

The topology of software risk in scientific models

3. Global hydrological and land use models

Arnald Puy

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1 Preliminary

```
# PRELIMINARY FUNCTIONS #####
#####

sensobol::load_packages(c("data.table", "tidyverse", "openxlsx", "scales",
                          "cowplot", "readxl", "ggrepel", "tidytext", "here",
                          "tidygraph", "igraph", "foreach", "parallel", "ggraph",
                          "tools", "purrr", "sensobol", "benchmarkme"))

# Create custom theme -----

theme_AP <- function() {
  theme_bw() +
    theme(panel.grid.major = element_blank(),
          panel.grid.minor = element_blank(),
          legend.background = element_rect(fill = "transparent", color = NA),
          legend.key = element_rect(fill = "transparent", color = NA),
          strip.background = element_rect(fill = "white"),
          legend.text = element_text(size = 7.3),
          axis.title = element_text(size = 10),
          legend.key.width = unit(0.4, "cm"),
          legend.key.height = unit(0.4, "cm"),
          legend.key.spacing.y = unit(0, "lines"),
          legend.box.spacing = unit(0, "pt"),
          legend.title = element_text(size = 7.3),
          axis.text.x = element_text(size = 7),
          axis.text.y = element_text(size = 7),
          axis.title.x = element_text(size = 7.3),
          axis.title.y = element_text(size = 7.3),
          plot.title = element_text(size = 8),
          strip.text.x = element_text(size = 7.4),
          strip.text.y = element_text(size = 7.4))
}

# Select color palette -----

color_languages <- c("fortran" = "steelblue", "python" = "lightgreen")

# Source all .R files in the "functions" folder -----

r_functions <- list.files(path = here("functions"),
                          pattern = "\\..R$", full.names = TRUE)

lapply(r_functions, source)

# Set seed -----
```

```
seed <- 123
```

2 Analysis

```
# CREATE DATASET #####

# Path to folder -----

path <- "./datasets/call_metrics"

# List CSV files -----

files <- list.files(path, pattern = "\\*.csv$", full.names = TRUE)

# Split by language -----

python_files <- grep("python", files, value = TRUE, ignore.case = TRUE)
fortran_files <- grep("fortran", files, value = TRUE, ignore.case = TRUE)

base_fortran <- file_path_sans_ext(basename(fortran_files))
base_python <- file_path_sans_ext(basename(python_files))

model_names_fortran <- models <- sub(".*_", "", base_fortran)
model_names_python <- models <- sub(".*_", "", base_python)

# Load and name files -----

python_list <- lapply(python_files, fread)
fortran_list <- lapply(fortran_files, fread)

names(python_list) <- model_names_python
names(fortran_list) <- model_names_fortran

# RBIND -----

make_callgraph <- function(lst, lang) {
  rbindlist(lst, idcol = "model") %>%
    .[, language := lang] %>%
    .[, .(file, model, language, `function`, call)] %>%
    setnames(., c("function", "call"), c("from", "to"))
}

python_callgraphs <- make_callgraph(python_list, "python")
fortran_callgraphs <- make_callgraph(fortran_list, "fortran")

all_callgraphs <- rbind(python_callgraphs, fortran_callgraphs)

# SOURCE CODE CLASSIFICATION BY FUNCTIONAL ROLE #####
```

```

# Strip leading "./models/"-----
all_callgraphs[, file_clean:= sub("^\\.models/", "", file)]

# Provenance: file must be inside ./models to be eligible at all-----
all_callgraphs[, in_model_tree:= !is.na(file) & nchar(file) > 0L &
  grepl("^\\.models/", file)]

# Rest = everything after first segment-----
all_callgraphs[, rest:= sub("^[/]+/", "", file_clean)]

# If rest starts with model_id/ then drop it (this is the duplicate)-----
all_callgraphs[, rest:= fifelse(startsWith(rest, paste0(tolower(model), "/")),
  sub("^[/]+/", "", rest), rest)]

# In case of triple nesting like vic/vic/vic/...-----
all_callgraphs[, rest:= fifelse(startsWith(rest, paste0(tolower(model), "/")),
  sub("^[/]+/", "", rest), rest)]

# Top_level-----
all_callgraphs[, top_level:= tstrsplit(rest, "/", fixed = TRUE, keep = 1L)]

# Useful generic external patterns (works across models)-----
all_callgraphs[, is_generic_external:= in_model_tree & (
  grepl("(^|/)(extern|external|externals|third[_-]?party|vendor|vendored)(/|$)",
    rest, ignore.case = TRUE) |
  grepl("(^|/)\.lib(/|$)", rest, ignore.case = TRUE))]

# CTSM-specific external (framework + FoX parser under cdeps + CESM shared infra)
all_callgraphs[, is_ctsm_external:= (model == "CTSM") & in_model_tree & (
  grepl("^cime(/|$)", rest) |
  grepl("^cime_config(/|$)", rest) |
  grepl("^components/cdeps(/|$)", rest) |      # FoX XML/SAX parser
  grepl("(^|/)share_esmf(/|$)", rest) |        # shared ESMF infrastructure
  grepl("^src/unit_test_shr(/|$)", rest)       # unit tests
)]

# Extra CTSM allowlist (tightens "model core" to land-model code)-----
CTSM_STRICT <- TRUE

all_callgraphs[, is_ctsm_allowed:= TRUE]

if (CTSM_STRICT) {
  all_callgraphs[model == "CTSM" & in_model_tree, is_ctsm_allowed :=
    grepl("^src(/|$)", rest) |
    grepl("^lilac(/|$)", rest) |
    grepl("^components/cism(/|$)", rest)]
}

```

```

## Component classification (order matters)-----
all_callgraphs[nchar(file) == 0L | is.na(file), component:= NA_character_]

all_callgraphs[!is.na(file) & nchar(file) > 0L, component:= fcase(

  # Anything not in ./models is external immediately
  !in_model_tree, "external_lib",

  # Generic external dirs (vendored/third_party/etc.)
  is_generic_external, "external_lib",

  # CI and vendored libraries
  top_level == ".github", "ci_cd",
  top_level == ".lib" | grepl("/\\.lib/", rest), "vendored_lib",

  # CIME / CESM / CDEPS infrastructure (framework)
  grepl("^cime/CIME/", rest), "framework",
  grepl("^cime_config/", rest), "framework",
  grepl("^cime/doc/", rest), "framework",
  grepl("^components/cdeps/cime_config/", rest), "framework",

  # CTSM-specific: treat cime + cdeps (incl FoX) as external/framework
  is_ctsm_external, "framework",

  # CTSM shared "shr_*" routines (framework by symbol)
  (model == "CTSM") & (grepl("^shr_", from) | grepl("^shr_", to)), "framework",

  # Tests (incl. SystemTests, case-insensitive)
  grepl("SystemTests/", rest, ignore.case = TRUE), "tests",
  grepl("/tests?/^tests?/", rest, ignore.case = TRUE), "tests",
  grepl("(^|/)unit_test", rest, ignore.case = TRUE), "tests",

  # Drivers: Fortran code clearly in drivers directories
  grepl("(^|/)drivers(/|$)", rest) & language == "fortran", "driver",

  # Couplers: NUOPC / LILAC / CESM / cpl
  grepl("cpl_|/cesm|/cpl|/cpl_", rest), "coupler",

  # CLI / tools: scripts, setup, CTSM python utilities
  grepl("tools/|scripts/|cli\\.py$|setup\\.py$", rest), "cli_or_tool",
  grepl("^python/ctsm/", rest), "cli_or_tool",

  # Everything else: model core
  default = "model_core"
)]

# include flag for risk calculations -----

```

```

# Key changes:
# - require in_model_tree
# - exclude external_lib/framework/tests/etc (already via component filter)
# - CTSM strict allowlist (optional)
all_callgraphs[, include_in_risk := (
  in_model_tree &
  language %chin% c("fortran", "python") &
  component %chin% c("model_core", "coupler") &
  (model != "CTSM" | !CTSM_STRICT | is_ctsm_allowed)
)]

# --- Sanity check: should now be FALSE for the SAX/FoX functions in CTSM
all_callgraphs[
  model == "CTSM" & include_in_risk &
  (grepl("sax|parseDTD|parsefile|parsestring|runParser", from, ignore.case = TRUE) |
   grepl("sax|parseDTD|parsefile|parsestring|runParser", to, ignore.case = TRUE)),
  .(from, to, file, rest, component)
]

## Empty data.table (0 rows and 5 cols): from,to,file,rest,component
# SUMmarize -----

all_callgraphs[, .N, component]

##      component      N
##      <char> <int>
## 1:      ci_cd       5
## 2: external_lib    100
## 3:   framework   8603
## 4:      tests     194
## 5: cli_or_tool    820
## 6:   model_core 15727
## 7:      driver     64
## 8:      coupler    299

all_callgraphs[, .N, include_in_risk]

##   include_in_risk      N
##   <lgcl> <int>
## 1:      FALSE   9786
## 2:      TRUE  16026

# Remove module calls -----

all_callgraphs <- all_callgraphs[!(from %in% "<module>")] %>%
  .[include_in_risk == TRUE]

# LOAD CYCLOMATIC COMPLEXITY VALUES FOR FUNCTIONS AND SUBROUTINES #####

```

```

cc_unique <- fread("./datasets/cyclomatic_complexity_functions.csv")

# CREATE NETWORK FROM CALL GRAPHS #####

all_graphs <- all_callgraphs[, .(graph = list(as_tbl_graph(.SD, directed = TRUE))),
                             model]

# ADD NODE METRICS #####

# Define the weights to characterize risky nodes -----

alpha <- 0.6 # Weight to cyclomatic complexity
beta  <- 0.3 # Weight to in-degree (impact of bug upstream)
gamma <- 0.1 # Weight to betweenness (critical bridge)

# Add node metrics -----

all_graphs[, graph:= Map(function(g, m) {

  comp_sub <- cc_unique[model == m]

  # mean cyclomatic complexity for this model & language -----

  mean_cyclo <- mean(comp_sub$cyclomatic_complexity, na.rm = TRUE)

  g %>%
    activate(nodes) %>%

    # Left join with dataset with cyclomatic complexity values -----

    left_join(comp_sub, by = "name") %>%

    # replace NA cyclomatic_complexity with model-language mean -----

    mutate(cyclomatic_complexity = if (!is.na(mean_cyclo)) {

      ifelse(is.na(cyclomatic_complexity), mean_cyclo, cyclomatic_complexity)

    } else {

      # if even the mean is NA (all NA in comp_sub), leave as-is

      cyclomatic_complexity

    }) %>%

    # Remove Python MODULE_AGG / CLASS_AGG nodes from this graph

```



```

# because they are not callable -----

filter(!(language == "python" & type %in% c("MODULE_AGG", "CLASS_AGG"))) %>%

# Calculation of key network metrics -----

mutate(type = type,
      indeg = centrality_degree(mode = "in"),
      outdeg = centrality_degree(mode = "out"),
      btw = centrality_betweenness(directed = TRUE, weights = NULL),
      cyclo_sc = rescale(cyclomatic_complexity),
      indeg_sc = rescale(indeg),
      btw_sc = rescale(btw),
      risk_score = alpha * cyclo_sc + beta * indeg_sc + gamma * btw_sc)
},
graph, model)]

# EXTRACT NODE DF #####

all_graphs[, node_df := lapply(graph, as_tibble, what = "nodes")]

# Export full node df -----

full_node_df <- all_graphs %>%
  mutate(node_df = purrr::map(node_df, ~ select(.x, -model, -language))) %>%
  unnest(node_df) %>%
  select(-graph) %>%
  data.table()

write.xlsx(full_node_df, "full_node_df.xlsx")

# COMPUTE ALL PATHS AND THEIR RISK SCORES #####

all_graphs[, paths_tbl := Map(all_paths_fun, node_df, graph)]

# Export full paths df -----

full_paths_df <- all_graphs %>%
  unnest(paths_tbl) %>%
  select(-c(graph, node_df))

write.xlsx(full_paths_df, "full_paths_df.xlsx")

# CONDUCT UNCERTAINTY AND SENSITIVITY ANALYSIS #####

# Define sample size and order of effects -----

```

```

N <- 2^11
order <- "first"

# Run the function (we remove the vic and python model implementation because
# there are not paths) -----

all_graphs[!model == "VIC", uncertainty_sensitivity:= Map(full_ua_sa_risk_fun, node_df, paths_1

# UNNEST APPROPRIATELY #####

unnested_df <- all_graphs %>%
  mutate(us_nodes = map(uncertainty_sensitivity, "nodes"),
         us_paths = map(uncertainty_sensitivity, "paths"))

# Create SA data frame -----

full_sa_df <- unnested_df %>%
  select(us_nodes) %>%
  unnest(cols = c(us_nodes)) %>%
  select(name, model, sensitivity_indices) %>%
  unnest(cols = c(sensitivity_indices))

# Export
fwrite(full_sa_df, "full_sa_df.csv")

# Create UA data frame -----

full_ua_df <- unnested_df %>%
  select(model, us_paths) %>%
  unnest(cols = c(us_paths)) %>%
  data.table()

# Export
fwrite(full_ua_df, "full_ua_df.csv")

# CALCULATE SOME DESCRIPTIVE METRICS #####

tmp <- data.table(full_paths_df)[, .(n_paths = .N), model] %>%
  .[order(-n_paths)]

tmp2 <- data.table(full_node_df)[, .(n_nodes = .N), model] %>%
  .[order(-n_nodes)]

# Path to node ratio: how interconnected the model is.
# Model_cc: Proxy for algorithmic complexity of model.
# Avg_path_length: Proxy for depth of dependency chains (risk-highway potential)

```

```

# Model fragility: more (error) propagation routes.
models_metrics <- merge(tmp, tmp2) %>%
  .[, `:=`(path_to_node_ratio = n_paths / n_nodes,
           model_cc = n_paths / log(n_nodes),
           avg_path_length = n_nodes / log(n_paths + 1),
           model_fragility_index = n_paths / (n_nodes * (n_nodes - 1)))]

models_metrics

# Read descriptive_stats_file -----

num_cols <- c("files", "functions", "modules", "lines", "lines_code", "lines_comments")

descriptive_stats <- data.table(read_xlsx("./datasets/descriptive_statistics/descriptive_statistics.xlsx"))

descriptive_stats <- dcast(melt(descriptive_stats, id.vars="model", measure.vars=num_cols),
  model ~ variable, value.var="value", fun.aggregate = function(z) sum(z, na.rm = TRUE))
  .[, lines_function := lines_code / functions]

all_descriptive_df <- merge(models_metrics, descriptive_stats)

# Sort by model -----
model_ordered <- all_descriptive_df[, sum(lines), model] %>%
  .[order(V1)]

# Plot descriptive measures per model -----

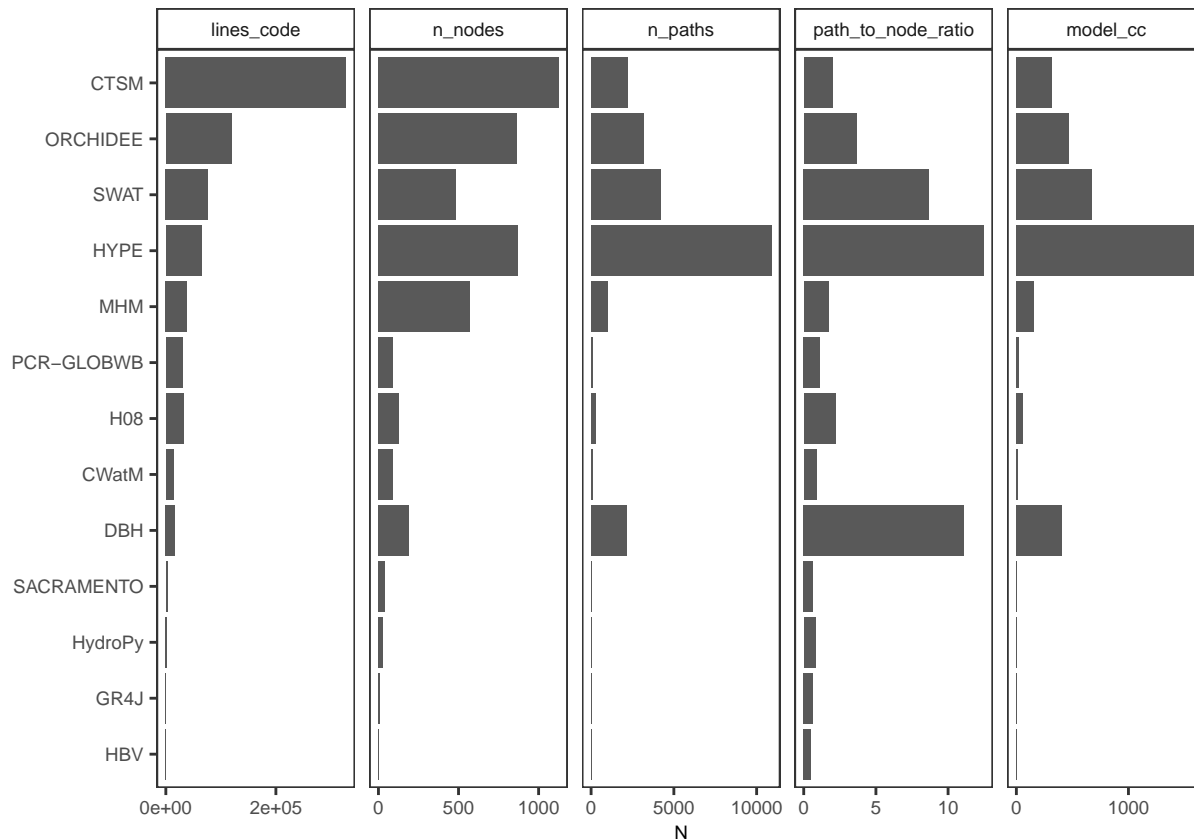
plot_descriptive <- melt(all_descriptive_df, measure.vars = c("lines_code", "n_nodes", "n_paths",
                                                             "path_to_node_ratio", "model_cc"))
  .[, model:= factor(model, levels = model_ordered[, model])] %>%
  ggplot(. , aes(model, value)) +
  geom_col() +
  coord_flip() +
  scale_y_continuous(breaks = breaks_pretty(n = 2)) +
  scale_fill_manual(values = color_languages, name = "") +
  facet_wrap(~ variable, ncol = 7, scales = "free_x") +
  labs(x = "", y = "N") +
  theme_AP() +
  theme(legend.position = c(0.1, 0.3))

## Warning in melt.data.table(all_descriptive_df, measure.vars = c("lines_code", :
## 'measure.vars' [lines_code, n_nodes, n_paths, path_to_node_ratio, ...] are not
## all of the same type. By order of hierarchy, the molten data value column will
## be of type 'double'. All measure variables not of type 'double' will be coerced
## too. Check DETAILS in ?melt.data.table for more on coercion.

```

```
plot_descriptive
```

```
## Warning: No shared levels found between `names(values)` of the manual scale and the
## data's fill values.
```



```
# METRICS AT THE FILE AND FUNCTION LEVEL #####

folder <- "./datasets/results_per_function"

# Get names of files -----

csv_files <- list.files(path = folder, pattern = "\\*.csv$", full.names = TRUE)

# Split into file_metrics and func_metrics -----

file_metric_files <- grep("file_metrics", csv_files, value = TRUE)
func_metric_files <- grep("func_metrics", csv_files, value = TRUE)

# Build one named list -----

list_metrics <- list(file_metrics = setNames(lapply(file_metric_files, fread),
                                                basename(file_metric_files)),
                    func_metrics = setNames(lapply(func_metric_files, fread),
                                                basename(func_metric_files)))
```

```

# Create function to combine files -----
make_combined <- function(subset_list, pattern) {
  rbindlist(subset_list[grepl(pattern, names(subset_list))], idcol = "source_file")
}

# Combine files -----

metrics_combined <- list(file_fortran = make_combined(list_metrics$file_metrics, "fortran"),
  file_python = make_combined(list_metrics$file_metrics, "python"),
  func_fortran = make_combined(list_metrics$func_metrics, "fortran"),
  func_python = make_combined(list_metrics$func_metrics, "python"))

# Functions to extract name of model and language from file -----

extract_model <- function(x)
  sub("^(file|func)_metrics_\\d+_[A-Za-z0-9-]+_(fortran|python).*", "\\2", x)

extract_lang <- function(x)
  sub("^(file|func)_metrics_\\d+_[A-Za-z0-9-]+_(fortran|python).*", "\\3", x)

# Extract name of model and language -----

metrics_combined <- lapply(metrics_combined, function(dt) {
  dt[, source_file := sub("\\.csv$", "", basename(source_file))]
  dt[, model := extract_model(source_file)]
  dt[, language := extract_lang(source_file)]
  dt
})

# Add column of complexity category -----

metrics_combined <- lapply(names(metrics_combined), function(nm) {
  dt <- as.data.table(metrics_combined[[nm]])
  if (grepl("^func_", nm) && "cyclomatic_complexity" %in% names(dt)) {
    dt[, complexity_category := cut(
      cyclomatic_complexity,
      breaks = c(-Inf, 10, 20, 50, Inf),
      labels = c("b1", "b2", "b3", "b4")
    )]
  }
  dt
}) |> setNames(names(metrics_combined))

# Define labels -----

```

```

lab_expr <- c(b1 = expression(C %in% "(" * 0 * ", 10" * "]" ),
             b2 = expression(C %in% "(" * 10 * ", 20" * "]" ),
             b3 = expression(C %in% "(" * 20 * ", 50" * "]" ),
             b4 = expression(C %in% "(" * 50 * ", " * infinity * "]" ))

# Define vector to exclude classes that are not functions -----
excluded_classes_vec <- c("MODULE_AGG", "CLASS_AGG")

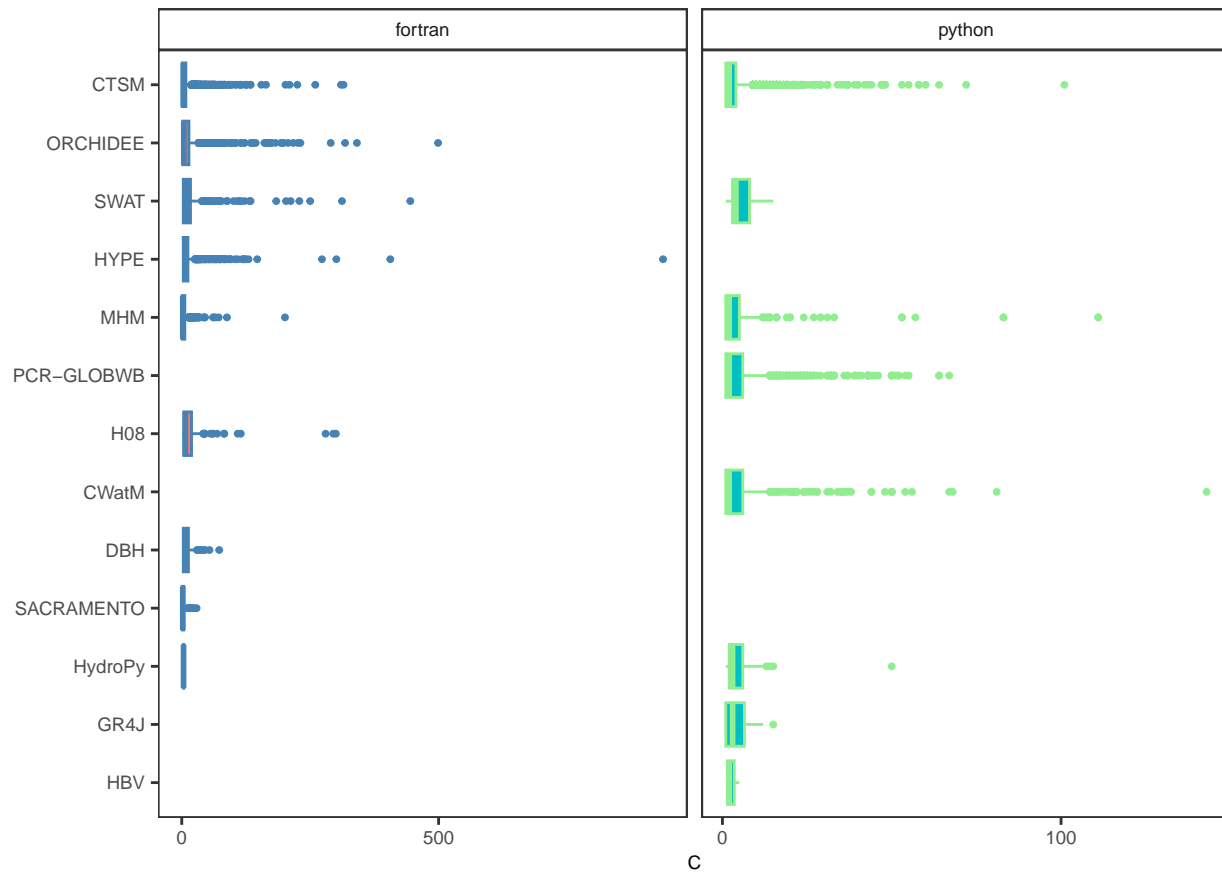
# PLOT #####

## ----plot_c_model, dependson="read_metrics_function_data", fig.height=2.2, fig.width=3.1----

plot_c_model <- metrics_combined[grep("^func_", names(metrics_combined))] %>%
  lapply(., function(x)
    x[, .(model, language, `function`, cyclomatic_complexity, loc, bugs, type)]) %>%
  rbindlist() %>%
  .[!type %in% excluded_classes_vec] %>%
  .[, model:= factor(model, levels = model_ordered[, model])] %>%
  na.omit() %>%
  ggplot(., aes(model, cyclomatic_complexity, fill = language, color = language)) +
  geom_boxplot(outlier.size = 0.7) +
  coord_flip() +
  scale_y_continuous(breaks = scales::breaks_pretty(n = 2)) +
  facet_wrap(~language, scales = "free_x") +
  labs(x = "", y = "C") +
  theme_AP() +
  scale_color_manual(values = color_languages) +
  theme(legend.position = "none",
        plot.margin = margin(0, 2, 0, 0))

plot_c_model

```



```
## ----plot_scatter_and_bar, dependson="read_metrics_function_data", fig.height=2.5, fig.width=10
```

```
# Scatterplot cyclomatic vs lines of code -----
```

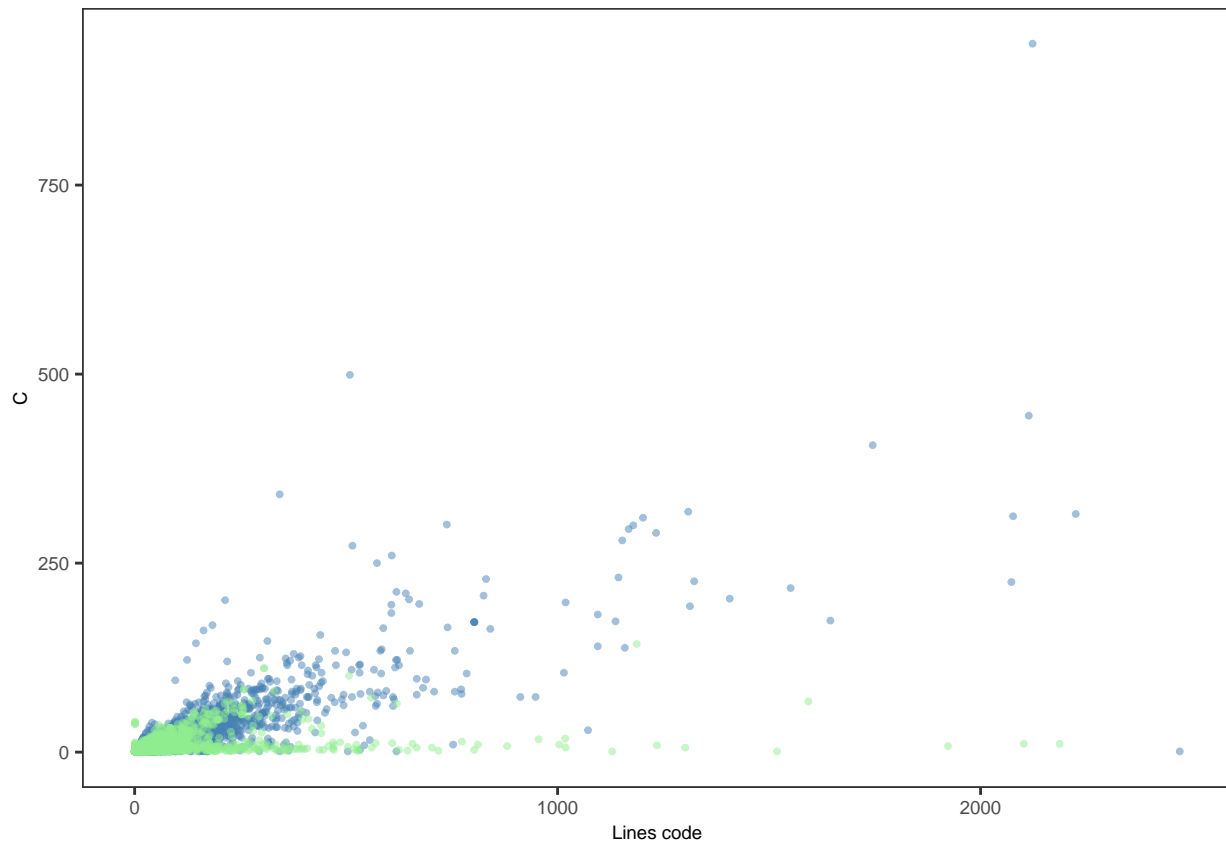
```
plot_c_vs_loc <- metrics_combined[grep("^func_", names(metrics_combined))] %>%
  lapply(., function(x) x[, .(loc, cyclomatic_complexity, language)]) %>%
  rbindlist() %>%
  ggplot(., aes(loc, cyclomatic_complexity, color = language)) +
  geom_point(alpha = 0.5, size = 0.7) +
  scale_x_continuous(breaks = breaks_pretty(n = 3)) +
  labs(x = "Lines code", y = "C") +
  scale_color_manual(values = color_languages) +
  theme_AP() +
  scale_x_continuous(breaks = breaks_pretty(n = 2)) +
  theme(legend.position = "none")
```

```
## Scale for x is already present.
```

```
## Adding another scale for x, which will replace the existing scale.
```

```
plot_c_vs_loc
```

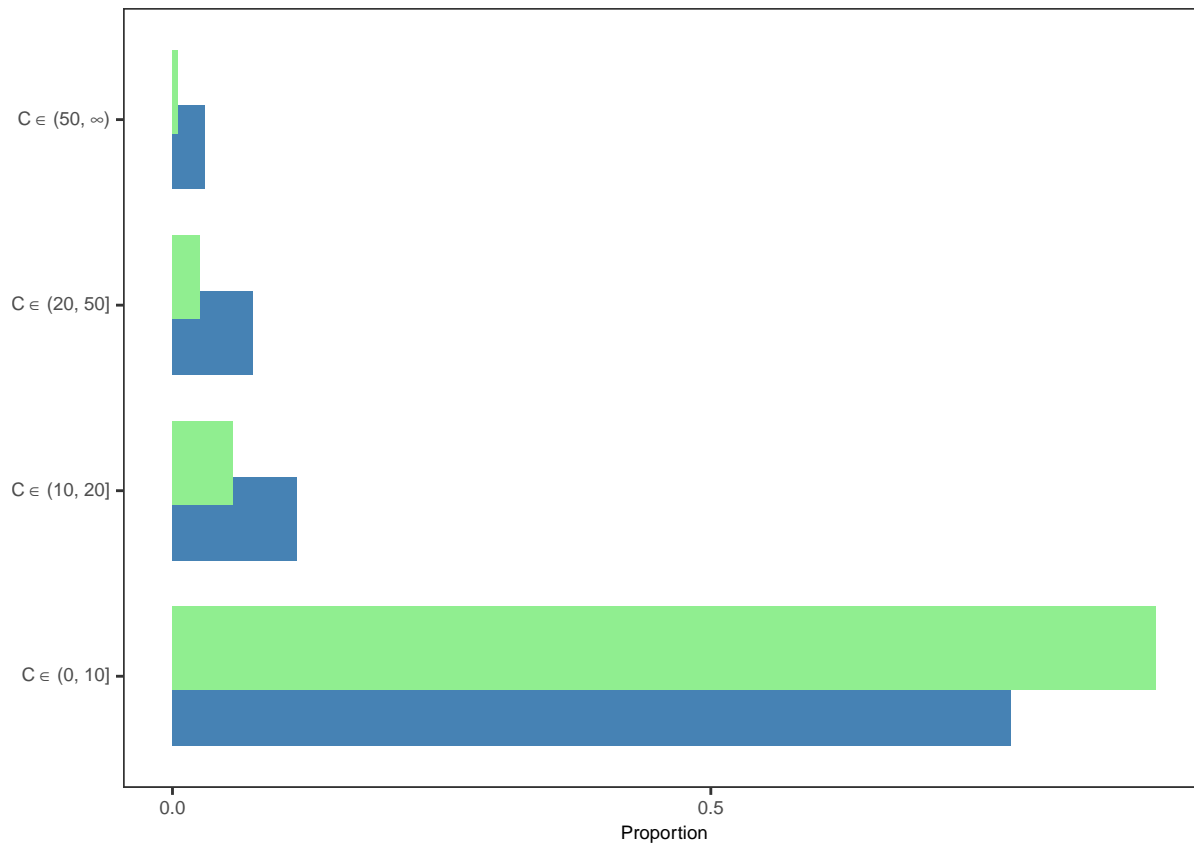
```
## Warning: Removed 1195 rows containing missing values or values outside the scale range
## (`geom_point()`).
```



```
# Count & proportion -----

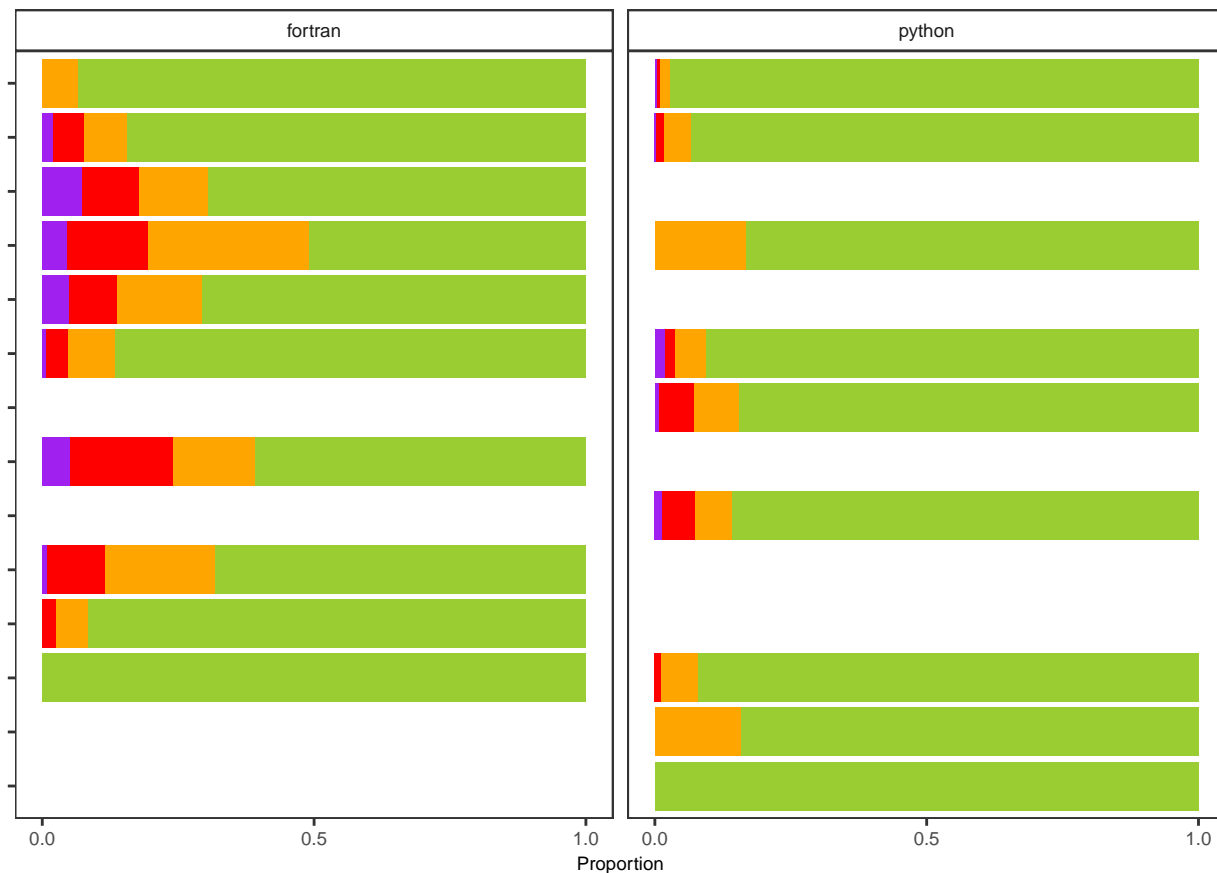
plot_bar_cyclomatic <- metrics_combined[grep("^func_", names(metrics_combined))] %>%
  lapply(., function(x) x[, .(complexity_category, language, type)]) %>%
  rbindlist() %>%
  .[!type %in% excluded_classes_vec] %>%
  .[, .N, .(complexity_category, language)] %>%
  .[, proportion := N / sum(N), language] %>%
  ggplot(., aes(complexity_category, proportion, fill = language)) +
  geom_bar(stat = "identity", position = position_dodge(0.6)) +
  scale_fill_manual(values = color_languages) +
  scale_y_continuous(breaks = scales::breaks_pretty(n = 3)) +
  scale_x_discrete(labels = lab_expr) +
  labs(x = "", y = "Proportion") +
  coord_flip() +
  theme_AP() +
  theme(legend.position = "none")

plot_bar_cyclomatic
```

```
plot_bar_category <- metrics_combined[grep("^func_", names(metrics_combined))] %>%
  lapply(., function(x)
    x[, .(model, language, complexity_category, type)] %>%
    rbindlist() %>%
    .[!type %in% excluded_classes_vec] %>%
    .[, model := factor(model, levels = model_ordered[, model])] %>%
    .[, .N, .(model, language, complexity_category)] %>%
    .[, proportion := N / sum(N), .(language, model)] %>%
    ggplot(., aes(model, proportion, fill = complexity_category)) +
    geom_bar(stat = "identity") +
    scale_fill_manual(values = c("yellowgreen", "orange", "red", "purple"),
                      labels = lab_expr,
                      name = "") +
    facet_wrap(~language) +
    labs(x = "", y = "Proportion") +
    coord_flip() +
    scale_y_continuous(breaks = scales::breaks_pretty(n = 3)) +
    theme_AP() +
    theme(legend.position = "none") +
    theme(axis.text.y = element_blank(),
          legend.text = element_text(size = 7),
          plot.margin = margin(0, 0, 0, 2))
```

plot_bar_category



3 Descriptive plots

```
# MERGE FIGURES #####
```

```
legend2 <- get_legend_fun(plot_bar_category + theme(legend.position = "top"))
```

```
## Warning: `is.ggplot()` was deprecated in ggplot2 3.5.2.
```

```
## i Please use `is_ggplot()` instead.
```

```
## This warning is displayed once every 8 hours.
```

```
## Call `lifecycle::last_lifecycle_warnings()` to see where this warning was
```

```
## generated.
```

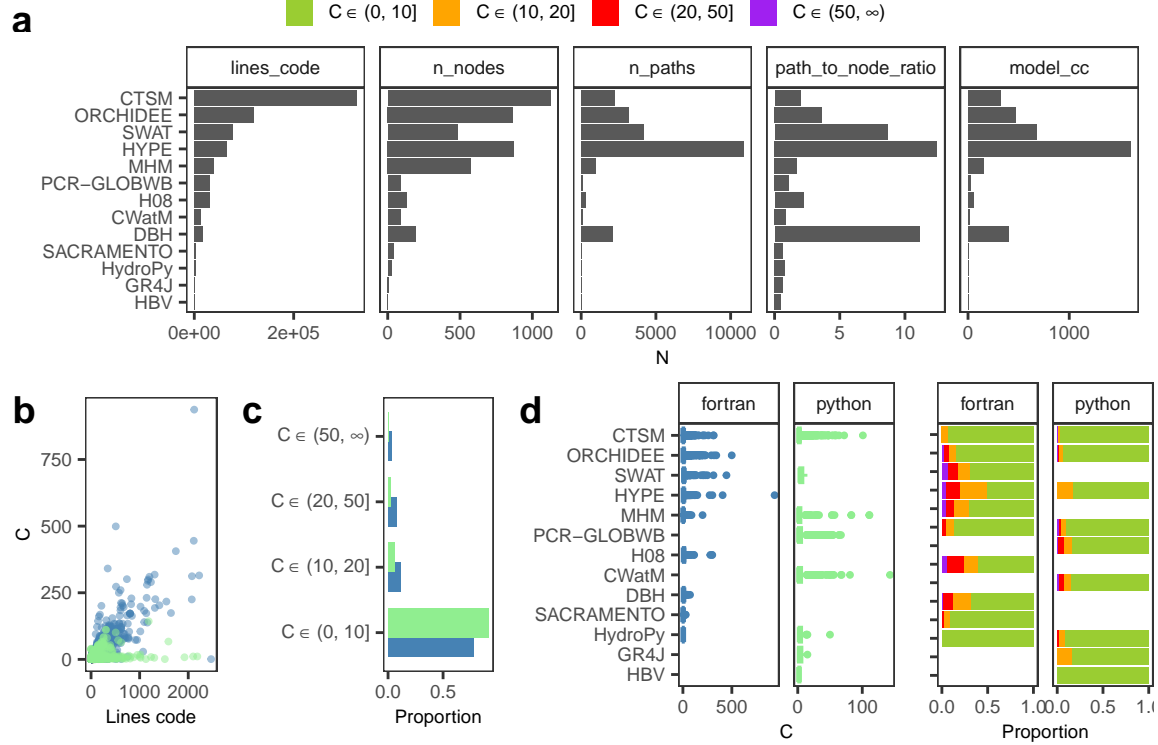
```
top_plot <- plot_grid(legend2, plot_descriptive, rel_heights = c(0.1, 0.9), ncol = 1,
  labels = "a")
```

```
## Warning: No shared levels found between `names(values)` of the manual scale and the
## data's fill values.
```

```
bottom <- plot_grid(plot_c_vs_loc, plot_bar_cyclomatic, plot_c_model,
  plot_bar_category, ncol = 4, rel_widths = c(0.2, 0.24, 0.34, 0.22),
  labels = c("b", "c", "d"))
```

```
## Warning: Removed 1195 rows containing missing values or values outside the scale range
## (`geom_point()`).
```

```
plot_grid(top_plot, bottom, ncol = 1, rel_heights = c(0.52, 0.48), align = "h",
          axis = "tb")
```



4 Figures

```
# PLOT FIGURES #####

# Define language of models -----

python_models <- c("CWatM", "GR4J", "HBV", "PCR-GLOBWB")
fortran_models <- c("ORCHIDEE", "HO8", "HYPE", "DBH", "SACRAMENTO")
python_and_fortran <- c("CTSM", "SWAT", "MHM", "HydroPy", "VIC")

all_graphs[, language := fcase(model %chin% python_models, "python",
                               model %chin% fortran_models, "fortran",
                               model %chin% python_and_fortran, "python+fortran",
                               default = NA_character_)]

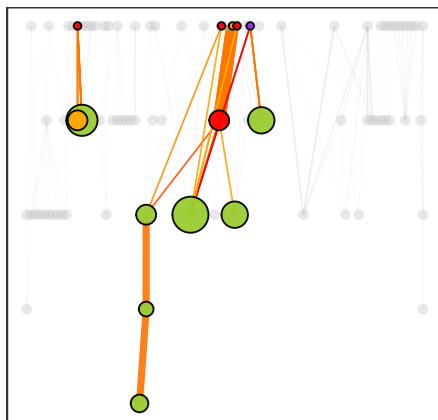
# Plot graphs -----

set.seed(seed)

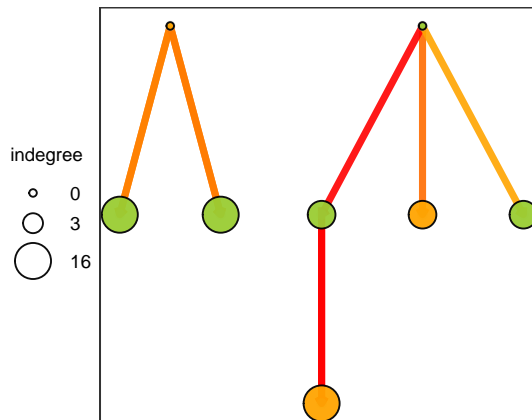
# Thickness of edge: frequency across top-10 riski paths
# Color of edge: mean risk of paths using that edge

all_graphs <- all_graphs[, plot_obj := mapapply(plot_top_paths_fun, call_g = graph,
                                                paths_tbl = paths_tbl, model.name = model,
                                                language = language, SIMPLIFY = FALSE)]
```

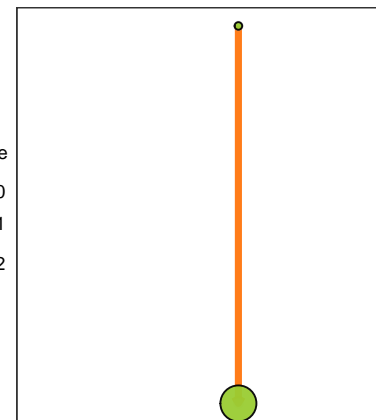
CWatM: python



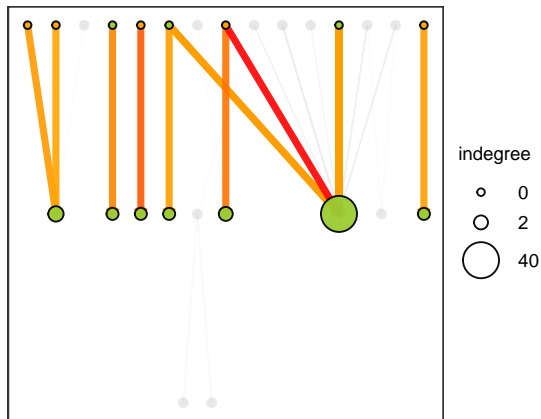
GR4J: python



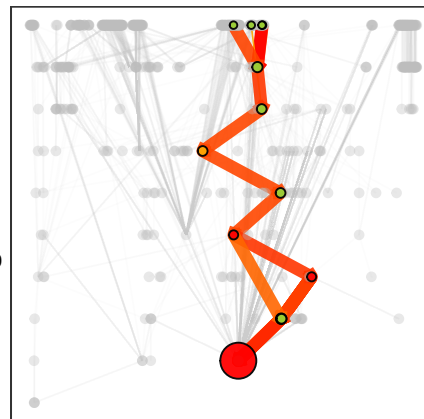
HBV: python



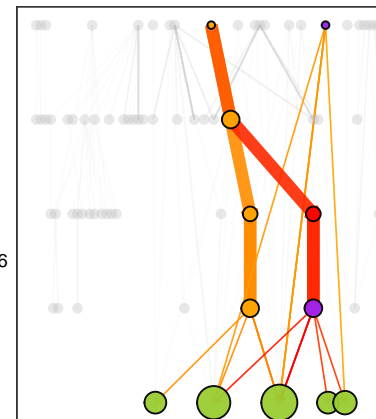
HydroPy: python+fortran



MHM: python+fortran

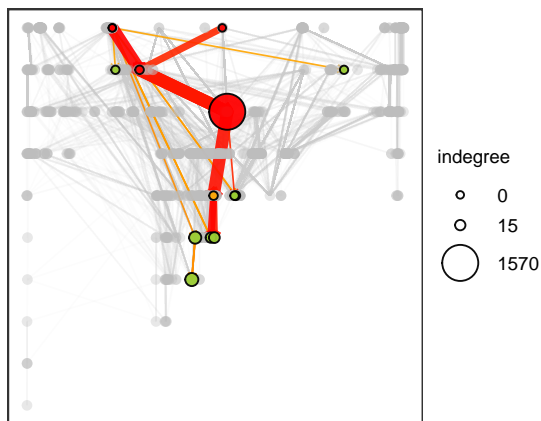


PCR-GLOBWB: python

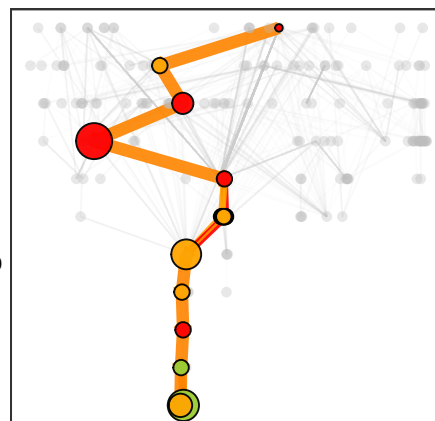


No paths in paths_tbl; skipping plot for: VIC

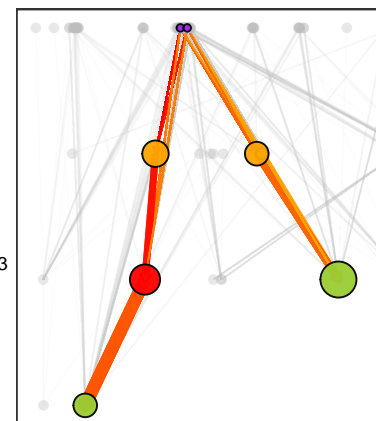
CTSM: python+fortran



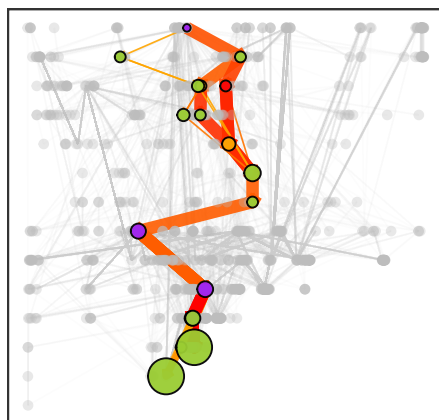
DBH: fortran



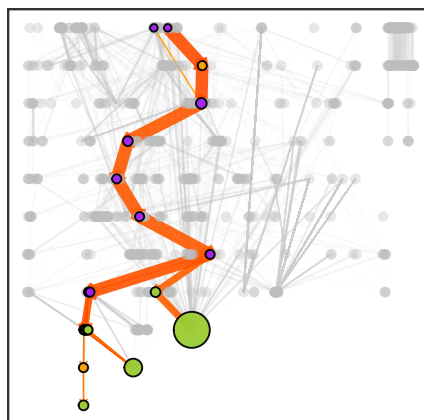
H08: fortran



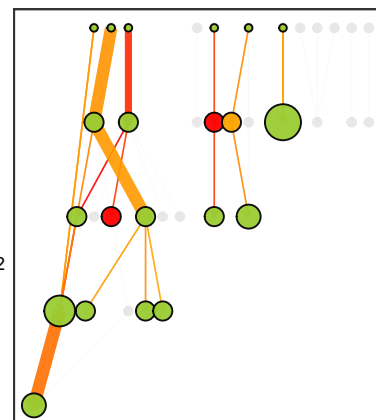
HYPE: fortran



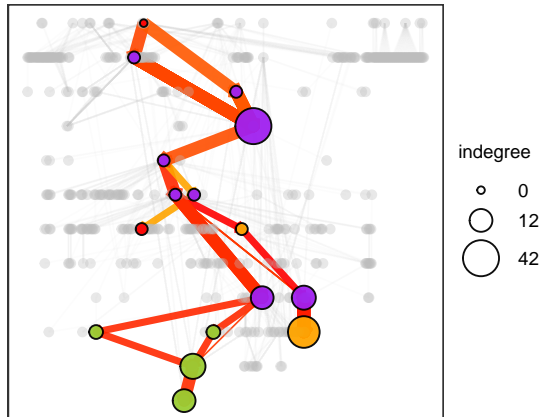
ORCHIDEE: fortran



SACRAMENTO: fortran



SWAT: python+fortran



PLOT OTHER FIGURES

```
selected_models <- data.table(model = c("CTSM", "PCR-GLOBWB", "DBH", "HYPE",
                                         "ORCHIDE", "SWAT", "CWatM", "MHM"))
```

Plot call graphs -----

```
tmp <- all_graphs[selected_models, on = .(model)]
plot_all_risky_paths <- plot_grid(plotlist = tmp$plot_obj, ncol = 2, align = "hv")
```

Plot risk_slope -----

```
a <- full_paths_df %>%
  data.table() %>%
  .[selected_models, on = .(model)] %>%
  .[order(-p_path_fail), .SD[1:10], model] %>%
  ggplot(., aes(reorder(model, risk_slope), risk_slope)) +
  geom_boxplot() +
  scale_y_continuous(breaks = pretty_breaks(n = 3)) +
  geom_hline(yintercept = 0, lty = 2, color = "red") +
  coord_flip() +
  labs(x = "", y = expression(theta[1*k])) +
  theme_AP()
```

Plot Gini metric -----

```
b <- full_paths_df %>%
  data.table() %>%
  .[selected_models, on = .(model)] %>%
  .[order(-p_path_fail), .SD[1:10], model] %>%
  ggplot(., aes(reorder(model, gini_node_risk), gini_node_risk)) +
  geom_boxplot() +
  coord_flip() +
  labs(x = "", y = expression(G[k])) +
```

```

theme_AP()

# Plot Si values -----

c <- full_sa_df %>%
  data.table() %>%
  .[selected_models, on = .(model)] %>%
  .[sensitivity == "Si", .(median = median(original, na.rm = TRUE)), .(model, parameters)] %>%
  ggplot(., aes(x = parameters, y = model, fill = median)) +
  geom_tile() +
  scale_fill_viridis_c(name = expression("Med(" * S[p] * ")"),
    limits = c(0, 1),
    breaks = c(0, 0.5, 1)) +
  scale_x_discrete(labels = c(a_raw = expression(alpha),
    b_raw = expression(beta),
    c_raw = expression(gamma))) +
  labs(x = NULL, y = NULL) +
  theme_AP() +
  theme(legend.position = "none")

# Plot interaction strength -----

tmp <- full_sa_df %>%
  data.table() %>%
  dcast(., name + model + parameters ~ sensitivity, value.var = "original",
    fun.aggregate= mean) %>%
  .[, interaction:= Ti - Si]

d <- tmp %>%
  .[selected_models, on = .(model)] %>%
  .[, .(median = median(interaction, na.rm = TRUE)), .(parameters, model)] %>%
  ggplot(., aes(x = parameters, y = model, fill = median)) +
  geom_tile() +
  scale_x_discrete(labels = c(a_raw = expression(alpha),
    b_raw = expression(beta),
    c_raw = expression(gamma))) +
  scale_fill_viridis_c(name = expression("Med(" * T[p] - S[p] * ")"),
    limits = c(0, 0.06),
    breaks = c(0, 0.03, 0.06),
    option = "magma") +
  labs(x = NULL, y = NULL) +
  theme_AP() +
  theme(legend.position = "none")

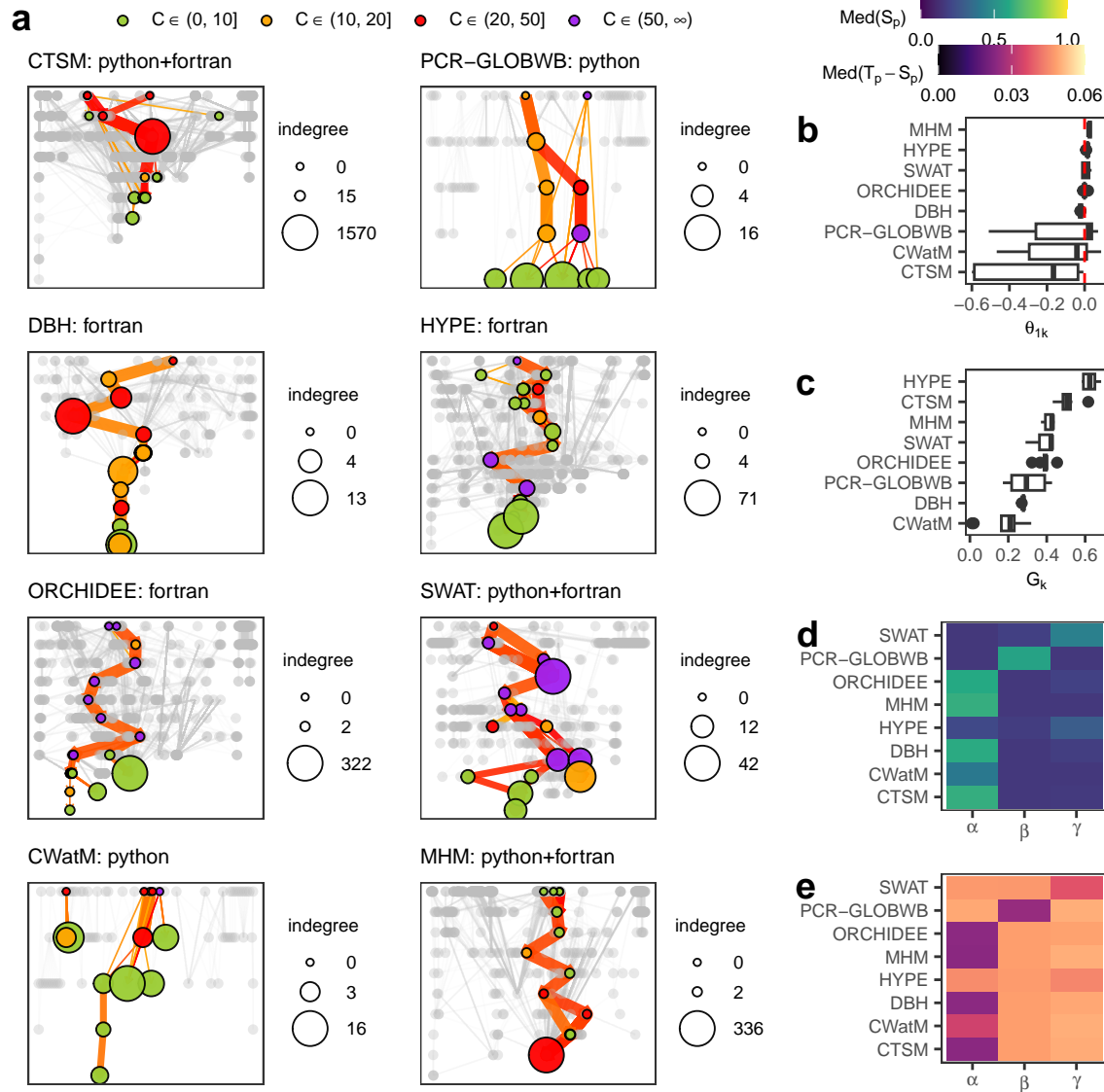
# MERGE #####

```

```

p_for_fill_legend <- all_graphs$plot_obj[[6]] +
  guides(size = "none", fill = guide_legend(title = ""))
fill_legend <- get_legend_fun(p_for_fill_legend + theme(legend.position = "top"))
plot_top_paths <- plot_grid(fill_legend, plot_all_risky_paths, ncol = 1, rel_heights = c(0.05,
  labels = "a"))
heatmap_legend <- get_legend(c + theme(legend.position = "top"))
ti_legend <- get_legend(d + theme(legend.position = "top"))
dada <- plot_grid(a, b, c, d, ncol = 1, labels = c("b", "c", "d", "e"))
all_legends <- plot_grid(heatmap_legend, ti_legend, ncol = 1)
right_plot <- plot_grid(all_legends, dada, ncol = 1, rel_heights = c(0.1, 0.9))
plot_grid(plot_top_paths, right_plot, ncol = 2, rel_widths = c(0.7, 0.3))

```



PATH-LEVEL RISK ACCOUNTED FOR THE TOP 5% NODES

```
setDT(full_paths_df)
```



```

# To long format -----
paths_long <- full_paths_df[, .(node = unlist(path_nodes),
  p_path_fail = p_path_fail,
  gini_node_risk = gini_node_risk,
  risk_slope = risk_slope,
  risk_mean = risk_mean,
  risk_sum = risk_sum),
  .(model, path_id)]

# Aggregate at function level -----
node_from_paths <- paths_long[, .(n_paths = .N,
  mean_p_path = mean(p_path_fail, na.rm = TRUE),
  max_p_path = max(p_path_fail, na.rm = TRUE),
  sum_p_path = sum(p_path_fail, na.rm = TRUE),
  mean_gini = mean(gini_node_risk, na.rm = TRUE),
  mean_slope = mean(risk_slope, na.rm = TRUE),
  mean_risksum = mean(risk_sum, na.rm = TRUE)),
  .(model, node)]

# Join with nodes -----
node_summary <- merge(node_from_paths, full_node_df, by.x = c("model", "node"),
  by.y = c("model", "name"), all.x = TRUE)

# Calculate risk mass -----
node_summary[, risk_mass := mean_p_path * n_paths]

# share of risk mass in top X% nodes, per model -----

top_share <- function(X = 0.05) {
  node_summary[!is.na(risk_mass) & risk_mass >= 0, {
    dt <- .SD[order(-risk_mass)]
    n_top <- max(1L, ceiling(.N * X))
    .(X = X, n_nodes = .N, n_top = n_top,
      share_risk_mass_topX = sum(dt$risk_mass[1:n_top]) / sum(dt$risk_mass))
  },
  model
]
}

# Run function -----

```

```
tmp <- top_share(0.05) %>%
  .[order(-share_risk_mass_topX)]
```

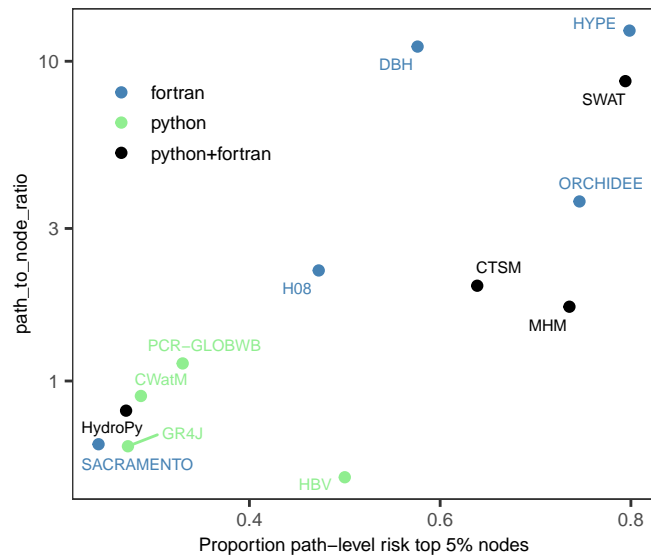
```
tmp
```

```
##      model      X n_nodes n_top share_risk_mass_topX
##      <char> <num>   <int> <num>           <num>
##  1:      HYPE  0.05     872   44           0.7985366
##  2:      SWAT  0.05     481   25           0.7942209
##  3: ORCHIDEE  0.05     865   44           0.7460796
##  4:      MHM   0.05     565   29           0.7355383
##  5:      CTSM  0.05    1124   57           0.6388660
##  6:      DBH   0.05     191   10           0.5763146
##  7:      HBV   0.05        2    1           0.5000000
##  8:      H08   0.05     129    7           0.4726677
##  9: PCR-GLOBWB 0.05      89    5           0.3300054
## 10:      CWatM 0.05      84    5           0.2862374
## 11:      GR4J   0.05       8    1           0.2726486
## 12:   HydroPy  0.05      26    2           0.2706942
## 13: SACRAMENTO 0.05      41    3           0.2418015
```

```
# Plot-----

color_languages_2 <- c("fortran" = "steelblue", "python" = "lightgreen",
  "python+fortran" = "black")

merge(all_descriptive_df, tmp, by = "model") %>%
  .[, language := fcase(model %chin% python_models, "python",
    model %chin% fortran_models, "fortran",
    model %chin% python_and_fortran, "python+fortran",
    default = NA_character_)] %>%
  ggplot(., aes(share_risk_mass_topX, path_to_node_ratio, color = language)) +
  geom_point() +
  scale_color_manual(values = color_languages_2, name = "") +
  geom_text_repel(aes(label = model), size = 2, max.overlaps = Inf, show.legend = FALSE) +
  scale_y_log10() +
  labs(x = "Proportion path-level risk top 5% nodes", y = "path_to_node_ratio") +
  theme_AP() +
  theme(legend.position = c(0.2, 0.8))
```



PLOT THE TOP 50 PATHS PER MODEL

```
tmp <- full_ua_df %>%
  .[order(-P_k_mean), .SD[1:50], model] %>%
  na.omit() %>%
  split(., .$model)
```

Plot in a for loop -----

```
out <- list()
```

```
for ( i in 1:length(tmp)) {
```

```
  out[[i]] <- ggplot(tmp[[i]], aes(P_k_mean, reorder(path_str, P_k_mean), color = risk_slope))
    geom_point(size = 1) +
    geom_errorbar(aes(xmin = P_k_q025, xmax = P_k_q975), height = 0.2) +
    scale_color_gradient2(low = "blue", mid = "grey80", high = "red", midpoint = 0,
                          name = expression(beta[k])) +
    labs(y = "Path ID", x = expression(P[k])) +
    theme_AP() +
    scale_x_continuous(breaks = breaks_pretty(n = 3),
                       limits = c(0, 1)) +
    theme(axis.text.y = element_text(size = 4),
          legend.position = "top") +
    ggtitle(names(tmp[i]))
```

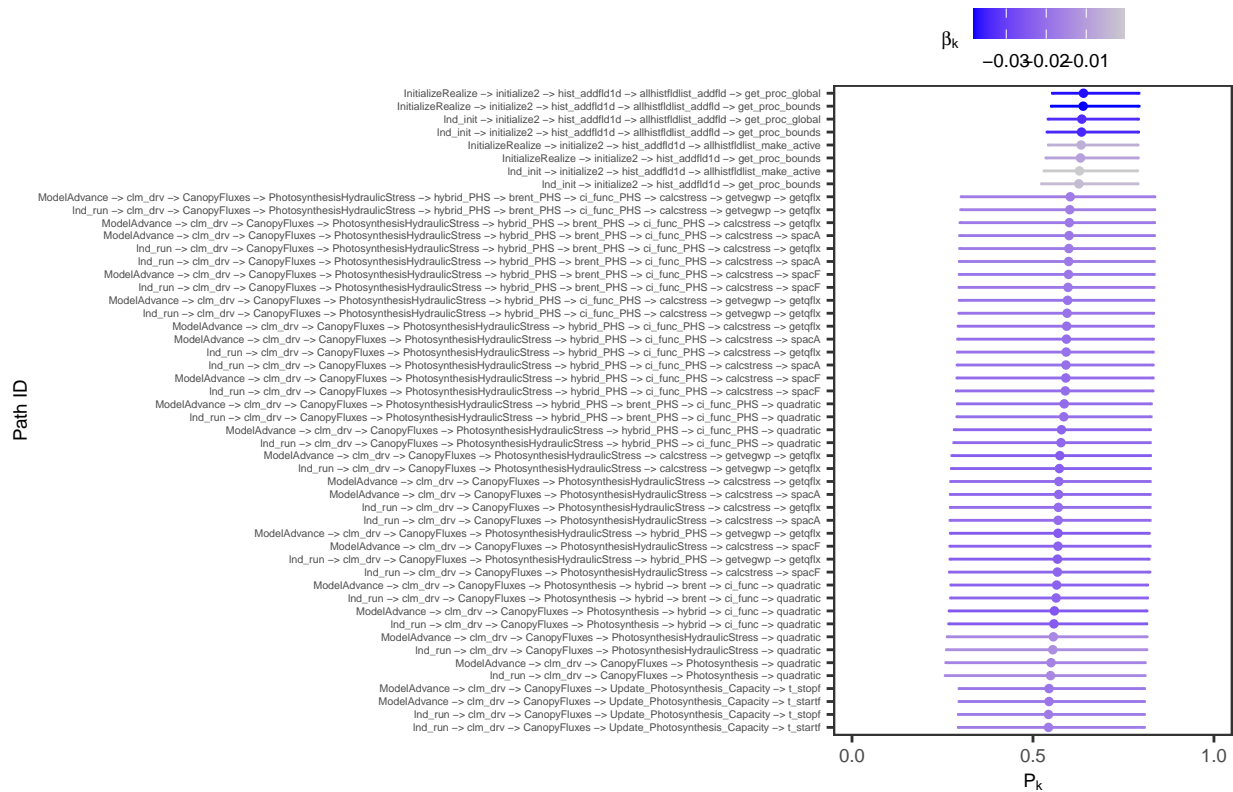
```
}
```

```
out
```

```
## [[1]]
```

```
## `height` was translated to `width`.
```

CTSM

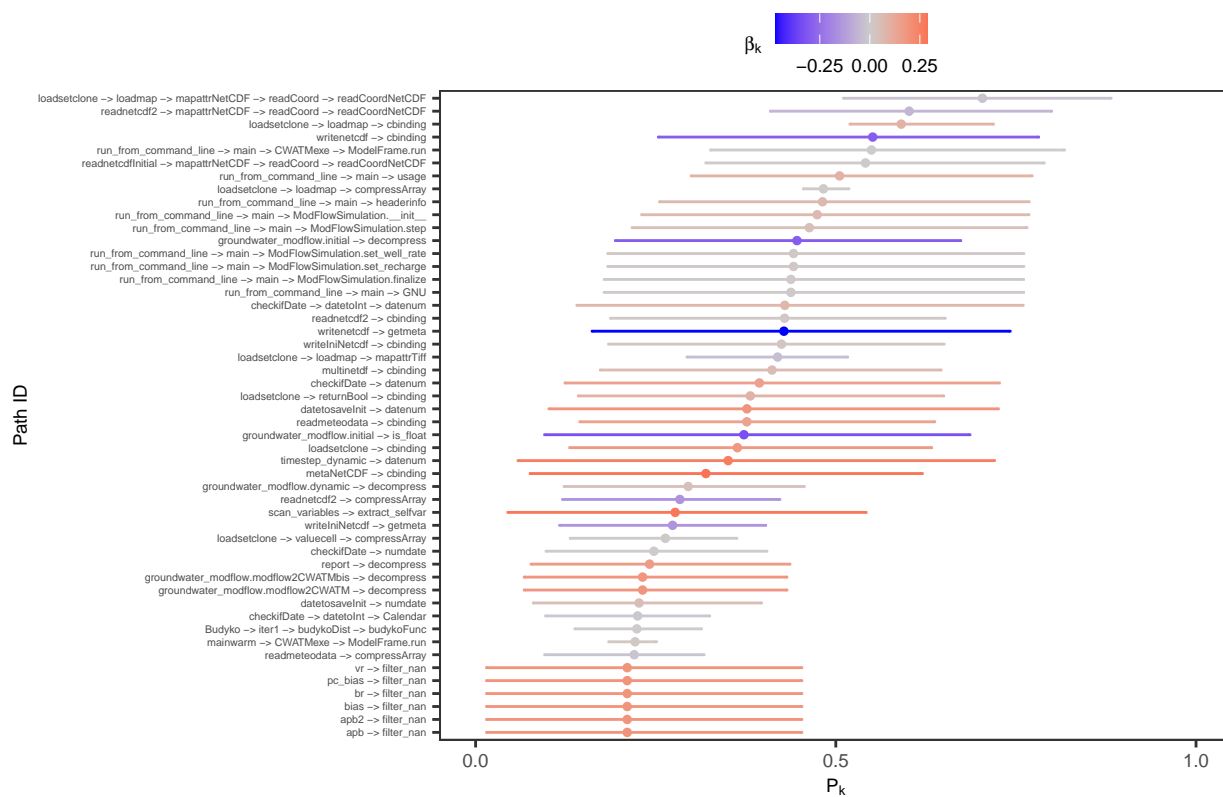


##

```
## [[2]]
```

```
## `height` was translated to `width`.
```

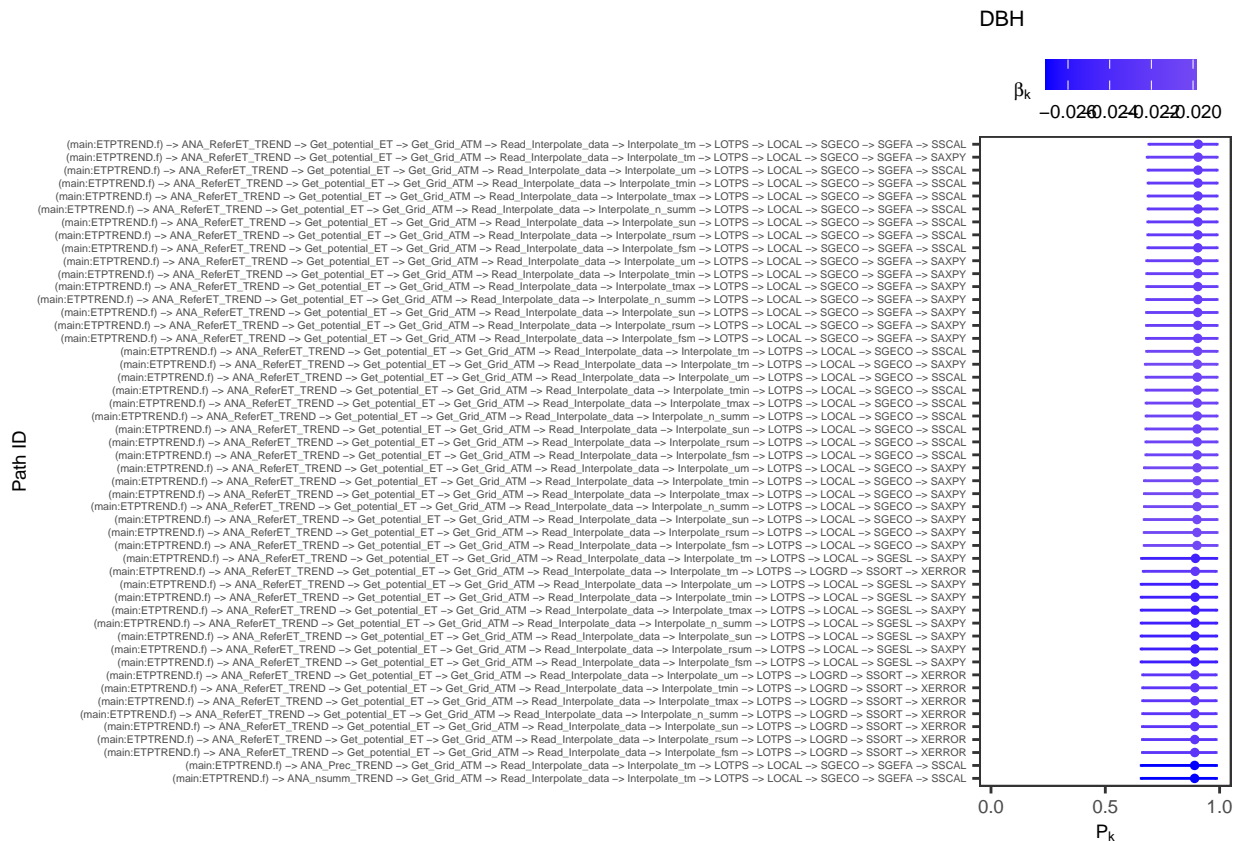
CWatM



##

[[3]]

`height` was translated to `width`.

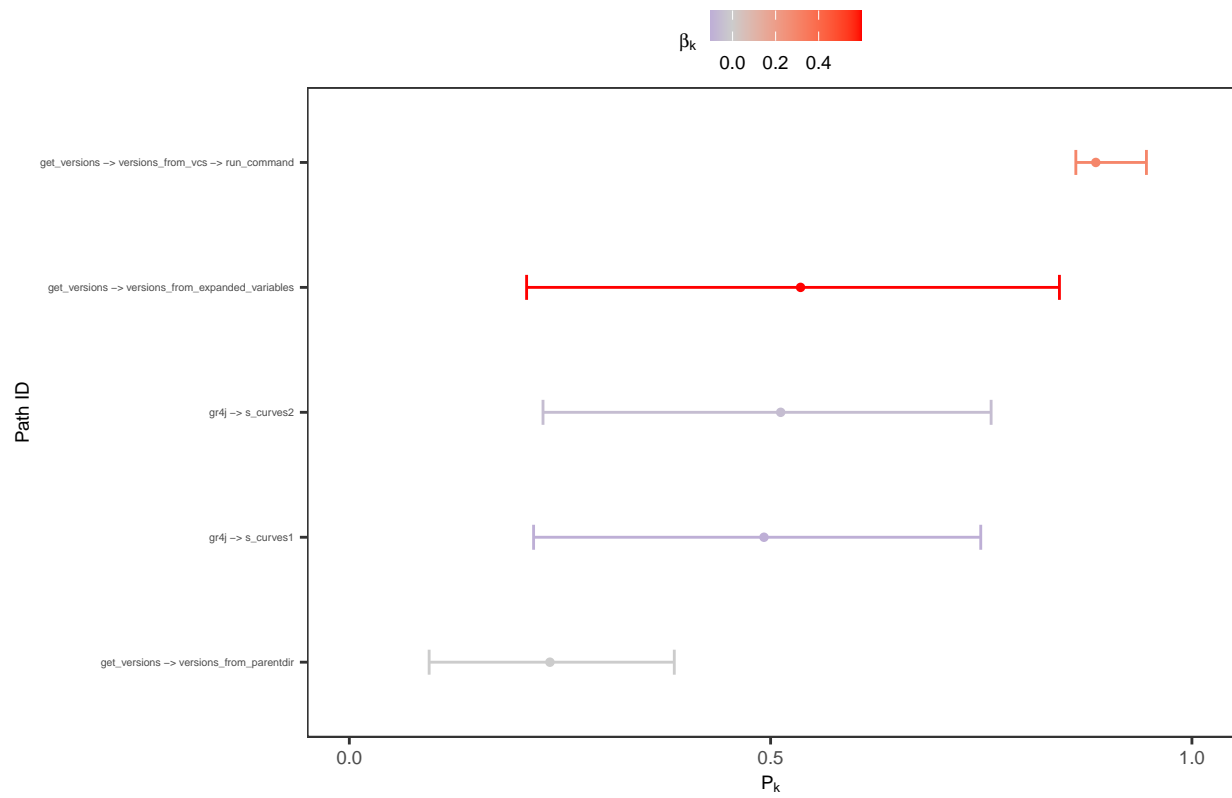


##

[[4]]

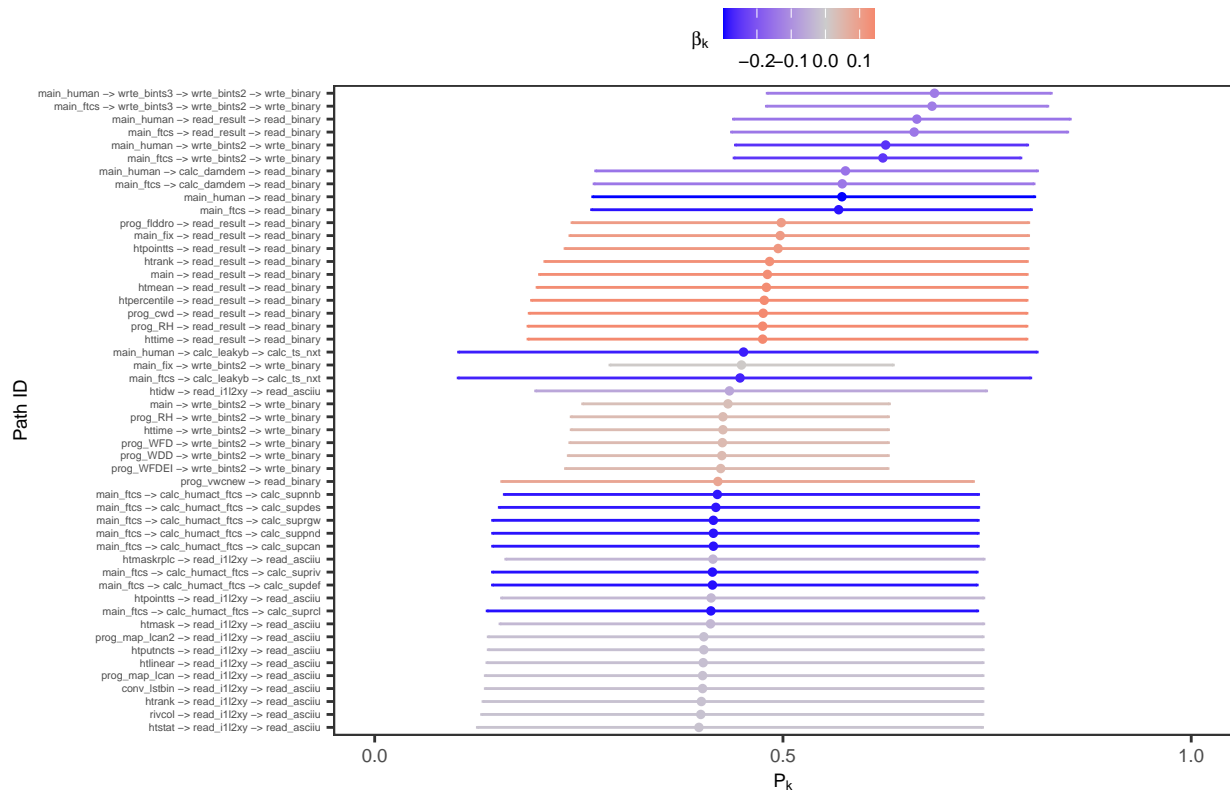
`height` was translated to `width`.

GR4J



```
##
## [[5]]
## `height` was translated to `width`.
```

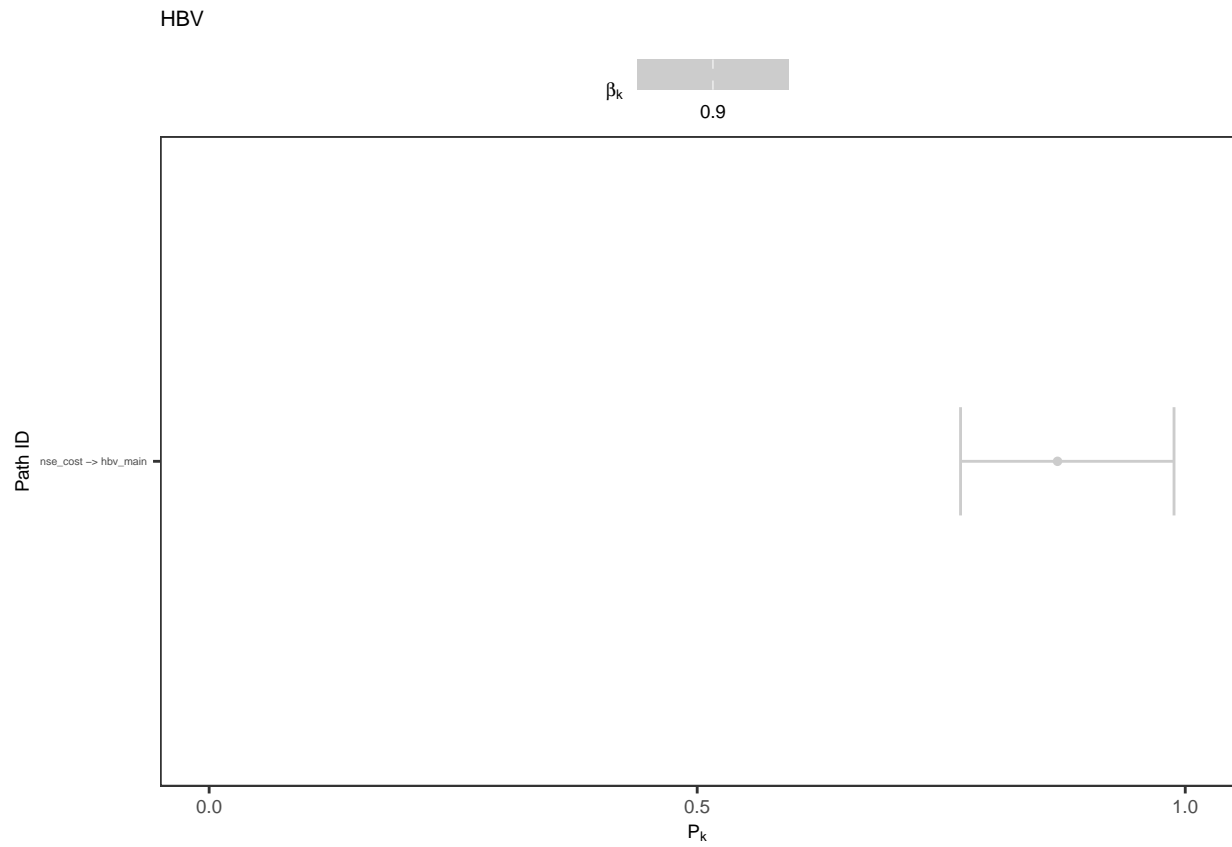
H08



##

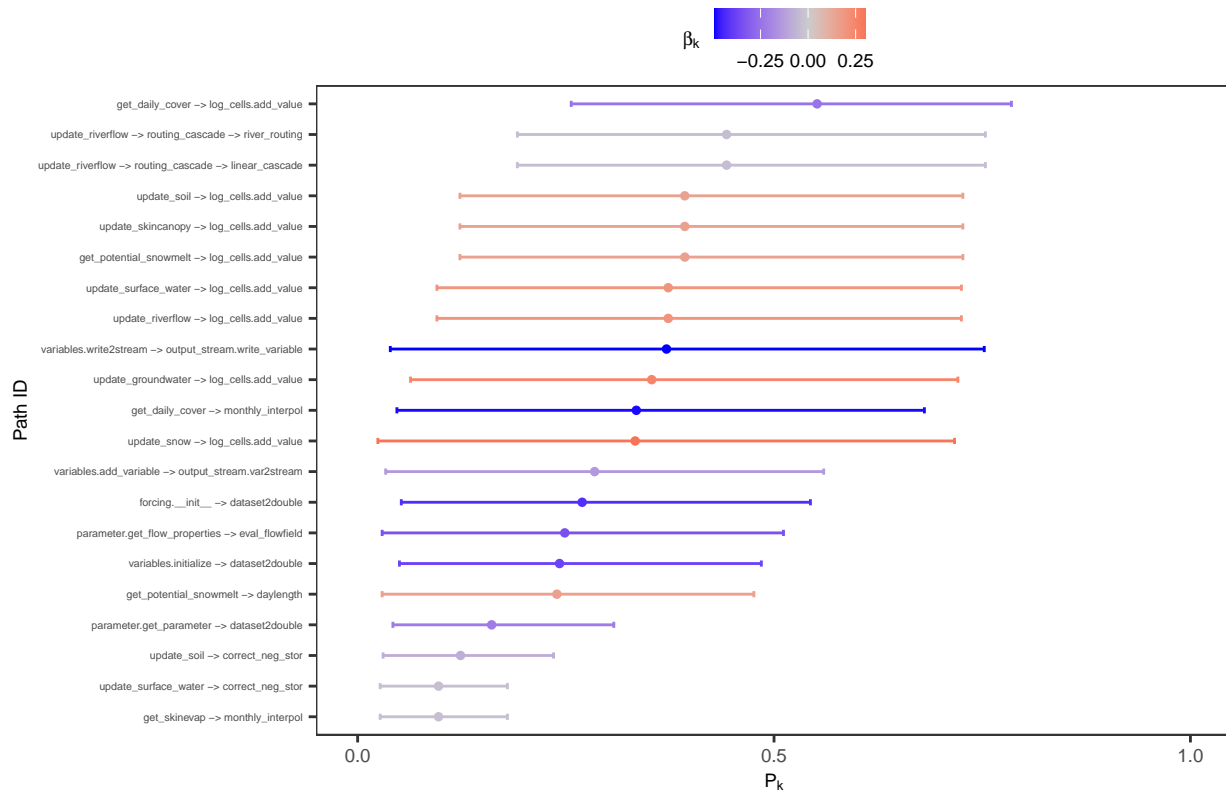
[[6]]

`height` was translated to `width`.



```
##  
## [[7]]  
## `height` was translated to `width`.
```

HydroPy



##

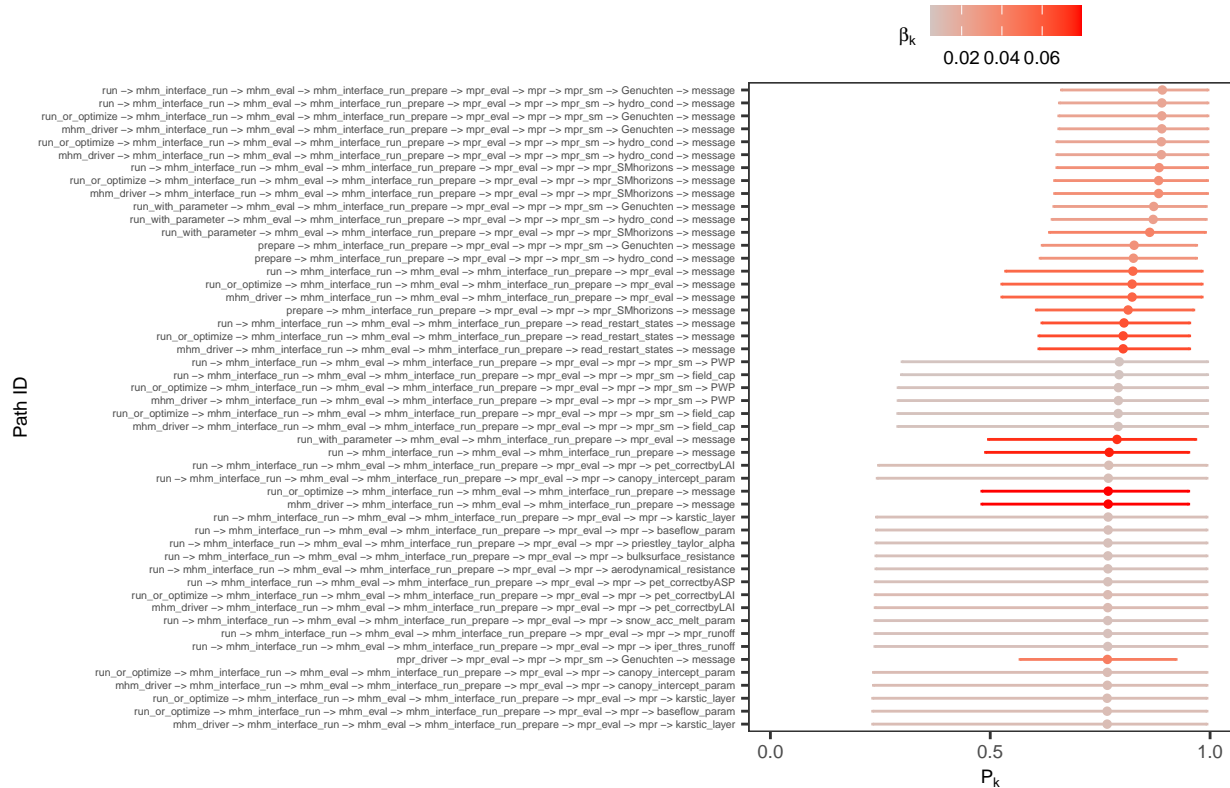
```
## [[8]]
```

```
## `height` was translated to `width`.
```

$$\beta_k$$
[illegible]

```
## `height` was translated to `width`.
```

MHM



```
## `height` was translated to `width`.
```

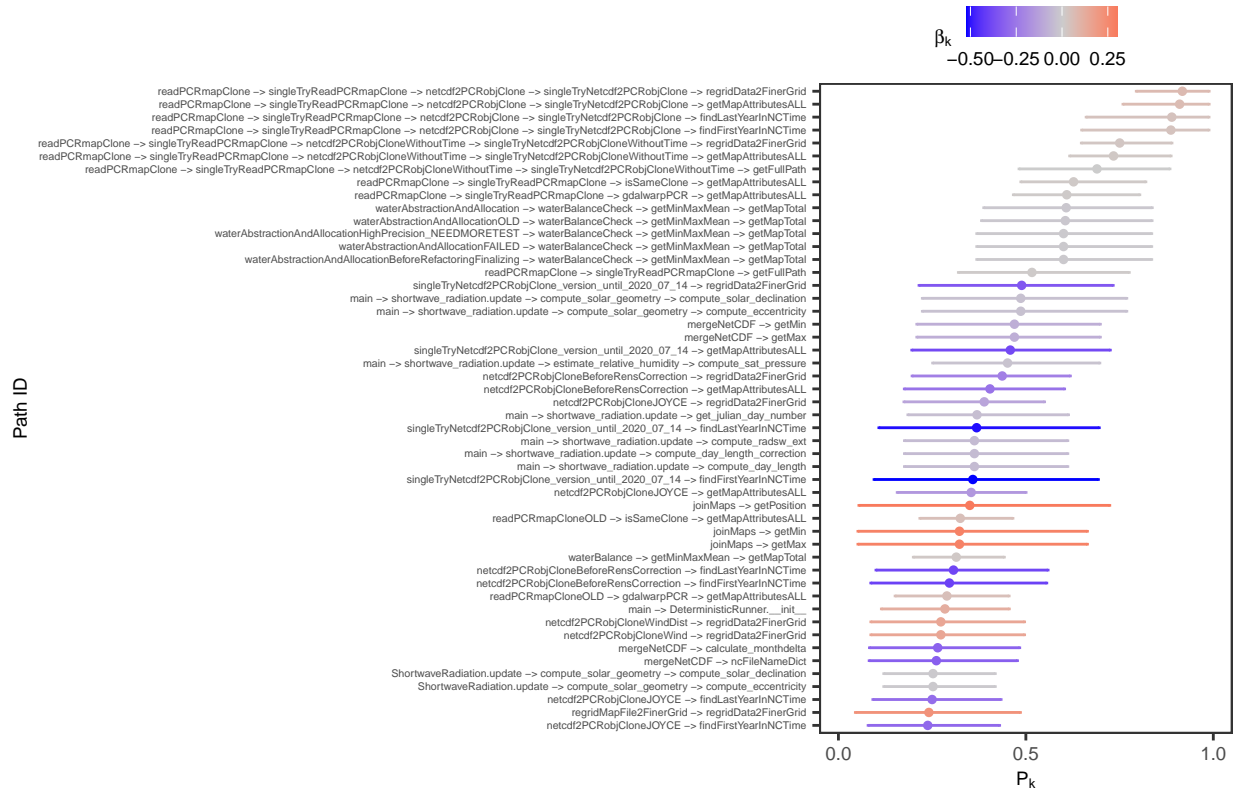
β_k 0.00

##

```
## `height` was translated to `width`.
```

$$P_k$$

PCR-GLOBWB

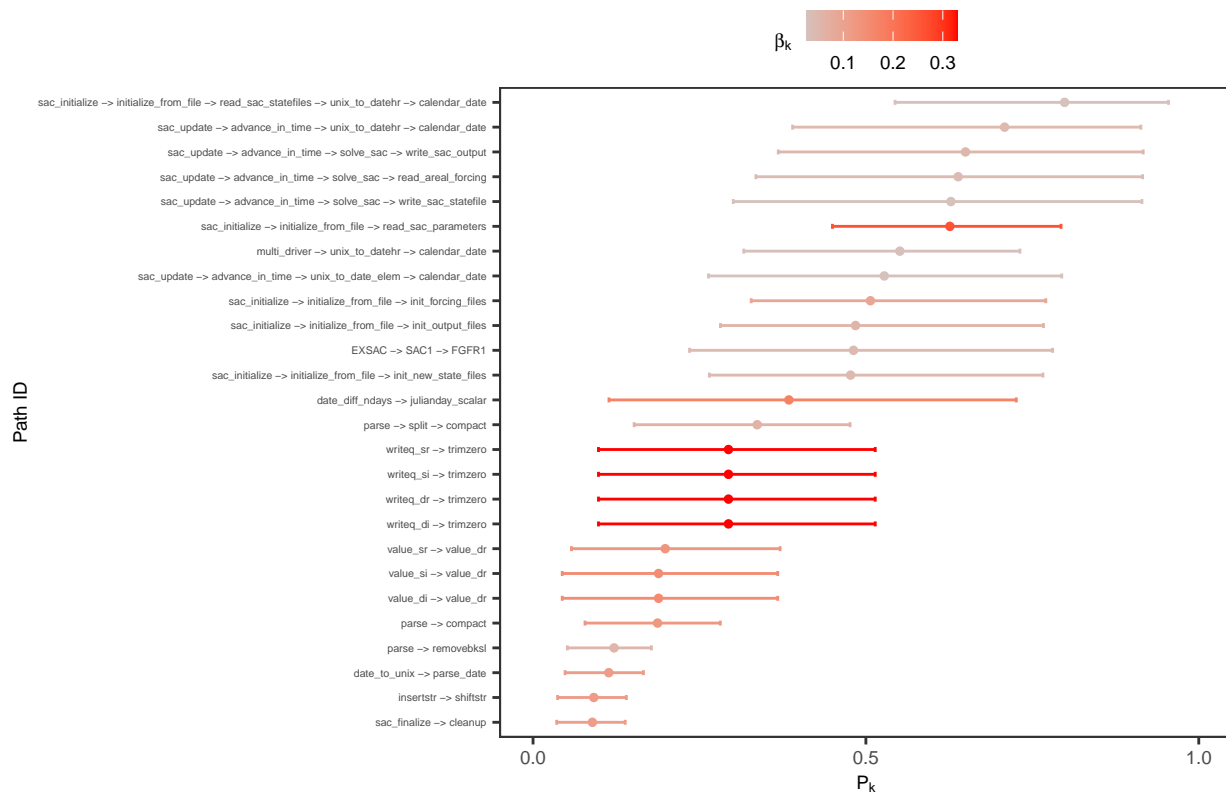


##

[[12]]

`height` was translated to `width`.

SACRAMENTO

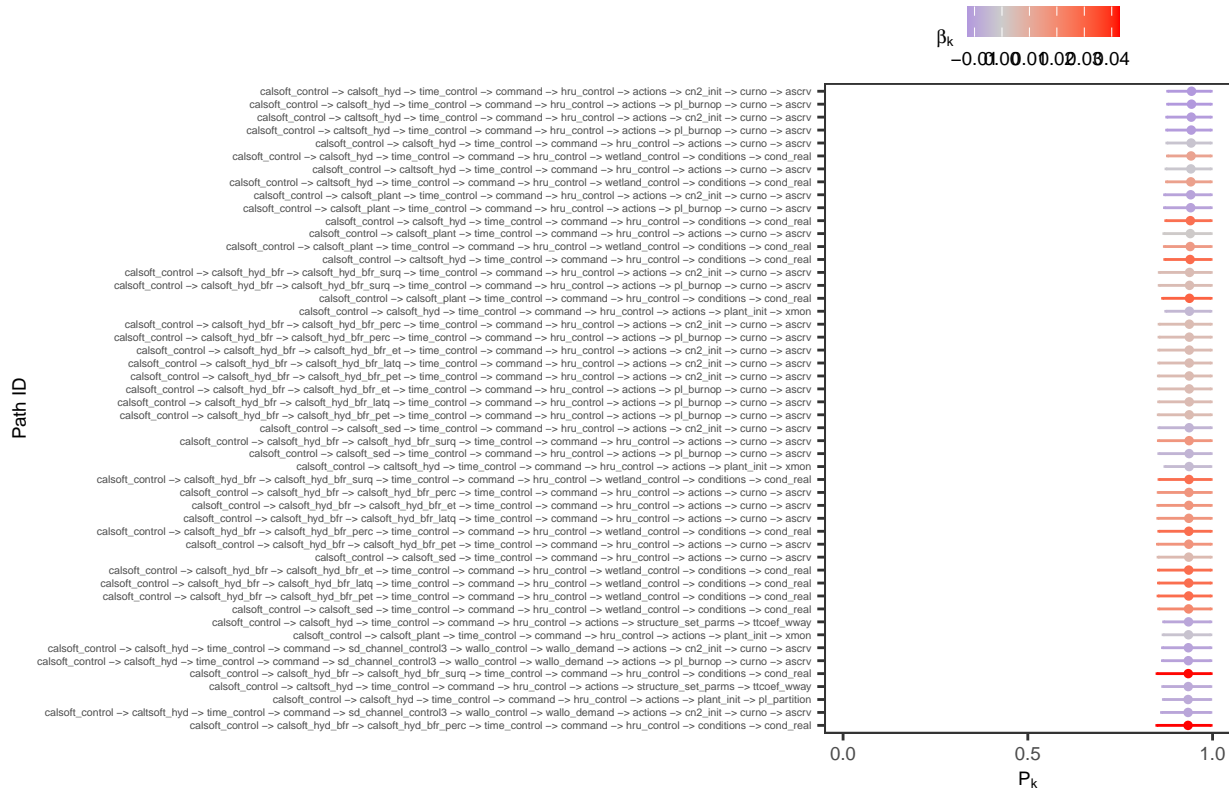


##

[[13]]

`height` was translated to `width`.

SWAT



```
# READ RANKING OF THE UA / SA DATASET #####

top_ten_overlap <- readRDS("top_ten_overlap.rds")

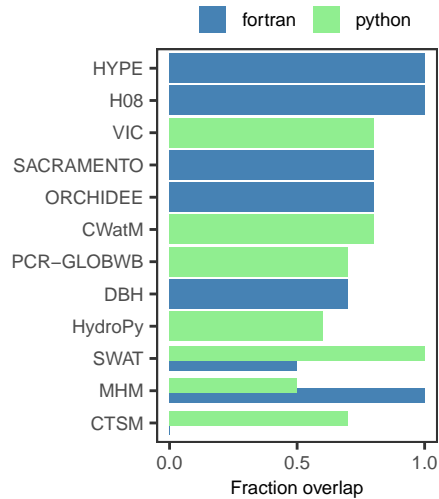
# ORDER AND FILTER OUT MODELS WITH LESS THAN 10 PATHS #####

model_names_ordered <- top_ten_overlap %>%
  .[n_paths > 10] %>%
  .[order(overlap_fraction)] %>%
  .[, model] %>%
  unique()

# PLOT #####

top_ten_overlap %>%
  .[n_paths > 10] %>%
  .[, model := factor(model, levels = model_names_ordered)] %>%
  ggplot(. , aes(model, overlap_fraction, fill = language)) +
  geom_bar(stat = "identity", position = position_dodge(0.6)) +
  scale_fill_manual(values = color_languages, name = "") +
  coord_flip() +
  scale_y_continuous(breaks = breaks_pretty(n = 3)) +
  labs(x = "", y = "Fraction overlap") +
```

```
theme_AP() +
theme(legend.position = "top")
```



```
# METRICS AT THE FILE AND FUNCTION LEVEL #####
```

```
folder <- "./datasets/git_logs"
```

```
# Get names of files -----
```

```
csv_files <- list.files(path = folder, pattern = "\\*.csv$", full.names = TRUE)
```

```
# Merge and flatten -----
```

```
logs_dt <- lapply(csv_files, fread) %>%
```

```
  lapply(., function(x) x[, .(model, language, `function`, cyclomatic_complexity, number_changes,
                             change_per_days, age_days)]) %>%
```

```
  rbindlist() %>%
```

```
  na.omit() %>%
```

```
  setnames(., "function", "node")
```

```
# Create and run function -----
```

```
spearman_fun <- function(dt) {
```

```
  cor.test(dt[, cyclomatic_complexity],
```

```
           dt[, number_changes], method = "spearman")$estimate[[1]]
```

```
}
```

```
rho_values <- logs_dt[, .(rho = spearman_fun(.SD)), model]
```

```
## Warning in cor.test.default(dt[, cyclomatic_complexity], dt[, number_changes],
```

```
## : Cannot compute exact p-value with ties
```

```
## Warning in cor.test.default(dt[, cyclomatic_complexity], dt[, number_changes],
```



```

## : Cannot compute exact p-value with ties
## Warning in cor.test.default(dt[, cyclomatic_complexity], dt[, number_changes],
## : Cannot compute exact p-value with ties
## Warning in cor.test.default(dt[, cyclomatic_complexity], dt[, number_changes],
## : Cannot compute exact p-value with ties

# Create dt to position labels -----

pos_df <- logs_dt %>%
  filter(is.finite(cyclomatic_complexity), is.finite(number_changes)) %>%
  group_by(model) %>%
  summarise(x = min(cyclomatic_complexity, na.rm = TRUE),
            y = max(number_changes, na.rm = TRUE), .groups = "drop")

# join rho and make label text -----

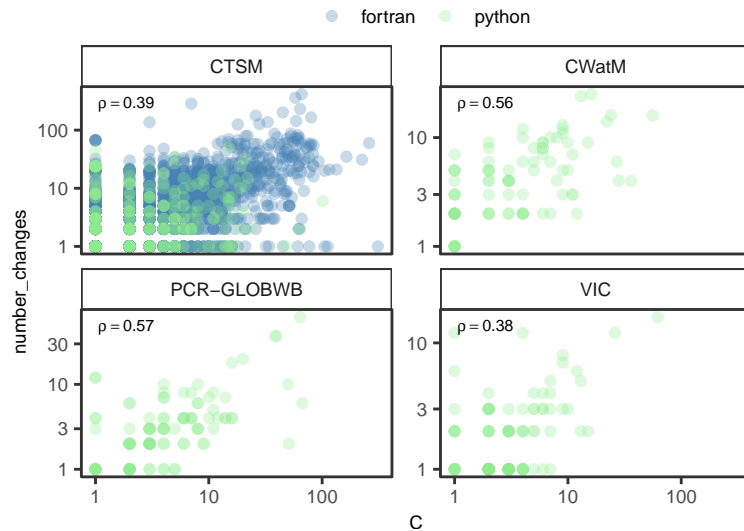
lab_df <- rho_values %>%
  left_join(pos_df, by = "model") %>%
  mutate(label = ifelse(is.na(rho), "rho==NA", paste0("rho==", round(rho, 2))))

# Plot -----

plot_logs <- logs_dt %>%
  ggplot(aes(cyclomatic_complexity, number_changes, color = language)) +
  geom_point(alpha = 0.3) +
  scale_color_manual(values = color_languages, name = "") +
  scale_x_log10() +
  scale_y_log10() +
  labs(x = expression(C), y = "number_changes") +
  facet_wrap(~model, scales = "free_y") +
  geom_text(data = lab_df %>% na.omit(),
            aes(x = x, y = y, label = label), inherit.aes = FALSE,
            hjust = -0.05, vjust = 1.2, size = 2, parse = TRUE) +
  coord_cartesian(clip = "off") +
  theme_AP() +
  theme(legend.position = "top")

plot_logs

```



ARE REFACTORING ACTIONS MORE COMMON IN RISKY PATHS?

```
models_to_check <- unique(logs_dt$model)
```

Extract name of functions in top ten risky paths per model -----

```
list_nodes <- full_paths_df %>%
  data.table() %>%
  .[order(-p_path_fail), .SD[1:10], model] %>%
  .[model %in% models_to_check] %>%
  na.omit() %>%
  .[, .(node = unlist(path_nodes)), model]
```

keep only unique risk nodes

```
risk_nodes <- unique(list_nodes[, .(model, node)])
risk_nodes[, high_risk_path := TRUE]
```

Filter out and keep only models that have log data -----

```
logs_dt <- logs_dt[model %in% models_to_check]
```

left join into logs_dt -----

```
logs_dt[risk_nodes, high_risk_path := i.high_risk_path,
  on = .(model, node)]
```

Turn NAs into FALSE -----

```
logs_dt[is.na(high_risk_path), high_risk_path := FALSE]
```

*# We now quantify effect size using the common-language statistic A,
defined as the probability that a randomly selected high-risk function*

has higher churn than a randomly selected non-risk function. -----

```
wilcox_dt <- logs_dt[is.finite(number_changes),
  {
    x <- number_changes[high_risk_path]
    y <- number_changes[!high_risk_path]

    n_x <- length(x)
    n_y <- length(y)

    if (n_x >= 3L && n_y >= 3L) {

      wt <- wilcox.test(x, y, alternative = "greater", exact = FALSE)

      # Wilcoxon rank-sum statistic (sum of ranks for x)-----
      W <- as.numeric(wt$statistic)

      # Convert to Mann-Whitney U -----
      U <- W - n_x * (n_x + 1) / 2

      # Common-language effect size (A statistic) -----
      A <- U / (n_x * n_y)

      .(
        p_value = wt$p.value,
        A_common_language = A,
        n_risk = n_x,
        n_nonrisk = n_y,
        median_risk = median(x),
        median_nonrisk = median(y)
      )

    } else {

      .(
        p_value = NA_real_,
        A_common_language = NA_real_,
        n_risk = n_x,
        n_nonrisk = n_y,
        median_risk = if (n_x) median(x) else NA_real_,
        median_nonrisk = if (n_y) median(y) else NA_real_
      )
    }
  },
model
```

]

```
wilcox_dt
```

```
##      model      p_value A_common_language n_risk n_nonrisk median_risk
##      <char>      <num>      <num>      <int>      <int>      <num>
## 1:      CTSM 0.9999999998      0.2263026      48      2044      1.0
## 2:      CWatM 0.0013898071      0.6284314      15      68      6.0
## 3: PCR-GLOBWB 0.0006109416      0.5789773      22      80      7.5
## 4:      VIC      NA      NA      0      101      NA
##      median_nonrisk
##      <num>
## 1:      4.5
## 2:      4.0
## 3:      3.0
## 4:      2.0
```

```
# WHICH PATHS IMPROVE IF A NODE'S RISK IS FIXED TO 0? #####
```

```
# how many nodes and paths to show in the heatmap -----
```

```
number_of_interest <- 30
```

```
N_nodes <- number_of_interest
```

```
K_paths <- number_of_interest
```

```
list_plots <- list_nodes <- list()
```

```
for (i in 1:nrow(all_graphs)) {
```

```
  if (i == 7) next
```

```
  model.name <- all_graphs$node_df[[i]]$model
```

```
  node_df <- all_graphs$node_df[[i]]
```

```
  paths_tbl <- all_graphs$paths_tbl[[i]]
```

```
# top-N nodes by failure probability -----
```

```
top_nodes <- node_df %>%
  arrange(desc(risk_score)) %>%
  slice_head(n = N_nodes)
```

```
# Return list of nodes -----
```

```
list_nodes[[i]] <- top_nodes %>%
  transmute(model = model,
            name = name,
            risk_score = risk_score,
            cyclomatic_complexity = cyclomatic_complexity,
```

```

        indegree = indeg,
        outdegree = outdeg,
        betweenness = btw) %>%
data.table::as.data.table()

# top-K paths by path failure probability -----

top_paths <- paths_tbl %>%
  arrange(desc(p_path_fail)) %>%
  slice_head(n = K_paths)

# COMPUTE #####

# make a named vector for fast lookup of risk_score by node name -----

p_map <- setNames(node_df$risk_score, node_df$name)

# Compute -----

delta_tbl <- map_dfr(seq_len(nrow(top_nodes)), function(i) {
  node_name <- top_nodes$name[i]

  map_dfr(seq_len(nrow(top_paths)), function(j) {
    pid <- top_paths$path_id[j]
    nodes_vec <- top_paths$path_nodes[[j]]
    P_orig <- top_paths$p_path_fail[j]

    # if node not in this path, improvement is zero -----

    if (!node_name %in% nodes_vec) {
      tibble(node = node_name,
              path_id = pid,
              deltaP = 0)
    } else {

      # failure probs along the path -----

      p_vec <- p_map[nodes_vec]

      # if node appears multiple times, fixing ANY of its appearances is enough:

      idxs <- which(nodes_vec == node_name)

      # we fix all occurrences (set all to zero) -----

      p_vec_fix <- p_vec
      p_vec_fix[idxs] <- 0

```

```

    P_fix <- if (length(p_vec_fix) == 0) 0 else 1 - prod(1 - p_vec_fix)
    tibble(node = node_name,
           path_id = pid,
           deltaP = P_orig - P_fix)
  }
})
})

# order factors so tha highest risk nodes are at the top and the highest
# risk paths are on the left -----

# Bright cells: nodes that are chokepoints for a given path.
# Rows with many bright cells: nodes whose refactoring will improve many
# risk paths (global chokepoints)
# Columns with few very bright cells: paths dominated by a single risky node
# Rows/columns with dark colors: diffuse risk.

delta_tbl <- delta_tbl %>%
  mutate(node = factor(node, levels = rev(top_nodes$name)),
         path_id = factor(path_id, levels = top_paths$path_id))

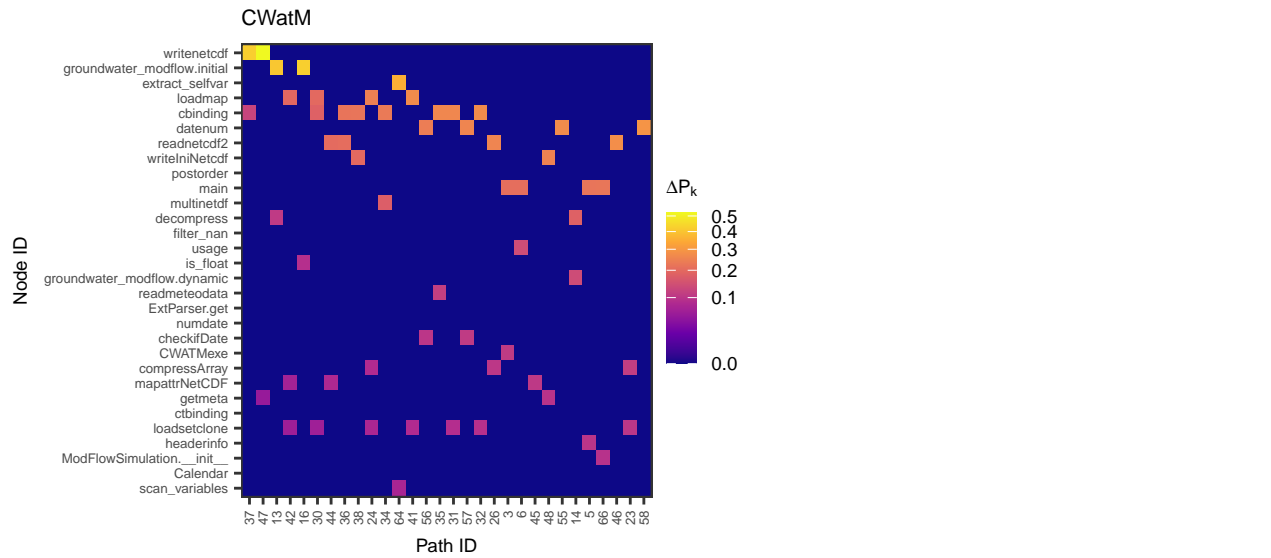
# Plote the tile figure -----

minP <- min(delta_tbl$deltaP, na.rm = TRUE)
medP <- median(delta_tbl$deltaP, na.rm = TRUE)
maxP <- max(delta_tbl$deltaP, na.rm = TRUE)

list_plots[[i]] <- ggplot(delta_tbl, aes(x = path_id, y = node, fill = deltaP)) +
  geom_tile() +
  scale_fill_viridis_c(option = "C",
                      name = expression(Delta * P[k]),
                      trans = "sqrt") +
  theme_AP() +
  theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust = 1, size = 5),
        axis.text.y = element_text(size = 5),
        axis.title.x = element_text(margin = margin(t = 5)),
        axis.title.y = element_text(margin = margin(r = 5))) +
  labs(x = "Path ID", y = "Node ID") +
  ggtitle(model.name)
}

for (i in 1:length(list_plots)) {
  print(list_plots[[i]])
  print(list_nodes[[i]])
}

```

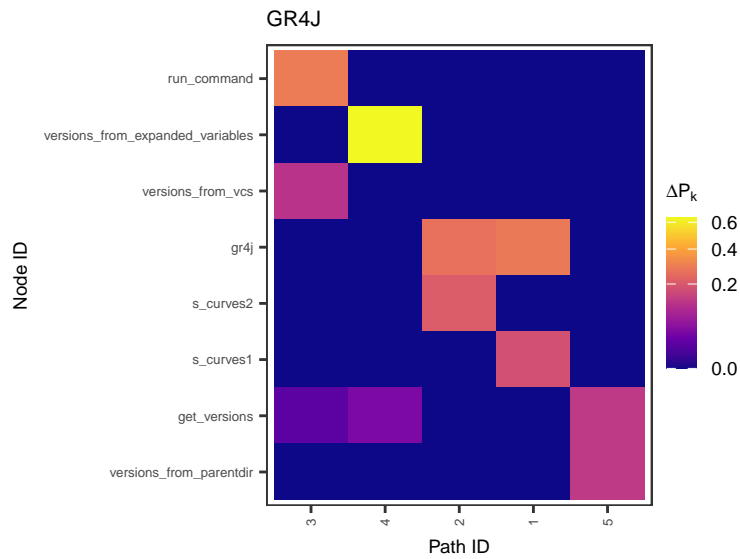


##	model	name	risk_score	cyclomatic_complexity
##	<char>	<char>	<num>	<num>
## 1:	CWatM	writenetcdf	0.6000000	56.0000
## 2:	CWatM	groundwater_modflow.initial	0.5127273	48.0000
## 3:	CWatM	extract_selfvar	0.3976077	36.0000
## 4:	CWatM	loadmap	0.3264593	22.0000
## 5:	CWatM	cbinding	0.3071770	6.0000
## 6:	CWatM	datenum	0.3000000	1.0000
## 7:	CWatM	readnetcdf2	0.2945455	28.0000
## 8:	CWatM	writeIniNetcdf	0.2836364	27.0000
## 9:	CWatM	postorder	0.2609569	9.0000
## 10:	<NA>	main	0.2520201	13.4878
## 11:	CWatM	multinetdf	0.2509091	24.0000
## 12:	CWatM	decompress	0.2173206	5.0000
## 13:	CWatM	filter_nan	0.1894737	1.0000
## 14:	<NA>	usage	0.1835990	13.4878
## 15:	<NA>	is_float	0.1835990	13.4878
## 16:	CWatM	groundwater_modflow.dynamic	0.1745455	17.0000
## 17:	CWatM	readmeteodata	0.1636364	16.0000
## 18:	CWatM	ExtParser.get	0.1613397	10.0000
## 19:	CWatM	numdate	0.1578947	1.0000
## 20:	CWatM	checkifDate	0.1527273	15.0000
## 21:	CWatM	CWATMexe	0.1500478	8.0000
## 22:	CWatM	compressArray	0.1432536	4.0000
## 23:	CWatM	mapattrNetCDF	0.1424880	2.0000
## 24:	CWatM	getmeta	0.1323445	3.0000
## 25:	CWatM	ctbinding	0.1323445	3.0000
## 26:	CWatM	loadsetclone	0.1309091	13.0000
## 27:	CWatM	headerinfo	0.1297608	10.0000
## 28:	CWatM	ModFlowSimulation.__init__	0.1248804	11.0000

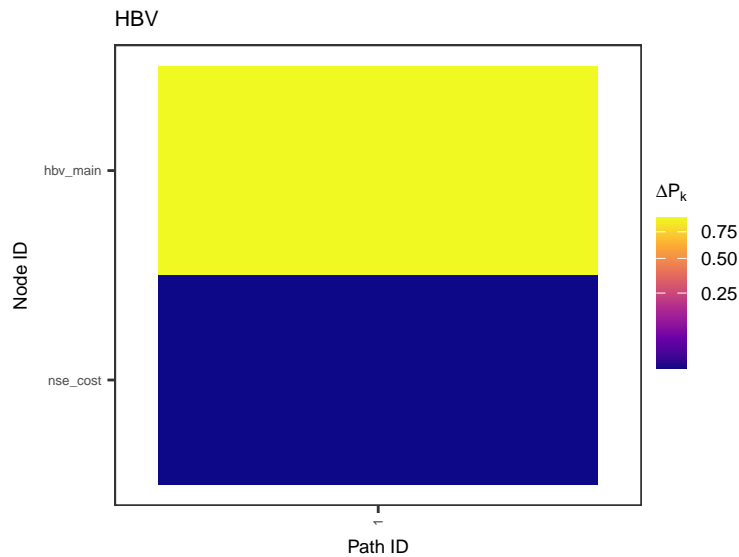
```

## 29:  CWatM          Calendar  0.1237321          8.0000
## 30:  CWatM          scan_variables  0.1200000        12.0000
##      model          name risk_score cyclomatic_complexity
##      <char>          <char>      <num>          <num>
##      indegree outdegree betweenness
##      <num>      <num>      <num>
##  1:      0          6          0.00
##  2:      0          8          0.00
##  3:      1          0          0.00
##  4:      3          5          4.75
##  5:     16          0          0.00
##  6:     19          0          0.00
##  7:      0          4          0.00
##  8:      0          5          0.00
##  9:     11          8          0.00
## 10:      1         10          9.50
## 11:      0          1          0.00
## 12:     11          0          0.00
## 13:     12          0          0.00
## 14:      3          0          0.00
## 15:      3          0          0.00
## 16:      0          3          0.00
## 17:      0          3          0.00
## 18:      4          0          0.00
## 19:     10          0          0.00
## 20:      0         23          0.00
## 21:      3          1          2.50
## 22:      7          0          0.00
## 23:      3          1          8.00
## 24:      7          0          0.00
## 25:      7          0          0.00
## 26:      0         12          0.00
## 27:      2          0          0.00
## 28:      1          0          0.00
## 29:      3          0          0.00
## 30:      0          1          0.00
##      indegree outdegree betweenness
##      <num>      <num>      <num>

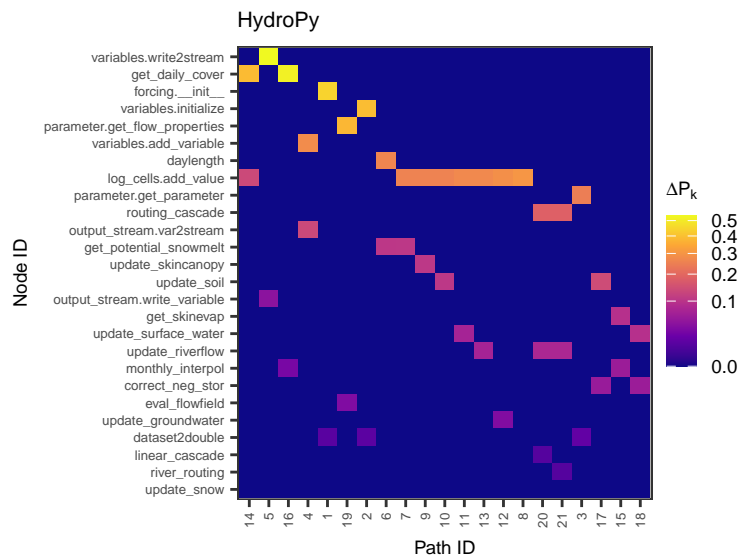
```

##	model	name	risk_score	cyclomatic_complexity
##	<char>	<char>	<num>	<int>
## 1:	GR4J	run_command	0.75	12
## 2:	GR4J	versions_from_expanded_variables	0.75	15
## 3:	GR4J	versions_from_vcs	0.55	9
## 4:	GR4J	gr4j	0.40	11
## 5:	GR4J	s_curves2	0.35	4
## 6:	GR4J	s_curves1	0.30	3
## 7:	GR4J	get_versions	0.15	6
## 8:	GR4J	versions_from_parentdir	0.15	3
##	indegree	outdegree	betweenness	
##	<num>	<num>	<num>	
## 1:	2	0	0	
## 2:	1	0	0	
## 3:	1	2	1	
## 4:	0	4	0	
## 5:	2	0	0	
## 6:	2	0	0	
## 7:	0	3	0	
## 8:	1	0	0	



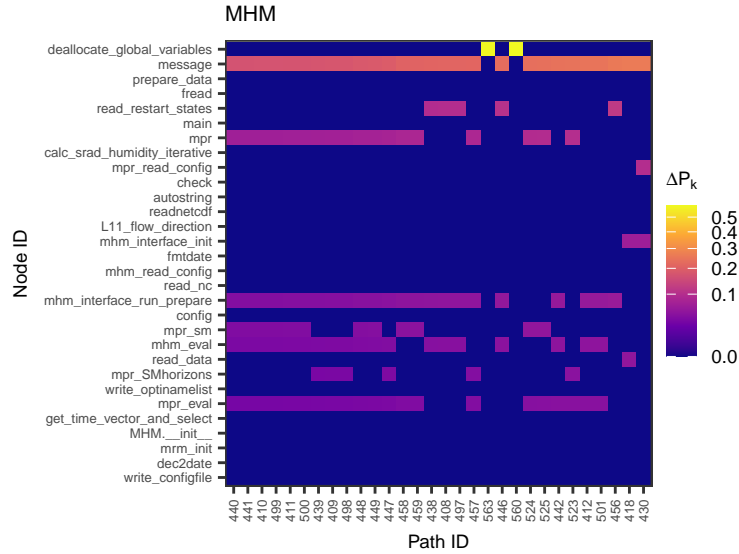
```
##      model      name risk_score cyclomatic_complexity indegree outdegree
##      <char>      <char>      <num>                <int>      <num>      <num>
## 1:    HBV hbv_main          0.95                    5          1          0
## 2:    HBV nse_cost          0.05                    1          0          1
##      betweenness
##      <num>
## 1:          0
## 2:          0
```



```
##      model      name risk_score cyclomatic_complexity
##      <char>      <char>      <num>                <num>
## 1: HydroPy      variables.write2stream 0.6000000      15.000000
## 2: HydroPy      get_daily_cover 0.5500000      14.000000
## 3: HydroPy      forcing.__init__ 0.4500000      12.000000
## 4: HydroPy      variables.initialize 0.4000000      11.000000
## 5: HydroPy      parameter.get_flow_properties 0.4000000      11.000000
```

## 6: HydroPy	variables.add_variable	0.3500000	10.000000
## 7: <NA>	daylength	0.3064583	8.979167
## 8: HydroPy	log_cells.add_value	0.3000000	3.000000
## 9: HydroPy	parameter.get_parameter	0.2500000	8.000000
## 10: HydroPy	routing_cascade	0.2075000	5.000000
## 11: HydroPy	output_stream.var2stream	0.2075000	7.000000
## 12: HydroPy	get_potential_snowmelt	0.1500000	6.000000
## 13: HydroPy	update_skincanopy	0.1500000	6.000000
## 14: HydroPy	update_soil	0.1500000	6.000000
## 15: HydroPy	output_stream.write_variable	0.1075000	5.000000
## 16: HydroPy	get_skinevap	0.1000000	5.000000
## 17: HydroPy	update_surface_water	0.1000000	5.000000
## 18: HydroPy	update_riverflow	0.1000000	5.000000
## 19: HydroPy	monthly_interpol	0.0650000	4.000000
## 20: HydroPy	correct_neg_stor	0.0650000	4.000000
## 21: HydroPy	eval_flowfield	0.0575000	4.000000
## 22: HydroPy	update_groundwater	0.0500000	4.000000
## 23: HydroPy	dataset2double	0.0225000	3.000000
## 24: HydroPy	linear_cascade	0.0150000	3.000000
## 25: HydroPy	river_routing	0.0150000	3.000000
## 26: HydroPy	update_snow	0.0000000	3.000000
##	model	name	risk_score cyclomatic_complexity
##	<char>	<char>	<num> <num>
##	indegree	outdegree	betweenness
##	<num>	<num>	<num>
## 1:	0	1	0
## 2:	0	2	0
## 3:	0	1	0
## 4:	0	1	0
## 5:	0	1	0
## 6:	0	1	0
## 7:	1	0	0
## 8:	40	0	0
## 9:	0	1	0
## 10:	1	4	2
## 11:	1	0	0
## 12:	0	3	0
## 13:	0	10	0
## 14:	0	7	0
## 15:	1	0	0
## 16:	0	1	0
## 17:	0	7	0
## 18:	0	5	0
## 19:	2	0	0
## 20:	2	0	0
## 21:	1	0	0
## 22:	0	3	0
## 23:	3	0	0

```
## 24:      2      0      0
## 25:      2      0      0
## 26:      0      8      0
##      indegree outdegree betweenness
##      <num>      <num>      <num>
```

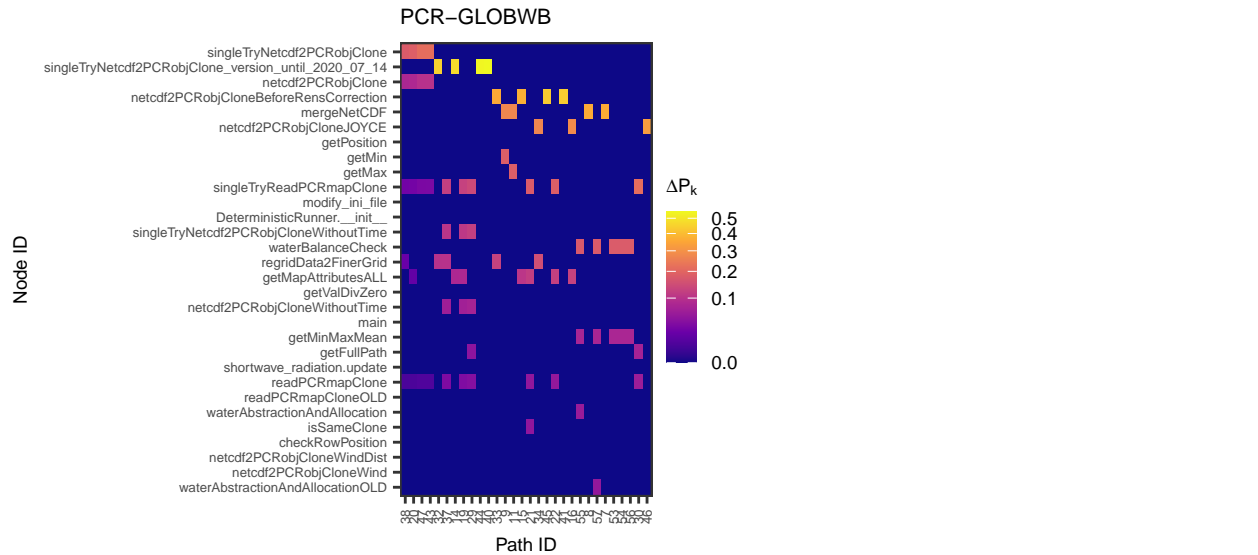


```
##      model      name risk_score cyclomatic_complexity
##      <char>      <char>      <num>      <num>
## 1:  MHM  deallocate_global_variables 0.60196813      201
## 2:  MHM      message 0.37200000      25
## 3:  MHM  prepare_data 0.26100000      88
## 4:  MHM      fread 0.24600000      83
## 5:  MHM  read_restart_states 0.21389286      72
## 6:  MHM      main 0.19500000      66
## 7:  MHM      mpr 0.19429071      35
## 8:  MHM  calc_srad_humidity_iterative 0.18689286      63
## 9:  MHM      mpr_read_config 0.18178571      61
## 10: MHM      check 0.17360714      5
## 11: MHM      autostring 0.16889286      57
## 12: MHM      readnetcdf 0.16297005      53
## 13: MHM      L11_flow_direction 0.13298246      45
## 14: MHM      mhm_interface_init 0.13245104      21
## 15: MHM      fmtdate 0.13200000      45
## 16: MHM      mhm_read_config 0.12850576      43
## 17: MHM      read_nc 0.12834562      27
## 18: MHM  mhm_interface_run_prepare 0.12465668      14
## 19: MHM      config 0.12300000      42
## 20: MHM      mpr_sm 0.11754877      32
## 21: MHM      mhm_eval 0.10778571      3
## 22: MHM      read_data 0.10685023      25
## 23: MHM      mpr_SMhorizons 0.10043049      34
## 24: MHM      write_optinamelist 0.09989286      34
```

```

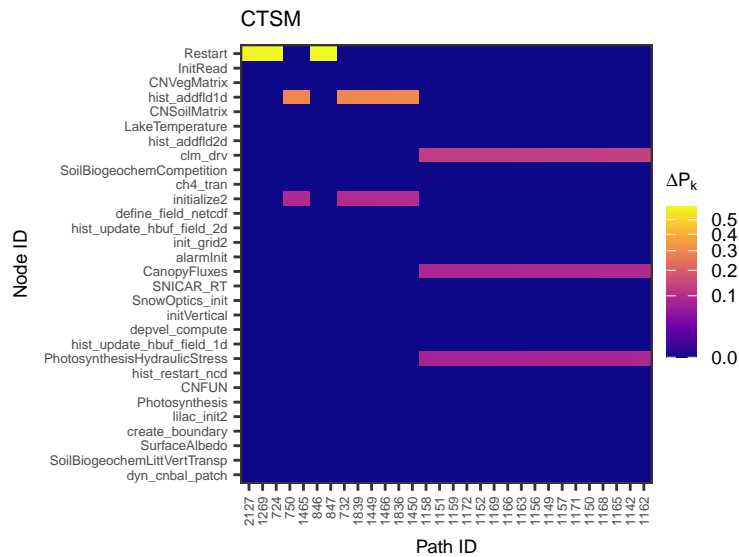
## 25:    MHM                                mpr_eval 0.09788249                4
## 26:    MHM    get_time_vector_and_select 0.09705415                18
## 27:    MHM                                MHM.__init__ 0.09000000        31
## 28:    MHM                                mrm_init 0.09000000         31
## 29:    MHM                                dec2date 0.08941475          4
## 30:    MHM                                write_configfile 0.08489286      29
##      model                                name risk_score cyclomatic_complexity
##      <char>                                <char>          <num>              <num>
##      indegree outdegree betweenness
##      <num>      <num>          <num>
## 1:      1          1      2.0000000
## 2:     336          0      0.0000000
## 3:      0         18      0.0000000
## 4:      0         16      0.0000000
## 5:      1          1      0.0000000
## 6:      0          4      0.0000000
## 7:      1         14 170.0000000
## 8:      1          0      0.0000000
## 9:      2          4      0.0000000
## 10:     181          0      0.0000000
## 11:      1          0      0.0000000
## 12:      6         34      3.0000000
## 13:      1          5      0.1666667
## 14:      2         23 131.4375000
## 15:      0         12      0.0000000
## 16:      1         35      3.0000000
## 17:      4          1  87.0000000
## 18:      2          5 156.0000000
## 19:      0          1      0.0000000
## 20:      1         14  44.0000000
## 21:      2          9 186.0000000
## 22:      2         25  61.5000000
## 23:      1          2      1.0000000
## 24:      1          2      0.0000000
## 25:      2          3 162.0000000
## 26:      1          8  84.0000000
## 27:      0          3      0.0000000
## 28:      0         38      0.0000000
## 29:     16          3 123.0000000
## 30:      1          4      0.0000000
##      indegree outdegree betweenness
##      <num>      <num>          <num>

```



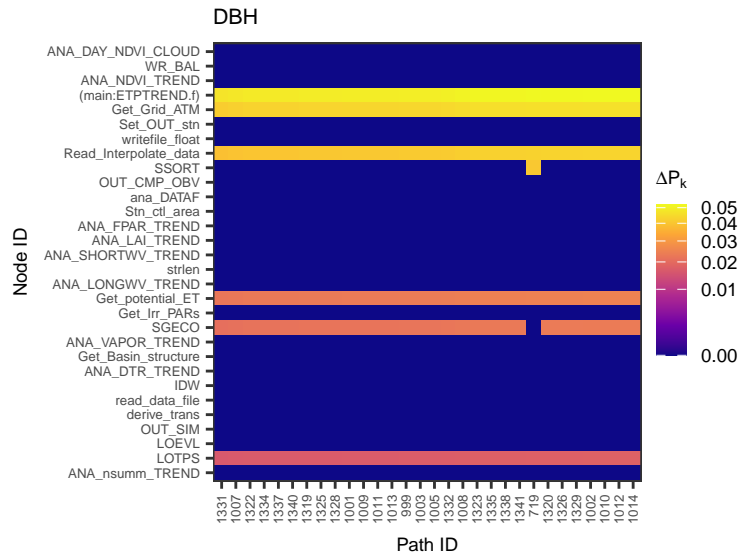
##	model	name	risk_score
##	<char>	<char>	<num>
## 1:	PCR-GLOBWB	singleTryNetcdf2PCRobjClone	0.70681818
## 2:	PCR-GLOBWB	singleTryNetcdf2PCRobjClone_version_until_2020_07_14	0.57272727
## 3:	PCR-GLOBWB	netcdf2PCRobjClone	0.52159091
## 4:	PCR-GLOBWB	netcdf2PCRobjCloneBeforeRensCorrection	0.45454545
## 5:	PCR-GLOBWB	mergeNetCDF	0.38181818
## 6:	PCR-GLOBWB	netcdf2PCRobjCloneJOYCE	0.34545455
## 7:	PCR-GLOBWB	getPosition	0.31818182
## 8:	PCR-GLOBWB	getMin	0.29318182
## 9:	PCR-GLOBWB	getMax	0.29318182
## 10:	PCR-GLOBWB	singleTryReadPCRmapClone	0.26136364
## 11:	PCR-GLOBWB	modify_ini_file	0.24090909
## 12:	PCR-GLOBWB	DeterministicRunner.__init__	0.23863636
## 13:	PCR-GLOBWB	singleTryNetcdf2PCRobjCloneWithoutTime	0.23409091
## 14:	PCR-GLOBWB	waterBalanceCheck	0.22954545
## 15:	PCR-GLOBWB	regridData2FinerGrid	0.22727273
## 16:	PCR-GLOBWB	getMapAttributesALL	0.18977273
## 17:	PCR-GLOBWB	getValDivZero	0.17500000
## 18:	PCR-GLOBWB	netcdf2PCRobjCloneWithoutTime	0.15795455
## 19:	<NA>	main	0.12413163
## 20:	PCR-GLOBWB	getMinMaxMean	0.11590909
## 21:	PCR-GLOBWB	getFullPath	0.10454545
## 22:	PCR-GLOBWB	shortwave_radiation.update	0.10340909
## 23:	PCR-GLOBWB	readPCRmapClone	0.09090909
## 24:	PCR-GLOBWB	readPCRmapCloneOLD	0.09090909
## 25:	PCR-GLOBWB	waterAbstractionAndAllocation	0.09090909
## 26:	PCR-GLOBWB	isSameClone	0.08863636
## 27:	PCR-GLOBWB	checkRowPosition	0.08409091
## 28:	PCR-GLOBWB	netcdf2PCRobjCloneWindDist	0.08181818
## 29:	PCR-GLOBWB	netcdf2PCRobjCloneWind	0.08181818
## 30:	PCR-GLOBWB	waterAbstractionAndAllocationOLD	0.08181818

##	model				name risk_score
##	<char>				<char> <num>
##	cyclomatic_complexity	indegree	outdegree	betweenness	
##	<num>	<num>	<num>	<num>	
## 1:	67.00000	2	6	9	
## 2:	64.00000	0	6	0	
## 3:	50.00000	1	2	7	
## 4:	51.00000	0	5	0	
## 5:	43.00000	0	40	0	
## 6:	39.00000	0	5	0	
## 7:	3.00000	24	0	0	
## 8:	3.00000	22	0	0	
## 9:	3.00000	22	0	0	
## 10:	16.00000	2	5	11	
## 11:	22.00000	4	0	0	
## 12:	8.00000	14	0	0	
## 13:	20.00000	2	5	4	
## 14:	8.00000	6	1	10	
## 15:	4.00000	16	0	0	
## 16:	4.00000	13	0	0	
## 17:	1.00000	14	0	0	
## 18:	14.00000	1	2	3	
## 19:	14.65448	0	28	0	
## 20:	5.00000	2	1	6	
## 21:	7.00000	4	0	0	
## 22:	2.00000	1	6	9	
## 23:	11.00000	0	2	0	
## 24:	11.00000	0	3	0	
## 25:	11.00000	0	3	0	
## 26:	6.00000	2	2	2	
## 27:	2.00000	6	0	0	
## 28:	10.00000	0	2	0	
## 29:	10.00000	0	2	0	
## 30:	10.00000	0	5	0	
##	cyclomatic_complexity	indegree	outdegree	betweenness	
##	<num>	<num>	<num>	<num>	
## NULL					
## NULL					



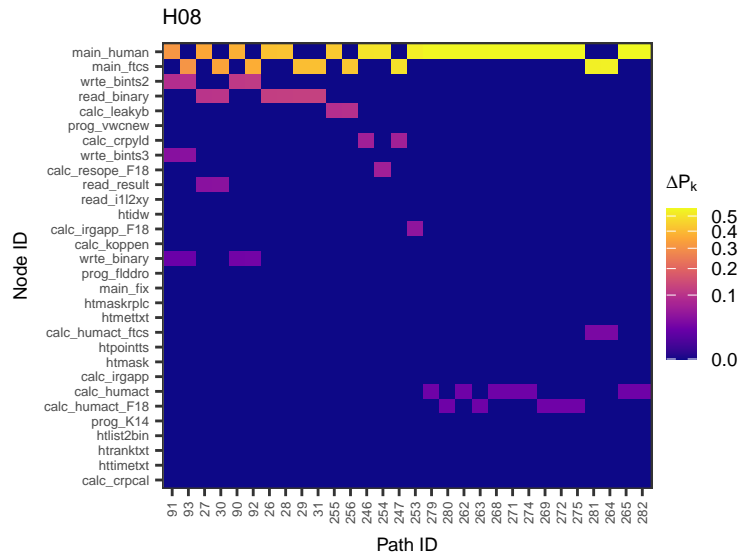
##	model	name	risk_score	cyclomatic_complexity
##	<char>	<char>	<num>	<int>
## 1:	CTSM	Restart	0.6000000	310
## 2:	CTSM	InitRead	0.5029126	260
## 3:	CTSM	CNVegMatrix	0.4355878	225
## 4:	CTSM	hist_addfld1d	0.4237577	63
## 5:	CTSM	CNSoilMatrix	0.3168444	164
## 6:	CTSM	LakeTemperature	0.2597794	134
## 7:	CTSM	hist_addfld2d	0.2307528	105
## 8:	CTSM	clm_drv	0.2265958	66
## 9:	CTSM	SoilBiogeochemCompetition	0.2240858	116
## 10:	CTSM	ch4_tran	0.2024723	105
## 11:	CTSM	initialize2	0.1914262	64
## 12:	CTSM	define_field_netcdf	0.1868026	97
## 13:	CTSM	hist_update_hbuf_field_2d	0.1795740	93
## 14:	CTSM	init_grid2	0.1747573	91
## 15:	CTSM	alarmInit	0.1712560	89
## 16:	CTSM	CanopyFluxes	0.1707223	80
## 17:	CTSM	SNICAR_RT	0.1617279	83
## 18:	CTSM	SnowOptics_init	0.1613561	84
## 19:	CTSM	initVertical	0.1613561	84
## 20:	CTSM	depvel_compute	0.1613561	84
## 21:	CTSM	hist_update_hbuf_field_1d	0.1595628	83
## 22:	CTSM	PhotosynthesisHydraulicStress	0.1586358	80
## 23:	CTSM	hist_restart_ncd	0.1573533	79
## 24:	CTSM	CNFUN	0.1484003	77
## 25:	CTSM	Photosynthesis	0.1480913	76
## 26:	CTSM	lilac_init2	0.1447588	74
## 27:	CTSM	create_boundary	0.1436893	75
## 28:	CTSM	SurfaceAlbedo	0.1398363	72
## 29:	CTSM	SoilBiogeochemLittVertTransp	0.1344685	70
## 30:	CTSM	dyn_cnbal_patch	0.1341717	70

##	model	name	risk_score	cyclomatic_complexity
##	<char>	<char>	<num>	<int>
##	indegree	outdegree	betweenness	
##	<num>	<num>	<num>	
##	1:	0	8	0.000000
##	2:	0	2	0.000000
##	3:	1	23	3.000000
##	4:	1570	3	22.700061
##	5:	1	30	1.000000
##	6:	1	4	9.000000
##	7:	142	3	11.299939
##	8:	2	184	673.714286
##	9:	1	6	4.000000
##	10:	2	2	1.000000
##	11:	2	93	462.935829
##	12:	1	35	1.372881
##	13:	1	2	5.000000
##	14:	0	18	0.000000
##	15:	2	0	0.000000
##	16:	1	32	115.428571
##	17:	10	4	4.000000
##	18:	1	1	0.000000
##	19:	1	2	0.000000
##	20:	1	0	0.000000
##	21:	1	1	1.000000
##	22:	1	4	34.000000
##	23:	3	15	35.866667
##	24:	2	9	3.000000
##	25:	2	2	14.000000
##	26:	1	9	19.000000
##	27:	0	4	0.000000
##	28:	1	14	12.000000
##	29:	1	1	2.000000
##	30:	1	0	0.000000
##	indegree	outdegree	betweenness	
##	<num>	<num>	<num>	



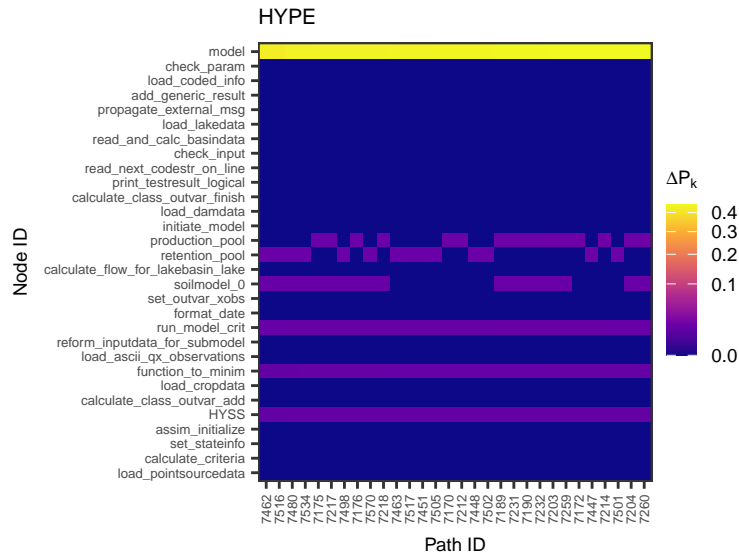
##	model	name	risk_score	cyclomatic_complexity	indegree
##	<char>	<char>	<num>	<num>	<num>
## 1:	DBH	ANA_DAY_NDVI_CLOUD	0.6000000	73	0
## 2:	DBH	WR_BAL	0.4446982	54	1
## 3:	DBH	ANA_NDVI_TREND	0.3666667	45	0
## 4:	DBH	(main:ETPTREND.f)	0.3583333	44	0
## 5:	DBH	Get_Grid_ATM	0.3312000	25	13
## 6:	DBH	Set_OUT_stn	0.3193265	39	1
## 7:	DBH	writefile_float	0.3166667	3	125
## 8:	DBH	Read_Interpolate_data	0.3151339	27	1
## 9:	DBH	SSORT	0.3025789	37	1
## 10:	DBH	OUT_CMP_OBV	0.2666667	33	0
## 11:	DBH	ana_DATAF	0.2421333	5	87
## 12:	DBH	Stn_ctl_area	0.2258489	26	7
## 13:	DBH	ANA_FPAR_TREND	0.2201400	27	1
## 14:	DBH	ANA_LAI_TREND	0.2201400	27	1
## 15:	DBH	ANA_SHORTWV_TREND	0.2120155	25	2
## 16:	DBH	strlen	0.2098000	4	77
## 17:	DBH	ANA_LONGWV_TREND	0.2096302	25	2
## 18:	DBH	Get_potential_ET	0.2087837	23	3
## 19:	DBH	Get_Irr_PARs	0.2026309	25	1
## 20:	DBH	SGECO	0.1990909	23	1
## 21:	DBH	ANA_VAPOR_TREND	0.1921883	23	2
## 22:	DBH	Get_Basin_structure	0.1778857	22	1
## 23:	DBH	ANA_DTR_TREND	0.1746124	21	1
## 24:	DBH	IDW	0.1745669	19	8
## 25:	DBH	read_data_file	0.1731813	21	1
## 26:	DBH	derive_trans	0.1693052	21	1
## 27:	DBH	OUT_SIM	0.1692559	21	1
## 28:	DBH	LOEVL	0.1690667	21	1
## 29:	DBH	LOTPS	0.1615691	11	8
## 30:	DBH	ANA_nsumm_TREND	0.1614932	20	1

##	model	name	risk_score	cyclomatic_complexity	indegree
##	<char>	<char>	<num>	<num>	<num>
##	outdegree	betweenness			
##	<num>	<num>			
##	1:	36	0.000000		
##	2:	55	3.530030		
##	3:	6	0.000000		
##	4:	50	0.000000		
##	5:	24	558.983471		
##	6:	8	1.452381		
##	7:	0	0.000000		
##	8:	11	537.000000		
##	9:	2	1.000000		
##	10:	20	0.000000		
##	11:	0	0.000000		
##	12:	2	4.000000		
##	13:	11	6.000000		
##	14:	11	6.000000		
##	15:	15	40.333333		
##	16:	0	0.000000		
##	17:	14	27.000000		
##	18:	9	102.016529		
##	19:	34	1.290909		
##	20:	10	74.666667		
##	21:	11	22.666667		
##	22:	8	2.715152		
##	23:	8	31.000000		
##	24:	1	30.000000		
##	25:	3	23.000000		
##	26:	1	1.333333		
##	27:	5	1.057692		
##	28:	0	0.000000		
##	29:	9	330.000000		
##	30:	8	4.247639		
##	outdegree	betweenness			
##	<num>	<num>			



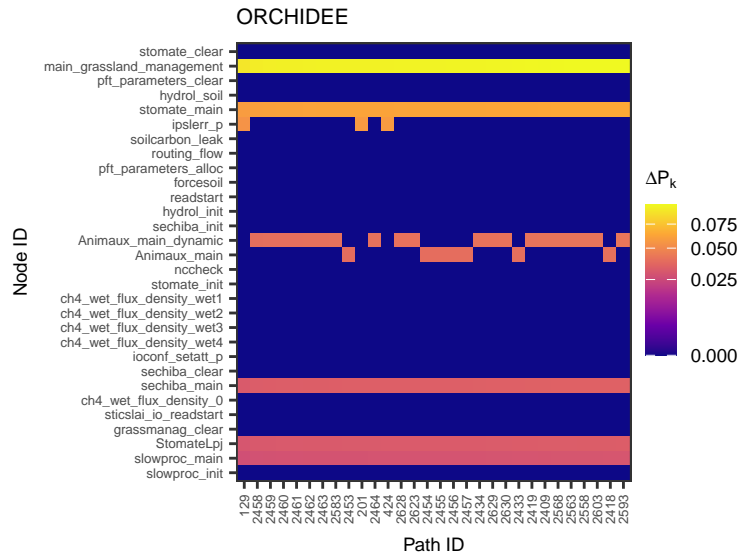
##	model	name	risk_score	cyclomatic_complexity	indegree	outdegree
##	<char>	<char>	<num>	<num>	<num>	<num>
## 1:	H08	main_human	0.60000000	300	0	377
## 2:	H08	main_ftcs	0.58996656	295	0	198
## 3:	H08	wrte_bints2	0.30693331	45	225	4
## 4:	H08	read_binary	0.30200669	2	356	0
## 5:	H08	calc_leakyb	0.24587925	115	5	1
## 6:	H08	prog_vwcnew	0.21672241	109	0	20
## 7:	H08	calc_crpild	0.16591259	82	4	0
## 8:	H08	wrte_bints3	0.16557439	18	156	2
## 9:	H08	calc_resope_F18	0.16539119	83	1	0
## 10:	H08	read_result	0.15398089	17	114	8
## 11:	H08	read_i1l2xy	0.14362012	3	47	2
## 12:	H08	htidw	0.13645485	69	0	7
## 13:	H08	calc_irgapp_F18	0.12124403	61	1	0
## 14:	H08	calc_koppen	0.11036789	56	0	4
## 15:	H08	wrte_binary	0.09522472	1	113	0
## 16:	H08	prog_flddro	0.08829431	45	0	7
## 17:	H08	main_fix	0.08428094	43	0	65
## 18:	H08	htmaskrplc	0.08227425	42	0	4
## 19:	H08	htmettxt	0.08026756	41	0	2
## 20:	H08	calc_humact_ftcs	0.07882306	27	1	27
## 21:	H08	htpointts	0.07625418	39	0	2
## 22:	H08	htmask	0.07424749	38	0	4
## 23:	H08	calc_irgapp	0.07191951	36	2	0
## 24:	H08	calc_humact	0.06319678	25	1	27
## 25:	H08	calc_humact_F18	0.06262282	26	1	27
## 26:	H08	prog_K14	0.06220736	32	0	6
## 27:	H08	htlist2bin	0.06020067	31	0	3
## 28:	H08	htranktxt	0.06020067	31	0	2
## 29:	H08	httimetxt	0.06020067	31	0	4
## 30:	H08	calc_crpcal	0.05819398	30	0	13

##	model	name	risk_score	cyclomatic_complexity	indegree	outdegree
##	<char>	<char>	<num>	<num>	<num>	<num>
##	betweenness					
##	<num>					
##	1:		0.0			
##	2:		0.0			
##	3:		9.0			
##	4:		0.0			
##	5:		4.0			
##	6:		0.0			
##	7:		0.0			
##	8:		0.0			
##	9:		0.0			
##	10:		8.0			
##	11:		31.0			
##	12:		0.0			
##	13:		0.0			
##	14:		0.0			
##	15:		0.0			
##	16:		0.0			
##	17:		0.0			
##	18:		0.0			
##	19:		0.0			
##	20:		8.0			
##	21:		0.0			
##	22:		0.0			
##	23:		0.0			
##	24:		4.4			
##	25:		3.6			
##	26:		0.0			
##	27:		0.0			
##	28:		0.0			
##	29:		0.0			
##	30:		0.0			
##	betweenness					
##	<num>					



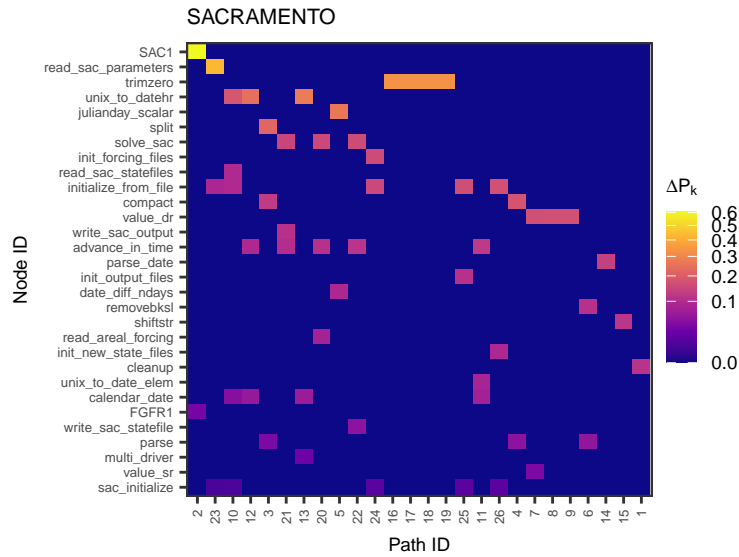
##	model	name	risk_score	cyclomatic_complexity
##	<char>	<char>	<num>	<int>
## 1:	HYPE	model	0.70652174	937
## 2:	HYPE	check_param	0.30571902	9
## 3:	HYPE	load_coded_info	0.26148852	406
## 4:	HYPE	add_generic_result	0.20269783	3
## 5:	HYPE	propagate_external_msg	0.20011706	10
## 6:	HYPE	load_lakedata	0.19377020	301
## 7:	HYPE	read_and_calc_basindata	0.17569388	273
## 8:	HYPE	check_input	0.13000780	4
## 9:	HYPE	read_next_codestr_on_line	0.12243032	17
## 10:	HYPE	print_testresult_logical	0.11540134	4
## 11:	HYPE	calculate_class_outvar_finish	0.11345596	3
## 12:	HYPE	load_damdata	0.09496386	147
## 13:	HYPE	initiate_model	0.09495934	125
## 14:	HYPE	production_pool	0.09260870	1
## 15:	HYPE	retention_pool	0.09064103	2
## 16:	HYPE	calculate_flow_for_lakebasin_lake	0.08848927	130
## 17:	HYPE	soilmodel_0	0.08524710	96
## 18:	HYPE	set_outvar_xobs	0.08345596	3
## 19:	HYPE	format_date	0.08334448	13
## 20:	HYPE	run_model_crit	0.08068341	9
## 21:	HYPE	reform_inputdata_for_submodel	0.07952274	123
## 22:	HYPE	load_ascii_qx_observations	0.07860718	121
## 23:	HYPE	function_to_minim	0.07821127	7
## 24:	HYPE	load_cropdata	0.07766727	120
## 25:	HYPE	calculate_class_outvar_add	0.07757525	4
## 26:	HYPE	HYSS	0.07500000	118
## 27:	HYPE	assim_initialize	0.07150415	109
## 28:	HYPE	set_stateinfo	0.06732999	104
## 29:	HYPE	calculate_criteria	0.06281391	64
## 30:	HYPE	load_pointsourcedata	0.06114986	94

##	model	name	risk_score	cyclomatic_complexity
##	<char>	<char>	<num>	<int>
##	indegree	outdegree	betweenness	
##	<num>	<num>	<num>	
##	1:	5	266	3771.000000
##	2:	230	1	22.279642
##	3:	1	159	21.449006
##	4:	154	1	20.597727
##	5:	149	0	0.000000
##	6:	1	29	5.964286
##	7:	1	13	1.152381
##	8:	98	1	9.753023
##	9:	86	0	0.000000
##	10:	87	0	0.000000
##	11:	86	0	0.000000
##	12:	1	15	2.630952
##	13:	6	16	288.333333
##	14:	71	0	0.000000
##	15:	69	0	0.000000
##	16:	1	16	169.416667
##	17:	6	47	623.104094
##	18:	63	0	0.000000
##	19:	58	0	0.000000
##	20:	1	17	2800.000000
##	21:	1	1	0.500000
##	22:	1	33	14.320635
##	23:	7	1	2460.000000
##	24:	1	10	3.049570
##	25:	58	0	0.000000
##	26:	0	136	0.000000
##	27:	1	63	36.542169
##	28:	1	0	0.000000
##	29:	7	83	501.500000
##	30:	1	25	8.678226
##	indegree	outdegree	betweenness	
##	<num>	<num>	<num>	



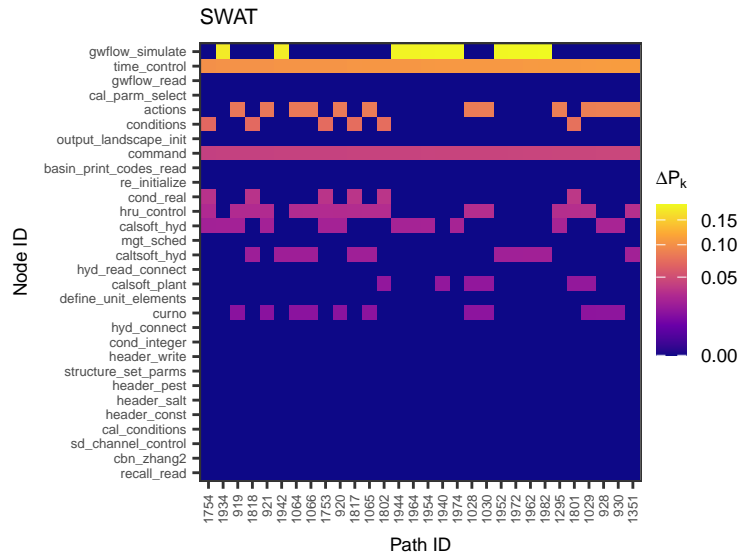
##	model	name	risk_score	cyclomatic_complexity
##	<char>	<char>	<num>	<int>
## 1:	ORCHIDEE	stomate_clear	0.6015020	499
## 2:	ORCHIDEE	main_grassland_management	0.4181497	318
## 3:	ORCHIDEE	pft_parameters_clear	0.4105702	341
## 4:	ORCHIDEE	hydrol_soil	0.3546497	290
## 5:	ORCHIDEE	stomate_main	0.3177095	193
## 6:	ORCHIDEE	ipslerr_p	0.3024096	3
## 7:	ORCHIDEE	soilcarbon_leak	0.2799343	231
## 8:	ORCHIDEE	routing_flow	0.2726577	226
## 9:	ORCHIDEE	pft_parameters_alloc	0.2611726	217
## 10:	ORCHIDEE	forcesoil	0.2481928	207
## 11:	ORCHIDEE	readstart	0.2382811	198
## 12:	ORCHIDEE	hydrol_init	0.2363009	196
## 13:	ORCHIDEE	sechiba_init	0.2347023	195
## 14:	ORCHIDEE	Animaux_main_dynamic	0.2316875	173
## 15:	ORCHIDEE	Animaux_main	0.2244936	182
## 16:	ORCHIDEE	nccheck	0.2146302	2
## 17:	ORCHIDEE	stomate_init	0.2104348	174
## 18:	ORCHIDEE	ch4_wet_flux_density_wet1	0.2069558	172
## 19:	ORCHIDEE	ch4_wet_flux_density_wet2	0.2069558	172
## 20:	ORCHIDEE	ch4_wet_flux_density_wet3	0.2069558	172
## 21:	ORCHIDEE	ch4_wet_flux_density_wet4	0.2069558	172
## 22:	ORCHIDEE	ioconf_setatt_p	0.2024471	2
## 23:	ORCHIDEE	sechiba_clear	0.2012048	168
## 24:	ORCHIDEE	sechiba_main	0.2001123	81
## 25:	ORCHIDEE	ch4_wet_flux_density_0	0.1985220	165
## 26:	ORCHIDEE	sticslai_io_readstart	0.1961124	163
## 27:	ORCHIDEE	grassmanag_clear	0.1937741	161
## 28:	ORCHIDEE	StomateLpj	0.1918230	105
## 29:	ORCHIDEE	slowproc_main	0.1773529	78
## 30:	ORCHIDEE	slowproc_init	0.1750749	134

##	model	name	risk_score	cyclomatic_complexity
##	<char>	<char>	<num>	<int>
##	indegree	outdegree	betweenness	
##	<num>	<num>	<num>	
## 1:	1	4	8.00000	
## 2:	1	14	495.00000	
## 3:	1	0	0.00000	
## 4:	1	24	77.50000	
## 5:	1	28	1198.60000	
## 6:	322	0	0.00000	
## 7:	2	3	13.50000	
## 8:	1	6	9.00000	
## 9:	1	0	0.00000	
## 10:	0	107	0.00000	
## 11:	1	3	0.00000	
## 12:	1	58	6.02381	
## 13:	1	1	0.50000	
## 14:	1	49	330.00000	
## 15:	1	12	77.00000	
## 16:	229	1	1.00000	
## 17:	1	2	15.00000	
## 18:	1	0	0.00000	
## 19:	1	0	0.00000	
## 20:	1	0	0.00000	
## 21:	1	0	0.00000	
## 22:	216	0	0.00000	
## 23:	0	10	0.00000	
## 24:	4	18	1402.65000	
## 25:	1	0	0.00000	
## 26:	1	0	0.00000	
## 27:	1	1	1.00000	
## 28:	1	36	920.00000	
## 29:	2	12	1160.25000	
## 30:	1	79	195.00000	
##	indegree	outdegree	betweenness	
##	<num>	<num>	<num>	



##	model	name	risk_score	cyclomatic_complexity	indegree
##	<char>	<char>	<num>	<num>	<num>
## 1:	SACRAMENTO	SAC1	0.66428571	29.000000	1
## 2:	SACRAMENTO	read_sac_parameters	0.54285714	24.000000	1
## 3:	SACRAMENTO	trimzero	0.32857143	7.000000	4
## 4:	SACRAMENTO	unix_to_datehr	0.31428571	3.000000	4
## 5:	SACRAMENTO	julianday_scalar	0.30000000	1.000000	6
## 6:	SACRAMENTO	split	0.28571429	12.000000	1
## 7:	SACRAMENTO	solve_sac	0.20000000	4.000000	1
## 8:	SACRAMENTO	init_forcing_files	0.20000000	8.000000	1
## 9:	SACRAMENTO	read_sac_statefiles	0.19285714	5.000000	1
## 10:	SACRAMENTO	initialize_from_file	0.19285714	3.000000	1
## 11:	SACRAMENTO	compact	0.18571429	5.000000	2
## 12:	SACRAMENTO	value_dr	0.17142857	2.000000	3
## 13:	SACRAMENTO	write_sac_output	0.15714286	6.000000	1
## 14:	SACRAMENTO	advance_in_time	0.15000000	1.000000	1
## 15:	SACRAMENTO	parse_date	0.13571429	5.000000	1
## 16:	SACRAMENTO	init_output_files	0.13571429	5.000000	1
## 17:	SACRAMENTO	date_diff_ndays	0.12857143	7.000000	0
## 18:	SACRAMENTO	removebksl	0.11428571	4.000000	1
## 19:	SACRAMENTO	shiftstr	0.11428571	4.000000	1
## 20:	SACRAMENTO	read_areal_forcing	0.11428571	4.000000	1
## 21:	SACRAMENTO	init_new_state_files	0.11428571	4.000000	1
## 22:	<NA>	cleanup	0.11055901	3.826087	1
## 23:	SACRAMENTO	unix_to_date_elem	0.10714286	3.000000	1
## 24:	SACRAMENTO	calendar_date	0.10000000	1.000000	2
## 25:	SACRAMENTO	FGFR1	0.09285714	3.000000	1
## 26:	SACRAMENTO	write_sac_statefile	0.07142857	2.000000	1
## 27:	<NA>	parse	0.06055901	3.826087	0
## 28:	SACRAMENTO	multi_driver	0.04285714	3.000000	0
## 29:	SACRAMENTO	value_sr	0.04285714	3.000000	0
## 30:	SACRAMENTO	sac_initialize	0.02142857	2.000000	0

##	model	name	risk_score	cyclomatic_complexity	indegree
##	<char>	<char>	<num>	<num>	<num>
##	outdegree	betweenness			
##	<num>	<num>			
##	1:	1		1	
##	2:	0		0	
##	3:	0		0	
##	4:	1		5	
##	5:	0		0	
##	6:	1		0	
##	7:	3		6	
##	8:	0		0	
##	9:	1		4	
##	10:	5		7	
##	11:	0		0	
##	12:	0		0	
##	13:	0		0	
##	14:	3		7	
##	15:	0		0	
##	16:	0		0	
##	17:	6		0	
##	18:	0		0	
##	19:	0		0	
##	20:	0		0	
##	21:	0		0	
##	22:	0		0	
##	23:	1		1	
##	24:	0		0	
##	25:	0		0	
##	26:	0		0	
##	27:	3		0	
##	28:	2		0	
##	29:	1		0	
##	30:	1		0	
##	outdegree	betweenness			
##	<num>	<num>			



##	model	name	risk_score	cyclomatic_complexity	indegree
##	<char>	<char>	<num>	<int>	<num>
## 1:	SWAT	gwflow_simulate	0.60955084	445	1
## 2:	SWAT	time_control	0.47567568	57	42
## 3:	SWAT	gwflow_read	0.42741313	312	1
## 4:	SWAT	cal_parm_select	0.42247597	250	12
## 5:	SWAT	actions	0.41668101	229	12
## 6:	SWAT	conditions	0.38678632	212	14
## 7:	SWAT	output_landscape_init	0.28011583	203	1
## 8:	SWAT	command	0.27139271	134	1
## 9:	SWAT	basin_print_codes_read	0.25444015	184	1
## 10:	SWAT	re_initialize	0.24961390	6	34
## 11:	SWAT	cond_real	0.23320463	15	30
## 12:	SWAT	hru_control	0.20981355	115	1
## 13:	SWAT	calsoft_hyd	0.17357035	122	1
## 14:	SWAT	mgt_sched	0.16674556	108	3
## 15:	SWAT	caltsoft_hyd	0.15977874	112	1
## 16:	SWAT	hyd_read_connect	0.13169463	35	12
## 17:	SWAT	calsoft_plant	0.12818783	89	1
## 18:	SWAT	define_unit_elements	0.12606178	15	15
## 19:	SWAT	curno	0.11663466	2	16
## 20:	SWAT	hyd_connect	0.11621622	87	0
## 21:	SWAT	cond_integer	0.11177606	15	13
## 22:	SWAT	header_write	0.10984556	77	1
## 23:	SWAT	structure_set_parms	0.10607235	26	10
## 24:	SWAT	header_pest	0.10444015	73	1
## 25:	SWAT	header_salt	0.10444015	73	1
## 26:	SWAT	header_const	0.10444015	73	1
## 27:	SWAT	cal_conditions	0.10190945	71	1
## 28:	SWAT	sd_channel_control	0.09324324	70	0
## 29:	SWAT	cbn_zhang2	0.09279399	64	1
## 30:	SWAT	recall_read	0.08687259	60	1

```
##      model      name risk_score cyclomatic_complexity indegree
##      <char>      <char>      <num>      <int>      <num>
##      outdegree betweenness
##      <num>      <num>
##  1:      5      70.00000
##  2:     13     2907.00000
##  3:      0      0.00000
##  4:     13      8.00000
##  5:     40     664.50000
##  6:     43     48.00000
##  7:      0      0.00000
##  8:     46     2457.00000
##  9:      0      0.00000
## 10:      0      0.00000
## 11:      0      0.00000
## 12:     67     1413.28571
## 13:     27      84.70924
## 14:     33     21.00000
## 15:     14     76.62500
## 16:      1      1.00000
## 17:     23     61.80435
## 18:      0      0.00000
## 19:      1     29.00000
## 20:     22      0.00000
## 21:      0      0.00000
## 22:      0      0.00000
## 23:      1     25.00000
## 24:      0      0.00000
## 25:      0      0.00000
## 26:      0      0.00000
## 27:     12      5.00000
## 28:     16      0.00000
## 29:      8     15.00000
## 30:      0      0.00000
##      outdegree betweenness
##      <num>      <num>
```

```
# FUNCTIONS TO SELECT THE TOP TEN RISKY PATHS PER MODEL AND
# PRINT THEM OUT FOR LATEX #####
```

```
tmp <- full_paths_df[order(-p_path_fail), .SD[1:10], model] %>%
  .[, .(model, path_str)] %>%
  na.omit() %>%
  split(., .$model)

to_tex_list_fun(tmp)
```

5 Session information

```
# SESSION INFORMATION #####
```

```
sessionInfo()
```

```
## R version 4.5.2 (2025-10-31)
## Platform: aarch64-apple-darwin20
## Running under: macOS Sequoia 15.6.1
##
## Matrix products: default
## BLAS: /System/Library/Frameworks/Accelerate.framework/Versions/A/Frameworks/vecLib.framework
## LAPACK: /Library/Frameworks/R.framework/Versions/4.5-arm64/Resources/lib/libRlapack.dylib;
##
## locale:
## [1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
##
## time zone: Europe/London
## tzcode source: internal
##
## attached base packages:
## [1] tools parallel stats graphics grDevices utils datasets
## [8] methods base
##
## other attached packages:
## [1] benchmarkme_1.0.8 sensobol_1.1.6 ggraph_2.2.2 foreach_1.5.2
## [5] igraph_2.2.1 tidygraph_1.3.1 here_1.0.2 tidytext_0.4.3
## [9] ggrepel_0.9.6 readxl_1.4.5 cowplot_1.2.0.9000 scales_1.4.0
## [13] openxlsx_4.2.8 lubridate_1.9.4 forcats_1.0.1 stringr_1.6.0
## [17] dplyr_1.1.4 purrr_1.2.0 readr_2.1.5 tidyr_1.3.2
## [21] tibble_3.3.0 ggplot2_4.0.1.9000 tidyverse_2.0.0 data.table_1.18.0
##
## loaded via a namespace (and not attached):
## [1] tidyselect_1.2.1 viridisLite_0.4.2 farver_2.1.2
## [4] viridis_0.6.5 S7_0.2.1 fastmap_1.2.0
## [7] tweenr_2.0.3 janeaustenr_1.0.0 digest_0.6.39
## [10] timechange_0.3.0 lifecycle_1.0.4 tokenizers_0.3.0
## [13] magrittr_2.0.4 compiler_4.5.2 rlang_1.1.6
## [16] yaml_2.3.12 knitr_1.51 labeling_0.4.3
## [19] graphlayouts_1.2.2 RColorBrewer_1.1-3 withr_3.0.2
## [22] grid_4.5.2 polyclip_1.10-7 iterators_1.0.14
## [25] MASS_7.3-65 tinytex_0.58 cli_3.6.5
## [28] crayon_1.5.3 rmarkdown_2.30 generics_0.1.4
## [31] RcppParallel_5.1.11-1 rstudioapi_0.17.1 httr_1.4.7
## [34] tzdb_0.5.0 cachem_1.1.0 ggforce_0.5.0
## [37] cellranger_1.1.0 vctrs_0.6.5 Matrix_1.7-4
## [40] hms_1.1.3 ineq_0.2-13 glue_1.8.0
## [43] benchmarkmeData_1.0.4 codetools_0.2-20 rngWELL_0.10-10
```

```
## [46] stringi_1.8.7      gtable_0.3.6      randtoolbox_2.0.5
## [49] pillar_1.11.1      htmltools_0.5.9   R6_2.6.1
## [52] zigg_0.0.2         Rdpack_2.6.4      doParallel_1.0.17
## [55] rprojroot_2.1.1    evaluate_1.0.5    lattice_0.22-7
## [58] rbibutils_2.4      SnowballC_0.7.1   Rfast_2.1.5.2
## [61] memoise_2.0.1      Rcpp_1.1.0        zip_2.3.3
## [64] gridExtra_2.3      xfun_0.55         pkgconfig_2.0.3
```

```
## Return the machine CPU -----
```

```
cat("Machine:      "); print(get_cpu()$model_name)
```

```
## Machine:
```

```
## [1] "Apple M1 Max"
```

```
## Return number of true cores -----
```

```
cat("Num cores:    "); print(detectCores(logical = FALSE))
```

```
## Num cores:
```

```
## [1] 10
```

```
## Return number of threads -----
```

```
cat("Num threads: "); print(detectCores(logical = FALSE))
```

```
## Num threads:
```

```
## [1] 10
```