2D Visualization of Immune System Cellular Protein Data by Nonlinear Dimensionality Reduction

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Abstract

We present in this paper a way to effectively visualize multi-dimensional immune system cellular data by means of nonlinear methods. We find that Stochastic Neighbor Embedding (SNE), and it's variations, t-SNE and s-SNE, to be most effective at successfully mapping clusters of points into a two dimensional embedding space while preserving both the structure between similar points and the disparity between different clusters. Using a centroid-based metric that relabels points according to the cluster centroid to which they are closest, we conclude that SNE works significantly better than linear and spectral methods. In additional, by using an optimization approach for SNE similar to Newton's Method, but with the Hessian of the objective function, $\nabla^2(E)$, replaced by its first term, we are able to run the SNE varients and EE two orders of magnitude faster than with standard optimization.

1. Introduction

1.1. Immune Cell Data

In the field of cancer immunology, scientists use the protein content of immune system cells as a way to identify a cells corresponding type. For example, immune system cells, which are contained in bone marrow, are comprised of a variety of cell types, and to a

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large degree are uniquely identifiable by the proteins they contain. Highly sophisticated methods have been developed that process cells and return information on the types and quantities of proteins expressed in those cells. This data can then be viewed by an expert in the field and categorized. The laborious process of viewing the different dimensions of protein expression and categorizing a cell's type is known as gating. Figure 1.1, below, taken from (Amir et al., 2013) shows this graphically.

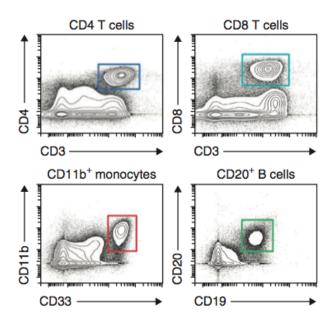


Figure 1. Strategy for cell gating: Two single dimensions of a cell are viewed at a time and through an iterative process the cell is classified

1.2. Project Goals and Metrics

It is of interest to cancer immunologists to find structure within multi-dimensional protein expression space and map it onto a lower dimensions (refered to henceforth as a map space) for ease of visualization and understanding. As cells change and evolve, so too do the types and quantities of the proteins they express. This leads to a shifting of their representation in multi-dimensional space which can be tracked. Dimensionality reduction of original biological data coupled with a metric for how well the projection represents the original data would provide biologists with a powerful tool for understanding the structure of their data. To address these challenges we demonstrate:

- The application of linear and non-linear methods of dimensionality reduction of multi-dimensional protein data
- A metric-based comparison of how each algorithm performs

2. Data Representation

2.1. Data Acquisition

Mass cytometry is a single-cell multiparametric protein detection technology based on inductively coupled plasma mass spectrometry. It is an extension of flow cytometry in which antibodies are tagged with isotopically pure rare earth elements allowing simultaneous measurement of greater than 40 parameters while circumventing the issue of spectral overlap. In single-cell droplet form, the cells are passed through an elemental mass spectrometer and an integrator to generate an $m \times p$ matrix where m is the number of cells processed and p is the number of distinct proteins contained in the cell set. These matrices are stored as .FCS files in online databanks which we have been granted access to. Figure 2.1 shows this process.

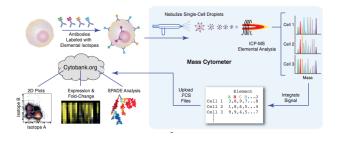


Figure 2. Overview of data acquisition, from extraction of cellular protein counts to storage in online databanks

Table 1. Classification of Cell Types and their Corresponding Sub-Types

CELL TYPES	Sub-types		
STEM CELL	HSC, MPP, CMP, GMP, MEP		
B Cells	PLASMA, PRE-B-I, PRE-B-II, IMMATURE, MATURE CD38 LOW, MATURE CD38 MID		
T Cells	Mature CD4+, Mature CD8+ Naive CD4+, Naive CD8+		
NK	-		
PDC	-		
Monocytes	CD11B - , CD11B HIGH, CD11B MID		

Table 1 lists the cell types and subtypes that are parsed using this method.

2.2. Feature Selection

The p=41 protein counts collected for each cell that passes through mass cytometry is comprised of both intracellular and surface proteins. These two types play fundamentally different roles in cell identification. Surface proteins are semi-permanent markers that last for significant periods of time relative to the lifetime of a cell, whereas intracellular proteins are highly transient and can change quickly. This is analogous to classifying a person based on where they live (semi-permanent) versus what they wore on a particular day (transient). Understanding this, we select as our feature space the n=17 surface protein markers of the cell data.

3. Methods

The datafiles provided contain cell counts on the order of tens of thousands where we consider each cell to be a point in \mathbb{R}^n . To simplify our algorithms and account for matrix size differences in the difference .FCS files we run our algorithms on equally sized portions of different cell data. In particular, if we let \mathbb{S} be the set of all cell sub-types as defined in Table 1, $S \in \mathbb{S}$ be some subset of interest with cardinality |S|, and N some fixed positive integer, then by taking N rows from each sub-type $s \in S$ we form a matrix $M \in \mathbb{R}^{N|S| \times n}$ on which we can run algorithms quickly and without giving unfair weighting to a particular cell sub-type.

We consider various algorithms which project sets of data in \mathbb{R}^n onto \mathbb{R}^2 for easy visualization.

- 3.1. Linear Methods
- 3.2. Spectral Methods
- 3.3. Nonlinear Methods

4. Electronic Submission

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dvips -Ppdf -tletter -G0 -o paper.ps paper.dvi ps2pdf paper.ps

Note that it is a zero following the "-G". This tells dvips to use the config.pdf file (and this file refers to a better font mapping).

Another alternative is to use the **pdflatex** program instead of straight LATEX. This program avoids the Type-3 font problem, however you must ensure that all of the fonts are embedded (use **pdffonts**). If they are not, you need to configure pdflatex to use a font map file that specifies that the fonts be embedded. Also

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We will continue the ICML tradition in which the authors are given the option of providing a short reaction to the initial reviews. These reactions will be taken into account in the discussion among the reviewers and area chairs.

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The final versions of papers accepted for publication should follow the same format and naming convention as initial submissions, except of course that the normal author information (names and affiliations) should be given. See Section 5.3.2 for details of how to format this.

The footnote, "Preliminary work. Under review by the International Conference on Machine Learning (ICML). Do not distribute." must be modified to "Appearing in *Proceedings of the 29th International Conference on Machine Learning*, Edinburgh, Scotland, UK, 2012. Copyright 2012 by the author(s)/owner(s)."

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The text of the paper should be formatted in two columns, with an overall width of 6.75 inches, height of 9.0 inches, and 0.25 inches between the columns. The left margin should be 0.75 inches and the top margin 1.0 inch (2.54 cm). The right and bottom margins will depend on whether you print on US letter or A4 paper, but all final versions must be produced for US letter size.

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The paper title should be set in 14 point bold type and centered between two horizontal rules that are 1 point thick, with 1.0 inch between the top rule and the top edge of the page. Capitalize the first letter of content words and put the rest of the title in lower case.

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You can use footnotes¹ to provide readers with additional information about a topic without interrupting the flow of the paper. Indicate footnotes with a number in the text where the point is most relevant. Place the footnote in 9 point type at the bottom of the column in which it appears. Precede the first footnote in a column with a horizontal rule of 0.8 inches.²

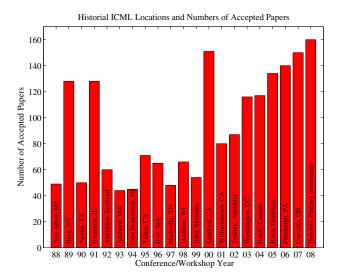


Figure 3. Historical locations and number of accepted papers for International Machine Learning Conferences (ICML 1993 – ICML 2008) and International Workshops on Machine Learning (ML 1988 – ML 1992). At the time this figure was produced, the number of accepted papers for ICML 2008 was unknown and instead estimated.

 $^{^{1}\}mathrm{For}$ the sake of readability, footnotes should be complete sentences.

²Multiple footnotes can appear in each column, in the same order as they appear in the text, but spread them across columns and pages if possible.

Algorithm 1 Bubble Sort Input: data x_i , size mrepeat Initialize noChange = true. for i = 1 to m - 1 do if $x_i > x_{i+1}$ then Swap x_i and x_{i+1} noChange = falseend if end for until noChange is true

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Table 2. Classification accuracies for naive Bayes and flexible Bayes on various data sets.

Data set	NAIVE	FLEXIBLE	Better?
Breast	95.9 ± 0.2	96.7 ± 0.2	\checkmark
CLEVELAND	83.3 ± 0.6	80.0 ± 0.6	×
Glass2	61.9 ± 1.4	83.8 ± 0.7	\checkmark
Credit	74.8 ± 0.5	78.3 ± 0.6	·
Horse	73.3 ± 0.9	69.7 ± 1.0	×
Meta	67.1 ± 0.6	76.5 ± 0.5	\checkmark
Pima	75.1 ± 0.6	73.9 ± 0.5	
VEHICLE	44.9 ± 0.6	61.5 ± 0.4	\checkmark

should be flush left.

Tables contain textual material that can be typeset, as contrasted with figures, which contain graphical material that must be drawn. Specify the contents of each row and column in the table's topmost row. Again, you may float tables to a column's top or bottom, and set wide tables across both columns, but place two-column tables at the top or bottom of the page.

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Authors should cite their own work in the third person in the initial version of their paper submitted for blind review. Please refer to Section 5.3 for detailed instructions on how to cite your own papers.

Use an unnumbered first-level section heading for the references, and use a hanging indent style, with the first line of the reference flush against the left margin and subsequent lines indented by 10 points. The references at the end of this document give examples for journal articles (Bendall et al., 2011), conference publications (Bendall et al., 2011), book chapters (Bendall

et al., 2011), books (Bendall et al., 2011), edited volumes (Bendall et al., 2011), technical reports (Crammer & Singer, 2002), and dissertations (Bendall et al., 2011).

Alphabetize references by the surnames of the first authors, with single author entries preceding multiple author entries. Order references for the same authors by year of publication, with the earliest first. Make sure that each reference includes all relevant information (e.g., page numbers).

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