

Bridging the Gap Between Clinical and Laboratory Label-free Two-photon Excitation Microscopy

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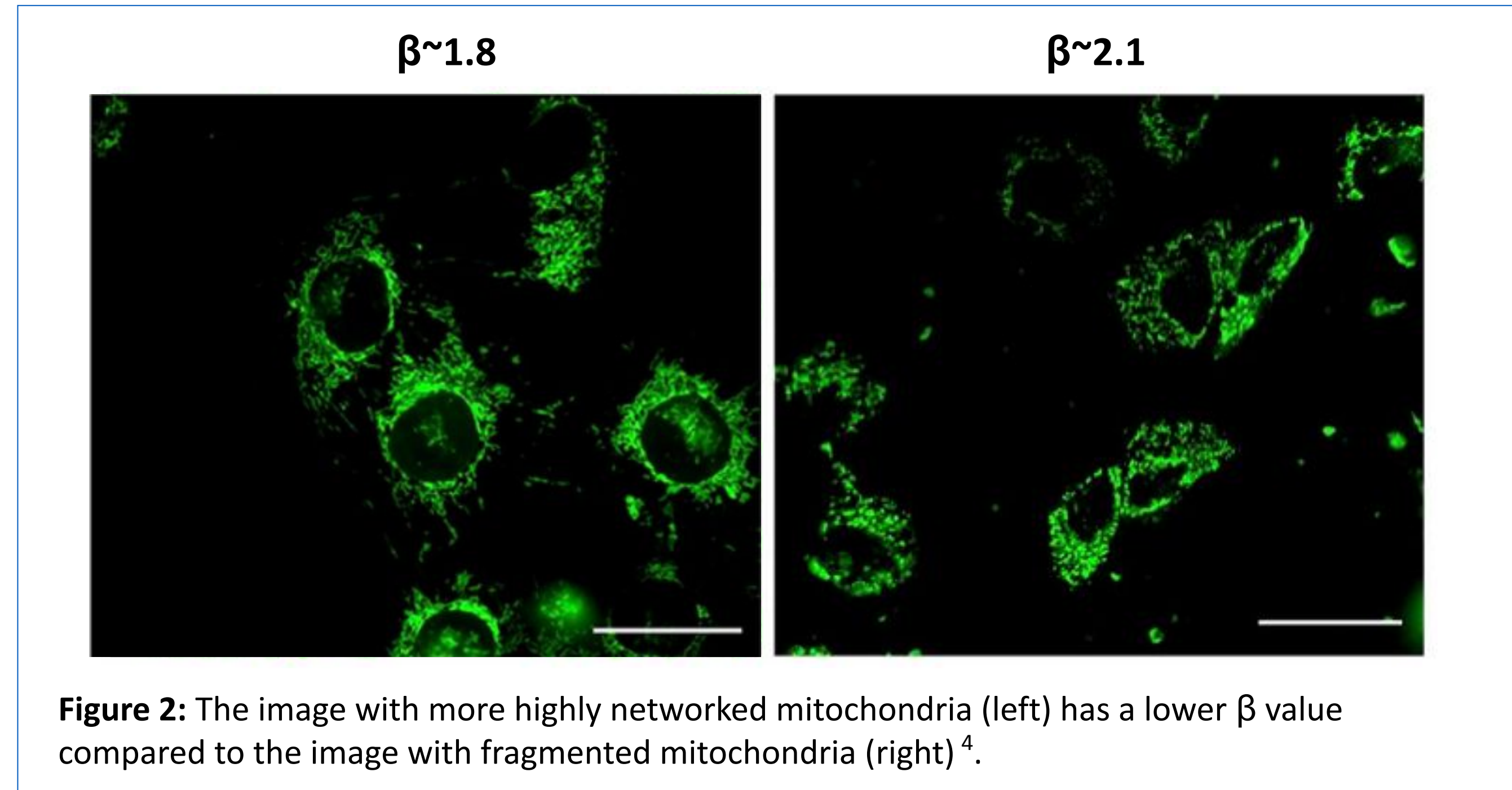
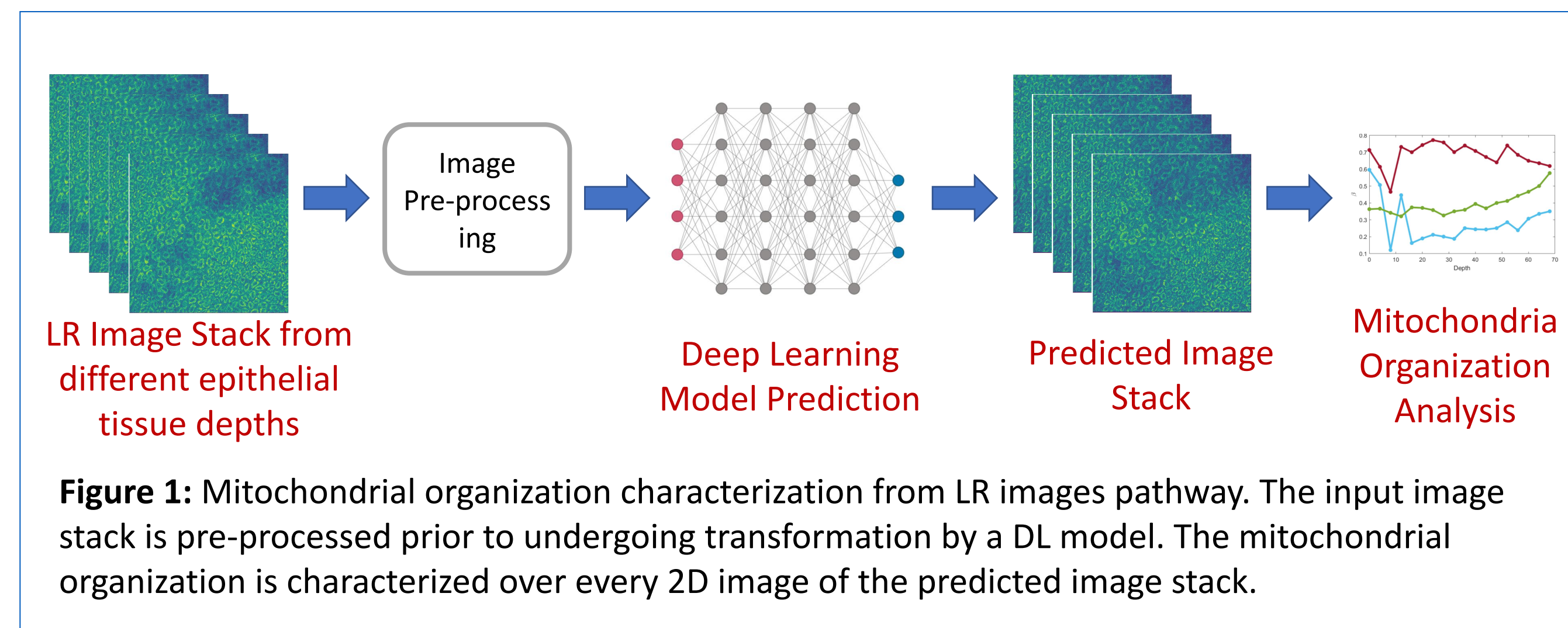
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Introduction

- Metabolic perturbation is a prevalent characteristic of many cancers, including cervical cancer¹.
- Mitochondrial organization (MO) correlates with cellular metabolic activity, making it a potential early cancer diagnostic biomarker².
- MO is quantitatively characterized using automated analysis of sub-micron resolution reduced nicotinamide adenine dinucleotide (phosphate) (NAD(P)H) two-photon excited fluorescence (TPEF) images³.
- Clinical label-free TPEF microendoscopes have lower resolution (LR) than lab-bench microscopes and need to be enhanced *in silico* to detect MO-associated precancerous metabolic changes.

Objective:

Developing a to achieve reliable mitochondrial organization assessments in low-resolution NAD(P)H images.

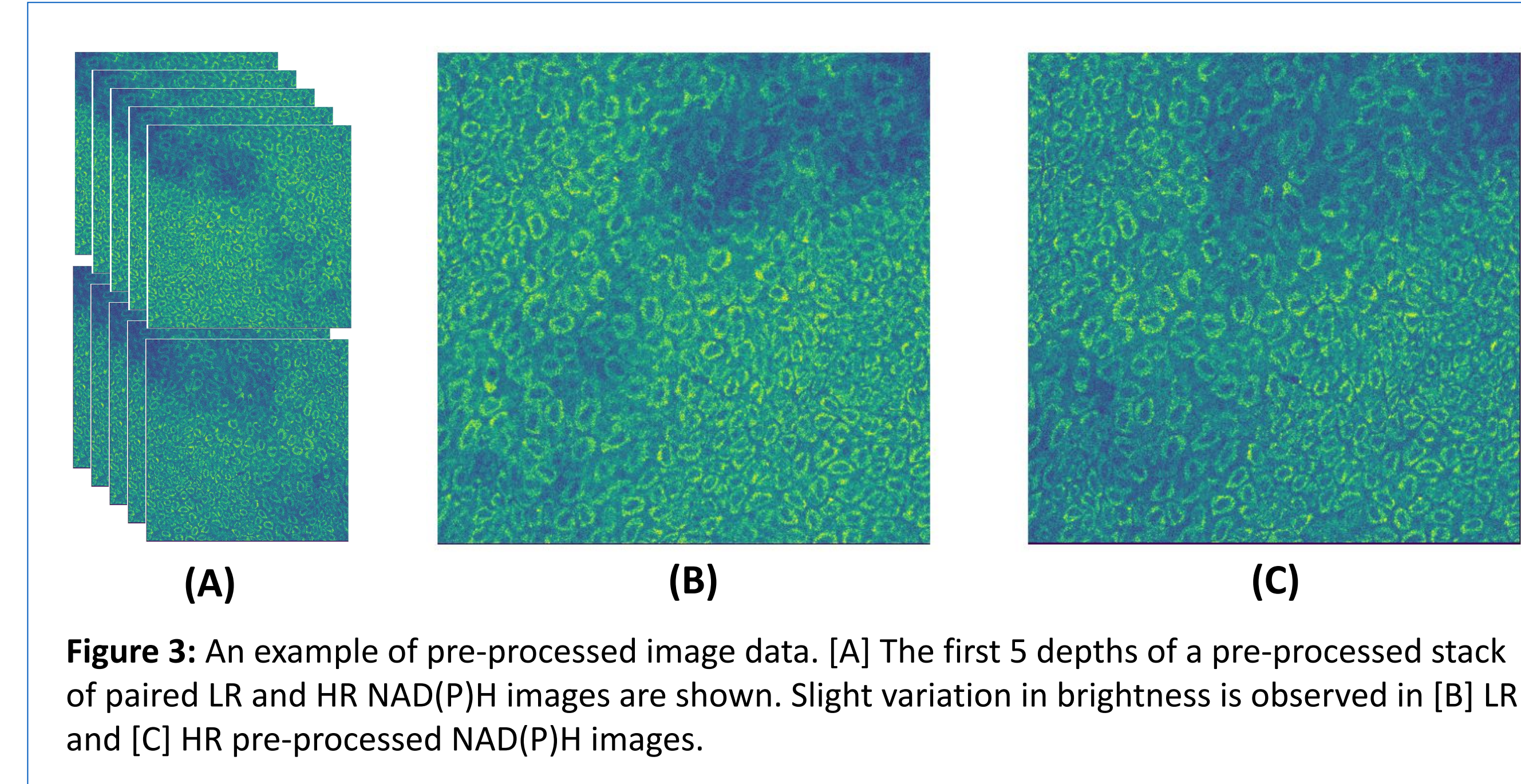


Methods

Dataset:

- 15 three-dimensional (3D) stacks of NAD(P)H images (512-by-512-pixels) are acquired from stratified squamous epithelial tissues using a TPEF microscope with 25X (LR) and 40X (HR) objectives and numerical apertures of 0.95 and 1.1, respectively.
- 238 images are preprocessed, registered as pairs of LR and high-resolution (HR) images, and split into training-validation sets with a 4:1 ratio.
- Images are patched into 3808 patches (128-by-128-pixels) for U-Net training.

Methods (continued)



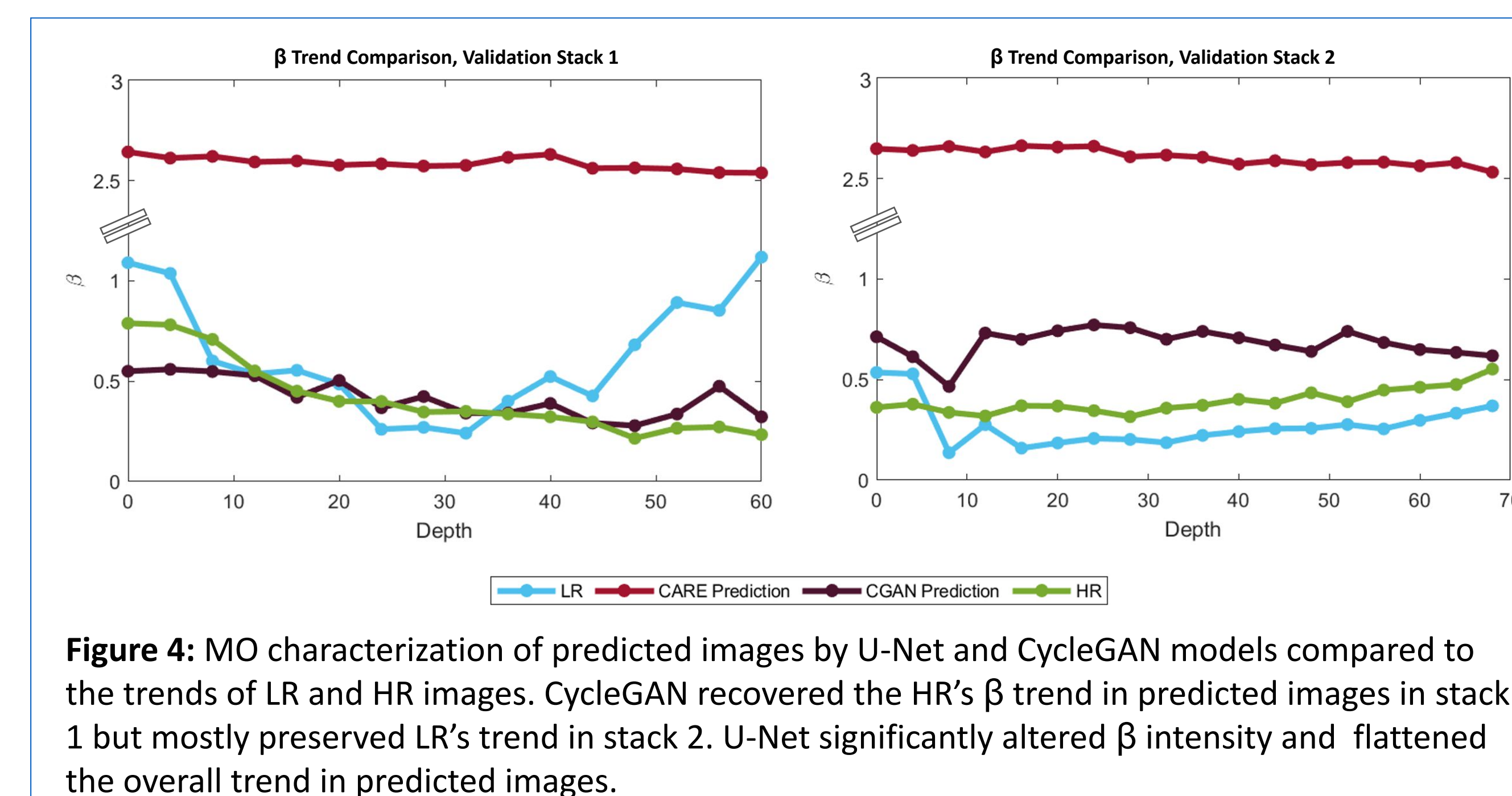
Training:

- Standard super-resolution and image restoration metrics are selected for training validation: structural similarity index (SSIM), peak signal-to-noise ratio (PSNR), and their variations.
- Two deep learning models are trained to transform LR images so that MO-associated metrics are similar to those from HR image analysis.
- A supervised deep convolutional neural network (CNN) based on U-net architecture is trained on image pairs⁵.
- An unsupervised generative adversary network (GAN) is trained on unpaired training and GT images. The model architecture is based on cycle-consistent adversarial networks (CycleGAN)⁶.
- Hyperparameter search and data augmentation is performed to improve training performance.

Analysis:

- Predicted images are visually inspected, and their image similarity to HR is quantitatively assessed.
- MO is characterized using the value of an inverse power law exponent, β , which is fit to the power spectral density of the NAD(P)H images⁷.
- β values of LR, predicted, and HR images are evaluated across depths of the tissue stack, and their respective MO trends are compared.

Results

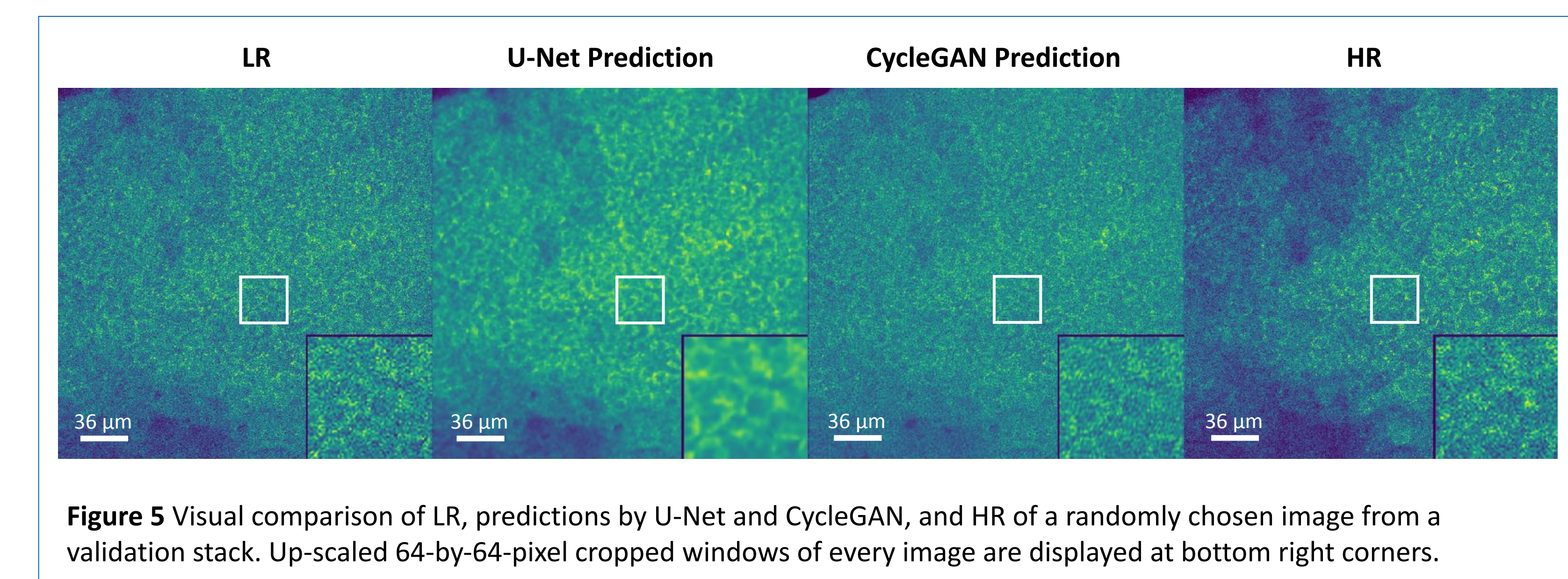


Results (continued)

- Manual inspection of image pairs revealed variability due to changes in the live specimen and non-ideal registration in some image pairs.
- U-Net and CycleGAN models improved the LR images to become more structurally similar to HR images as SSIM increased; however, we observed a decrease in PSNR.
- Visual analysis showed that the U-Net substantially blurred predicted images while CycleGAN reduced the signal contrast in predicted images.
- β trends across all images in image stacks predicted by the U-Net were not recovered. The recovered β trends are nearly flat in all evaluated stacks.
- β trend recovery in predicted image stacks by CycleGAN was inconclusive as MO characterization yielded inconsistent results over evaluated stacks.

Table 1: Quantitative analyses results averaged over the validation dataset

	SSIM	PSNR	β Value
LR	0.261	17.2	0.449
HR	—	—	0.407
U-Net	0.283	16.8	2.60
CycleGAN	0.299	16.2	0.550



Discussion

- Changes in SSIM and PSNR metrics do not correlate with improved MO characterization.
- The U-Net model performed poorly on MO analysis over the given dataset. We believe that variations and artifacts in paired images limited the performance of the U-Net.
- CycleGAN improved MO characterization accuracy based on analysis of LR images.
- We hypothesize that CycleGAN's better performance is attributed to the unsupervised training approach, which was not affected by variations in paired images.
- Future studies will focus on collecting new *in vivo* data which will minimize variations in paired images such as tissue shrinkage. Further investigation of unsupervised deep learning methods will be performed on the new datasets.

References:

- [1] Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science*. 2009;324(5930):1029-1033. [2] Xylas J, Varone A, Quinn KP, et al. Noninvasive assessment of mitochondrial organization in three-dimensional tissues reveals changes associated with cancer development. *Int J Cancer*. 2015;136(2):322-332. [3] Liu Z, Pouli D, Alonzo CA, et al. Mapping metabolic changes by noninvasive, multiparametric, high-resolution imaging using endogenous contrast. *Sci Adv*. 2018;4(3):eaap9302. [4] Jheng HF, Tsai PJ, Guo SM, et al. Mitochondrial fission contributes to mitochondrial dysfunction and insulin resistance in skeletal muscle. *Mol Cell Biol*. 2012;32(2):309-319. [5] Weigert M, Schmidt U, Boothe T, et al. Content-aware image restoration: pushing the limits of fluorescence microscopy. *Nat Methods*. 2018;15(12):1090-1097. [6] Kearney V, Ziemer BP, Perry A, et al. Attention-Aware Discrimination for MR-to-CT Image Translation Using Cycle-Consistent Generative Adversarial Networks. *Radiol Artif Intell*. 2020;2(2):e190027. [7] Pouli D, Balu M, Alonzo CA, et al. Imaging mitochondrial dynamics in human skin reveals depth-dependent hypoxia and malignant potential for diagnosis. *Sci Transl Med*. 2016;8(367):367ra169.