

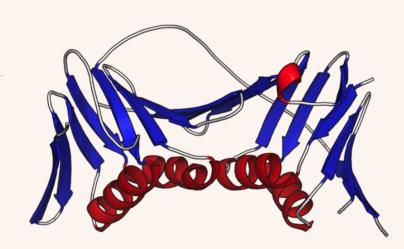
ALPHA HELICES

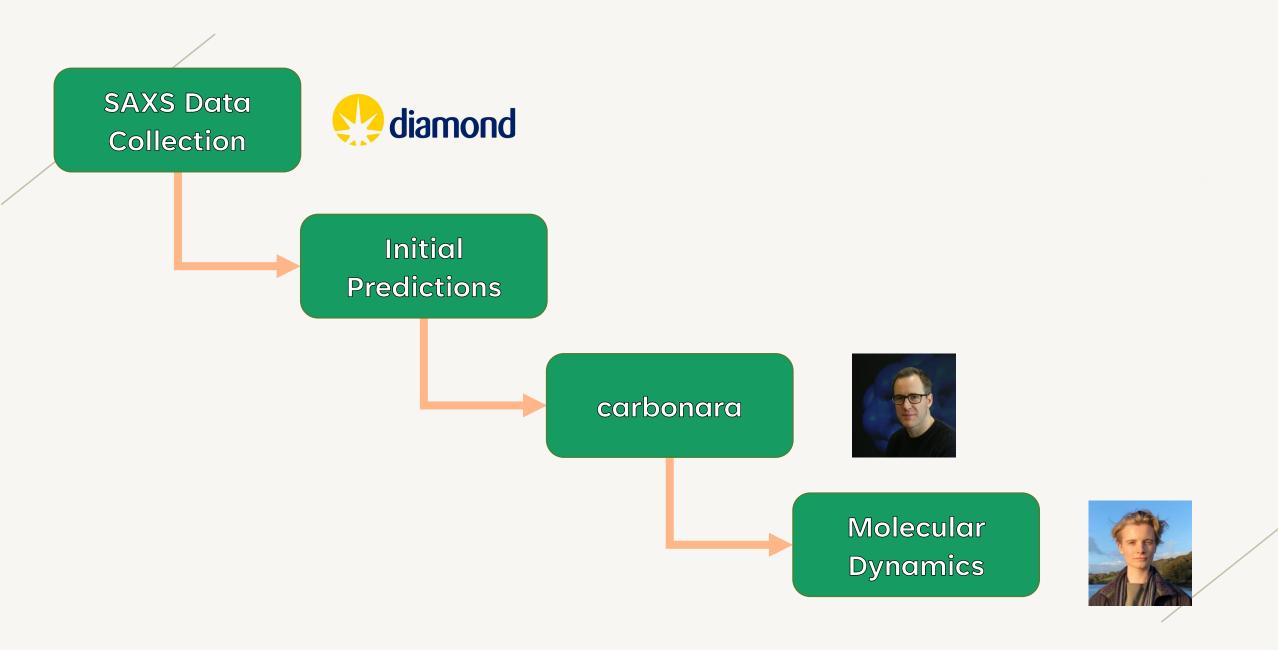
SECONDARY
STRUCTURE

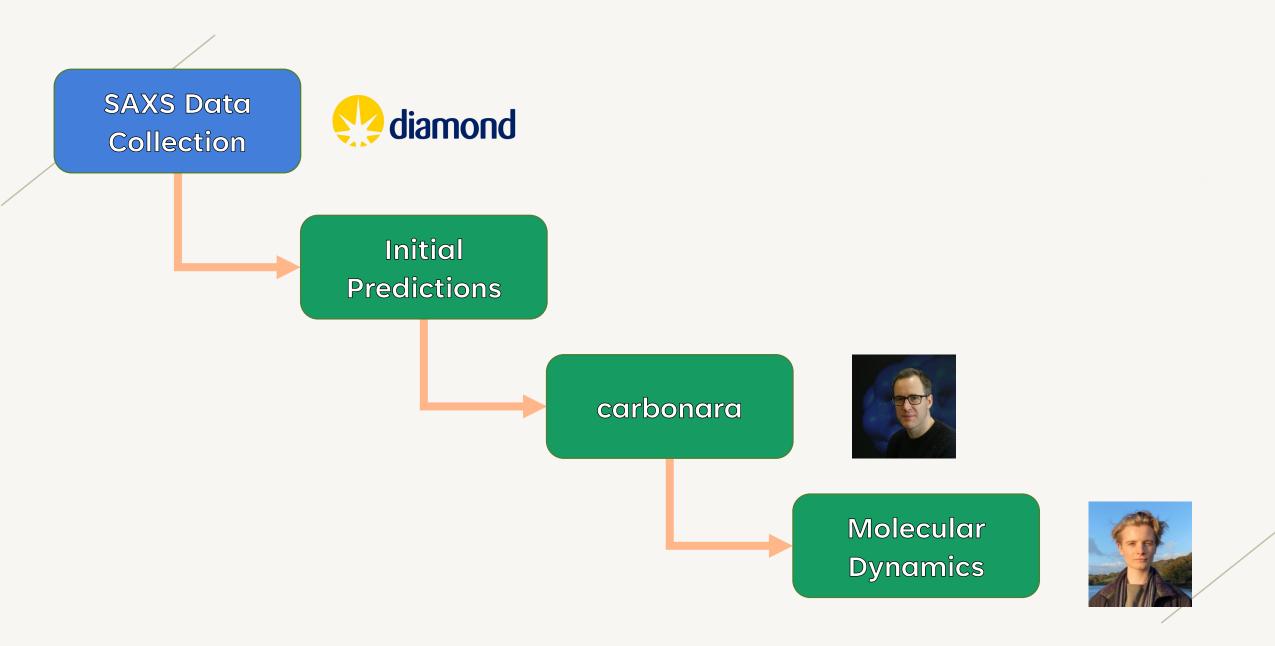
BETA-SHEETS

TERTIARY STRUCTURE

START EASY: WHAT IS A PROTEIN?

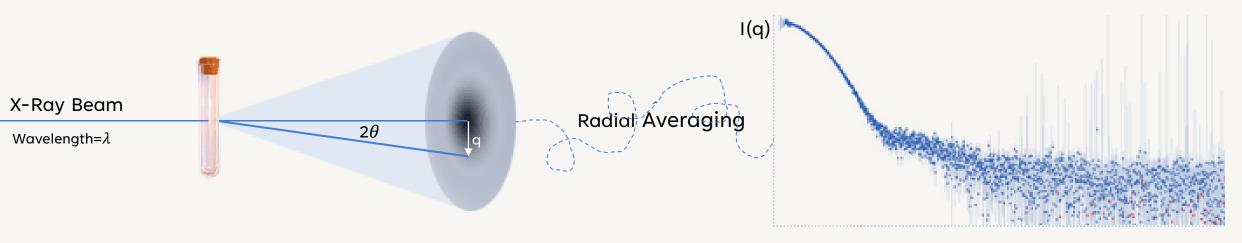




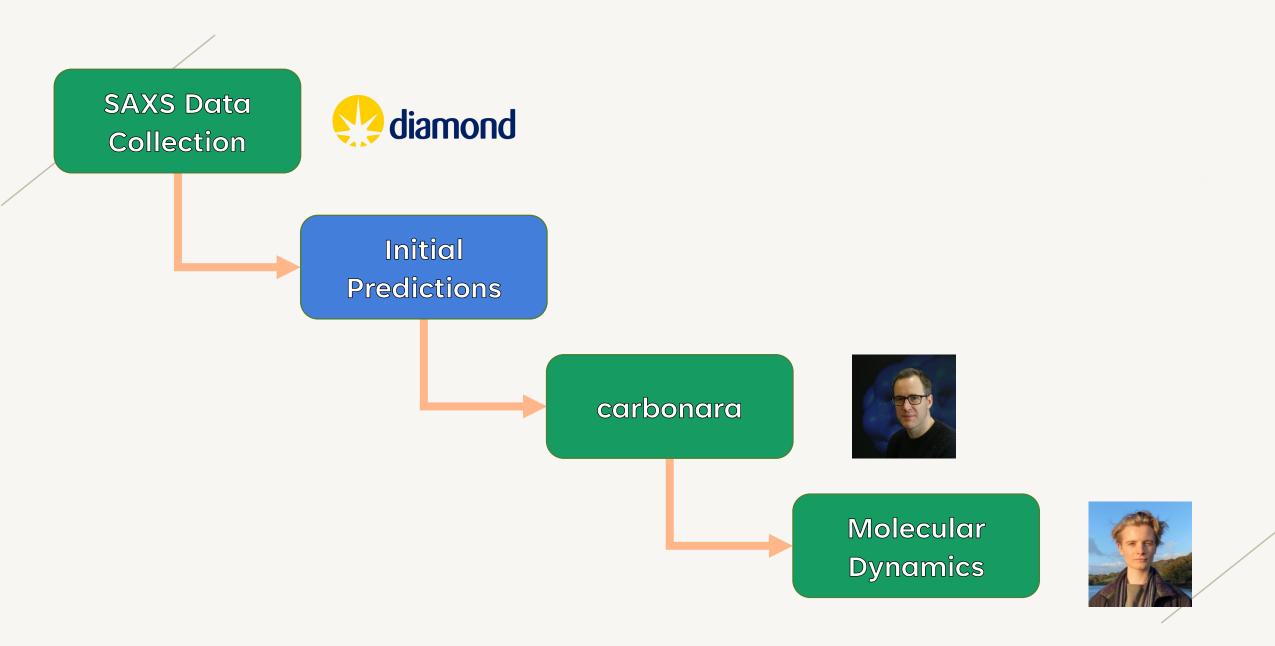




### SMALL ANGLE X-RAY SCATTERING



 $q=4\pi \sin \theta/\lambda$ 

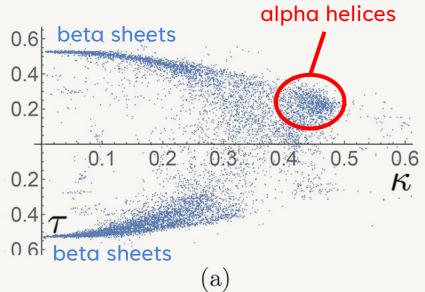


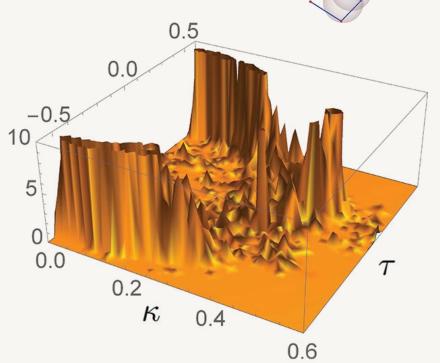
Tell me the secondary structure, I'll build a backbone curve

We can produce **locally** realistic protein curves using just two parameters:

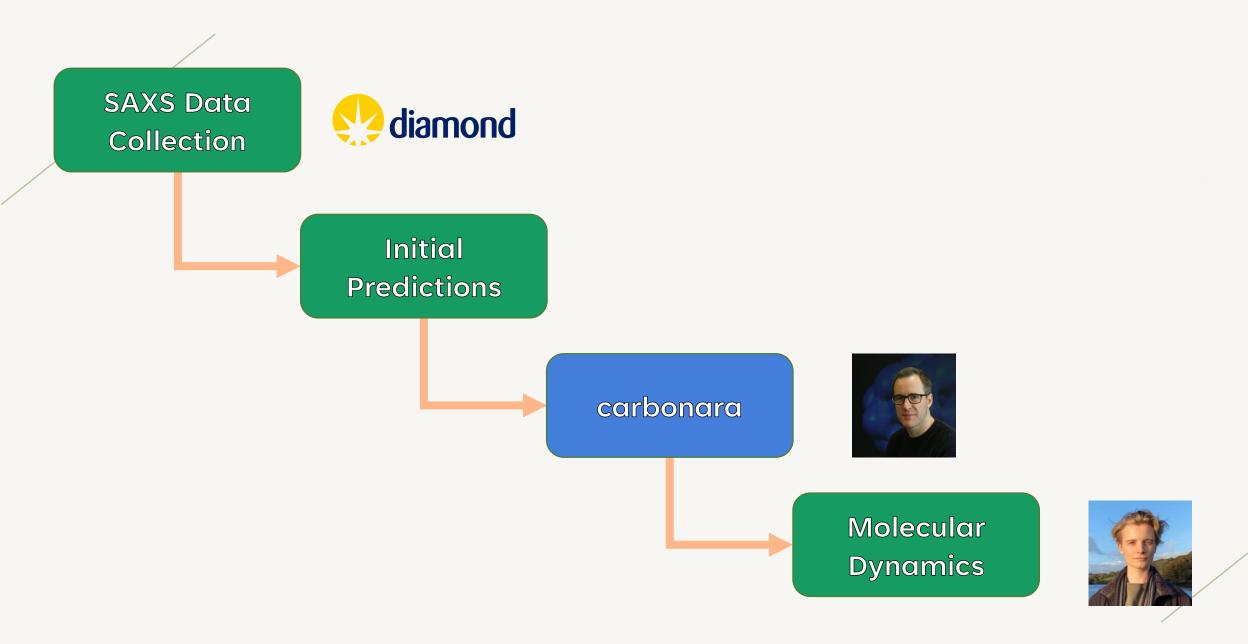
**Curvature** ( $\kappa$ ) ~ how tightly wound is my curve.

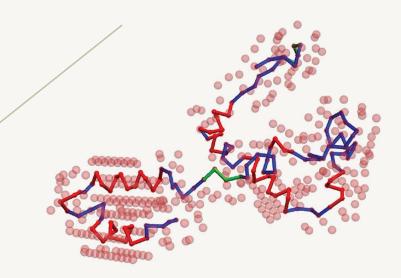
**Torsion**  $(\tau)$  ~ how helical is my curve.



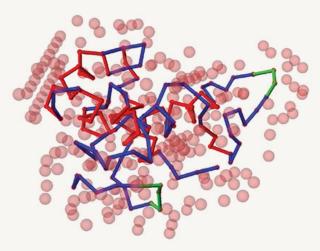


(b)

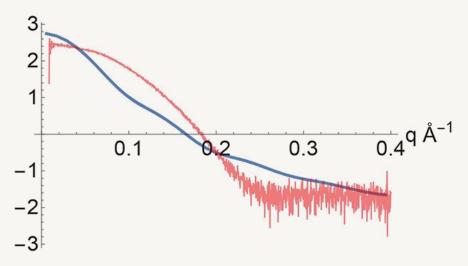




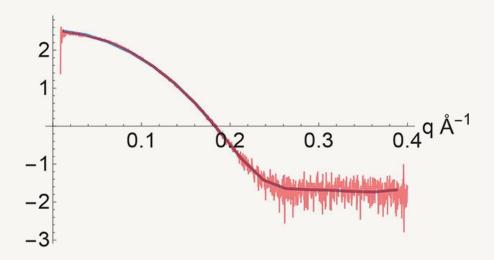
A starting configuration. Alpha helices and beta strands are shown in red and green respectively.



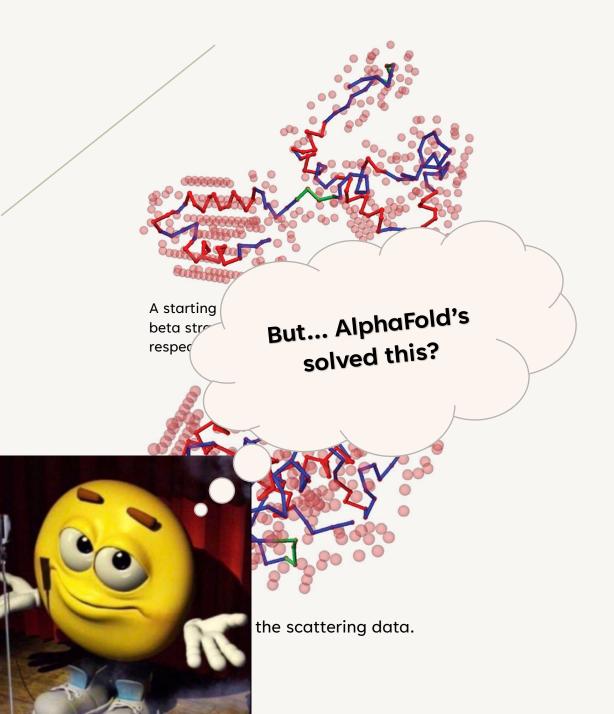
A good fit to the scattering data.

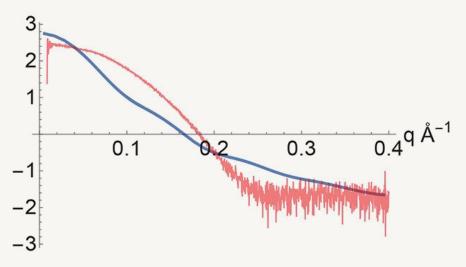


In red: The experimental scattering data. In blue: The scattering profile of the starting configuration (not good...)

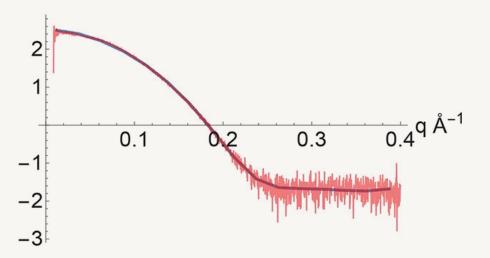


In red: The experimental scattering data. In blue: The scattering profile of the final configuration (good!)



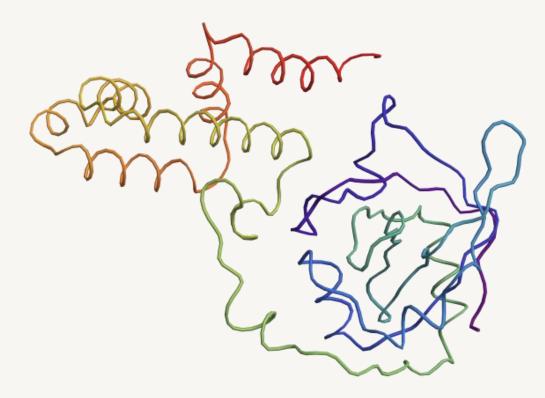


In red: The experimental scattering data. In blue: The scattering profile of the starting configuration (not good...)

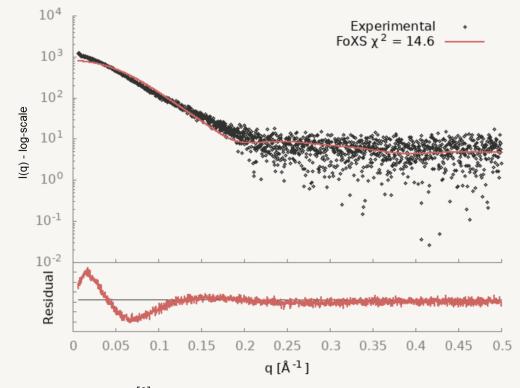


In red: The experimental scattering data. In blue: The scattering profile of the final configuration (good!)

# NOT THIS TIME ALPHAFOLD!

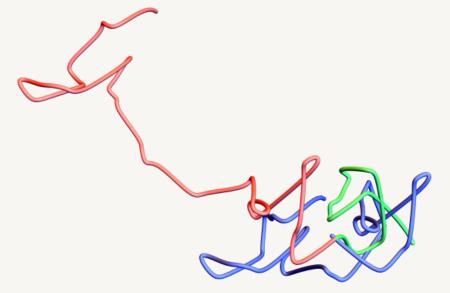


The crystallographic (=AlphaFold<sup>[1]</sup> predicted) structure.



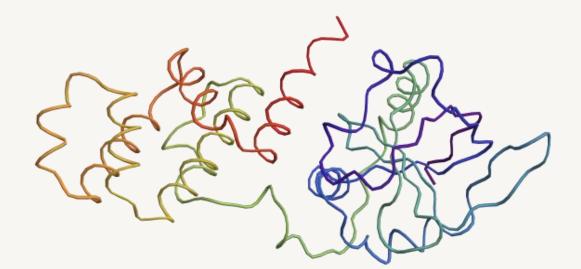
In red, the FoXS<sup>[2]</sup> scattering profile for the crystallographic structure. This is a poor fit, especially at low q, indicating the global structure is not right.

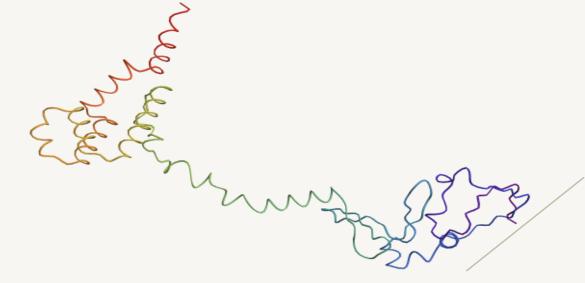
<sup>[1]</sup> Jumper, J et al. Highly accurate protein structure prediction with AlphaFold. Nature (2021).



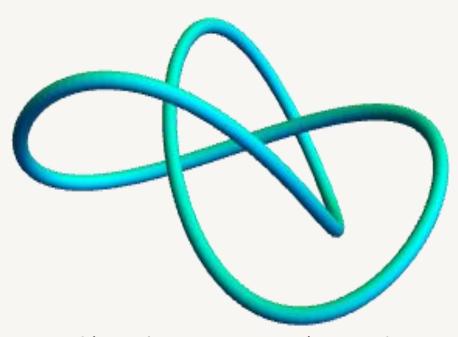
Two good fits to the scattering data.

Where they agree is shown in green. The conformations on the left and right are shown in blue and red respectively

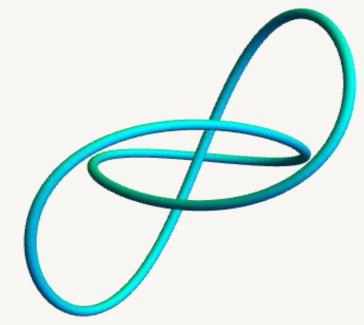




# Average Crossing Number (ACN)



From this angle, we see 4 crossings on the trefoil knot.

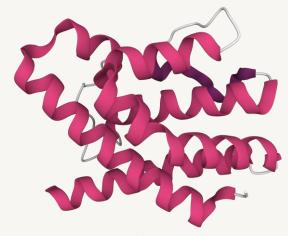


From this angle, we see 3 crossings on the trefoil knot.

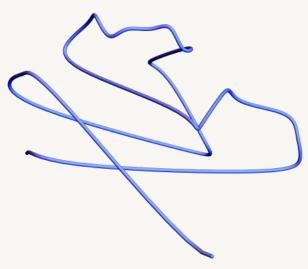
#### **SMOOTHING**

- Given the secondary structure fingerprint of a protein.
- Take the start point (c $\alpha$  molecule) of each SSE.
- Construct the discrete curve given by this subset of points (and the end points of the protein).

By smoothing in this way, we get a much clearer picture of the global entanglement of the protein, and work on a resolution closer to that of most SAXS data.

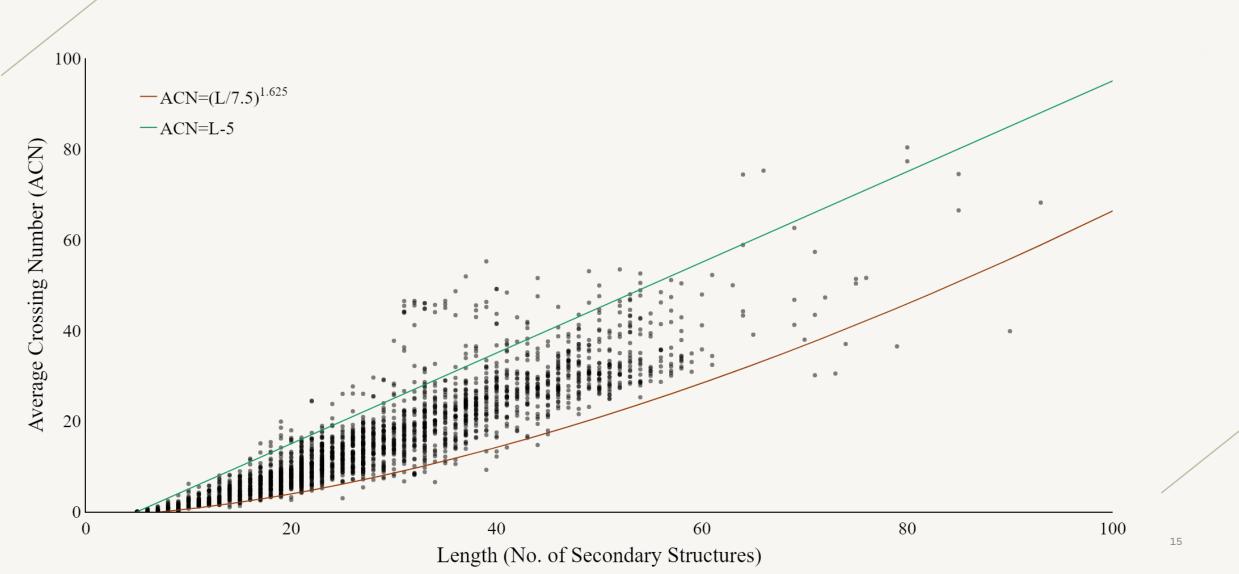


Cartoon representation of PDB: 20EB. In pink are the alpha-helices and in white the flexible linkers.

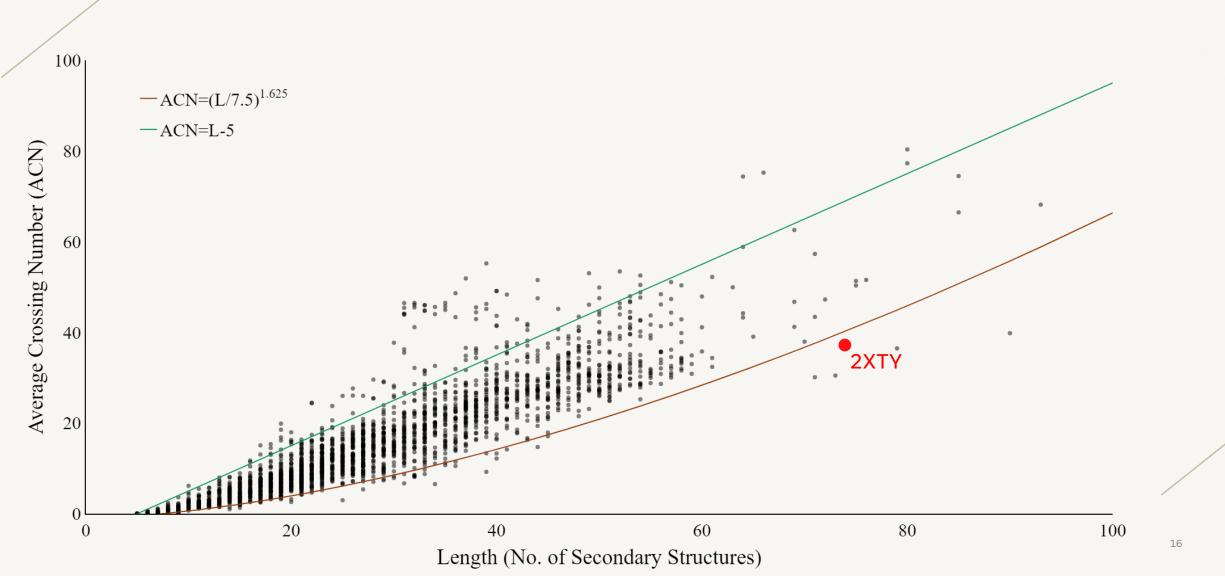


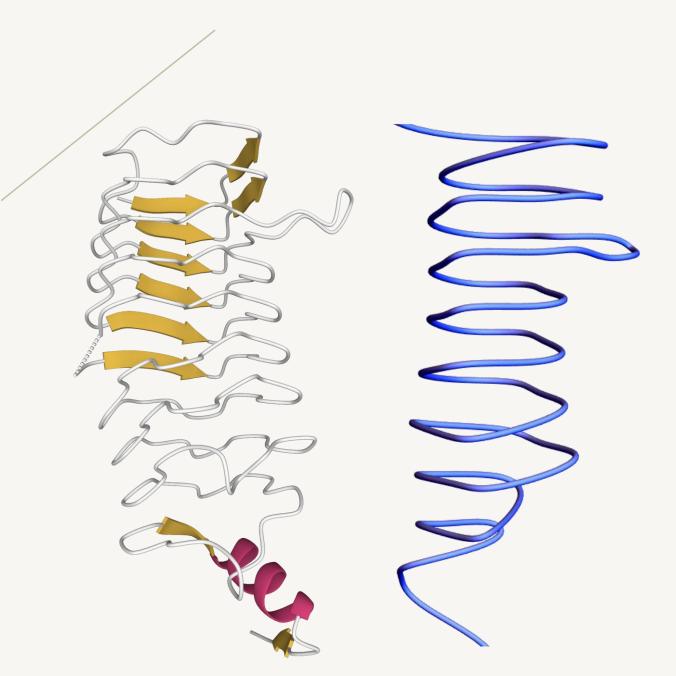
Our smoothed representation of 20EB. Note the lack of real global entanglement.

An empirical bound on the average crossing number for a representative sample of >2000 proteins from the PDB



An empirical bound on the average crossing number for a representative sample of >2000 proteins from the PDB

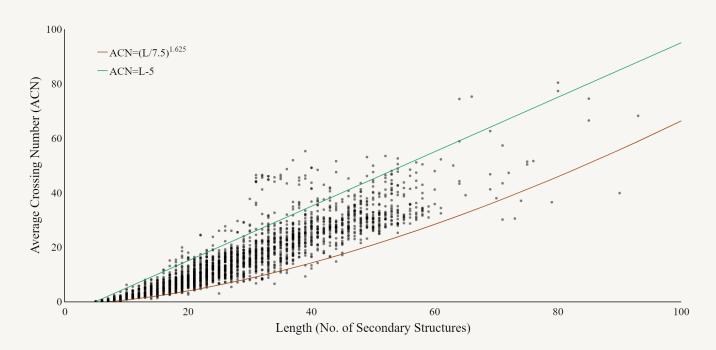


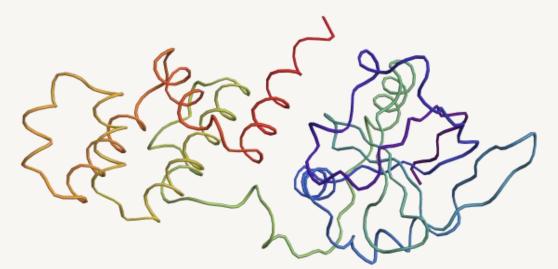


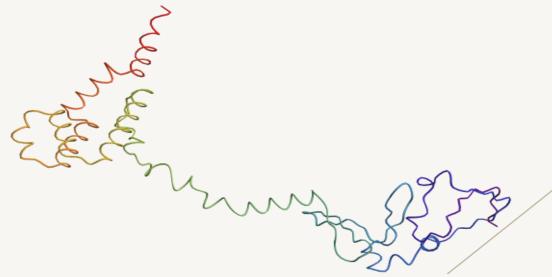
#### 2XTY

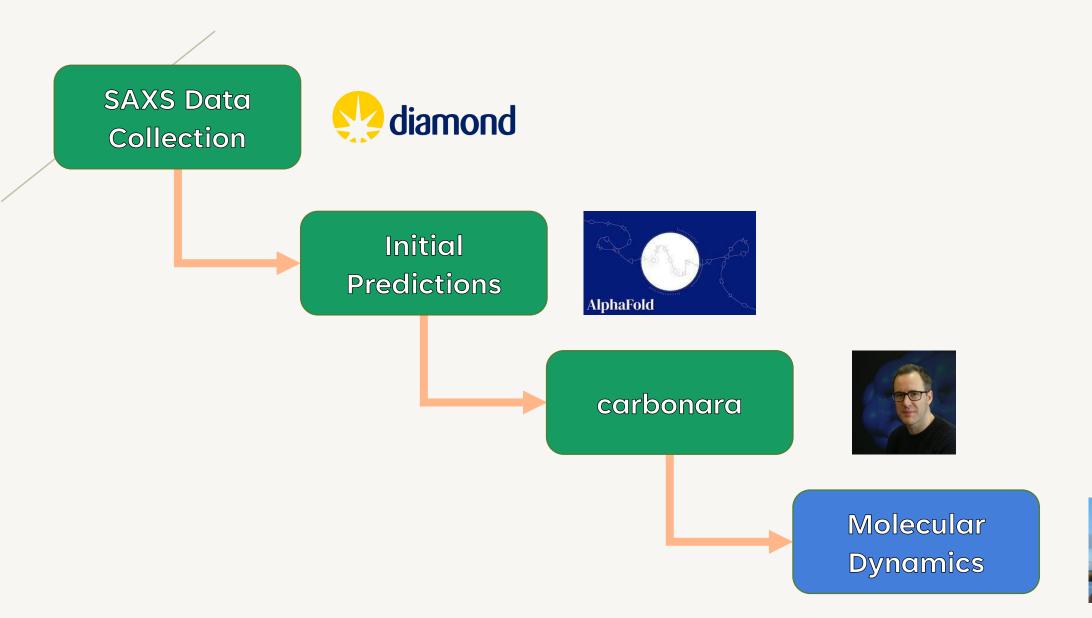
Vetting, M.W., Hegde, S.S., Wang, M., Jacoby, G.A., Hooper, D.C., Blanchard, J.S. (2011) J Biol Chem 286: 25265

- Secondary structure assignment not always reliable
- Here, DSSP finds many single amino acid long  $\beta$ -strands, opposite those seen in yellow in the left figure.
- With the correct length, this falls well inside the lower bound
- With initial predictions from e.g. AlphaFold, not an issue, the secondary structure assignment is clean.











carbonara is able to produce novel structures which fit solution scattering data.

The Average Crossing Number (ACN) of proteins is bounded from above and below.

CONCLUSIONS

The ACN ensures we make realistic predictions, which behave well in molecular dynamics simulations.













## THANK YOU FOR LISTENING

Any questions?