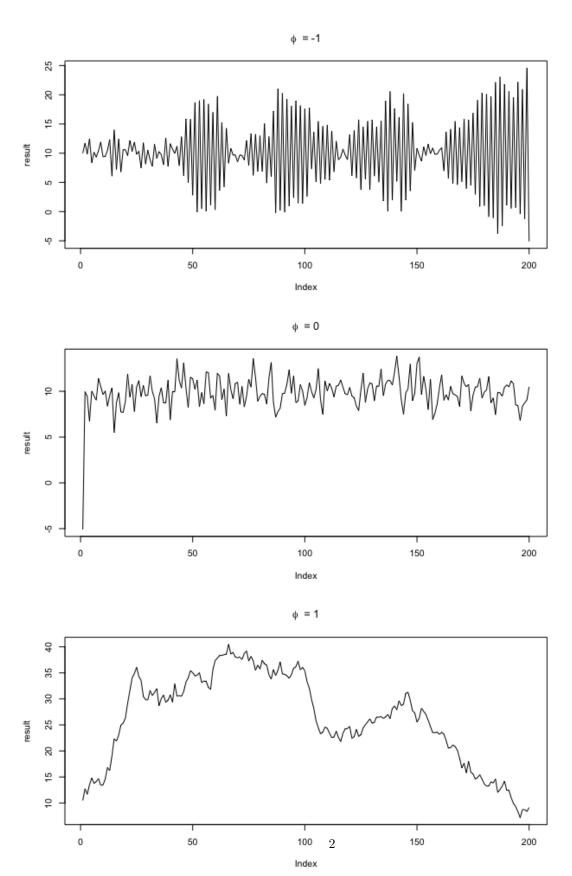
TDDE07 - Lab 4

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1 - Time series models in Stan

1a) AR(1)-process



It is clear that ϕ has a large effect on $x_{1:T}$. The figure above depicts 3 different scenarios where ϕ is equal to -1, 0 and 1. In the first example, we have that

$$x t = \mu - (x t-1 - \mu) + \epsilon_t$$

As can be seen in the figure above the values of x_t oscillate between positive and negatiive values when $\phi = -1$. This is because ϕ changes the sign of x_t in each iteration.

When $\phi = 0$ we have that

$$x t = \mu + \epsilon_t$$

In this case the value of x_t only depends on μ and ϵ_t , meaning the values will be close to μ . In the last example we have that

$$x t = \mu + (x t-1 - \mu) + \epsilon_t$$

In this case, x_t value depends on the previous value but won't change sign in each iteration as long as the error is fairly small.

1b)

Using our AR-function defined in a) resulted in the following values for the posterior mean and the upper and lower limits of the 95~% credible intreval.

phi = 0.95	Posterior mean	Lower limit	Upper limit
mu	NA	NA	NA
sigma2	2.143	1.752	2.593
phi	0.968	0.944	1.000

phi=0.30	Posterior mean	Lower limit	Upper limit
mu	10.188	9.874	10.497
sigma2	2.176	1.790	2.654
phi	0.328	0.194	0.464

The convergence of ϕ and μ can be seen by plotting the parameters in the same graph. The red dot represent the posterior mean of each parameter. The shape of the joint distribution of ϕ and μ is the same for different initial values of ϕ .

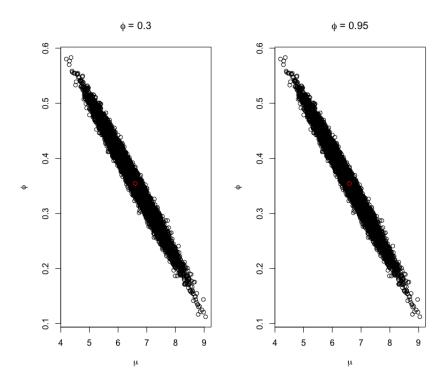


Figure 1: Convergence of parameters

The chains are mixing fairly well for $\phi=0.3$, with convergence towards the true values ($\mu=10,\sigma^2=2,\phi=0.3$), while $\phi=0.95$ results in relatively poor mixing for all but σ^2 , with no actual convergence for μ .

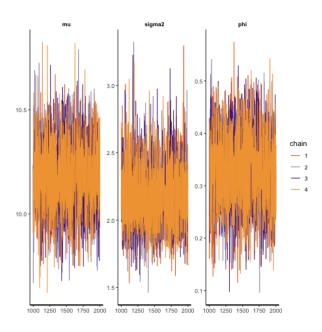


Figure 2: Sample trajectories for $\phi=0.3$

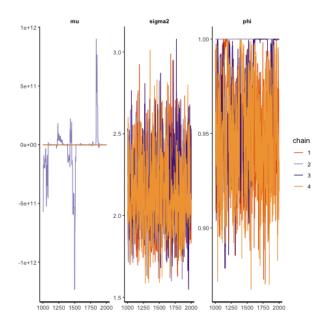


Figure 3: Sample trajectories for $\phi = 0.3$

1c)

The figure below shows a plot of the data as well as the posterior mean and a 95 % credible interval for the letaent intensity.

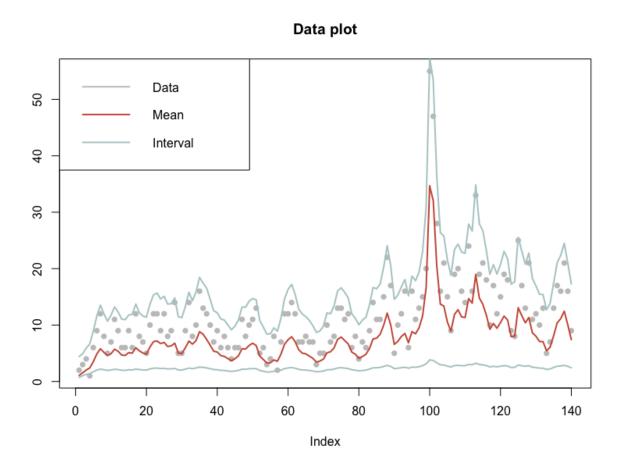


Figure 4: Task C

1d)

Here the prior for σ^2 has been modified to be informative. We set the prior to be

$$\sigma^2 \sim Gamma(1, 1000)$$

Since the prior for sigma has a mean close to 0 and low variance, the effect of the error in the regression model has decreased.

The modified prior had an effect on the posterior mean to be less sensitive to the data. This is an expected outcome since the prior, by being more informative, had a larger impact on the posterior.

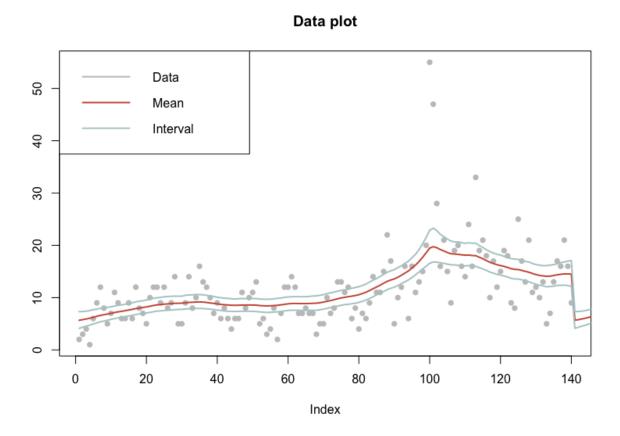


Figure 5: Task D

Code - 1

```
library('rstan')
######## A ########
mu <- 10
sigma_sq <- 2
T <- 200
x <- mu
par(mfrow=c(3,1))
AR <- function(mu, prev_x, phi, sigma_sq) {
 error <- rnorm(1, 0, sqrt(sigma_sq))</pre>
 return (mu + phi * (prev_x - mu) + error)
phis <- c(-1,0.1,0.95)
for(phi in phis) {
 result = c(x)
  for (i in 2:T) {
   x <- AR(mu, x, phi, sigma_sq)
   result <- c(result, x)
 plot(result, type = 'line', main = bquote(phi ~ ' =' ~ .(phi)))
####### B ########
mu <- 10
sigma_sq <- 2
T <- 200
x <- mu
phis <-c(0.3, 0.95)
X = c()
for(phi in phis) {
 result \leftarrow c(x)
 for (i in 2:T) {
   x <- AR(mu, x, phi, sigma_sq)
   result <- c(result, x)
 }
 X <- rbind(X, result)</pre>
model <- stan_model('StanNormalModel.stan')</pre>
warmup=1000
iter=2000
niter = 4*(iter-warmup)
```

```
fitX <- sampling(model, data = list(T=200, x=X[1,]), iter = iter, warmup = warmup)</pre>
fitY <- sampling(model, data = list(T=200, x=X[2,]), iter = iter, warmup = warmup)</pre>
# Print the fitted model
print(fitX,digits_summary=3) # Extract posterior samples
print(fitY,digits_summary=3) # Extract posterior samples
postDrawsX <- extract(fitX)</pre>
postDrawsY <- extract(fitX)</pre>
par(mfrow=c(1,2))
plot(postDrawsX$mu, postDrawsX$phi, main = expression(phi ~ '= 0.3'), ylab=expression(phi), xlab=expres
lines(mean(postDrawsX$mu), mean(postDrawsX$phi), type='p', col="red")
plot(postDrawsX$mu, type='1')
# legend('topright', legend=expression(mean(postDrawsX$mu) ~ ',' ~ mean(postDrawsX$phi)))
plot(postDrawsY$mu, postDrawsY$phi, main = expression(phi ~ '= 0.95'), ylab=expression(phi), xlab=expre
lines(mean(postDrawsY$mu), mean(postDrawsY$phi), type='p', col="red")
\# legend('topright', legend=expression(().mean(postDrawsY$mu) ~ ',' ~ ().mean(postDrawsY$phi)))
# plot convergence
png('plots/lab4_b_convergence_x.png')
traceplot(fitX)
dev.off()
png('plots/lab4_b_convergence_y.png')
traceplot(fitY)
dev.off()
######## C #########
library(ggplot2)
data <- read.table('campy.txt', header=TRUE)</pre>
N <- length(data$c)</pre>
model <- stan_model('StanPoissonModel.stan')</pre>
fit <- sampling(model, data = list(N=N, c=data$c), iter = 2000, warmup = 1000)
# Print the fitted model
print(fit,digits_summary=3) # Extract posterior samples
postDraws <- extract(fit)</pre>
x <- postDraws$x</pre>
xMean <- c()
thetaUp <- c()
thetaLow <- c()
for (i in 1:length(x[1,])) {
  xMean <- c(xMean, exp(mean(x[,i])))</pre>
  thetaUp <- c(thetaUp, exp(quantile(x[,i], probs = 0.975)))
  thetaLow <- c(thetaLow, exp(quantile(x[,i], probs = 0.025)))
# draws <- c()
```

```
# for(i in 1:140) {
# draws \leftarrow c(draws, rpois(1, exp(xMean[i])))
# }
draws <- rpois(140, exp(xMean))</pre>
plot(data$c, pch=16, col='gray77', ylab='', main='Data plot')
lines(xMean, col='lightcyan3', lwd=2)
lines(thetaUp, col='lightcyan3', lwd=2)
lines(thetaLow, col='coral3', lwd=2)
legend('topleft',legend = c('Data', 'Mean','Interval'),
       col = c('gray77', 'coral3', 'lightcyan3'), lwd=2)
# qeom_ribbon(mapping = aes(seq(1,140,1), ymin = thetaLow, ymax = thetaUp), alpha = 0.25)
# Do traceplots of the first chain
par(mfrow = c(1,1))
# plot(postDraws$mu[1:(1000)], type="l", ylab="mu", main="Traceplot")
# Do automatic traceplots of all chains
# traceplot(fit)
# Bivariate posterior plots
pairs(fit)
# plot(fit)
######## D #########
data <- read.table('campy.txt', header=TRUE)</pre>
N <- length(data$c)</pre>
model <- stan_model('StanPoissonModelD.stan')</pre>
fit <- sampling(model, data = list(N=N, c=data$c), iter = 2000, warmup = 1000)
# Print the fitted model
print(fit,digits_summary=3) # Extract posterior samples
postDraws <- extract(fit)</pre>
x <- postDraws$x
xMean <- c()
thetaUp <- c()
thetaLow <- c()
for (i in 1:length(x[1,])) {
  xMean <- c(xMean, mean(exp(x[,i])))</pre>
 thetaUp <- c(thetaUp, quantile(exp(x[,i]), probs = 0.975))
  thetaLow <- c(thetaLow, quantile(exp(x[,i]), probs = 0.025))
plot(data$c, pch=16, col='gray77', ylab='', main='Data plot')
lines(xMean, col='coral3', lwd=2)
lines(thetaUp, col='lightcyan3', lwd=2)
lines(thetaLow, col='lightcyan3', lwd=2)
```

Stan Model for b)

```
data {
  int<lower=0> T; // Number of observations
 real x[T];
parameters {
 real mu;
 real<lower=0> sigma2;
 real<lower=-1, upper=1> phi;
}
model {
 //priors
 // mu ~ normal(10,2);
 // phi ~ uniform(-1,1);
 // sigma2 ~ scaled_inv_chi_square(1,2); // Scaled-inv-chi2 with nu
 //
 //likelihood
 for (t in 2:T)
    x[t] \sim normal(mu + phi * (x[t-1] - mu), sqrt(sigma2));
```

Stan Model for c)

```
data {
  int<lower=0> N; // Number of observations
  int c[N];
parameters {
 real mu;
 real<lower=-1,upper=1> phi;
 real<lower=0> sigma;
 real<lower=0> x[N]; // Data points
transformed parameters {
  real lambda[N];
  lambda = exp(x);
}
model {
  //likelihood
  for(n in 1:N)
    c[n] ~ poisson(lambda[n]);
  //prior
 for(n in 2:N)
    x[n] \sim normal(mu + phi * (x[n-1] - mu), sigma);
}
```

Stan Model for d)

```
data {
  int<lower=0> N; // Number of observations
  int c[N];
parameters {
 real mu;
 real phi;
 real<lower=0> sigma;
 real<lower=0> x[N]; // Data points
transformed parameters {
  real lambda[N];
  lambda = exp(x);
}
model {
  //likelihood
  for(n in 1:N)
    c[n] ~ poisson(lambda[n]); // Poisson
  //prior
  mu ~ normal(10,2);
  phi ~ normal(0, 1);
  sigma ~ gamma(1,1000);
 for(n in 2:N)
    x[n] \sim normal(mu + phi * (x[n-1] - mu), sigma);
```