

S-Plus workshop

7-9 and 14-16 January students.washington.edu/arnima/s

Syllabus

Tue 7 Introduction

Import data, summarize, regression, plots, export graphs

Wed 8 Basic statistics

Descriptive statistics, significance tests, linear models

Thu 9 Linear models

Anova, LM, GLM, loess

Tue 14 Graphics

Types, multipanel, export graphs

Wed 15 Data manipulation

Data objects, describe, extract, sort, manipulate

Thu 16 Programming

Functions, import/export, project management, packages



Today: Linear models

1 Object anatomy

Im, summary

2 Regression plots

plot, loess, boxplot, coplot, interaction.plot, diagnostic plots

3 Auxiliary functions

extract elements, build models, predict, diagnose, transform

4 Exercise

weight loss

Fetch data and create models

```
library(MASS)
#R: data(mammals, cabbages)
#S: mammals <- mammals
#S: cabbages <- cabbages
mammals.lm <- lm(log(brain)~log(body), data=mammals)
cabbages.aov <- aov(VitC~Cult+Date, data=cabbages)
cabbages.lm <- lm(VitC~HeadWt, data=cabbages)
cabbages.ancova <- lm(VitC~HeadWt+Cult*Date, data=cabbages)</pre>
```

Object anatomy - How they print

mammals.lm

```
Call:
lm(formula = log(brain) ~ log(body), data = mammals)

Coefficients:
(Intercept) log(body)
    2.1348    0.7517

#S: Degrees of freedom: 62 total; 60 residual
#S: Residual standard error: 0.6942947
```

Object anatomy - How they print

```
summary(mammals.lm)
#R: summary(mammals.lm, cor=T)
 Call:
 lm(formula = log(brain) ~ log(body), data = mammals)
 Residuals:
      Min
               10 Median
                           30
                                        Max
 -1.71550 -0.49228 -0.06162 0.43597 1.94829
 Coefficients:
             Estimate Std. Error t value Pr(>|t|)
 (Intercept) 2.13479 0.09604 22.23 <2e-16 ***
                        0.02846 26.41 <2e-16 ***
 log(body)
           0.75169
 Signif. codes: 0 `***' 0.001 `**' 0.01 `*' 0.05 `.' 0.1 ` ' 1
 Residual standard error: 0.6943 on 60 degrees of freedom
 Multiple R-Squared: 0.9208, Adjusted R-squared: 0.9195
 F-statistic: 697.4 on 1 and 60 DF, p-value: < 2.2e-16
 Correlation of Coefficients:
           (Intercept)
 log(body) - 0.3964
```



Object anatomy - What's inside

```
mammals.lm$call
mammals.lm$coe
mammals.lm$fit
mammals.lm$res
mammals.lm$rank
mammals.lm$df.res
```

Object anatomy - What's inside

```
names(summary(mammals.lm))

coefficients  # parameter estimates and t test of β=0
r.squared
correlation  # between parameter estimates

summary(mammals.lm)$coe

x <- summary(mammals.lm)
x$coe

x$r.s
x$cor</pre>
```

Symbols - Formula notation

Symbols - Formula notation

```
lm(y~1)  # estimate intercept only, null model
lm(y~-1+x)  # estimate slope, fix intercept at 0
lm(offset(y-3)~-1+x)  # estimate slope, fix intercept at 3
lm(y~offset(3*x))  # estimate intercept, fix slope at 3
?formula
```

Symbols - () [] { }

```
f(x)  # Pass argument x to function f

x[i]  # Extract element i from vector x

{cmd}  # Lump commands together as a block, used when programming
```

Regression plots

Scatterplot and friends

```
plot(log(mammals$body), log(mammals$brain))
abline(mammals.lm)
points(5, 0)
points(5, 0, cex=2)
lines(c(6,4,5), c(0,1,-1))
x.human <- log(mammals$body)[row.names(mammals)=="Human"]
x.human
y.human <- log(mammals$brain)[row.names(mammals)=="Human"]
points(x.human, y.human, pch=3, cex=2)
text(x.human, y.human+0.5, "me")</pre>
```

Smoothing with loess

```
#R: library(modreg)
plot(log(mammals$body), log(mammals$brain))
mammals.loess <- loess(log(brain)~log(body), data=mammals)
mammals.loess
summary(mammals.loess)
names(mammals.loess)
call  # recipe, what we can type to create this model
fitted
mammals.loess$fit
cbind(log(mammals), mammals.loess$fit)</pre>
```

Smoothing with loess

```
points(log(mammals$body), mammals.loess$fit, col=6)
lines(log(mammals$body), mammals.loess$fit, col=6)

x <- log(mammals$body)
y <- mammals.loess$fit
plot(log(mammals$body), log(mammals$brain))
lines(x[order(x)], y[order(x)])</pre>
```

Box plot

```
boxplot(cabbages$VitC)
boxplot(split(cabbages$VitC, cabbages$Date))
```

Conditioning plot

Interaction plot

interaction.plot(cabbages\$Cult, cabbages\$Date, cabbages\$VitC)

Plot influence diagnostics

```
par(mfrow=c(2,3))
#R: par(mfrow=c(2,2))
plot(mammals.lm)
par(mfrow=c(1,1))
plot(mammals.lm$fit, mammals.lm$res)
abline(h=0)
identify(mammals.lm$fit, mammals.lm$res, row.names(mammals))
```

Auxiliary functions

Formal extraction of elements

```
coef(mammals.lm)  # same as mammals.lm$coef

fitted(mammals.lm)  # same as mammals.lm$fitted

residuals(mammals.lm)  # select one of five different kinds of residuals

args(residuals.lm)

deviance(mammals.lm)  # GLM context, for lm this is SSE=sum(mammals.lm$res^2)
```

Model building and selection



Predict from new data

```
new.cabbage <- data.frame(Cult="c39", Date="d16", HeadWt=4.0)
predict(cabbages.plain, new.cabbage)
predict(cabbages.full, new.cabbage)
exp(predict(mammals.lm, data.frame(body=100)))</pre>
```

Influence diagnostics

See slide: Plot influence diagnostics

Transform response variable

```
mammals.plain <- update(mammals.lm, brain~body)
library(MASS)
plot(boxcox(mammals.plain))  # evaluate 1/Y, log(Y), Y, Y^2, ...
plot(logtrans(mammals.plain))  # evaluate log(Y+0.1), log(Y+1), ...</pre>
```

Models related to Im and aov

```
?glm
?gam #R: library(mgcv)
?nls #R: library(nls)
```

Caveats

I recommend never using attach() on data frames.

Extract residuals from Im and aov objects using the lazy \$res, but use formal residuals(x,type="") for other models.

Exercise: weight loss

```
library(MASS)
#R: data(wtloss)
wtloss <- wtloss</pre>
```

Analyze the data:

- Fit a model that goes through the data reasonably well
- Paste a table and graph into Word
- Bonus question: one might be interested in predicting the person's weight after two years at the health clinic