

More complex models produce more uncertain estimates

R code

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```

# PRELIMINARY FUNCTIONS -----

# Function to read in all required packages in one go:
loadPackages <- function(x) {
  for(i in x) {
    if(!require(i, character.only = TRUE)) {
      install.packages(i, dependencies = TRUE)
      library(i, character.only = TRUE)
    }
  }
}

# Load the packages
loadPackages(c(
  "data.table", "tidyverse", "parallel", "doParallel", "deSolve",
  "sensobol", "crone", "KScorrect", "cowplot", "wesanderson",
  "Rfast", "RcppAlgos", "scales", "pracma", "grid"))

# Create custom theme
theme_AP <- function() {
  theme_bw() +
    theme(panel.grid.major = element_blank(),
          panel.grid.minor = element_blank(),
          legend.background = element_rect(fill = "transparent",
                                            color = NA),
          legend.key = element_rect(fill = "transparent",
                                     color = NA),
          strip.background = element_rect(fill = "white"))
}

# Set checkpoint

dir.create(".checkpoint")
library("checkpoint")

checkpoint("2021-05-10",
          R.version = "4.0.3",
          checkpointLocation = getwd())

```

1 PSACOIN model

```
# PSACOIN MODEL -----

psacoin0_fun <- function(R_0, X_B, K_DB, K_DG, D_G0, X_G,
                        alpha_G, V_G, W, W_M, t) {

  # WASTE MODEL -----
  #-----

  # Constants
  I_0 <- 2.035e-5
  lambda <- 1.07e-5
  S <- 1.2e6
  Q <- 2e+08

  tau_0 <- Q / (R_0 * S)
  I <- I_0 * exp(-lambda * t)
  heaviside_out <- ifelse((t - tau_0) < 0, 1, 0)
  R_wf <- R_0 * (1 - heaviside_out)
  Fw <- R_wf * I * S

  # BUFFER MODEL -----
  #-----

  # Constants
  p_B <- 1.85e3
  e_B <- 0.099
  D_B <- 0.03

  R_B <- 1 + ((1 - e_B) * K_DB * p_B) / e_B
  tau_B <- (X_B^2 * R_B) / (4 * D_B)
  F_B <- ifelse(t < tau_B, 0, Fw * (t - tau_B) * exp(-lambda * tau_B))

  # GEOSPHERE MODEL -----
  #-----

  # Constants
  p_G <- 2e3
  epsilon_G <- 0.3

  D_G <- D_G0 + alpha_G * V_G
  R_G <- 1 + p_G / epsilon_G * (1 - epsilon_G) * K_DG

  alpha <- X_G
  beta <- 2 * sqrt(D_G / R_G)
  gamma <- V_G / R_G
```

```

a <- (gamma / beta)^2
b <- -((2 * alpha * gamma) / beta^2 + 1)
c <- (alpha / beta)^2

tau_H <- (-b + sqrt(b^2 - 4 * a * c)) / (2 * a)
tau_L <- (-b - sqrt(b^2 - 4 * a * c)) / (2 * a)

t_prim <- ((t - tau_L - tau_B) / (tau_H + tau_0 - tau_L)) * tau_0 + tau_B
F_G <- ifelse(t < (tau_B + tau_L) |
              t > (tau_B + tau_H + tau_0), 0,
              tau_0 / (tau_H + tau_0 - tau_L) * F_B * t_prim * exp(-lambda * (t - t_prim)))

# BIOSPHERE MODEL -----
#-----

# Constant
A <- 2.04e11
D <- 2.3e-9

C <- (F_G * A) / W
H <- C * W_M * D

#####

return(list(Fw, F_B, F_G, H))
}

# SETTINGS PSACOIN -----

N <- 2^15
R <- 10^3
matrices <- c("A", "B", "AB", "BA")
order <- "third"
first <- "azzini"
total <- "azzini"
n_cores <- detectCores() * 0.75

# RUN PSACOIN IN EACH COMPARTMENT -----

# BUFFER -----

params.buffer <- c("R_0", "X_B", "K_DB")
mat <- sobol_matrices(params = params.buffer, N = N, scrambling = 1,
                     matrices = matrices, order = order)

# Precise distributions

```

```

mat[, "R_0"] <- qlunif(mat[, "R_0"], 10^-2.57, 10^1.11)
mat[, "X_B"] <- qunif(mat[, "X_B"], 0.5, 5)
mat[, "K_DB"] <- qlnorm(mat[, "K_DB"], -2.38, 0.143) # Selenium

y.buffer <- psacoin0_fun(R_0 = mat[, "R_0"],
                        X_B = mat[, "X_B"],
                        K_DB = mat[, "K_DB"],
                        K_DG = NULL,
                        D_GO = NULL,
                        X_G = NULL,
                        alpha_G = NULL,
                        V_G = NULL,
                        W = NULL,
                        W_M = NULL,
                        t = 10^7)[[2]]

# GEOSPHERE -----

params.geo <- c("R_0", "X_B", "K_DB", "K_DG", "D_GO", "X_G", "alpha_G", "V_G")
mat <- sobol_matrices(params = params.geo, N = N, scrambling = 1,
                     matrices = matrices, order = order)

mat[, "R_0"] <- qlunif(mat[, "R_0"], 10^-2.57, 10^1.11)
mat[, "X_B"] <- qunif(mat[, "X_B"], 0.5, 5)
mat[, "K_DB"] <- qlnorm(mat[, "K_DB"], -2.38, 0.143) # Selenium
mat[, "K_DG"] <- qlnorm(mat[, "K_DG"], -3.38, 0.3) # Selenium
mat[, "D_GO"] <- qnorm(mat[, "D_GO"], 0.04, 0.001)
mat[, "X_G"] <- qunif(mat[, "X_G"], 10^3, 10^4)
mat[, "alpha_G"] <- qlunif(mat[, "alpha_G"], 10^0.3, 10^2.3)
mat[, "V_G"] <- qlunif(mat[, "V_G"], 10^-3, 10^-1)

y.geo <- psacoin0_fun(R_0 = mat[, "R_0"],
                     X_B = mat[, "X_B"],
                     K_DB = mat[, "K_DB"],
                     K_DG = mat[, "K_DG"],
                     D_GO = mat[, "D_GO"],
                     X_G = mat[, "X_G"],
                     alpha_G = mat[, "alpha_G"],
                     V_G = mat[, "V_G"],
                     W = NULL,
                     W_M = NULL,
                     t = 10^7)[[3]]

# BIOSPHERE -----

params <- c("R_0", "X_B", "K_DB", "K_DG", "D_GO", "X_G",
           "alpha_G", "V_G", "W", "W_M")

```

```

mat <- sobol_matrices(params = params, N = N, scrambling = 1,
                     matrices = matrices, order = order)

mat[, "R_0"] <- qlunif(mat[, "R_0"], 10^-2.57, 10^1.11)
mat[, "X_B"] <- qunif(mat[, "X_B"], 0.5, 5)
mat[, "K_DB"] <- qlnorm(mat[, "K_DB"], -2.38, 0.143) # Selenium
mat[, "K_DG"] <- qlnorm(mat[, "K_DG"], -3.38, 0.3) # Selenium
mat[, "D_GO"] <- qnorm(mat[, "D_GO"], 0.04, 0.001)
mat[, "X_G"] <- qunif(mat[, "X_G"], 10^3, 10^4)
mat[, "alpha_G"] <- qlunif(mat[, "alpha_G"], 10^0.3, 10^2.3)
mat[, "V_G"] <- qlunif(mat[, "V_G"], 10^-3, 10^-1)
mat[, "W"] <- qunif(mat[, "W"], 5 * 10^5, 5 * 10^6)
mat[, "W_M"] <- qunif(mat[, "W_M"], 0.7, 0.9)

```

```

y.bio <- psacoin0_fun(R_0 = mat[, "R_0"],
                     X_B = mat[, "X_B"],
                     K_DB = mat[, "K_DB"],
                     K_DG = mat[, "K_DG"],
                     D_GO = mat[, "D_GO"],
                     X_G = mat[, "X_G"],
                     alpha_G = mat[, "alpha_G"],
                     V_G = mat[, "V_G"],
                     W = mat[, "W"],
                     W_M = mat[, "W_M"],
                     t = 10^7)

```

ARRANGE OUTPUT OF THE COMPARTMENTS -----

```

model.levels <- c("Waste", "Buffer", "Geosphere", "Biosphere")

names(y.bio) <- model.levels

full.output <- do.call(cbind, y.bio) %>%
  data.table()

A <- full.output %>%
  .[1:N]

```

RUN MODEL DYNAMICS -----

```

mat <- sobol_matrices(params = params, N = 2^11, matrices = "A")

mat[, "R_0"] <- qlunif(mat[, "R_0"], 10^-2.57, 10^1.11)
mat[, "X_B"] <- qunif(mat[, "X_B"], 0.5, 5)
mat[, "K_DB"] <- qlnorm(mat[, "K_DB"], -2.38, 0.143) # Selenium
mat[, "K_DG"] <- qlnorm(mat[, "K_DG"], -3.38, 0.3) # Selenium
mat[, "D_GO"] <- qnorm(mat[, "D_GO"], 0.04, 0.001)
mat[, "X_G"] <- qunif(mat[, "X_G"], 10^3, 10^4)

```

```

mat[, "alpha_G"] <- qlunif(mat[, "alpha_G"], 10^0.3, 10^2.3)
mat[, "V_G"] <- qlunif(mat[, "V_G"], 10^-3, 10^-1)
mat[, "W"] <- qunif(mat[, "W"], 5 * 10^5, 5 * 10^6)
mat[, "W_M"] <- qunif(mat[, "W_M"], 0.7, 0.9)

t <- seq(1, 8, 0.05)

y <- mclapply(t, function(t)
  psacoin0_fun(R_0 = mat[, "R_0"],
              X_B = mat[, "X_B"],
              K_DB = mat[, "K_DB"],
              K_DG = mat[, "K_DG"],
              D_GO = mat[, "D_GO"],
              X_G = mat[, "X_G"],
              alpha_G = mat[, "alpha_G"],
              V_G = mat[, "V_G"],
              W = mat[, "W"],
              W_M = mat[, "W_M"],
              t = 10^t),
  mc.cores = n_cores)

# ARRANGE OUTPUT -----

out <- lapply(y, function(x)
  do.call(cbind, x) %>%
  data.table)

names(out) <- 10^t

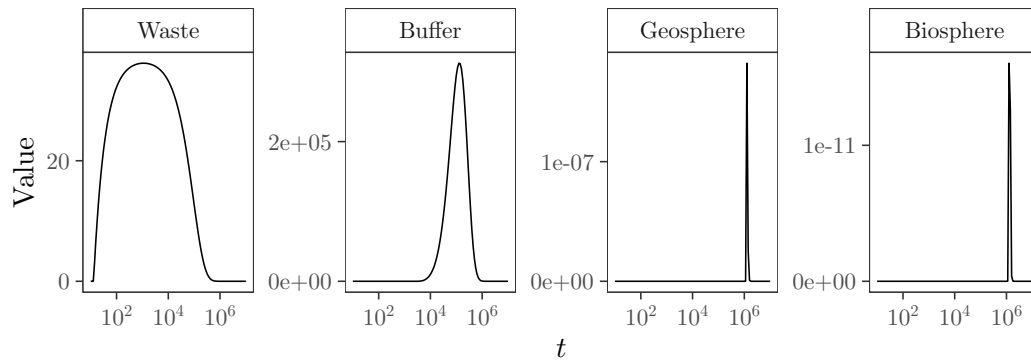
out.plot <- rbindlist(out, idcol = "time") %>%
  setnames(., c("time", paste("V", 1:4, sep = "")),
           c("time", model.levels)) %>%
  melt(., measure.vars = model.levels) %>%
  .[, time:= as.numeric(time)] %>%
  .[, mean(value), .(time, variable)] %>%
  .[time >= 10^1 & time <= 10^7]

# PLOT DYNAMICS PSACOIN -----

plot.psacoin <- ggplot(out.plot, aes(time, V1, group = variable)) +
  geom_line() +
  facet_wrap(~variable, scales = "free_y", ncol = 4) +
  scale_x_log10(labels = scales::trans_format("log10", scales::math_format(10^.x))) +
  scale_y_continuous(breaks = pretty_breaks(n = 2)) +
  labs(x = "$t$", y = "Value") +
  theme_AP()

plot.psacoin

```



```
# SENSITIVITY ANALYSIS -----

ind.psacoin <- list()
for(i in 2:4) {
  if(i == 2) {
    y <- y.buffer
    params <- params.buffer
  } else if(i == 3) {
    y <- y.geo
    params <- params.geo
  } else {
    y <- y.bio[[4]]
    params <- c("R_0", "X_B", "K_DB", "K_DG", "D_GO", "X_G",
                "alpha_G", "V_G", "W", "W_M")
  }
  ind.psacoin[[i]] <- sobol_indices(Y = y, N = N, params = params,
                                   first = first, total = total,
                                   R = R, boot = TRUE,
                                   parallel = "multicore", order = order,
                                   ncpus = n_cores, matrices = matrices)
}

# ARRANGE DATA -----

out <- lapply(ind.psacoin, function(x) x$results)[2:4]
names(out) <- c("Buffer", "Geosphere", "Biosphere")

ind.psacoin <- rbindlist(out, idcol = "compartment") %>%
  .[, compartment:= factor(compartment, levels = c("Buffer", "Geosphere", "Biosphere"))]

# Sum of first, second and third-order effects in each compartment
ind.psacoin[!sensitivity == "Ti", sum(original), compartment]
```

```
##      compartment      V1
## 1:      Buffer 1.0000174
## 2:    Geosphere 0.4548629
## 3:    Biosphere 0.3758309
```



```

psacoin.cv <- A[, lapply(.SD, function(x)
  sd(x, na.rm = TRUE) / mean(x, na.rm = TRUE)), .SDcols = (model.levels)] %>%
melt(., measure.vars = model.levels, value.name = "cv")

psacoin.kt <- ind.psacoin[sensitivity == "Ti" & original > 0.05] %>%
  .[, .(kt = length(unique(parameters))), compartment] %>%
  rbind(., list("Waste", 1)) %>%
  setnames(., "compartment", "variable")

psacoin.cv.kt <- merge(psacoin.cv, psacoin.kt, by = "variable") %>%
  .[, model:= "PSACOIN"]

print(psacoin.cv.kt)

```

```

##      variable      cv kt  model
## 1:      Waste 1.799618  1 PSACOIN
## 2:      Buffer 2.363077  2 PSACOIN
## 3: Geosphere 7.194928  5 PSACOIN
## 4: Biosphere 9.051070  6 PSACOIN

```

PLOT PSACOIN CV -----

```

psacoin.cv.plot <- ggplot(psacoin.cv.kt, aes(variable, cv, group = 1)) +
  geom_line() +
  geom_point(aes(size = kt)) +
  scale_x_discrete(guide = guide_axis(n.dodge = 2)) +
  scale_size_continuous(name = "$k_t$", range = c(1, 4)) +
  labs(x = "", y = "CV") +
  theme_AP() +
  theme(legend.position = "none")

```

PLOT -----

```

psacoin.ks.plot <- ind.psacoin[!sensitivity == "Ti"] %>%
  .[, sum(original), .(sensitivity, compartment)] %>%
  ggplot(., aes(compartment, V1, fill = sensitivity, order = sensitivity)) +
  geom_col(position = position_stack(reverse = TRUE), color = "black") +
  labs(x = "", y = "$\\sum S_i + \\sum S_{i,j} + \\sum S_{i,j,l}$") +
  scale_fill_manual(name = "Sensitivity",
    labels = c("$S_i$", "$S_{i,j}$", "$S_{i,j,l}$"),
    values = wes_palette("Cavalcanti1")) +
  geom_hline(yintercept = 0.99, lty = 2) +
  scale_x_discrete(guide = guide_axis(n.dodge = 2)) +
  theme_AP() +
  theme(legend.position = "none",
    axis.title.y = element_text(size = 7)) +
  annotation_custom(textGrob("p", gp = gpar(col = "red")),
    xmin = 3.5, xmax = 3.5, ymin = 0.98, ymax = 0.98)

```

```
# PLOT ALL FIGURES PSACOIN -----
```

```
psacoin.plots <- plot_grid(psacoin.cv.plot,  
                           psacoin.ks.plot, ncol = 2, labels = "auto")
```

```
# EXPORT PSACOIN SENSITIVITY INDICES -----
```

```
fwrite(ind.psacoin, "ind.psacoin.csv")
```

2 The Bateman equations

```
# BATEMAN EQUATIONS -----

bateman <- function(t, state, parameters) {
  with(as.list(c(state, parameters)), {
    dx1 <- - lambda1 * x1
    dx2 <- - lambda2 * x2 + lambda1 * x1
    dx3 <- - lambda3 * x3 + lambda2 * x2
    dx4 <- - lambda4 * x4 + lambda3 * x3
    dx5 <- - lambda5 * x5 + lambda4 * x4
    dx6 <- - lambda6 * x6 + lambda5 * x5
    dx7 <- - lambda7 * x7 + lambda6 * x6
    dx8 <- - lambda8 * x8 + lambda7 * x7
    dx9 <- - lambda9 * x9 + lambda8 * x8
    dx10 <- - lambda10 * x10 + lambda9 * x9
    list(c(dx1, dx2, dx3, dx4, dx5, dx6, dx7, dx8, dx9, dx10))
  })
}

# BATEMAN SETTINGS -----

# settings
t <- 20
x1 <- 100
sample.size <- 2 ^ 12
times <- seq(0, 20, 0.01)
params <- c(100, rep(0, 9))
names(params) <- paste("x", 1:10, sep = "")

# SAMPLE MATRIX -----

mat <- sensobol::sobol_matrices(matrices = "A", N = sample.size,
                              params = paste("lambda", 1:10, sep = ""), type = "R")
mat <- apply(mat, 2, function(x) KScorrect::qlunif(x, 0.001, 10))

# RUN DIFFERENTIAL EQUATIONS -----

Y <- data.table(ode(y = params,
                  times = times,
                  func = bateman,
                  parms = colMeans(mat)))

# PLOT BATEMAN DYNAMICS -----

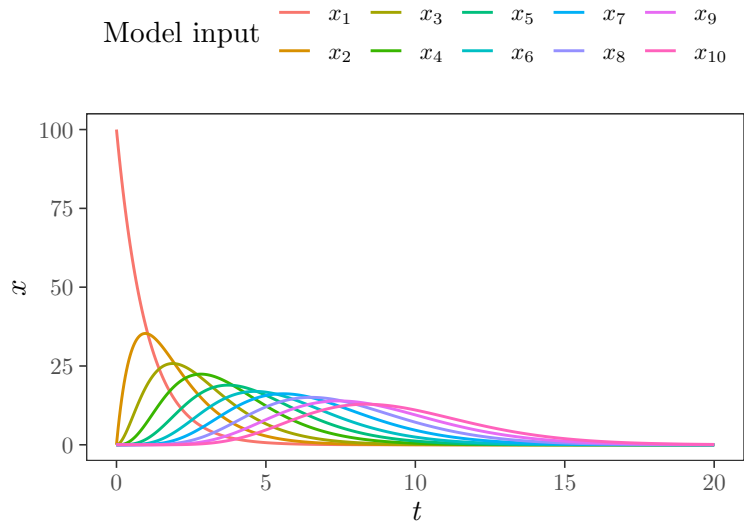
plot.bateman <- melt(Y, measure.vars = paste("x", 1:10, sep = "")) %>%
  ggplot(., aes(time, value, color = variable)) +
  geom_line(size = 1) +
  labs(x = expression(italic(t)), y = "$x$") +
```

```

scale_color_discrete(name = "Model input",
                      labels = paste("$x_{", 1:10, "}$", sep = "")) +
theme_AP() +
theme(legend.position = "top")

```

```
plot.bateman
```



```

# DEFINE BATEMAN EQUATIONS FOR K -----

# Define Bateman equations for xk
bateman_fun <- function(x1, lambda, t) {
  out <- list()
  for(i in 1:length(lambda)) {
    out[[i]] <- (lambda / (lambda - lambda[i]))
  }
  alpha <- unlist(lapply(out, function(x) prod(x[!is.infinite(x)])))
  xk <- x1 / lambda[length(lambda)] * sum(lambda * alpha * exp(1) ^ (-lambda * t))
  return(xk)
}

# Wrapper to run bateman equations rowwise
bateman_rowwise <- function(x1, mat, t) {
  out <- vector()
  for(i in 1:nrow(mat)) {
    out[i] <- bateman_fun(x1 = x1, lambda = mat[i, ], t = t)
  }
  return(out)
}

# DEFINE SETTINGS -----

t <- 10
x1 <- 100

```

```

k <- seq(2, t, 1)
sample.size <- 2 ^ 15
matrices <- "A"

# RUN BATEMAN IN PARALLEL -----

# Define parallel computing
n_cores <- detectCores() * 0.75
cl <- makeCluster(n_cores)
registerDoParallel(cl)

y <- foreach(i = 1:length(k),
             .packages = c("sensobol", "data.table", "KScorrect")) %dopar%
{
  params <- paste("x", 1:k[i], sep = "")
  set.seed(2)
  tmp <- sensobol::sobol_matrices(matrices = matrices, N = sample.size, params = params,
                                type = "R")
  mat <- apply(tmp, 2, function(x) KScorrect::qlunif(x, 0.001, 10))
  out <- bateman_rowwise(x1 = x1, mat = mat, t = k[i])
}

# Stop parallel cluster
stopCluster(cl)

# ARRANGE OUTPUT -----

# Arrange data
tmp <- lapply(y, data.table::data.table)
names(tmp) <- k
dt <- data.table::rbindlist(tmp, idcol = "k")
dt <- dt[, k:= as.numeric(k)]
A <- dt[, .SD[1:(2 * sample.size)], k]

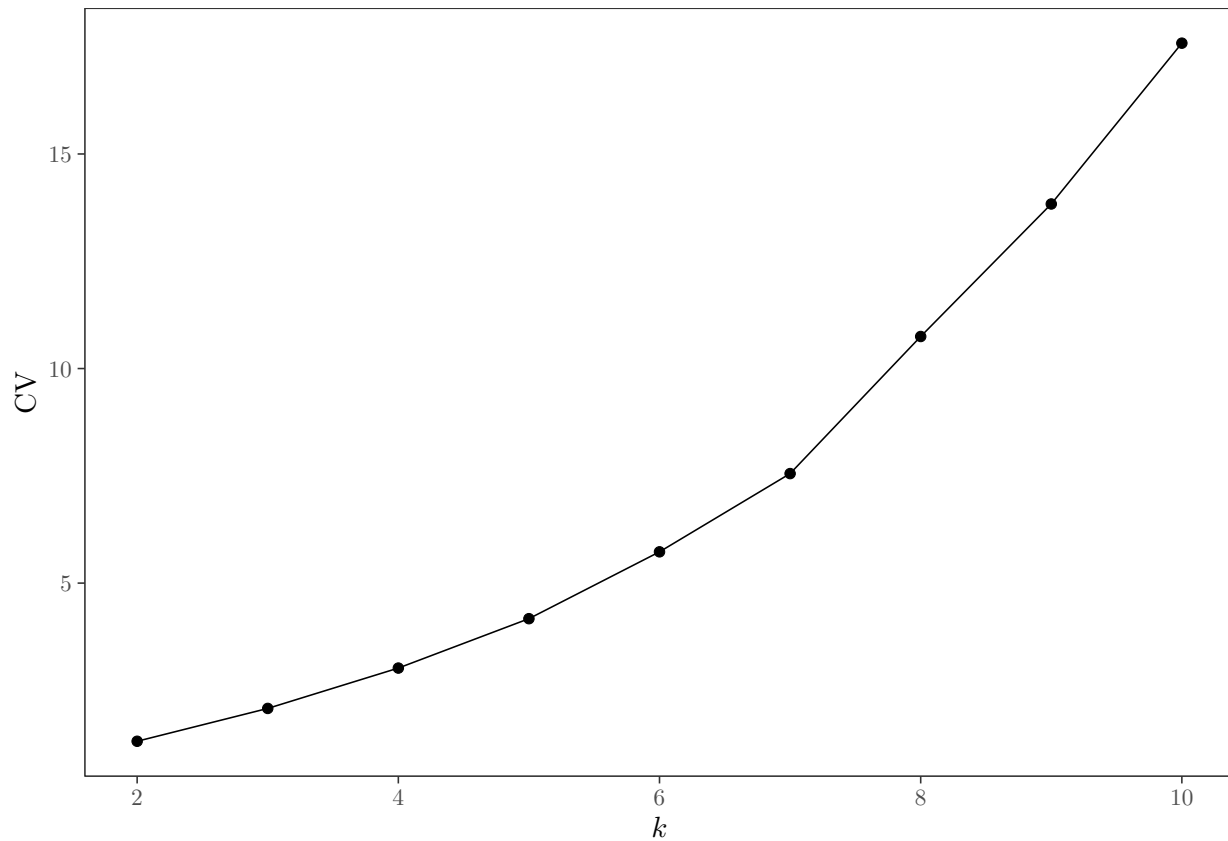
# Compute coefficient of variation and median absolute deviation
dt.stat <- A[, .(cv = sd(V1, na.rm = TRUE) / mean(V1, na.rm = TRUE)), k]

# COEFFICIENT OF VARIATION -----

cv.bateman <- ggplot(dt.stat, aes(k, cv)) +
  geom_line() +
  geom_point() +
  labs(x = "$k$", y = "CV") +
  theme_AP()

cv.bateman

```



```
# SENSITIVITY ANALYSIS -----

order <- "third"
k <- 3:10
sample.size <- 2 ^ 15
matrices <- c("A", "B", "AB", "BA")
first <- "azzini"
second <- "azzini"
#-----

# Define parallel computing
n_cores <- detectCores() * 0.75
cl <- makeCluster(n_cores)
registerDoParallel(cl)

ind.bateman <- foreach(i = 1:length(k),
  .packages = c("sensobol", "data.table", "KScorrect")) %dopar%
{
  params <- paste("X", 1:k[i], sep = "")
  mat <- sensobol::sobol_matrices(matrices = matrices, N = sample.size,
    params = params, order = order,
    scrambling = 1)
  mat <- apply(mat, 2, function(x) KScorrect::qlunif(x, 0.001, 10))
  y <- bateman_rowwise(x1 = x1, mat = mat, t = k[i])
}
```

```

    sobol_indices(matrices = matrices, Y = y, N = sample.size,
                  params = params, order = order, first = first,
                  total = total, R = R, boot = TRUE)
  }

# Stop parallel cluster
stopCluster(cl)

# ARRANGE BATEMAN SENSITIVITY ANALYSIS -----

# Compute coefficient of variation and median absolute deviation
bateman.cv <- A[, .(cv = sd(V1, na.rm = TRUE) / mean(V1, na.rm = TRUE)), k]

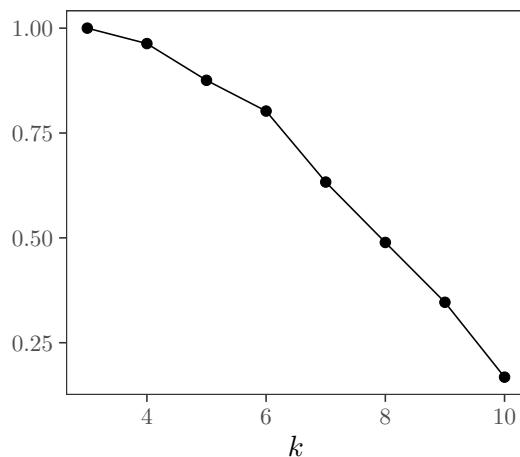
out <- lapply(ind.bateman, function(x) x$results)
names(out) <- k

ind.bateman <- rbindlist(out, idcol = "dim")

# SUM UP TO THIRD-ORDER EFFECTS-----

ind.bateman[!sensitivity == "Ti", sum(original), dim] %>%
  .[, dim:= as.numeric(dim)] %>%
  ggplot(. , aes(dim, V1)) +
  geom_line() +
  labs(x = "$k$", y = "") +
  geom_point() +
  theme_AP()

```



```

# PLOT BATEMAN CV -----

bateman.kt <- ind.bateman[sensitivity == "Ti" & original > 0.05] %>%
  .[, .(kt= length(unique(parameters))), dim] %>%
  setnames(., "dim", "variable") %>%
  .[, variable:= as.numeric(variable)]

```

```

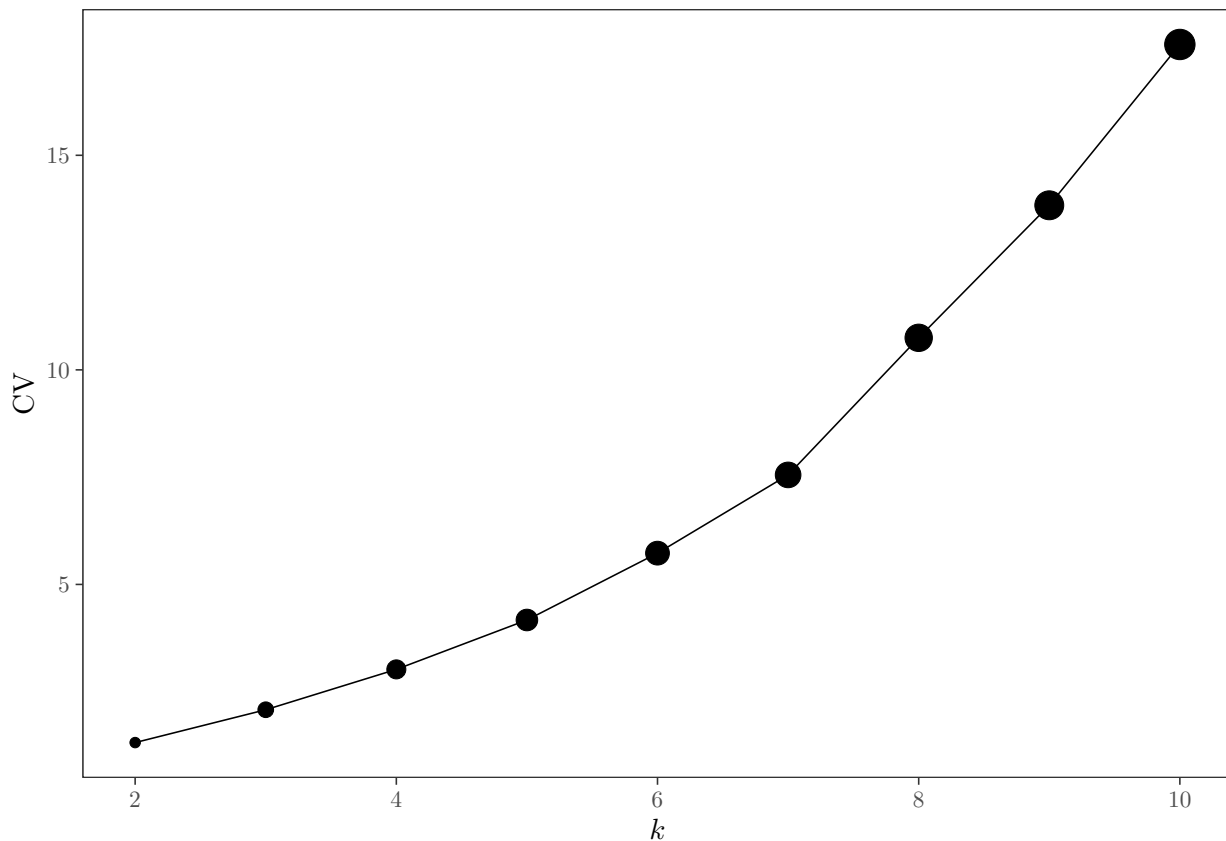
bateman.cv <- bateman.cv[, kt:= c(2, bateman.kt$kt)]

bateman.cv.plot <- ggplot(bateman.cv, aes(k, cv)) +
  geom_line() +
  geom_point(aes(size = kt)) +
  geom_point() +
  scale_size_continuous(name = "$k_t$", range = c(1, 5)) +
  labs(x = "$k$", y = "CV") +
  theme_AP() +
  theme(legend.position = "none")

legend.kt <- get_legend(bateman.cv.plot + theme(legend.position = "top"))

bateman.cv.plot

```



```

# PLOT BATEMAN -----

high.order.bateman <- ind.bateman[!sensitivity == "Ti"] %>%
  .[, sum(original), .(sensitivity, dim)] %>%
  .[, dim:= as.numeric(dim)] %>%
  ggplot(., aes(dim, V1, fill = sensitivity)) +
  geom_col(position = position_stack(reverse = TRUE), color = "black") +
  labs(x = "$k$", y = "$\\sum S_i + \\sum S_{i,j} + \\sum S_{i,j,l}$") +
  geom_hline(yintercept = 0.99, lty = 2) +

```

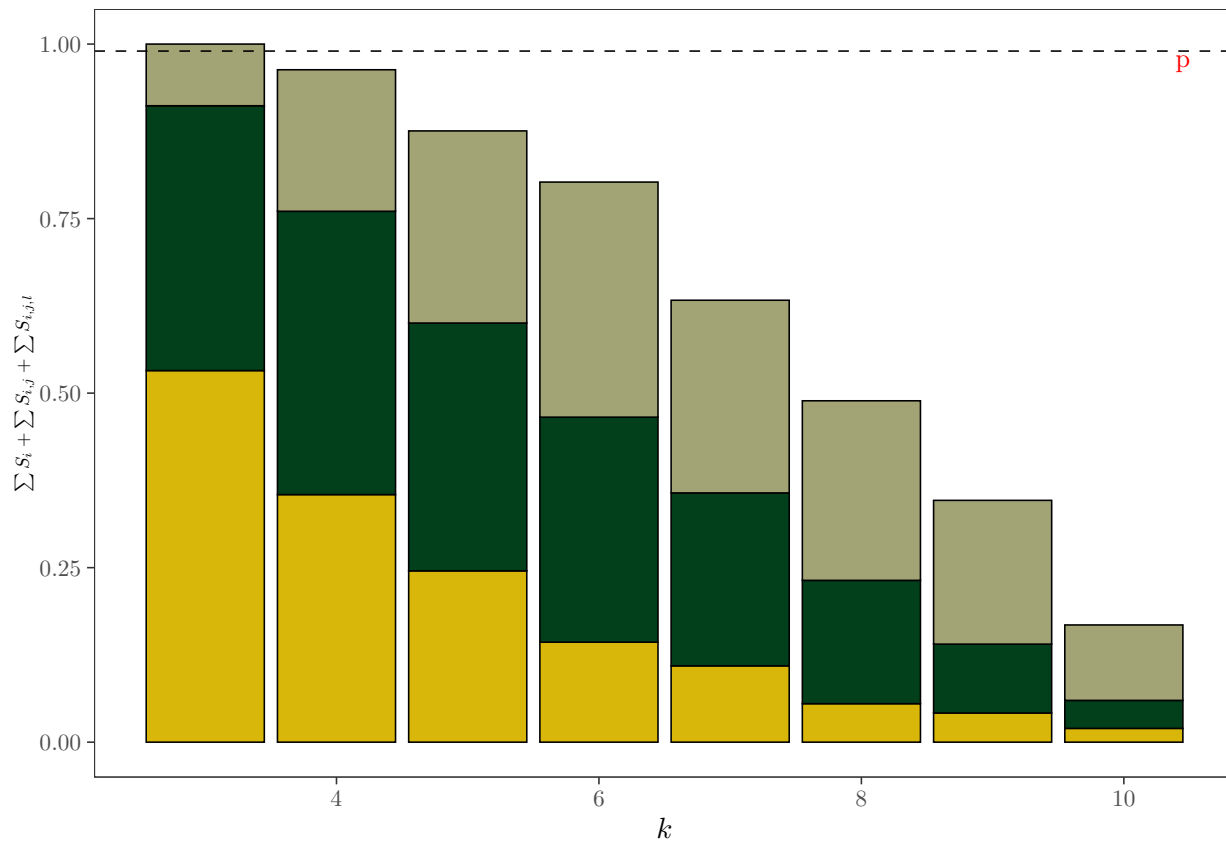


```

scale_fill_manual(name = "",
                  labels = c("$S_i$", "$S_{i,j}$", "$S_{i,j,l}$"),
                  values = wes_palette("Cavalcanti1")) +
annotation_custom(textGrob("p", gp = gpar(col = "red")),
                  xmin = 10.45, xmax = 10.45, ymin = 0.98, ymax = 0.98) +
theme_AP() +
theme(legend.position = "none",
      axis.title.y = element_text(size = 7))

```

high.order.bateman



```
ind.bateman[sensitivity == "Ti" & original > 0.05, length(unique(parameters)), dim]
```

```

##      dim V1
## 1:    3  3
## 2:    4  4
## 3:    5  5
## 4:    6  6
## 5:    7  7
## 6:    8  8
## 7:    9  9
## 8:   10 10

```

```

legend.ks <- get_legend(high.order.bateman + theme(legend.position = "top",
                                                    legend.key.size = unit(1, 'lines'),
                                                    legend.spacing.x = unit(0.2, "cm")))

```

PLOT BATEMAN2 -----

```

bateman.plots <- plot_grid(bateman.cv.plot, high.order.bateman,
                           ncol = 2, labels = c("c", "d"))

```

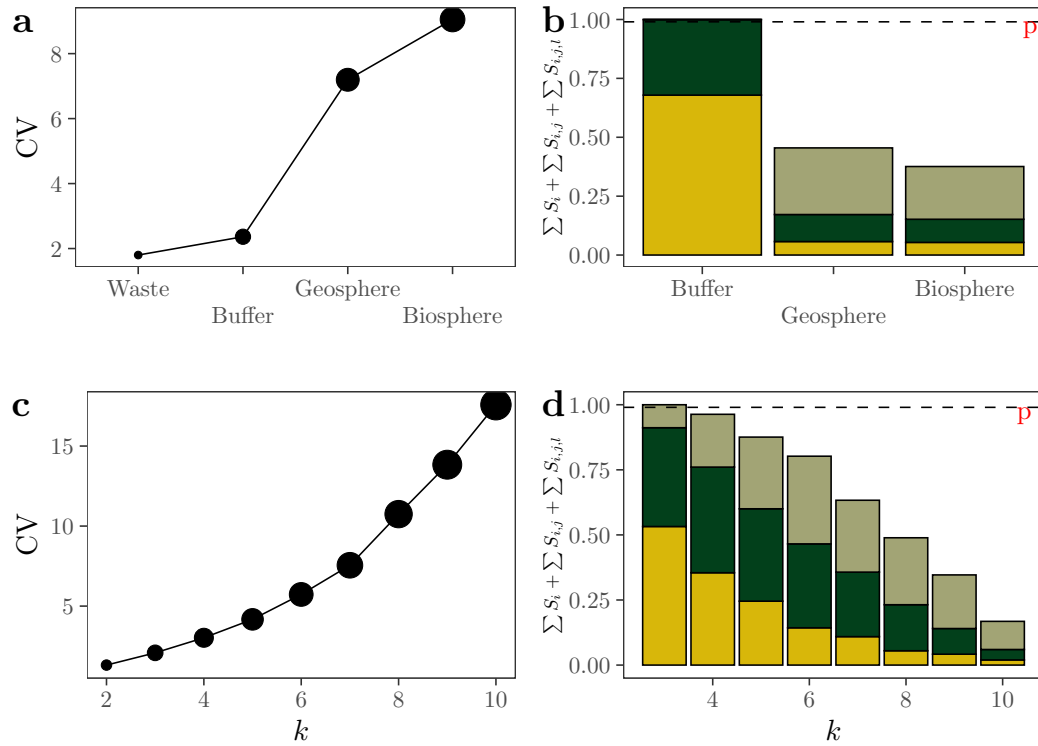
MERGE ALL PLOTS -----

```

all.legends <- plot_grid(legend.kt, legend.ks, ncol = 2)
all.plots <- plot_grid(psacoin.plots, bateman.plots, ncol = 1)
plot_grid(all.legends, all.plots, ncol = 1, rel_heights = c(0.1, 0.8))

```

k_t • 2 ● 4 ● 6 ● 8 ● 10 S_i $S_{i,j}$ $S_{i,j,t}$



EXPORT BATEMAN SENSITIVITY INDICES -----

```

fwrite(ind.bateman, "ind.bateman.csv")

```

3 COVID-19 models

```
# PRELIMINARIES -----

# Define timesteps (5 years)
times <- seq(1, 5 * 52, 1)

# Read input data
R0.listOrig <- fread("NY.csv")
R0.list.a <- R0.listOrig$hku1

shift <- 0
R0.list.shifted <- circshift(R0.list.a, -shift)
R0.list <- rep(R0.list.shifted, length = length(times))

# COVID MODELS -----

# SIR(S) with seasonality
sir_s_fun <- function(t, state, parameters) {

  with(as.list(c(state, parameters)), {

    beta <- R0.list[t] * gamma

    dSP <- mu * N - (beta * SP * (IP + alpha * IS) / N) - mu * SP
    dIP <- (beta * SP * (IP + alpha * IS) / N) - (gamma + mu) * IP
    dR <- gamma * (IP + IS) - (delta + mu) * R
    dSS <- delta * R - epsilon * (beta * SS * (IP + alpha * IS) / N) - mu * SS
    dIS <- epsilon * (beta * SS * (IP + alpha * IS) / N) - (gamma + mu) * IS
    list(c(dSP, dIP, dR, dSS, dIS))
  })
}

# SIR(S) with seasonality and vaccination
sir_s_vaccination_fun <- function(t, state, parameters) {

  with(as.list(c(state, parameters)), {

    beta <- R0.list[t] * gamma

    svax <- 0
    if(t > tvax){
      svax <- 1
    }

    dSP <- mu * N - (beta * SP * (IP + alpha * IS) / N) - mu * SP - svax * nu * SP
    dIP <- (beta * SP * (IP + alpha * IS) / N) - (gamma + mu) * IP
    dR <- gamma * (IP + IS) - (delta + mu) * R
    dSS <- delta * R - epsilon * (beta * SS * (IP + alpha * IS) / N) - mu * SS +
```

```

    delta_vax * V - svax * nu * SS
    dIS <- epsilon * (beta * SS * (IP + alpha * IS) / N) - (gamma + mu) * IS
    dV <- svax * nu * (SP + SS) - delta_vax * V - mu * V
    list(c(dSP, dIP, dR, dSS, dIS, dV))
  })
}

# Extended SIR(S)
sir_s_extended <- function(t, state, parameters) {

  with(as.list(c(state, parameters)), {
    beta <- R0.list[t] * gamma

    svax <- 0
    if(t > tvax){
      svax <- 1
    }

    dSP <- mu - (beta * SP * (IP + alpha * IS + alphaV * IV +
      alpha1 * IS1 + alpha2 * IS2)) - mu * SP - svax * nu * SP
    dIP <- (beta * SP * (IP + alpha * IS + alphaV *
      IV + alpha1 * IS1 + alpha2 * IS2)) - (gamma + mu) * IP
    dR <- gamma * (IP + IS + IV + IS1 + IS2) - (delta + mu) * R
    dSS <- delta * R - epsilon * (beta * SS * (IP + alpha * IS + alphaV * IV + alpha1 * IS1 +
      alpha2 * IS2)) - mu * SS - svax * nu * SS
    dIS <- epsilon * (beta * SS * (IP + alpha * IS + alphaV *
      IV + alpha1 * IS1 + alpha2 * IS2)) - (gamma + mu) * IS
    dV1 <- svax * nu * (SP + d * SS) - epsilonV1 *
      beta * V1 * (IP + alpha * IS + alphaV * IV + alpha1 * IS1 + alpha2 * IS2) -
      (omega + rho1 + mu) * V1
    dV2 <- (1 - d) * svax * nu * SS + omega * V1 - epsilonV2 * beta * V2 *
      (IP + alpha * IS + alphaV * IV + alpha1 * IS1 + alpha2 * IS2) - (rho2 + mu) * V2
    dIV <- (epsilonV1 * beta * V1 + epsilonV2 *
      beta * V2) * (IP + alpha * IS + alphaV *
      IV + alpha1 * IS1 + alpha2 * IS2) - (gamma + mu) * IV
    dSS1 <- rho1 * V1 - epsilon1 * beta * SS1 *
      (IP + alpha * IS + alphaV * IV + alpha1 * IS1 + alpha2 * IS2) - mu * SS1
    dSS2 <- rho2 * V2 - epsilon2 * beta * SS2 *
      (IP + alpha * IS + alphaV * IV + alpha1 * IS1 + alpha2 * IS2) - mu * SS2
    dIS1 <- epsilon1 * beta * SS1 *
      (IP + alpha * IS + alphaV * IV + alpha1 * IS1 + alpha2 * IS2) - (gamma + mu) * IS1
    dIS2 <- epsilon2 * beta * SS2 *
      (IP + alpha * IS + alphaV * IV + alpha1 * IS1 + alpha2 * IS2) - (gamma + mu) * IS2
    list(c(dSP, dIP, dR, dSS, dIS, dV1, dV2, dIV, dSS1, dSS2, dIS1, dIS2))
  })
}

```

```

# MERGE ALL MODELS -----

covid_models <- list(sir_s_fun, sir_s_vaccination_fun, sir_s_extended)
type_model <- c("SIR(S)", "SIR(S)vaccination", "SIR(S)extended")
names(covid_models) <- type_model

# INITIAL VALUES -----

N <- 1 # Total population
I0 <- 1e-9
SP <- 1 - I0 # Fully susceptible individuals
IP <- I0 # Individuals infected
R <- 0 # Immune individuals due to recovery
SS <- 0 # Individuals whose immunity has waned at rate delta
IS <- 0 # Individuals with secondary infection
V <- 0 # Individuals vaccinated
V1 <- V2 <- IV <- SS1 <- SS2 <- IS1 <- IS2 <- 0

# Define list of initial values
all_y <- list(
  c(SP = SP, IP = IP, R = R, SS = SS, IS = IS),
  c(SP = SP, IP = IP, R = R, SS = SS, IS = IS, V = V),
  c(SP = SP, IP = IP, R = R, SS = SS, IS = IS, V1 = V1,
    V2 = V2, IV = IV, SS1 = SS1, SS2 = SS2, IS1 = IS1, IS2 = IS2)
)

# Name them
names(all_y) <- type_model

# CONSTANTS -----

gamma <- 7 / 5 # Recovery rate from primary and secondary infections ]
delta <- 1 / (1 * 52) # Wane rate of full immunity from natural infection
mu <- 1 / (50 * 52) # birth rate at which individuals enter the susceptible class SP

d <- 0.5
epsilonV1 <- 0.1 # First level of immune protection
epsilonV2 <- 0.05 # Second level of immune protection
omega <- 0
rho2 <- rho1 <- 0 # Vaccinal immunity wanes at rho1 and rho2
delta_vax <- 1
epsilon1 <- epsilon2 <- 0.7 # Effect of vaccine 1 and 2; fixed
# after observing that their uncertainty (epsilon1=U(0.5, 0.9);
# epsilon2 = U(0.7, 0.9) does not contribute output uncertainty)

# DEFINE SETTINGS -----

sample.size <- 2^10

```

```

matrices <- c("A", "B", "AB", "BA")
order <- "third"
first <- "azzini"
total <- "azzini"
R <- 10^3

# DEFINE MATRICES -----

# SIR(S) with seasonality
#-----
params <- c("epsilon", "alpha")
# order = second because there are only two uncertain parameters
mat1 <- sobol_matrices(N = sample.size, params = params, scrambling = 1,
                      matrices = matrices, order = "second")

mat1[, "epsilon"] <- qunif(mat1[, "epsilon"], 0.4, 1)
mat1[, "alpha"] <- qunif(mat1[, "alpha"], 0.8, 1)

mat.dt1 <- data.table(mat1[, `:=` (alpha1 = alpha, alpha2 = alpha, alphaV = alpha)])
mat.dt1 <- cbind(mat.dt1, gamma, delta, mu, epsilonV1, epsilonV2, omega)

# SIR(S) with vaccination
#-----
# params <- c("epsilon", "alpha", "nu", "delta_vax", "tvax")
params <- c("epsilon", "alpha", "nu", "tvax")
mat2 <- sobol_matrices(N = sample.size, params = params, scrambling = 1,
                      matrices = matrices, order = order)

mat2[, "epsilon"] <- qunif(mat2[, "epsilon"], 0.4, 1)
mat2[, "alpha"] <- qunif(mat2[, "alpha"], 0.8, 1)
mat2[, "nu"] <- qunif(mat2[, "nu"], 0.001, 0.009)
# mat2[, "delta_vax"] <- qunif(mat2[, "delta_vax"], 0.5, 2)
mat2[, "tvax"] <- floor(qunif(mat2[, "tvax"], 48, 78))

mat.dt2 <- data.table(mat2[, `:=` (alpha1 = alpha, alpha2 = alpha, alphaV = alpha)])
mat.dt2 <- cbind(mat.dt2, gamma, delta, mu, epsilonV1, epsilonV2, omega)

# SIR(S) extended
#-----
# params <- c("epsilon", "alpha", "nu", "delta_vax", "tvax", "epsilon1", "epsilon2")
params <- c("epsilon", "alpha", "nu", "tvax")
mat <- sobol_matrices(N = sample.size, params = params, scrambling = 1,
                     matrices = matrices, order = order)

mat[, "epsilon"] <- qunif(mat[, "epsilon"], 0.4, 1)
mat[, "alpha"] <- qunif(mat[, "alpha"], 0.8, 1)
mat[, "nu"] <- qunif(mat[, "nu"], 0.001, 0.009)

```

```

# mat[, "delta_vax"] <- qunif(mat[, "delta_vax"], 0.5, 2)
mat[, "tvax"] <- floor(qunif(mat[, "tvax"], 48, 78))
# mat[, "epsilon1"] <- qunif(mat[, "epsilon1"], 0.5, 0.9)
# mat[, "epsilon2"] <- qunif(mat[, "epsilon2"], 0.5, 0.7)

# MERGE ALL MATRICES -----

all.mat <- list(mat1, mat2, mat)
all.mat <- lapply(all.mat, data.table) %>%
  lapply(., function(x)
    x[, `:=` (alpha1 = alpha, alpha2 = alpha, alphaV = alpha)]) %>%
  lapply(., function(x) cbind(x, gamma, delta, mu, epsilonV1, epsilonV2, omega,
    delta_vax, epsilon1, epsilon2))

names(all.mat) <- type_model

# RUN MODELS FOR DYNAMICS -----

out <- list()
for (i in names(covid_models)) {
  out[[i]] <- data.table(ode(y = all_y[[i]],
    times = times,
    func = covid_models[[i]],
    parms = colMeans(all.mat[[i]])))
}

# PLOT DYNAMICS -----

bottom <- lapply(out, function(x)
  melt(x, measure.vars = colnames(x)[-1])) %>%
  rbindlist(., idcol = "Model") %>%
  .[, Model := factor(Model, levels = type_model)] %>%
  ggplot(., aes(time, value, color = variable)) +
  geom_line(size = 1) +
  labs(x = expression(italic(t)),
    y = expression(italic(N))) +
  facet_wrap(~ Model) +
  scale_color_discrete(name = "Variable",
    labels = c("$S_P$", "$I_P$", "$R$", "$S_S$",
      "$I_S$", "$V$", "$V_1$", "$V_2$",
      "$I_V$", "$S_{S1}$", "$S_{S2}$", "$I_{S1}$",
      "$I_{S2}$")) +
  theme_AP() +
  theme(legend.position = "none")

legend <- get_legend(bottom + theme(legend.position = "top",
  legend.text = element_text(
    margin = margin(l = -5, unit = "pt")))) +

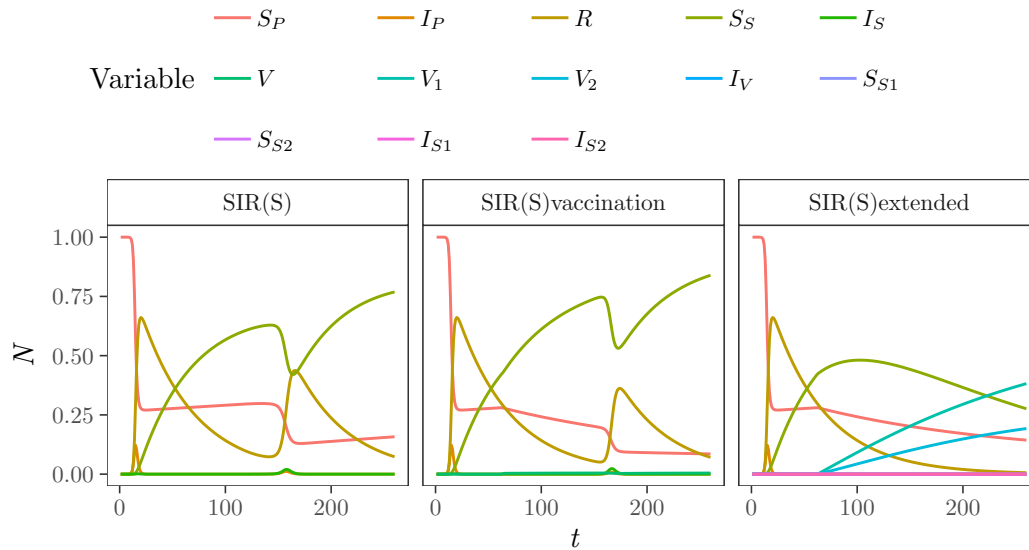
```

```

    guides(color = guide_legend(nrow = 3, byrow=TRUE)))
dynamics.covid <- plot_grid(legend, bottom, ncol = 1, rel_heights = c(0.3, 0.7))

dynamics.covid

```



```

# RUN MODEL IN PARALLEL -----

# Define parallel computing
n_cores <- makeCluster(floor(detectCores() * 0.75))
registerDoParallel(n_cores)

y <- foreach(j = type_model) %:%
  foreach(i = 1:nrow(all.mat[[j]]), .combine = "rbind",
    .packages = "deSolve") %dopar%
  {
    .GlobalEnv$R0.list <- R0.list
    .GlobalEnv$N <- N
    .GlobalEnv$d <- d
    .GlobalEnv$rho1 <- rho1
    .GlobalEnv$rho2 <- rho2
    ode(y = all_y[[j]],
      times = times,
      func = covid_models[[j]],
      parms = all.mat[[j]][i, ])
  }

# Stop clusters
stopCluster(n_cores)

# ARRANGE DATA -----

common.compartments <- c("SP", "IP", "R", "SS", "IS")

```



```

dt.covid <- lapply(y, function(x) data.table(x) %>%
  .[, .SD, .SDcols = c("time", common.compartments)])

names(dt.covid) <- type_model

# Dataset for uncertainty analysis
A <- rbindlist(dt.covid, idcol = "model") %>%
  .[, .SD[1:((sample.size * 2) * length(times))], model] %>%
  .[, model:= factor(model, levels = type_model)] %>%
  melt(., measure.vars = common.compartments)

# Full dataset
out <- rbindlist(dt.covid, idcol = "model") %>%
  melt(., measure.vars = common.compartments) %>%
  .[, model:= factor(model, levels = type_model)]

# PLOT CV -----

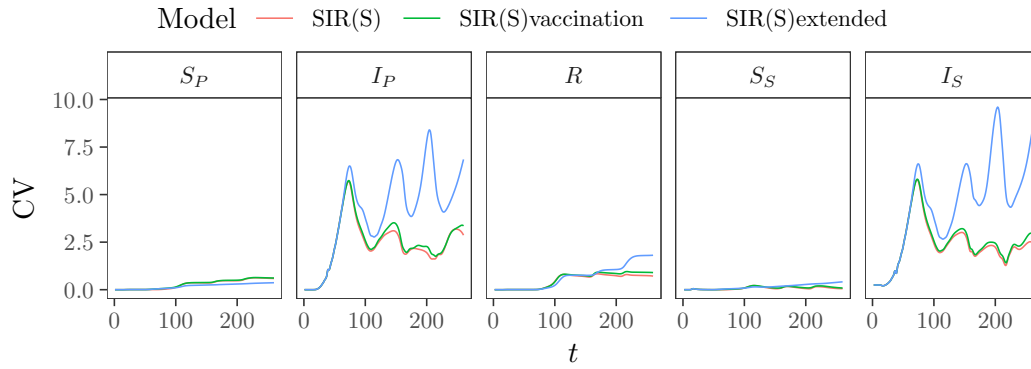
state_names <- c(
  "SP" = "$S_P$",
  "IP" = "$I_P$",
  "R" = "$R$",
  "SS" = "$S_S$",
  "IS" = "$I_S$"
)

covid.cv <- A[, .(cv = sd(value) / mean(value)), .(model, time, variable)] %>%
  ggplot(., aes(time, cv, group = model, color = model)) +
  geom_line() +
  facet_wrap(~variable, ncol = 5, labeller = as_labeller(state_names)) +
  labs(x = "$t$", y = "CV") +
  theme_AP() +
  scale_color_discrete(name = "Model") +
  theme(legend.position = "none")

legend <- get_legend(covid.cv + theme(legend.position = "top"))
cv.covid <- plot_grid(legend, covid.cv, ncol = 1, rel_heights = c(0.1, 0.9))

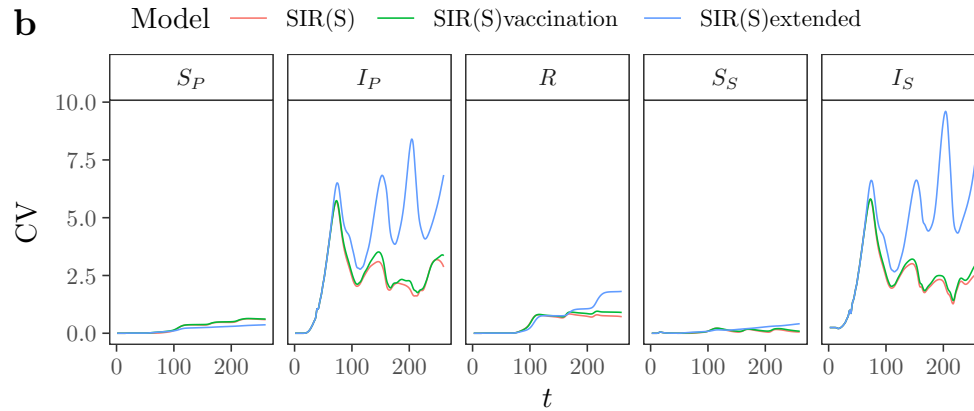
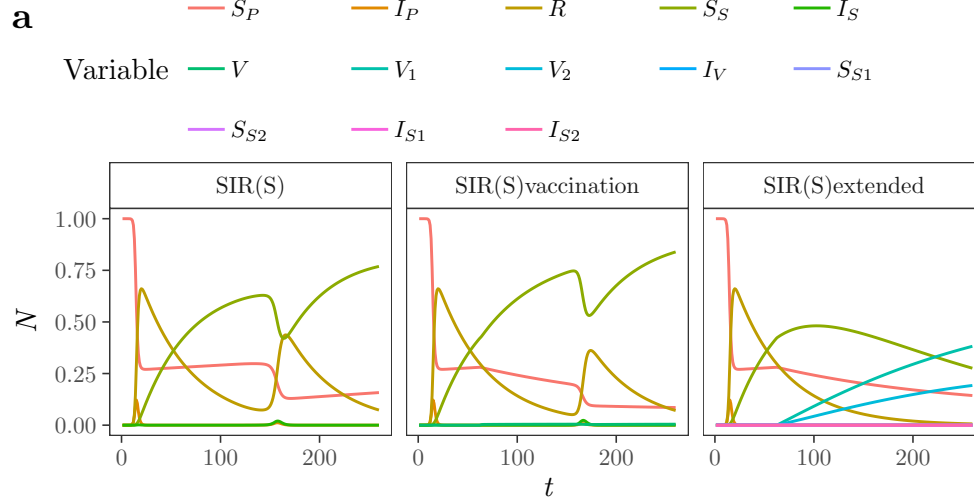
cv.covid

```



MERGE DYNAMICS AND CV COVID MODEL -----

```
plot_grid(dynamics.covid, cv.covid, labels = "auto", ncol = 1,
          rel_heights = c(0.55, 0.45))
```



SENSITIVITY ANALYSIS -----

```
timeSteps <- seq(10, 260, 10)

prove <- lapply(dt.covid, function(x)
  melt(x, measure.vars = common.compartments) %>%
```

```

      .[time %in% timeSteps])

names(prove) <- type_model

ind <- list()
for(i in names(prove)) {
  if(i == "SIR(S)") {
    params <- c("$\\epsilon$", "$\\alpha$")
    order <- "second"
  } else if(i == "SIR(S)vaccination") {
    params <- c("$\\epsilon$", "$\\alpha$", "$\\nu$", "$t_{vax}$")
    order <- "third"
  } else if(i == "SIR(S)extended") {
    params <- c("$\\epsilon$", "$\\alpha$", "$\\nu$", "$t_{vax}$")
    order <- "third"
  }
  ind[[i]] <- prove[[i]][, sobol_indices(Y = value, N = sample.size, matrices = matrices,
                                         params = params, order = order,
                                         first = first, total = total, boot = TRUE,
                                         R = R, parallel = "multicore",
                                         ncpus = detectCores() * 0.75)$results,

                                         .(time, variable))]
}

```

```

## [1] "All values of t are equal to 0.999999999960785 \n Cannot calculate confidence interval"
## [1] "All values of t are equal to 0.999999999967852 \n Cannot calculate confidence interval"
## [1] "All values of t are equal to 0.999999999960825 \n Cannot calculate confidence interval"
## [1] "All values of t are equal to 0.99999999996784 \n Cannot calculate confidence interval"
## [1] "All values of t are equal to 0.999999999960821 \n Cannot calculate confidence interval"
## [1] "All values of t are equal to 0.999999999967854 \n Cannot calculate confidence interval"

```

PLOT SENSITIVITY INDICES -----

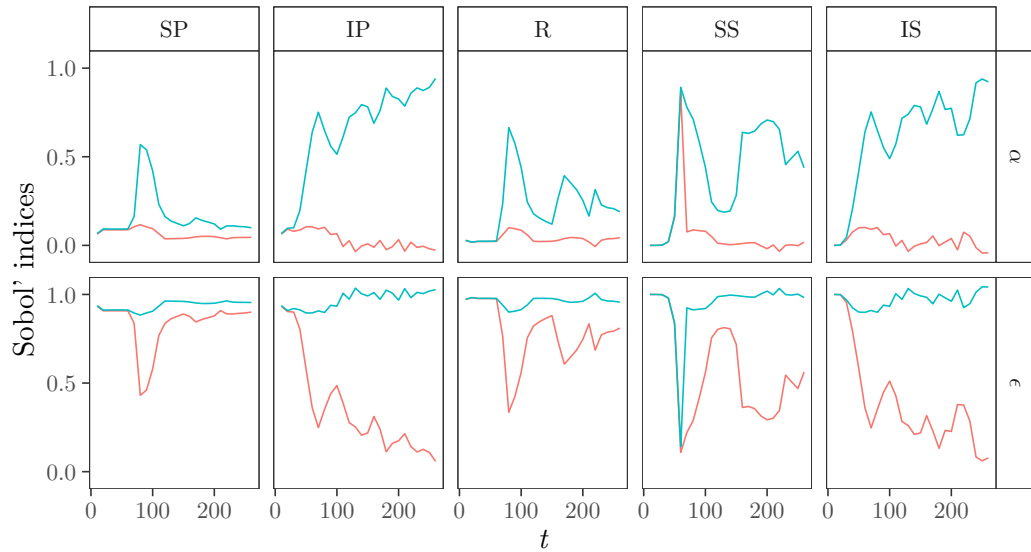
```

plot.ind <- list()
for(i in names(ind)) {
  plot.ind[[i]] <- ggplot(ind[[i]][sensitivity %in% c("Si", "Ti")],
                          aes(time, original, fill = sensitivity,
                              color = sensitivity, group = sensitivity)) +

    geom_line() +
    guides(linetype = FALSE, color = FALSE) +
    facet_grid(parameters ~ variable) +
    scale_y_continuous(breaks = scales::pretty_breaks(n = 3)) +
    labs(x = expression(italic(t)),
         y = "Sobol' indices") +
    theme_AP() +
    theme(legend.position = "top")
}

```

```
plot.ind[[1]]
```

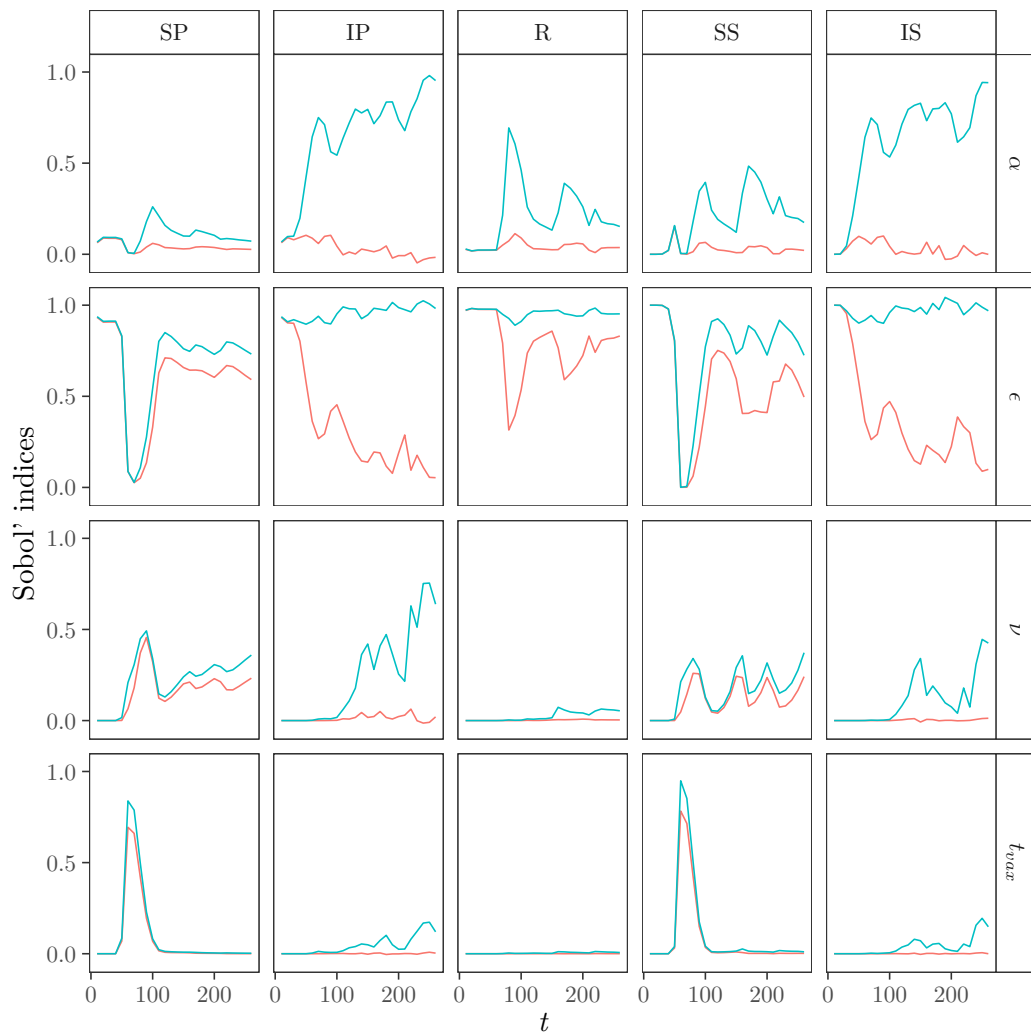


```
# PLOT SENSITIVITY INDICES -----
```

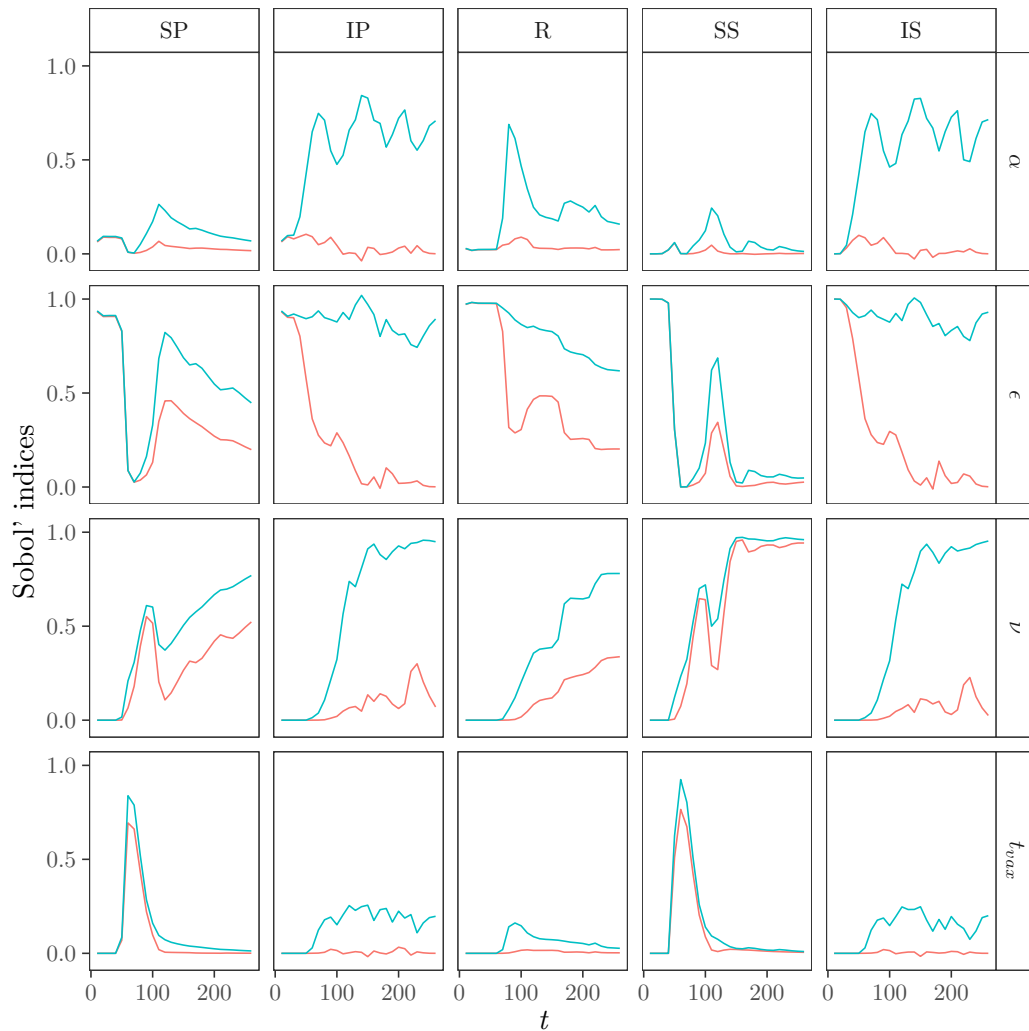
```
plot.ind <- list()
for(i in names(ind)) {
  plot.ind[[i]] <- ggplot(ind[[i]][sensitivity %in% c("Si", "Ti")],
    aes(time, original, fill = sensitivity,
        color = sensitivity, group = sensitivity)) +
    geom_line() +
    guides(linetype = FALSE, color = FALSE) +
    facet_grid(parameters ~ variable) +
    scale_y_continuous(breaks = scales::pretty_breaks(n = 3)) +
    labs(x = expression(italic(t)),
        y = "Sobol' indices") +
    theme_AP() +
    theme(legend.position = "top")
}
```

```
lapply(2:3, function(x) plot.ind[[x]])
```

```
## [[1]]
```



[[2]]



```
# EXPORT SOBOLE' INDICES COVID -----
```

```
covid.indices <- rbindlist(ind, idcol = "model")
fwrite(covid.indices, "ind.covid.csv")
```

```
# PLOT COVID KT -----
```

```
x.labels <- c("$S_P$", "$I_P$", "$R$", "$S_S$", "$I_S$")
```

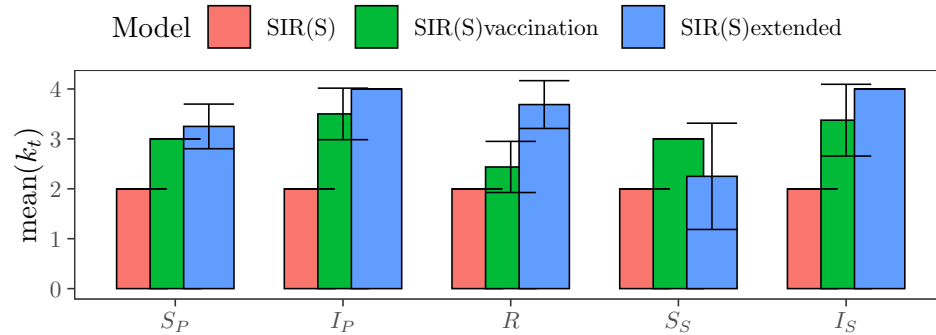
```
covid.kt <- covid.indices[sensitivity == "Ti" & original > 0.05,
                          length(unique(parameters)), .(model, time, variable)] %>%
  .[time > 100, .(mean = mean(V1), sd = sd(V1)), .(model, variable)] %>%
  .[, model:= factor(model, levels = type_model)] %>%
  ggplot(., aes(variable, mean, fill = model)) +
  geom_bar(stat = "identity", position = position_dodge(0.6), color = "black") +
  geom_errorbar(aes(ymin = mean - sd, ymax = mean + sd),
               position = position_dodge(0.6)) +
  scale_fill_discrete(name = "Model") +
  scale_x_discrete(labels = x.labels) +
```

```

theme_AP() +
labs(x = "", y = "mean($k_t$)") +
theme(legend.position = "none")

legend.covid.kt <- get_legend(covid.kt + theme(legend.position = "top"))
plot.covid.kt <- plot_grid(legend.covid.kt, covid.kt, ncol = 1, rel_heights = c(0.15, 0.85))
plot.covid.kt

```



```

# PLOT COVID CV AND COVID KT -----

covid.cv.kt <- plot_grid(cv.covid, plot.covid.kt, labels = "auto", ncol = 1)

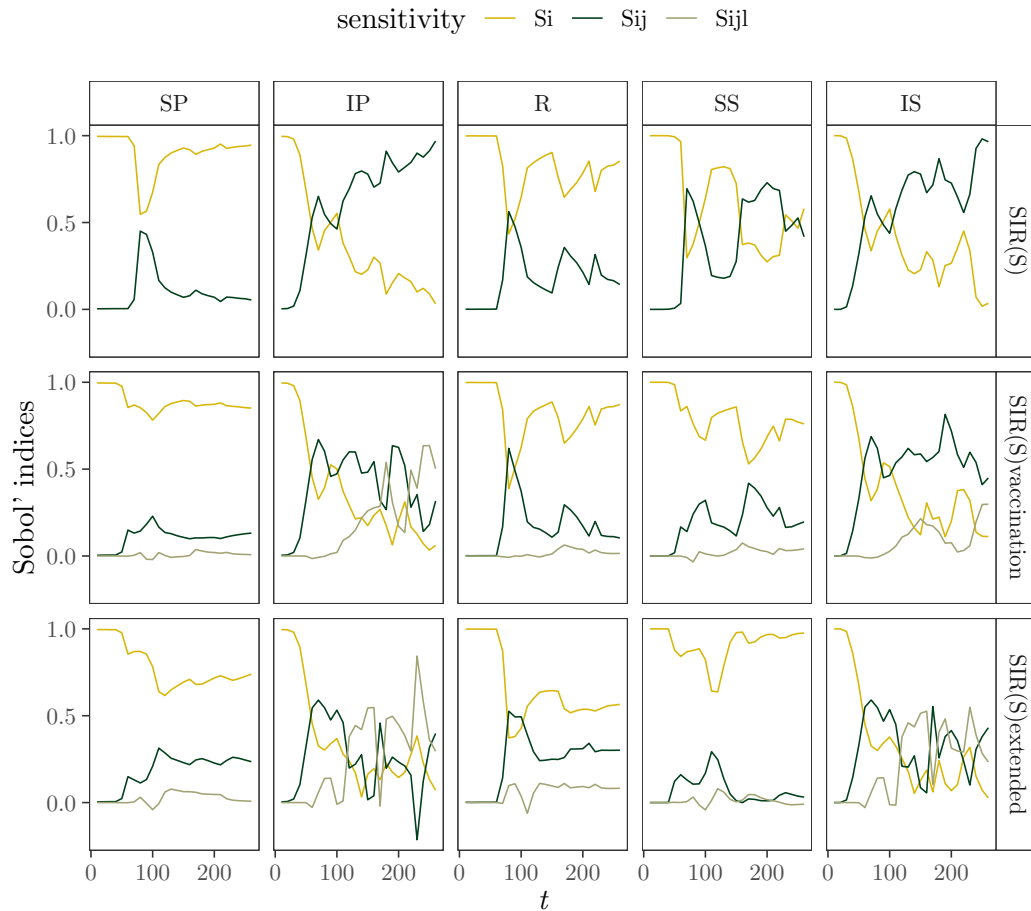
```

```

# COVID SUM FIRST, SECOND AND THIRD -ORDER -----

covid.indices %>%
  .[!sensitivity == "Ti"] %>%
  .[, sum(original), .(sensitivity, time, variable, model)] %>%
  .[, model:= factor(model, levels = type_model)] %>%
  ggplot(., aes(time, V1, color = sensitivity, group = sensitivity)) +
  geom_line() +
  facet_grid(model ~ variable) +
  scale_y_continuous(breaks = scales::pretty_breaks(n = 3)) +
  scale_color_manual(values = wes_palette("Cavalcanti")) +
  labs(x = expression(italic(t)),
       y = "Sobol' indices") +
  theme_AP() +
  theme(legend.position = "top")

```



PLOT SUM OF SI, SIJ AND SIJL AT T = 200 -----

```
time.selected <- 200
```

```
covid.ks <-
  rbindlist(ind, idcol = "model") %>%
  .[time == time.selected] %>%
  .[!sensitivity == "Ti"] %>%
  .[, sum(original), .(model, variable, sensitivity)] %>%
  .[, model:= factor(model, levels = type_model)] %>%
  ggplot(., aes(variable, V1, fill = sensitivity)) +
  geom_col(position = position_stack(reverse = TRUE), color = "black") +
  labs(x = "", y = "$\\sum S_i + \\sum S_{i,j} + \\sum S_{i,j,l}$") +
  geom_hline(yintercept = 0.99, lty = 2) +
  scale_fill_manual(name = "",
    labels = c("$S_i$", "$S_{i,j}$", "$S_{i,j,l}$"),
    values = wes_palette("Cavalcanti1")) +
  annotation_custom(textGrob("p", gp = gpar(col = "red")),
    xmin = 1, xmax = 1, ymin = 0.98, ymax = 0.98) +
  facet_wrap(~model) +
  scale_x_discrete(labels = c("SP" = "$S_P$",
    "IP" = "$I_P$",
```



```

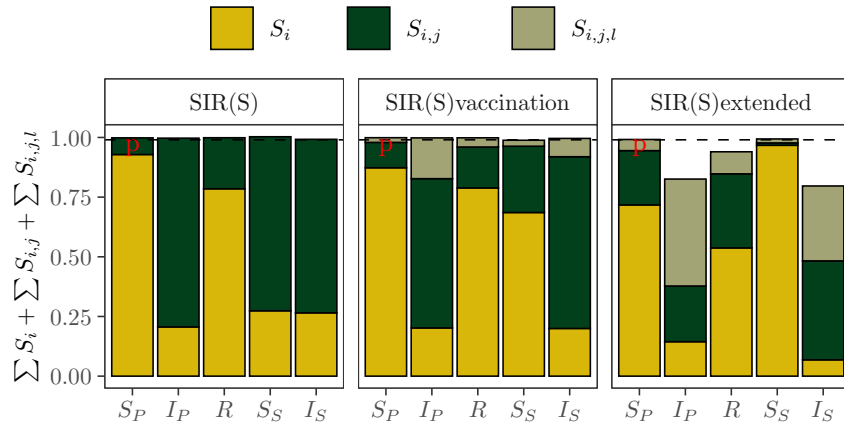
      "R" = "$R$",
      "SS" = "$S_S$",
      "IS" = "$I_S$")) +
  theme_AP() +
  theme(legend.position = "none",
        axis.title.y = element_text(size = 9))

legend.covid.ks <- get_legend(covid.ks + theme(legend.position = "top"))

covid.ks <- plot_grid(legend.covid.ks, covid.ks, ncol = 1, rel_heights = c(0.15, 0.85))

covid.ks

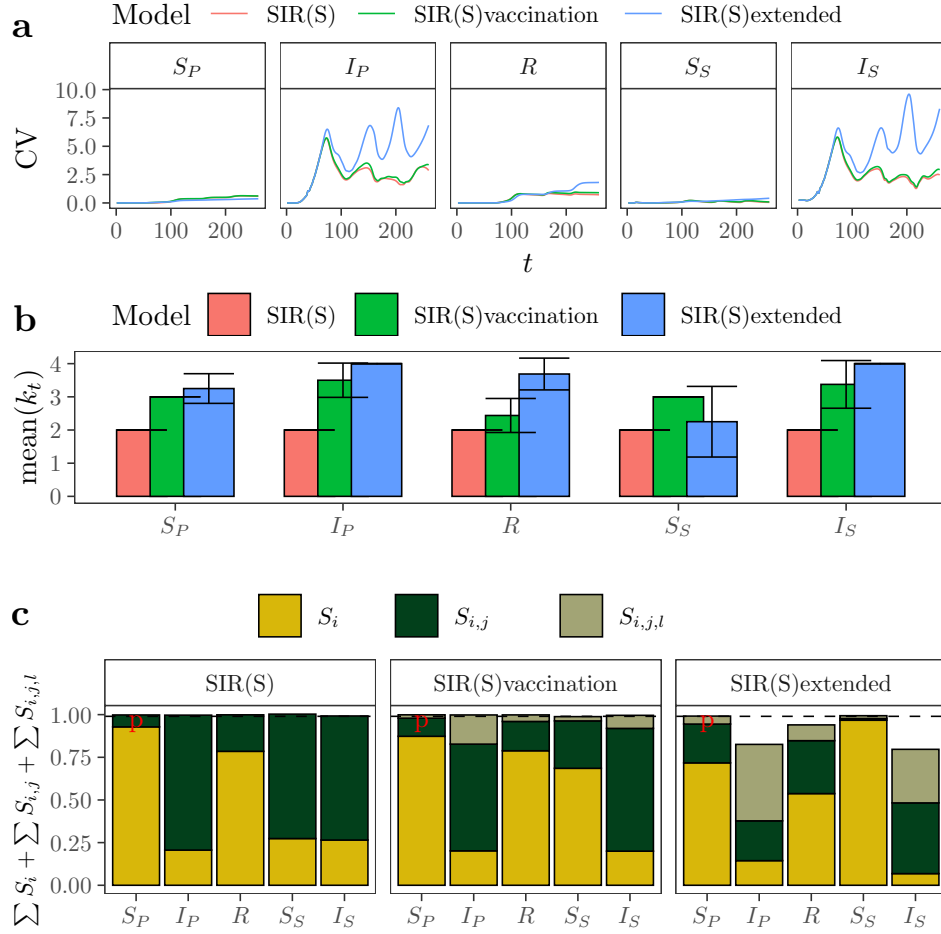
```



```

plot_grid(covid.cv.kt, covid.ks, ncol = 1,
          labels = c("", "c"), rel_heights = c(0.6, 0.4))

```



4 A model on irrigation water withdrawals

```
# DEFINE THE MODELS -----

run_model <- function(trigger, output, mat) {

  # Priestley Taylor
  if(trigger == "priestley") {
    et0 <- mat[, "alpha"] * ((mat[, "Delta"] * mat[, "A"]) / (mat[, "gamma"] + mat[, "Delta"]))

    # Penman Monteith
  } else if(trigger == "penman") {
    et0 <- (0.408 * mat[, "Delta"] * mat[, "A"] +
            mat[, "gamma"] * (900 / (mat[, "T_a"] + 273)) * mat[, "w"] * mat[, "v"]) /
            (mat[, "Delta"] + mat[, "gamma"] * (1 + 0.34 * mat[, "w"]))
  }

  if(output == "et0") {
    final <- et0
  }
}
```

```

} else if(output == "etc") {
  final <- mat[, "k_c"] * et0

} else if(output == "water") {
  final <- (mat[, "I_a"] * (mat[, "k_c"] * et0 - mat[, "P"])) /
    (mat[, "E_a"] * mat[, "E_c"] * mat[, "M_f"])
  final <- final / 10^3 # Output is in m3 ha
}

return(final)
}

# Model to run rowwise
run_model_rowwise <- function(I_a, Delta, A, gamma, T_a, w, v, output,
                              k_c, P, E_a, E_c, M_f, alpha, X1) {

  # Priestley Taylor
  if(X1 == 1) {
    et0 <- alpha * ((Delta * A) / (gamma + Delta))

    # Penman Monteith
  } else if(X1 == 2) {
    et0 <- (0.408 * Delta * A + gamma * (900 / (T_a + 273)) * w * v) /
      Delta + gamma * (1 + 0.34 * w)

  } else {
    stop("trigger should be either 1 or 2")
  }

  crop.reference <- k_c * et0
  water.withdrawal <- (I_a * (crop.reference - P)) / (E_a * E_c * M_f)

  if(output == "et0") {
    final <- et0
  } else if(output == "etc") {
    final <- crop.reference
  } else if(output == "water") {
    final <- water.withdrawal / 10^3 # Output is in m3 ha
  }
  return(final)
}

# FUNCTION TO CREATE MATRICES -----
create_matrices <- function(trigger, output) {

```

```

if(trigger == "priestley") {
  params <- c("alpha", "Delta", "A", "gamma")

} else if(trigger == "penman") {
  params <- c("Delta", "A", "gamma", "T_a", "w", "v")

} else if(trigger == "all") {
  params <- c("Delta", "A", "gamma", "T_a", "w", "v", "alpha", "X1")
}

if(output == "et0") {
  params <- params

} else if(output == "etc") {
  params <- c(params, "k_c")

} else if(output == "water") {
  params <- c(params, "k_c", "I_a", "E_a", "E_c", "M_f", "P")
}

mat <- sobol_matrices(N = sample.size, params = params, order = order,
                     matrices = matrices, scrambling = 1)
return(mat)
}

# FUNCTION TO TRANSFORM MATRICES -----

Delta <- 0.08725467
Gamma <- 0.06658597
T_air <- 11
v <- 0.4115359

transform_matrices <- function(trigger, mat, output) {

  if(trigger == "priestley") {
    mat[, "alpha"] <- qunif(mat[, "alpha"], 1.13, 1.38)
    mat[, "Delta"] <- qunif(mat[, "Delta"], Delta + Delta * -0.005, Delta + Delta * 0.005)
    mat[, "gamma"] <- qunif(mat[, "gamma"], Gamma + Gamma * -0.001, Gamma + Gamma * 0.001)
    mat[, "A"] <- qunif(mat[, "A"], 350 + 350 * -0.15, 350 + 350 * 0.15)

  } else if(trigger == "penman") {
    mat[, "Delta"] <- qunif(mat[, "Delta"], Delta + Delta * -0.005, Delta + Delta * 0.005)
    mat[, "gamma"] <- qunif(mat[, "gamma"], Gamma + Gamma * -0.001, Gamma + Gamma * 0.001)
    mat[, "A"] <- qunif(mat[, "A"], 350 + 350 * -0.15, 350 + 350 * 0.15)
    mat[, "T_a"] <- qunif(mat[, "T_a"], T_air + T_air * -0.01, T_air + T_air * 0.01)
    mat[, "w"] <- qunif(mat[, "w"], 2.81 + 2.81 * -0.05, 2.81 + 2.81 * 0.05)
  }
}

```

```

    mat[, "v"] <- qunif(mat[, "v"], v + v * -0.04, v + v * 0.04)

  } else if(trigger == "all") {
    mat[, "Delta"] <- qunif(mat[, "Delta"], Delta + Delta * -0.005, Delta + Delta * 0.005)
    mat[, "gamma"] <- qunif(mat[, "gamma"], Gamma + Gamma * -0.001, Gamma + Gamma * 0.001)
    mat[, "A"] <- qunif(mat[, "A"], 350 + 350 * -0.15, 350 + 350 * 0.15)
    mat[, "T_a"] <- qunif(mat[, "T_a"], T_air + T_air * -0.01, T_air + T_air * 0.01)
    mat[, "w"] <- qunif(mat[, "w"], 2.81 + 2.81 * -0.05, 2.81 + 2.81 * 0.05)
    mat[, "v"] <- qunif(mat[, "v"], v + v * -0.04, v + v * 0.04)
    mat[, "alpha"] <- qunif(mat[, "alpha"], 1.13, 1.38)
    mat[, "X1"] <- floor(mat[, "X1"] * (2 - 1 + 1)) + 1

  }

  if(output == "et0") {
    mat <- mat

  } else if(output == "etc") {
    mat[, "k_c"] <- qunif(mat[, "k_c"], 0.4564315, 1.144222)

  } else if(output == "water") {
    mat[, "k_c"] <- qunif(mat[, "k_c"], 0.4564315, 1.144222)
    mat[, "I_a"] <- qunif(mat[, "I_a"], 42.932, 144.5515)
    mat[, "E_a"] <- qunif(mat[, "E_a"], 0.49, 0.88)
    mat[, "E_c"] <- qunif(mat[, "E_c"], 0.64, 0.96)
    mat[, "M_f"] <- qunif(mat[, "M_f"], 0.5, 0.97)
    mat[, "P"] <- qunif(mat[, "P"], 0, 0.1)

  }
  return(mat)
}

# MERGE ALL FUNCTIONS -----

full_model <- function(trigger, output) {
  mat <- create_matrices(trigger = trigger, output = output)
  mat <- transform_matrices(trigger = trigger, mat = mat, output = output)
  out <- run_model(trigger = trigger, output = output, mat = mat)
  return(out)
}

# DEFINE SETTINGS -----

sample.size <- 2^12
order <- "third"
matrices <- c("A", "B", "AB")
first <- "saltelli"

```

```

total <- "jansen"
R <- 10^3

# RUN MODELS -----

et0.formulae <- c("priestley", "penman")
outputs <- c("et0", "etc", "water")

# Define parallel computing
n_cores <- makeCluster(floor(detectCores() * 0.75))
registerDoParallel(n_cores)

y <- foreach(j = et0.formulae) %:%
  foreach(l = outputs,
    .packages = "sensobol") %dopar%
  {
    full_model(trigger = j, output = l)
  }

# Stop parallel computing
stopCluster(n_cores)

# RUN MODELS (all) -----

trigger <- "all"
output <- "water"

mat <- create_matrices(trigger = trigger, output = output)
mat <- data.table(transform_matrices(trigger = trigger, mat = mat, output = output))

# Define parallel computing
n_cores <- makeCluster(floor(detectCores() * 0.75))
registerDoParallel(n_cores)

y.all <- foreach(j = c("et0", "etc", "water")) %:%
  foreach(i = 1:nrow(mat), .combine = "rbind") %dopar%
  {
    run_model_rowwise(I_a = mat[i, "I_a"],
      Delta = mat[i, "Delta"],
      A = mat[i, "A"],
      gamma = mat[i, "gamma"],
      T_a = mat[i, "T_a"],
      w = mat[i, "w"],
      v = mat[i, "v"],
      k_c = mat[i, "k_c"],
      P = mat[i, "P"],
      E_a = mat[i, "E_a"],
      E_c = mat[i, "E_c"],
      alpha = mat[i, "alpha"],

```

```

        M_f = mat[i, "M_f"],
        X1 = mat[i, "X1"],
        output = j)
    }

# Stop parallel computing
stopCluster(n_cores)

# ARRANGE RESULTS -----

for(i in 1:2) {
  names(y[[i]]) <- outputs
}

names(y) <- et0.formulae
names(y.all) <- outputs

# CREATION OF THE A MATRIX -----

# First round of simulations
A <- list()
for(i in names(y)) {
  for(j in names(y[[i]])) {
    A[[i]][[j]] <- data.table(y[[i]][[j]][1:(2 * sample.size)])
  }
}

A <- lapply(A, function(x) rbindlist(x, idcol = "output")) %>%
  rbindlist(., idcol = "et0.formulae")

# Second round of simulations (all)
out.all <- lapply(y.all, function(x) data.table(x)) %>%
  rbindlist(., idcol = "output") %>%
  .[, et0.formulae:= "all"]

out.all <- setcolorder(out.all, c("et0.formulae", "output", "V1"))

A.all <- out.all[, .SD[1:(sample.size * 2)], output]

A.water <- rbind(A, A.all)

# SENSITIVITY ANALYSIS -----

ind <- list()

for(i in names(y)) {
  for(j in names(y[[i]])) {
    if(i == "priestley") {

```

```

    params <- c("alpha", "Delta", "A", "gamma")
  } else if(i == "penman") {
    params <- c("Delta", "A", "gamma", "T_a", "w", "v")
  }
  if(j == "et0") {
    params <- params
  } else if(j == "etc") {
    params <- c(params, "k_c")
  } else if(j == "water") {
    params <- c(params, "k_c", "I_a", "E_a", "E_c", "M_f", "P")
  }
  ind[[i]][[j]] <- sobol_indices(matrices = matrices, Y = y[[i]][[j]],
                                N = sample.size, params = params, order = order,
                                first = first, total = total,
                                R = R, boot = TRUE, parallel = "multicore")$results
}
}

ind.all <- out.all[, sobol_indices(matrices = matrices,
                                   Y = V1, params = colnames(mat), N = sample.size,
                                   order = order, R = R, boot = TRUE, parallel = "multicore",
                                   first = first, total = total)$results, .(et0.formulae, output)]

# ARRANGE SENSITIVITY DATA -----

ind.dt <- lapply(ind, function(x) rbindlist(x, idcol = "output")) %>%
  rbindlist(., idcol = "et0.formulae")

# merge
all.ind <- rbind(ind.dt, ind.all)[, original:= ifelse(original < 0, 0, original)]

# CREATE PLOTS -----

water.kt <- all.ind[sensitivity == "Ti" & original > 0.05] %>%
  .[, .(kt = length(unique(parameters))), .(et0.formulae, output)]

water.kt

##      et0.formulae output kt
## 1:      priestley    et0  2
## 2:      priestley    etc  2
## 3:      priestley  water  5
## 4:         penman    et0  1
## 5:         penman    etc  2
## 6:         penman  water  5
## 7:             all    et0  2
## 8:             all    etc  3
## 9:             all  water  6

```



```

water.cv <- A.water[, .(cv = sd(V1) / mean(V1)), .(et0.formulae, output)]

# KT and CV plot
# KT and CV plot
a <- merge(water.kt, water.cv, by = c("et0.formulae", "output")) %>%
  .[, et0.formulae:= factor(et0.formulae, levels = c("penman", "priestley", "all"))] %>%
  ggplot(. , aes(output, cv, color = et0.formulae, group = et0.formulae)) +
  geom_line() +
  geom_point(aes(size = kt)) +
  scale_color_manual(name = "$ET_0$ formula",
                     values = wes_palette("Zissou1"),
                     labels = c("Penman-Monteith",
                                "Priestley-Taylor",
                                "Uncertain")) +
  scale_size_continuous(name = "$k_t$",
                        breaks = c(1, 3, 6)) +
  scale_x_discrete(labels = c("Reference \n evapotranspiration",
                              "Crop \n evapotranspiration",
                              "Irrigation \n water withdrawal"),
                   guide = guide_axis(n.dodge = 2)) +
  labs(x = "", y = "CV") +
  theme_AP()

# ks plots

facet_labels <- c(
  "penman" = "Penman-Monteith",
  "priestley" = "Priestley-Taylor",
  "all" = "Uncertain"
)

b <- all.ind[!sensitivity == "Ti", sum(original), .(et0.formulae, output, sensitivity)] %>%
  .[, et0.formulae:= factor(et0.formulae, levels = c("penman", "priestley", "all"))] %>%
  ggplot(. , aes(output, V1, fill = sensitivity)) +
  geom_col(position = position_stack(reverse = TRUE), color = "black") +
  scale_fill_manual(name = "",
                    labels = c("$S_i$", "$S_{i,j}$", "$S_{i,j,l}$"),
                    values = wes_palette("Cavalcanti1")) +
  scale_x_discrete(labels = c("Reference \n evapotranspiration",
                              "Crop \n evapotranspiration",
                              "Irrigation \n water withdrawal"),
                   guide = guide_axis(n.dodge = 2)) +
  scale_y_continuous(breaks = pretty_breaks(n = 3)) +
  geom_hline(yintercept = 0.99, lty = 2) +
  labs(x = "", y = "$\\sum S_i + \\sum S_{i,j} + \\sum S_{i,j,l}$") +
  annotation_custom(textGrob("p", gp = gpar(col = "red")),
                    xmin = 0.3, xmax = 1, ymin = 0.98, ymax = 0.98) +

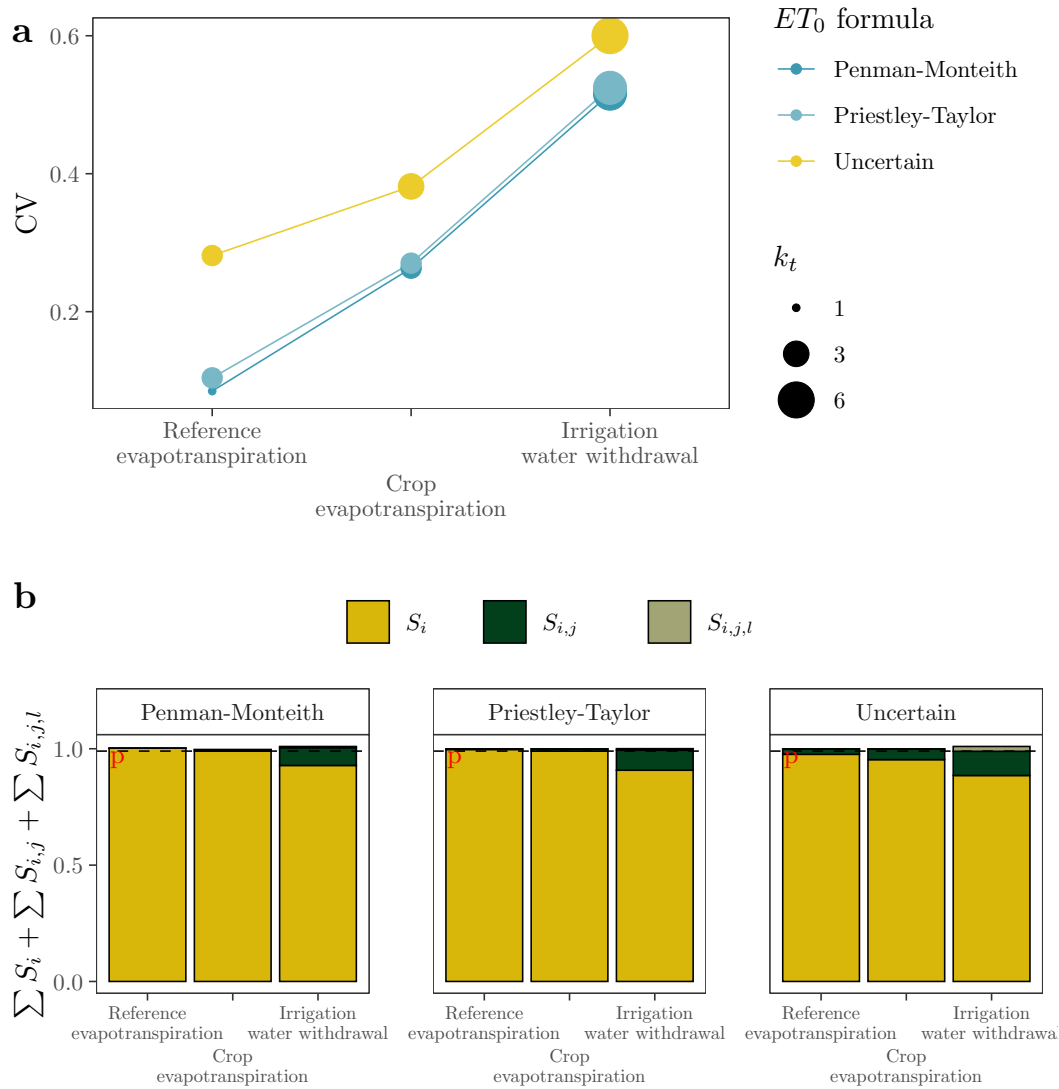
```

```

facet_wrap(~et0.formulae, labeller = as_labeller(facet_labels), ncol = 3) +
theme_AP() +
theme(legend.position = "top",
      panel.spacing = unit(2, "lines"),
      axis.text.x = element_text(size = 7))

plot_grid(a, b, ncol = 1, labels = "auto")

```



```

all.ind[!sensitivity == "Ti", round(sum(original), 3), .(et0.formulae, output, sensitivity)]

```

```

##      et0.formulae output sensitivity   V1
##  1:  priestley    et0          Si 0.997
##  2:  priestley    et0          Si,j 0.002
##  3:  priestley    et0          Si,j,l 0.000
##  4:  priestley    etc          Si 0.990
##  5:  priestley    etc          Si,j 0.010
##  6:  priestley    etc          Si,j,l 0.000

```

```
## 7: priestley water Si 0.908
## 8: priestley water Sij 0.086
## 9: priestley water Sijl 0.007
## 10: penman et0 Si 1.002
## 11: penman et0 Sij 0.000
## 12: penman et0 Sijl 0.000
## 13: penman etc Si 0.989
## 14: penman etc Sij 0.007
## 15: penman etc Sijl 0.000
## 16: penman water Si 0.928
## 17: penman water Sij 0.076
## 18: penman water Sijl 0.005
## 19: all et0 Si 0.976
## 20: all et0 Sij 0.023
## 21: all et0 Sijl 0.000
## 22: all etc Si 0.953
## 23: all etc Sij 0.045
## 24: all etc Sijl 0.001
## 25: all water Si 0.885
## 26: all water Sij 0.104
## 27: all water Sijl 0.021
## et0.formulae output sensitivity V1
```

```
# EXPORT WATER SENSITIVITY INDICES -----
```

```
fwrite(all.ind, "ind.water.csv")
```

5 The Sobol' G function

```
# DEFINE MODELS -----
```

```
# Analytical indices of G function -----
```

```
g_analytical <- function(a) {
  # Si
  Vi <- (1 / 3) / (1 + a)^2
  V <- prod((1 + Vi)) - 1
  Si.analytical <- Vi / V

  # Ti
  Vt <- vector()
  for(i in 1:length(Vi)) {
    Vt[i] <- Vi[i] * (prod(1 + Vi[-i]))
  }
  Ti.analytical <- Vt / V
  out <- c(Si.analytical, Ti.analytical)
  return(out)
}
```

```

# G Function -----
g_fun <- function(X, a, epsilon) {
  set.seed(epsilon)
  a <- sample(a, size = ncol(X), prob = prob, replace = TRUE)
  y <- 1
  for (j in 1:ncol(X)) {
    y <- y * (abs(4 * X[, j] - 2) + a[j]) / (1 + a[j])
  }
  analytical <- g_analytical(a)
  output <- list(y, analytical)
  names(output) <- c("output", "analytical")
  return(output)
}

# Model with G function
model_g <- function(N, k, epsilon, order) {
  params <- paste("X", 1:k, sep = "")
  mat <- sobol_matrices(N = N, params = params, order = order, scrambling = 1)
  y <- g_fun(X = mat, a = a, epsilon = epsilon)
  ind <- sobol_indices(Y = y$output, N = N, params = params, order = order)
  mae <- ind$results[sensitivity %in% c("Si", "Ti")] [
    , abs(mean(original - y$analytical)), sensitivity][, V1]
  cv <- sd(y$output) / mean(y$output)
  sum.si <- ind$si.sum
  k.t <- ind$results[sensitivity == "Ti" & original > 0.05, length(unique(parameters))]
  Sij <- ind$results[sensitivity == "Sij", sum(original)]
  Sijl <- ind$results[sensitivity == "Sijl", sum(original)]
  return(c(cv, sum.si, k.t, Sij, Sijl, mae))
}

# DEFINE SETTINGS -----

# Settings
params <- c("k", "epsilon")
N <- 2^10
N.internal <- 2^12
order <- "third"
matrices <- "A"
a <- c(0, 1, 4.5, 9, 99)
prob <- c(0.4, 0.3, 0.2, 0.05, 0.05)

# Creation of sample matrix
mat <- sobol_matrices(N = N, params = params, matrices = matrices, scrambling = 1)

# maximum number of explored inputs
max.k <- 20

```

```

# Transformation to appropriate distributions
mat[, "k"] <- floor(mat[, "k"] * (max.k - 3 + 1) + 3)
mat[, "epsilon"] <- floor(mat[, "epsilon"] * (N - 1 + 1) + 1)

# RUN SOBOL' G MODEL -----

# Define parallel computing
n_cores <- detectCores() * 0.75
cl <- makeCluster(n_cores)
registerDoParallel(cl)

# Run model
Y <- foreach(i=1:nrow(mat), .packages = "sensobol",
             .combine = "rbind") %dopar%
{
  model_g(N = N.internal,
          k = mat[[i, "k"]],
          epsilon = mat[[i, "epsilon"]],
          order = order)
}

# Stop parallel cluster
stopCluster(cl)

# ARRANGE SOBOL' G DATA -----

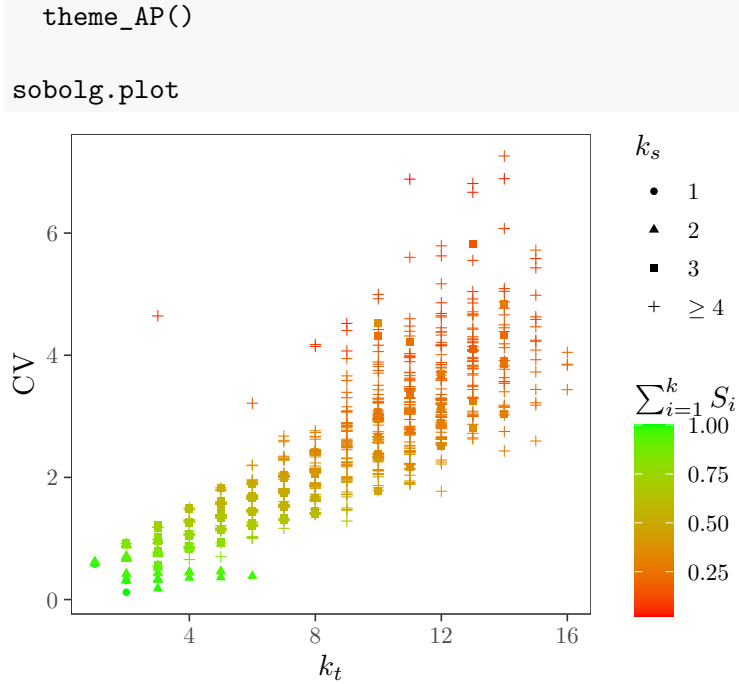
dt <- data.table(Y) %>%
  setnames(., paste("V", 1:7, sep = ""),
           c("CV", "sum.si", "k.t", "sij", "sijl", "mae.si", "mae.ti")) %>%
  cbind(mat, .) %>%
  .[, k.s:= ifelse(sum.si >= 0.99, 1,
                  ifelse(sum.si + sij >= 0.99, 2,
                        ifelse((sum.si + sij + sijl) >= 0.99, 3, 4)))] %>%
  .[, k.s:= factor(k.s)] %>%
  .[, up.second:= sum.si + sij] %>%
  .[, up.third:= sum.si + sij + sijl]

fwrite(dt, "dt.sobol.csv")

# PLOT -----

sobolg.plot <- ggplot(dt, aes(k.t, CV, color = sum.si, shape = k.s)) +
  geom_point(size = 1) +
  labs(x = "$k_t$", y = "CV") +
  scale_color_gradient(low = "red", high = "green",
                      name = "$\\sum_{i=1}^k S_i$") +
  scale_shape_discrete(name = "$k_s$",
                      labels = c("1", "2", "3", "$\\geq 4$")) +

```



6 The metafunction

```
# DEFINE SETTINGS -----
```

```
# Settings
params <- c("k", "epsilon", "n")
N <- 2^11
matrices <- "A"
```

```
# Creation of sample matrix
mat <- sobol_matrices(N = N, params = params, matrices = matrices, scrambling = 1)
```

```
# maximum number of explored inputs
max.k <- 15
```

```
# Transformation to appropriate distributions
mat[, "k"] <- floor(mat[, "k"] * (max.k - 2 + 1) + 2)
mat[, "epsilon"] <- floor(mat[, "epsilon"] * (N - 1 + 1) + 1)
set.seed(666)
mat[, "n"] <- sapply(mat[, "k"], function(x) sample(2:x, 1))
mat[, "n"] <- ifelse(mat[, "n"] == 1, 2, mat[, "n"]) # Correct and force 1 to be 2
```

```
# PLOT METAFUNCTION -----
```

```
function_list <- list(
  Linear = function(x) x,
  Quadratic = function(x) x ^ 2,
```

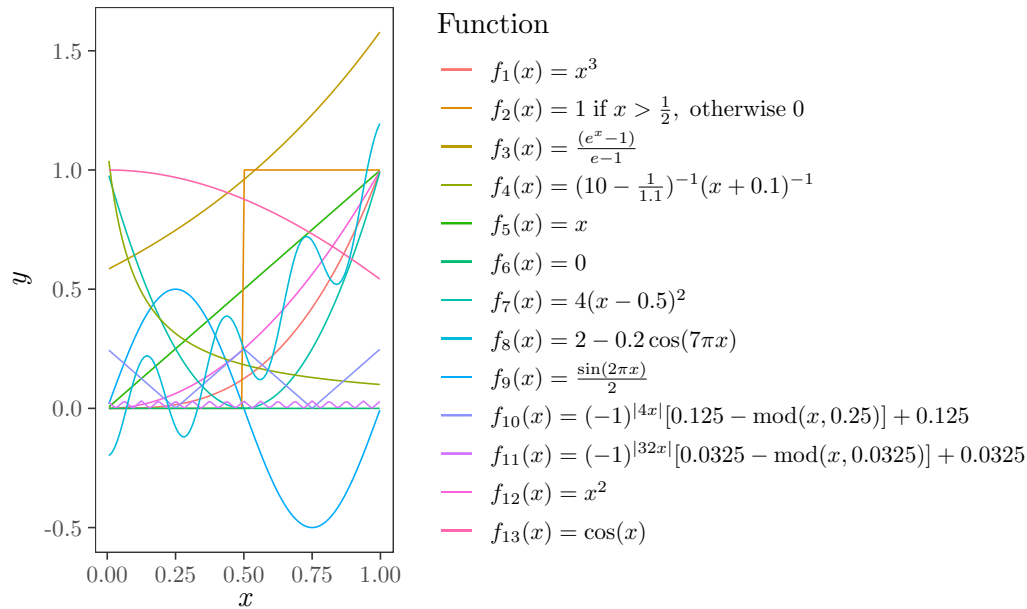
```

Cubic = function(x) x ^ 3,
Exponential = function(x) exp(1) ^ x / (exp(1) - 1),
Periodic = function(x) sin(2 * pi * x) / 2,
Discontinuous = function(x) ifelse(x > 0.5, 1, 0),
Non.monotonic = function(x) 4 * (x - 0.5) ^ 2,
Inverse = function(x) (10 - 1 / 1.1) ^ -1 * (x + 0.1) ^ - 1,
No.effect = function(x) x * 0,
Trigonometric = function(x) cos(x),
Piecewise.large = function(x) ((-1) ^ as.integer(4 * x) *
                                (0.125 - (x %% 0.25)) + 0.125),
Piecewise.small = function(x) ((-1) ^ as.integer(32 * x) *
                                (0.03125 - 2 * (x %% 0.03125)) + 0.03125) / 2,
Oscillation = function(x) x ^ 2 - 0.2 * cos(7 * pi * x)
)

plot.metafunction <- ggplot(data.frame(x = runif(100)), aes(x)) +
  map(1:length(function_list), function(nn) {
    stat_function(fun = function_list[[nn]],
                  geom = "line",
                  aes_(color = factor(names(function_list[nn]))))
  }) +
  labs(color= "Function", linetype = "Function",
       x = expression(italic(x)),
       y = expression(italic(y))) +
  scale_color_discrete(labels = c("$f_1(x) = x^3$",
                                   "$f_2(x) = 1 \\hspace{1mm} \\mbox{if} \\hspace{1mm} x > \\frac{1}{2}$",
                                   "$f_3(x) = \\frac{(e^x - 1)}{e-1}$",
                                   "$f_4(x) = (10-\\frac{1}{1.1})^{-1}(x + 0.1)^{-1}$",
                                   "$f_5(x) = x$",
                                   "$f_6(x) = 0$",
                                   "$f_7(x) = 4(x - 0.5)^2$",
                                   "$f_8(x) = 2 - 0.2 \\cos(7 \\pi x)$",
                                   "$f_9(x) = \\frac{\\sin(2 \\pi x)}{2}$",
                                   "$f_{10}(x) = (-1)^{|4x|} [0.125- \\mbox{mod}(x, 0.25)] + 0.125$",
                                   "$f_{11}(x) = (-1)^{|32x|} [0.0325-\\mbox{mod}(x, 0.0325)] + 0.0325$",
                                   "$f_{12}(x) = x^2$",
                                   "$f_{13}(x) = \\cos(x)$")) +
  theme_AP() +
  theme(legend.text.align = 0)

plot.metafunction

```



```
# DEFINE MODEL -----

# Define metafunction
meta_fun <- function(data, epsilon, n) {

  # Define list of functions included in metafunction
  function_list <- list(
    Linear = function(x) x,
    Quadratic = function(x) x ^ 2,
    Cubic = function(x) x ^ 3,
    Exponential = function(x) exp(1) ^ x / (exp(1) - 1),
    Periodic = function(x) sin(2 * pi * x) / 2,
    Discontinuous = function(x) ifelse(x > 0.5, 1, 0),
    Non.monotonic = function(x) 4 * (x - 0.5) ^ 2,
    Inverse = function(x) (10 - 1 / 1.1) ^ -1 * (x + 0.1) ^ - 1,
    No.effect = function(x) x * 0,
    Trigonometric = function(x) cos(x),
    Piecewise.large = function(x) ((-1) ^ as.integer(4 * x) * (0.125 - (x %% 0.25)) + 0.125),
    Piecewise.small = function(x) ((-1) ^ as.integer(32 * x) * (0.03125 - 2 * (x %% 0.03125)) - 0.03125),
    Oscillation = function(x) x ^ 2 - 0.2 * cos(7 * pi * x)
  )

  # Sample list of functions
  set.seed(epsilon)
  all_functions <- sample(names(function_list), ncol(data), replace = TRUE)

  # Compute model output first order effects
  mat.y <- sapply(seq_along(all_functions), function(x)
    function_list[[all_functions[x]]](data[, x]))
}
```



```

# Compute first-order effects
y1 <- Rfast::rowsums(mat.y)

# Define matrix with all possible interactions up to the n-th order
interactions <- lapply(2:n, function(x) RcppAlgos::comboGeneral(1:n, x, nThreads = 4))

out <- lapply(1:length(interactions), function(x) {
  lapply(1:nrow(interactions[[x]]), function(y) {
    Rfast::rowprods(mat.y[, interactions[[x]][y, ]])
  })
})

y2 <- lapply(out, function(x) do.call(cbind, x)) %>%
  do.call(cbind, .) %>%
  Rfast::rowsums(.)

y <- y1 + y2

return(y)
}

# Merge metafunction with model
model <- function(data, epsilon, n) {

  k <- ncol(data)

  if (n > k) {
    stop("level_interactions should be smaller or equal than \n
         the number of parameters")
  }
  y <- meta_fun(data = data, epsilon = epsilon, n = n)

  return(y)
}

# Define final model
model_fun <- function(k, epsilon, N, n) {
  params <- paste("X", 1:k, sep = "")
  mat <- sobol_matrices(N = N, params = params,
                       matrices = "A", scrambling = 1)
  y <- model(data = mat, epsilon = epsilon, n = n)
  cv <- sd(y) / mean(y)
  return(cv)
}

```

```

# RUN MODEL -----

# Define parallel computing
n_cores <- detectCores() * 0.75
cl <- makeCluster(n_cores)
registerDoParallel(cl)

N.internal <- 2^9
high.order <- c("n", "k")

# Compute
Y <- foreach(j = high.order) %:%
  foreach(i=1:nrow(mat), .packages = c("Rfast", "sensobol", "dplyr", "RcppAlgos"),
    .combine = "rbind") %dopar%
  {
    model_fun(k = mat[[i, "k"]],
              epsilon = mat[[i, "epsilon"]],
              n = mat[[i, j]],
              N = N.internal)
  }

# Run by fixing the model dimensionality at 15
Y.k <- foreach(i=1:nrow(mat), .packages = c("Rfast", "sensobol", "dplyr", "RcppAlgos"),
  .combine = "rbind") %dopar%
  {
    model_fun(k = max.k,
              epsilon = mat[[i, "epsilon"]],
              n = mat[[i, "n"]],
              N = N.internal)
  }

# Stop parallel cluster
stopCluster(cl)

# ARRANGE MODEL OUTPUT -----

names(Y) <- high.order

dt <- lapply(Y, data.table) %>%
  lapply(., function(x) cbind(mat, x)) %>%
  rbindlist(., idcol = "high.order") %>%
  .[, n:= ifelse(high.order == "n", n, k)] %>%
  setnames(., "V1", "CV") %>%
  setnames(., "n", "order.interactions") %>%
  .[, ID:= 1:.N]

```

```
dt.k <- cbind(mat, data.table(Y.k)) %>%
  setnames(., "V1", "CV")
```

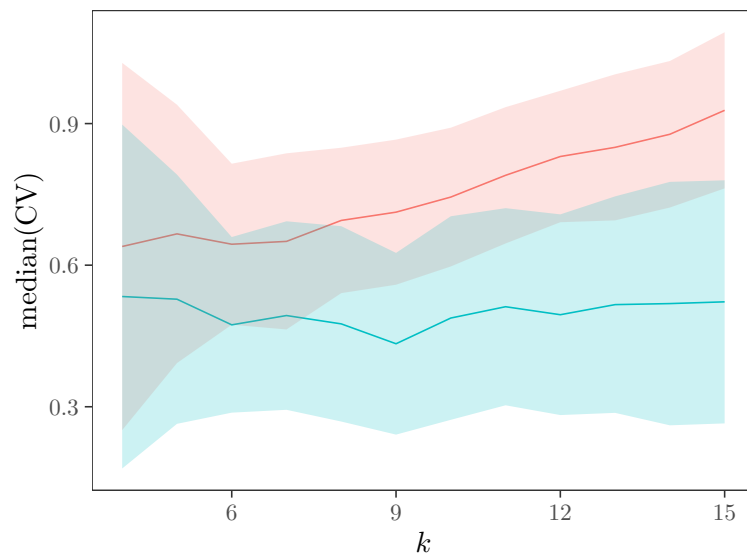
```
fwrite(dt, "dt.csv")
fwrite(dt.k, "dt.k.csv")
```

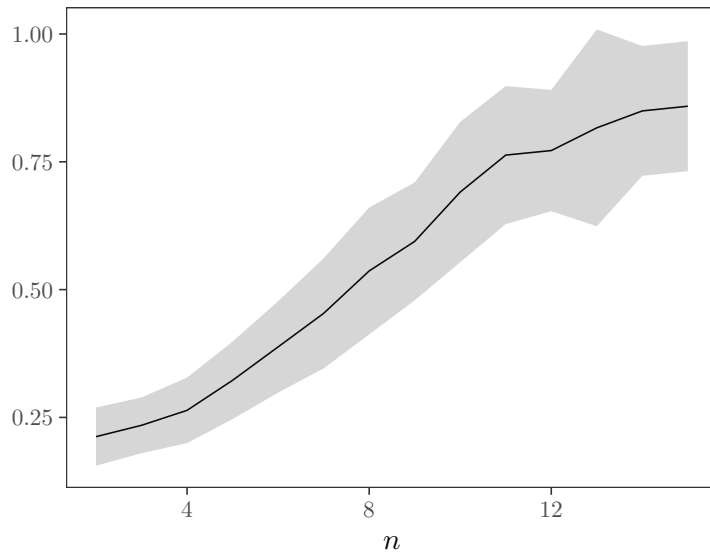
PLOTS -----

```
a <- dt[k > 3, .(median = median(CV, na.rm = TRUE), sd = sd(CV, na.rm = TRUE)), .(k, high.order)]
ggplot(., aes(k, median, fill = high.order)) +
  geom_line(aes(color = high.order)) +
  geom_ribbon(aes(y = median, ymin = median - sd, ymax = median + sd), alpha = 0.2) +
  labs(x = "$k$", y = "median(CV)") +
  scale_color_discrete(name = "Interactions",
    labels = c("Up to the $k$-th order",
      "Up to the $n$-th order for $k=15$")) +
  scale_fill_discrete(name = "Interactions",
    labels = c("Up to the $k$-th order",
      "Up to the $n$-th order for $k=15$")) +
  theme_AP() +
  theme(legend.position = "none")
```

```
b <- dt.k[, .(median = median(CV, na.rm = TRUE), sd = sd(CV, na.rm = TRUE)), n] %>%
  ggplot(., aes(n, median)) +
  geom_line() +
  geom_ribbon(aes(y = median, ymin = median - sd, ymax = median + sd), alpha = 0.2) +
  labs(x = "$n$", y = "") +
  theme_AP()
```

a



b

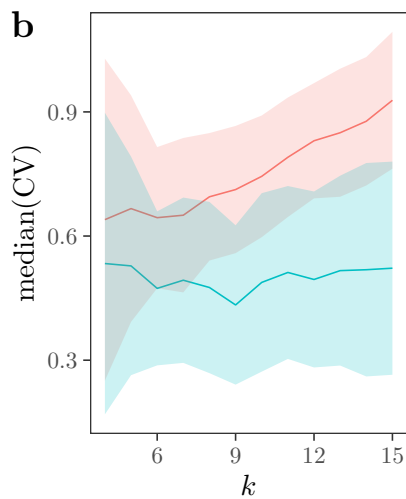
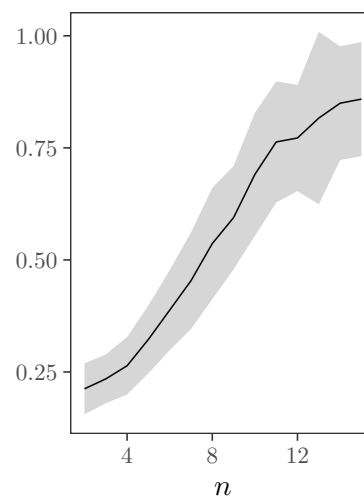
MERGE PLOTS -----

```

legend <- get_legend(a + theme(legend.position = "top"))
bottom <- plot_grid(a, b, ncol = 2, labels = c("b", "c"))
meta.plot <- plot_grid(legend, bottom, ncol = 1, rel_heights = c(0.1, 0.9))
meta.plot

```

teractions ■ Up to the k -th order ■ Up to the n -th order for $k = 15$

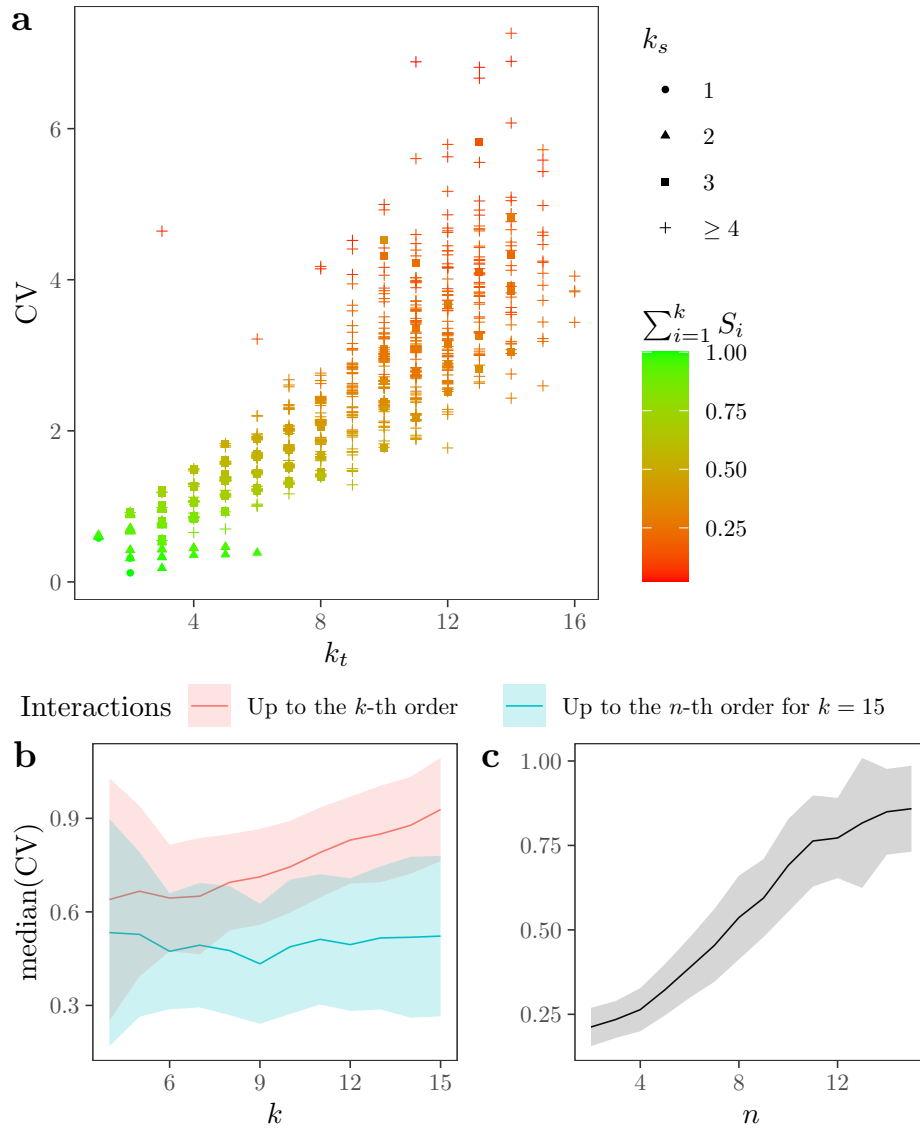
b**c**

MERGE WITH SOBOL' G PLOT -----

```

plot_grid(sobolg.plot, meta.plot, ncol = 1, labels = c("a", ""),
          rel_heights = c(0.6, 0.4))

```



```
plot_grid(plot.metafunction, meta.plot, rel_heights = c(0.65, 0.35),
          rel_widths = c(0.7, 0.3), ncol = 1)
```

tion

$$f_1(x) = x^3$$

$$f_2(x) = 1 \text{ if } x > \frac{1}{2}, \text{ otherwise } 0$$

$$f_3(x) = \frac{(e^x - 1)}{e - 1}$$

$$f_4(x) = (10 - \frac{1}{1.1})^{-1}(x + 0.1)^{-1}$$

$$f_5(x) = x$$

$$f_6(x) = 0$$

$$f_7(x) = 4(x - 0.5)^2$$

$$f_8(x) = 2 - 0.2 \cos(7\pi x)$$

$$f_9(x) = \frac{\sin(2\pi x)}{2}$$

$$f_{10}(x) = (-1)^{\lfloor 4x \rfloor} [0.125 - \text{mod}(x, 0.25)] + 0.125$$

$$f_{11}(x) = (-1)^{\lfloor 32x \rfloor} [0.0325 - \text{mod}(x, 0.0325)] + 0.0325$$

$$f_{12}(x) = x^2$$

$$f_{13}(x) = \cos(x)$$

