

IN-BIOS[9,5]000 2018

Data pre-processing

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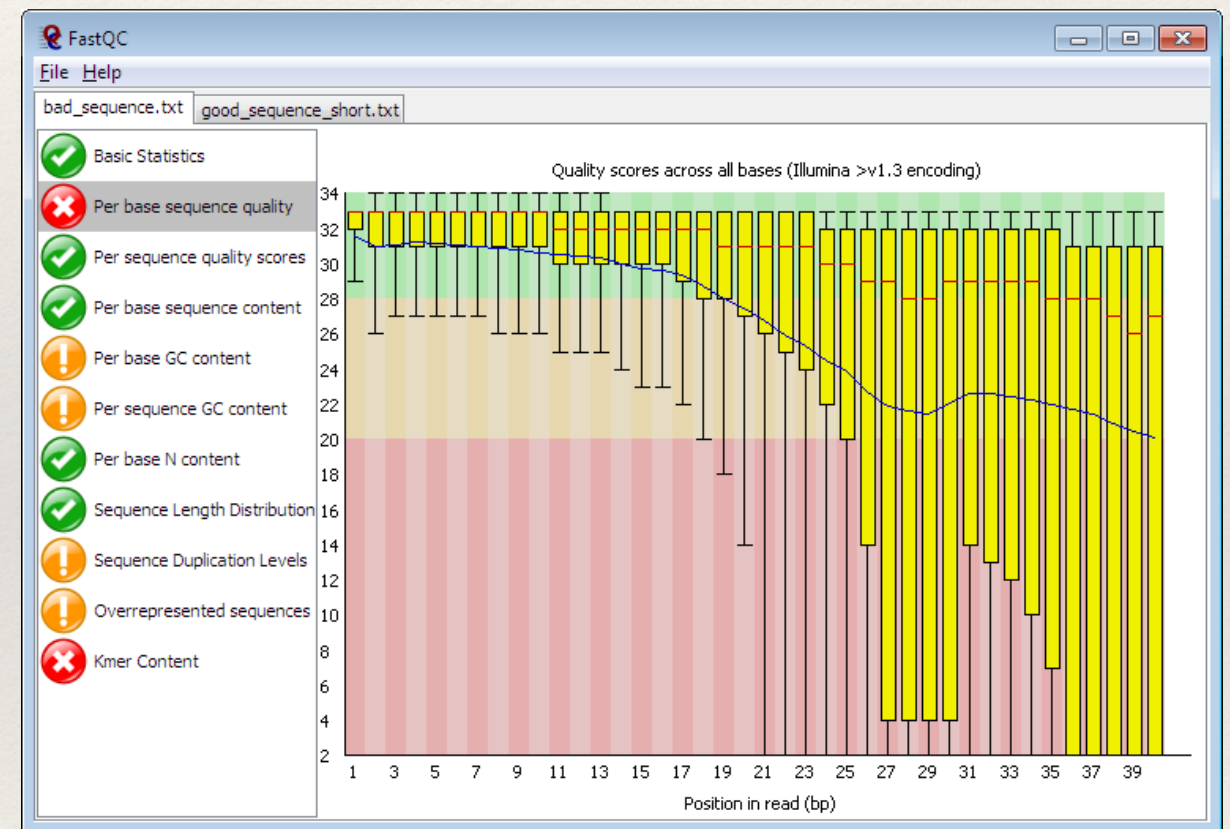
Norwegian Sequencing Centre
OUS, Ullevål, Oslo

Data pre-processing

- ❖ Quality control
- ❖ Why should we pre-process a sequence data
- ❖ Tools available
- ❖ Hands-on exercise

FastQC

- ❖ GUI, command line based
 - ❖ Import of data from BAM, SAM or FastQ files
 - ❖ Providing a quick overview to tell you in which areas there may be problems
 - ❖ Summary graphs and tables
 - ❖ HTML based permanent report
- ❖ requires Java



FastQC; MultiQC

- ❖ Video tutorial:

- ❖ <https://www.youtube.com/watch?v=bz93ReOv87Y>

- ❖ Example reports:

- ❖ <http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>

- ❖ MultiQC

- ❖ <https://www.youtube.com/watch?v=BbScv9TcaMg>

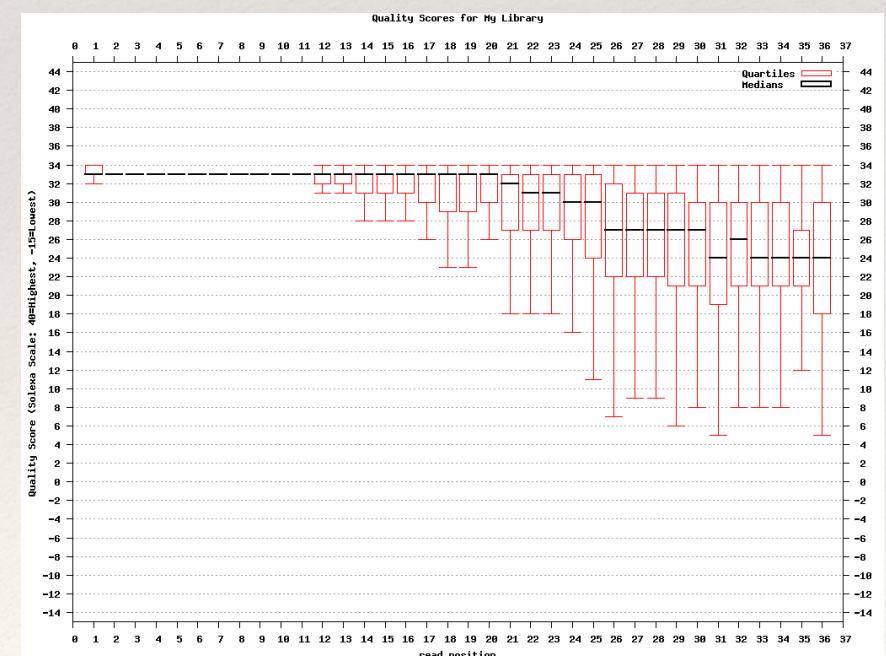
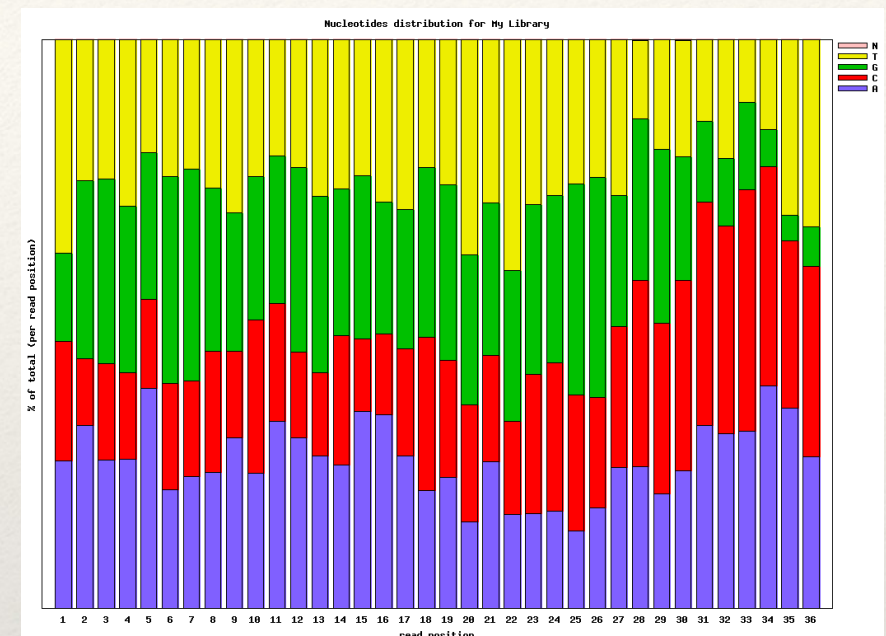
- ❖ Not just for summarising FastQC reports but much more.....

FASTX-Toolkit

- ❖ Command line tool
 - ❖ Unix-based
 - ❖ FastQ/ A short-reads pre-processing tools
 - ❖ FASTQ-to-FASTA
 - ❖ FASTQ/ A Quality Statistics
 - ❖ FASTQ Quality chart
 - ❖ FASTQ/ A Nucleotide Distribution chart
 - ❖ FASTQ/ A Clipper
 - ❖ FASTQ/ A Renamer
 - ❖ FASTQ/ A Trimmer
 - ❖ FASTQ/ A Collapser
 - ❖ FASTQ/ A Artifacts Filter
 - ❖ FASTQ Quality Filter
 - ❖ FASTQ/ A Reverse Complement
 - ❖ FASTA Formatter
 - ❖ FASTA nucleotides changer
 - ❖ FASTA Clipping Histogram
 - ❖ FASTX Barcode Splitter

FASTX-Toolkit

- ❖ Command line usage:
 - ❖ http://hannonlab.cshl.edu/fastx_toolkit/commandline.html
- ❖ Remember to use ‘-Q 33’ as a parameter



FastQC & FASTX toolkit



To do

- ❖ Run FastQC on data
 - ❖ Review the results
 - ❖ Discuss
-
- ❖ Run your preferred FASTX toolkit tool

FastQ pre-processing

- ❖ Remove/Trim adapters
- ❖ Remove/Trim low quality reads
- ❖ Remove reads from spike-ins
 - ❖ PhiX for Illumina sequencing
- ❖ Trimmomatic*
- ❖ cutadapt
- ❖ PRINSEQ
- ❖ Make sure you understand what is going on under the hood

Do this if necessary

<http://www.usadellab.org/cms/index.php?page=trimmomatic>

Trimmomatic

- ❖ Quick start:

- ❖ Paired End:

- ❖ `java -jar trimmomatic-0.35.jar PE -phred33 input_forward.fq.gz
input_reverse.fq.gz output_forward_paired.fq.gz
output_forward_unpaired.fq.gz output_reverse_paired.fq.gz
output_reverse_unpaired.fq.gz ILLUMINACLIP:TruSeq3-PE.fa:
2:30:10 LEADING:3 TRAILING:3 SLIDINGWINDOW:4:15 MINLEN:36`

- ❖ Single End:

- ❖ `java -jar trimmomatic-0.35.jar SE -phred33 input.fq.gz
output.fq.gz ILLUMINACLIP:TruSeq3-SE:2:30:10 LEADING:3
TRAILING:3 SLIDINGWINDOW:4:15 MINLEN:36`

Trimmomatic

- ❖ ILLUMINACLIP
 - ❖ Cut adapter and other Illumina-specific sequences from the read
 - ❖ Adapter file location
- ❖ SLIDINGWINDOW
 - ❖ Perform a sliding window trimming, cutting once the average quality within the window falls below a threshold.
- ❖ LEADING
 - ❖ Cut bases off the start of a read, if below a threshold quality
- ❖ TRAILING
 - ❖ Cut bases off the end of a read, if below a threshold quality
- ❖ CROP
 - ❖ Cut the read to a specified length
- ❖ HEADCROP
 - ❖ Cut the specified number of bases from the start of the read
- ❖ MINLEN
 - ❖ Drop the read if it is below a specified length

Trimmomatic



To do

- ❖ Run trimmomatic on paired end data
- ❖ Review the results
- ❖ Discuss