

Fabric-elasticity Relationships of Femoral Head Trabecular Bone are Similar in Type 2 Diabetes and Non-Diabetic Individuals

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ABSTRACT

Type 2 diabetes (T2D) is a chronic disease leading to an elevated glucose level in the blood and increased fracture risk. Because T2D individuals tend to have normal to higher areal bone mineral density (aBMD) than healthy individuals, their fracture risk is often underestimated. As an alternative, high-resolution peripheral quantitative computed tomography (HR-pQCT) is an attractive tool to investigate bone morphology in vivo and estimate fracture risk. Based on HR-pQCT scans, bone strength can be estimated using micro finite element (μ FE) analysis or homogenised finite element (hFE) analysis. While μ FE is computationally expensive, hFE provides an accurate estimation of bone mechanical properties within reasonable efforts. However, the hFE scheme is based on relationships between the local fabric (anisotropy) and elasticity. These relationships have been shown to hold for healthy controls as well as in the case of osteogenesis imperfecta. Nevertheless, whether these relationships are also valid for T2D-diagnosed patients remains unclear. Therefore, the present work aims to compare fabric-elasticity relationships between T2D and non-diabetic controls.

The present study collected 56 trabecular bone cores from the femoral head of 28 T2D and 28 control donors. These cadaveric samples were scanned in a micro-CT system at an isotropic 14.8 μ m voxel size. Three cubic regions of interest (ROIs) were selected in each scan. The resolution of these ROIs was downsampled by a factor of 4, mimicking clinical HR-pQCT resolution, and the ROIs were subsequently segmented. Standard morphometric parameters were computed from the segmented ROIs using medtool (v4.8; Dr. Pahr Ingenieure e.U., Pfaffstätten, Austria). Additionally, their fabric tensor and their apparent stiffness tensors were computed. The ROIs were compared between T2D and control regarding their morphometric and mechanical properties. Finally, ROIs were matched between T2D and control for bone volume fraction (ρ) and degree of anisotropy. The matched dataset allowed the comparison of fabric-elasticity relationships between T2D and control samples.

No significant difference was observed between T2D and control samples, both regarding their morphology and their mechanical properties. Specifically, fabric-elasticity relationships were shown to hold for both the control and the T2D groups. A comparison of the resulting exponents related to ρ and DA has highlighted weakly different trends but no significant difference between T2D and control samples.

In conclusion, trabecular bone architecture was similar between T2D and non-T2D donors. Additionally, fabric-elasticity relationships, i.e. morphology-mechanical relationships, are also similar between donors with and without diabetes. Accordingly, HR-pQCT-based hFE analysis could also be used for estimating the bone mechanical properties of T2D patients and for their fracture risk assessment.

1. Introduction

Fragility fractures are, by definition, bones that break under a load which shouldn't have caused a fracture in healthy conditions [14]. Such fractures are a worldwide burden, causing high morbidity, mortality, and high costs for the health care system [5, 13]. The lifetime risk of fragility fracture is relatively high, lying between 40-50% for women and 13-22% for men [12]. This fracture risk is even higher in adults diagnosed with type 2 diabetes (T2D) [29, 2].

Abbreviations: AGEs, advanced glycation endproducts ROI, region of interest; μ CT, micro-computed tomography; Type 2 diabetes, T2D; High-resolution peripheral quantitative computed tomography, HR-pQCT; Areal bone mineral density, aBMD; Dual-energy X-ray absorptiometry, DXA; Finite elements analysis, FEA; Trabecular thickness, Tb.Th.; Trabecular spacing, Tb.Sp.; Trabecular number, Tb.N.; Mean intercept length, MIL; Degree of anisotropy, DA; Coefficient of variation, CV

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T2D is a chronic disease where cells become insulin resistant, leading to a lower glucose uptake by the cell and thus, an elevated glucose level in the blood [25]. T2D conditions also lead to an increase of advanced glycation endproducts (AGEs) in bone collagen [18]. AGEs is shown to interfere with reducing bone formation [1] and bone resorption [34] leading to a reduced bone turnover. Additionally, AGEs form non-enzymatic cross-links across and within collagen fibers, leading to more brittle bone matrix and decreasing toughness [33]. Thus, T2D can lead an increased bone brittleness and potentially also reduced quality [26].

Currently, fracture risk is usually assessed based on areal bone mineral density (aBMD) from dual-energy X-ray absorptiometry (DXA) measurement performed either at the lumbar spine or the femoral neck [20]. However, T2D patients tend to have normal to higher aBMD than healthy individuals [17]. This trend might arise from the fact that T2D

patients tend to present a higher body mass index, thus, triggering an osteogenic response. Therefore, T2D-associated skeletal fragility is underestimated in clinics [27].

An alternative to DXA is high-resolution peripheral quantitative computed tomography (HR-pQCT). Indeed, as opposed to DXA, which provides areal size-dependent measurement, HR-pQCT provides 3-dimensional size-independent quantitative measurement [36]. The high resolution provided by HR-pQCT allows for the analysis of the trabecular and cortical bone phases separately. Additionally, HR-pQCT images can be used as a basis to perform finite element analysis (FEA), allowing the estimation of local bone mechanical properties, which can, in turn, be used for fracture risk assessment [4]. FEA can either be performed using so-called micro FE (μ FE) approach or homogenised FE (hFE). While μ FE convert each voxel to a hexahedral element, thus leading to high computational costs, hFE makes use of local bone volume fraction (ρ) and anisotropy (fabric) to assess bone properties within a reasonable computational effort [23]. Evidences have shown high correlations between hFE prediction and experimental tests on fresh frozen samples [35, 10, 3, 28, 32]. Thus, HR-pQCT-based hFE can complement DXA in fracture risk estimations. However, hFE relies on bone quality at the structural level, i.e. fabric-elasticity relationships developed for functionally adapted bone [38]. It was shown that these fabric-elasticity relationships are valid for the trabecular bone in the radius, the vertebra, and the femur [9, 24] and hold even in the case of osteogenesis imperfecta (OI) [31]. Thus, the present study aims to compare the trabecular bone microstructure of non-diabetic and T2D bone samples and test the hypothesis of similar fabric-elasticity relationships. Similar fabric-elasticity relationships will allow to further extend the application of HR-pQCT-based hFE to T2D bone at least in the linear elastic regime.

2. Material and Methods

This technical note can be seen as an extension of previously published work on healthy and OI bones [31]. Thus, most of the methods are similar and will be summarized here.

2.1. Participants, Samples, and Imaging

The control (Ctrl) and diabetic (T2D) groups consisted of 28 donors each. The control group was composed of 14 males and 14 females aged between 51 and 97 years old at death with a mean age of 73 ± 13 (mean \pm standard deviation). The diabetic group also counted 14 males and 14 females aged between 54 and 97 years old at death, with a mean age of 75 ± 13 . **Here we would like to have a description of the diabetic conditions like type, glucose level, duration, and treatment. The idea is to describe the clinical info that we have about the two cohorts and cite the ones that we don't have in the limitations** A cylindrical sample of about 10 mm in diameter and 15 mm in height was collected from each donor's femoral head (left or right). After harvesting, samples were flushed with distilled water to remove excess marrow and embedded in polymethylmethacry-

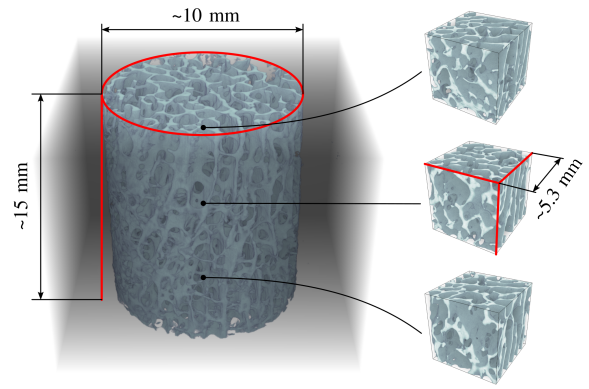


Figure 1: 3-dimensional rendering of a typical trabecular core sample and three cubic regions of interest.

late (PMMA). The embedded samples were imaged by micro-computed tomography (μ CT50, Scanco Medical AG) at an isotropic voxel size of $14.8 \mu\text{m}$ with standardized scanning settings (voltage of 60 kVp, 900 μA , 100 ms integration time).

2.2. Region of Interest

In each scanned sample, three cubic regions of interest (ROIs) of 5.3 mm were selected. For this, the image was divided into three stacks of 5.3 mm (top, centre, and bottom), and a ROI was selected at the centre of mass of the stack. A 3-dimensional rendering of a typical sample and its three cubic ROIs is shown in Figure 1.

2.3. Morphological Analysis

Image analysis was performed using medtool (v4.8; Dr. Pahr Ingenieure e.U., Pfaffstätten, Austria) The pipeline was defined as segmentation, cleaning, and morphometry. The segmentation was performed using a single threshold based on the average Otsu threshold [21] of all the scans. The cleaning step consisted of removing isolated islands resulting from the single threshold segmentation. Then, standard trabecular morphometric parameters were computed. Namely, the bone volume fraction (ρ), trabecular thickness (Tb.Th.), trabecular spacing (Tb.Sp.), and trabecular number (Tb.N.). Additionally, the fabric tensor \mathbf{M} was computed using the mean intercept length (MIL) method [19]. The ROI's degree of anisotropy (DA) was computed by dividing the fabric tensor's highest eigenvalue by the lowest. A last morphological parameter, the coefficient of variation (CV), assessing the homogeneity of mass distribution within the ROI was computed as defined by Panyasantisuk et al. [24].

2.4. Numerical Analysis

After morphological analysis, each ROI underwent numerical homogenization. For this, μ FE analyses were performed using ABAQUS 2023. The model was built using a direct voxel conversion approach to fully integrated linear brick elements (C3D8). Each element was assigned a Young's modulus of 10 GPa and a Poisson's ratio of 0.3. The ho-

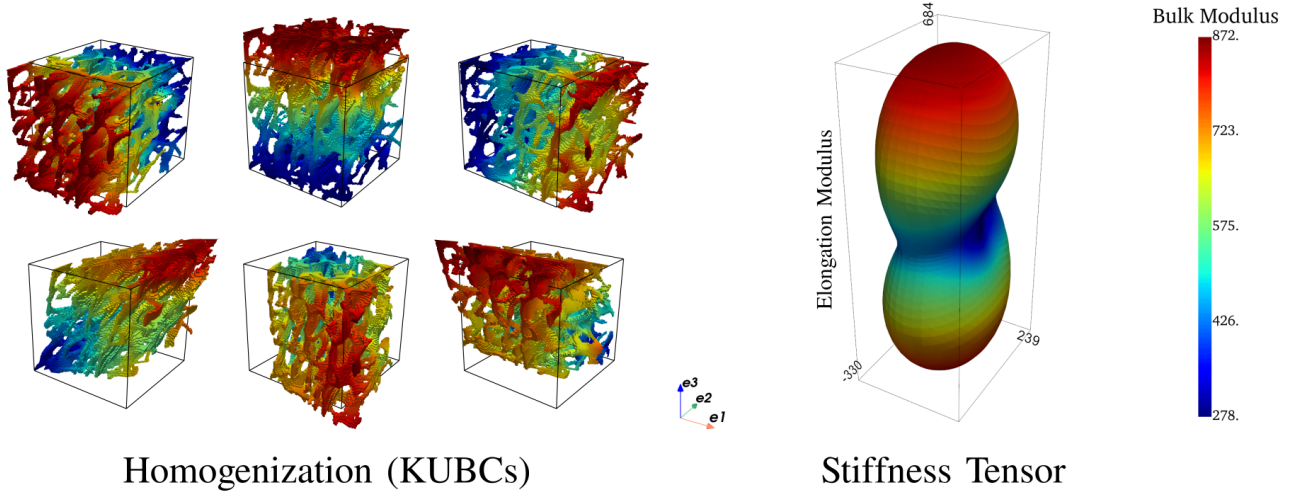


Figure 2: Schematic representation of the homogenization process leading to the ROI's stiffness tensor. Note that the deformation is amplified by multiple orders of magnitude in the illustration.

mogenization scheme was composed of three uni-axial tests and three pure shear tests using kinematic uniform boundary conditions (KUBCs) [24]. Then, the ROI's homogenized stiffness tensor \mathbb{S} was computed from these six independent loadcases. This step is illustrated in Figure 2. Finally, the resulting stiffness tensor was transformed into the fabric coordinate system and projected onto orthotropy, leading to 12 non-zero symmetric components.

2.5. Group Comparison

The first comparison between the control and diabetic groups was regarding their morphology. For this, the Mann-Whitney test was performed on samples having a bone volume fraction (ρ) lower than 0.5. Indeed, a ρ higher than 0.5 cannot really be considered as purely trabecular bone. A p-value lower than 0.05 was considered meaning a significant difference between the groups.

The second comparison was regarding the stiffness tensor components after transformation into the fabric coordinate system and projection onto orthotropy. Mann-Whitney test was again used, and a p-value lower than 0.05 was considered significant. This comparison was performed on samples with a $\rho < 0.5$ and a coefficient of variation (CV) lower than 0.263. Indeed, this CV threshold was determined by Panyasantisuk et al. [24] as a limit for the homogeneity assumption. A higher CV involves heterogeneous mass distribution, which violates the representative volume element (RVE) homogeneity assumption [7] underlying our calculations.

The third comparison of the two groups was performed by fitting the orthotropic stiffness tensor to the Zysset-Curnier model [37] and comparing the resulting parameters and their 95% confidence interval (95% CI). Briefly, this model expresses the stiffness tensor \mathbb{S} based on the bone volume fraction ρ , fabric tensor eigenvalues $m_1 < m_2 < m_3$, three stiffness constants λ_0 , λ'_0 , and μ_0 and two exponents k and l . The

fitting procedure consisted of a multiple linear regression which was performed on the logarithmic space as shown in Equation 1, where λ_{ij} and μ_{ij} are the components of the stiffness tensor, $\lambda^* = \lambda_0 + 2\mu_0$, and ϵ_i the residuals. However, as k and l are exponents, it is necessary to impose values for further fabric-elasticity relationship comparison. These imposed values can either be on the exponents (as done in previous work [31]) or on the stiffness constants. In the present technical note, it was chosen to impose values for λ_0 , λ'_0 , and μ_0 , thus assuming identical tissue properties, and compare the resulting k and l . The imposed values were determined by performing the fit on the control and diabetic groups pooled together.

$$\ln \begin{pmatrix} \lambda_{11} \\ \lambda_{12} \\ \lambda_{13} \\ \lambda_{21} \\ \lambda_{22} \\ \lambda_{23} \\ \lambda_{31} \\ \lambda_{32} \\ \lambda_{33} \\ \mu_{23} \\ \mu_{31} \\ \mu_{12} \end{pmatrix} = \begin{pmatrix} 1 & 0 & 0 & \ln(\rho) & \ln(m_1^2) \\ 0 & 1 & 0 & \ln(\rho) & \ln(m_1 m_2) \\ 0 & 1 & 0 & \ln(\rho) & \ln(m_1 m_3) \\ 0 & 1 & 0 & \ln(\rho) & \ln(m_2 m_1) \\ 1 & 0 & 0 & \ln(\rho) & \ln(m_2^2) \\ 0 & 1 & 0 & \ln(\rho) & \ln(m_2 m_3) \\ 0 & 1 & 0 & \ln(\rho) & \ln(m_3 m_1) \\ 0 & 1 & 0 & \ln(\rho) & \ln(m_3 m_2) \\ 1 & 0 & 0 & \ln(\rho) & \ln(m_3^2) \\ 0 & 0 & 1 & \ln(\rho) & \ln(m_2 m_3) \\ 0 & 0 & 1 & \ln(\rho) & \ln(m_3 m_1) \\ 0 & 0 & 1 & \ln(\rho) & \ln(m_1 m_2) \end{pmatrix} \begin{pmatrix} \ln(\lambda^*) \\ \ln(\lambda'_0) \\ \ln(\mu_0) \\ k \\ l \end{pmatrix} + \begin{pmatrix} \epsilon_1 \\ \epsilon_2 \\ \epsilon_3 \\ \epsilon_4 \\ \epsilon_5 \\ \epsilon_6 \\ \epsilon_7 \\ \epsilon_8 \\ \epsilon_9 \\ \epsilon_{10} \\ \epsilon_{11} \\ \epsilon_{12} \end{pmatrix} \quad (1)$$

The fit quality was assessed using the adjusted Pearson correlation coefficient squared (R_{adj}^2) and relative norm error of fourth-order tensors (NE). This relative norm error allows to quantify the accuracy of the fit. Thus, the multiple linear regression was performed in three different steps.

1. Multiple linear regression with control and diabetic pooled together

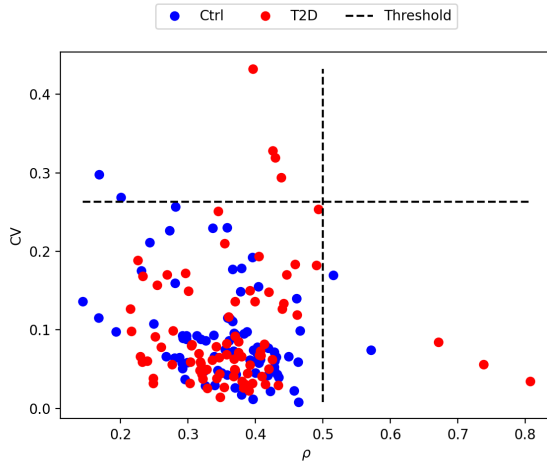


Figure 3: Coefficient of variation (CV) as function of the bone volume fraction (ρ) of the selected regions of interest (ROIs). The dashed black lines show the threshold used to filter ROIs that do not meet the model assumptions.

2. Multiple linear regression with control group or diabetic group separated allowing fit quality comparison
3. Multiple linear regression with control group or diabetic group separated and imposing λ_0 , λ'_0 , and μ_0 from step 1, allowing exponents (k and l) comparison

In order to properly compare the two groups, it is necessary to perform the multiple linear regression on similar ranges of values. In this regard, additionally to filter ROIs according to ρ and CV, a matching was performed between the ROIs of both groups for ρ and degree of anisotropy (DA), where each ROI of the diabetic group was matched to the closest control ROI regarding ρ and DA. To summarize, three subsets were used for the different comparisons:

1. Morphological: ROIs having a $\rho < 0.5$
2. Mechanical: ROIs have a $\rho < 0.5$ and $CV < 0.263$
3. Morphology-mechanical relations: diabetic-control matched ROIs having both a $\rho < 0.5$ and $CV < 0.263$

3. Results

Figure 3 shows the distribution of the ROIs selected according to the bone volume fraction (ρ) and coefficient of variation (CV). Filtering ROIs with a bone volume fraction that is too high leads to the exclusion of two controls and three diabetic ROIs. The further filtering according to CV leads to the additional exclusion of two control and four diabetic ROIs. Finally, matching ROIs for ρ and DA leads to 76 ROIs in each group.

3.1. Morphology and Mechanics

Morphological comparisons are shown in Table 1. For each of the analysed trabecular parameters, the interquartile range (0.25-0.75) presents a significant overlap between the control and diabetic groups. This overlap is confirmed by the p-value resulting from the Mann-Whitney test being higher

Table 1

Summary of the morphological parameters and comparison. The first column shows the variable assessed, the second column is the p-value resulting from the Mann-Whitney test, and the third and fourth columns show the median value for each group with their interquartile range.

Variable	p-value	Ctrl	T2D
ρ	0.77	0.37 [0.30-0.40]	0.36 [0.31-0.41]
Tb.N.	0.06	1.04 [0.96-1.12]	1.02 [0.93-1.06]
Tb.Th.	0.41	0.30 [0.28-0.32]	0.31 [0.28-0.33]
Tb.Sp.	0.09	0.66 [0.59-0.74]	0.68 [0.63-0.75]
Tb.Sp.SD	0.50	0.07 [0.07-0.08]	0.07 [0.07-0.09]
DA	0.29	1.69 [1.59-1.82]	1.66 [1.54-1.81]
CV	0.94	0.07 [0.05-0.11]	0.07 [0.05-0.14]

than the significant level of 0.05 for all computed variables. Component-wise comparisons of the stiffness tensors resulting from homogenisation are available in Appendix A. As for the morphological comparison, none of the variables compared (stiffness components) shows a significant difference (p-value always lower than 0.05) between the control and the diabetic group.

3.2. Morphology-Mechanical Relationships

The multiple linear regression fitting control and diabetic group pooled together to the Zysset-Curnier model lead to a R^2_{adj} of 0.97 and a norm error of 0.08. The fits performed on the individual groups are shown in Figure 4. The control group reaches a R^2_{adj} of 0.97 and a norm error of 0.08, as for the regression using the two groups pooled together. The multiple linear regression performed on the samples from diabetic patients alone reaches a slightly higher R^2_{adj} of 0.98 and a norm error of 0.07.

Finally, a comparison of the values obtained for the k and l exponents for restricted multiple linear regressions are shown in Figure 5. Pooling control and diabetic group together lead to k and l values of 1.7 (1.68-1.71) and 0.65 (0.64-0.67) (value and 95% CI), respectively. The separation of the groups leads to slightly higher values for the control group than for the diabetic group, but their 95% CI overlap. The control versus the diabetic group shows a k of 1.71 (1.70-1.71) versus 1.69 (1.69-1.70) and a l of 0.67 (0.64-0.69) versus 0.64 (0.62-0.66), respectively.

4. Discussion and Conclusion

This technical note investigates relationships between morphology and elasticity of trabecular bone in diabetic conditions as compared to control. This work can be seen as an extension of previous similar work [31] comparing osteogenesis imperfecta conditions to healthy controls. Briefly, the trabecular morphology of regions of interest (ROIs) from femoral head samples is analysed. Then, numerical homogenisation allows to compare the apparent stiffness of these ROIs. Finally, the fitting to a fabric-based orthotropic model [37]

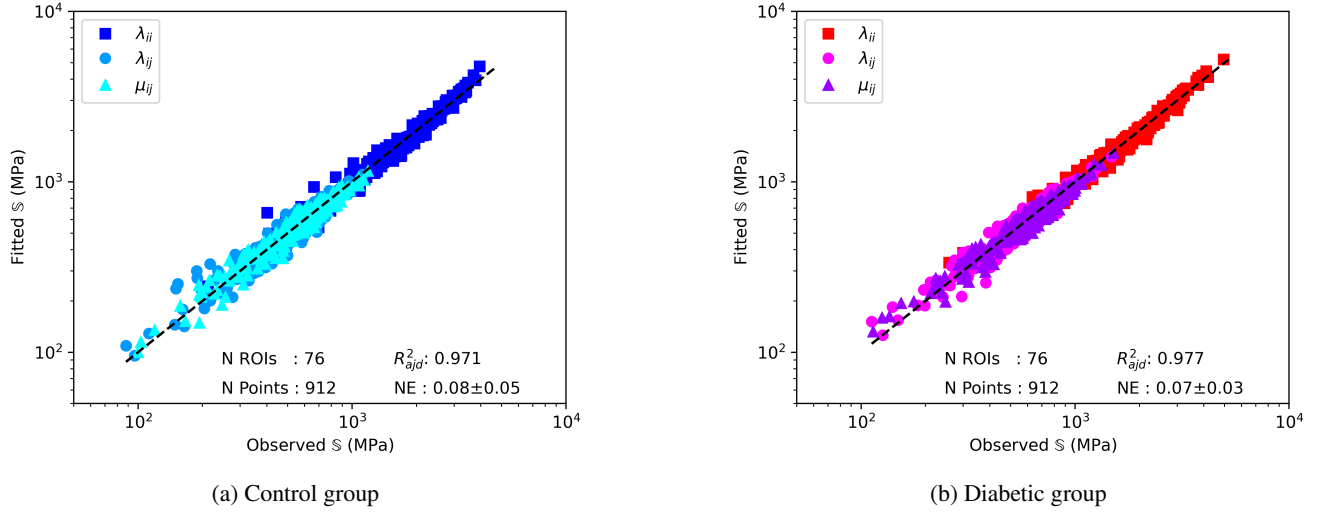


Figure 4: Results of the multiple linear regression performed on the individual groups. Observed \mathbb{S} is the stiffness tensor resulting from numerical homogenization, and fitted \mathbb{S} is the stiffness tensor predicted by the theoretical model.

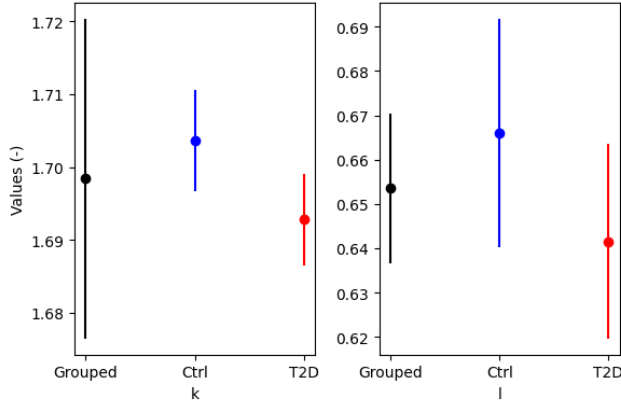


Figure 5: The resulting exponents and their 95% confidence interval of the restricted multiple linear regression performed on the grouped and individual groups.

allows comparison of fabric-elasticity, i.e. morphology-mechanical relationships between samples from the control or the diabetic group.

To be considered as trabecular bone regions, ROIs presenting a bone volume fraction (ρ) higher than 0.5 were excluded from the analysis. The presence of cortical bone within these excluded ROIs might come from the original anatomical location of the samples and, to a reduced extent, the choice of the single threshold value for segmentation. The comparison of morphological parameters between the control and diabetic groups shows no significant difference. These results mostly agree with similar morphological comparisons performed using HR-pQCT [6, 30, 8, 22, 27, 11]. While some of these studies have shown no differences between control and T2D [30, 27, 11], the significant p-values were comprised between 0.05 and 0.01 in the studies of [6][6] and Paccou et al.[22] and significant differences vanished after adjustment for body mass index (BMI) in the study of Farr

et al.[8]. In the present study, specifically ρ , Tb. Th., DA and CV are highly unlikely to be different. Thus, the morphology of femoral head trabecular bone samples presented in this work is highly similar in diabetic conditions as compared to control.

Homogenisation of the cubic ROIs requires that the bone mass is homogeneously distributed with the volume. Therefore, ROIs presenting a CV higher than a previously defined threshold of 0.263 [24] were filtered out. The component-wise comparison of the resulting stiffness tensors shows no differences between ROIs from control or diabetic patients. This result complements the morphological comparison, showing that, beyond similar morphology, the mechanical behavior resulting from the bone structural quality is similar between diabetic and control samples.

After morphological and mechanical comparison, the relationships between morphology and mechanics are compared using the Zysset-Curnier model [37]. As performed in previous work [31], ROIs are matched for ρ and DA to perform the fit on similar ranges for both groups. The fit quality assessed using R^2_{adj} and NE is in the expected range, i.e. similar as observed in other studies [9, 24, 31]. Moreover, these quality parameters are quite similar among the different data sets used for the regression (grouped, control, and diabetic), representing a first step indicating similar fabric-elasticity relationships between control and diabetic conditions. Going one step further, stiffness parameters (λ_0 , λ'_0 , and μ_0) are fixed to a common value, assuming identical bulk tissue properties, and the resulting exponents (k and k) with their 95% CI are compared. The two exponents tend to be lower for the diabetic group than the control group, but their 95% CI overlap. Thus, the existence of a common value for both groups cannot be excluded. Additionally, the 95% CI overlap for l , which is directly linked to the fabric, shows a relatively better overlap, including the value of the other group. On the other hand, the 95% CI overlap for the k exponent is

relatively worse, but it is linked to the bone volume fraction ρ . Indeed, the k exponent tends to be closer to 1 for diabetic group than for the control group, indicating even a more efficient organisation of the bone mass. While both the k and l exponents seems to deviate slightly between the groups, their value remains remarkably close. Altogether, these results allow to assume that fabric-elasticity relationships are similar between control and diabetes-diagnosed individuals.

Nevertheless, this study presents some limitations that need to be addressed. First, the sample were harvested only from the femoral head, thus limiting the generalization of the results to other anatomical locations. However, multiples studies have shown that the fabric-elasticity relationships are extremely robust and hold similarly across anatomical locations [9, 24, 31]. Second limitations are the downsampling of the original ROIs and the filtering steps. Indeed, the downsampling was performed to mimic the resolution of HR-pQCT, which is the clinical tool used for bone health assessment. This downsampling lead to bulkier structures, which might have slightly modified the morphological and mechanical properties. However, this downsampling was performed on both the control and diabetic ROIs similarly, thus not affecting the comparison between the two groups. Regarding the filtering steps, the exclusion of ROIs with a bone volume fraction higher than 0.5 is necessary to ensure that the ROIs are purely trabecular bone. Additionally, the filtering step removing highly heterogeneous ROIs is also a necessary step because, as stated in the original work [31], "homogeneity is a necessary condition underlying the computational homogenisation of mechanical properties and the results of heterogeneous samples must be disregarded". Third and finally, (some clinical data that are unknown) are not available regarding the two cohorts. However, these missing information are not thought to have a significant impact on the results of the present study.

To conclude, the present work shows that trabecular bone from the femoral head of diabetic conditions presents similar morphology, mechanics, and fabric-elasticity relationships compared to control. As mentioned in the introduction, these fabric-elasticity relationships are a basic assumption of HR-pQCT-based hFE simulations. Therefore, the present results suggest using HR-pQCT for bone health assessment and monitoring in diabetes-diagnosed patients.

Declaration of competing interest

We wish to confirm that there are no known conflicts of interest associated with this publication and there has been no significant financial support for this work that could have influenced its outcome.

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Data availability statement

The data that support the findings of this study are available on request. The data are not publicly available due to privacy/ethical restrictions. The scripts used for the analyses performed in the present study are available on Github: <https://github.com/artorg-unibe-ch/FABTIB>

Research ethics

We further confirm that any aspect of the work covered in this manuscript that has involved human patients has been conducted with the ethical approval of all relevant bodies and that such approvals are acknowledged within the manuscript.

CRediT author statement

Mathieu Simon: Data Curation, Formal analysis, Investigation, Methodology, Software, Visualization, Writing - original draft. **Sasidhar Uppuganti:** Data Curation, Resources, Writing - review and editing. **Jeffrey S Nyman:** Conceptualization, Funding acquisition, Project administration, Resources, Validation, Writing - review and editing. **Philippe Zysset:** Conceptualization, Funding acquisition, Methodology, Project administration, Resources, Supervision, Validation, Writing - review and editing.

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A. Mechanical Comparison

The results of the mechanical comparison for each non-zero component of the apparent stiffness tensor resulting from homogenisation are shown in Figure 6.

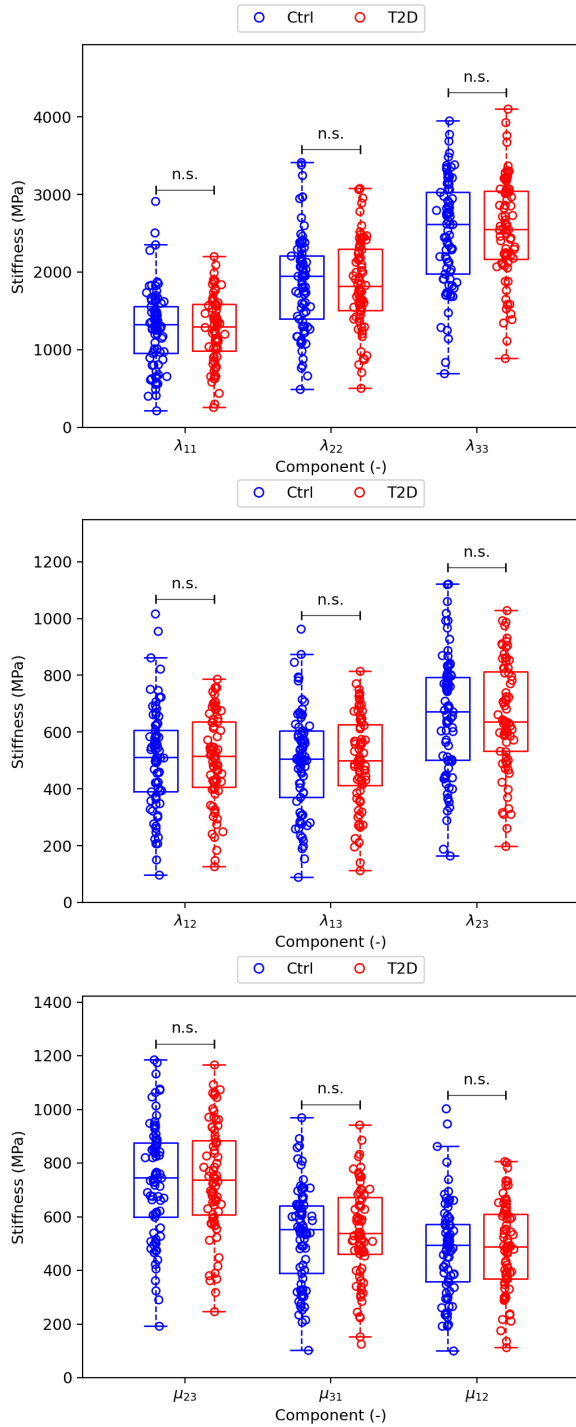


Figure 6: Comparison of the stiffness tensor components between the control and the diabetic group. N.s. stands for non-significant p-value (>0.05) resulting from Mann-Whitney test