

Irrigation's real impact on global water and food security

R code

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```

# PRELIMINARY FUNCTIONS #####

sensobol::load_packages(c("openxlsx", "data.table", "tidyverse", "bibliometrix",
                          "igraph", "ggraph", "cowplot", "tidygraph", "benchmarkme",
                          "parallel", "wesanderson", "scales", "countrycode"))

# Create custom theme
theme_AP <- function() {
  theme_bw() +
    theme(panel.grid.major = element_blank(),
          panel.grid.minor = element_blank(),
          legend.background = element_rect(fill = "transparent",
                                            color = NA),
          legend.key = element_rect(fill = "transparent",
                                     color = NA),
          strip.background = element_rect(fill = "white"),
          legend.margin = margin(0.5, 0.1, 0.1, 0.1),
          legend.box.margin = margin(0.2, -4, -7, -7),
          plot.margin = margin(3, 4, 0, 4),
          legend.text = element_text(size = 8),
          axis.title = element_text(size = 10),
          legend.key.width = unit(0.4, "cm"),
          legend.key.height = unit(0.4, "cm"),
          legend.title = element_text(size = 9))
}

```

```

# CREATION OF VECTORS WITH NAMES #####

database <- c("wos", "scopus", "dimensions")
topic <- c("water", "food")

# Create all possible combinations
combinations <- expand.grid(database = database, topic = topic)

# Combine the vectors with an underscore
file.name <- paste(combinations$database, "dt", combinations$topic, sep = "_")

# READ IN THE DATA #####

# Loop to create the file names -----
for (i in 1:length(file.name)) {

  database.type <- str_extract(file.name, "(wos|scopus|dimensions)")

  if(isTRUE(database.type[i] == "wos")) {

    file.name[i] <- paste(file.name[i], "bib", sep = ".")

  } else {

    file.name[i] <- paste(file.name[i], "csv", sep = ".")

  }

}

# vector with new column names -----

new_colnames <- c("doi", "authors", "year", "title", "journal", "abstract", "database")
to_lower <- c("authors", "title", "journal", "abstract")

# Loop to read in the datasets -----

out <- list()

for (i in 1:length(file.name)) {

  database.type <- str_extract(file.name[i], "(wos|scopus|dimensions)")

  if(isTRUE(database.type == "wos")) {

    out[[i]] <- convert2df(file = file.name[i],
                        dbsource = "wos",

```

```

        format = "bibtex") %>%
data.table() %>%
.[, .(DI, AU, PY, TI, SO, AB)] %>%
.[, database:= "wos"]

} else if (isTRUE(database.type == "dimensions")) {

  out[[i]] <- fread(file.name[i], skip = 1) %>%
    .[, .(DOI, Authors, PubYear, Title, `Source title`, Abstract)] %>%
    .[, database:= "dimensions"]

} else if(isTRUE(database.type == "scopus")) {

  out[[i]] <- fread(file.name[i]) %>%
    .[, .(DOI, Authors, Year, Title, `Source title`, Abstract)] %>%
    .[, database:= "scopus"]
}

setnames(out[[i]], colnames(out[[i]]), new_colnames) %>%
.[, (to_lower):= lapply(.SD, tolower), .SDcols = (to_lower)] %>%
.[, abstract:= sub("references.*", "", abstract)]

}

```

```

##
## Converting your wos collection into a bibliographic dataframe
##
##
## Warning:
## In your file, some mandatory metadata are missing. Bibliometrix functions may not work properly
##
## Please, take a look at the vignettes:
## - 'Data Importing and Converting' (https://www.bibliometrix.org/vignettes/Data-Importing-and-Converting)
## - 'A brief introduction to bibliometrix' (https://www.bibliometrix.org/vignettes/Introduction-to-Bibliometrix)
##
##
## Missing fields:  C1 CR
## Done!
##
##
## Converting your wos collection into a bibliographic dataframe
##
##
## Warning:
## In your file, some mandatory metadata are missing. Bibliometrix functions may not work properly
##
## Please, take a look at the vignettes:

```

```
## - 'Data Importing and Converting' (https://www.bibliometrix.org/vignettes/Data-Importing-and-Converting)
## - 'A brief introduction to bibliometrix' (https://www.bibliometrix.org/vignettes/Introduction-to-Bibliometrix)
##
##
## Missing fields: C1 CR
## Done!
```

```
names(out) <- combinations$topic

# CLEAN THE DATASETS #####

# Arrange -----

dt <- rbindlist(out, idcol = "topic")

tmp <- split(dt, list(dt$topic, dt$database))

cols_to_merge_by <- c("doi", "year", "title", "journal", "abstract")

dt.water <- merge(merge(tmp$water.dimensions, tmp$water.scopus, by = cols_to_merge_by,
                        all = TRUE), tmp$water.wos, by = cols_to_merge_by,
                  all = TRUE)

dt.food <- merge(merge(tmp$food.dimensions, tmp$food.scopus, by = cols_to_merge_by,
                      all = TRUE), tmp$food.wos, by = cols_to_merge_by,
                 all = TRUE)

# Filter out duplicated studies by doi -----

tmp.list <- list(dt.water, dt.food)
duplicated.dois <- final.dt <- list()

for (i in 1:length(tmp.list)) {

  duplicated.dois[[i]] <- duplicated(tmp.list[[i]]$doi, incomparables = NA, na.rm = TRUE)
  final.dt[[i]] <- tmp.list[[i]][!duplicated.dois[[i]][, location.belief.system := "abstract"]]

}

names(final.dt) <- topic

# Check if there is any duplicated doi -----

any(duplicated(final.dt$food$doi, na.rm = TRUE, incomparables = NA))

## [1] FALSE

# Export to xlsx -----
```

```
for (i in names(final.dt)) {

  write.xlsx(final.dt[[i]][, .(doi, year, title, abstract, location.belief.system)],
            paste("final.dt", names(final.dt[i]), "xlsx", sep = "."))
}
```

1 Retrieve all corpus

1.1 Abstract corpus

```
final.dt.water.screened <- data.table(read.xlsx("final.dt.water_screened.xlsx"))
final.dt.food.screened <- data.table(read.xlsx("final.dt.food_screened.xlsx"))
screened.dt <- list(final.dt.water.screened, final.dt.food.screened)
names(screened.dt) <- c("water", "food")
```

```
lapply(screened.dt, function(x) x[, .N, screening])
```

```
## $water
##   screening      N
##   <char> <int>
## 1:      F    168
## 2:      T    163
##
## $food
##   screening      N
##   <char> <int>
## 1:      F    465
## 2:      T     39
```

```
# Export for close-reading only the references that do include
# the belief system in the abstract -----
```

```
for (i in names(screened.dt)) {

  screened.dt[[i]][screening == "T"] %>%
    unique(., by = "title") %>%
    .[, .(doi, title, year)] %>%
    write.xlsx(., paste("abstract.corpus", i, "xlsx", sep = "."))

}
```

1.2 Policy corpus

```
# LOAD IN DIMENSIONS DATASETS (POLICY TEXT) #####

# Function to load and preprocess data -----
```

```

load_and_preprocess_data <- function(file_path, topic) {
  fread(file_path, skip = 1)[, topic := topic]
}

colnames.full.text <- c("doi", "year", "title", "journal", "topic")
keywords <- c("water", "irrigat")

# Load data -----

dt.policy.water <- load_and_preprocess_data("dimensions_dt_policy.csv", "water")
dt.policy.food <- load_and_preprocess_data("dimensions_dt_policy_food.csv", "food")

dimensions.full.text.policy <- rbind(dt.policy.food, dt.policy.water) %>%
  .[, .(`Policy document ID`, PubYear, Title, `Publishing Organization`,
    `Sustainable Development Goals`, `Source Linkout`, topic)]

dimensions.full.text.policy[, .N, topic]

##      topic      N
##      <char> <int>
## 1:   food 10573
## 2:  water  3455

# Create a logical condition for pattern matching using grepl
pattern_condition_policy <- sapply(keywords, function(keyword)
  grepl(keyword, dimensions.full.text.policy$Title, ignore.case = TRUE))

# Combine conditions with OR using rowSums
matching.rows.policy <- dimensions.full.text.policy[rowSums(pattern_condition_policy) > 0]

matching.rows.policy[, .N, topic]

##      topic      N
##      <char> <int>
## 1:   food   750
## 2:  water   450

# Export -----

for (i in c("water", "food")) {
  matching.rows.policy[topic == i] %>%
    write.xlsx(paste("policy.corpus", i, "xlsx", sep = "."))
}

```

1.3 Full text corpus

```
# LOAD IN DIMENSIONS DATASET (FULL TEXT) #####
```

```
full.text.corpus.water <- fread("full.text.corpus.water.csv")
```

2 Split full text corpus for analysis

```
# SPLIT THE DATASET INTO N FOR RESEARCH #####
```

```
# Function to split dataset in n chunks -----
```

```
split_dt_fun <- function(dt, num_parts) {
```

```
  split_dt <- list()
```

```
  # Calculate the number of rows in each part
```

```
  rows_per_part <- nrow(dt) %/% num_parts
```

```
  # Split the data.table into roughly equal parts
```

```
  for (i in 1:num_parts) {
```

```
    start_row <- (i - 1) * rows_per_part + 1
```

```
    end_row <- i * rows_per_part
```

```
    if (i == num_parts) {
```

```
      end_row <- nrow(dt)
```

```
    }
```

```
    split_dt[[i]] <- dt[start_row:end_row, ]
```

```
  }
```

```
  return(split_dt)
```

```
}
```

```
# Create the datasets for close reading -----
```

```
times.nanxin <- 2
```

```
times.arnald <- 1
```

```
nanxin <- paste(rep("nanxin", times.nanxin), 1:times.nanxin, sep = "")
```

```
arnald <- paste(rep("arnald", times.arnald), 1:times.arnald, sep = "")
```

```
names_surveyors <- c(arnald, nanxin, "seth", paste("student", 1:4, sep = ""))
```

```
n_surveyors <- length(names_surveyors)
```

```
survey.dt.split <- split_dt_fun(dt = full.text.corpus.water, num_parts = n_surveyors)
```

```
names(survey.dt.split) <- names_surveyors
```



```

# Export -----

for (i in 1:length(survey.dt.split)) {

  write.xlsx(survey.dt.split[[i]],
             file = paste0(names(survey.dt.split)[i], ".dt", ".xlsx"))

}

```

3 Network analysis

```

# CREATE VECTORS TO READ IN AND CLEAN THE DATASETS #####

tmp <- list()
names.files <- c("WORK", "NETWORK")
topics <- c("water", "food")
corpus <- c("abstract.corpus", "policy.corpus", "full.text.corpus")
cols_of_interest <- c("title", "author", "claim", "citation")

# Paste all possible combinations of names -----

combs <- expand.grid(corpus = corpus, topics = topics, approach = names.files)
all.files <- paste(paste(paste(combs$corpus, combs$topics, sep = "."), combs$approach, sep = ".
                        "xlsx", sep = "."))

# READ IN DATASETS AND TURN TO LOWERCAPS #####

tmp <- list()

for (i in 1:length(all.files)) {

  tmp[[i]] <- data.table(read.xlsx(all.files[i]))

  if (!str_detect(all.files[i], "NETWORK")) {

    tmp[[i]][, title:= tolower(title)]

  } else {

    tmp[[i]][, (cols_of_interest):= lapply(.SD, tolower), .SDcols = (cols_of_interest)]
  }
}

names(tmp) <- all.files

sub(".*\\.([^\.\.]+)_.*", "\\1", all.files)

```

```
## [1] "water" "water" "water" "food" "food" "food" "water" "water" "water"
## [10] "food" "food" "food"

# CLEAN AND MERGE DATASETS #####

# Work datasets -----

dataset.works <- all.files[str_detect(all.files, "_WORK")]
dataset.works.topics <- sub(".*\\.(\\.[^\\.]*)_.*", "\\1", dataset.works)

tmp.works <- tmp[dataset.works]
names(tmp.works) <- dataset.works.topics
lapply(tmp.works, function(dt) dt[, .(doi, title, claim.in.text)]) %>%
  rbindlist(., idcol = "topic") %>%
  .[, .N, .(topic, claim.in.text)]

##      topic claim.in.text      N
##      <char>      <char> <int>
## 1:  water              F    715
## 2:  water             <NA>    402
## 3:  water              T    471
## 4:  water      Paywalled      9
## 5:  water        Russian      1
## 6:  water        French      1
## 7:  water         Indian      1
## 8:  water      Ukranian      1
## 9:  water    Portuguese      1
## 10:  food             <NA>     82
## 11:  food              T      33
## 12:  food              F    524

# Network datasets -----

dataset.networks <- all.files[str_detect(all.files, "NETWORK")]
dataset.networks.topics <- sub(".*\\.(\\.[^\\.]*)_.*", "\\1", dataset.networks)

tmp2 <- tmp[dataset.networks]
names(tmp2) <- dataset.networks.topics

network.dt <- rbindlist(tmp2, idcol = "topic") %>%
  .[, policy:= grepl("^policy", doi)]

# Retrieve year -----

network.dt[, year:= as.integer(sub(".* (\\d{4})[a-z]?$", "\\1", author))]

## Warning in eval(jsub, SEnv, parent.frame()): NAs introduced by coercion

# move policy to author -----
```

```

network.dt[, author:= ifelse(policy == TRUE, doi, author)]

# CHECK NUMBER OF FAO AQUASTAT CITES #####

aquastat.cites <- network.dt[citation %like% "fao aquastat"] %>%
  .[, .N, .(citation, topic)]

aquastat.cites

##           citation  topic      N
##           <char> <char> <int>
##  1: fao aquastat 2006  water      1
##  2:      fao aquastat  water     12
##  3: fao aquastat 2010  water      5
##  4: fao aquastat 2020  water      3
##  5: fao aquastat 2011  water      2
##  6: fao aquastat 2012  water      4
##  7: fao aquastat 2021  water      2
##  8: fao aquastat 2017  water      1
##  9: fao aquastat 2015  water      2
## 10: fao aquastat 2019  water      3
## 11: fao aquastat 2016  water      6
## 12: fao aquastat 2014  water      1
## 13: fao aquastat 2023  water      1
## 14: fao aquastat 2018  water      1
## 15: fao aquastat 2004  water      1
## 16: fao aquastat 2005  water      1
## 17:      fao aquastat  food      5

oldest.aquastat.cite <- min(as.integer(sub(".* (\\d{4})[a-z]?$", "\\1",
                                           aquastat.cites$citation)),
                             na.rm = TRUE)

## Warning: NAs introduced by coercion

# WRITE LOOKUP TABLE TO CHECK ALREADY RETRIEVED STUDIES #####

lookup.dt <- network.dt[, .(doi, title, author, topic)] %>%
  .[order(title)] %>%
  unique(.)

lookup.dt[, .(number.rows = nrow(.SD)), topic]

##      topic number.rows
##      <char>      <int>
## 1:  water          779
## 2:   food           48

```

```

# Export lookup tables -----

write.xlsx(lookup.dt, "lookup.dt.xlsx")
write.xlsx(lookup.dt[topic == "water"], "lookup.water.dt.xlsx")
write.xlsx(lookup.dt[topic == "food"], "lookup.food.dt.xlsx")

# Remove the year from mentions to FAO Aquastat -----

pattern <- "\\b(?:19|20)\\d{2}\\b" # Matches years between 1900 and 2099

for (col in c("citation", "author")) {
  matches <- grepl("^fao aquastat\\s+\\d+$", network.dt[[col]], ignore.case = TRUE)
  network.dt[matches, (col) := gsub("\\d+", "", network.dt[[col]][matches], perl = TRUE)]
  network.dt[, (col) := trimws(network.dt[[col]])]
}

# Rename columns -----

setnames(network.dt, c("author", "citation"), c("from", "to"))

# Rename category -----

network.dt[, category:= ifelse(!classification == "F", "Uncertain", "Fact")]

# Create copy and remove duplicated -----

network.dt.claim <- copy(network.dt)
network.dt.claim <- unique(network.dt.claim,
                           by = c("from", "to", "document.type", "nature.claim"))

fwrite(network.dt.claim, "network.dt.claim.csv")

# Convert all to lower caps -----

network.dt <- network.dt[, .(from, to, year, document.type, nature.claim,
                             classification, category, topic)]
cols_to_change <- colnames(network.dt)
network.dt[, (cols_to_change):= lapply(.SD, trimws), .SDcols = (cols_to_change)]

# PLOT DESCRIPTIVE STATISTICS #####

total.rows <- network.dt[, .(number.rows = nrow(.SD)), topic]

# Check proportion of studies by nature of claim -----

network.dt.claim[, .N, .(nature.claim, topic)] %>%
  merge(., total.rows, by = "topic") %>%

```

```

[, fraction:= N / number.rows] %>%
print()

## Key: <topic>
##      topic      nature.claim      N number.rows      fraction
##      <char>      <char> <int>      <int>      <num>
## 1:   food      no citation    22        51 0.43137255
## 2:   food citation backup    19        51 0.37254902
## 3:   food      no claim      2        51 0.03921569
## 4:   food      <NA>          1        51 0.01960784
## 5: water citation backup   534       921 0.57980456
## 6: water      modelling     21       921 0.02280130
## 7: water      no citation   257       921 0.27904452
## 8: water      <NA>         29       921 0.03148751
## 9: water      no claim     72       921 0.07817590

# Count document type by nature of claim -----

a <- network.dt[, .N, .(nature.claim, document.type, topic)] %>%
merge(., total.rows, by = "topic") %>%
[, proportion:= N / number.rows] %>%
na.omit() %>%
ggplot(., aes(reorder(nature.claim, proportion), proportion)) +
coord_flip() +
geom_bar(stat = "identity") +
facet_grid(topic~document.type) +
scale_y_continuous(breaks = breaks_pretty(n = 2)) +
labs(x = "", y = "Fraction") +
theme_AP()

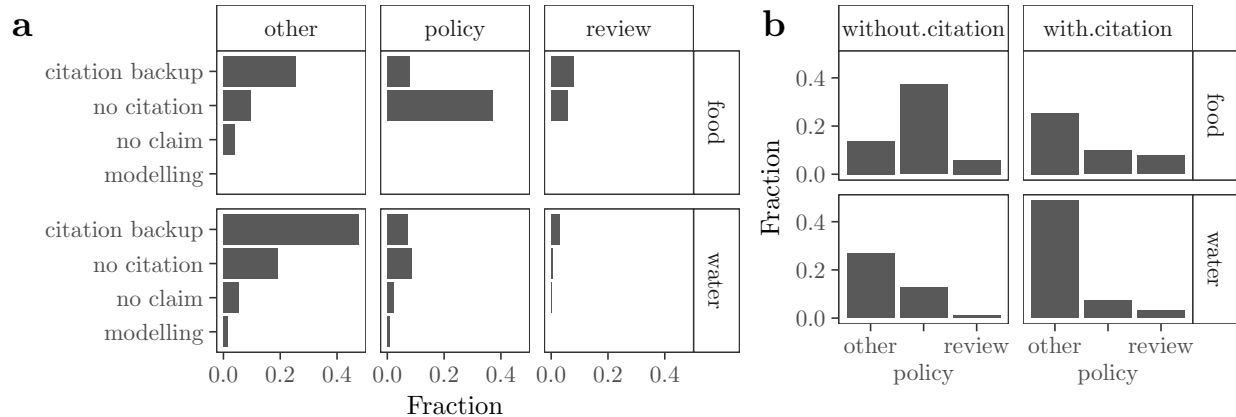
# Count how many documents make the claim and cite / do not cite,
# by document.type -----

b <- network.dt[, .(without.citation = sum(is.na(to)),
                    with.citation = .N - sum(is.na(to))), .(document.type, topic)] %>%
melt(., measure.vars = c("without.citation", "with.citation")) %>%
merge(., total.rows, by = "topic") %>%
[, proportion:= value / number.rows] %>%
ggplot(., aes(document.type, proportion)) +
geom_bar(stat = "identity") +
scale_y_continuous(breaks = breaks_pretty(n = 2)) +
scale_x_discrete(guide = guide_axis(n.dodge = 2)) +
labs(x = "", y = "Fraction") +
facet_grid(topic~variable) +
theme_AP()

# merge -----

```

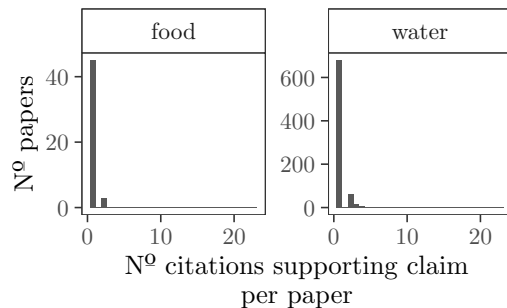
```
plot_grid(a, b, ncol = 2, rel_widths = c(0.6, 0.4), labels = "auto")
```



PLOT DISTRIBUTION OF CITATION SUPPORTING THE CLAIM

```
network.dt[, .N, .(from, topic)] %>%
  .[order(-N)] %>%
  ggplot(., aes(N)) +
  geom_histogram() +
  facet_wrap(~topic, scale = "free_y") +
  scale_x_continuous(breaks = breaks_pretty(n = 3)) +
  scale_y_continuous(breaks = breaks_pretty(n = 3)) +
  theme_AP() +
  labs(x = "N° citations supporting claim \n per paper", y = "N° papers")
```

`stat_bin()` using `bins = 30`. Pick better value with `binwidth`.



3.1 Network metrics

CALCULATE NETWORK METRICS

only complete cases -----

```
network.dt.complete <- network.dt[complete.cases(network.dt$to), ]
split.networks <- split(network.dt.complete, network.dt.complete$topic)
```

Export-----

```

write.xlsx(network.dt.complete, "network.dt.complete.xlsx")

# Transform to graph -----

citation_graph <- lapply(split.networks, function(dt)
  graph_from_data_frame(d = dt, directed = TRUE))

## Warning in graph_from_data_frame(d = dt, directed = TRUE): In `d` `NA` elements
## were replaced with string "NA"

# Calculate network metrics -----

lapply(citation_graph, function(x) edge_density(x))

## $food
## [1] 0.02083333
##
## $water
## [1] 0.001534518

# Modularity:
# - c.1: Strong community structure, where nodes within groups are highly connected.
# - c. -1: Opposite of community structure, where nodes between groups are more connected.
# - c. 0: Indicates absence of community structure or anti-community structure in the network.
wtc <- lapply(citation_graph, function(x) cluster_walktrap(x))
lapply(wtc, function(x) modularity(x))

## $food
## [1] 0.7944215
##
## $water
## [1] 0.8471048

network_metrics <- lapply(citation_graph, function(x)
  data.table(node = V(x)$name,

    # Degree of a node: The number of connections or
    # edges linked to that node.
    # It represents how well-connected or central a
    # node is within the graph.
    degree = degree(x, mode = "in"),

    degree.out = degree(x, mode = "out"),

    # Betweenness centrality of a node: Measures the
    # extent to which a node lies on the shortest
    # paths between all pairs of other nodes in the graph.
    # Nodes with high betweenness centrality act as
    # bridges or intermediaries, facilitating

```

```

# communication and information flow between other nodes.
betweenness = betweenness(x),

# Closeness centrality of a node: Measures how
# close a node is to all other nodes in the graph,
# taking into account the length of the shortest paths.
# Nodes with high closeness centrality are able to
# efficiently communicate or interact with other
# nodes in the graph.
closeness = closeness(x),
pagerank = page_rank(x)$vector)
)

# Define the max number of rows
max.number <- 3

degree.nodes <- lapply(network_metrics, function(dt) dt[order(-degree)][1:max.number])
degree.nodes.out <- lapply(network_metrics, function(dt) dt[order(-degree.out)][1:max.number])
betweenness.nodes <- lapply(network_metrics, function(dt) dt[order(-betweenness)][1:max.number])
pagerank.nodes <- lapply(network_metrics, function(dt) dt[order(-closeness)][1:max.number])

degree.nodes

```

```

## $food
##           node degree degree.out betweenness closeness  pagerank
##           <char>  <num>      <num>      <num>      <num>      <num>
## 1:      fao aquastat      5          0          0      NaN 0.09168305
## 2: meier et al 2018      1          1          1 1.0000000 0.02743993
## 3:  wang et al 2012      1          1          2 0.3333333 0.03562376
##
## $water
##           node degree degree.out betweenness closeness  pagerank
##           <char>  <num>      <num>      <num>      <num>      <num>
## 1:      fao aquastat     46          0      0.00000      NaN 0.063790805
## 2: siebert et al 2010     12          3     37.33333 0.3333333 0.008580413
## 3:  molden et al 2007     10          1     22.00000 0.3333333 0.009077773

```

```
degree.nodes.out
```

```

## $food
##           node degree degree.out betweenness closeness  pagerank
##           <char>  <num>      <num>      <num>      <num>      <num>
## 1:      niu et al 2023      0          2          0      0.25 0.01925609
## 2: borsato et al 2020      0          2          0      0.50 0.01925609
## 3:      borin 2023      0          2          0      0.25 0.01925609
##
## $water
##           node degree degree.out betweenness  closeness  pagerank

```



```
##           <char> <num>         <num>         <num>         <num>         <num>
## 1:      wada 2015      0          23          0 0.02702703 0.0009265398
## 2: wada et al 2014      1           9         10 0.09090909 0.0010840515
## 3: wada et al 2016      0           8          0 0.06250000 0.0009265398
```

```
betweenness.nodes
```

```
## $food
##           node degree degree.out betweenness closeness pagerank
##           <char> <num>         <num>         <num>         <num>         <num>
## 1:      wang et al 2012      1           1          2 0.33333333 0.03562376
## 2: hanjra and qureshi 2010      1           1          2 1.00000000 0.04953629
## 3:      meier et al 2018      1           1          1 1.00000000 0.02743993
```

```
## $water
##           node degree degree.out betweenness closeness pagerank
##           <char> <num>         <num>         <num>         <num>         <num>
## 1: boretti and rosa 2019      4           4         44.000000 0.05555556 0.003682996
## 2:      siebert et al 2010     12           3         37.333333 0.33333333 0.008580413
## 3:      molden et al 2007     10           1         22.000000 0.33333333 0.009077773
```

```
pagerank.nodes
```

```
## $food
##           node degree degree.out betweenness closeness pagerank
##           <char> <num>         <num>         <num>         <num>         <num>
## 1: okorogbona et.al 2018      0           1          0          1 0.01925609
## 2:      du preez et al 2018      0           1          0          1 0.01925609
## 3:      meier et al 2018      1           1          1          1 0.02743993
```

```
## $water
##           node degree degree.out betweenness closeness pagerank
##           <char> <num>         <num>         <num>         <num>         <num>
## 1: sharma and irmak 2012      0           1          0          1 0.0009265398
## 2:      world bank 2007      3           1         11          1 0.0076566564
## 3:      brajovic et al 2015      0           1          0          1 0.0009265398
```

3.2 Network plots

```
# ADD FEATURES TO NODES #####
```

```
# Retrieve a vector with the node names -----
```

```
graph <- lapply(split.networks, function(nt)
  tidygraph::as_tbl_graph(nt, directed = TRUE))
```

```
## Warning in graph_from_data_frame(x, directed = directed): In `d` `NA` elements
## were replaced with string "NA"
```

```

vec.names <- lapply(graph, function(graph)
  graph %>%
    activate(nodes) %>%
    pull() %>%
    data.table(name = .))

# Merge with info from the network.dt -----

tmp.network <- split(network.dt, network.dt$topic)

vec.nature.claim <- list()

for(i in names(tmp.network)) {

  vec.nature.claim[[i]] <- merge(merge(vec.names[[i]], unique(tmp.network[[i]][, .(from, year,
    by.x = "name", by.y = "from", all.x = TRUE),
    unique(tmp.network[[i]][, .(from, document.type, classification)
    by.x = "name", by.y = "from", all.x = TRUE)
})

# Merge with the correct order -----

order_indices <- final.vec.nature.claim <- final.vec.document.type <-
  final.vec.year <- final.vec.classification <- final.vec.category <- list()

for (i in names(vec.names)) {

  order_indices[[i]] <- match(vec.names[[i]]$name, vec.nature.claim[[i]]$name)
  final.vec.nature.claim[[i]] <- vec.nature.claim[[i]][order_indices[[i]], ] %>%
    .[, nature.claim]
  final.vec.document.type[[i]] <- vec.nature.claim[[i]][order_indices[[i]], ] %>%
    .[, document.type]
  final.vec.year[[i]] <- vec.nature.claim[[i]][order_indices[[i]], ] %>%
    .[, year] %>%
    as.numeric()
  final.vec.classification[[i]] <- vec.nature.claim[[i]][order_indices[[i]], ] %>%
    .[, classification]
  final.vec.category[[i]] <- vec.nature.claim[[i]][order_indices[[i]], ] %>%
    .[, category]
}

# Attach to the graph -----

graph.final <- list()

for (i in names(graph)) {

```

```

graph.final[[i]] <- graph[[i]] %>%
  activate(nodes) %>%
  mutate(nature.claim = final.vec.nature.claim[[i]],
         document.type = final.vec.document.type[[i]],
         year = final.vec.year[[i]],
         degree = network_metrics[[i]]$degree,
         classification = final.vec.classification[[i]],
         category = final.vec.category[[i]],
         degree.out = network_metrics[[i]]$degree.out,
         betweenness = network_metrics[[i]]$betweenness,
         pagerank = network_metrics[[i]]$pagerank)
}

# NUMBER OF NODES #####

lapply(graph.final, function(graph) V(graph))

## $food
## + 33/33 vertices, named, from c33b012:
## [1] okorogbona et.al 2018      du preez et al 2018
## [3] niu et al 2023            meier et al 2018
## [5] lobell et al 2006         rosa 2022
## [7] rolle et al 2021          mitchell et al 2018
## [9] wang et al 2012           hanjra and qureshi 2010
## [11] de pascale et al 2011     borsato et al 2020
## [13] borin 2023                turrall et al 2010
## [15] meier et al 2017          policy.1666264
## [17] policy.1869122            policy.1898497
## [19] policy.1667934            fernandez-cirelli et al 2009
## + ... omitted several vertices
##
## $water
## + 597/597 vertices, named, from 43d1a43:
## [1] sharma and irmak 2012
## [2] doreau et al 2012
## [3] world water assessment programme 2009
## [4] world bank 2007
## [5] brajovic et al 2015
## [6] rivers et al 2015
## [7] kijne 2005
## [8] hafeez and khalid awan 2022
## [9] dunkelman et al 2017
## [10] nordin et al 2013
## + ... omitted several vertices

```

```
# NUMBER OF EDGES #####
```

```
lapply(graph.final, function(graph) ecount(graph))
```

```
## $food  
## [1] 22  
##  
## $water  
## [1] 546
```

```
# PROPORTION OF ALL PATHS THAT PASS THROUGH FIVE HIGHEST BETWEENNESS NODES #####
```

```
lapply(graph.final, function(graph) {  
  
  bc <- betweenness(graph)  
  nodes_of_interest <- sort(bc, decreasing = TRUE)[1:5]  
  total_paths <- choose(vcount(graph), 2) # Total number of paths  
  total_paths  
  sum(nodes_of_interest) / total_paths  
  
})
```

```
## $food  
## [1] 0.01136364  
##  
## $water  
## [1] 0.0007789694
```

```
# PROPORTION OF LINKS CONNECTED TO THE 5 NODES WITH HIGHEST DEGREE #####
```

```
lapply(graph.final, function(graph) {  
  
  dg <- degree(graph)  
  nodes_of_interest_degree <- sort(dg, decreasing = TRUE)[1:5]  
  total_edges <- ecount(graph) # Total number of edges  
  sum(nodes_of_interest_degree) / total_edges  
  
})
```

```
## $food  
## [1] 0.5909091  
##  
## $water  
## [1] 0.1941392
```

```
# PLOT NETWORK #####
```

```
seed <- 123  
selected_colors <- c("darkblue", "lightgreen", "orange", "red", "grey")
```

```

# by nature of claim -----

# Label the nodes with highest degree -----

p1 <- p2 <- p3 <- p4 <- list()

for(i in names(graph.final)) {

  set.seed(seed)

  p1[[i]] <- ggraph(graph.final[[i]], layout = "igraph", algorithm = "nicely") +
    geom_edge_link(arrow = arrow(length = unit(1.8, 'mm')),
      end_cap = circle(1, "mm")) +
    geom_node_point(aes(color = nature.claim, size = degree)) +
    geom_node_text(aes(label = ifelse(degree >= min(degree.nodes[[i]]$degree), name, NA)),
      repel = TRUE, size = 2.2) +
    labs(x = "", y = "") +
    scale_color_manual(name = "",
      values = selected_colors) +
    theme_AP() +
    theme(axis.text.x = element_blank(),
      axis.ticks.x = element_blank(),
      axis.text.y = element_blank(),
      axis.ticks.y = element_blank(),
      legend.position = "right")
}

p1

```

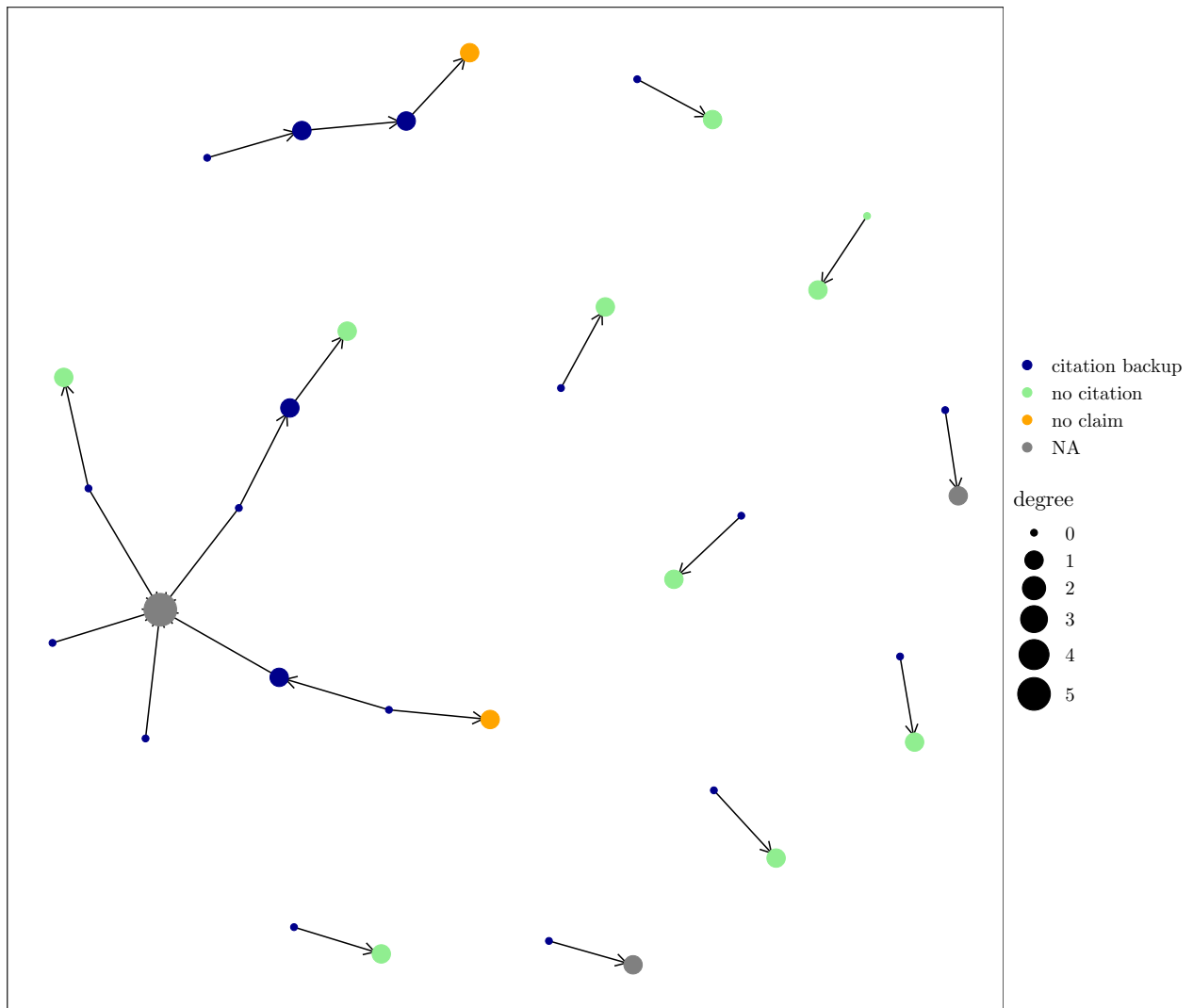
```
## $food
```

```

## Warning: Using the `size` aesthetic in this geom was deprecated in ggplot2 3.4.0.
## i Please use `linewidth` in the `default_aes` field and elsewhere instead.
## This warning is displayed once every 8 hours.
## Call `lifecycle::last_lifecycle_warnings()` to see where this warning was
## generated.

## Warning: Removed 33 rows containing missing values (`geom_text_repel()`).

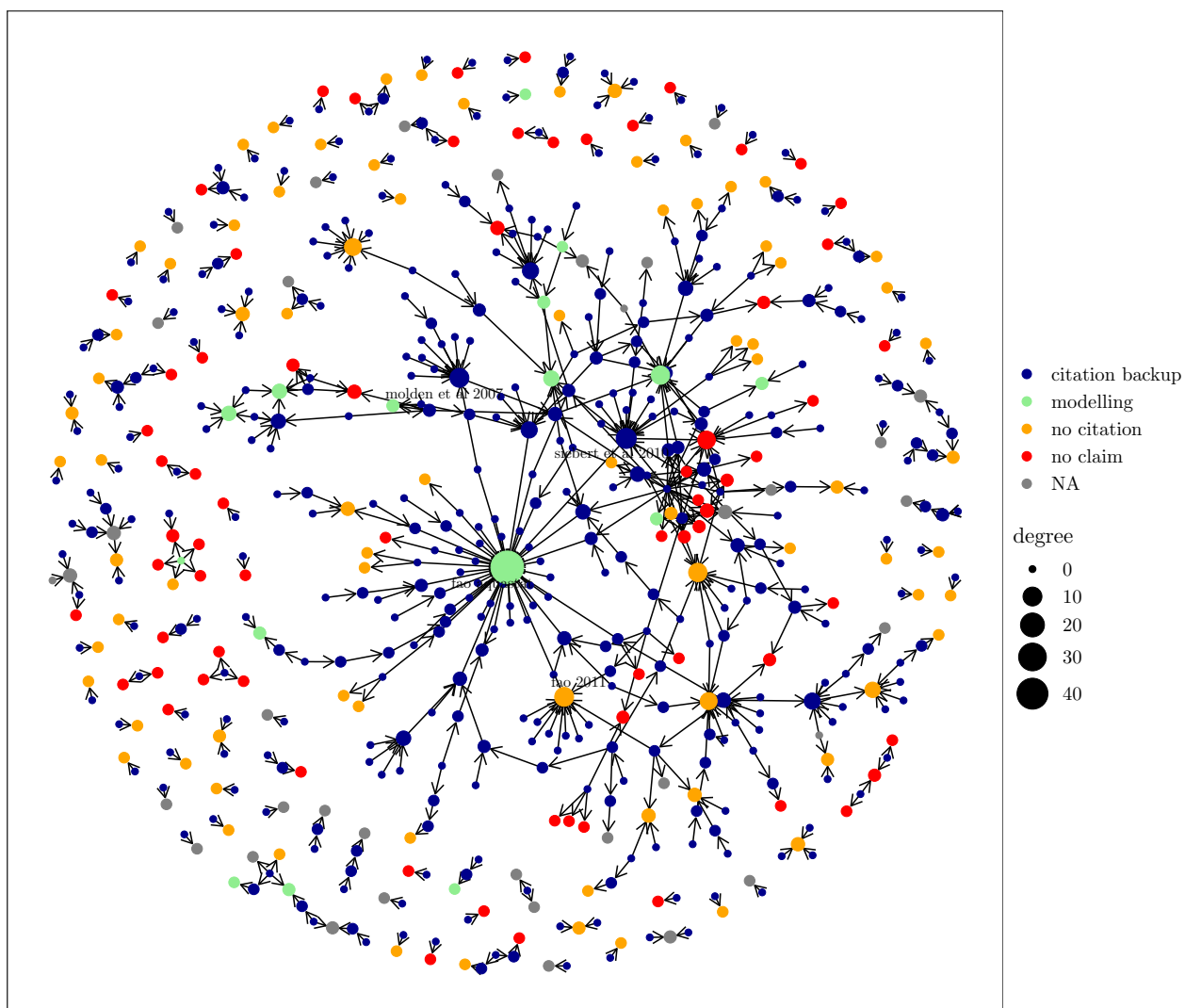
```



##

\$water

Warning: Removed 593 rows containing missing values (`geom_text_repel()`).



```
# Label the nodes with highest betweenness -----
for (i in names(graph.final)) {
  set.seed(seed)

  p2[[i]] <- ggraph(graph.final[[i]], layout = "igraph", algorithm = "nicely") +
    geom_edge_link(arrow = arrow(length = unit(1.8, 'mm')),
      end_cap = circle(1, "mm")) +
    geom_node_point(aes(color = nature.claim, size = betweenness)) +
    geom_node_text(aes(label = ifelse(betweenness >= min(betweenness.nodes[[i]]$betweenness),
      name, NA)),
      repel = TRUE, size = 2.2) +
    labs(x = "", y = "") +
    scale_color_manual(name = "",
      values = selected_colors) +
    theme_AP() +

```

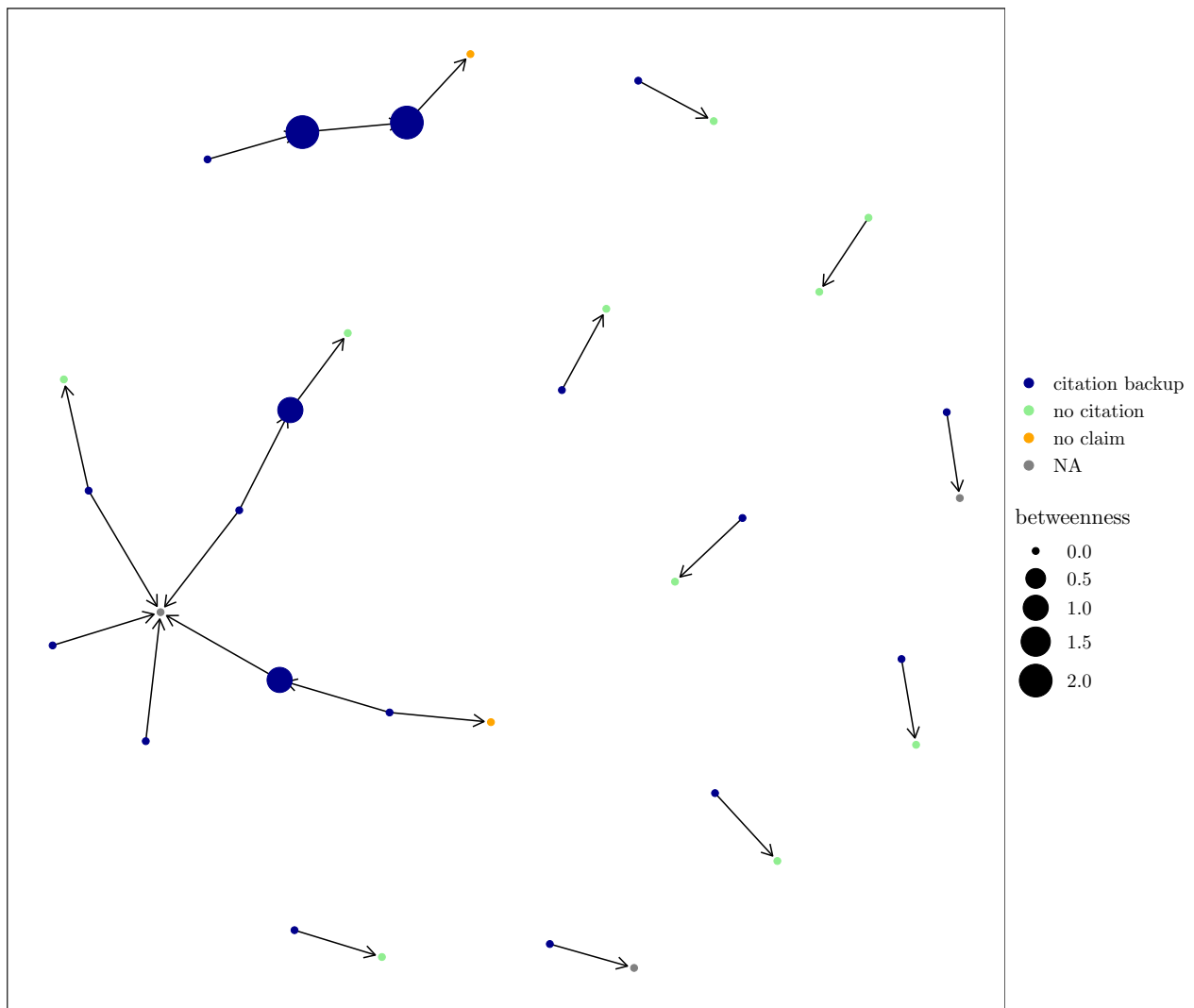
```

    theme(axis.text.x = element_blank(),
          axis.ticks.x = element_blank(),
          axis.text.y = element_blank(),
          axis.ticks.y = element_blank(),
          legend.position = "right")
  }
  p2

```

```
## $food
```

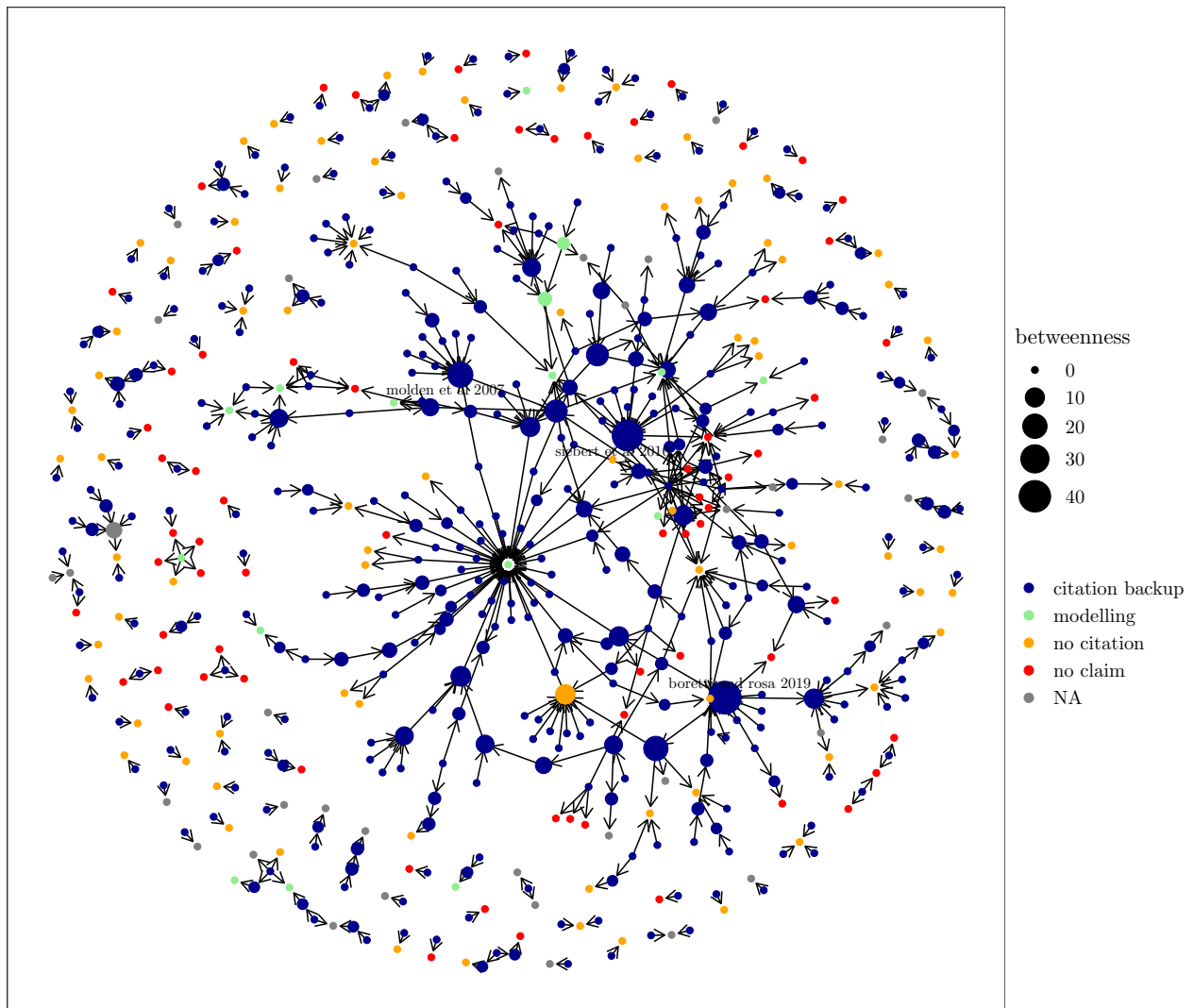
```
## Warning: Removed 33 rows containing missing values (`geom_text_repel()`).
```



```
##
```

```
## $water
```

```
## Warning: Removed 594 rows containing missing values (`geom_text_repel()`).
```

```
# by document.type-----
for (i in names(graph.final)) {
  set.seed(seed)

  p3[[i]] <- ggraph(graph.final[[i]], layout = "igraph", algorithm = "nicely") +
    geom_edge_link(arrow = arrow(length = unit(1.8, 'mm')),
      end_cap = circle(1, "mm")) +
    geom_node_point(aes(color = document.type, size = degree)) +
    geom_node_text(aes(label = ifelse(degree >= min(degree.nodes[[i]]$degree), name, NA)),
      repel = TRUE, size = 2.2) +
    labs(x = "", y = "") +
    scale_color_discrete(name = "") +
    theme_AP() +
    theme(axis.text.x = element_blank(),
      axis.ticks.x = element_blank(),
```

```

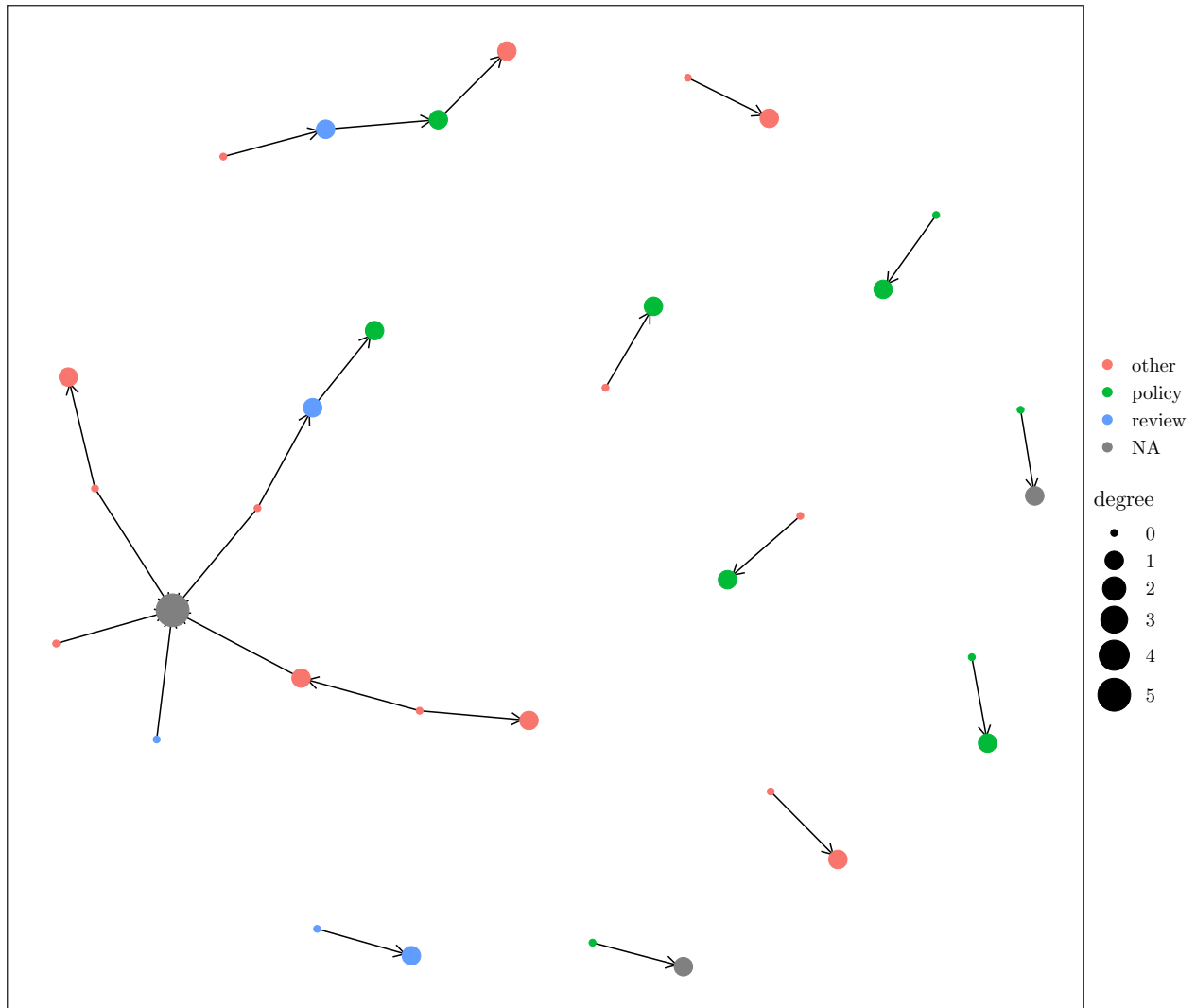
    axis.text.y = element_blank(),
    axis.ticks.y = element_blank(),
    legend.position = "right")
}

```

p3

```
## $food
```

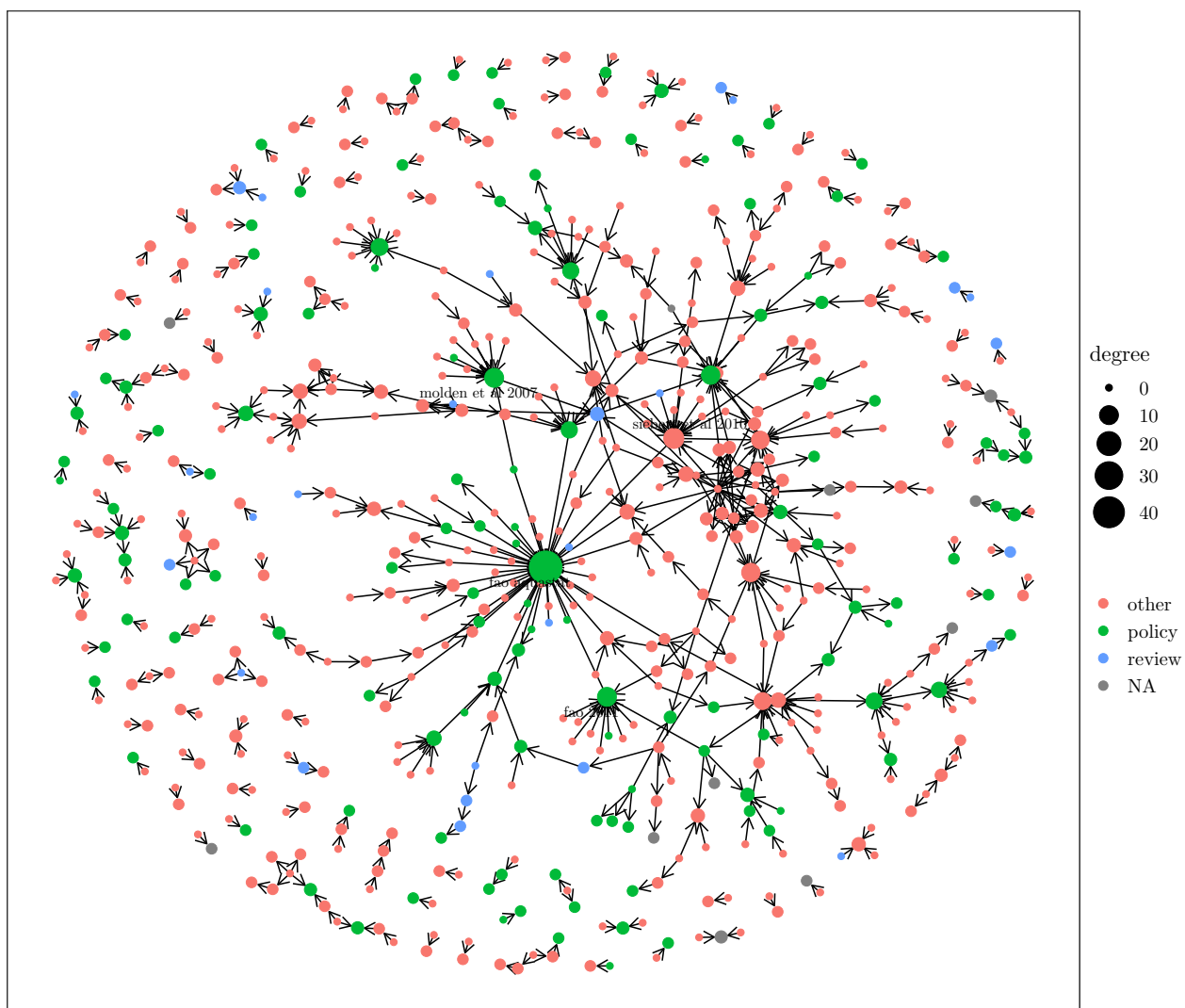
```
## Warning: Removed 33 rows containing missing values (`geom_text_repel()`).
```



```
##
```

```
## $water
```

```
## Warning: Removed 593 rows containing missing values (`geom_text_repel()`).
```



Label nodes that are modelling exercises -----

```
for (i in names(graph.final)) {

  set.seed(seed)

  p4[[i]] <- ggraph(graph.final[[i]], layout = "igraph", algorithm = "nicely") +
    geom_edge_link(arrow = arrow(length = unit(1.8, 'mm')),
      end_cap = circle(1, "mm")) +
    geom_node_point(aes(color = nature.claim)) +
    geom_node_text(aes(label = ifelse(nature.claim == "modelling", name, NA)),
      repel = TRUE, size = 2.2) +
    labs(x = "", y = "") +
    scale_color_manual(name = "",
      values = selected_colors) +
    theme_AP() +
    theme(axis.text.x = element_blank(),
```

```

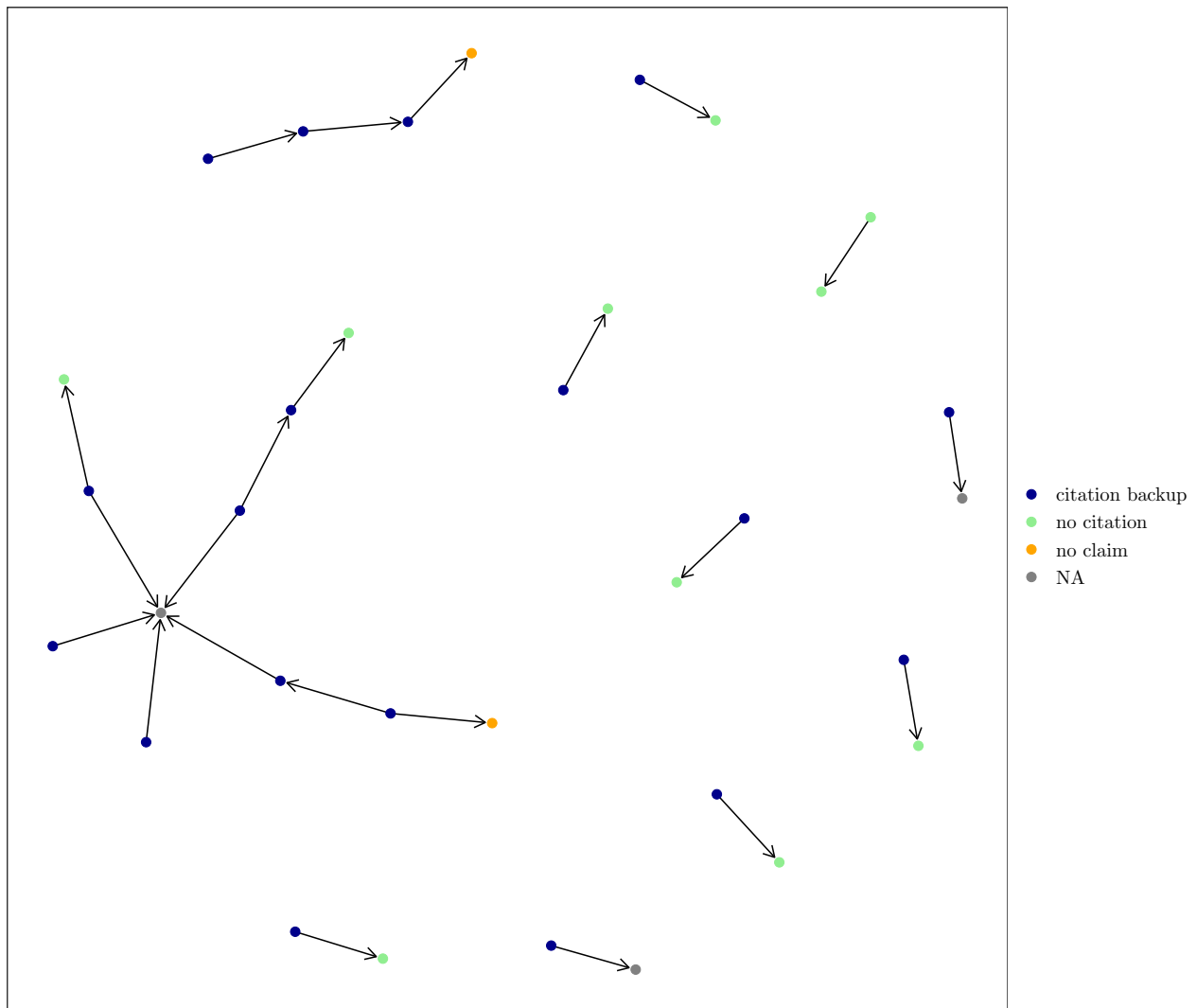
axis.ticks.x = element_blank(),
axis.text.y = element_blank(),
axis.ticks.y = element_blank(),
legend.position = "right")
}

```

p4

```
## $food
```

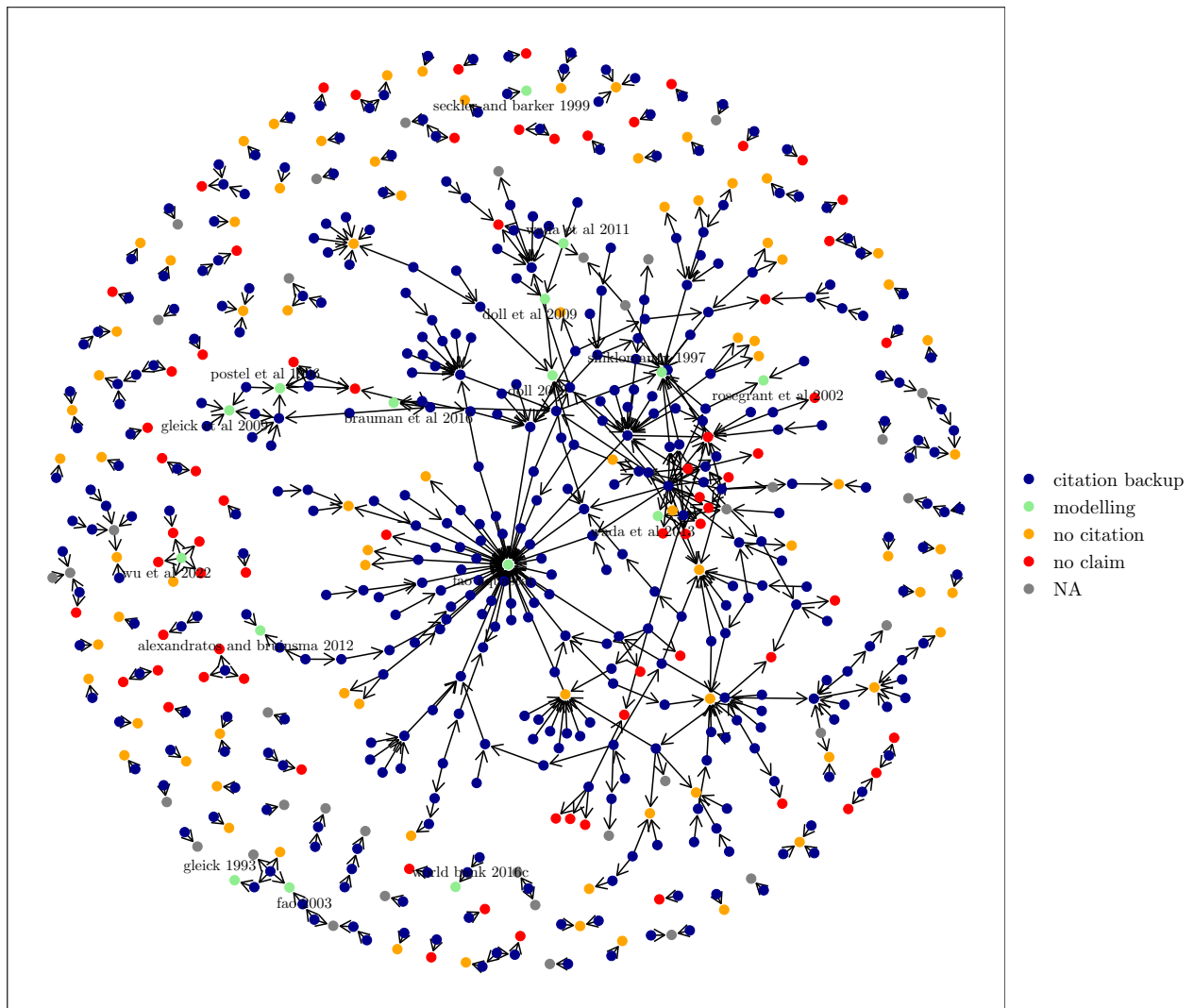
```
## Warning: Removed 33 rows containing missing values (`geom_text_repel()`).
```



```
##
```

```
## $water
```

```
## Warning: Removed 581 rows containing missing values (`geom_text_repel()`).
```



3.3 Uncertainties turned into facts

COUNT PROPORTION OF NODES THAT STATE AS FACT A CLAIM UTTERED AS UNCERTAIN

```
uncertainty_plot_fun <- function(graph) {
```

Extract name of all studies -----

```
all.names <- graph %>%
  activate(nodes) %>%
  pull(name)
```

Extract name of studies stating claim as fact -----

```
f.names <- graph %>%
  activate(nodes) %>%
  data.frame() %>%
```

```

filter(classification == "F") %>%
pull(name)

# Add names to edges -----

add.names.edges <- graph %>%
  activate(edges) %>%
  mutate(from.name = all.names[from],
         to.name = all.names[to])

# Calculate, for each study stating claim as fact, the studies it cites -----
out.classes <- lapply(f.names, function(x) {

  out_nodes <- add.names.edges %>%
    activate(edges) %>%
    filter(from.name == x) %>%
    pull(to.name)

})

# unlist names of studies cited by studies uttering claim as fact -----

di <- sort(unlist(out.classes))

# Extract only those that do not state claim as fact -----

nodes.no.fact <- graph %>%
  activate(nodes) %>%
  data.frame() %>%
  data.table() %>%
  .[name %in% di] %>%
  .[!classification == "F"] %>%
  .$name

name.edges <- add.names.edges %>%
  activate(edges) %>%
  data.frame() %>%
  filter(from.name %in% f.names & to.name %in% nodes.no.fact) %>%
  .[, c("from.name", "to.name")] %>%
  c() %>%
  unlist() %>%
  unique(.)

output <- add.names.edges %>%
  activate(nodes) %>%
  filter(name %in% name.edges) %>%
  activate(edges) %>%

```

```

    filter(from.name %in% name.edges & to.name %in% name.edges)

    return(output)
}

# PLOT GRAPH UNCERTAINTIES TURNED INTO FACTS #####

out <- lapply(graph.final, function(x) uncertainty_plot_fun(x))

p7 <- list()

for (i in names(out)) {

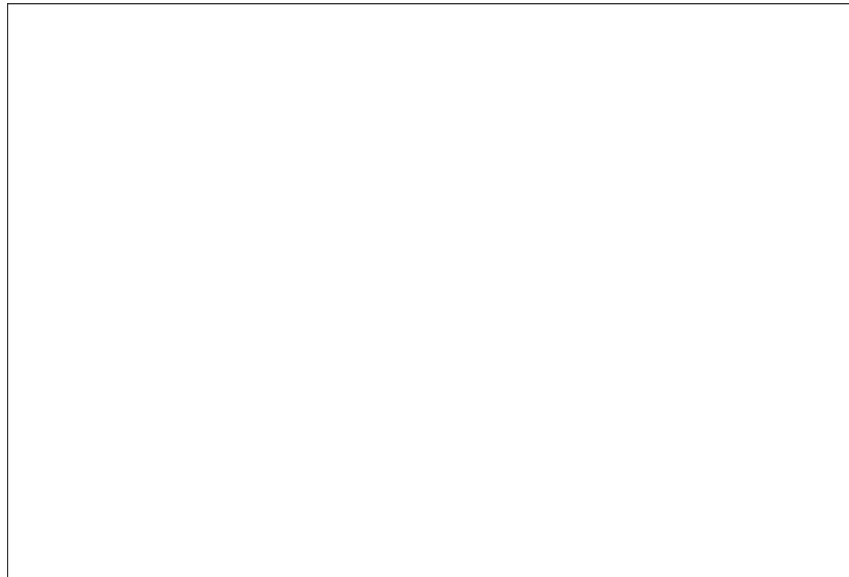
  set.seed(seed)

  p7[[i]] <- ggraph(out[[i]], layout = "igraph", algorithm = "nicely") +
    geom_edge_link(arrow = arrow(length = unit(1.8, 'mm')),
                  end_cap = circle(1, "mm")) +
    geom_node_point(aes(color = category, size = degree, shape = classification)) +
    scale_color_manual(values = c("lightgreen", "orange")) +
    scale_shape_discrete(labels = c("Approximate", "Fact", "Lower threshold", "Range", "Upper threshold")) +
    geom_node_text(aes(label = ifelse(degree >= min(degree.nodes[[i]]$degree), name, NA)),
                  repel = TRUE, size = 2.2) +
    labs(x = "", y = "") +
    theme_AP() +
    theme(axis.text.x = element_blank(),
          axis.ticks.x = element_blank(),
          axis.text.y = element_blank(),
          axis.ticks.y = element_blank(),
          legend.position = "right")
}

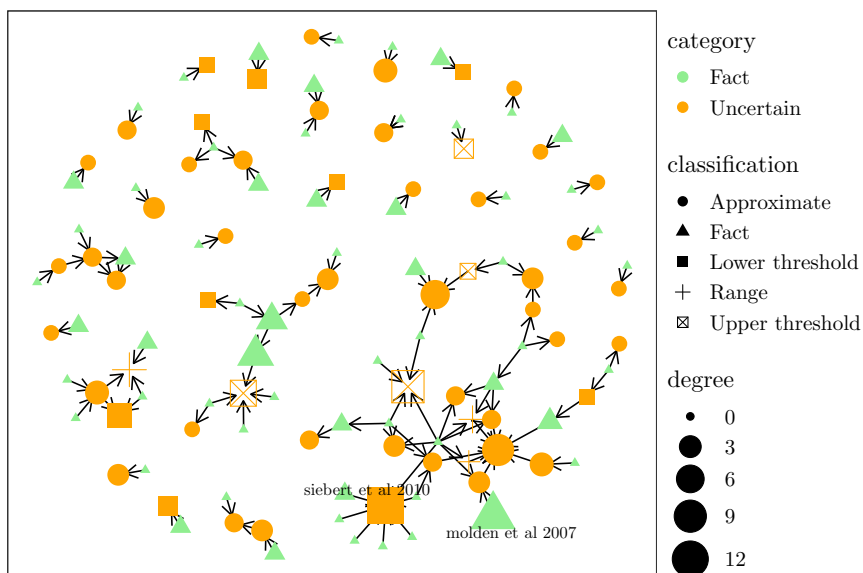
p7

## $food

```



```
##
## $water
## Warning: Removed 117 rows containing missing values (`geom_text_repel()`).
```



```
# FUNCTION TO CALCULATE ALL PATHS BETWEEN PAIRS OF NODES #####
calculate_paths <- function(graph) {
  # Convert to igraph -----
  igraph_graph <- as.igraph(graph)
  # Get all unique pairs of nodes -----
```



```

node_pairs <- expand.grid(from = V(igraph_graph), to = V(igraph_graph))
node_pairs <- node_pairs[node_pairs$from != node_pairs$to, ]

# Function to calculate all simple paths between a pair of nodes-----

calculate_paths <- function(from, to) {
  paths <- all_simple_paths(igraph_graph, from = from, to = to)
  lapply(paths, names)
}

# Apply the function to all node pairs and unnest the results-----

all_paths <- node_pairs %>%
  rowwise() %>%
  mutate(paths = list(calculate_paths(from, to))) %>%
  unnest(cols = c(paths))

out <- sum(sapply(all_paths$paths, function(x) length(x)))

return(out)
}

# CALCULATE ALL PATHS / PATHS TURNING HYPOTHESIS INTO FACTS #####

all.paths <- hypothesis.into.facts.paths <- list()

for (i in names(graph.final)) {

  all.paths[[i]] <- calculate_paths(graph.final[[2]])
  hypothesis.into.facts.paths[[i]] <- uncertainty_plot_fun(graph.final[[2]]) %>%
    calculate_paths(.)
}

# Print results: proportion of paths turning uncertainties into facts -----

for (i in names(all.paths)) {
  print(hypothesis.into.facts.paths[[i]] / all.paths[[i]])
}

## [1] 0.1432161
## [1] 0.1432161

```

3.4 Network through time

```

# PREPARE DATA TO PLOT NETWORK THROUGH TIME #####

# Extract vector with names -----

location_aquastat <- graph.final[[2]] %>%
  activate(nodes) %>%
  data.frame() %>%
  pull(name) %>%
  grep("aquastat", .)

# Extract vector with years -----

v_years <- graph.final[[2]] %>%
  activate(nodes) %>%
  data.frame() %>%
  pull(year)

# Substitute fao aquastat without year with the oldest aquastat citation -----

v_years[location_aquastat] <- oldest.aquastat.cite

# Find NA values -----

na_indices <- is.na(v_years)
sum(na_indices)

## [1] 17

# Generate random values to replace NA -----

random_values <- sample(2000:2020, sum(na_indices), replace = TRUE)

# Replace NA with random values -----

v_years[na_indices] <- random_values

# Define the coordinates-----

y_positions <- runif(length(v_years), min = -3, max = 3) # Random y-axis positions
layout <- cbind(v_years, y_positions) # Use actual years for x-axis
layout_matrix <- as.matrix(layout)
colnames(layout_matrix) <- c("x", "y")

# PLOT NETWORK THROUGH TIME #####

# Set seed -----

set.seed(seed)

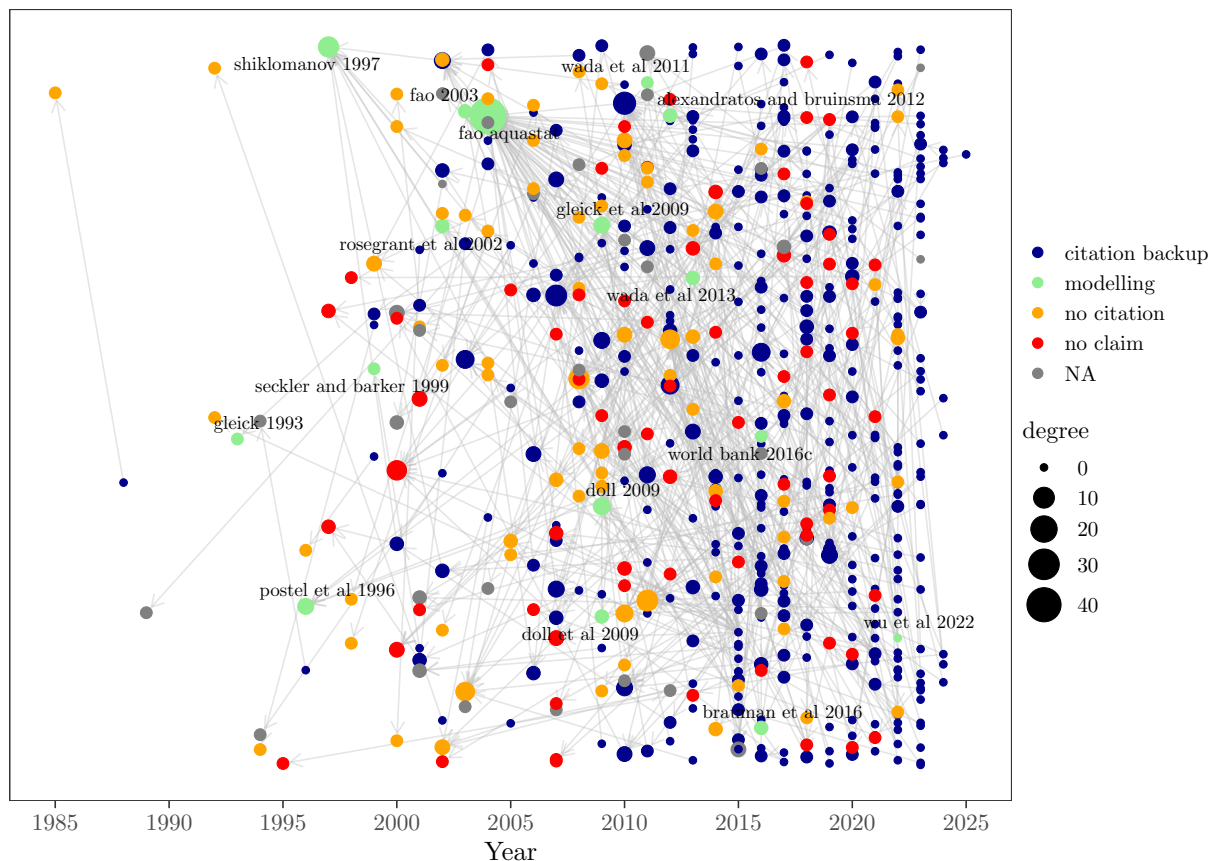
```

```
# Plot -----

ggraph(graph.final[[2]], layout = layout_matrix, algorithm = "nicely") +
  geom_edge_link(arrow = arrow(length = unit(1.8, "mm")),
    end_cap = circle(1, "mm"),
    color = "grey",
    alpha = 0.4) +
  geom_node_point(aes(color = nature.claim, size = degree)) +
  geom_node_text(aes(label = ifelse(nature.claim == "modelling", name, NA)),
    repel = TRUE, size = 2.5) +
  scale_color_manual(name = "",
    values = selected_colors) +
  scale_x_continuous(name = "Year",
    limits = range(v_years),
    breaks = seq(min(v_years),
      max(v_years), by = 5)) +

  labs(x = "Year", y = "") +
  theme_AP() +
  theme(axis.text.y = element_blank(),
    axis.ticks.y = element_blank())
```

Warning: Removed 581 rows containing missing values (`geom_text_repel()`).



```
# FUNCTION TO PLOT EVOLUTION OF NETWORK THROUGH TIME #####
```

```
network_through_time_fun <- function(graph, Year, seed) {
```

```
# Extract all names -----
```

```
all.names <- graph %>%
  activate(nodes) %>%
  pull(name)
```

```
# Add names to edges -----
```

```
add.names.edges <- graph %>%
  activate(edges) %>%
  mutate(from.name = all.names[from],
         to.name = all.names[to])
```

```
# Extract nodes by year -----
```

```
names.targeted <- add.names.edges %>%
  activate(edges) %>%
  filter(year < Year) %>%
  data.frame() %>%
  .[, c("from.name", "to.name")] %>%
  c() %>%
  unlist() %>%
  unique(.)
```

```
name.nodes <- add.names.edges %>%
  activate(nodes) %>%
  filter(name %in% names.targeted) %>%
  activate(edges) %>%
  filter(from.name %in% names.targeted & to.name %in% names.targeted)
```

```
set.seed(seed)
```

```
# Plot -----
```

```
out <- ggraph(name.nodes, layout = "igraph", algorithm = "nicely") +
  geom_edge_link(arrow = arrow(length = unit(1, 'mm')),
                end_cap = circle(0.3, "mm")) +
  geom_node_point(aes(color = nature.claim), size = 0.5) +
  geom_node_text(aes(label = ifelse(nature.claim == "modelling", name, NA)),
                repel = TRUE, size = 2.2) +
  scale_color_manual(name = "",
                    values = selected_colors) +
  labs(x = "", y = "") +
```

```

    theme_AP() +
    theme(axis.text.x = element_blank(),
          axis.ticks.x = element_blank(),
          axis.text.y = element_blank(),
          axis.ticks.y = element_blank(),
          legend.position = "none")

    return(out)
}

# DEFINE YEARS OF INTEREST #####

years.vector <- c(seq(2000, 2020, 10), 2024)

# RUN FUNCTION #####

plots.through.time <- list()

for (i in names(graph.final)) {

  plots.through.time[[i]] <- lapply(years.vector, function(year)
    network_through_time_fun(graph = graph.final[[i]], Year = year, seed = seed) +
    ggtitle(year))
}

# PLOT #####

# Extract legend -----

legend.plot <- list()

for (i in names(plots.through.time)) {

  legend.plot[[i]] <- get_legend(plots.through.time[[i]][[length(plots.through.time[[i]])]] +
    theme(legend.position = "top"))
}

## Warning: Removed 27 rows containing missing values (`geom_text_repel()`).
## Warning: Removed 567 rows containing missing values (`geom_text_repel()`).
# Plot -----

bottom <- out.plot <- list()

for (i in names(plots.through.time)) {

  bottom[[i]] <- do.call(plot_grid, c(plots.through.time[[i]],

```

```

                                nrow = floor(length(years.vector) / 2))
out.plot[[i]] <- plot_grid(legend.plot[[i]],
                           bottom[[i]], ncol = 1, rel_heights = c(0.1, 0.9))
}

```

```

## Warning: Removed 2 rows containing missing values (`geom_text_repel()`).
## Warning: Removed 17 rows containing missing values (`geom_text_repel()`).
## Warning: Removed 27 rows containing missing values (`geom_text_repel()`).
## Warning: Removed 9 rows containing missing values (`geom_text_repel()`).
## Warning: Removed 86 rows containing missing values (`geom_text_repel()`).
## Warning: Removed 371 rows containing missing values (`geom_text_repel()`).
## Warning: Removed 567 rows containing missing values (`geom_text_repel()`).

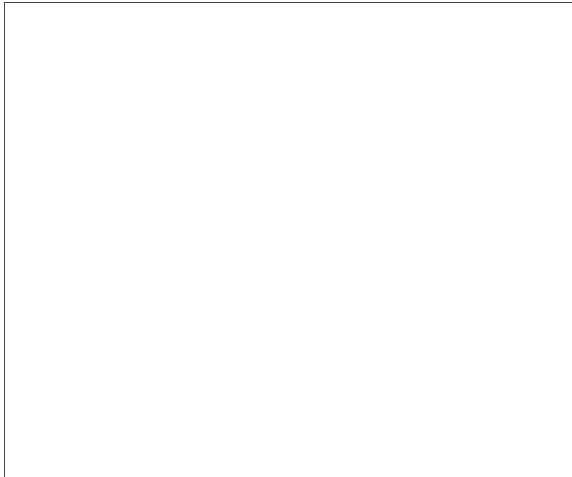
```

```
out.plot
```

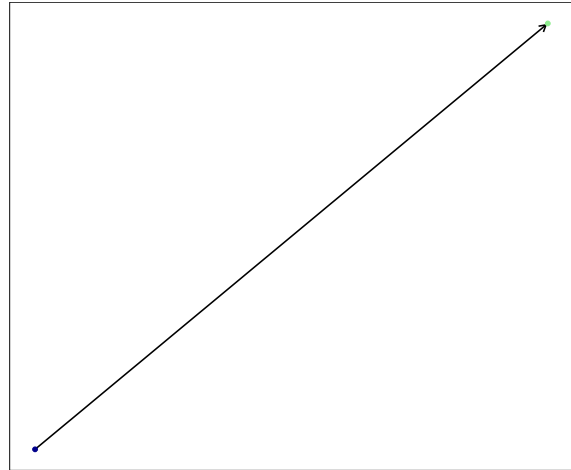
```
## $food
```

• citation backup • no citation • no claim • NA

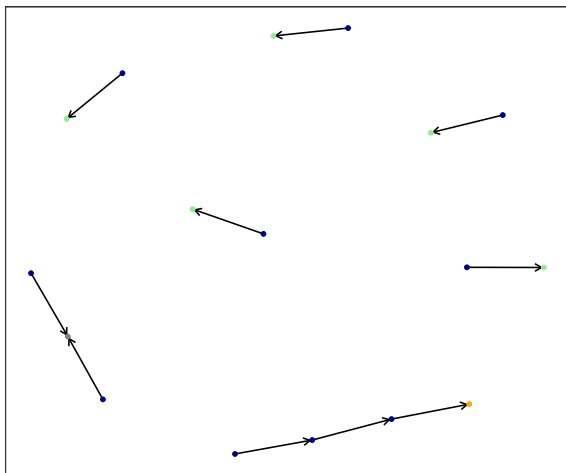
2000



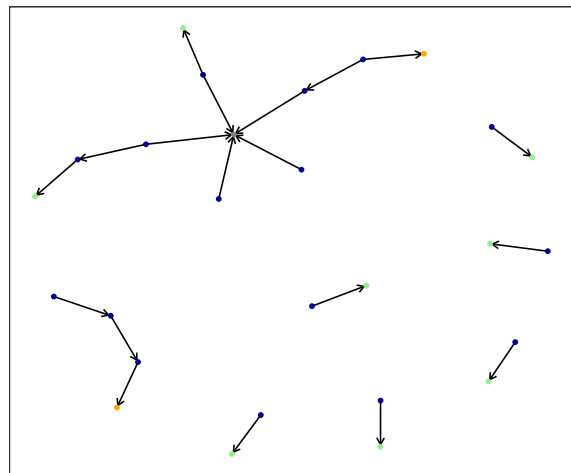
2010



2020



2024

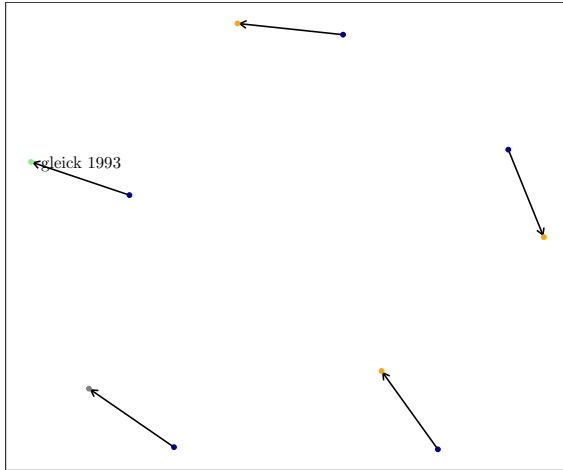


##

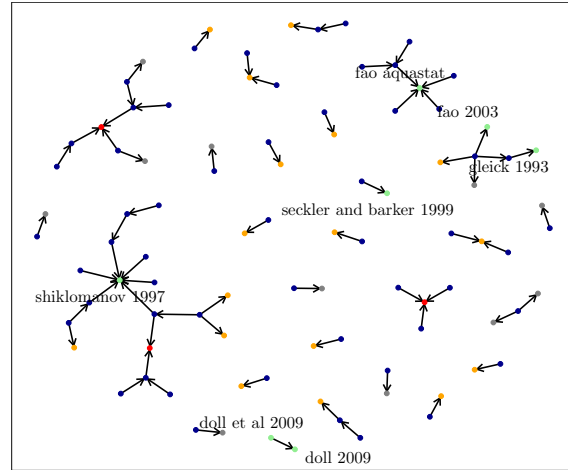
\$water

• citation backup • modelling • no citation • no claim • NA

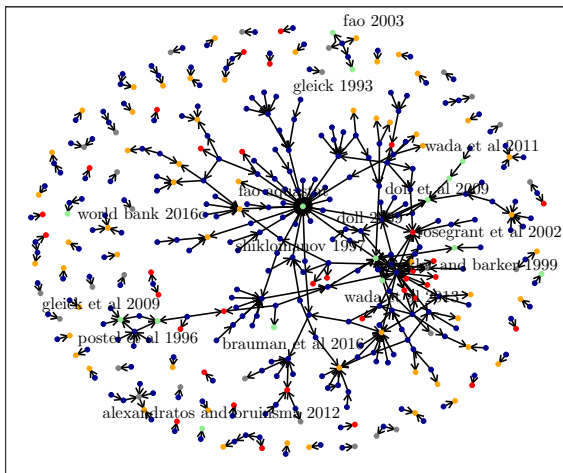
2000



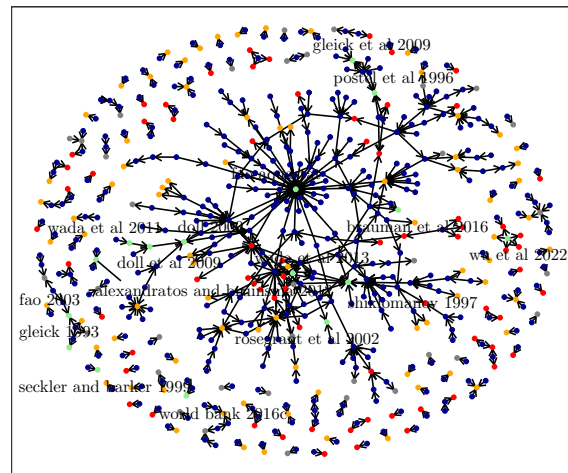
2010



2020



2024



4 Analysis of paths

4.1 “no claim” or “no citation” paths

```
# COUNT THE NUMBER OF NODES WITH PATHS ULTIMATELY LEADING TO NODES
# THAT DO NOT MAKE THE CITATION #####

# Function: loop through each node that do not make the claim to find all nodes
# connected to it -----

nodes_to_no_claim_node_fun <- function(g, terminal_nodes) {

  if (!is.igraph(g)) {
    g <- as.igraph(g)
  }
}
```



```

}

all_predecessors <- vector("list", length(terminal_nodes))

for (i in seq_along(terminal_nodes)) {

  terminal_node <- terminal_nodes[i]
  predecessors <- subcomponent(g, terminal_node, mode = "in")
  all_predecessors[[i]] <- predecessors
}

unique_predecessors <- unique(names(unlist(all_predecessors)))

return(unique_predecessors)

}

# CALCULATE

# Extract name of all nodes -----
all_nodes <- lapply(graph.final, function(graph)
  graph %>%
    activate(nodes) %>%
    pull(name))

# Extract name of nodes that do not make the claim -----

no.claim_nodes <- lapply(graph.final, function(graph)
  graph %>%
    activate(nodes) %>%
    filter(degree.out == 0 & nature.claim == "no claim") %>%
    pull(., "name"))

# Extract name of nodes that do not make the claim and those that make
# the claim but do not cite anybody -----

no.claim.and.no.citation.nodes <- lapply(graph.final, function(graph)
  graph %>%
    activate(nodes) %>%
    filter(degree.out == 0 & nature.claim == "no claim" | nature.claim == "no citation" ) %>%
    pull(., "name"))

# Run the function -----

tmp <- list()

for(i in names(graph.final)) {

```

```

tmp[[i]] <- lapply(list(no.claim_nodes[[i]],
                        no.claim.and.no.citation.nodes[[i]]), function(x)
  sort(nodes_to_no_claim_node_fun(graph.final[[i]], terminal_nodes = x)))
}

for(i in names(graph.final)) {
  names(tmp[[i]]) <- c("path ending in no claim",
                      "path ending in no claim or no citation")
}

tmp

```

```

## $food
## $food$path ending in no claim`
## [1] "hanjra and qureshi 2010" "mitchell et al 2018"
## [3] "molden et al 2010"      "niu et al 2023"
## [5] "siebert and doll 2010"  "wang et al 2012"
##
## $food$path ending in no claim or no citation`
## [1] "borin 2023" "borsato et al 2020"
## [3] "chartzoulakis and bertaki 2015" "de pascale et al 2011"
## [5] "du preez et al 2018" "evans and sadler 2008"
## [7] "fao 2002" "fao 2003"
## [9] "fao 2007b" "fernandez-cirelli et al 2009"
## [11] "hanjra and qureshi 2010" "lobell et al 2006"
## [13] "mitchell et al 2018" "molden et al 2010"
## [15] "niu et al 2023" "okorogbona et.al 2018"
## [17] "policy.1667934" "policy.1898497"
## [19] "rolle et al 2021" "salmon et al 2015"
## [21] "siebert and doll 2010" "turrall et al 2010"
## [23] "wang et al 2012" "world bank 2020"
## [25] "world bank 2021"
##
##
## $water
## $water$path ending in no claim`
## [1] "abbot et al 2019" "acosta et al 2016"
## [3] "ahmed et al 2022" "alcamo et al 2007"
## [5] "antia 2022" "badrul masud et al 2019"
## [7] "bar et al 2015" "barreto and amaral 2018"
## [9] "besharat et al 2020" "biemans et al 2011"
## [11] "bjornlund et al 2013" "bondeau et al 2007"
## [13] "boretti and rosa 2019" "braun et al 2022"
## [15] "calzadilla et al 2010" "carmona et al 2017"
## [17] "carvalho 2019" "chai et al 2016"
## [19] "chirone et al 2022" "clay 2004"
## [21] "coelho et al 2012" "cristache et al 2018"

```

## [23] "d'odorico et al 2019"	"de graaf et al 2017"
## [25] "deikman et al 2012"	"dirwai et al 2022"
## [27] "doll et al 2014"	"droppers et al 2020"
## [29] "du et al 2015"	"eckert and kovalevska 2021"
## [31] "elmoneim badr et al 2021"	"epri 2002"
## [33] "faiz alam et al 2023"	"falkenmark 2013"
## [35] "falkenmark et al 1997"	"fao 2007b"
## [37] "fao 2016"	"fao 2018c"
## [39] "fao 2020"	"friha et al 2022"
## [41] "gan et al 2013"	"gerten et al 2007"
## [43] "gheewala et al 2014"	"giordano 2007"
## [45] "gleick and palaniappan 2010"	"gleick et al 2011"
## [47] "gleick et al 2018"	"gordon et al 2010"
## [49] "gorjian et al 2020"	"gorjian et al 2022"
## [51] "grigas et al 2023"	"gumidyala et al 2020"
## [53] "gustinasari et al 2020"	"hanasaki et al 2008"
## [55] "hanasaki et al 2008b"	"harun and hanafiah 2018"
## [57] "hochmuth et al 2015"	"hoekstra 2003"
## [59] "hoekstra and mekonnen 2012"	"hofste et al 2019"
## [61] "holdren 2008"	"howden et al 2013"
## [63] "huang et al 2023"	"huo et al 2022"
## [65] "iwmi 2000"	"jaramillo and destouni 2015"
## [67] "jez et al 2016"	"jiang et al 2017"
## [69] "johnson et al 2001"	"jury and vaux jr 2005"
## [71] "kaba gurmessa and assefa 2023"	"kabir et al 2023"
## [73] "karimi et al 2019"	"kaur saggi and jain 2022"
## [75] "khan and hanjra 2009"	"kiani et al 2023"
## [77] "kilemo 2022"	"kong et al 2017"
## [79] "kumar dubey et al 2021"	"kumar ravi et al 2023"
## [81] "laluet et al 2024"	"lamastra et al 2014"
## [83] "li and long 2019"	"li et al 2023b"
## [85] "liu and yang 2010"	"liu et al 2016"
## [87] "marston et al 2018"	"martinez sosa et al 2023"
## [89] "mcdermid et al 2023"	"meghan salmon et al 2015"
## [91] "mekonnen and hoekstra 2012"	"mekonnen et al 2015"
## [93] "menceloglu et al 2022"	"mendonca et al 2023"
## [95] "mohanty et al 2018"	"moldovan et al 2022"
## [97] "nahar sumiya and khatun 2016"	"nnadi et al 2015"
## [99] "oladosu et al 2019"	"oladosu et al 2022"
## [101] "opio et al 2011"	"othmani et al 2021"
## [103] "ozdogan et al 2010b"	"ozturk et al 2022"
## [105] "pang et al 2021"	"payero et al 2006"
## [107] "pellegrini et al 2016"	"pena-arancibia et al 2016"
## [109] "perry et al 2017"	"pimentel et al 1995"
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```

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## [129] "scanlon et al 2007"
## [131] "scanlon et al 2023"
## [133] "sepaskhah and ahmadi 2010"
## [135] "shiklomanov 2000"
## [137] "siebert et al 2005"
## [139] "siebert et al 2015"
## [141] "steward and bernard 2006"
## [143] "tiwari et al 2023"
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## [147] "unesco 2001"
## [149] "united nations 2003"
## [151] "united nations 2022"
## [153] "velis et al 2017"
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## [159] "wajima 2018"
## [161] "wbcsd 2009"
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## [165] "world bank 2001"
## [167] "worldometers 2019"
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## [177] "zeng et al 2022"
## [179] "zhang et al 2022"
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## [4] "abou zaki et al 2018"
## [5] "ackerman 2015"
## [6] "acosta et al 2016"
## [7] "adama et al 2020"
## [8] "adhikari et al 2021"
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## [10] "alan rotz 2020"
## [11] "alcamo et al 2007"
## [12] "alvarez et al 2004"
## [13] "anderson et al 2017"
## [14] "angaleeswari et al 2021"
## [15] "antia 2022"
"redhu and jain 2023"
"rivera et al 2017"
"rodriguez et al 2022"
"sadoff et al 2020"
"sahray et al 2023"
"scanlon et al 2017"
"schreinemachers and tipraqsa 2012"
"seyboth and plattner 2014"
"shtull-trauring et al 2016"
"siebert et al 2010"
"singh et al 2024"
"tabunshikov et al 2021"
"tortell 2020"
"turner 2008"
"united nations 1998"
"united nations 2021"
"velez sanchez et al 2023"
"vorosmarty et al 2000"
"wada 2015"
"wada et al 2016"
"walter et al 2017"
"weatherhead and howden 2009"
"wmo 1997"
"world bank 2017"
"wri 2000"
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"yilmazkuday et al 2021"
"young et al 2019"
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[88] "epri 2002"
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[110] "firdayati et al 2022"
[111] "fitton et al 2019"

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 ## [114] "friha et al 2022"
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 ## [121] "giordano 2007"
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 ## [132] "grigas et al 2023"
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 ## [135] "gustinasari et al 2020"
 ## [136] "gwp nd"
 ## [137] "haddeland et al 2013"
 ## [138] "hanasaki et al 2008"
 ## [139] "hanasaki et al 2008b"
 ## [140] "hannah 2017"
 ## [141] "harun and hanafiah 2018"
 ## [142] "he et al 2023"
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 ## [144] "hochmuth et al 2015"
 ## [145] "hoekstra 2003"
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 ## [147] "hofste et al 2019"
 ## [148] "hofwegen and svendsen 2000"
 ## [149] "holdren 2008"
 ## [150] "howden et al 2013"
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 ## [154] "iaastd 2009"
 ## [155] "ingrao et al 2023"
 ## [156] "ipcc 2007"
 ## [157] "iwmi 2000"
 ## [158] "jagermeyr et al 2017"
 ## [159] "jaramillo and destouni 2015"

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 ## [162] "jez et al 2016"
 ## [163] "jiang et al 2017"
 ## [164] "jimenez-arias et al 2023"
 ## [165] "johansson et al 2015"
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 ## [167] "jury and vaux jr 2005"
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 ## [312] "schulte et al 2014"
 ## [313] "seckler et al 1998"
 ## [314] "sepaskhah and ahmadi 2010"
 ## [315] "seyboth and plattner 2014"
 ## [316] "shang et al 2024"
 ## [317] "shiklomanov 1999"
 ## [318] "shiklomanov 2000"
 ## [319] "shiklomanov and rodde 2003"
 ## [320] "shtull-trauring et al 2016"
 ## [321] "siebert and doll 2010"
 ## [322] "siebert et al 2005"
 ## [323] "siebert et al 2010"
 ## [324] "siebert et al 2013"
 ## [325] "siebert et al 2015"
 ## [326] "singh et al 2022"
 ## [327] "singh et al 2024"
 ## [328] "sonen capital 2016"
 ## [329] "song and song 2023"
 ## [330] "sophocleous 2004"
 ## [331] "spiegel international 2009"
 ## [332] "steduto et al 2018"
 ## [333] "steffen 2017"
 ## [334] "steward and bernard 2006"
 ## [335] "swatuk et al 2018"
 ## [336] "tabunshikov et al 2021"
 ## [337] "tavares et al 2022"
 ## [338] "ti et al 2021"
 ## [339] "tian et al 2022"
 ## [340] "tiwari et al 2023"
 ## [341] "tortell 2020"
 ## [342] "tsiropoulos et al 2022"
 ## [343] "tsur 2005"
 ## [344] "tunc and sahin 2025"
 ## [345] "tuninetti et al 2015"
 ## [346] "turner 2008"
 ## [347] "unctad 2011"
 ## [348] "unep 2011"
 ## [349] "unesco 2001"
 ## [350] "unesco 2006"
 ## [351] "unesco 2014"

[352] "unesco 2017"
 ## [353] "united nations 1998"
 ## [354] "united nations 2003"
 ## [355] "united nations 2015"
 ## [356] "united nations 2021"
 ## [357] "united nations 2022"
 ## [358] "united nations 2023"
 ## [359] "velez sanchez et al 2023"
 ## [360] "velis et al 2017"
 ## [361] "velpuri et al 2009"
 ## [362] "vorosmarty et al 2000"
 ## [363] "vorosmarty et al 2005"
 ## [364] "vorosmarty et al 2010"
 ## [365] "wada 2015"
 ## [366] "wada et al 2013b"
 ## [367] "wada et al 2014"
 ## [368] "wada et al 2016"
 ## [369] "wajima 2018"
 ## [370] "walter et al 2017"
 ## [371] "wbcsd 2009"
 ## [372] "weatherhead and howden 2009"
 ## [373] "who 2014"
 ## [374] "williams et al 2017"
 ## [375] "wilson 2013"
 ## [376] "winpenny et al 2010"
 ## [377] "wisser et al 2008"
 ## [378] "wisser et al 2010"
 ## [379] "wmo 1997"
 ## [380] "wood et al 2000"
 ## [381] "world bank 1992"
 ## [382] "world bank 2001"
 ## [383] "world bank 2017"
 ## [384] "world bank 2021"
 ## [385] "world watch institute 2004"
 ## [386] "world water assessment programme 2003"
 ## [387] "world water assessment programme 2014"
 ## [388] "worldometers 2019"
 ## [389] "wri 1994"
 ## [390] "wri 2000"
 ## [391] "wu et al 2022"
 ## [392] "wwap 2018"
 ## [393] "wwf 2006"
 ## [394] "xing yuan et al 2024"
 ## [395] "xu et al 2020"
 ## [396] "ye et al 2023"
 ## [397] "yilmazkuday et al 2021"
 ## [398] "yin et al 2022"
 ## [399] "young et al 2019"

```
## [400] "yu et al 2019"
## [401] "zeman et al 2006"
## [402] "zeng et al 2022"
## [403] "zhang et al 2015"
## [404] "zhang et al 2015b"
## [405] "zhang et al 2022"
## [406] "zhao et al 2022"
## [407] "zhou et al 2022"
## [408] "zhuo et al 2022"

# Calculate proportions -----

out <- list()

for(i in names(tmp)) {
  out[[i]] <- lapply(tmp[[i]], function(x) length(x) / length(all_nodes[[i]]))
}

out

## $food
## $food$path ending in no claim`
## [1] 0.1818182
##
## $food$path ending in no claim or no citation`
## [1] 0.7575758
##
##
## $water
## $water$path ending in no claim`
## [1] 0.3015075
##
##
## $water$path ending in no claim or no citation`
## [1] 0.6834171
```

4.2 Calculation of amplification

```
# CREATE FUNCTION TO CHECK AMPLIFICATION #####

# amplification measure for paper P: defined as the number of
# citation-paths originating at P and terminating at all other papers,
# except for paths of length 1 flowing directly to modelling papers.

amplification_fun <- function(graph) {

  # Convert tbl_graph to igraph object -----

  ig <- as.igraph(graph)
```

```

nature_claims <- V(ig)$nature.claim

# initialize counter to store results for each paper -----
results <- numeric(vcount(ig))

# Loop over each paper -----

for (P in V(ig)) {

  # Initialize counter for valid paths
  path_count <- 0

  # Traverse through all nodes and count paths avoiding direct "modelling"
  for (target in V(ig)) {

    if (P != target) {

      all_paths <- all_simple_paths(ig, from = P, to = target, mode = "out")

      # Filter out paths of length 1 that end in a "modelling" node
      valid_paths <- Filter(function(path) {
        !(length(path) == 2 && nature_claims[path[2]] == "modelling")
      }, all_paths)

      path_count <- path_count + length(valid_paths)
    }
  }

  results[P] <- path_count
}

return(results)
}

# RUN AMPLIFICATION FUNCTION #####

amplification.indices <- lapply(graph.final, function(graph)
  amplification_fun(graph))

# Calculate average amplification index of the networks -----
# (e.g., the number paths initiated by the average paper
# leading to studies that do # not flow directly to "primary" data)
lapply(amplification.indices, function(x) mean(x))

## $food
## [1] 0.6060606

```

```
##
## $water
## [1] 1.355109

# PLOT DISTRIBUTION OF AMPLIFICATION INDEXES #####

plot.amplification <- list()

for (i in names(amplification.indices)) {

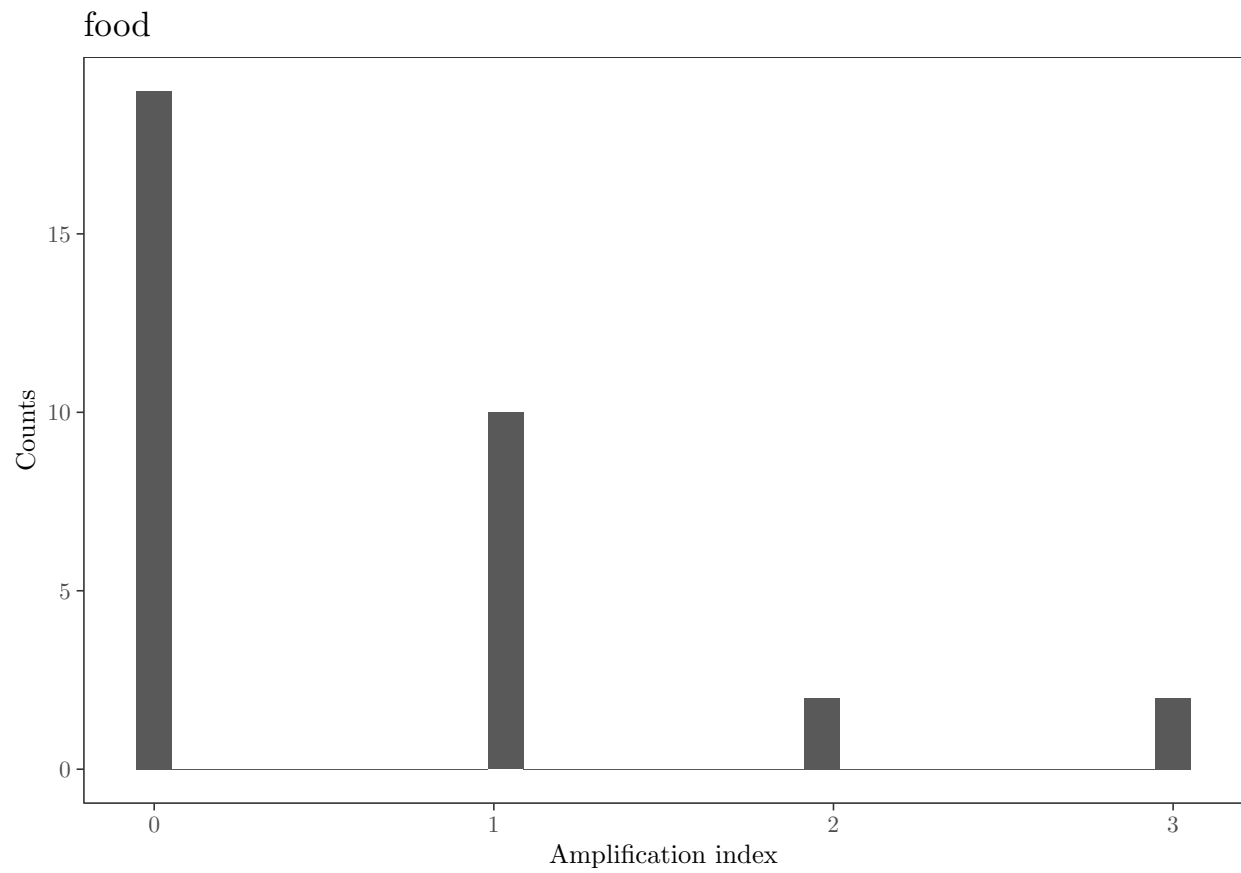
  plot.amplification[[i]] <- amplification.indices[[i]] %>%
    data.frame("index" = .) %>%
    ggplot(., aes(index)) +
    geom_histogram() +
    theme_AP() +
    labs(y = "Counts", x = "Amplification index") +
    ggtitle(names(amplification.indices[i]))

}

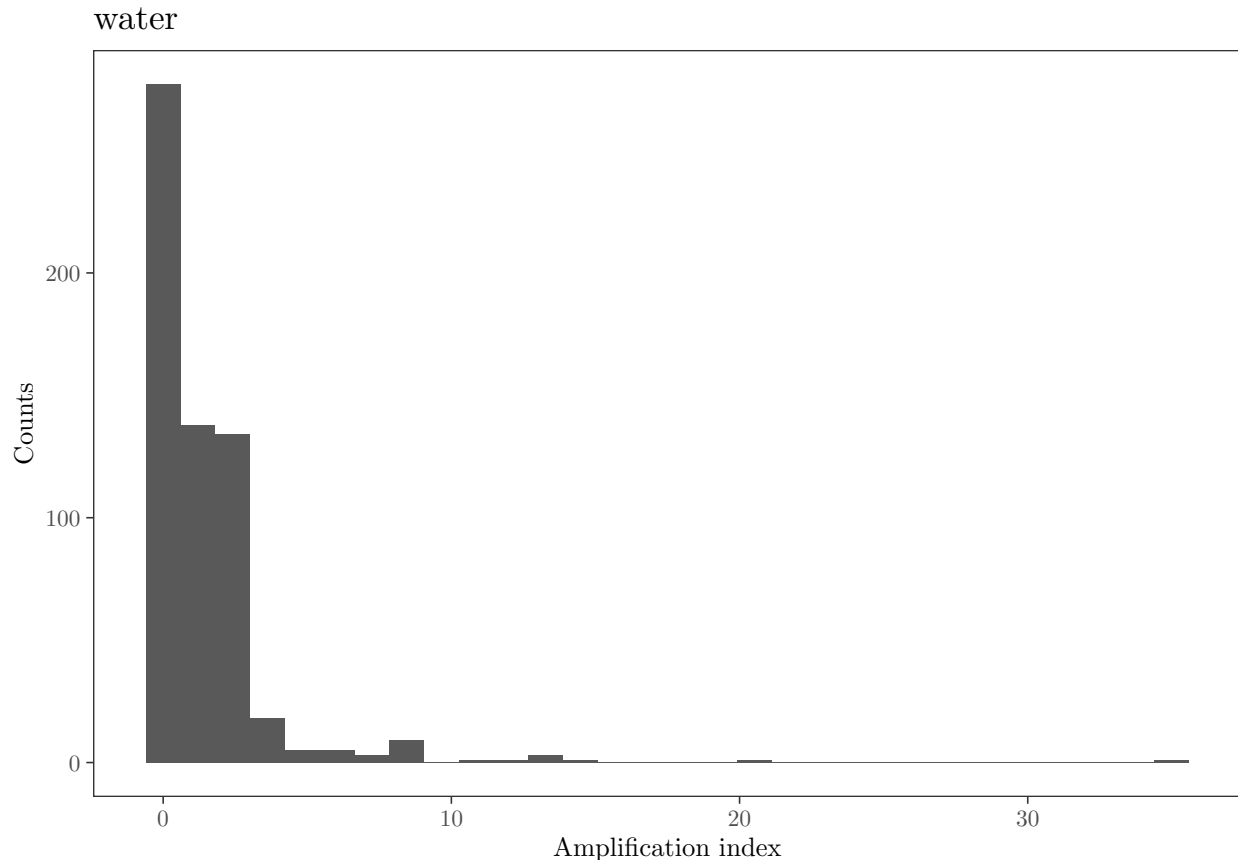
plot.amplification

## $food

## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



```
##  
## $water  
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```

5 Study of Aquastat values

```
# STUDY OF AQUASTAT PERCENTAGES #####
```

```
# Read in aquastat dataset -----
```

```
aquastat.dt <- read.xlsx("aquastat_dt.xlsx") %>%
  data.table() %>%
  .[Year == 2020] %>%
  setnames(., c("Value", "Area"), c("percentage", "country")) %>%
  .[, .(country, percentage)] %>%
  .[, data:= "aquastat 2020"] %>%
  .[, country:= countrycode(country, origin = "country.name", destination = "country.name")]
```

```
## Warning: Some values were not matched unambiguously: Australia and New Zealand
```

```
## Warning: Some strings were matched more than once, and therefore set to <NA> in the result:
```

```
aquastat.dt[, continent:= countrycode(country, origin = "country.name", destination = "continent.name")]
```

```
# Read in world resources institute dataset -----
```

```
wri <- fread("world_resources_institut_guide_to_the_global_environment_1994.csv") %>%
```

```

.[order(country)] %>%
.[, data:= "wri 1994"] %>%
.[, country:= countrycode(country, origin = "country.name", destination = "country.name")]

## Warning: Some values were not matched unambiguously: , Cote d'Ivoire
wri[, continent:= countrycode(country, origin = "country.name", destination = "continent")]

## Warning: Some values were not matched unambiguously: Czechoslovakia, Yugoslavia
# Compare distributions -----

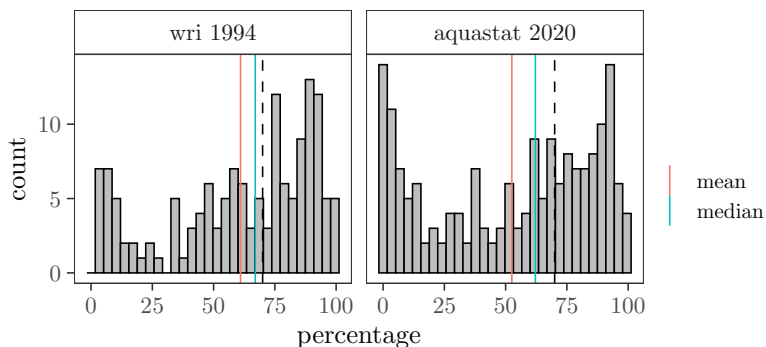
dt.comparison <- rbind(aquastat.dt, wri) %>%
.[, data:= factor(data, levels = c("wri 1994", "aquastat 2020"))]

dt.stats.comparison <- dt.comparison[, .(mean = mean(percentage, na.rm = TRUE),
                                          median = median(percentage, na.rm = TRUE)), data] %>%
melt(., measure.vars = c("mean", "median"))

ggplot(dt.comparison, aes(percentage)) +
  geom_histogram(color = "black", fill = "grey") +
  facet_wrap(~data) +
  geom_vline(data = dt.stats.comparison, aes(xintercept = value, color = variable)) +
  scale_color_discrete(name = "") +
  geom_vline(xintercept = 70, lty = 2) +
  theme_AP()

## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
## Warning: Removed 1 rows containing non-finite values (`stat_bin()`).

```



```

# At the country level -----

tmp <- aquastat.dt[wri, on = c("country", "continent")] %>%
.[, .(country, continent, percentage, i.percentage)] %>%
setnames(., c("percentage", "i.percentage"), c("aquastat 2020", "wri 1994")) %>%
melt(., measure.vars = c("aquastat 2020", "wri 1994")) %>%
.[, country:= ifelse(country == "Trinidad & Tobago", "Trinidad and Tobago", country)] %>%
na.omit() %>%

```

```
split(., .$continent)
```

```
## Warning in melt.data.table(., measure.vars = c("aquastat 2020", "wri 1994")):  
## 'measure.vars' [aquastat 2020, wri 1994] are not all of the same type. By order  
## of hierarchy, the molten data value column will be of type 'double'. All  
## measure variables not of type 'double' will be coerced too. Check DETAILS in  
## ?melt.data.table for more on coercion.
```

```
out <- list()
```

```
for(i in names(tmp)) {
```

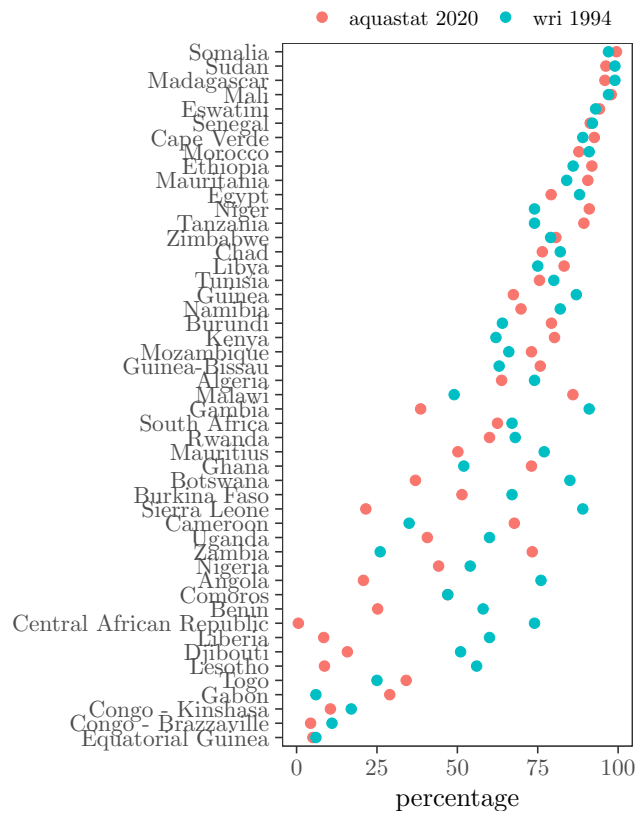
```
  out[[i]] <- ggplot(tmp[[i]], aes(reorder(country, value),  
                                   value, color = variable)) +  
  
    coord_flip() +  
    scale_color_discrete(name = "") +  
    geom_point() +  
    theme_AP() +  
    theme(legend.position = "top") +  
    labs(x = "", y = "percentage") +  
    ggtitle(names(tmp[i]))
```

```
}
```

```
out
```

```
## $Africa
```

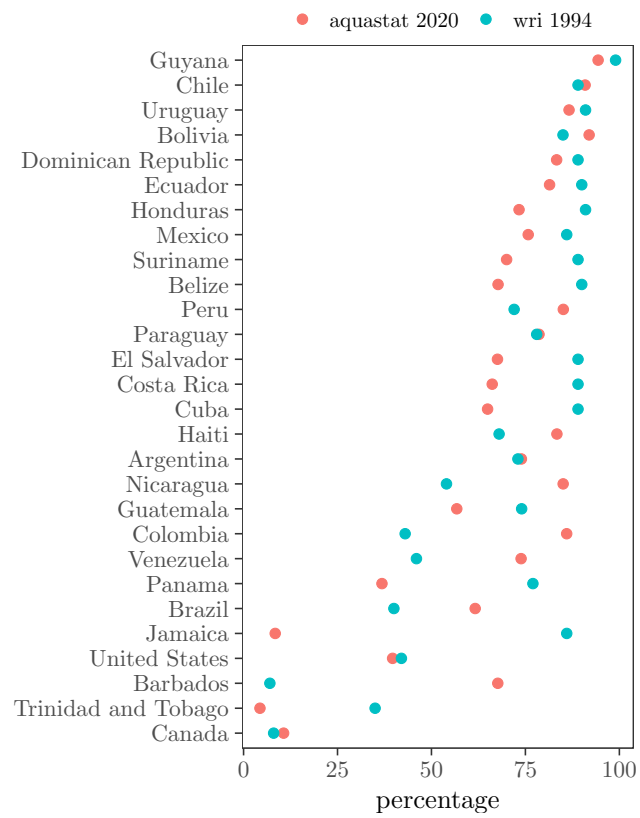
Africa



##

\$Americas

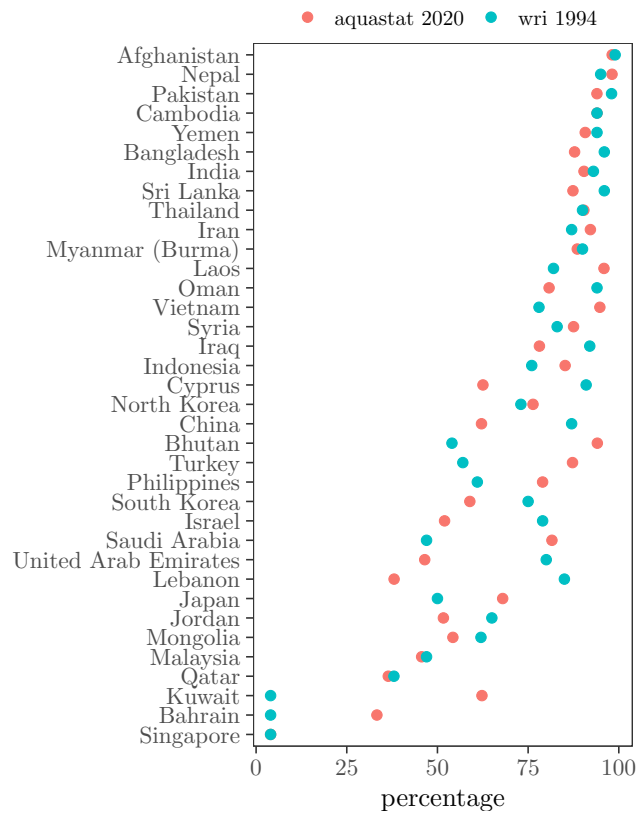
Americas



##

\$Asia

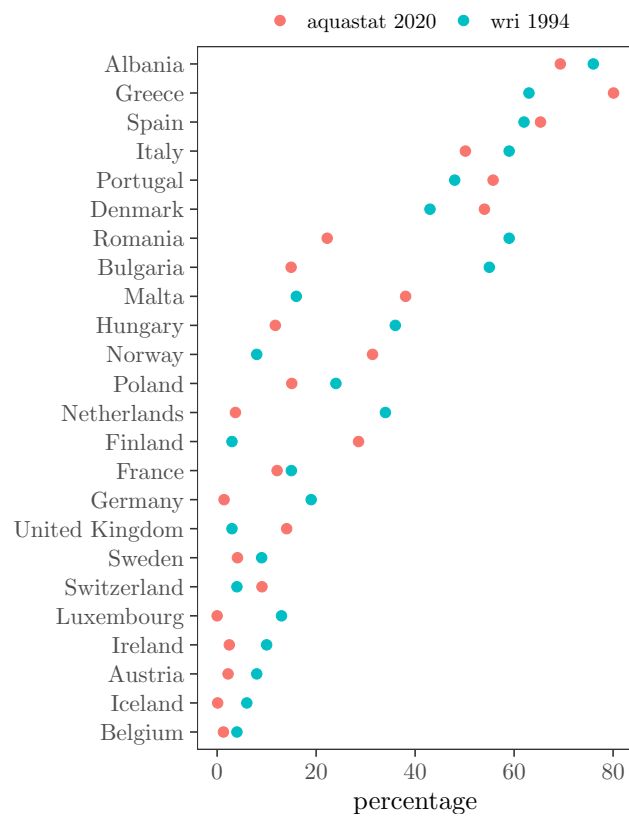
Asia



##

\$Europe

Europe



##

\$Oceania

Oceania



6 Session information

```
# SESSION INFORMATION #####
```

```
sessionInfo()
```

```
## R version 4.3.3 (2024-02-29)
## Platform: aarch64-apple-darwin20 (64-bit)
## Running under: macOS Sonoma 14.2.1
##
## Matrix products: default
## BLAS: /Library/Frameworks/R.framework/Versions/4.3-arm64/Resources/lib/libRblas.0.dylib
## LAPACK: /Library/Frameworks/R.framework/Versions/4.3-arm64/Resources/lib/libRlapack.dylib;
##
## locale:
## [1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
##
## time zone: Europe/London
## tzcode source: internal
##
## attached base packages:
## [1] parallel stats graphics grDevices utils datasets methods
## [8] base
##
## other attached packages:
## [1] scales_1.3.0 wesanderson_0.3.6 benchmarkme_1.0.8 tidygraph_1.3.0
## [5] cowplot_1.1.1 ggraph_2.1.0 igraph_1.6.0 bibliometrix_4.0.1
## [9] lubridate_1.9.2 forcats_1.0.0 stringr_1.5.1 dplyr_1.1.4
## [13] purrr_1.0.2 readr_2.1.4 tidyr_1.3.0 tibble_3.2.1
## [17] ggplot2_3.4.4 tidyverse_2.0.0 data.table_1.14.99 openxlsx_4.2.5.2
##
## loaded via a namespace (and not attached):
## [1] Rdpack_2.6 gridExtra_2.3 readxl_1.4.2
## [4] rlang_1.1.3 magrittr_2.0.3 tidytext_0.4.1
## [7] compiler_4.3.3 vctrs_0.6.5 crayon_1.5.2
## [10] pkgconfig_2.0.3 fastmap_1.1.1 ellipsis_0.3.2
## [13] labeling_0.4.3 utf8_1.2.4 promises_1.2.0.1
## [16] rmarkdown_2.21 tzdb_0.3.0 tinytex_0.45
## [19] bit_4.0.5 xfun_0.39 jsonlite_1.8.4
## [22] flashClust_1.01-2 highr_0.10 SnowballC_0.7.1
## [25] later_1.3.0 tweenr_2.0.2 cluster_2.1.6
## [28] R6_2.5.1 stringi_1.8.3 RColorBrewer_1.1-3
## [31] cellranger_1.1.0 estimability_1.4.1 iterators_1.0.14
## [34] Rcpp_1.0.12 knitr_1.42 filehash_2.4-5
## [37] httpuv_1.6.9 rentrez_1.2.3 Matrix_1.6-5
## [40] timechange_0.2.0 tidyselect_1.2.0 viridis_0.6.4
## [43] rstudioapi_0.15.0 stringdist_0.9.10 pubmedR_0.0.3
## [46] yaml_2.3.7 codetools_0.2-19 doParallel_1.0.17
```

```
## [49] lattice_0.22-5      plyr_1.8.8           shiny_1.7.4
## [52] withr_3.0.0         benchmarkmeData_1.0.4 coda_0.19-4
## [55] evaluate_0.20       polyclip_1.10-6      zip_2.3.0
## [58] pillar_1.9.0        janeaustenr_1.0.0    foreach_1.5.2
## [61] DT_0.27             plotly_4.10.1        generics_0.1.3
## [64] vroom_1.6.1         hms_1.1.3           munsell_0.5.0
## [67] sensobol_1.1.4      xtable_1.8-4         leaps_3.1
## [70] glue_1.7.0          tikzDevice_0.12.4    emmeans_1.8.5
## [73] scatterplot3d_0.3-43 lazyeval_0.2.2       tools_4.3.3
## [76] tokenizers_0.3.0    mvtnorm_1.1-3        graphlayouts_1.0.2
## [79] XML_3.99-0.14       grid_4.3.3          rbibutils_2.2.16
## [82] rscopus_0.6.6       colorspace_2.1-0     dimensionsR_0.0.3
## [85] ggforce_0.4.1       bibliometrixData_0.3.0 cli_3.6.2
## [88] fansi_1.0.6         viridisLite_0.4.2    gtable_0.3.4
## [91] digest_0.6.34       ggrepel_0.9.5        FactoMineR_2.8
## [94] htmlwidgets_1.6.2   farver_2.1.1         htmltools_0.5.5
## [97] factoextra_1.0.7    lifecycle_1.0.4      http_1.4.5
## [100] multcompView_0.1-9  mime_0.12            bit64_4.0.5
## [103] MASS_7.3-60.0.1
```

```
## Return the machine CPU
```

```
cat("Machine:      "); print(get_cpu())$model_name)
```

```
## Machine:
```

```
## [1] "Apple M1 Max"
```

```
## Return number of true cores
```

```
cat("Num cores:    "); print(detectCores(logical = FALSE))
```

```
## Num cores:
```

```
## [1] 10
```

```
## Return number of threads
```

```
cat("Num threads: "); print(detectCores(logical = FALSE))
```

```
## Num threads:
```

```
## [1] 10
```