



# Understanding genotype-phenotype relationships using the model plant Arabidopsis thaliana

**Arthur Korte 01.03.2021** 

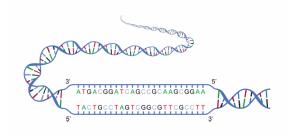




# Genotype



# **Phenotype**



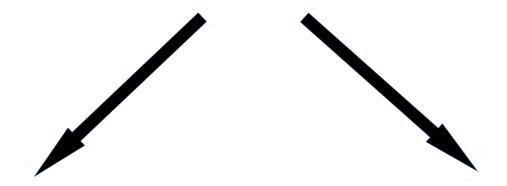






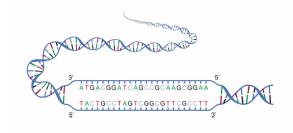


### **Environment**



Genotype













#### **Environment**

Adaptation Natural selection

Genotype Phenotype

Genome-wide association studies (GWAS)

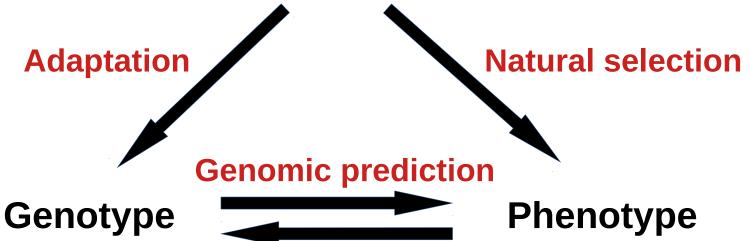








### **Environment**





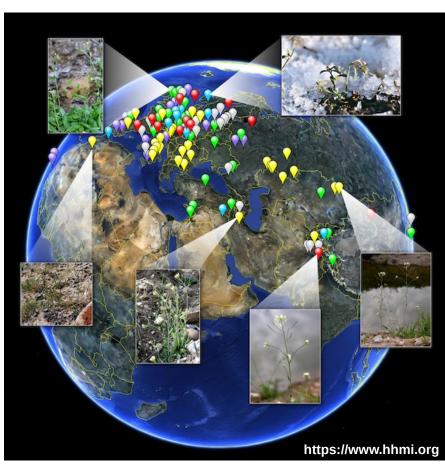






# Arabidopsis thaliana





Arabidopsis is a prime model for Evolutionary Genomics (small genome, adapted to the local environment, occurs as natural inbred)



# Arabidopsis thaliana





# **The 1001 Genomes Project**

#### 1001 Genomes

A Catalog of Arabidopsis thaliana Genetic Variation

Home Data	Providers Acces	ssions To	ools So	oftware Da	ata Center A	About
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• Full genome sequence of 1,135 different natural ecotypes

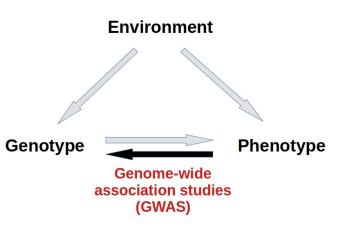
10M SNPs and 500k structural variants in a genome with ~125 M Bp





### Genome-wide association studies (GWAS)





Associations between genotype and phenotype



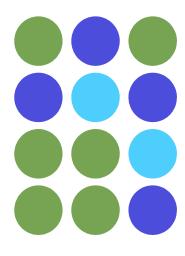
# Genome-wide association studies (GWAS)



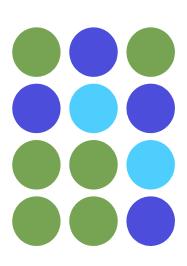


Associations between genotype and phenotype



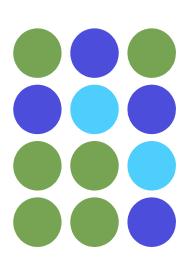






```
• • • AGCCTG TTGCACTAAGACT • • •
• • • AGCCTG TTGCACTAAGACT • • •
• • • AGCCTG TTGCACTAAGAGT • • •
• • • AGCCTG TTGCACTAAGACT • • •
• • • AGCCTGAGTGTGCACTAAGAGT• • •
• • • AGCCTGAGTGTGCACTAAGAGT• • •
• • • AGCCTGAGTGTGTACTAAGACT • • •
• • • AGCCTGAGTGTGTACTAAGAGT• • •
• • • AGCCTGAGTGTGTACTAAGACT • • •
• • • AGCCTGAGTGTGTACTAAGAGT• • •
• • • AGCCTGAGTGTGTACTAAGACT • • •
• • • AGCCTGAGTGTGTACTAAGACT • • •
```





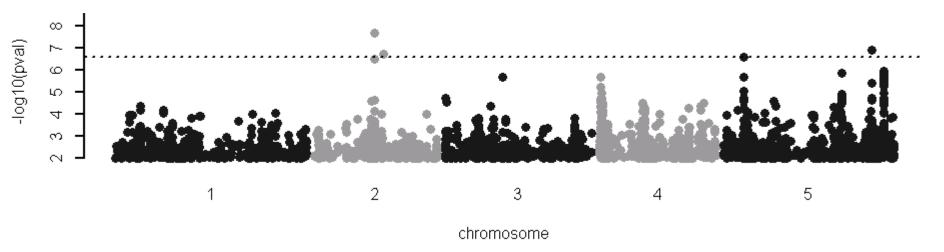
```
TGCACTAAGAC
• • • AGCCTG
                 TG<mark>CA</mark>CTAAGA<mark>C</mark>
• • • AGCCTG
                 TGCACTAAGAGT
AGCCTG
• • • AGCCTG — — TGCACTAAGAC
AGCCTGAGTGTGCACTAAGAGT
• • • • AGCCTGAGTGTGCACTAAGAG
• • • • AGCCTGAGTGTGTACTAAGAC
• • • AGCCTGAGTGTACTAAGAG
• • • AGCCTGAGTGTGTACTAAGAC
• • • • AGCCTGAGTGTACTAAGAG
• • • AGCCTGAGTGTACTAAGAC
• • • AGCCTGAGTGTGTACTAAGA
```



Calculate a statistical test for each SNP using linear mixed models

$$Y = X\beta + u + \epsilon$$
,  $u \sim N(0, \sigma_g K)$ ,  $\epsilon \sim N(0, \sigma_e I)$ 

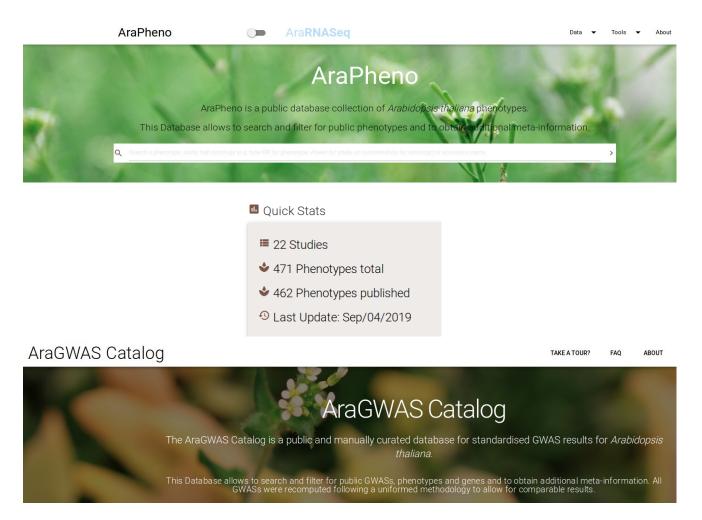
We get a p-value for each SNP that can be plotted in a Manhattan plot



Goal: To identify markers that are significantly associated with a given trait permutation-based significance threshold (Freudenthal et al. 2019)



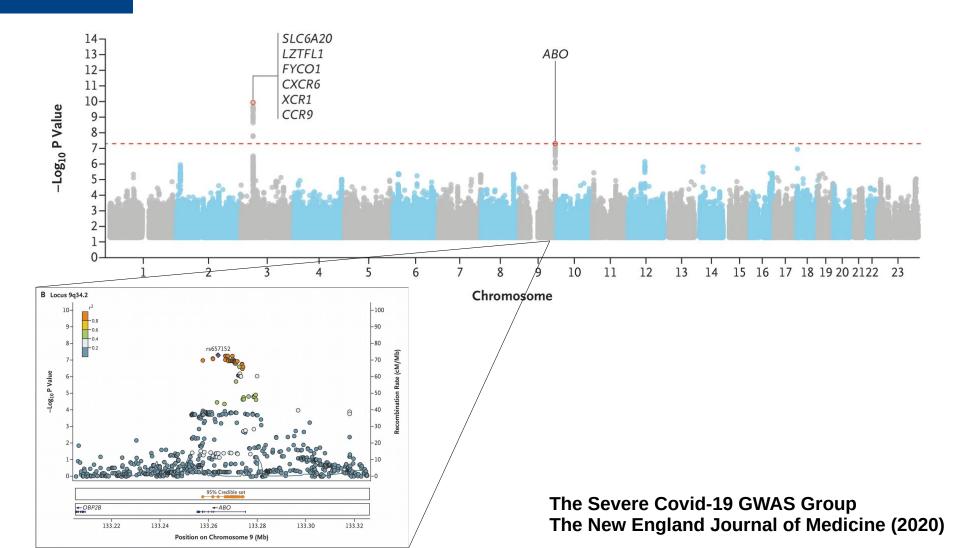
#### **AraPHENO and AraGWAS**



Seren et al. NAR (2017), Togninalli et al. NAR (2018) & Togninalli et al. NAR (2020)



#### **GWAS on Covid-19 severeness**





#### Gene x environment interactions

# Genotype Phenotype Genome-wide

**GWAS**: Genome - Phenotype

association studies

(GWAS)



#### **HHS Public Access**

Author manuscript

Nat Genet. Author manuscript; available in PMC 2013 March 01.

Published in final edited form as:

Nat Genet. 2012 September; 44(9): 1066-1071. doi:10.1038/ng.2376.

#### A mixed-model approach for genome-wide association studies of correlated traits in structured populations

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#### **Abstract**

Genome-wide association studies (GWAS) are a standard approach for studying the genetics of natural variation. A major concern in GWAS is the need to account for the complicated dependence-structure of the data both between loci as well as between individuals. Mixed models have emerged as a general and flexible approach for correcting for population structure in GWAS. Here we extend this linear mixed model approach to carry out GWAS of correlated phenotypes, deriving a fully parameterized multi-trait mixed model (MTMM) that considers both the within-trait and between-trait variance components simultaneously for multiple traits. We apply this to human cohort data for correlated blood lipid traits from the Northern Finland Birth Cohort 1966, and demonstrate greatly increased power to detect pleiotropic loci that affect more than one blood lipid trait. We also apply this to an Arabidopsis dataset for flowering measurements in two different locations, identifying loci whose effect depends on the environment.

Intraduction

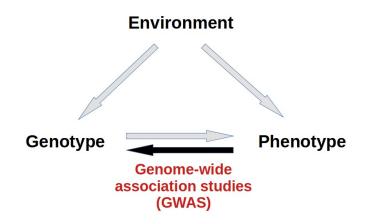
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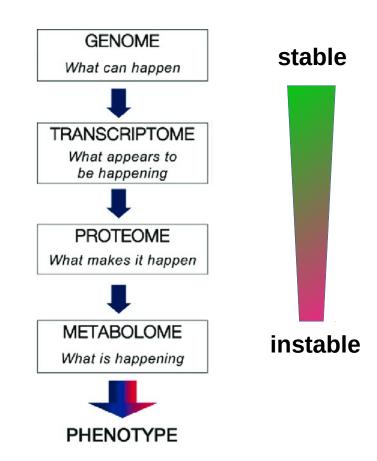
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**GWAS**: Genome - Phenotype





**GENOME** 

What can happen

**TRANSCRIPTOME** What appears to be happening

stable

**Full genomes >1000 ecotypes** (1001 genomes consortium, 2016)

**PROTEOME** What makes it happen **METABOLOME** instable What is happening **PHENOTYPE** 

> 500 different phenotypes

(Seren et al., 2017 & Togninalli et al. 2020)



Full genomes >1000 ecotypes (1001 genomes consortium, 2016)

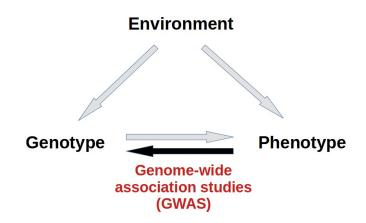
Complete transcriptomes (28 k genes) for 80 ecotypes (Clauw et. al. 2016) & for~ 700 ecotypes (Kawakatsu *et al.* 2016)

**GENOME** stable What can happen TRANSCRIPTOME What appears to be happening **PROTEOME** What makes it happen **METABOLOME** instable What is happening **PHENOTYPE** 

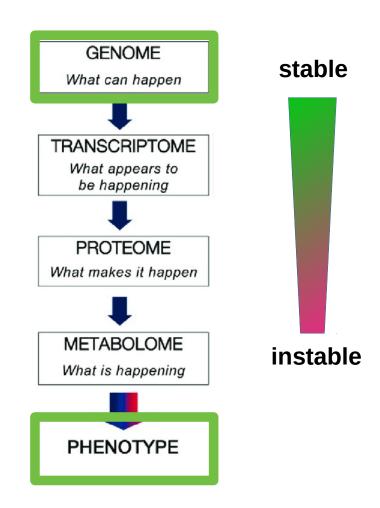
> 500 different phenotypes

(Seren et al.,2017 & Togninalli et al. 2020)

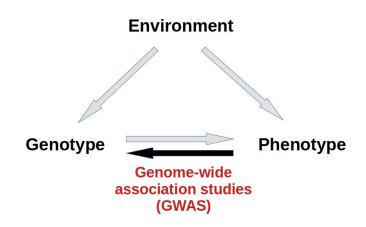




**GWAS**: Genome - Phenotype

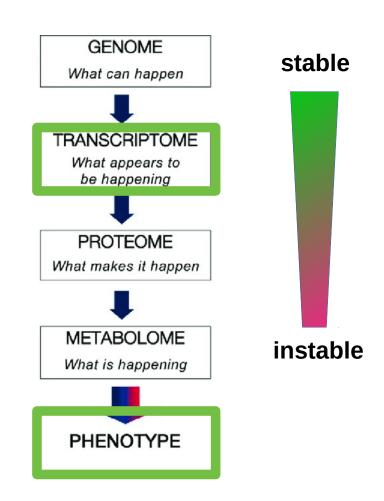






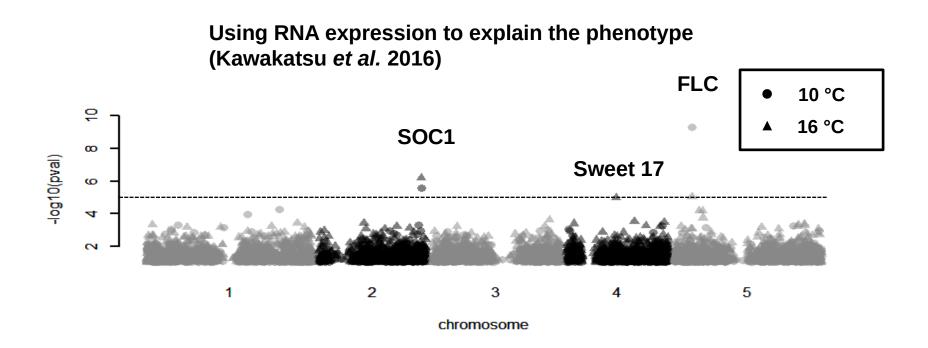
**GWAS**: Genome - Phenotype

**TWAS**: Transcriptome - Phenotype



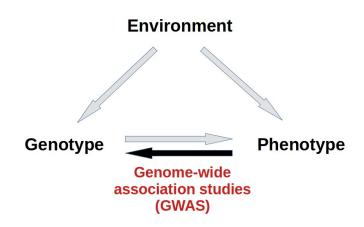


# TWAS (Transcriptome – Phenotype) for Flowering Time



**Beyond GWAS: Understanding the function of pathways** 

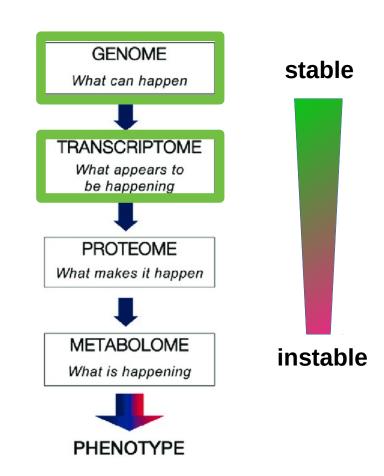




**GWAS**: Genome - Phenotype

**TWAS**: Transcriptome - Phenotype

**eGWAS:** Genome - Transcriptome

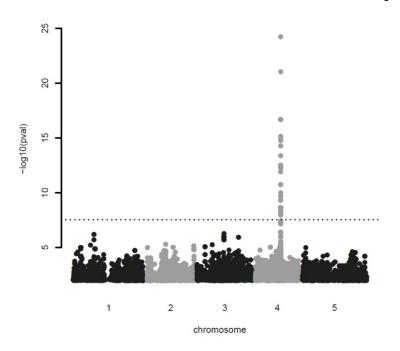




### eGWAS (Genome - Transcriptome)

Data: 80 ecotypes grown under drought and control conditions (VIB Ghent), RNAseq for all of them.

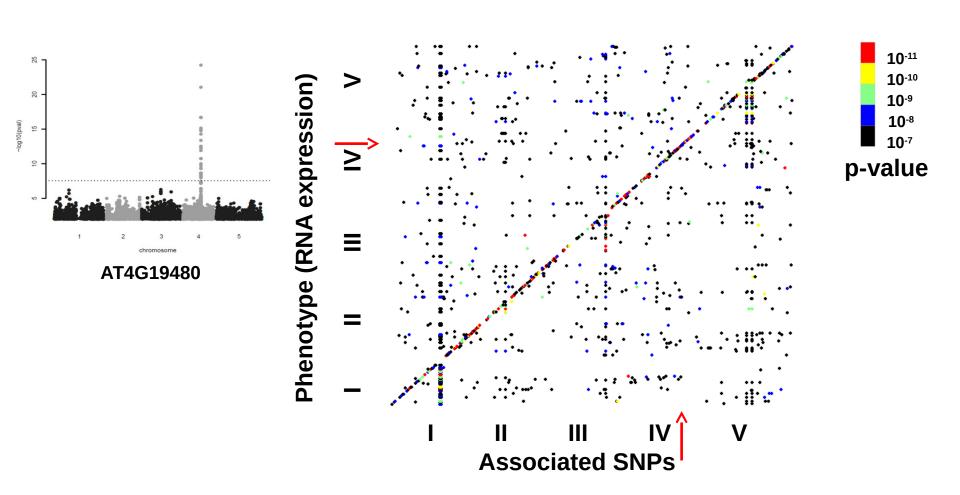
Use the expression values of each gene as molecular phenotype



A SNP on Chromosome 4 affects the expression of the gene AT4G19480 How does this affect the phenotype?



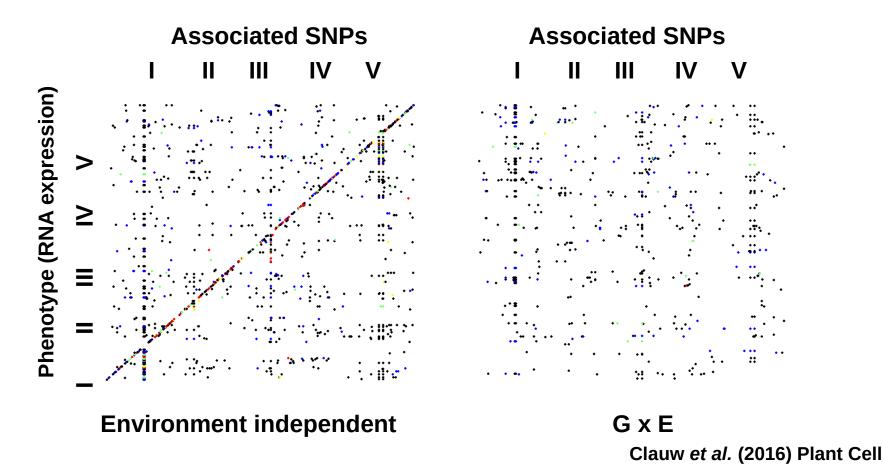
# eGWAS, summary plot of 28k analyses





#### eGWAS with a multivariate model

Joint analyses of RNAseq data from control and drought-stressed samples Method: Korte et al. (2012) NatGen





# Global genetic heterogeneitity in adaptive traits



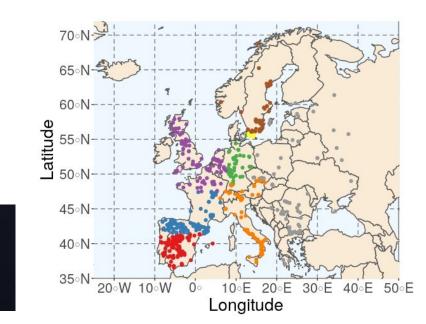


#### Global genetic heterogeneity in adaptive traits

We measured flowering time in a growth chamber at 10°C for 888 European Arabidopsis ecotypes







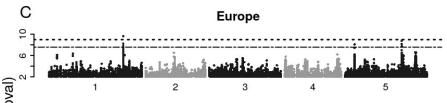


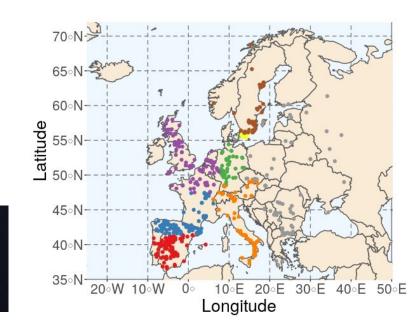
#### Global genetic heterogeneity in adaptive traits

# We measured flowering time at 10°C In 888 European Arabidopsis ecotypes









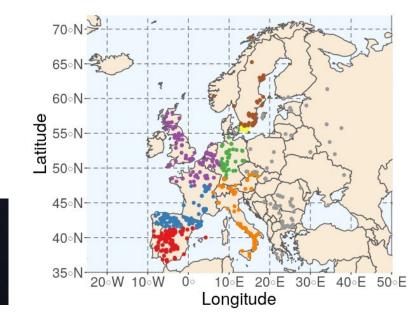


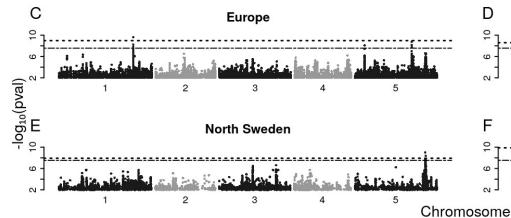
#### Global genetic heterogeneity in adaptive traits

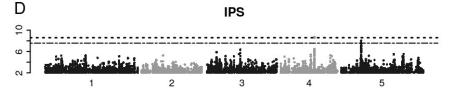
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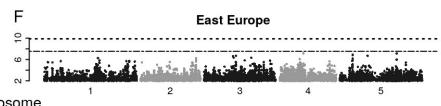










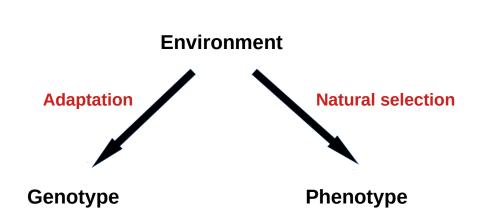


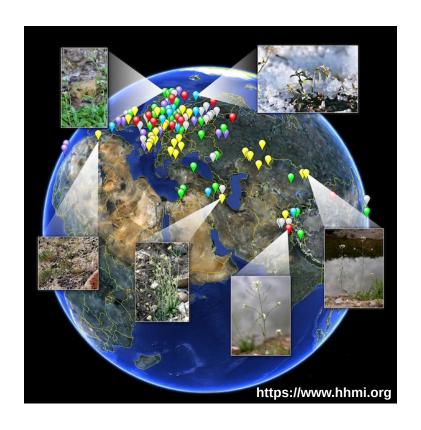
López Arboleda et al. BioRxiv



# **Genotype – Phenotype relationship**

Is Flowering time optimized by different genes in different local subsets?

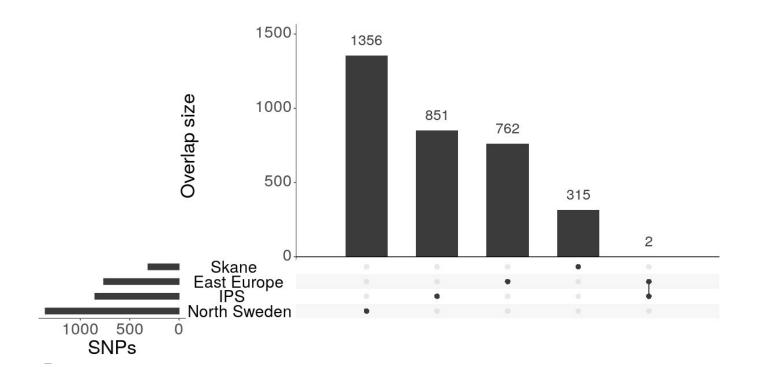






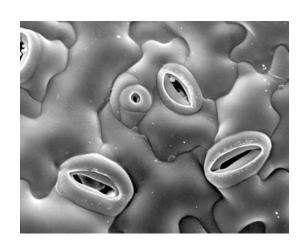
# Different genetic architecture of flowering time in different subsets

Overlap of SNPs that are associated with a p-value <10<sup>-4</sup> in the respective subset

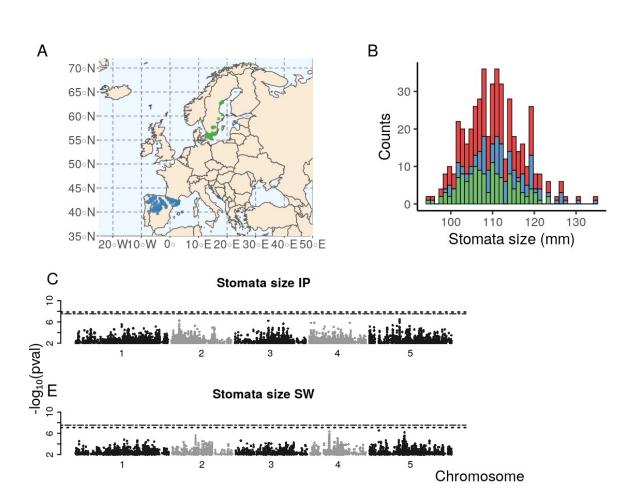




# Different genetic architecture in many adaptive traits

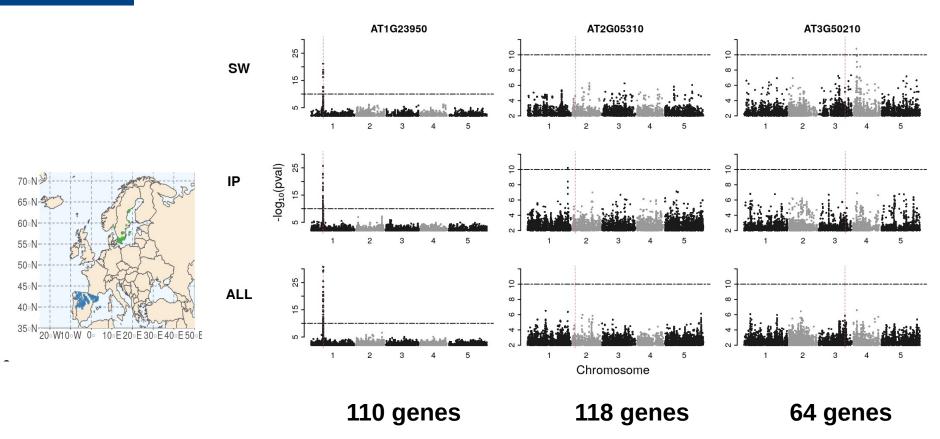


Stomata size was measured in a set of 330 European Arabidopsis ecotypes Dittberner *et al.* 2018



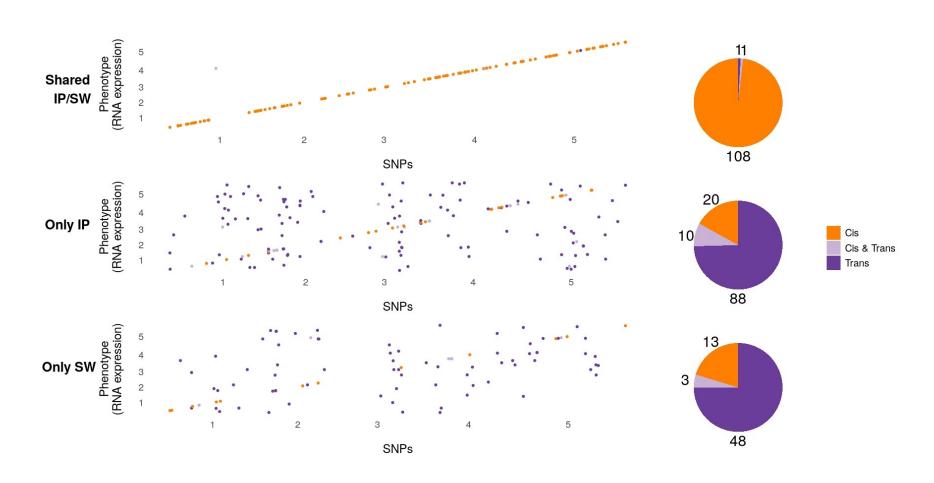


# eGWAS in different local populations



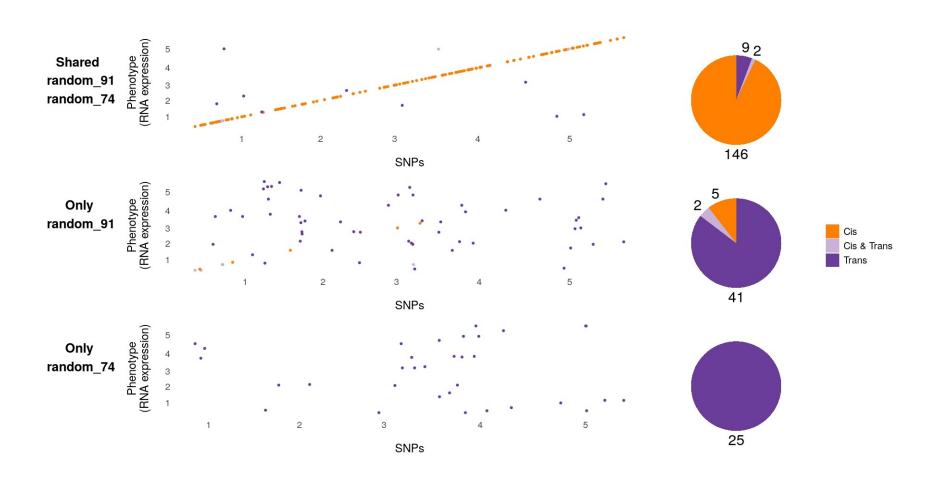


# eGWAS in different local populations



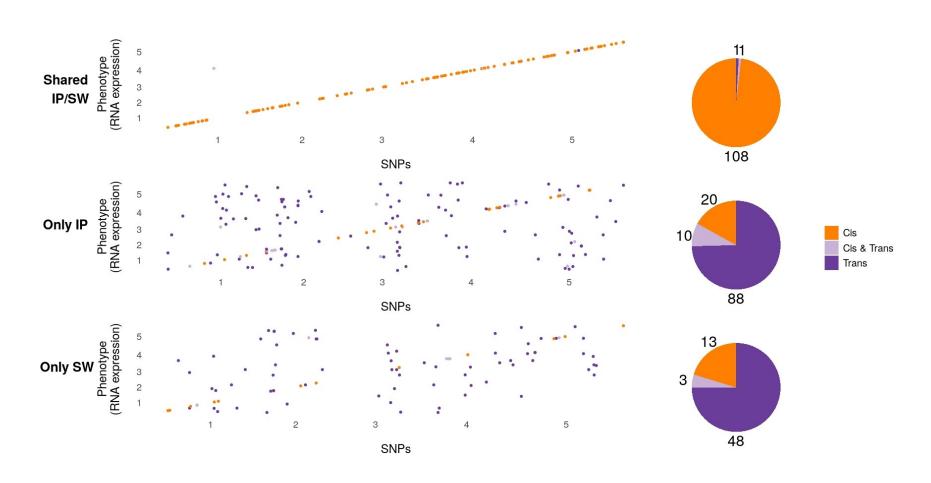


# eGWAS in different non-local populations





## eGWAS in different local populations





# **Summary I**

What you conclude from GWAS will depend on where you sample True for humans as well (e.g. genetic architecture for skin color differ dramatically between Iceland and Cape Verde Islands)



# **Summary II**

Can we infer how local adaptation works?

Putative local adaptive regulatory alleles are mostly regulated in *trans* 



# Genomic variation in the A. thaliana population (1135 accessions)

10,709,466 SNPs segregate in the population

1,854,599 SNPs are located in coding regions

28,148 SNPs lead to a premature STOP codon



Nearly 10,000 genes are knock-out in at least 1 accession



### **Co-occurence of premature Stop codons**

Gene 1	Gene 2	gene1_count	gene2_count	Co - occurrence	Lower border	Upper border	P-value
AT1G66920	AT3G59750	341	251	122	100	157	0.03
AT4G36140	AT5G45150	312	356	266	138	196	1.7e-57
AT3G05685	AT3G63320	444	114	38	55	96	5.6e-16

Many genes are knocked-out together more (or less) often than expected



# **Co-occurence of premature Stop codons**

	prematSTOP
Knocked-out together more often (over)	2917
Knocked-out together less frequently (under)	138



## **Co-occurence of premature Stop codons**

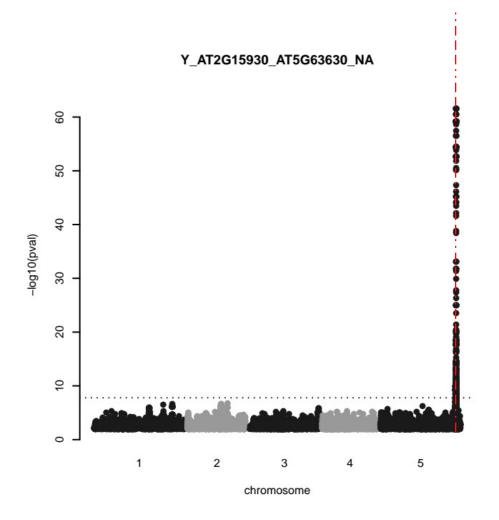
	prematSTOP	SYN	NON_SYN
Knocked-out together more often (over)	2917	1216	1086
Knocked-out together less frequently (under)	138	46	59

# More connections for functional variants compared to (potential) neutral variants

Anwar et al. In prep.



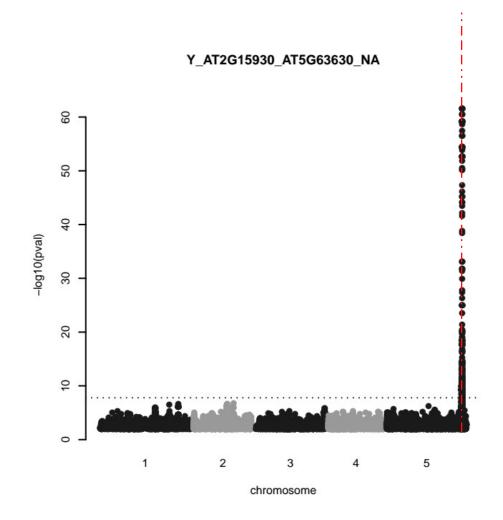
Create a binary trait (0|1) which indicates if in a given accessions both genes contain functional premature STOP codons or if only one gene is knocked out





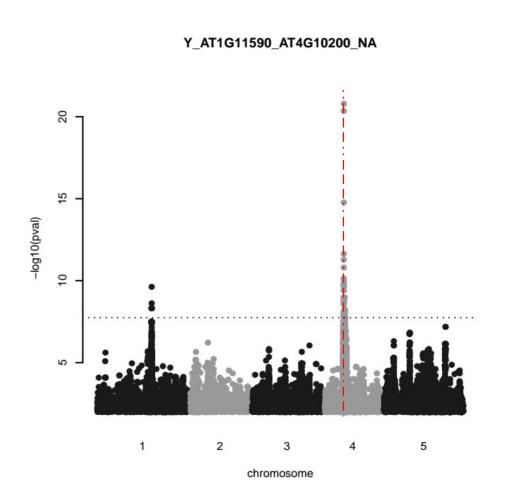
Create a binary trait (0|1) which indicates if in a given accessions both genes contain functional premature STOP codons or if only one gene is knocked out

Signal in cis = expected ( 5569/5834 for over and 209/276 for under)





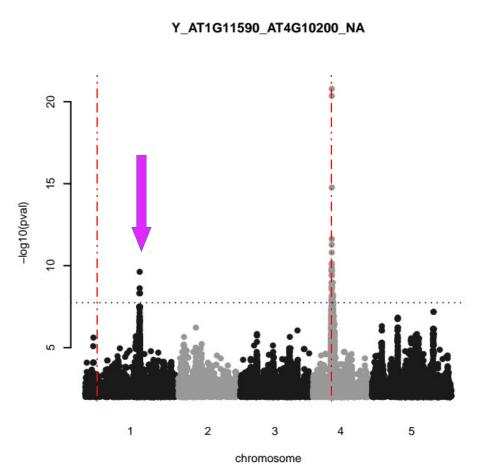
Create a binary trait (0|1) which indicates if in a given accessions both genes contain functional premature STOP codons or if only one gene is knocked out





Create a binary trait (0|1)
which indicates if in a
given accessions both
genes contain functional
premature STOP
codons or if only one
gene is knocked out

Additional signal in trans (212/5834 for over and 8/276 for under)

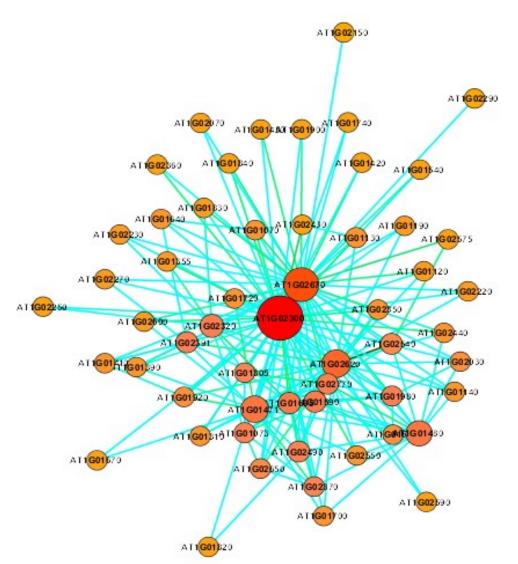


We can map genes that are responsible for the co-occurence

Anwar et al. In prep.



### Network of genes that are knocked-out together





In collaboration with Pascal Braun (INET, HMGU)



# **Summary III**

Tremendous amount of Natural Variation in signaling pathways

If we want to understand G-P, we need to understand the underlying gene regulatory networks





