexual, many of these just reflect HWE), and very limited differentiation among the 5 L-isolates.

Branch lengths of course depend on filtering criteria used (and, of course, full vs Chr1 differ by about a factor of 10), but the tree *topology* appears to be fairly stable. Various versions are drawn below, exactly to explore how robust this story is. I think we should go with “medium stringency” SNP filtering (based on un-qfiltered reads).

NOTE: Much of this analysis make less sense for q-filtered read data, since (a) the point of the SNP filtering was to try to correct for noise in the raw reads, which may (or may not; haven’t looked closely, yet) be largely fixed by qfiltering (e.g., “loose” or no SNP filtering may be more appropriate, post-q-filtering, esp. if we had re-run SAMTools to call SNPs based on the q-filtered reads), and (b) tree topology *does* appear to change, in that Gyre’s coverage has been so sharply reduced by qfiltering that it clearly stands aside from the others (and that’s confirmed by bootstrap), but this also seems to be clearly a technical rather than a biological artifact. SO, code below will run on q-filtered data, but *is not tuned to it*. Likewise, most comments in the prose below were made to describe the un-q-filtered data, and *are misleading and in some cases flatly wrong* for qfiltered data, but it doesn’t seem worthwhile to bother with a rewrite...

Trees are coded in newick format, which doesn’t seem to tolerate line-breaks; print with line-wrap:.

```
# wrap a long char string across multiple lines in printout
cat.hardwrap <- function(str,width=80){
  while(nchar(str)>width){
    cat(substr(str,1,width),'\n')
    str <- substr(str,width+1,nchar(str))
  }
  cat(str,'\n')
}
```

Trees are built as follows. Code for drawing, especially, is specific to the topology of the medium tree, and placement of some of the figure elements have been hand-optimized for this case; drawings for the other variants will not be as pretty.

```

# set up for tree figs

# the newick parser in ape seems to be confused by commas and parens in
# tip names, and blanks are not allowed, so replace by *, <, >, _, resp.
newick.name <- function(name){
  name <- gsub(' ', '_', name, fixed=TRUE)
  name <- gsub(',', '*', name, fixed=TRUE)
  name <- gsub('(', '<', name, fixed=TRUE)
  name <- gsub(')', '>', name, fixed=TRUE)
  return(name)
}

# undo above changes
newick.name.undo <- function(name){
  #name <- gsub('_', ' ', name, fixed=TRUE) # unnecessary; ape plot routine handles this one
  name <- gsub('*', ',', name, fixed=TRUE)
  name <- gsub('<', '(', name, fixed=TRUE)
  name <- gsub('>', ')', name, fixed=TRUE)
  return(name)
}

# make a newick string from tree; see it below
# 'pre' is prefixed to ccmpid; 'nb' optionally included;
# 'alt' can be used instead of pre/ccmp/nb/where for less formal labeling
# 'newstyle'=T => new node label: [nb_]where[(pre-less-id)]
# 'newstyle'=F => old node label: [nb_][pre id]where
newickize <- function(tree,pre='CCMP',nb=TRUE,alt=F,newstyle=TRUE){
  if(is.null(tree$where)){
    # not a leaf; paste together newick from subtrees
    sub1 <- newickize(tree$sub1,pre=pre,nb=nb,alt=alt,newstyle=newstyle)
    sub2 <- newickize(tree$sub2,pre=pre,nb=nb,alt=alt,newstyle=newstyle)
    new <- paste('(', sub1, ',', sub2, ')', sep='')
    if(!is.null(tree$length)){
      # internal node, add length
      return(paste(new, ':', tree$length, sep=''))
    } else {
      # top level; escape blanks and add trailing ';'
      return(paste(gsub(' ', '_', new), ';', sep=''))
    }
  } else {
    # a leaf; build label and branch length
    if(alt){
      # label is just alt; if alt omitted, default to where
      new <- newick.name(ifelse(is.null(tree$alt), tree$where, tree$alt))
    } else {
      if(newstyle){
        # new node label = [nb_]where[(pre-less-id)]
        new <- ifelse(nb && !is.null(tree$nb), paste(tree$nb, '_', sep=''), '')
        new <- newick.name(paste(new, tree$where, sep=''))
        new <- ifelse(is.null(tree$id), new, paste(new, '_', tree$id, ')', sep=''))
        new <- newick.name(new)
      } else {
        # old style node label = [nb_][pre id]where
        new <- ifelse(nb && !is.null(tree$nb), paste(tree$nb, '_', sep=''), '')
        new <- ifelse(is.null(tree$id), new, paste(new, pre, tree$id, '_', sep=''))
        new <- newick.name(paste(new, tree$where, sep=''))
      }
    }
    #add length to either
    new <- paste(new, ':', tree$length, sep='')
  }
  return(new)
}

# Make a tree as nested lists, **based on the chr1, count2 topology**, but using any of the counts.
# Internal nodes have subtrees sub1/2 and length
# Root has sub1/2, but no length
# Leaves have where, length, optionally, id, alt, nb. (Omit id for 'outgroup'. Use 'alt' for less formal
# labeling in cartoon version; it defaults to 'where'. Use 'nb' to add abcde annotations for legend.)
# The single parameter v is any of the 4 count vectors contained in pat.summaries (most conveniently
# indexed in octal). E.g., make.tree(pat.summaries['count2']) reproduces the count2 tree.
# (This was previously built by hand-pasting the edge lengths; tree.by.hand is retained in appendix
# for comparison, & its counts are in comments below).
#
make.tree <- function(v){
  pat.count <- function(pat, pat.counts=v){return(pat.counts[1+strtoi(pat,8)])}
  thetree <-
    list(
      sub1 = list(
        sub1 = list(
          sub1 = list(id=3367, length=pat.count('002'), where='Venice, Italy', alt='Venice'), #8813

```

```

sub2 = list(id=1013, length=pat.count('020'), where='Wales, UK'), #9652
length=pat.count('022')), #9365
sub2 = list(
  sub1 = list(
    sub1 = list(
      sub1 = list(id=1007, length=pat.count('100'), nb='e', where='Virginia, USA'), #30
      sub2 = list(id=1012, length=pat.count('040'), nb='d', where='Perth, W. Australia', alt='Perth'), #61
      length=pat.count('140')), #19
    sub2 = list(
      sub1 = list(id=1015, length=pat.count('004'), nb='c', where='Washington, USA', alt='Puget Sound'), #207
      sub2 = list(id=1335, length=pat.count('001'), nb='b', where='New York, USA', alt='NY'), #41
      length=pat.count('005')), #18
      length=pat.count('145')), #1005
      sub2 = list(id=1014, length=pat.count('010'), nb='a', where='N. Pacific Gyre'), #61
      length=pat.count('155')), #3912
      length=pat.count('177')), #7054
    sub2 = list(length=0, where='outgroup')
  )
)
return(thetree)
}

```

Code to plot a tree given newick description. Again, code is somewhat general, but has some specializations tied to the medium-stringency, full-genome, un-qfiltered data.

```

# run following 2 lines after an R upgrade
# update.packages()
# install.packages("ape")
library(ape)
show.tree <- function(newick.str=newick.medium,
  col.edge = 'darkblue', lwd.edge = 2,
  col.elabel='darkblue',
  col.arrow = 'red', lwd.arrow=1.5, cex.elabel=0.8, font.elabel=3,
  col.clade = 'black', lwd.clade=1, cex.clade = 1.0, font.clade = 3,
  col.legend='beige', cex.legend=0.8,
  col.tip = 'darkblue', font.tip = 4,
  plusx=FALSE, pltdebug=FALSE, total.snps=consistent.count[2],
  straight.arrow=FALSE){

  ####
  #
  # ADJUST NEWICK & GET LENGTHS, COORDINATES
  #
  newick.str.noout <- sub('outgroup','_',newick.str) # Hide outgroup ('_' prints as blank)
  the.tree <- read.tree(text=newick.str.noout)

  ## nasty hack: ape's newick parser seems to be confused by commas, () in tip labels, so
  ## newickize replaced them by '*<>'; before plotting, I want to convert them back, and hope
  ## this doesn't break anything else... And if a revised version of ape changes the internal
  ## representation of a tree, this may need to be redone.
  the.tree$tip.label <- newick.name.undo(the.tree$tip.label)

  # extract branch lengths as char string of comma-separated numbers via pattern matching hack:
  # lengths always preceded by colon
  lengths.ch <- strsplit(paste(newick.str,':'),'[^0-9][^:]*:')[[1]]

  # then convert to ints, dropping empty string at front
  lengths.int <- scan(what=integer(),quiet=T,sep=',',text=lengths.ch[-1])

  # then to data frame with named rows; a..g are terminal branches; others are internal.
  # a..e match legend in plot; f/g = wales/italy. lengths appear in postfix order of
  # newick tree, and ape draws the 1st of them at the bottom of the plot.
  lmed <- data.frame(lengths=lengths.int,
    row.names=c('g','f','fg','e','d','de','c','b','bc','bcde','a','abcde','all','out'))

  # extract counts needed for legend:
  #leg.counts <- c( 61, 41,207, 61, 30, 1005, 18, 19) #by hand, medium chr1
  leg.counts <- lmed[c('a','b','c','d','e','bcde','bc','de'),1]
  discord <- total.snps - sum(lmed$lengths)

  #tree.labels <- list( ## x,y,text; coords are all picked by eye
  # 3000, 3.62, paste(lmed['all',1], 'shared by 7', sep='\n'), # 7054
  # 8900, 5.75, paste(lmed['abcde',1], 'by 5', sep='\n'), # 3912
  # 12000, 1.50, paste(lmed['fg',1], 'shared by 2', sep='\n'), # 9365
  # 21000, 2.00, paste(lmed['f',1], 'only\nin Wales'), # 9652
  # 21000, 1.00, paste(lmed['g',1], 'only\nin Italy'), # 8813
  # 11500, 4.50, '*')
  # automating x-placement, below; retain above for comparison...
  tip <- integer(7) # x coords of tree tips
  tip[1] <-sum(lmed[c('all','fg','g'),1])

```



```

tip[2] <-sum(lmed[c('all','fg','f'),1])
tip[3] <-sum(lmed[c('all','abcde','bcde','de','e'),1])
tip[4] <-sum(lmed[c('all','abcde','bcde','de','d'),1])
tip[5] <-sum(lmed[c('all','abcde','bcde','bc','c'),1])
tip[6] <-sum(lmed[c('all','abcde','bcde','bc','b'),1])
tip[7] <-sum(lmed[c('all','abcde','a'),1])

inode <- integer(5) # x coords of (some) internal nodes
inode[1] <- 0 # root
inode[2] <- lmed['all',1] # lca of all
inode[3] <- sum(lmed[c('all','fg'),1]) # lca H-clade
inode[4] <- sum(lmed[c('all','abcde'),1]) # lca L-clade
inode[5] <- sum(lmed[c('all','abcde','bcde'),1]) # lca L-clade, nonGyre
tree.labels <- list( ## x,y,text; y coords partially picked by eye
  sum(inode[c(1,2)])/2, 3.62, paste(lmed['all',1], 'shared by 7', sep='\n'), # 7054
  sum(inode[c(2,4)])/2, 5.75, paste(lmed['abcde',1], 'by 5', sep='\n'), # 3912
  sum(inode[c(2,3)])/2, 1.50, paste(lmed['fg',1], 'shared by 2', sep='\n'), # 9365
  (inode[3]+tip[2])/2, 2.00, paste(lmed['f',1], 'only\nin 1013'), # 9652
  (inode[3]+tip[1])/2, 1.00, paste(lmed['g',1], 'only\nin 3367'), # 8813
  sum(inode[c(4,5)])/2, 4.35, '* ')

tree.labels <- list( ## x,y,text; y coords partially picked by eye
  sum(inode[c(1,2)])/2, 3.62, paste(lmed['all',1], 'in 7', sep='\n'), # 7054
  sum(inode[c(2,4)])/2, 5.75, paste(lmed['abcde',1], 'in 5', sep='\n'), # 3912
  sum(inode[c(2,3)])/2, 1.50, paste(lmed['fg',1], 'in 2', sep='\n'), # 9365
  (inode[3]+tip[2])/2, 2.00, paste(lmed['f',1], 'only\nin 1013'), # 9652
  (inode[3]+tip[1])/2, 1.00, paste(lmed['g',1], 'only\nin 3367'), # 8813
  sum(inode[c(4,5)])/2, 4.35, '* ')

####
#
# BOGUS PLOT
#
# a messy bit: need string widths to set xlim; but strwidth needs x-scale so must plot first.
# M plot completely invisible, overlay 2nd plot via par(new=F...) .
#
# PROVISIONALLY set x.lim here at about 30% wider than tree; fine tune it for the real plot
# based on strwidth(tip labels) below.
#
provisional.tree.x.lim <- 1.3 * max(tip) # <= PROVISIONAL plot width
plot(0,0, type='n', bty='n', xaxt='n', yaxt='n', xlab='', ylab='', xlim=c(0,provisional.tree.x.lim), ylim=c(0,7))

tiplabel.x <- integer(7)
for(i in 1:7){
  # see warning above about internals of the tree; labels have '_', printed as ' '.
  tiplabel.x[i] <- tip[i]+strwidth(gsub('_', ' ', the.tree$tip.label[i], fixed=T), font=font.tip)
}

# visually show tip coords & max x to debug placement issues
plt.debug <- function(tree.x.lim, tip, tiplabel.x, spx=NULL, spy=NULL){
  if(pltdebug){ # F to hide/T to show debug
    cat('Tip labels:', paste(the.tree$tip.label, sep='', collapse='/'), '\n')
    axis(2) # useful only for placing labels
    for(i in 1:7){
      points(c(tip[i], tiplabel.x[i]), c(i,i)) # debug: do I have right tip coordinates?
    }
    lines(rep(tree.x.lim,2), c(0,7)) # where is right edge?
    if(!is.null(spx)){
      points(spx, spy) # show spline control points, for tweaking
    }
  }
}

plt.debug(provisional.tree.x.lim, tip, tiplabel.x)

label.end.H <- max(tiplabel.x[1:2])
label.end.L <- max(tiplabel.x[3:7])
clade.dx <- strwidth('x') # space between clade marker line and its label
xdel <- 3*clade.dx # space between labeled clade tips and marker line

tree.x.lim <- 1.03*(max(tiplabel.x)+xdel) # <= FINAL plot width
tree.y.lim <- 7
if(pltdebug){cat('Plot width hacking:', provisional.tree.x.lim, tree.x.lim, tree.x.lim/1.03/max(tip), clade.dx)}

par(new=T) # I.e., NOT starting a new plot

####
#

```

```

# REAL PLOT
#
plot(the.tree,
     x.lim = c(0, tree.x.lim),
     y.lim = c(0, tree.y.lim),
     font=font.tip, label.offset=100,          # bold-italic, nudged slightly right
     tip.color=col.tip, edge.color=col.edge,
     edge.width=lwd.edge,
     edge.lty=c(1,1,1,1, 1, 1,1,1,1,1,1,0)    # 5th is bottleneck edge; 14th is outgroup
)
lines(00+c(0,0),c(3.5,6),col='white',lwd=6)    # Hide vertical line to outgroup
axis(1, pos=0.25, at=seq(0,25,by=5)*10^round(log10(max(tip)/25)))

if(pltdebug){text(tip[1]+100, 1.0, 'Venice, Italy (3367)', adj=0, font=font.tip)}

####
#
# BOTTLENECK ANNOTATION
#
# spline/ellipse control points (spy/y) & tweaks thereto (dx/y)
dx <- 0.01 * tree.x.lim
dy <- .04
spx <- c(7400, 7400, 9900, 10500) # by eye, chr1, for comparison
spx <- c(inode[2]+dx,inode[2]+dx,inode[4]-3*dx,inode[4]-dx)
spy <- c( 3.8,  3.9,  5.6-dy,  5.6-dy)

plt.debug(tree.x.lim, tip, tiplabel.x, spx, spy)

if(T){
  #ellipse version, defined by rect thru 2 middle pts of spx/y
  spf<-function(x){
    ifelse(x <= spx[2], spy[1],
           ifelse(x >= spx[3], spy[4],
                  spy[2]+(spy[3]-spy[2])*sqrt(pmax(0,1-((x-spx[3])/(spx[3]-spx[2]))^2))))
  }
} else {
  # spline version
  spf <- splinefun(spx,spy,method='hyman')
}
serx <- seq(spx[1],spx[length(spx)],length.out=50)
sery <- spf(serx)
tailx <- spx[1]
taily <- spy[1]
headx <- spx[4]
heady <- spy[4]
textx <- (headx+tailx)/2+(headx-tailx)*(-.01)
texty <- (heady+taily)/2+(heady-taily)*(-.10)
bottle.txt <- "inbreeding\nLoH / LoS"
if(!straight.arrow){
  arrows(headx,heady,headx+tree.x.lim*1e-3,heady, length=.1,col=col.arrow,lwd=lwd.arrow)
  lines(rev(serx), rev(sery), lty=c(5,1),col=col.arrow, lwd=lwd.arrow)
  textangle <- 66
  textadj <- c(0,0)
} else {
  # Tweak from 7-node to base of L-clade;
  # center text, rotated, on diagonal towards L-clade; ditto the straight arrow.
  llx <- inode[2] # the aforementioned rectangle
  urx <- inode[4]
  lly <- 3.62
  ury <- 5.75
  # rect(llx,lly,urx,ury) # show rect for debug
  textx <- (llx+urx)/2 # center text
  texty <- (lly+ury)/2
  textangle <- atan(grconvertY(ury-lly,to='dev')/grconvertX(urx-llx,to='dev'))*360/(2*pi)
  textadj <- c(0.50, 0.43) #tweak position; ".5" = center in x , ".43" raises, THEN rotate.
  alpha <- .78 # fraction along diag at which arrow begins
  beta <- .95 # ... and ends
  arrows((1-alpha)*llx + alpha*urx,
         (1-alpha)*lly + alpha*ury,
         (1-beta)*llx + beta*urx,
         (1-beta)*lly + beta*ury, length=.1,col=col.arrow,lwd=lwd.arrow,angle=25)
}
if(T){
  text(textx, texty, bottle.txt, srt=textangle, font=font.arrow, cex=cex.arrow,
       col=col.arrow, adj=textadj)
} else {
  # experiment at wrapping text along curved path; unpretty, but retain for now, maybe revisit
  bottlec <- strsplit(bottle,split=NULL)[[1]]
  for(i in 1:length(bottlec)){
    text(xser[i],yser[i],bottlec[i], srt=65, font=4, cex=.7, col=col.arrow)
  }
}

```

```

}
}

####
#
# CLADE ANNOTATION
#
clade.L.x <- label.end.L + xdel
clade.H.x <- label.end.H + xdel
dy <- .33
lines(rep(clade.L.x,2),c(3-dy,7+dy),lwd=lwd.clade,col=col.clade)
lines(rep(clade.H.x,2),c(1-dy,2+dy),lwd=lwd.clade,col=col.clade)
text(clade.L.x+clade.dx,5.0,'L-clade',srt=90,font=font.clade,cex=cex.clade,col=col.clade)
text(clade.H.x+clade.dx,1.5,'H-clade',srt=90,font=font.clade,cex=cex.clade,col=col.clade)

####
#
# LEGEND
#
# parameter plusx controls whether we try to annotate b/c (+) and d/e (x) sharing in tree; I think
# it looks cluttered, rather than adding clarity, so I vote no, but code is here, in case. "Logic,"
# if any, for my symbol choice is that + overlaid on x looks like the * at the next level; this
# analogy is more visible if we use pch 3/4/8 rather than Courier or Helvetica chars, but probably
# should use same in both tree & legend, which will take a modicum of additional work.
legend.text <- c('a: only in 1014 ',
                'b: only in 1335 ',
                'c: only in 1015 ',
                'd: only in 1012 ',
                'e: only in 1007 ',
                '*: shared by bcde',
                paste(ifelse(plusx,'+:',' '), 'shared by b/c '),
                paste(ifelse(plusx,'x:',' '), 'shared by d/e ')
)

legend.text <- c('a: only in 1014 ',
                'b: only in 1335 ',
                'c: only in 1015 ',
                'd: only in 1012 ',
                'e: only in 1007 ',
                '*: in bcde ',
                paste(ifelse(plusx,'+:',' '), 'in bc '),
                paste(ifelse(plusx,'x:',' '), 'in de '),
                'Discordant SNPs '
)

legend.text <- paste(legend.text,format(c(leg.counts,discord),width=4),sep=' - ')
legend.text <- paste(legend.text,' ') # add a little more right margin in box
opar <- par(family="mono",cex=cex.legend)
legend('topright', legend=legend.text, cex=cex.legend, inset=c(0.05,0), bg=col.legbox, box.col=col.legbox)
par(opar)
if(plusx){
  points(tree.labels[[16]],tree.labels[[17]]+.14,pch=8,col=col.elabel)
  points(tree.labels[[16]]+200,tree.labels[[17]]+1,pch=3,col=col.elabel)
  points(tree.labels[[16]]+200,tree.labels[[17]]-1,pch=4,col=col.elabel)
}

####
#
# EDGE LENGTHS
#
for(i in seq(1,length(tree.labels)-ifelse(plusx,5,2),by=3)){
  if(F){ # T for \n in edge labels; F to remove (except "by 5")
    text(tree.labels[[i]], tree.labels[[i+1]], tree.labels[[i+2]])
  } else {
    # points(tree.labels[[i]], tree.labels[[i+1]], pch=3,col='green') # for debugging
    text(tree.labels[[i]], tree.labels[[i+1]], sub('\n([z])',' \\1', tree.labels[[i+2]]),
         pos=3, offset=.4, font=font.elabel, col=col.elabel,cex=cex.elabel)
  }
}
}
if(FALSE){#for debug convenience
pdf(paperfig.path, width=8,height=5,onfile=TRUE,family='Helvetica',fonts='Courier',pointsize=10)
show.tree(newick.medium, total.snps=consistent.count[2], pltdebug=F, straight.arrow=T)
dev.off()
}

caption <- function(stringency,which.tables=which.snp.tables(string.val=F)){
  caption.where <- '(UNKNOWN genome subset).'
  if(which.tables[1]=='Chr1') {caption.where <- 'on Chr1.'}
}

```

```

if(which.tables[1]=='full') {caption.where <- 'genome-wide.'}
if(which.tables[1]=='trunc'){caption.where <- 'all Chrs.'}
cap.stringency <- c(
  'loose SNP filters.',
  'medium SNP filters.',
  'strict SNP filters.',
  'unfiltered SNPs.')
cap <- paste('Tree based on', which.tables[2], 'reads and', cap.stringency[stringency],
  '``Lengths\\`\\` are numbers of shared/private SNPs', caption.where)

return(cap)
}

```

Trees based on all four SNP filtering criteria are shown below. Their topologies are exactly the same, although the branch lengths are different. In all four, the length of the branch labeled “*” is probably inflated by lower coverage and higher error rate in 1014, which may mask further legitimate sharing between it and the other L-isolates. The branch lengths among the other 4 are too short for their topology to be convincing without a more rigorous analysis (e.g., a bootstrap test), but detail there is irrelevant to the story.

My sense is that the “medium” version is the best for the paper, made here and shown in Fig 1. In theory, this should look exactly like Fig 3, but something is apparently different between Knitr and direct-to-pdf. (Increasing fig.width in Knitr’s chunk headers from 8 (as in the pdf call below) to 9 helps somewhat, but probably still best to make the paper fig directly rather than via Knitr.)

```

###
#
# MAKE PROTOTYPE PDF FOR PAPER, *AND* SAVE DATA NEEDED TO BUILD IT
#
w.s.t. <- which.snp.tables()
if(w.s.t. == 'trunc-unfiltered'){
  rda.Description <- 'This .rda contains data to generate Fig 3; see shared.snps.rnw for details.'
  save(rda.Description, w.s.t., pat.summaries, consistent.count, file='Fig3-data.rda')
  paperfig.path <- paste('figs-mine/paperfig-medium-tree-', w.s.t., '--Fig3proto.pdf', sep='')
} else {
  paperfig.path <- paste('figs-mine/paperfig-medium-tree-', w.s.t., '.pdf', sep='')
}
pdf(paperfig.path, width=8,height=5,onefile=TRUE,family='Helvetica',fonts='Courier',pointsize=10)
newick.medium <- newickize(make.tree(pat.summaries[, 'count2']))
show.tree(newick.medium, total.snps=consistent.count[2], pltdebug=F, straight.arrow=T)
dev.off()

# pdf
# 2

```

```

# fig.paths for knitr chunks below; .h for "hand-made" trees; plain for automatic chr1/full versions
myfigpath <- paste(getwd(), '/figs-knitr/newick-', which.snp.tables(), '-', sep='')
myfigpath.h <- paste(getwd(), '/figs-knitr/newick-', sep='')

```

Figure 2, i.e., criteria [[1]]:

```

newick.loose <- newickize(make.tree(pat.summaries[, 'count1']))
show.tree(newick.loose, total.snps=consistent.count[1])

```

Figure 3, i.e. [[2]]:

```

# newick.medium <- newickize(tree.by.hand)
# simple.newick.medium <- newickize(tree.by.hand, alt=TRUE)
newick.medium <- newickize(make.tree(pat.summaries[, 'count2']))
simple.newick.medium <- newickize(make.tree(pat.summaries[, 'count2']), alt=TRUE)
show.tree(newick.medium, total.snps=consistent.count[2])

```

Figure 4, i.e. [[3]]:

```

newick.strict <- newickize(make.tree(pat.summaries[, 'count3']))
show.tree(newick.strict, total.snps=consistent.count[3])

```

Figure 5, i.e. [[4]]:

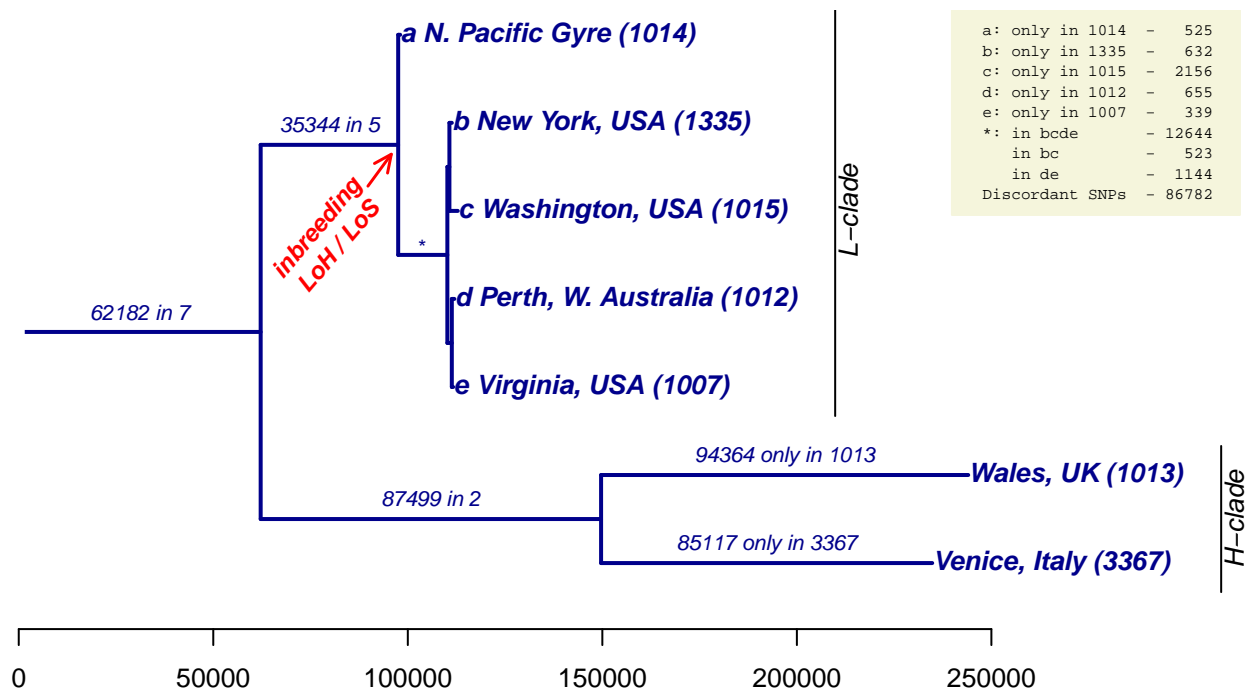


Figure 1: Proposed fig. for paper: Tree based on qfiltered reads and medium SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs.

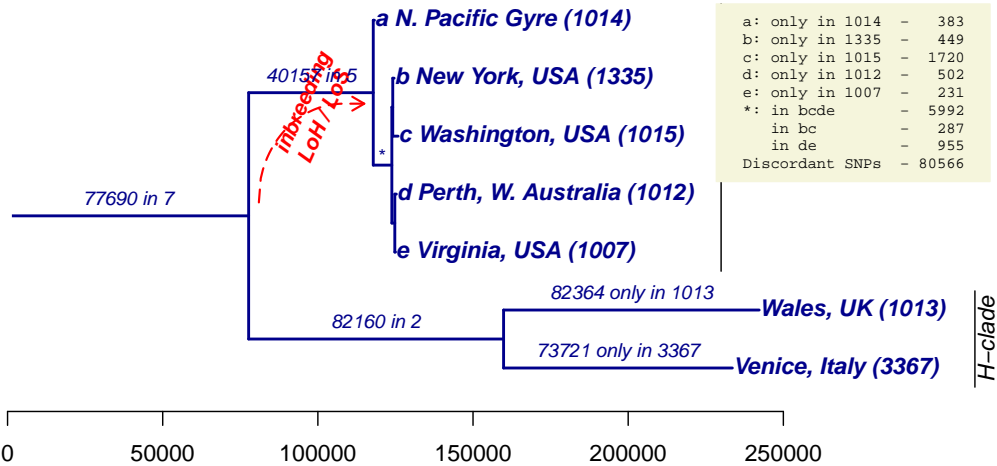


Figure 2: Tree based on qfiltered reads and loose SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs.

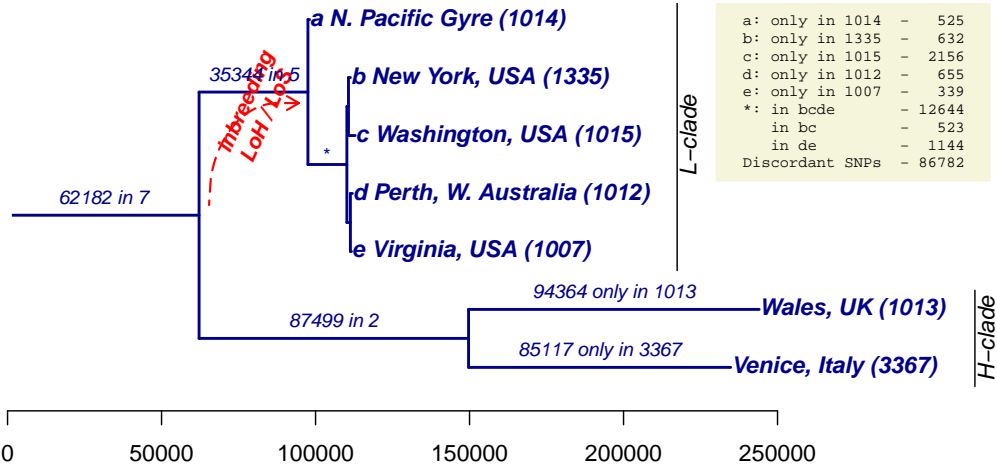


Figure 3: Tree based on qfiltered reads and medium SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs.

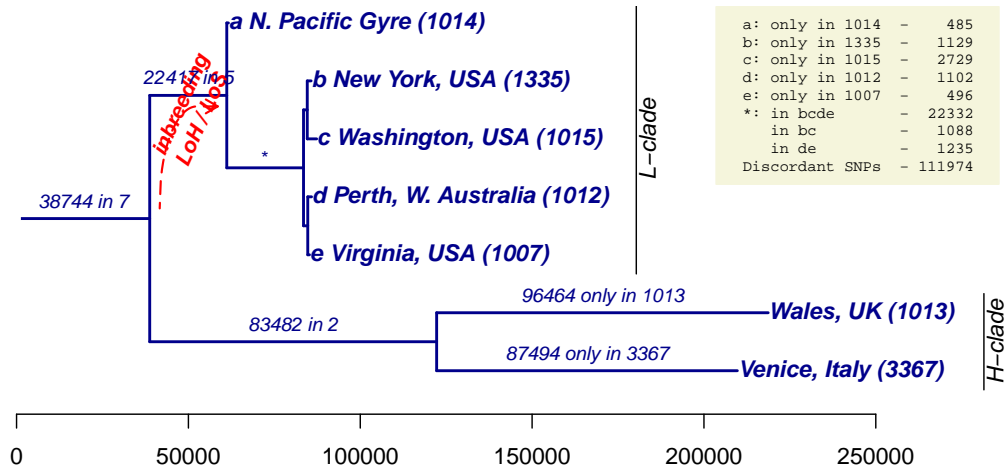


Figure 4: Tree based on qfiltered reads and strict SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs.

```
newick.unfiltered <- newickize(make.tree(pat.summaries[, 'count4']))
show.tree(newick.unfiltered, total.snps=consistent.count[4])
```

Some other versions of the trees are included in the appendix.

Counts for all tree edges in the medium tree:

```
#pat.summaries[c(128,110,102,6,97,19,9,2,5,33,65,17,3),]
tree.edges <- c(128,110,102,6,97,19,9,2,5,33,65,17,3)-1
non.edges <- setdiff(0:127, tree.edges)
sg.edges <- showgroup(restrict.to=tree.edges) ; sg.edges
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 2	001	1							X	449	632	1129	2260
# 3	002	1						X		73721	85117	87494	105614
# 5	004	1					X			1720	2156	2729	4608
# 6	005	2					X		X	287	523	1088	1407
# 9	010	1				X				383	525	485	1231
# 17	020	1		X						82364	94364	96464	113191
# 19	022	2			X			X		82160	87499	83482	58009
# 33	040	1		X						502	655	1102	2450
# 65	100	1	X							231	339	496	2005
# 97	140	2	X	X						955	1144	1235	2144
# 102	145	4	X	X			X		X	5992	12644	22332	23189
# 110	155	5	X	X		X	X		X	40157	35344	22417	30602
# 128	177	7	X	X	X	X	X	X	X	77690	62182	38744	15186
# Total										366611	383124	359197	361896

Counts for the top 10 discordant patterns, i.e., SNPs whose sharing pattern does not match any of the bifurcations in the tree:

```
tenth <- sort(showgroup(restrict.to=non.edges)[- (length(non.edges)+1), 'count2'], decreasing=T)[10]
sg.non.edges <- showgroup(restrict.to=non.edges, c2.thresh = tenth) ; sg.non.edges
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 32	037	5			X	X	X	X	X	2247	1877	1193	620
# 104	147	5	X	X			X	X	X	2073	4042	6741	10001
# 109	154	4	X	X		X	X			1227	1390	1065	1738
# 112	157	6	X	X		X	X	X	X	13239	10814	6862	12202



And percent of discordant SNPs:

In short, the sharing pattern observed at 86782 or 18.5% of the 469906 medium-stringency consistent SNPs positions observed across all 7 isolates are discordant with the medium tree. (The strict tree has slightly more.)

A majority of the discordant SNPs fall into one of three patterns: 6-way sharing excluding Gyre (likely a technical artifact since the low coverage in Gyre reduces our power to detect SNPs there), or 6-way sharing excluding one of the two H-isolates (likely a reflection of sexuality in the H-clade—SNP positions in a population in Hardy-Weinberg equilibrium are fairly likely to be homozygous for the reference allele in a given individual).

```
third.biggest <- sort(showgroup(pat.summaries,6)[-8,'count2'],decreasing=T)[3]
big.three <- showgroup(pat.summaries,6,c2.thresh = third.biggest); big.three
```

#	Pat	ShrBy	1007	1012	1013	1014	1015	3367	1335	count1	count2	count3	count4
# 112	157	6	X	X		X	X	X	X	13239	10814	6862	12202
# 120	167	6	X	X	X		X	X	X	11742	21003	35227	15091
# 126	175	6	X	X	X	X	X		X	16884	13630	8608	12697
# Other	(4 rows	w/	c2	<	10814)			3808	3670	2680	1964
# Total										45673	49117	53377	41954


```
big.three.frac <- sum(big.three[1:3,'count2'])/discordv$count2; big.three.frac
# [1] 0.5236915
```

I.e., 52.4% of discordant SNPs fall into one of these three categories.

Out of curiosity: what is the ratio of full genome to Chr 1 branch lengths. Except for the shortest few, generally $\approx 10x$, as expected given the length of Chr 1:

```
# (vectors derived by editing Newick strings, and in that order)
print(
  c(Italy=86155, Wales=95697, IW=89598, Virg=330, Aust=632, VA=1296,
    Puget=2113, NY=658, PNY=480, four=10059, Gyre=568, five=39517, all=69526) /
  c(Italy=8813, Wales=9652, IW=9365, Virg=30, Aust=61, VA=19,
    Puget=207, NY=41, PNY=18, four=1005, Gyre=61, five=3912, all= 7054),
  digits=3)

# Italy Wales IW Virg Aust VA Puget NY PNY four Gyre five all
# 9.78 9.91 9.57 11.00 10.36 68.21 10.21 16.05 26.67 10.01 9.31 10.10 9.86

round(genome.length.constants()$genome.length.trunc / genome.length.constants()$chr1.length, digits=4)

# [1] 10.2879
```

9 Semi-Automated Tree-Building

Slightly formalizing the process above: Look for the bifurcation of the 7 strains that maximizes the number of shared SNPs *within* each side of the partition while minimizing the number and fraction of SNPs that are shared by subsets that include at least one strain on each side of the partition. The 2/5 split is the winner, with 6418 SNPs in conflict with that partition (16% of the 39842 SNPs not shared by all 7; Chr1 data). The runner-up places the Gyre in a group by itself (7079 = 18% in conflict).

```
treepart <- function(p.summ=pat.summaries, root=127, verbose=T, stringency='count2'){
  root.shared <- p.summ[root+1,stringency]
  df<-NULL
  for(i in 1:floor(root/2)){
    if(bitwAnd(i,root)==i && i < root-i){
      l1 <- showgroup(p.summ,subset=i,split=NULL,proper.subset=F,total=T)
      l <- l1[nrow(l1),stringency]
      r1 <- showgroup(p.summ,subset=root-i,split=NULL,proper.subset=F,total=T)
      r <- r1[nrow(r1),stringency]
      c1 <- showgroup(p.summ,subset=root,split=i,proper.subset=T,total=T)
      c <- c1[nrow(c1),stringency]
      df <- rbind(df, data.frame(pat=i,left=l,right=r,both=l+r,cross=c,all=l+r+c,ratio=c/(l+r+c),
                                best=' ',stringsAsFactors=F))
    }
  }
  df$pat<-as.octmode(df$pat)
  maxl <- which.max(df$left)
  maxr <- which.max(df$right)
  maxb <- which.max(df$both)
  minc <- which.min(df$cross)
  minr <- which.min(df$ratio)
  df$best[c(maxl,maxr,maxb,minc,minr)] <- '<'
  df$best[maxl] <- paste(df$best[maxl], 'L') # max Left
  df$best[maxr] <- paste(df$best[maxr], 'R') # max Right
  df$best[maxb] <- paste(df$best[maxb], 'B') # max Both (L+R)
  df$best[minc] <- paste(df$best[minc], 'C') # min Cross
  df$best[minr] <- paste(df$best[minr], 'O') # min ratio (Cross/(Left+Right+Cross))
  if(verbose){
    same <- all(maxl==c(maxr,maxb,minc,minr))
    cat('root:', format(as.octmode(root),width=3),
        '; shared:', root.shared,
        '. max l', format(as.octmode(df$pat[maxl]),width=3),
        ', max r', format(as.octmode(df$pat[maxr]),width=3),
```

```

        ', max both', format(as.octmode(df$pat[maxb]),width=3),
        ', min cross', format(as.octmode(df$pat[minc]),width=3),
        ', min ratio', format(as.octmode(df$pat[minr]),width=3),
        '\nAll the same?:', same,
        '\n')
    cat('\n')
  }
  return(df)
}

```

```
treepart()
```

```

# root: 177 ; shared: 62182 . max l 077 , max r 010 , max both 010 , min cross 010 , min ratio 010 .
# All the same?: FALSE
#
```

#	pat	left	right	both	cross	all	ratio	best
# 1	01	1210	289197	290407	117895	408302	0.2887446	
# 2	02	85695	179062	264757	143545	408302	0.3515658	
# 3	03	86625	105697	192322	215980	408302	0.5289712	
# 4	04	2734	279522	282256	126046	408302	0.3087078	
# 5	05	3889	274296	278185	130117	408302	0.3186783	
# 6	06	87989	100546	188535	219767	408302	0.5382462	
# 7	07	89639	98363	188002	220300	408302	0.5395516	
# 8	10	1103	332135	333238	75064	408302	0.1838443	< R B C O
# 9	11	2221	282010	284231	124071	408302	0.3038707	
# 10	12	86269	123335	209604	198698	408302	0.4866447	
# 11	13	87911	102474	190385	217917	408302	0.5337152	
# 12	14	3398	276093	279491	128811	408302	0.3154797	
# 13	15	6023	273221	279244	129058	408302	0.3160847	
# 14	16	88762	98932	187694	220608	408302	0.5403060	
# 15	17	92437	97647	190084	218218	408302	0.5344524	
# 16	20	94942	165462	260404	147898	408302	0.3622270	
# 17	21	95741	96070	191811	216491	408302	0.5302227	
# 18	22	267558	61350	328908	79394	408302	0.1944492	
# 19	23	269213	8703	277916	130386	408302	0.3193372	
# 20	24	97293	91454	188747	219555	408302	0.5377270	
# 21	25	98831	89139	187970	220332	408302	0.5396300	
# 22	26	270478	4870	275348	132954	408302	0.3256266	
# 23	27	273828	3327	277155	131147	408302	0.3212010	
# 24	30	95536	112945	208481	199821	408302	0.4893951	
# 25	31	97054	93029	190083	218219	408302	0.5344549	
# 26	32	268332	21310	289642	118660	408302	0.2906182	
# 27	33	271905	6449	278354	129948	408302	0.3182644	
# 28	34	98145	89824	187969	220333	408302	0.5396324	
# 29	35	101895	88428	190323	217979	408302	0.5338671	
# 30	36	271890	3636	275526	132776	408302	0.3251907	
# 31	37	280857	2716	283573	124729	408302	0.3054822	
# 32	40	1233	283707	284940	123362	408302	0.3021342	
# 33	41	1957	273177	275134	133168	408302	0.3261507	
# 34	42	86469	103226	189695	218607	408302	0.5354052	
# 35	43	87577	98843	186420	221882	408302	0.5434262	
# 36	44	3668	272853	276521	131781	408302	0.3227537	
# 37	45	5508	269028	274536	133766	408302	0.3276153	
# 38	46	89196	97572	186768	221534	408302	0.5425739	
# 39	47	91873	95971	187844	220458	408302	0.5399386	
# 40	50	1782	275846	277628	130674	408302	0.3200425	
# 41	51	3049	271596	274645	133657	408302	0.3273484	
# 42	52	87080	99819	186899	221403	408302	0.5422530	
# 43	53	88991	97918	186909	221393	408302	0.5422285	
# 44	54	4434	270058	274492	133810	408302	0.3277231	
# 45	55	8497	268202	276699	131603	408302	0.3223178	
# 46	56	90128	96225	186353	221949	408302	0.5435903	
# 47	57	95836	95357	191193	217109	408302	0.5317363	
# 48	60	95713	93634	189347	218955	408302	0.5362575	
# 49	61	96684	89422	186106	222196	408302	0.5441952	
# 50	62	268717	6909	275626	132676	408302	0.3249458	
# 51	63	270814	3906	274720	133582	408302	0.3271647	

```
# 52 64 98527 88440 186967 221335 408302 0.5420865
# 53 65 101114 86697 187811 220491 408302 0.5400194
# 54 66 272743 2629 275372 132930 408302 0.3255678
# 55 67 278683 1453 280136 128166 408302 0.3139000
# 56 70 96340 90579 186919 221383 408302 0.5422041
# 57 71 98107 88583 186690 221612 408302 0.5427649
# 58 72 269573 4578 274151 134151 408302 0.3285583
# 59 73 273771 3195 276966 131336 408302 0.3216639
# 60 74 99536 87112 186648 221654 408302 0.5428678
# 61 75 105295 86101 191396 216906 408302 0.5312391
# 62 76 274535 1596 276131 132171 408302 0.3237089
# 63 77 288224 917 289141 119161 408302 0.2918453 < L
```

Comparing the 5/2 split to the second-place NPG/rest split (below), the former has fewer pattern instances in conflict with the split (6418 vs 7079), as well as somewhat more random distribution of the conflicting patterns (92 vs 62 rows), whereas the 1/6 split has the majority of its conflicts (3912 of 7079, or 55%) concentrated in one pattern—the 5 NE strains. Collectively, these seem to favor the 5/2 split as the correct “history.”

```
showgroup(pat.summaries,split=strtoi('022'), subset=127, proper.subset=T, c2.thresh=100)
```

```
#      Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 4      003      2                X      X      994    298    532    587
# 7      006      2                X      X      624    138    282    590
# 8      007      3                X      X      104    197    371    557
# 12     013      3                X      X      257    226    146    338
# 16     017      4                X      X      390    329    211    564
# 18     021      2                X      X      998    167    337    402
# 20     023      3                X      X      1274   558   1020   533
# 21     024      2                X      X      686    195    410    625
# 22     025      3                X      X      135    216    466    522
# 23     026      3                X      X      793    431    763    789
# 24     027      4                X      X      461    759   1423    771
# 26     031      3                X      X      268    233    151    361
# 27     032      3                X      X      698    131     74     86
# 28     033      4                X      X      1139   973    574    306
# 29     034      3                X      X      103    119     91    219
# 30     035      4                X      X      578    509    329    503
# 31     036      4                X      X      345    320    227    211
# 32     037      5                X      X      2247   1877   1193    620
# 35     042      2                X      X      503    119    254    394
# 39     046      3                X      X      58     154    425    604
# 40     047      4                X      X      127    256    708    708
# 48     057      5                X      X      221    219    189    324
# 49     060      2                X      X      627    116    237    388
# 51     062      3                X      X      703    269    454    469
# 52     063      4                X      X      151    184    332    194
# 53     064      3                X      X      53     184    458    601
# 54     065      4                X      X      122    284    731    582
# 55     066      4                X      X      217    489   1025    851
# 56     067      5                X      X      556   1015   2544   1151
# 62     075      5                X      X      195    187    210    328
# 63     076      5                X      X      106    130    124    128
# 64     077      6                X      X      850    847    827    485
# 83     122      3      X          X      X      501    162    165    354
# 86     125      4      X          X      X      67     113    223    283
# 87     126      4      X          X      X      99     198    268    425
# 88     127      5      X          X      X      225    321    501    482
# 94     135      5      X          X      X      126    117     88    235
# 96     137      6      X          X      X      405    324    240    333
# 99     142      3      X      X          X      386    409    463    755
# 103    146      4      X      X          X      196    510    969   1795
# 104    147      5      X      X          X      2073   4042   6741  10001
# 111    156      5      X      X          X      565    575    410    735
# 112    157      6      X      X          X      13239  10814   6862  12202
# 113    160      3      X      X      X      337     375    399    712
# 115    162      4      X      X      X      1848   1932   1828   1014
# 116    163      5      X      X      X      255     271    328    316
```

```
# 117 164 4 X X X X 237 627 1053 1752
# 118 165 5 X X X X 2726 5022 8440 9715
# 119 166 5 X X X X X 902 1976 3172 2688
# 120 167 6 X X X X X 11742 21003 35227 15091
# 125 174 5 X X X X X 659 682 468 782
# 126 175 6 X X X X X X 16884 13630 8608 12697
# 127 176 6 X X X X X X 2422 2412 1566 1032
# Other ( 39 rows w/ c2 < 100 ) 3209 1750 1921 4847
# Total 75686 79394 97058 94037
```

```
showgroup(pat.summaries, split=strtoi('010'), subset=127, proper.subset=T, c2.thresh=100)
```

```
# Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 10 011 2 X 515 486 317 827
# 12 013 3 X X 257 226 146 338
# 13 014 2 X X 133 139 132 417
# 14 015 3 X X X 1041 984 660 1389
# 16 017 4 X X X 390 329 211 564
# 26 031 3 X X X 268 233 151 361
# 27 032 3 X X X 698 131 74 86
# 28 033 4 X X X 1139 973 574 306
# 29 034 3 X X X 103 119 91 219
# 30 035 4 X X X X 578 509 329 503
# 31 036 4 X X X X 345 320 227 211
# 32 037 5 X X X X X 2247 1877 1193 620
# 46 055 4 X X X X 606 696 668 971
# 48 057 5 X X X X 221 219 189 324
# 62 075 5 X X X X 195 187 210 328
# 63 076 5 X X X X X 106 130 124 128
# 64 077 6 X X X X X 850 847 827 485
# 78 115 4 X X X X 141 139 122 604
# 94 135 5 X X X X 126 117 88 235
# 96 137 6 X X X X X 405 324 240 333
# 109 154 4 X X X X 1227 1390 1065 1738
# 110 155 5 X X X X X 40157 35344 22417 30602
# 111 156 5 X X X X X 565 575 410 735
# 112 157 6 X X X X X 13239 10814 6862 12202
# 125 174 5 X X X X X 659 682 468 782
# 126 175 6 X X X X X X 16884 13630 8608 12697
# 127 176 6 X X X X X X 2422 2412 1566 1032
# Other ( 35 rows w/ c2 < 100 ) 2151 1232 1130 3730
# Total 87668 75064 49099 72767
```

Below is the full summary of shared SNPs that do *not* directly correspond to tree splits, e.g. deep coalescence, independent coincident mutations, false positives/false negatives in the shared SNP calls, loss of SNPs in hemizygous regions, etc. (Additionally, SAMTools' SNP calls exclude positions it judges to be homozygous, and I think it operates without regard to the reference sequence, so homozygous nonreference positions, while rare except in IT/Wales, often are not called SNPs by SAMTools, but are relevant for this analysis. Provided the position is called a SNP in some other isolate, the consistency filtering we've done above should recover it, but this is still worth keeping in mind when examining the data.)

First, here are SNPs that “coalesce” on the branch from the LCA of bcde, i.e., shared among some nonempty, proper subset of bcde other than bc or de. There are 8 such patterns: any of the 4 choose 3 trios plus any of the 4 pairs having exactly one of bc.

```
sg4 <- showgroup(pat.summaries, subset=strtoi('0145'), split=5, proper.subset = F)
sg4
```

```
# Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 34 041 2 X 42 92 313 368
# 37 044 2 X X 69 279 1001 1809
# 38 045 3 X X 202 593 1970 1656
# 66 101 2 X X 29 47 73 314
# 69 104 2 X X 39 122 329 1196
# 70 105 3 X X 78 181 396 805
# 98 141 3 X X 78 149 258 519
```

```
# 101 144 3 X X X 383 1176 2395 4432
# 102 145 4 X X X X 5992 12644 22332 23189
# Total 6912 15283 29067 34288

sg4n <- nrow(sg4)
sg4pct <- round(sg4$count2[sg4n-1]/sg4$count2[sg4n]*100,1)
sg4pct

# [1] 82.7
```

So, of the 15283 SNPs found only in bcde, 82.7% have a sharing pattern consistent with the given tree structure.

Similarly, we analyze patterns relative to the root of the L-clade (14 patterns—any nonempty proper subset of bcde together with a):

```
sg5 <- showgroup(pat.summaries, subset=stoi('0155'), split=8, proper.subset = F)
sg5

# Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 10 011 2 X 515 486 317 827
# 13 014 2 X X 133 139 132 417
# 14 015 3 X X X 1041 984 660 1389
# 41 050 2 X X 13 24 53 105
# 42 051 3 X X X 52 57 74 126
# 45 054 3 X X X 20 78 133 292
# 46 055 4 X X X 606 696 668 971
# 73 110 2 X X 12 11 29 150
# 74 111 3 X X X 6 11 8 139
# 77 114 3 X X X 12 36 56 365
# 78 115 4 X X X 141 139 122 604
# 105 150 3 X X X 28 51 55 238
# 106 151 4 X X X 43 69 54 220
# 109 154 4 X X X X 1227 1390 1065 1738
# 110 155 5 X X X X 40157 35344 22417 30602
# Total 44006 39515 25843 38183

sg5n <- nrow(sg5)
sg5pct <- round(sg5$count2[sg5n-1]/sg5$count2[sg5n]*100,1)
```

I.e., of the 39515 SNPs found only in abcde, 89.4% have a sharing pattern consistent with the given tree structure.

Finally, how many SNPs have patterns inconsistent with the 5-2 split, i.e., include at least one strain on each side of the 5-2 split, but not shared by all 7?

```
sg7 <- showgroup(pat.summaries, subset=127, split=stoi('022'), proper.subset=F)
sg7

# Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 4 003 2 X X 994 298 532 587
# 7 006 2 X X 624 138 282 590
# 8 007 3 X X X 104 197 371 557
# 11 012 2 X X 565 49 32 93
# 12 013 3 X X X 257 226 146 338
# 15 016 3 X X X 46 60 49 152
# 16 017 4 X X X 390 329 211 564
# 18 021 2 X X 998 167 337 402
# 20 023 3 X X 1274 558 1020 533
# 21 024 2 X X 686 195 410 625
# 22 025 3 X X X 135 216 466 522
# 23 026 3 X X X 793 431 763 789
# 24 027 4 X X X 461 759 1423 771
# 25 030 2 X X 609 69 47 93
# 26 031 3 X X X 268 233 151 361
# 27 032 3 X X X 698 131 74 86
# 28 033 4 X X X 1139 973 574 306
# 29 034 3 X X X 103 119 91 219
# 30 035 4 X X X 578 509 329 503
```

# 31	036	4			X	X	X	X		345	320	227	211
# 32	037	5			X	X	X	X	X	2247	1877	1193	620
# 35	042	2		X				X		503	119	254	394
# 36	043	3		X				X	X	56	86	151	133
# 39	046	3		X			X	X		58	154	425	604
# 40	047	4		X			X	X	X	127	256	708	708
# 43	052	3		X		X		X		8	13	17	22
# 44	053	4		X		X		X	X	35	34	26	56
# 47	056	4		X		X	X	X		26	44	50	88
# 48	057	5		X		X	X	X	X	221	219	189	324
# 49	060	2		X	X					627	116	237	388
# 50	061	3		X	X				X	52	80	131	115
# 51	062	3		X	X			X		703	269	454	469
# 52	063	4		X	X			X	X	151	184	332	194
# 53	064	3		X	X		X			53	184	458	601
# 54	065	4		X	X		X		X	122	284	731	582
# 55	066	4		X	X		X	X		217	489	1025	851
# 56	067	5		X	X		X	X	X	556	1015	2544	1151
# 57	070	3		X	X	X				22	9	14	24
# 58	071	4		X	X	X			X	9	20	7	28
# 59	072	4		X	X	X		X		41	36	21	31
# 60	073	5		X	X	X		X	X	94	72	62	38
# 61	074	4		X	X	X	X			20	46	51	116
# 62	075	5		X	X	X	X		X	195	187	210	328
# 63	076	5		X	X	X	X	X		106	130	124	128
# 64	077	6		X	X	X	X	X	X	850	847	827	485
# 67	102	2	X					X		351	67	96	351
# 68	103	3	X					X	X	24	34	46	143
# 71	106	3	X				X	X		32	66	109	377
# 72	107	4	X				X	X	X	58	84	129	330
# 75	112	3	X			X		X		10	11	8	26
# 76	113	4	X			X		X	X	7	9	5	66
# 79	116	4	X			X	X	X		8	8	11	101
# 80	117	5	X			X	X	X	X	48	32	25	241
# 81	120	2	X		X					432	76	98	309
# 82	121	3	X		X				X	22	22	34	73
# 83	122	3	X		X			X		501	162	165	354
# 84	123	4	X		X			X	X	63	98	91	124
# 85	124	3	X		X		X			43	88	152	400
# 86	125	4	X		X		X		X	67	113	223	283
# 87	126	4	X		X		X	X		99	198	268	425
# 88	127	5	X		X		X	X	X	225	321	501	482
# 89	130	3	X		X	X				9	9	9	27
# 90	131	4	X		X	X			X	6	3	0	52
# 91	132	4	X		X	X		X		19	21	10	38
# 92	133	5	X		X	X		X	X	31	28	15	52
# 93	134	4	X		X	X	X			18	17	22	143
# 94	135	5	X		X	X	X		X	126	117	88	235
# 95	136	5	X		X	X	X	X		34	56	26	106
# 96	137	6	X		X	X	X	X	X	405	324	240	333
# 99	142	3	X	X				X		386	409	463	755
# 100	143	4	X	X				X	X	37	58	103	190
# 103	146	4	X	X			X	X		196	510	969	1795
# 104	147	5	X	X			X	X	X	2073	4042	6741	10001
# 107	152	4	X	X		X		X		18	27	15	67
# 108	153	5	X	X		X		X	X	39	27	26	96
# 111	156	5	X	X		X	X	X		565	575	410	735
# 112	157	6	X	X		X	X	X	X	13239	10814	6862	12202
# 113	160	3	X	X	X					337	375	399	712
# 114	161	4	X	X	X				X	74	96	113	207
# 115	162	4	X	X	X			X		1848	1932	1828	1014
# 116	163	5	X	X	X			X	X	255	271	328	316
# 117	164	4	X	X	X		X			237	627	1053	1752
# 118	165	5	X	X	X		X		X	2726	5022	8440	9715
# 119	166	5	X	X	X		X	X		902	1976	3172	2688
# 120	167	6	X	X	X		X	X	X	11742	21003	35227	15091
# 121	170	4	X	X	X	X				13	18	15	69

```
# 122 171 5 X X X X X 41 19 13 70
# 123 172 5 X X X X X 58 71 45 86
# 124 173 6 X X X X X 131 87 47 114
# 125 174 5 X X X X X 659 682 468 782
# 126 175 6 X X X X X X 16884 13630 8608 12697
# 127 176 6 X X X X X X 2422 2412 1566 1032
# 128 177 7 X X X X X X 77690 62182 38744 15186
# Total 153376 141576 135802 109223

sg7n <- nrow(sg7)
sg7pct <- round(sg7$count2[sg7n-1]/sg7$count2[sg7n]*100,1)
sg7pct

# [1] 43.9
```

A more compact version of that table, showing only the larger counts:

```
thresh <- signif(.02 * sg7$count2[sg7n],1)
thresh

# [1] 3000

showgroup(pat.summaries, subset=127, split=stoi('022'), proper.subset=F, c2.thresh = thresh)

# Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 104 147 5 X X X X X 2073 4042 6741 10001
# 112 157 6 X X X X X 13239 10814 6862 12202
# 118 165 5 X X X X X 2726 5022 8440 9715
# 120 167 6 X X X X X 11742 21003 35227 15091
# 126 175 6 X X X X X 16884 13630 8608 12697
# 128 177 7 X X X X X 77690 62182 38744 15186
# Other ( 87 rows w/ c2 < 3000 ) 29022 24883 31180 34331
# Total 153376 141576 135802 109223
```

So, of the 141576 SNPs found both in the L- and H-clades, 43.9% have a sharing pattern consistent with the given tree structure, i.e., are found in all 7 isolates. Among the others, three patterns dominate—(i) the 6-way pattern excluding the Gyre is the largest, plausibly explained by 7-way sharing from which the Gyre drops out due to low coverage/high error rate, (ii) the 6-way excluding Italy, and (iii) ditto for Wales. Origin of the later two cases is unclear, but may partly reflect Hardy-Weinberg—some positions that are *population-level* SNPs in those isolates will be homozygous-reference in the CCMP founder cell for IT or Wales. If I take the 7-way shared SNP count (69526) as a surrogate approximating the number of population-level SNPs in either IT or Wales that are shared with the L-clade, then I might expect, based on HWE, roughly half that number to be lost (become homozygous) in IT, and a similar number in Wales. However, the observed counts of these positions are lower by $\approx 20K$ than I might have guessed from HWE, perhaps suggesting that IT and Wales are distinct populations, each with a pool of many thousand private polymorphisms.

In aggregate:

```
untreelike <-
  sg7$count2[sg7n]-sg7$count2[sg7n-1] +
  sg5$count2[sg5n]-sg5$count2[sg5n-1] +
  sg4$count2[sg4n]-sg4$count2[sg4n-1]
untreelike

# [1] 86204

consistent.count[2]

# [1] 469906

unpct <- round(untreelike/consistent.count[2]*100,1)
unpct

# [1] 18.3
```

Overall, based on this data, I take the following to be obvious: (a) separation of the the H-isolates from the L-isolates (and from each other??), and (b) near-identity of the L-isolates. Due to the small counts, the exact topology among the L-isolates (esp. bcde) is uncertain, but *any* topology there is consistent with the asexual/clonal/global-expansion hypothesis, so there is little point in examining this subtree more carefully. Again, we believe the (apparent) slight separation of the Gyre from the other L-isolates is largely driven by technical artifacts (lower coverage/higher error rates) in the sequencing rather than by biological effects. However, the discord between Gyre SNPs and others is the major substantive ambiguity in the offered tree. Nevertheless, in the next section we show by a bootstrap analysis that the offered placement of Gyre with respect to the other 4 L-isolates is strongly supported by the data.

9.1 Bootstrap

```
n2 <- sum(pattern.counts[[2]][,2]); n2
# [1] 469906
```

```
boot.sample <- sample(0:127,n2,replace=T,prob=pattern.counts[[2]][,2])
str(boot.sample)

#   int [1:469906] 18 2 109 18 16 2 127 18 2 25 ...

boot.count <- mytable(boot.sample,c(0,127))
boot.count[c(1:4,125:128),] # show a few rows

#      val count
# [1,]    0   600
# [2,]    1   668
# [3,]    2 84782
# [4,]    3   317
# [5,]  124   714
# [6,] 125 13529
# [7,] 126  2444
# [8,] 127 62227

boot.counts <- list(NULL,boot.count,NULL) # dummy list with just c2 summaries
cor(pattern.counts[[2]][,2],boot.counts[[2]][,2]) # just curious - how correlated are they?

# [1] 0.9999912

boot.summaries <- pat.summary(boot.counts)
showgroup(boot.summaries,c2.thresh=400) #show a few rows

#      Pat ShrBy 1007 1012 1013 1014 1015 3367 1335 count1 count2 count3 count4
# 1     000     0
# 2     001     1
# 3     002     1
# 5     004     1
# 6     005     2
#           X
#           X
#           X
```


# 9	010	1				X				NA	496	NA	NA
# 10	011	2				X			X	NA	522	NA	NA
# 14	015	3				X	X		X	NA	962	NA	NA
# 17	020	1			X					NA	94351	NA	NA
# 19	022	2			X				X	NA	87810	NA	NA
# 20	023	3			X				X	NA	622	NA	NA
# 23	026	3			X			X	X	NA	416	NA	NA
# 24	027	4			X			X	X	NA	756	NA	NA
# 28	033	4			X	X			X	NA	1024	NA	NA
# 30	035	4			X	X	X		X	NA	548	NA	NA
# 32	037	5			X	X	X	X	X	NA	1910	NA	NA
# 33	040	1			X					NA	696	NA	NA
# 38	045	3			X			X		NA	590	NA	NA
# 46	055	4			X		X	X		NA	672	NA	NA
# 55	066	4			X	X		X	X	NA	477	NA	NA
# 56	067	5			X	X		X	X	NA	984	NA	NA
# 64	077	6			X	X	X	X	X	NA	836	NA	NA
# 97	140	2	X	X						NA	1098	NA	NA
# 101	144	3	X	X				X		NA	1208	NA	NA
# 102	145	4	X	X				X		NA	12366	NA	NA
# 103	146	4	X	X				X	X	NA	512	NA	NA
# 104	147	5	X	X				X	X	NA	4120	NA	NA
# 109	154	4	X	X			X	X		NA	1402	NA	NA
# 110	155	5	X	X			X	X		NA	35560	NA	NA
# 111	156	5	X	X			X	X	X	NA	606	NA	NA
# 112	157	6	X	X			X	X	X	NA	10853	NA	NA
# 115	162	4	X	X	X				X	NA	1963	NA	NA
# 117	164	4	X	X	X			X		NA	624	NA	NA
# 118	165	5	X	X	X			X		NA	5137	NA	NA
# 119	166	5	X	X	X			X	X	NA	1934	NA	NA
# 120	167	6	X	X	X			X	X	NA	20694	NA	NA
# 125	174	5	X	X	X	X		X		NA	714	NA	NA
# 126	175	6	X	X	X	X	X		X	NA	13529	NA	NA
# 127	176	6	X	X	X	X	X	X	X	NA	2444	NA	NA
# 128	177	7	X	X	X	X	X	X	X	NA	62227	NA	NA
# Other	(88 rows	w/	c2	<	400)			NA	10498	NA	NA
# Total										NA	469906	NA	NA

Tree partition analysis (and how to pluck out only the best rows based on 3 smallest cross counts and “best” criteria):

```
tp <- treepart(boot.summaries, root=127) ; tp

# root: 177 ; shared: 62227 . max l 077 , max r 010 , max both 010 , min cross 010 , min ratio 010 .
# All the same?: FALSE
# pat left right both cross all ratio best
# 1 01 1268 289265 290533 117746 408279 0.2883959
# 2 02 85382 179137 264519 143760 408279 0.3521122
# 3 03 86367 105773 192140 216139 408279 0.5293904
# 4 04 2783 279686 282469 125810 408279 0.3081471
# 5 05 3963 274240 278203 130076 408279 0.3185959
# 6 06 87716 100567 188283 219996 408279 0.5388374
# 7 07 89418 98307 187725 220554 408279 0.5402041
# 8 10 1096 331755 332851 75428 408279 0.1847462 < R B C O
# 9 11 2286 281996 284282 123997 408279 0.3037065
# 10 12 85939 123202 209141 199138 408279 0.4877498
# 11 13 87673 102514 190187 218092 408279 0.5341739
# 12 14 3422 276174 279596 128683 408279 0.3151840
# 13 15 6086 273191 279277 129002 408279 0.3159653
# 14 16 88479 98916 187395 220884 408279 0.5410124
# 15 17 92218 97618 189836 218443 408279 0.5350336
# 16 20 94951 165296 260247 148032 408279 0.3625756
# 17 21 95790 95849 191639 216640 408279 0.5306175
# 18 22 267543 61374 328917 79362 408279 0.1943818
# 19 23 269321 8779 278100 130179 408279 0.3188481
# 20 24 97330 91176 188506 219773 408279 0.5382912
# 21 25 98899 88783 187682 220597 408279 0.5403094
# 22 26 270489 4932 275421 132858 408279 0.3254098
# 23 27 273958 3311 277269 131010 408279 0.3208835
# 24 30 95505 112516 208021 200258 408279 0.4904930
# 25 31 97115 92767 189882 218397 408279 0.5349210
# 26 32 268291 21112 289403 118876 408279 0.2911636
```

```
# 27 33 272091 6515 278606 129673 408279 0.3176088
# 28 34 98142 89531 187673 220606 408279 0.5403315
# 29 35 101992 88090 190082 218197 408279 0.5344311
# 30 36 271866 3665 275531 132748 408279 0.3251404
# 31 37 281103 2719 283822 124457 408279 0.3048332
# 32 40 1296 283851 285147 123132 408279 0.3015879
# 33 41 2061 273142 275203 133076 408279 0.3259438
# 34 42 86211 103290 189501 218778 408279 0.5358542
# 35 43 87379 98842 186221 222058 408279 0.5438879
# 36 44 3768 273016 276784 131495 408279 0.3220714
# 37 45 5635 268963 274598 133681 408279 0.3274256
# 38 46 88975 97624 186599 221680 408279 0.5429620
# 39 47 91719 95936 187655 220624 408279 0.5403756
# 40 50 1819 275928 277747 130532 408279 0.3197127
# 41 51 3179 271605 274784 133495 408279 0.3269700
# 42 52 86807 99874 186681 221598 408279 0.5427612
# 43 53 88822 97953 186775 221504 408279 0.5425310
# 44 54 4519 270157 274676 133603 408279 0.3272346
# 45 55 8615 268167 276782 131497 408279 0.3220763
# 46 56 89905 96267 186172 222107 408279 0.5440079
# 47 57 95668 95360 191028 217251 408279 0.5321141
# 48 60 95744 93371 189115 219164 408279 0.5367996
# 49 61 96761 89126 185887 222392 408279 0.5447059
# 50 62 268745 6953 275698 132581 408279 0.3247314
# 51 63 270977 3919 274896 133383 408279 0.3266957
# 52 64 98594 88179 186773 221506 408279 0.5425359
# 53 65 101214 86346 187560 220719 408279 0.5406083
# 54 66 272780 2683 275463 132816 408279 0.3253070
# 55 67 278829 1435 280264 128015 408279 0.3135478
# 56 70 96339 90329 186668 221611 408279 0.5427930
# 57 71 98221 88296 186517 221762 408279 0.5431629
# 58 72 269584 4636 274220 134059 408279 0.3283514
# 59 73 274043 3224 277267 131012 408279 0.3208884
# 60 74 99588 86839 186427 221852 408279 0.5433833
# 61 75 105462 85769 191231 217048 408279 0.5316169
# 62 76 274561 1642 276203 132076 408279 0.3234945
# 63 77 288510 925 289435 118844 408279 0.2910853 < L
```

```
otp <- order(tp[, 'cross'])[1:3] # 3 smallest 'cross' counts
btp <- which(tp[, 'best'] != '') # 'best' by Left/Right/Both/Cross/ratio
toptp <- unique(c(otp, btp, 18, 8)) # above, plus 5/2, 6/1 splits
print(tp[toptp,]) # show the winners
```

```
# pat left right both cross all ratio best
# 8 10 1096 331755 332851 75428 408279 0.1847462 < R B C O
# 18 22 267543 61374 328917 79362 408279 0.1943818
# 1 01 1268 289265 290533 117746 408279 0.2883959
# 63 77 288510 925 289435 118844 408279 0.2910853 < L
```

Now repeat the above nboot times, and summarize results:

```
nboot <- params$nboot # default from params set in section 2
nboot <- ((nboot+2) %/% 4) * 4 + 1 # summary is cleaner if n mod 4 == 1, so int median/quartiles
cat('***\n*** Doing', nboot, 'bootstrap replicates.\n***\n')

# ***
# *** Doing 101 bootstrap replicates.
# ***

bcor <- numeric(nboot)
b52cross <- integer(nboot)
b61cross <- integer(nboot)
brev <- logical(nboot)
for(i in 1:nboot){
  boot.sample <- sample(0:127, n2, replace=T, prob=pattern.counts[[2]][,2])
  boot.count <- mytable(boot.sample, c(0, 127))
  boot.counts <- list(NULL, boot.count, NULL) # dummy list with just c2 summaries
  boot.summaries <- pat.summary(boot.counts)
  tp <- treepart(boot.summaries, root=127, verbose=F)
  bcor[i] <- cor(pattern.counts[[2]][,2], boot.counts[[2]][,2]) # just curious - how correlated are they?
  b52cross[i] <- tp[18, 'cross']
  b61cross[i] <- tp[ 8, 'cross']
```

```

brev[i] <- (b52cross[i] > b61cross[i])
if(brev[i]){
  # show the unexpected ones; probably breaks w/ cache
  otp <- order(tp[, 'cross'])[1:3]
  btp <- which(tp[, 'best'] != '')
  totp <- unique(c(otp, btp, 18, 8))
  print(tp[tottp,])
}
}

# pat left right both cross all ratio best
# 8 10 1160 332014 333174 75109 408283 0.1839631 < R B C O
# 18 22 267271 61464 328735 79548 408283 0.1948354
# 1 01 1221 288865 290086 118197 408283 0.2894977
# 63 77 288099 927 289026 119257 408283 0.2920940 < L
# pat left right both cross all ratio best
# 8 10 1094 331785 332879 75081 407960 0.1840401 < R B C O
# 18 22 267351 61178 328529 79431 407960 0.1947029
# 1 01 1193 289019 290212 117748 407960 0.2886263
# 63 77 288087 923 289010 118950 407960 0.2915727 < L
# pat left right both cross all ratio best
# 8 10 1077 332566 333643 74877 408520 0.1832885 < R B C O
# 18 22 267919 61400 329319 79201 408520 0.1938730
# 1 01 1193 289455 290648 117872 408520 0.2885342
# 63 77 288578 897 289475 119045 408520 0.2914056 < L
# pat left right both cross all ratio best
# 8 10 1099 331968 333067 74994 408061 0.1837813 < R B C O
# 18 22 267149 61552 328701 79360 408061 0.1944807
# 1 01 1222 288655 289877 118184 408061 0.2896234
# 63 77 287835 915 288750 119311 408061 0.2923852 < L
# pat left right both cross all ratio best
# 8 10 1094 331997 333091 75318 408409 0.1844181 < R B C O
# 18 22 267448 61849 329297 79112 408409 0.1937078
# 1 01 1198 289032 290230 118179 408409 0.2893643
# 63 77 288274 936 289210 119199 408409 0.2918618 < L
# pat left right both cross all ratio best
# 8 10 1085 332159 333244 75184 408428 0.1840814 < R B C O
# 18 22 267430 61424 328854 79574 408428 0.1948299
# 1 01 1229 288886 290115 118313 408428 0.2896790
# 63 77 288071 913 288984 119444 408428 0.2924481 < L
# pat left right both cross all ratio best
# 8 10 1092 332052 333144 75376 408520 0.1845099 < R B C O
# 18 22 267464 61343 328807 79713 408520 0.1951263
# 1 01 1170 289311 290481 118039 408520 0.2889430
# 63 77 288125 932 289057 119463 408520 0.2924288 < L
# pat left right both cross all ratio best
# 8 10 1084 332108 333192 74966 408158 0.1836691 < R B C O
# 18 22 267710 61217 328927 79231 408158 0.1941185
# 1 01 1242 289256 290498 117660 408158 0.2882707
# 63 77 288517 886 289403 118755 408158 0.2909535 < L
# pat left right both cross all ratio best
# 8 10 1119 332162 333281 75485 408766 0.1846656 < R B C O
# 18 22 267660 61604 329264 79502 408766 0.1944927
# 1 01 1196 289451 290647 118119 408766 0.2889648
# 63 77 288117 983 289100 119666 408766 0.2927494 < L
# pat left right both cross all ratio best
# 8 10 1109 332181 333290 75171 408461 0.1840347 < R B C O
# 18 22 267410 61476 328886 79575 408461 0.1948166
# 1 01 1199 289122 290321 118140 408461 0.2892320
# 63 77 288130 942 289072 119389 408461 0.2922898 < L
# pat left right both cross all ratio best
# 8 10 1114 331931 333045 75374 408419 0.1845507 < R B C O
# 18 22 267525 61921 329446 78973 408419 0.1933627
# 1 01 1234 289028 290262 118157 408419 0.2893034
# 63 77 288165 971 289136 119283 408419 0.2920604 < L
# pat left right both cross all ratio best
# 8 10 1092 331886 332978 75297 408275 0.1844272 < R B C O
# 18 22 267181 61582 328763 79512 408275 0.1947511

```

```

# 1 01 1199 288894 290093 118182 408275 0.2894667
# 63 77 287855 898 288753 119522 408275 0.2927488 < L
# pat left right both cross all ratio best
# 8 10 1126 331522 332648 75408 408056 0.1847982 < R B C O
# 18 22 266928 61673 328601 79455 408056 0.1947159
# 1 01 1245 288534 289779 118277 408056 0.2898548
# 63 77 287610 949 288559 119497 408056 0.2928446 < L
# pat left right both cross all ratio best
# 8 10 1041 332369 333410 74817 408227 0.1832730 < R B C O
# 18 22 267763 61191 328954 79273 408227 0.1941885
# 1 01 1162 289338 290500 117727 408227 0.2883861
# 63 77 288333 898 289231 118996 408227 0.2914947 < L
# pat left right both cross all ratio best
# 8 10 1164 332149 333313 75134 408447 0.1839504 < R B C O
# 18 22 267737 61479 329216 79231 408447 0.1939811
# 1 01 1247 289361 290608 117839 408447 0.2885050
# 63 77 288530 923 289453 118994 408447 0.2913328 < L
# pat left right both cross all ratio best
# 8 10 1127 332179 333306 74791 408097 0.1832677 < R B C O
# 18 22 267615 61330 328945 79152 408097 0.1939539
# 1 01 1259 289192 290451 117646 408097 0.2882795
# 63 77 288472 923 289395 118702 408097 0.2908671 < L
# pat left right both cross all ratio best
# 8 10 1059 331695 332754 75177 407931 0.1842885 < R B C O
# 18 22 266826 61315 328141 79790 407931 0.1955968
# 1 01 1218 288717 289935 117996 407931 0.2892548
# 63 77 287605 891 288496 119435 407931 0.2927824 < L
# pat left right both cross all ratio best
# 8 10 1113 332352 333465 74951 408416 0.1835163 < R B C O
# 18 22 268249 61175 329424 78992 408416 0.1934106
# 1 01 1176 289774 290950 117466 408416 0.2876136
# 63 77 288882 889 289771 118645 408416 0.2905004 < L
# pat left right both cross all ratio best
# 8 10 1105 331707 332812 75289 408101 0.1844862 < R B C O
# 18 22 267191 61398 328589 79512 408101 0.1948341
# 1 01 1213 289018 290231 117870 408101 0.2888256
# 63 77 288122 968 289090 119011 408101 0.2916214 < L
# pat left right both cross all ratio best
# 8 10 1106 332248 333354 74938 408292 0.1835402 < R B C O
# 18 22 267667 61280 328947 79345 408292 0.1943340
# 1 01 1247 289422 290669 117623 408292 0.2880855
# 63 77 288256 935 289191 119101 408292 0.2917054 < L
# pat left right both cross all ratio best
# 8 10 1112 332321 333433 74941 408374 0.1835107 < R B C O
# 18 22 267698 61208 328906 79468 408374 0.1945961
# 1 01 1234 289478 290712 117662 408374 0.2881231
# 63 77 288460 886 289346 119028 408374 0.2914681 < L
# pat left right both cross all ratio best
# 8 10 1067 332182 333249 74826 408075 0.1833634 < R B C O
# 18 22 267830 61482 329312 78763 408075 0.1930111
# 1 01 1175 289535 290710 117365 408075 0.2876064
# 63 77 288417 888 289305 118770 408075 0.2910494 < L
# pat left right both cross all ratio best
# 8 10 1123 332364 333487 74821 408308 0.1832465 < R B C O
# 18 22 267954 61230 329184 79124 408308 0.1937851
# 1 01 1236 289533 290769 117539 408308 0.2878685
# 63 77 288562 949 289511 118797 408308 0.2909495 < L
# pat left right both cross all ratio best
# 8 10 1046 331901 332947 75232 408179 0.1843113 < R B C O
# 18 22 267330 61161 328491 79688 408179 0.1952281
# 1 01 1174 289077 290251 117928 408179 0.2889125
# 63 77 287867 921 288788 119391 408179 0.2924967 < L
# pat left right both cross all ratio best
# 8 10 1136 332210 333346 74916 408262 0.1834998 < R B C O
# 18 22 267418 61060 328478 79784 408262 0.1954235
# 1 01 1173 289301 290474 117788 408262 0.2885108
# 63 77 288040 870 288910 119352 408262 0.2923417 < L

```

#	pat	left	right	both	cross	all	ratio	best
# 8	10	1092	332546	333638	75071	408709	0.1836784	< R B C O
# 18	22	267576	61853	329429	79280	408709	0.1939766	
# 1	01	1189	289391	290580	118129	408709	0.2890296	
# 63	77	288227	893	289120	119589	408709	0.2926018	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1116	331900	333016	75085	408101	0.1839863	< R B C O
# 18	22	267508	61256	328764	79337	408101	0.1944053	
# 1	01	1262	289186	290448	117653	408101	0.2882938	
# 63	77	288136	924	289060	119041	408101	0.2916949	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1076	332146	333222	74983	408205	0.1836896	< R B C O
# 18	22	267828	61309	329137	79068	408205	0.1936968	
# 1	01	1143	289247	290390	117815	408205	0.2886172	
# 63	77	288442	890	289332	118873	408205	0.2912091	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1061	332297	333358	75054	408412	0.1837703	< R B C O
# 18	22	267597	61312	328909	79503	408412	0.1946637	
# 1	01	1193	289214	290407	118005	408412	0.2889362	
# 63	77	288063	885	288948	119464	408412	0.2925085	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1041	332188	333229	74801	408030	0.1833223	< R B C O
# 18	22	267732	61104	328836	79194	408030	0.1940887	
# 1	01	1144	289333	290477	117553	408030	0.2880989	
# 63	77	288289	919	289208	118822	408030	0.2912090	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1134	331704	332838	74970	407808	0.1838365	< R B C O
# 18	22	267111	61420	328531	79277	407808	0.1943979	
# 1	01	1232	288955	290187	117621	407808	0.2884225	
# 63	77	288191	927	289118	118690	407808	0.2910438	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1091	331789	332880	75217	408097	0.1843116	< R B C O
# 18	22	267288	61621	328909	79188	408097	0.1940421	
# 1	01	1270	289086	290356	117741	408097	0.2885123	
# 63	77	288116	939	289055	119042	408097	0.2917003	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1118	331550	332668	75272	407940	0.1845173	< R B C O
# 18	22	267262	61409	328671	79269	407940	0.1943153	
# 1	01	1193	288873	290066	117874	407940	0.2889494	
# 63	77	287753	918	288671	119269	407940	0.2923690	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1107	331953	333060	74953	408013	0.1837025	< R B C O
# 18	22	267920	60897	328817	79196	408013	0.1941017	
# 1	01	1194	289418	290612	117401	408013	0.2877384	
# 63	77	288605	894	289499	118514	408013	0.2904662	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1133	332284	333417	74786	408203	0.1832079	< R B C O
# 18	22	267548	61139	328687	79516	408203	0.1947952	
# 1	01	1217	289374	290591	117612	408203	0.2881214	
# 63	77	288301	924	289225	118978	408203	0.2914677	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1164	331676	332840	75409	408249	0.1847133	< R B C O
# 18	22	267218	61567	328785	79464	408249	0.1946459	
# 1	01	1230	289089	290319	117930	408249	0.2888678	
# 63	77	287957	991	288948	119301	408249	0.2922261	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1148	331936	333084	74919	408003	0.1836236	< R B C O
# 18	22	267492	61070	328562	79441	408003	0.1947069	
# 1	01	1229	289018	290247	117756	408003	0.2886155	
# 63	77	288037	905	288942	119061	408003	0.2918140	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1115	332064	333179	75078	408257	0.1838989	< R B C O
# 18	22	267679	61479	329158	79099	408257	0.1937481	
# 1	01	1239	289119	290358	117899	408257	0.2887862	
# 63	77	288288	933	289221	119036	408257	0.2915712	< L
#	pat	left	right	both	cross	all	ratio	best
# 8	10	1077	331731	332808	75096	407904	0.1841021	< R B C O

```

# 18 22 267283 61294 328577 79327 407904 0.1944747
# 1 01 1174 288798 289972 117932 407904 0.2891170
# 63 77 287840 891 288731 119173 407904 0.2921594 < L
# pat left right both cross all ratio best
# 8 10 1110 332205 333315 74724 408039 0.1831296 < R B C O
# 18 22 267699 61192 328891 79148 408039 0.1939717
# 1 01 1147 289270 290417 117622 408039 0.2882617
# 63 77 288498 896 289394 118645 408039 0.2907688 < L
# pat left right both cross all ratio best
# 8 10 1174 332154 333328 75231 408559 0.1841374 < R B C O
# 18 22 267486 61640 329126 79433 408559 0.1944223
# 1 01 1223 289263 290486 118073 408559 0.2889987
# 63 77 288171 916 289087 119472 408559 0.2924229 < L
# pat left right both cross all ratio best
# 8 10 1056 332415 333471 75010 408481 0.1836316 < R B C O
# 18 22 267522 61017 328539 79942 408481 0.1957056
# 1 01 1212 289197 290409 118072 408481 0.2890514
# 63 77 288093 881 288974 119507 408481 0.2925644 < L
# pat left right both cross all ratio best
# 8 10 1145 331829 332974 75131 408105 0.1840972 < R B C O
# 18 22 267139 61335 328474 79631 408105 0.1951238
# 1 01 1197 288976 290173 117932 408105 0.2889747
# 63 77 287604 993 288597 119508 408105 0.2928364 < L
# pat left right both cross all ratio best
# 8 10 1091 332036 333127 75511 408638 0.1847870 < R B C O
# 18 22 267495 61343 328838 79800 408638 0.1952829
# 1 01 1176 289294 290470 118168 408638 0.2891753
# 63 77 288205 883 289088 119550 408638 0.2925572 < L
# pat left right both cross all ratio best
# 8 10 1122 332241 333363 75029 408392 0.1837181 < R B C O
# 18 22 267707 61407 329114 79278 408392 0.1941223
# 1 01 1197 289339 290536 117856 408392 0.2885855
# 63 77 288307 944 289251 119141 408392 0.2917320 < L
# pat left right both cross all ratio best
# 8 10 1027 331856 332883 75051 407934 0.1839783 < R B C O
# 18 22 267542 60990 328532 79402 407934 0.1946442
# 1 01 1156 289237 290393 117541 407934 0.2881373
# 63 77 287942 890 288832 119102 407934 0.2919639 < L
# pat left right both cross all ratio best
# 8 10 1080 332197 333277 75099 408376 0.1838967 < R B C O
# 18 22 267559 61270 328829 79547 408376 0.1947886
# 1 01 1199 289257 290456 117920 408376 0.2887535
# 63 77 288517 868 289385 118991 408376 0.2913761 < L
# pat left right both cross all ratio best
# 8 10 1119 332005 333124 75131 408255 0.1840296 < R B C O
# 18 22 267285 61687 328972 79283 408255 0.1941997
# 1 01 1234 289004 290238 118017 408255 0.2890767
# 63 77 287841 971 288812 119443 408255 0.2925696 < L
# pat left right both cross all ratio best
# 8 10 1081 331731 332812 75434 408246 0.1847758 < R B C O
# 18 22 267354 61334 328688 79558 408246 0.1948776
# 1 01 1197 288844 290041 118205 408246 0.2895436
# 63 77 287927 885 288812 119434 408246 0.2925540 < L
# pat left right both cross all ratio best
# 8 10 1093 332022 333115 75225 408340 0.1842215 < R B C O
# 18 22 267666 61426 329092 79248 408340 0.1940736
# 1 01 1204 289443 290647 117693 408340 0.2882230
# 63 77 288320 927 289247 119093 408340 0.2916516 < L
# pat left right both cross all ratio best
# 8 10 1055 331379 332434 75436 407870 0.1849511 < R B C O
# 18 22 267115 61588 328703 79167 407870 0.1940986
# 1 01 1152 288871 290023 117847 407870 0.2889327
# 63 77 287951 902 288853 119017 407870 0.2918013 < L
# pat left right both cross all ratio best
# 8 10 1105 332302 333407 74890 408297 0.1834204 < R B C O
# 18 22 268082 61200 329282 79015 408297 0.1935233
# 1 01 1181 289606 290787 117510 408297 0.2878052

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# 63 77 288711 904 289615 118682 408297 0.2906757 < L
# pat left right both cross all ratio best
# 8 10 1078 332142 333220 75209 408429 0.1841422 < R B C O
# 18 22 267283 61519 328802 79627 408429 0.1949592
# 1 01 1178 288980 290158 118271 408429 0.2895754
# 63 77 287983 903 288886 119543 408429 0.2926898 < L
# pat left right both cross all ratio best
# 8 10 1127 332038 333165 75031 408196 0.1838112 < R B C O
# 18 22 267354 61364 328718 79478 408196 0.1947055
# 1 01 1264 288958 290222 117974 408196 0.2890131
# 63 77 288250 925 289175 119021 408196 0.2915781 < L
# pat left right both cross all ratio best
# 8 10 1087 332264 333351 75074 408425 0.1838134 < R B C O
# 18 22 267382 61626 329008 79417 408425 0.1944470
# 1 01 1229 289078 290307 118118 408425 0.2892036
# 63 77 288071 917 288988 119437 408425 0.2924331 < L
# pat left right both cross all ratio best
# 8 10 1128 332817 333945 74557 408502 0.1825132 < R B C O
# 18 22 267873 60794 328667 79835 408502 0.1954336
# 1 01 1151 289597 290748 117754 408502 0.2882581
# 63 77 288560 941 289501 119001 408502 0.2913107 < L
# pat left right both cross all ratio best
# 8 10 1127 332385 333512 75240 408752 0.1840725 < R B C O
# 18 22 268226 61185 329411 79341 408752 0.1941055
# 1 01 1174 289575 290749 118003 408752 0.2886909
# 63 77 289012 917 289929 118823 408752 0.2906970 < L
# pat left right both cross all ratio best
# 8 10 1148 332272 333420 75027 408447 0.1836885 < R B C O
# 18 22 268023 61145 329168 79279 408447 0.1940986
# 1 01 1195 289731 290926 117521 408447 0.2877264
# 63 77 288467 923 289390 119057 408447 0.2914870 < L
# pat left right both cross all ratio best
# 8 10 1102 331761 332863 74937 407800 0.1837592 < R B C O
# 18 22 267268 61140 328408 79392 407800 0.1946837
# 1 01 1192 289036 290228 117572 407800 0.2883080
# 63 77 288146 909 289055 118745 407800 0.2911844 < L
# pat left right both cross all ratio best
# 8 10 1080 332026 333106 74915 408021 0.1836057 < R B C O
# 18 22 267540 61120 328660 79361 408021 0.1945022
# 1 01 1203 288877 290080 117941 408021 0.2890562
# 63 77 288033 934 288967 119054 408021 0.2917840 < L
# pat left right both cross all ratio best
# 8 10 1102 331970 333072 75098 408170 0.1839871 < R B C O
# 18 22 267329 61629 328958 79212 408170 0.1940662
# 1 01 1225 289057 290282 117888 408170 0.2888208
# 63 77 287975 914 288889 119281 408170 0.2922336 < L
# pat left right both cross all ratio best
# 8 10 1086 332224 333310 75231 408541 0.1841455 < R B C O
# 18 22 267226 61514 328740 79801 408541 0.1953317
# 1 01 1211 289182 290393 118148 408541 0.2891950
# 63 77 288177 924 289101 119440 408541 0.2923574 < L
# pat left right both cross all ratio best
# 8 10 1032 331970 333002 75095 408097 0.1840126 < R B C O
# 18 22 267177 61161 328338 79759 408097 0.1954413
# 1 01 1239 288566 289805 118292 408097 0.2898625
# 63 77 287622 870 288492 119605 408097 0.2930798 < L
# pat left right both cross all ratio best
# 8 10 1079 332309 333388 75123 408511 0.1838947 < R B C O
# 18 22 267445 61607 329052 79459 408511 0.1945088
# 1 01 1221 289226 290447 118064 408511 0.2890106
# 63 77 288076 948 289024 119487 408511 0.2924940 < L
# pat left right both cross all ratio best
# 8 10 1151 332468 333619 74727 408346 0.1829992 < R B C O
# 18 22 267745 61343 329088 79258 408346 0.1940952
# 1 01 1258 289274 290532 117814 408346 0.2885151
# 63 77 288441 959 289400 118946 408346 0.2912873 < L
# pat left right both cross all ratio best

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# 8 10 1082 332319 333401 74760 408161 0.1831630 < R B C O
# 18 22 267397 61180 328577 79584 408161 0.1949819
# 1 01 1205 289038 290243 117918 408161 0.2889007
# 63 77 288157 908 289065 119096 408161 0.2917868 < L
# pat left right both cross all ratio best
# 8 10 1120 332303 333423 75026 408449 0.1836851 < R B C O
# 18 22 267792 61632 329424 79025 408449 0.1934758
# 1 01 1246 289464 290710 117739 408449 0.2882588
# 63 77 288399 929 289328 119121 408449 0.2916423 < L
# pat left right both cross all ratio best
# 8 10 1074 331813 332887 74955 407842 0.1837844 < R B C O
# 18 22 267249 61101 328350 79492 407842 0.1949088
# 1 01 1172 288819 289991 117851 407842 0.2889624
# 63 77 287898 910 288808 119034 407842 0.2918630 < L
# pat left right both cross all ratio best
# 8 10 1116 332412 333528 75202 408730 0.1839894 < R B C O
# 18 22 268008 61191 329199 79531 408730 0.1945808
# 1 01 1212 289682 290894 117836 408730 0.2882979
# 63 77 288610 889 289499 119231 408730 0.2917109 < L
# pat left right both cross all ratio best
# 8 10 1113 332371 333484 75068 408552 0.1837416 < R B C O
# 18 22 267733 61405 329138 79414 408552 0.1943792
# 1 01 1190 289535 290725 117827 408552 0.2884015
# 63 77 288431 915 289346 119206 408552 0.2917768 < L
# pat left right both cross all ratio best
# 8 10 1109 332385 333494 74803 408297 0.1832073 < R B C O
# 18 22 267603 61327 328930 79367 408297 0.1943855
# 1 01 1147 289047 290194 118103 408297 0.2892576
# 63 77 288254 971 289225 119072 408297 0.2916308 < L
# pat left right both cross all ratio best
# 8 10 1065 332101 333166 75043 408209 0.1838348 < R B C O
# 18 22 267045 61240 328285 79924 408209 0.1957919
# 1 01 1171 288784 289955 118254 408209 0.2896898
# 63 77 287749 885 288634 119575 408209 0.2929259 < L
# pat left right both cross all ratio best
# 8 10 1118 332168 333286 74889 408175 0.1834728 < R B C O
# 18 22 267943 61004 328947 79228 408175 0.1941030
# 1 01 1197 289557 290754 117421 408175 0.2876732
# 63 77 288642 884 289526 118649 408175 0.2906817 < L
# pat left right both cross all ratio best
# 8 10 1059 331847 332906 75171 408077 0.1842079 < R B C O
# 18 22 267922 61036 328958 79119 408077 0.1938825
# 1 01 1217 289434 290651 117426 408077 0.2877545
# 63 77 288736 873 289609 118468 408077 0.2903080 < L
# pat left right both cross all ratio best
# 8 10 1125 331670 332795 75154 407949 0.1842240 < R B C O
# 18 22 267221 61269 328490 79459 407949 0.1947768
# 1 01 1232 288707 289939 118010 407949 0.2892764
# 63 77 287716 924 288640 119309 407949 0.2924606 < L
# pat left right both cross all ratio best
# 8 10 1103 331801 332904 75038 407942 0.1839428 < R B C O
# 18 22 267190 61273 328463 79479 407942 0.1948292
# 1 01 1230 288729 289959 117983 407942 0.2892151
# 63 77 287880 926 288806 119136 407942 0.2920415 < L
# pat left right both cross all ratio best
# 8 10 1126 332034 333160 75395 408555 0.1845406 < R B C O
# 18 22 267653 61154 328807 79748 408555 0.1951953
# 1 01 1187 289163 290350 118205 408555 0.2893246
# 63 77 288407 957 289364 119191 408555 0.2917380 < L
# pat left right both cross all ratio best
# 8 10 1072 332112 333184 75332 408516 0.1844040 < R B C O
# 18 22 267519 61638 329157 79359 408516 0.1942617
# 1 01 1168 288977 290145 118371 408516 0.2897585
# 63 77 288145 913 289058 119458 408516 0.2924194 < L
# pat left right both cross all ratio best
# 8 10 1080 332189 333269 75366 408635 0.1844335 < R B C O
# 18 22 267485 61696 329181 79454 408635 0.1944376

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# 1 01 1226 289339 290565 118070 408635 0.2889376
# 63 77 288420 901 289321 119314 408635 0.2919818 < L
# pat left right both cross all ratio best
# 8 10 1108 332120 333228 75153 408381 0.1840267 < R B C O
# 18 22 267504 61357 328861 79520 408381 0.1947201
# 1 01 1189 289019 290208 118173 408381 0.2893695
# 63 77 288109 905 289014 119367 408381 0.2922932 < L
# pat left right both cross all ratio best
# 8 10 1098 332205 333303 75119 408422 0.1839250 < R B C O
# 18 22 267400 61362 328762 79660 408422 0.1950434
# 1 01 1201 289183 290384 118038 408422 0.2890099
# 63 77 288083 909 288992 119430 408422 0.2924181 < L
# pat left right both cross all ratio best
# 8 10 1107 332488 333595 75053 408648 0.1836617 < R B C O
# 18 22 267864 61188 329052 79596 408648 0.1947789
# 1 01 1232 289595 290827 117821 408648 0.2883190
# 63 77 288670 933 289603 119045 408648 0.2913143 < L
# pat left right both cross all ratio best
# 8 10 1089 332348 333437 74991 408428 0.1836089 < R B C O
# 18 22 267691 61499 329190 79238 408428 0.1940073
# 1 01 1230 289245 290475 117953 408428 0.2887975
# 63 77 288379 915 289294 119134 408428 0.2916891 < L
# pat left right both cross all ratio best
# 8 10 1097 332088 333185 75191 408376 0.1841220 < R B C O
# 18 22 267788 61182 328970 79406 408376 0.1944434
# 1 01 1181 289568 290749 117627 408376 0.2880360
# 63 77 288256 923 289179 119197 408376 0.2918805 < L
# pat left right both cross all ratio best
# 8 10 1061 332365 333426 74856 408282 0.1833439 < R B C O
# 18 22 267729 61114 328843 79439 408282 0.1945689
# 1 01 1214 289462 290676 117606 408282 0.2880509
# 63 77 288438 891 289329 118953 408282 0.2913501 < L
# pat left right both cross all ratio best
# 8 10 1193 332170 333363 75205 408568 0.1840697 < R B C O
# 18 22 267575 61551 329126 79442 408568 0.1944401
# 1 01 1263 289024 290287 118281 408568 0.2895014
# 63 77 288318 984 289302 119266 408568 0.2919122 < L
# pat left right both cross all ratio best
# 8 10 1075 331808 332883 75602 408485 0.1850790 < R B C O
# 18 22 267350 61522 328872 79613 408485 0.1948982
# 1 01 1249 288909 290158 118327 408485 0.2896728
# 63 77 288015 935 288950 119535 408485 0.2926301 < L
# pat left right both cross all ratio best
# 8 10 1096 332054 333150 74968 408118 0.1836920 < R B C O
# 18 22 267575 61269 328844 79274 408118 0.1942428
# 1 01 1203 288980 290183 117935 408118 0.2889728
# 63 77 288029 893 288922 119196 408118 0.2920626 < L
# pat left right both cross all ratio best
# 8 10 1032 332458 333490 75104 408594 0.1838108 < R B C O
# 18 22 268134 61251 329385 79209 408594 0.1938575
# 1 01 1147 289365 290512 118082 408594 0.2889959
# 63 77 288839 826 289665 118929 408594 0.2910689 < L
# pat left right both cross all ratio best
# 8 10 1046 332410 333456 75054 408510 0.1837262 < R B C O
# 18 22 268428 61296 329724 78786 408510 0.1928619
# 1 01 1162 289771 290933 117577 408510 0.2878191
# 63 77 288824 884 289708 118802 408510 0.2908179 < L
# pat left right both cross all ratio best
# 8 10 1131 331692 332823 75027 407850 0.1839573 < R B C O
# 18 22 267152 61364 328516 79334 407850 0.1945176
# 1 01 1189 288729 289918 117932 407850 0.2891553
# 63 77 287800 959 288759 119091 407850 0.2919971 < L
# pat left right both cross all ratio best
# 8 10 1131 332442 333573 74636 408209 0.1828377 < R B C O
# 18 22 267639 61123 328762 79447 408209 0.1946233
# 1 01 1194 289285 290479 117730 408209 0.2884062
# 63 77 288364 934 289298 118911 408209 0.2912993 < L

```

```

#   pat   left  right  both  cross  all    ratio    best
# 8   10   1074 331823 332897 75058 407955 0.1839860 < R B C O
# 18  22  267257 61656 328913 79042 407955 0.1937518
# 1   01   1290 288935 290225 117730 407955 0.2885858
# 63  77  287869   921 288790 119165 407955 0.2921033      < L
#   pat   left  right  both  cross  all    ratio    best
# 8   10   1133 332501 333634 75107 408741 0.1837521 < R B C O
# 18  22  267785 61367 329152 79589 408741 0.1947174
# 1   01   1234 289554 290788 117953 408741 0.2885764
# 63  77  288428   944 289372 119369 408741 0.2920407      < L
#   pat   left  right  both  cross  all    ratio    best
# 8   10   1129 331681 332810 75431 408241 0.1847708 < R B C O
# 18  22  267425 61507 328932 79309 408241 0.1942701
# 1   01   1234 288951 290185 118056 408241 0.2891821
# 63  77  288225   916 289141 119100 408241 0.2917394      < L
#   pat   left  right  both  cross  all    ratio    best
# 8   10   1091 331783 332874 75234 408108 0.1843483 < R B C O
# 18  22  267568 61491 329059 79049 408108 0.1936963
# 1   01   1170 289024 290194 117914 408108 0.2889284
# 63  77  288080   875 288955 119153 408108 0.2919644      < L
#   pat   left  right  both  cross  all    ratio    best
# 8   10   1119 332517 333636 74829 408465 0.1831956 < R B C O
# 18  22  267585 61249 328834 79631 408465 0.1949518
# 1   01   1228 289215 290443 118022 408465 0.2889403
# 63  77  288271   931 289202 119263 408465 0.2919785      < L
#   pat   left  right  both  cross  all    ratio    best
# 8   10   1153 331638 332791 75567 408358 0.1850509 < R B C O
# 18  22  267163 61499 328662 79696 408358 0.1951621
# 1   01   1218 288839 290057 118301 408358 0.2896992
# 63  77  287842   953 288795 119563 408358 0.2927897      < L
#   pat   left  right  both  cross  all    ratio    best
# 8   10   1070 331896 332966 75345 408311 0.1845285 < R B C O
# 18  22  267415 61638 329053 79258 408311 0.1941118
# 1   01   1226 289127 290353 117958 408311 0.2888925
# 63  77  288003   924 288927 119384 408311 0.2923850      < L
#   pat   left  right  both  cross  all    ratio    best
# 8   10   1100 332138 333238 75067 408305 0.1838503 < R B C O
# 18  22  267449 61084 328533 79772 408305 0.1953736
# 1   01   1206 289294 290500 117805 408305 0.2885221
# 63  77  288095   938 289033 119272 408305 0.2921150      < L
#   pat   left  right  both  cross  all    ratio    best
# 8   10   1106 332217 333323 74881 408204 0.1834401 < R B C O
# 18  22  267869 60926 328795 79409 408204 0.1945326
# 1   01   1247 289350 290597 117607 408204 0.2881084
# 63  77  288314   920 289234 118970 408204 0.2914474      < L

# summarize:
corsummary <- t(as.matrix(c(summary(bcor),sd=sd(bcor))))
row.names(corsummary) <- 'bcor'
bdelta <- b61cross-b52cross
brevp <- 100*brev # make it percent reversed instead of logical
thesummary <- rbind(summary(b52cross),summary(b61cross),summary(c(bdelta)),summary(brevp))
row.names(thesummary) <- c('b52cross', 'b61cross', 'b61-b52', '% rev')
thesummary <- cbind(thesummary, sd=c(sd(b52cross),sd(b61cross),sd(bdelta),sd(brevp)))

```

SUMMARY: In 101 bootstrap replicates, we saw 101 samples with the 6/1 split having fewer conflicts than the 5/2 split, and the minimum separation between them was ≈ -19 sigma, hence highly statistically significant.

```

# 'opt' hacking is trying to force knitr to show more digits of bcor in summary, as Rstudio does, but
# it still fails... Bottom line is the correlation seems to be > .999 in all samples, rounds to 1.0,
# as seen in this run of 1001 samples cut/paste from Rstudio:
#           Min.      1st Qu.      Median      Mean      3rd Qu.      Max.      sd
# bcor      " 0.9998" " 0.9999" " 0.9999" " 0.9999" " 1" " 1" " 0.00003462"
# > max(bcor)
# [1] 0.9999915
o.opts <- options(digits=7,width=127)
format(rbind(corsummary,thesummary),scientific=F,digits=4,drop0trailing=T)

```

```

#           Min.           1st Qu.           Median           Mean           3rd Qu.           Max.
# bcor      " 0.999981368" " 0.999992195" " 0.999994211" " 0.999993689" " 0.999995819" " 0.99999
# b52cross   "78763"      "79238"      "79409"      "79396.445544554" "79547"      "79942"
# b61cross   "74557"      "74953"      "75078"      "75089.237623762" "75217"      "75602"
# b61-b52    "-5278"      "-4455"      "-4324"      "-4307.207920792" "-4105"      "-3599"
# % rev      " 100"      " 100"      " 100"      " 100"      " 100"      " 100"
#
#           sd
# bcor      " 0.000003173"
# b52cross   " 235.554260214"
# b61cross   " 207.05314045"
# b61-b52    " 282.072945063"
# % rev      " 0"
options(o.opts)

```

Based on this, it is reasonable to claim that we are confident that the tree topology is as shown in the earlier figures, with the exception of the exact order of the splits with the 4 NE coastal isolates.

10 Notes

This section is a random brain dump of limitations of the current analysis, ideas for improvements, etc. In the main, these may not be worth doing, unless we see significant holes or get pushed by reviewers, etc, but I wanted to catalog before we forget them.

Noise: Various sources of “noise” in the data:

1. Read errors, low read depth — perhaps fixed by medium/strict thresholding
2. Deep coalescence
3. Skew because 1335 is the reference. (Julie notes we could partially fix this by remapping based on discovered SNPs, tho that wouldn't fix gross misassembly in 1335, e.g. collapsed or misordered tandem duplicates, or segments missing in 1335 that are present in one or more other strains, etc.; much harder to fix those, let's just hope they are rare...)
4. Varying error rates and sequencing depth among the 7. E.g., plausibly the 1000 SNPs shared by 4 but not by Gyre are a result of lower read depth (we missed a SNP that is actually present) and/or higher error rates (causing a position to appear inconsistent in gyre) in the gyre data. I can't think of a way to correct for this effect. It might be possible, perhaps by simulation, to estimate the size of the effect and see whether it could explain ≈ 1000 SNPs.
5. Varying numbers of founder cells in the sequencing cultures. (Again, I made some attempts at modeling this, but nothing very satisfactory yet.)
6. Tri-allelic positions where stochastic fluctuation in sequence sampling promotes the rare allele to prominence. (Julie replies: “isn't this the same as more than one founder cell? If they are diploid there should only ever be two alleles, unless there were random and very rare, thus unlikely, trisomy events?” I agree, but it is a concrete example of an effect of multiple founders that might be important. Not sure this is the most important such effect...)
7. Gaps/indels - alignments are likely to be of lower quality in the vicinity of an indel, so, maybe lower coverage/more SNPs. We ignored them. Does this add any systematic bias? e.g. if one strain had more indels than another, would this confound other analyses? unclear. Julie suggested a paper titled “Barking up the wrong tree-length: yada yada yada gap penalties”; maybe relevant?

Other Items/Potential To Dos:

1. any spacial structure to various sub-classes?
2. after top level split, should I reanalyze halves of partition in isolation? said another way, I think the tree-building is sensible, but not sure it's optimal.

3. if we believe no sex, then I think gain of SNP should be more common than loss of SNP, since the later can only happen by (a) mutation reverting to reference, (b) second mutation matching nonreference, (c) homologous repair (look for blocks of LOH), or (d) false negative e.g. from low read depth. Does tree-building appropriately weight the gain vs loss cases? (Does it even care?)
4. should we weight coding and/or nonsynonymous SNPs more heavily? Julie says “you do not want to weight the coding or nonsynonymous/coding SNPs because for time you want the more clock-like neutral mutations.” I.e., I got this backwards. Maybe should redo tree based on noncoding SNPs only.
5. We could also do an actual parsimony analysis based on 2-state model (homozygous-ref vs not), but I’m not quite sure how to handle this in a mixed sex/nosex case.
6. Might be interesting to look at sharing just within (shared?) deserts. Given tree model above and expectation that bottleneck followed split of H- from L-clades, I would expect little or no sharing of L-clade desert SNPs with H-clade; sharing between It/Wales might suggest “desert” is actually a region under strong purifying selection (e.g. a gene); sharing/non-sharing within L-clade deserts might suggest more about evo history of the 5.

11 Appendix: Old Trees, etc.

Tangents, old stuff of historical interest at best, etc..

11.1 HWE Sharing

Tangent: As a function of nonref allele freq, assuming HWE, what is probability that nonref allele will be seen in k strains, $0 \leq k \leq 4$ (Fig 6).

```
myfigpath.h <- paste(getwd(), '/figs-knitr/', sep='')

```

```
p <- (0:20)/20
q <- 1-p
r <- 2*p*q+p^2
plot(p, 1*q^0*r^4, type='b', pch='4', ylab="share prob")
points(p, 4*q^2*r^3, type='b', pch='3')
points(p, 6*q^4*r^2, type='b', pch='2')
points(p, 4*q^6*r^1, type='b', pch='1')
points(p, 1*q^8*r^0, type='b', pch='0')
```

11.2 Old Tree Stuff

All based on un-q-filtered reads.

The first pass at the tree analysis was the Chr1 tree, *loose criteria* (c1); it is rendered via <http://iubio.bio.indiana.edu/treeapp/treeprint-form.html> as Fig 7, and in newick format is:

```
newick.chr1.loose <- '(((tp3367_Italy:4551,tp1013_Wales:4954):5920,((tp1007_Virginia:10,tp1012_Australia:29):9,
cat.hardwrap(newick.chr1.loose)

# (((tp3367_Italy:4551,tp1013_Wales:4954):5920,((tp1007_Virginia:10,tp1012_Austra
# lia:29):9,(tp1015_Puget_Sound:90,tp1335_NY:13):11):320,tp1014_Gyre:22):3484):859
# 3,outgroup:0);
```

Chr 1 tree based on *medium criteria* (c2) has exactly the same topology is, although the branch lengths are different. As noted earlier, the length of the branch labeled “*” is probably inflated by lower coverage and higher error rate in 1014, which may mask further legitimate sharing between it and the other L-isolates. The branch lengths among the other 4 are too short for its topology to be convincing without a more rigorous analysis (e.g., a bootstrap test).

Chr1 tree, medium criteria, in newick format:

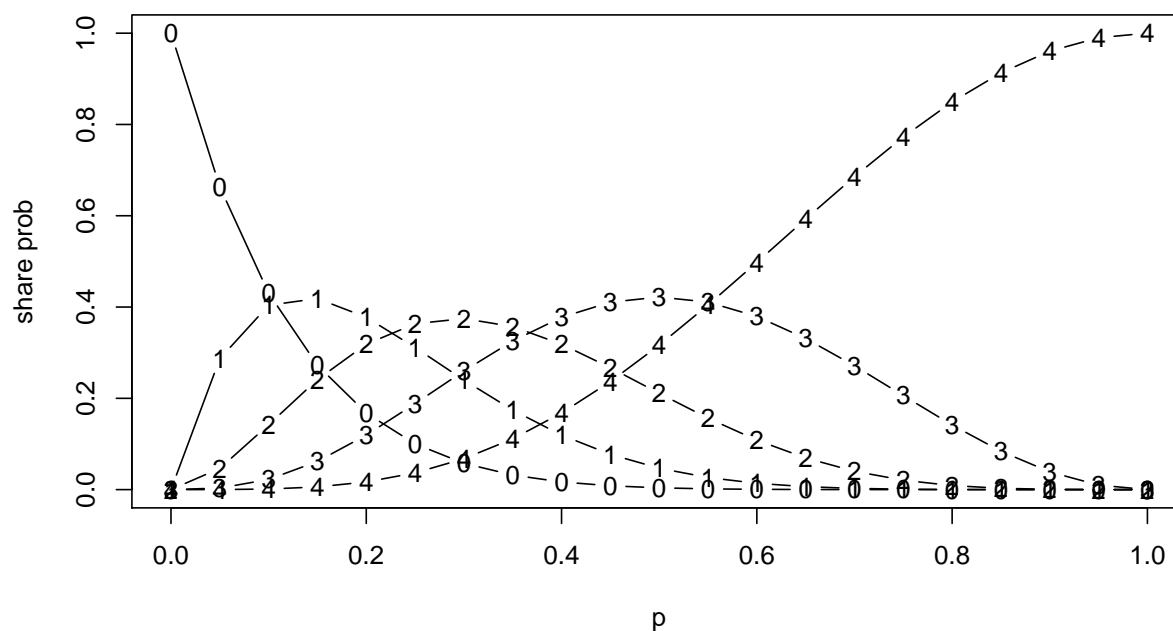


Figure 6: Sharing Probability

```
newick.chr1.med <- '(((tp3367_Italy:8813,tp1013_Wales:9652):9365,(((e_tp1007_Virginia:30,d_tp1012_Australia:61):19,
cat.hardwrap(newick.chr1.med)
```

```
# (((tp3367_Italy:8813,tp1013_Wales:9652):9365,(((e_tp1007_Virginia:30,d_tp1012_Au
# stralia:61):19,(c_tp1015_Puget_Sound:207,b_tp1335_NY:41):18):1005,a_tp1014_Gyre:
# 61):3912):7054,outgroup:0);
```

NOTE: In early code, tree was not being recalculated; it was defined by constants in the following code chunk, hand-copied from the analysis above.

```
# tree parameters as nested lists
# Internal nodes have subtrees sub1/2 and length
# Root has sub1/2, but no length
# Leaves have where, length, optionally, id, alt, nb. (Omit id for 'outgroup'. Use 'alt' for less formal
# labeling in cartoon version; it defaults to 'where'. Use 'nb' to add abcde annotations for legend.)
# This hand-made version is now subsumed by make.tree; retained for comparison
tree.by.hand <-
list(
  sub1 = list(
    sub1 = list(
      sub1 = list(id=3367, length=8813, where='Venice, Italy', alt='Venice'),
      sub2 = list(id=1013, length=9652, where='Wales, UK'),
      length=9365),
    sub2 = list(
      sub1 = list(
        sub1 = list(
          sub1 = list(id=1007, length=30, nb='e', where='Virginia, USA'),
          sub2 = list(id=1012, length=61, nb='d', where='Perth, W. Australia', alt='Perth'),
          length=19),
        sub2 = list(
          sub1 = list(id=1015, length=207, nb='c', where='Washington, USA', alt='Puget Sound'),
          sub2 = list(id=1335, length=41, nb='b', where='New York, USA', alt='NY'),
          length=18),
        length=1005),
      sub2 = list(id=1014, length=61, nb='a', where='N. Pacific Gyre'),
```

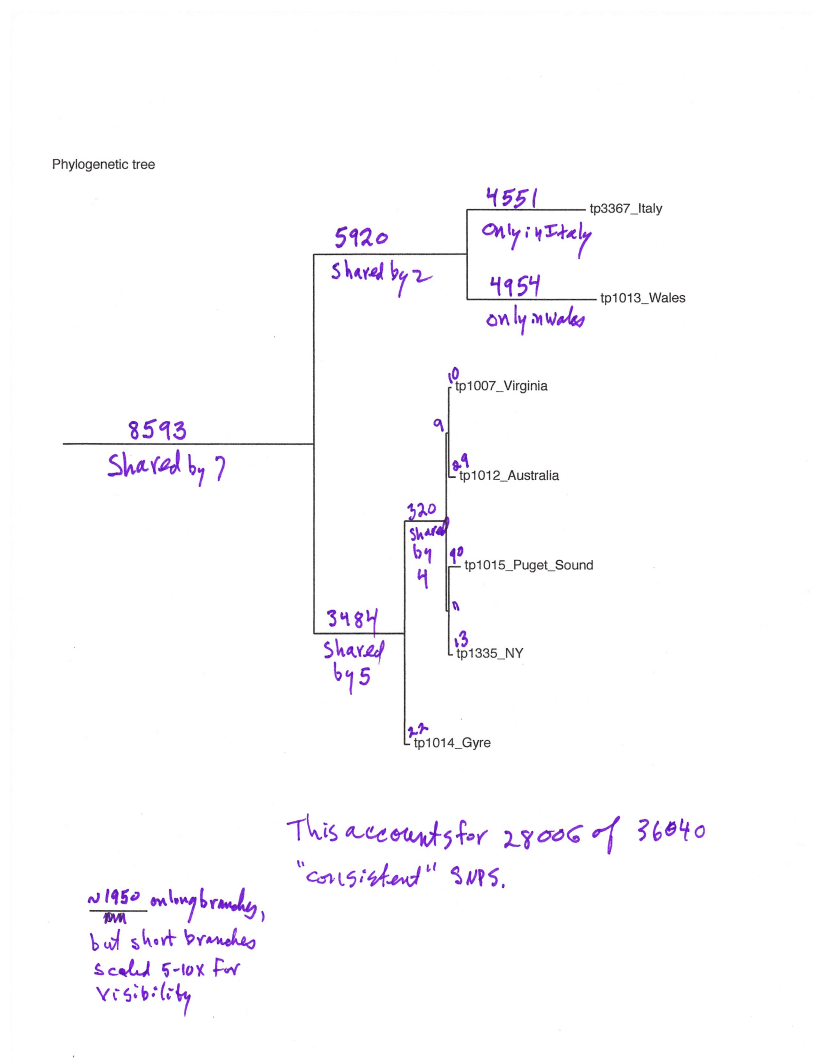


Figure 7: Inferred Tree, based on Chr1, un-q-filtered reads, loose criteria. (Note: to visually resolve the edges among the 5, their lengths were scaled by 5x – 10x in this figure, but not in the newick description shown in the text.)

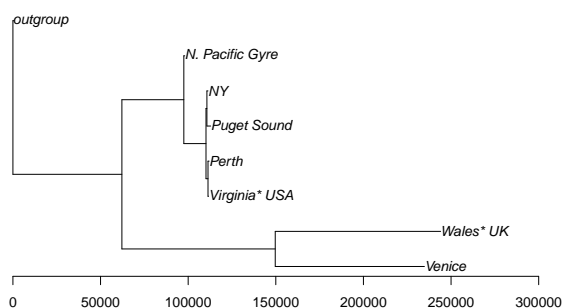


Figure 8: Tree based on qfiltered reads and medium SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs. (no edge labels, nolegend)

```
length=3912),
length=7054),
sub2 = list(length=0, where='outgroup')
)

# historical, format example, and debug help:
oldwick.medium <- '(((CCMP3367_Italy:8813,CCMP1013_Wales:9652):9365,((e_CCMP1007_Virginia:30,d_CCMP1012_Australia:61):19,(c_CCMP
# with simpler labeling for cartoon
simple.oldwick.medium <- '(((Italy:8813,Wales:9652):9365,((Virginia:30,Australia:61):19,(Puget:207,NY:41):18):1005,Gyre:61):3912
cat.hardwrap(oldwick.medium)

# (((CCMP3367_Italy:8813,CCMP1013_Wales:9652):9365,((e_CCMP1007_Virginia:30,d_CCM
# P1012_Australia:61):19,(c_CCMP1015_Puget_Sound:207,b_CCMP1335_NY:41):18):1005,a_
# CCMP1014_NPG:61):3912):7054,outgroup:0);

cat.hardwrap(simple.oldwick.medium)

# (((Italy:8813,Wales:9652):9365,((Virginia:30,Australia:61):19,(Puget:207,NY:41
# :18):1005,Gyre:61):3912):7054,outgroup:0);
```

Two other versions of the tree, for possible use in figs in the main paper.

Figure 8: [** as of 10/4/2015, this fig and next have stray stars on virginia, wales labels; probably due to hacking with commas in newick; not worth fixing unless we resurrect these trees for some purpose, but if so, see use of newick.name.undo in show.tree as probable fix. **]

```
tree.scale <- ifelse(which.snp.tables(string.val=F)[1]=='Chr1', 1, 10)
tree.x.lim <- 3e4 * tree.scale
the.simple.tree <- read.tree(text=simple.newick.medium)
plot(the.simple.tree, x.lim = tree.x.lim)
axis(1)
```

Figure 9:

```
plot(the.simple.tree, x.lim = tree.x.lim)
axis(1, (0:4)*7000*tree.scale, (0:4)*7000*tree.scale)
```

At some much earlier point, Tony ran the whole-genome version of the then-current code above, and manually entered tree branch lengths/legend for the resuting tree, shown in Fig 10. Code above can now automatically generate such a tree, but retain the following for comparison. The basic story seems clear—same topology and branch lengths scaled by about 10x, which is completely reasonable given that Chr1 is about 10% of the genome. Note that this tree is not being recalculated; it is defined by constants in the following code chunk.

```
fullgenome.newick.medium <- '(((3367_Italy:86155,1013_Wales:95697):89598,((e_1007_VA:330,d_1012_Australia:632):1296,(c_1015_WA:2
cat.hardwrap(fullgenome.newick.medium)
```

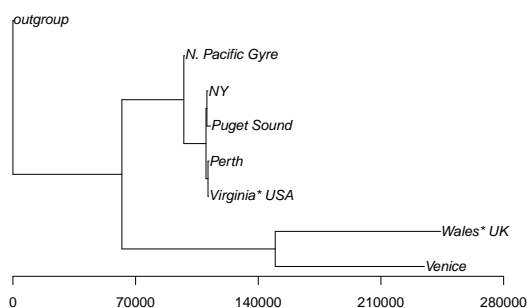


Figure 9: Tree based on qfiltered reads and medium SNP filters. “Lengths” are numbers of shared/private SNPs all Chrs. (no edge labels, no legend, short scale bar)

```
# ((3367_Italy:86155,1013_Wales:95697):89598,(((e_1007_VA:330,d_1012_Australia:63
# 2):1296,(c_1015_WA:2113,b_1335_NY:658):480):10059,a_1014_NPG:568):39517):69526,o
# utgroup:0);

legend.text <- c('a: only in 1014 ',
                'b: only in 1335 ',
                'c: only in 1015 ',
                'd: only in 1012 ',
                'e: only in 1007 ',
                '*: shared by bcde',
                '  shared by b/c ',
                '  shared by d/e ')

fullgenome.tree.x.lim <- 300000
fullgenome.counts <- c( 568, 658, 2113, 632, 330, 10059, 480, 1296 )
fullgenome.legend.text <- paste(legend.text,format(fullgenome.counts,width=5),sep=' - ')
fullgenome.tree.labels <- list( ## x,y,text
  41000,3.63,'69526\nshared by 7',
  90000,5.75,'39517\nby 5 (**)',
  115000,1.5, '89598\nshared by 2',
  210000,2.0, '95697 only\nin Wales',
  210000,1.0, '86155 only\nin Italy',
  113500,4.6, '*')
```

Figure 10:

```
library(ape)
the.fullgenome.tree <- read.tree(text=fullgenome.newick.medium)
plot(the.fullgenome.tree, x.lim = fullgenome.tree.x.lim)
axis(1) # ; axis(2) useful only for placing labels
opar <- par(family='mono',cex=.8)
legend('topright', legend=fullgenome.legend.text)
par(opar)
for(i in seq(1,length(fullgenome.tree.labels)-2,by=3)){
  text(fullgenome.tree.labels[[i]], fullgenome.tree.labels[[i+1]], fullgenome.tree.labels[[i+2]])
}
```

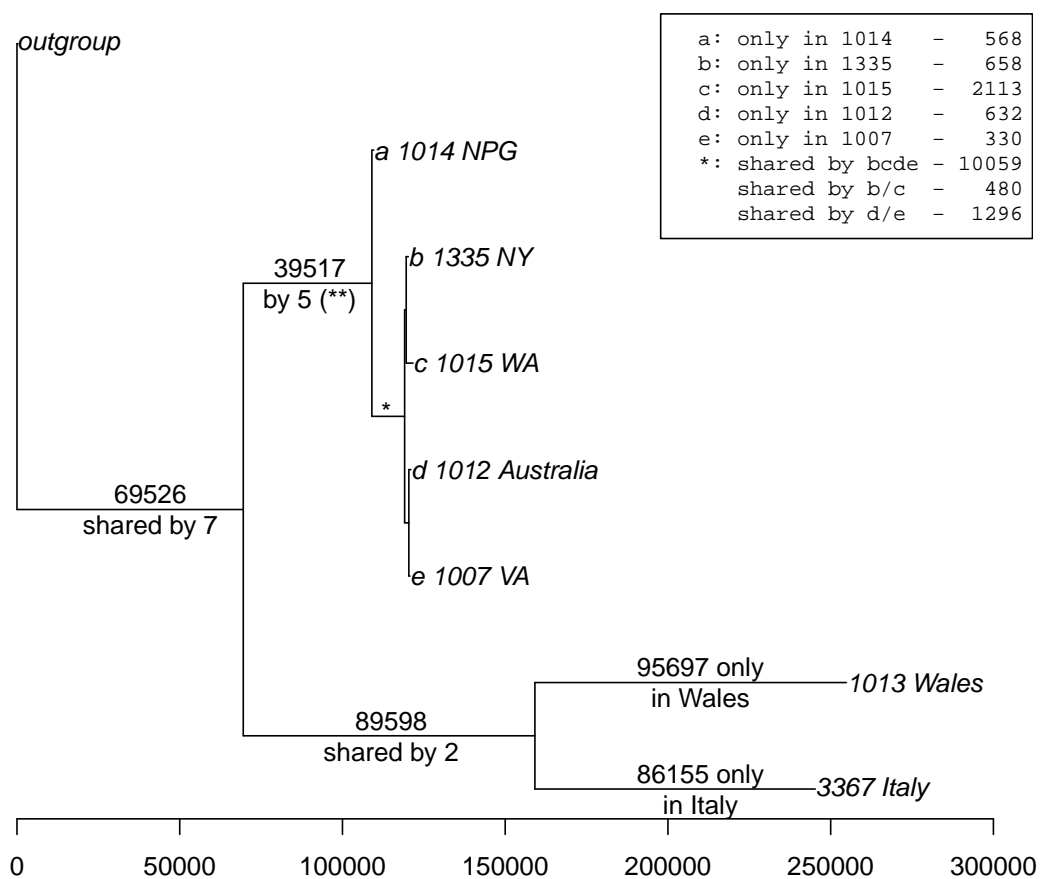



Figure 10: Tree based on unfiltered reads and medium SNP filters. “Lengths” are numbers of shared/private SNPs genome-wide. (By-hand legacy version)

